Genomic structural equation modeling of impulsivity and risk-taking traits reveals three latent factors distinctly associated with brain structure and development

Mari Shishikura[1], Lang Liu[1,2], Eric Yu[1,2], Rui Zhu[1,2], Laura Vilar-Ribó[3], Renata B Cupertino, Abraham A. Palmer[3,4], Sandra Sanchez-Roige, Uku Vainik[6,7], Filip Morys[1], Ziv Gan-Or[1,2,8], Bratislav Misic[1], Alain Dagher[1,8]

1. Montreal Neurological Institute, McGill University, Montreal, Canada
2. Department of Human Genetics, McGill University, Montreal, Canada
3. Department of Psychiatry, University of California San Diego, La Jolla, CA, USA
4. Institute for Genomic Medicine, University of California San Diego, La Jolla, CA, USA
5. Division of Genetic Medicine, Department of Medicine, Vanderbilt University, Nashville, TN, USA
6. Institute of Psychology, University of Tartu, Tartu, Estonia
7. Institute of Genomics, University of Tartu, Tartu, Estonia
8. Department of Neurology and Neurosurgery, McGill University, Montreal, Canada

Abstract

Background

Impulsivity is a multifaceted transdiagnostic trait that emerges in childhood. Research has identified genetic loci and brain systems associated with different facets of impulsivity and risk-taking. However, how these genetic underpinnings overlap across different facets, and how they are associated with brain development during childhood remain unknown.

Methods

Using genomic structural equation modeling on 17 impulsivity and risk-taking traits, we identified latent factors capturing overlapping genetic architecture. We then calculated polygenic scores for these factors using Adolescent Brain Cognitive Development Study data (N = 4,142) and examined their associations with brain structure, development, and behavior in children aged 9-14 years. Finally, we tested whether socioeconomic status modulated the associations between latent polygenic scores and brain structures.

Results

We identified three distinct genetic latent factors, which we label lack of self-control, reward drive, and sensation seeking. In children, polygenic scores for the three factors showed associations with distinct brain patterns: lack of self-control associated with reduced prefrontal cortical thickness, reward drive with increased subcortical cellular density, and sensation seeking with increased cortical surface area and white matter integrity. Longitudinally, lack of self-control predicted slower white matter development. The association between polygenetic score for lack of self-control and white matter mean diffusivity was modulated by socioeconomic status.

Conclusions

We identified three genetically distinct dimensions of impulsivity and risk-taking with separable neurodevelopmental origins. These genetic predispositions manifested as distinct brain patterns as early as ages 9–10. Environmental experience modulated some of the genetic effects on brain development.

Introduction

Impulsivity refers to acting quickly without sufficient consideration for consequences (1,2) and can also encompass the tendency to take risks (3,4). Impulsivity is a transdiagnostic trait observed in many psychiatric disorders, including attention-deficit/hyperactivity disorder (5,6), substance use disorders (7–9), bipolar disorder (10–12), and schizophrenia (13,14).

Impulsivity is typically assessed using self-reported questionnaires, such as the Barratt Impulsiveness Scale (BIS-11) (15) and the UPPS-P (16,17). Alternatively, laboratory tasks, such as the Delay Discounting task, can be used to measure one's preference for smaller immediate rewards over larger delayed rewards (18). In addition, risky behaviors—such as speeding, alcohol consumption, and smoking—are often evaluated alongside impulsivity, as they reflect a tendency to engage in activities with potential negative health consequences (19).

Impulsivity is therefore a multifaceted construct (20–27), with several theories proposed to characterize its underlying dimensions. For example, dual-process models identify two competing systems: one for cognitive control, which regulates imprudent impulses, and one for motivational drive, which promotes reward or sensation seeking (28–32).

Neuroimaging studies mirror the multidimensional nature of impulsivity, identifying multiple brain structures and networks associated with facets of impulsivity. Trait impulsivity has been associated with lower gray matter volume in the prefrontal cortex (33). Meanwhile, task-based functional MRI research found that impulsive decision-making is associated with greater activation of ventromedial prefrontal cortex and ventral striatum, while inhibition of impulsive responses recruits regions in fronto-parietal control networks (34,35).

Twin studies have shown that impulsivity and risk-taking traits are heritable, with heritability estimates varying between 30 and 60% (36–42). Genome-wide association studies (GWASs) have further identified key loci associated with different impulsivity traits near genes implicated in neurobiological processes (43–46). However, how these genetic predispositions manifest in the brain during development is largely unknown.

Identifying the mechanisms underpinning impulsivity and risk-taking in early adolescence is of interest, as this period is marked by heightened impulsivity and increased engagement in risky behaviors (47–49). Moreover, since most individuals have not yet experienced prolonged substance use during early adolescence (50–52), this stage offers a unique opportunity to disentangle the brain mechanisms driving impulsivity and risk-taking from the effects of substance exposure.

The aim of the present study was three-fold. First, we aimed to identify genetic constructs underlying different facets of impulsivity and risk-taking by applying genomic structural equation modeling to 17 relevant traits from published GWASs (44–46,53,54). Although the genetic architecture of impulsivity has been previously explored (55–58),

we extended this work by jointly analyzing risk-taking and impulsivity traits using a data-driven framework. Second, we examined whether the genetic predisposition for impulsivity and risk-taking is associated with brain structures and cognitive function during early development. To this end, we utilized data from the Adolescent Brain and Cognitive Development [SM] (ABCD) Study, the largest genetic and neuroimaging dataset of youth to date. Finally, since socioeconomic status (SES) has been linked to both impulsivity and brain development (59–62), we examined how genetic predisposition for impulsivity and SES factors such as parental education and household income jointly contribute to differences in brain structures among children and adolescents.

Materials and Methods

GWAS of impulsivity and risk-taking behavior

We used published GWAS summary statistics of 17 traits related to impulsivity and risk-taking behaviors (Table. 1). We included Body Mass Index (BMI) and Education Attainment (EA) for the initial analyses, given their relevance to impulsivity (63–68). All GWASs were derived from individuals of genetically inferred European ancestry.

Table 1 GWASs related to impulsivity and risk-taking behaviors
We used summary statistics made available by the original papers, except for Risk Tolerance and Educational Attainment. For these, we conducted meta-GWAS, following the methods described in the original paper. This is due to the limitation of data availability (See Supplementary Information). SNP h2: SNP-based heritability. None of these GWAS included individuals from the ABCD cohort.

Phenotype	Sample Size	SNP h2	Phenotype Description	Reference
BIS Attentional	124,739	0.070	Inability to focus attention or concentrate	Sanchez-Roige et al. (2023) (45)
BIS Motor	124,104	0.067	Tendency to act without thinking	Sanchez-Roige et al. (2023) (45)
BIS Nonplanning	123,509	0.094	Lack of planning or forethought	Sanchez-Roige et al. (2023) (45)
UPPS-P Lack of Premeditation	132,667	0.066	Tendency to not plan ahead and act without thinking	Sanchez-Roige et al. (2023) (45)
UPPS-P Lack of Perseverance	133,517	0.064	Inability to sustain attention or motivation	Sanchez-Roige et al. (2023) (45)
UPPS-P Positive Urgency	132,132	0.061	To act rashly under positive emotion	Sanchez-Roige et al. (2023) (45)
UPPS-P Negative Urgency	132,559	0.079	To act rashly under negative emotion	Sanchez-Roige et al. (2023) (45)
UPPUP Sensation Seeking	132,395	0.097	Inclination to seek novel and thrilling experiences	Sanchez-Roige et al. (2023) (45)
Delay Discounting	23,127	0.122	Choice between smaller	Sanchez-Roige et

			current versus larger delayed reward	al. (2018) (44)
Risk Tolerance	1,415,869	0.051	Self-reported tendency to take risks	Linnér et al. (2019), Becker et al. (2021) (46,53)
Smoking Initiation	632,802	0.078	Whether an individual has ever smoked regularly	Liu et al. (2019) (54)
Drinks per week	537,349	0.042	Number of alcoholic drinks consumed per week	Liu et al. (2019) (54)
Automobile Speeding Propensity	404,291	0.079	Frequency of driving above speed limit on the motorway	Linnér et al. (2019) (46)
Number of Sexual Partners	370,711	0.128	Number of sexual partners in their lifetime	Linnér et al. (2019) (46)
Drug Exposures	124,739	0.108	The number of 11 different classes of drugs an individual has used	Sanchez-Roige et al. (2023) (45)
Body mass index	681,275	0.060	Weight (kg) divided by height (m) squared	Yengo et al. (2018) (69)
Educational Attainment	3,037,499	0.078	Highest level of education achieved	Okbay et al. (2022) (70)

Genomic Structural Equation Modeling (genomic SEM)

Genomic SEM R package v0.0.5 (71) was used to build and test factor models, following our previous work (55–58,72). While previous hypothesis-driven models successfully captured aspects of the genetic architecture of impulsivity, here we adopted a data-driven (hypothesis-free) approach (73) to further explore how impulsivity and risk-taking traits are organized jointly (See Supplementary Information for detailed descriptions). Model fit was evaluated using comparative fit indices (CFI) and standardized root mean square residuals (SRMR).

<u>Multivariate GWAS</u>

Genomic SEM R package (71) was used to conduct multivariate GWAS, where SNP associations with the latent factors derived from genomic modeling were estimated. The effective sample size was calculated as described by Mallard et al. (74). For subsequent analyses, we omitted SNPs that had Q_{SNP} p-values below 5e-8 (See Supplementary Information).

The SNP2GENE function from the Functional Mapping and Annotation (FUMA) v1.5.2 pipeline (75) was used to identify genomic risk loci and lead SNPs.

Gene-based analyses

We used MAGMA v1.08 (76) to conduct gene association, gene-set enrichment, and gene property analyses for each latent factor. These analyses tested whether GWAS-implicated genes were preferentially associated with specific gene-sets, expression in particular tissues, or expression during specific developmental epochs. (See Supplementary Information for reference datasets. For the developmental epoch analysis, arthritis GWAS (77) was used as a negative control to support that associations reflected impulsivity-specific patterns, not general developmental expression trends.

ABCD dataset

Participants

To examine how the genetic predisposition of impulsivity and risk-taking unfolds during development, we used data from the ABCD Study ®. This is a longitudinal cohort study, with genetic, demographic, behavioral, and brain-imaging data collected across 21 sites in the US (78–80). We used neuroimaging and genetic data from the ABCD 5.1 release. The participants were recruited at ages 9 to 10 and undergo follow-up MRI scans every 2 years. We included participants of genetically inferred European ancestry to match the population of the GWASs. This totaled 4,142 participants at baseline (mean age: 119.4 months, females: 1,958), 3,260 participants at 2-year-follow up (mean age: 143.6 months, females: 1,451), and 1,346 participants at 4-year-followup (mean age: 169.2 months, females: 606). Subjects with at least two measurements were included in the longitudinal analysis (N= 3,552, females: 1591).

Neuroimaging data

Neuroimaging data was preprocessed by the ABCD consortium as described elsewhere (81). We used cortical surface area and thickness measures derived from Desikan-Killiany (DK) Atlas (82), and white matter (WM) tracts labeled using AtlasTrack (83). Fractional anisotropy (FA) and mean diffusivity (MD) were obtained for each white matter tract. We also included hindered normalized isotropic component (HNI), representing extracellular diffusion, and restricted normalized isotropic component (RNI), representing intracellular diffusion, for subcortical structures (81,84,85). We harmonized imaging measures using longComBat to account for bias arising from different scanning sites (86). Imaging data that did not pass the ABCD recommended QC was excluded (81). In total, we analyzed 234 regional brain measures: cortical surface area and thickness (68 regions each), subcortical volume (16 structures), subcortical HNI and RNI (14 each), and FA/MD across 27 white matter tracts.

Genetic data

To derive individual-level genetic predisposition for impulsivity and risk-taking (see below), we analyzed the genotype data from ABCD participants. These were obtained using the Affymetrix Axiom Smokescreen Array platform (87,88). Individuals

with a high rate of genotype missingness (0.05), high or low rate of heterozygosity (F-statistic <-0.15 or >0.15), mismatched sex labeling, and high relatedness ($\pi>0.125$) were excluded from the sample. We also removed data from genotyping batch 461. Ancestry outliers were detected using HapMap3 data in R v4.0.1. See Supplementary Information for detailed descriptions.

Behavioral and Cognitive data

To examine whether genetic predisposition captured relevant phenotypes, we looked into impulsivity-related measures in the ABCD dataset. These included the UPPS-P Impulsive Behavior Scale for children (80) and the Delay Discounting task (89–91). We also included the Behavioral Inhibition Scale and Behavioral Activation Scale (BIS/BAS), which assess sensitivity to punishment and reward (80,92), and the Externalizing and Internalizing Behavior Composite Score from the Child Behavior Checklist (CBCL), completed by parents (80). Lastly, we included BMI in our analyses, given its well-documented association with impulsivity (63–68).

We further explored associations with broader domains of cognitive function. We included the Little Man Task (visuospatial processing) (89), Wills Problem Solving Task (problem-solving skills) (93–95), Matrix Reasoning from the Wechsler Intelligence Scale for Children-V (nonverbal reasoning) (89), and the Emotional Stroop Task (96). Additionally, we calculated a composite executive function score based on NIH Toolbox tasks, following previous studies (89,97–100).

Most measures in our analysis come from the baseline data collection, except the Delay Discounting Task and Wills Problem Solving Task, which were from the 1-year follow-up.

Socioeconomic Status (SES) of Parents

We assessed socioeconomic status using average parental education (in years), household income, marital status, and the Area Deprivation Index (ADI)—a composite measure of neighborhood disadvantage based on 17 socioeconomic indicators, where higher scores indicate greater deprivation (101,102).

Polygenic Score (PGS) calculation

PGSs for the latent factors identified in genomic SEM were calculated using the PRS-CS pipeline (103), accounting for linkage disequilibrium (LD) using 1000 Genomes Phase 3 reference data.

Longitudinal modeling of neuroimaging data

To assess the developmental trajectory of brain structures, we constructed a latent growth curve model (LGCM) (104,105) for each brain measure (5 global measures and 234 regional measures), using *lavaan* package (106) in R (107). Each

model estimated group-level intercepts and slopes, as well as individual trajectories. Percentage change per annum was calculated as slope/intercept×100. All models had a good fit with CFIs above 0.90 and SRMRs below 0.10.

Partial Least Square

We used Partial Least Squares (PLS) analysis to identify latent patterns of brain structure associated with latent factor PGSs in a multivariate framework. PLS decomposes the covariance between two datasets—in this case, PGS value and brain features—into latent variables that maximally capture their shared variance (108–110). Before analysis, both datasets were z-scored, and brain features were residualized for age, sex, 15 genetic principal components (PCs), and genotyping batch. The significance of latent factors was tested via permutation testing. A Bootstrap Ratio (BSR) was calculated for each variable to capture both the strength and the reliability of its contribution to the latent pattern. Cross-validation was performed to assess robustness and generalizability (111,112). Detailed methods are provided in Supplementary Information.

The same PLS framework was applied to longitudinal brain change data, with the brain matrix replaced by annual percent change in brain features.

Interaction analysis with PGSs and SES

Linear regression models with an interaction term (PGS × SES) were constructed to test whether SES modulated the relation between PGS and global brain measurement. Each model was adjusted for age, sex, 15 genetic PCs, and genotyping batch. The interaction term was tested for significance, and false discovery rate correction was applied for multiple comparison (See Supplementary Information for detailed descriptions).

Dominance Analysis

We used dominance analysis to assess the relative contribution of genetic and environmental predictors to global brain measurements in separate multiple regression models (113,114). Genetic predictors included latent factor PGSs, while environmental predictors comprised parental education, household income, marital status, and ADI. Outcomes included cortical surface area, cortical thickness, subcortical volume, white matter fractional anisotropy, mean diffusivity, and subcortical cellularity (HNI, RNI).

Results

Genetic architecture of Impulsivity and Risk-Taking

To examine genetic relationships among impulsivity-related traits, we estimated genetic correlations across 17 GWASs—15 indexing impulsivity and risk-taking, plus BMI and educational attainment (EA) due to their established associations with impulsivity (Table 1). Most of the traits were significantly correlated (average $|r_g| = 0.30$), with stronger correlations within certain subgroups (max$|r_g|$ between Drug Exposure and Smoking Initiation ($r_g = 0.83$, SE = 0.02), and lower or not significant correlations for a few others (min$|r_g|$ between UPPS-P Lack of Perseverance and Sensation Seeking ($r_g = -0.0013$, SE = 0.04). Hierarchical clustering on the correlation matrix suggested three subgroups (Fig. 1A). EA was negatively correlated with most impulsivity traits except for positive associations with Automobile Speeding Propensity, Risk Tolerance, and Sensation Seeking, which clustered together.

We then modeled genetic covariances of all traits using Genomic SEM. We determined that a three factor model best explained the genetic covariance (Fig. 1B, 1C; CFI=0.907, SRMR=0.088).

The first latent factor appeared to capture *lack of self-control*. This included Barratt Impulsiveness Scale subscales, UPPS-P Premeditation, Positive Urgency, Negative Urgency, and Delay Discounting. The second factor can be labeled as *reward drive*, including Ever smoker, Drinks per week, Number of sexual partners, and Drug Exposure. Finally, the third factor can be conceptualized as *sensation seeking* tendency, encompassing Risk Tolerance, UPPS-P Sensation Seeking, and Automobile Speeding Propensity.

Figure 1 Genetic relationships among impulsivity and risk-taking behaviors
A) Genetic correlation matrix among 17 phenotypes of interest. **B**) Scatterplot of standardized factor loadings from explanatory factor analysis. **C**) Genomic SEM model with three latent factors. Residual covariances are omitted for visual clarity (See Fig. S1 in Supplementary Information).

Multivariate GWAS

Multivariate GWAS identified 2, 66, and 45 genomic risk loci for *lack of self-control*, *reward drive*, and *sensation seeking*, respectively (Table. 2; Fig.2; Table. S1-3). Almost all loci were previously reported in the individual GWASs (Table. S4-6), except

for one locus in *reward drive* near *TNRC6B* (rs139911, $p = 1.23\times10^{-8}$) and 6 loci in *sensation seeking* near genes *SCARNA11* (rs61183738, $p = 4.53\times10^{-8}$), *AGBL4* (rs3121532, $p = 2.53\times10^{-8}$), *ASH1L* (rs12041534, $p = 1.15\times10^{-8}$), *RP11-328D5.1* (rs1204678, $p = 2.16\times10^{-8}$), *AC079135.1:ASB18* (rs10929169, $p = 4.05\times10^{-9}$), and *CCDC171* (rs9298741, $p = 1.77\times10^{-8}$). Among these, loci with hit SNPs rs3121532 and rs1204678 have not been implicated in previous GWASs, while others are implicated in BMI, EA, or Schizophrenia (Table. S4-6).

All three factors showed associations with loci near *CADM2* and *FOXP2*. SNPs near *CADM2* and *FOXP2* genes have been robustly implicated in previous impulsivity and risk-taking GWASs (45,46,115,116). Only 6 associations were shared between *reward drive* and *sensation seeking* (Table S2,3).

Table 2 Genome-wide significant loci associated with lack of self-control, reward drive, and sensation seeking

For reward drive and sensation seeking, only top 10 most significant loci are shown. List of all significant loci (66 for *reward drive* and 45 for *sensation seeking*) are shown in Supplementary Tables S2 and S3, respectively.

rsID	chr	pos	Non effect	effect	MAF	P value	Beta	SE	Nearest gene	function
Lack of self-control										
rs1897699	3	85489304	T	A	0.38	6.18E-10	0.020	0.0032	*CADM2*	intronic
rs936146	7	114294405	C	G	0.41	1.18E-08	-0.018	0.0032	*FOXP2*	intronic
Reward drive										
rs3802848	11	112912387	C	A	0.44	9.90E-27	-0.021	0.0015	*NCAM1*	intronic
rs6788098	3	85624131	T	A	0.37	2.58E-26	0.022	0.0016	*CADM2*	intronic
rs1143702	1	44086831	T	C	0.36	3.52E-16	0.017	0.0016	*PTPRF*	exonic
rs6493275	15	47676579	T	A	0.21	3.05E-14	-0.018	0.0018	*SEMA6D:CTD-2050N2.1*	ncRNA intronic
rs542883	2	45143382	C	G	0.42	4.11E-14	-0.015	0.0015	*RP11-89K21.1*	intergenic
rs1549212	5	166996722	T	C	0.36	8.59E-14	0.015	0.0016	*TENM2*	intronic
rs1591915	10	104646849	T	C	0.42	1.03E-13	-0.015	0.0015	*C10orf32-ASMT:AS3MT*	intronic
rs266059	2	104084908	G	A	0.50	5.71E-13	0.014	0.0015	*AC092568.1*	intergenic
rs2088088	6	98887788	T	C	0.34	1.83E-12	-0.015	0.0016	*RP11-436D23.1*	intergenic

rs1899896	8	93201036	T	C	0.31	5.54E-12	-0.015	0.0016	*RP11-777J24.1*	intergenic
Sensation seeking										
rs6790699	3	85624189	G	A	0.37	4.85E-54	0.021	0.0010	*CADM2*	intronic
rs4841503	8	11024326	C	A	0.47	1.53E-26	0.014	0.0001	*XKR6*	intronic
rs3764002	12	108618630	T	C	0.26	2.04E-15	0.012	0.0011	*WSCD2*	exonic
rs62474683	7	115020725	G	A	0.47	3.07E-15	0.010	0.0010	*RP11-222O23.1*	intergenic
rs13095117	3	18745500	G	A	0.40	1.15E-14	0.010	0.0010	*AC144521.1*	ncRNA intronic
rs7618871	3	136400420	T	G	0.43	1.47E-12	0.009	0.0010	*STAG1*	intronic
rs12413688	10	99095544	G	A	0.37	1.87E-12	0.009	0.0010	*RP11-452K12.4*	ncRNA intronic
rs4551267	7	127175236	T	C	0.45	2.51E-12	0.009	0.0010	*GCC1*	intergenic
rs1015511	7	114142250	G	A	0.48	2.56E-12	-0.009	0.0010	*FOXP2*	intronic
rs556560	5	102623924	G	A	0.34	5.10E-12	-0.009	0.0010	*C5orf30*	intergenic

Figure 2 Multivariate GWAS results for the three latent factors
A-C) Manhattan plots for *lack of self-control* **(A)**, *reward drive* **(B)**, and *sensation seeking* **(C)**. The nearest genes for the lead SNPs are labelled (top 10 significant loci for *reward drive* and *sensation seeking*).

Gene-based analyses

MAGMA identified 2, 150, and 139 genes associated with *lack of self-control*, *reward drive*, and *sensation seeking*, respectively (Table. S7-9). *CADM2* and *FOXP2* were associated with all three factors, consistent with the GWAS results, while an additional 129 and 115 genes were associated with *reward drive* and *sensation seeking*, beyond the GWAS implicated loci.

Reward drive was enriched for genes linked to neuronal development, while *sensation seeking* was enriched for synaptic properties (Table. S10-12). No significant enrichment was found for *lack of self-control* after multiple comparison corrections. Tissue-specific analysis showed broad enrichment in brain tissues across all three factors (Fig. S2). Finally, BrainSpan-based gene property analyses revealed that *reward drive* was significantly enriched for genes preferentially expressed during early, early-mid, and late-mid prenatal period ($p = 1.17 \times 10^{-3}$, $p = 2.23 \times 10^{-5}$, $p = 1.14 \times 10^{-4}$, respectively), while *sensation seeking* was enriched for genes expressed in late infancy ($p = 1.19 \times 10^{-4}$, Fig. 3).

Figure 3 Gene expression enrichment for each developmental epoch
The y-axis refers to the significance on a -log$_{10}$ scale and the horizontal dashed line denotes significance after multiple comparison correction. Arthritis is shown as a control.

Polygenic analyses of impulsivity and risk-taking in children and adolescents
<u>Behavioral Phenotypes</u>

PGS for *lack of self-control*, *reward drive*, and *sensation seeking* were all generally associated with impulsivity measures and risky behaviors, although they diverged in specific association patterns (Fig. 4, Table. S13). *Lack of self-control* PGS showed significant associations with Delay Discounting (β = 0.031, p_{FDR} = 0.045) and UPPS-P subscales except Sensation Seeking (standardized β = 0.038–0.063). *Reward drive* PGS showed the largest association with Externalizing Behavior Scores (β = 0.096, p_{FDR} = 9.45×10^{-10}) and was associated with all UPPS-P subscales (β range between 0.050 and 0.074). *Sensation seeking* PGS was significantly associated with only Sensation Seeking (β = 0.075, p_{FDR} = 3.33×10^{-6}) and Lack of Planning (β = 0.061, p_{FDR} = 1.22×10^{-4}) among UPPS-P subscales. Furthermore, we observed associations beyond impulsivity and risk-taking traits. All three PGSs were negatively associated with the Wills Problem Solving Scale, which captures cognitive resilience in problem solving (β = −0.053, p_{FDR} = 7.88×10^{-4}, β = −0.062, p_{FDR} = 6.55×10^{-5}, β = −0.032, p_{FDR} = 0.0453, respectively).

Figure 4 Association between PGS for lack of self-control, reward drive, and sensation seeking and behavioral and cognitive phenotypes in the ABCD cohort
Age, sex, 15 genetic PCs, and genotyping batch were corrected for all linear regression analyses. Asterisk denotes significance after FDR correction. Measures are from baseline collection (age 9-10) except for Delay Discounting and Wills problem solving (from 1 year follow-up, age 10-11). BIS: Behavioral Inhibition Scale, BAS: Behavioral Activation Scale, RWD: Reward Responsiveness, DRV: Drive, FUN: Fun Seeking.

<u>Brain Measures</u>

Using PLS analysis, we examined whether each PGS was associated with a distinct pattern of brain structural characteristics in the ABCD dataset.

Lack of self-control PGS was significantly associated with a latent brain pattern (p_{perm} = 0.010; Fig. 5A), characterized by reduced cortical thickness of right hemisphere precentral cortex (BSR = −4.01), left cuneus (BSR = −3.44) and insula (BSR = −3.18), and reduced surface area of left insula (BSR = −3.62) and right pars orbitalis (BSR = −3.03; Table. S14).

Reward drive PGS and brain measurements also showed a significant association (p_{perm} = 0.040; Fig. 5B, Table. S15). The identified brain pattern was characterized by thinner lateral orbitofrontal cortex (BSR_{right} = −3.42, BSR_{left} = −2.32), larger surface area in pars opercularis (BSR_{right} = 2.97, BSR_{left} = 2.26) and insula (BSR_{right} = 2.79, BSR_{left} = 2.18), and higher RNI, i.e., higher cellular density (117,118), in right caudate (BSR = 3.13) and accumbens (BSR_{right} = 2.91, BSR_{left} = 2.29).

Finally, *sensation seeking* PGS also significantly associated with a latent brain pattern (p_{perm} = 0.014; Fig. 5C, Table. S16). The brain pattern was characterized by higher FA in left inferior longitudinal fasciculus (BSR = 3.85), superior corticostriate (BSR_{left} = 3.32, BSR_{right} = 2.02), and corticospinal tracts (BSR_{left} = 2.95, BSR_{right} = 2.24), and larger surface area in rostral middle frontal cortex (BSR_{left} = 3.49, BSR_{right} = 2.49), insula (BSR_{left} = 3.25, BSR_{right} = 2.40), and pars opercularis (BSR_{left} = 3.25, BSR_{right} = 2.63).

We ran cross-validation to confirm the generalizability of the PLS analyses. The resulting mean out-of-sample brain-PGS correlation was significant for all three analyses (*lack of self*-control PGS: r(1028) = 0.042, p_{perm} = 0.0198; *reward drive* PGS: r(1028) = 0.050, p_{perm} = 0.0346; *sensation seeking* PGS: r(1028) = 0.039, p_{perm} = 0.0099).

Figure 5 Brain patterns associated with genetic predisposition for impulsivity and risk taking

Bootstrap ratios from the PLS analyses between brain regional measures and **A**) *lack of self-control* PGS, **b**) *reward drive* PGS, and **c**) *sensation seeking* PGS. Regions with Bootstrap Ratio (BSR) values exceeding the threshold corresponding to p < 0.05 are highlighted, indicating significant contribution of the brain measures to the associations. HNI: hindered normalized isotropic component, RNI: restricted normalized isotropic component, FA: fractional anisotropy, MD: mean diffusivity, lh: left hemisphere, rh: right hemisphere.

Brain Development

We also tested whether the PGSs were related to the brain developmental trajectory. *Lack of self-control* PGS showed a significant association with changes in brain structure (p_{perm} = 0.020; Table. S17). As each measurement has a distinct direction of change during this period (Fig. 6A), we translated the BSR to the level of development (Fig. 6B).

Figure 6 Brain development trajectory and the directional interpretation of PLS
A) Annual percentage change for regional brain measurements derived from latent growth curve models. **B**) Schematic panel of how developmental direction and Bootstrap ratio (BSR) translates to levels of development. A positive BSR in regions with positive growth (top left), or a negative BSR in regions with negative change (bottom right) indicates that higher PGS is associated with bigger change (accelerated development). Conversely, a negative BSR in regions with increasing values (top right), or a positive BSR in regions with negative developmental change (bottom left) indicates that having higher PGS is associated with smaller change (slowed development). HNI: hindered normalized isotropic component, RNI: restricted normalized isotropic component, FA: fractional anisotropy, MD: mean diffusivity, lh: left hemisphere, rh: right hemisphere.

The association with *lack of self-control* PGS was characterized by smaller increase in WM FA values—most prominent in the right inferior frontal superior frontal tract (BSR = −3.40), left uncinate (BSR = −3.24), and cingulate cingulum (BSR$_{left}$ = −3.17, BSR$_{right}$ = −3.05)—as well as smaller decrease in HNI of right amygdala (BSR = 2.22), putamen (BSR = 2.06), and left hypothalamus (BSR = 2.10; Fig. 7, Table S14). It also involved greater cortical thinning in left pericalcarine (BSR = −3.22) and more shrinking of right lateral occipital cortex (BSR = −2.98) and pars opercularis (BSR = −2.75). Cross-validation confirmed generalizability (r(785) = 0.036, p_{perm} = 0.0396).

We found no significant association between *reward drive* or *sensation seeking* PGS and developmental changes in brain measurements.

Figure 7 Brain developmental patterns associated with genetic predisposition for lack of self-control
Bootstrap ratios (BSRs) from PLS analysis between regional developmental trajectories and *lack of self-control* PGS. The figure is color-coded to reflect whether *lack of self-control* PGS is associated with bigger growth (pink) or smaller growth (green). BSR values exceeding the threshold corresponding to $p < 0.05$ are highlighted. HNI: hindered normalized isotropic component, RNI: restricted normalized isotropic component, FA: fractional anisotropy, MD: mean diffusivity, lh: left hemisphere, rh: right hemisphere.

Relation with Socioeconomic status

Interaction Analysis

We explored whether the effect of the PGS depended on socioeconomic status. The association between *lack of self-control* PGS and overall WM MD was moderated by ADI ($\beta = 0.057$, $p_{FDR} = 0.039$; Fig. 8A). Further inspection revealed that individuals

with higher ADI had a positive association between PGS and WM MD, whereas those with lower ADI showed a negative association (Fig. 8B).

Dominance Analysis

We used dominance analysis to ascertain the relative contributions of genetics and environment to brain measures. Variance in mean cortical thickness and total subcortical volume was best captured by *lack of self-control* PGS (69.6% and 47.0%, respectively), while subcortical cellularity was best explained by *reward drive* PGS (HNI: 62.1%, RNI:35.3%). Notably, parental education contributed the most to total surface area (48.8%), and ADI and parental marital status were the biggest contributors for WM FA (24.9%) and WM MD (46.3%), respectively (Fig.8C).

Dominance analysis on longitudinal data showed that change of mean WM FA was best described by *lack of self-control* PGS (47.1%; Fig. 8D). ADI contributed most to change in average subcortical cellularity values (HNI: 42.0%, RNI:73.7%), as well as to change in WM MD (46.3%).

Figure 8 Interacting and collective effect of genetic predisposition and socioeconomic status on the brain
A) Scatterplot of association between *lack of self-control* PGS and WM MD, in relation to ADI quantile. Each point is color-coded according to the ADI quantile to which it belongs. The regression line represents the association for each quantile. **B**) Bar plot showing the association coefficient between *lack of self-control* PGS and WM MD at

different ADI quantiles. The y axis shows the standardized beta estimates from linear regressions ran between *lack of self-control* PGS and WM MD within each ADI quantile. **C-D**) Dominance analysis illustrating the contribution of each variable (i.e., PGS or SES measure) to **C**) global brain measures at baseline or **D**) developmental change in global brain measures. The percent contribution of each input variable is calculatged as the variable□s dominance relative to the total fit of the model. WM: White Matter, MD: Mean Diffusivity, ADI: Area Deprivation Index.

Discussion

Using genomic SEM on 17 impulsivity and risk-taking traits, we identified three latent factors—which we labeled *lack of self-control*, *reward drive*, and *sensation seeking*—with both distinct and overlapping genetic associations. Furthermore, the polygenic scores for these factors were associated with specific brain morphology patterns during childhood. Finally, we demonstrated that socioeconomic factors can moderate the relationship between genetic predisposition and brain morphometry.

Our genomic SEM recapitulates the dual-process conceptualization of impulsivity, comprising "top-down" (poor cognitive control) and "bottom-up" (motivation) processes (28–32,119,120). Moreover, it further separates the "bottom-up" system into *reward drive* and *sensation seeking* factors. Gene enrichment analyses also revealed that genes implicated in *reward drive* were more preferentially involved in neurogenesis, whereas those associated with *sensation seeking* related to synaptic properties. Consistent with this, *reward drive* was enriched for genes expressed during the prenatal period, when cortical neurogenesis occurs (121,122), while *sensation seeking* showed enrichment for genes expressed in late infancy, when synapse formation and pruning peak (123–126). Finally, *reward drive* PGS was mostly associated with subcortical nuclei, while *sensation seeking* PGS was reflected in cortical and white matter structure in children. These results collectively suggest that the motivational aspect of impulsivity can be separated into two components with distinct genetic and neurodevelopmental origins.

While the GWAS data used in our modeling were derived from adults, the genetic predisposition was already observable at the behavioral level in children. *Lack of self-control* PGS was indeed related to impulsivity in children—especially with UPPS-P subscales reflecting poor planning and high urgency. Similarly, *sensation seeking* PGS was associated with the UPPS-P Sensation Seeking scale itself in children. While manifestations of *reward drive* (drug use and sexual partners) are not present in children, its PGS was associated with externalizing behavior.

The three latent genetic factors showed distinct brain associations in children. *Lack of self-control* PGS was associated with reduced cortical thickness and surface

area, most prominently in the frontal lobes. This aligns with previous findings linking lower self-control to reduced frontal gray matter across the lifespan (33,100,127–129), and supports the role of frontal lobes in self-regulation and executive function, as shown in functional neuroimaging (130,131) and lesion studies (132).

Reward drive PGS showed associations mainly with heightened cellularity of subcortical nuclei and reduced cortical thickness in orbitofrontal cortex. The subcortical and limbic structures associated with *reward drive* PGS, namely putamen, nucleus accumbens, amygdala, caudate, and orbitofrontal cortex are central to reward processing (133) and implicated in substance use (134–136). While a study in adults found reduced subcortical volumes to be associated with a reward-related phenotype (137), our study in children showed higher cellular density. As most children have not yet used addictive substances, this brain pattern may reflect a predisposing neurological state, whereas the reduction in volume observed in adults may be the result of substance use itself.

The *sensation seeking* PGS was associated with increased cortical surface area and higher white matter FA. This is consistent with evidence directly linking sensation seeking measures to cortical surface area and white matter integrity (103,138,139). Cortical surface expansion and white matter myelination have been shown to co-occur in children, pointing to a shared neurodevelopmental origin that underpins cognitive functions (140). We also found that traits composing the *sensation seeking* factor exhibited positive genetic correlations with Educational Attainment. These results underscore *sensation seeking*'s unique nature and may suggest that this genetic factor is less likely to be associated with later negative outcomes.

Leveraging the longitudinal design of the ABCD Study, we found that *lack of self-control* PGS was associated with slower rate of increase in white matter FA, which likely captures myelination and axonal development (141). This association pattern differed from the cross-sectional results, suggesting dynamic genetic effects on brain development.

While the three factors showed distinct genetic and neurodevelopmental associations, they also showed similarities, particularly the implication *of CADM2* and *FOXP2*. *CADM2* encodes a synaptic cell adhesion protein and mediates synaptic plasticity. Previous work has shown its consistent association with impulsivity and risk-taking, as well as with a wide range of phenotypes such as anxiety and migraine (45). *FOXP2* encodes a transcription factor that regulates the embryonic expression of various genes, including those involved in neuronal growth and synaptic plasticity (127). This gene is highly expressed in cortex and basal ganglia (128) and plays a role in the synaptic wiring of the cortico-basal ganglia circuit (129), a critical network for the cognitive functions.

We also demonstrated the importance of socioeconomic factors in relation to genetic predisposition. Specifically, higher *lack of self-control* PGS was linked to reduced white matter integrity in children from more deprived environments, showing that the relation between genetic risk and the brain can vary according to environmental context. Furthermore, dominance analysis revealed different relative contributions of genetic and environmental factors in brain structure in children (59–62). For cortical thickness, subcortical volume, and subcortical cellularity, PGSs for the three factors had larger total contributions than the total of SES factors, whereas for cortical surface area and white matter integrity, SES factors contributed more than PGSs.

Our model differed from previous genomic SEM models on impulsivity, mainly due to the data-driven approach and the inclusion of risk-taking traits (55,142). For example, Gustavson et al. (55) constructed a 5-factor model that differentiates the UPPS-P subscales, also demonstrating that Delay Discounting constitutes a separate factor. Our model, on the other hand, grouped Lack of Premeditation, Negative and Positive Urgency, and Delay Discounting—which were genetically correlated—as the *lack of self-control* factor. Our data-driven procedure rather picked up the genetic overlap of these traits and highlighted the significant differences from the other traits included.

There are limitations in our study. First, our analyses were limited to individuals of European ancestry, limiting generalizability. This highlights the need to include more diverse populations in future studies (143). Next, the GWAS effective sample sizes differed across the latent factors, which made it difficult to compare results from gene-based analyses. Specifically, the *lack of self-control* factor had a smaller sample size, making it unclear whether the absence of overlap in the enrichment analyses truly represented the difference from the other two factors. Additionally, while our PLS analyses showed collective characteristics of the brain structure associated with each PGS, it remains unclear whether these reflect a unitary neurobiological influence or different independent developmental processes. Finally, this study only focused on brain-PGS associations that were observed across both sexes. How the effect differs between males and females should be explored in the future, as they may show divergent neurodevelopmental patterns.

In sum, this study highlighted the multifaceted nature of impulsivity and risk-taking at the genetic level. Furthermore, we show that the genetic predisposition for these traits is manifested in the brain during early development, with each facet involving different brain structures. This underscores that impulsivity and risk-taking comprise multiple dimensions that tap into different brain mechanisms, which are shaped by unique genetic and neurodevelopmental factors.

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The ABCD data repository grows and changes over time. The ABCD data used in this report came from 10.15154/z563-zd24. DOIs can be found at http://dx.doi.org/10.15154/z563-zd24.

Reference

1. Bakhshani NM. Impulsivity: A Predisposition Toward Risky Behaviors. Int J High Risk Behav Addict. 2014 Jun 1;3(2):e20428.

2. Dalley JW, Robbins TW. Fractionating impulsivity: neuropsychiatric implications. Nat Rev Neurosci. 2017 Mar;18(3):158–71.

3. Meda SA, Stevens MC, Potenza MN, Pittman B, Gueorguieva R, Andrews MM, et al. Investigating the behavioral and self-report constructs of impulsivity domains using principal component analysis. Behavioural Pharmacology. 2009 Sep;20(5–6):390–9.

4. Kozak K, Lucatch AM, Lowe DJE, Balodis IM, MacKillop J, George TP. The neurobiology of impulsivity and substance use disorders: implications for treatment. Ann N Y Acad Sci. 2019 Sep;1451(1):71–91.

5. Moeller FG, Barratt ES, Dougherty DM, Schmitz JM, Swann AC. Psychiatric Aspects of Impulsivity. AJP. 2001 Nov;158(11):1783–93.

6. Miller DJ, Derefinko KJ, Lynam DR, Milich R, Fillmore MT. Impulsivity and Attention Deficit-Hyperactivity Disorder: Subtype Classification Using the UPPS Impulsive Behavior Scale. J Psychopathol Behav Assess. 2010 Sep 1;32(3):323–32.

7. Schneider S, Peters J, Bromberg U, Brassen S, Miedl SF, Banaschewski T, et al. Risk Taking and the Adolescent Reward System: A Potential Common Link to Substance Abuse. AJP. 2012 Jan;169(1):39–46.

8. De Wit H. Impulsivity as a determinant and consequence of drug use: a review of underlying processes. Addiction Biology. 2009 Jan;14(1):22–31.

9. Cupertino RB, Medland SE, Ottino-Gonzalez J, Cao Z, Juliano A, Pancholi D, et al. Impulsivity behaviors and white matter mediate the relationship between genetic risk for cannabis use disorder and early cannabis use in adolescents. Addiction. 2025 May;120(5):984–96.

10. Swann AC, Lijffijt M, Lane SD, Steinberg JL, Moeller FG. Increased trait-like impulsivity and course of illness in bipolar disorder. Bipolar Disorders. 2009 May;11(3):280–8.

11. Mason L, O'Sullivan N, Montaldi D, Bentall RP, El-Deredy W. Decision-making and trait impulsivity in bipolar disorder are associated with reduced prefrontal regulation of striatal reward valuation. Brain. 2014 Aug;137(8):2346–55.

12. Ramírez-Martín A, Ramos-Martín J, Mayoral-Cleries F, Moreno-Küstner B, Guzman-Parra J. Impulsivity, decision-making and risk-taking behaviour in bipolar disorder: a systematic review and meta-analysis. Psychol Med. 2020 Oct;50(13):2141–53.

13. Ouzir M. Impulsivity in schizophrenia: A comprehensive update. Aggression and Violent Behavior. 2013 Mar;18(2):247–54.

14. Hoptman MJ. Impulsivity and aggression in schizophrenia: a neural circuitry perspective with implications for treatment. CNS Spectr. 2015 Jun;20(3):280–6.

15. Patton JH, Stanford MS, Barratt ES. Factor structure of the barratt impulsiveness scale. Journal of Clinical Psychology. 1995;51(6):768–74.

16. Whiteside SP, Lynam DR. The Five Factor Model and impulsivity: using a structural model of personality to understand impulsivity. Personality and Individual Differences. 2001 Mar;30(4):669–89.

17. Cyders MA, Littlefield AK, Coffey S, Karyadi KA. Examination of a short English version of the UPPS-P Impulsive Behavior Scale. Addictive Behaviors. 2014 Sep;39(9):1372–6.

18. Koffarnus MN, Bickel WK. A 5-trial adjusting delay discounting task: Accurate discount rates in less than one minute. Experimental and Clinical Psychopharmacology. 2014;22(3):222–8.

19. Isles AR, Winstanley CA, Humby T. Risk taking and impulsive behaviour: fundamental discoveries, theoretical perspectives and clinical implications. Phil Trans R Soc B. 2019 Feb 18;374(1766):20180128.

20. Huang Y, Luan S, Wu B, Li Y, Wu J, Chen W, et al. Impulsivity is a stable, measurable, and predictive psychological trait. Proc Natl Acad Sci USA. 2024 Jun 11;121(24):e2321758121.

21. MacKillop J, Gray JC, Weafer J, Sanchez-Roige S, Palmer AA, de Wit H. Genetic influences on delayed reward discounting: A genome-wide prioritized subset approach. Experimental and Clinical Psychopharmacology. 2019 Feb;27(1):29–37.

22. Creswell KG, Wright AGC, Flory JD, Skrzynski CJ, Manuck SB. Multidimensional assessment of impulsivity-related measures in relation to externalizing behaviors. Psychol Med. 2019 Jul;49(10):1678–90.

23. Sharma L, Markon KE, Clark LA. Toward a theory of distinct types of "impulsive" behaviors: A meta-analysis of self-report and behavioral measures. Psychological Bulletin. 2014;140(2):374–408.

24. Vassileva J, Conrod PJ. Impulsivities and addictions: a multidimensional integrative framework informing assessment and interventions for substance use disorders. Phil Trans R Soc B. 2019 Feb 18;374(1766):20180137.

25. Dick DM, Smith G, Olausson P, Mitchell SH, Leeman RF, O'Malley SS, et al. REVIEW: Understanding the construct of impulsivity and its relationship to alcohol use disorders. Addiction Biology. 2010 Apr;15(2):217–26.

26. Evenden JL. Varieties of impulsivity. Psychopharmacology. 1999 Oct 21;146(4):348–61.

27. Griffin SA, Lynam DR, Samuel DB. Dimensional conceptualizations of impulsivity. Personality Disorders: Theory, Research, and Treatment. 2018 Jul;9(4):333–45.

28. Steinberg L. A social neuroscience perspective on adolescent risk-taking. Developmental Review. 2008 Mar;28(1):78–106.

29. Casey BJ, Getz S, Galvan A. The adolescent brain. Developmental Review. 2008 Mar;28(1):62–77.

30. Shulman EP, Smith AR, Silva K, Icenogle G, Duell N, Chein J, et al. The dual systems model: Review, reappraisal, and reaffirmation. Developmental Cognitive Neuroscience. 2016 Feb 1;17:103–17.

31. Steinberg L. A dual systems model of adolescent risk-taking. Developmental Psychobiology. 2010;52(3):216–24.

32. Shulman EP, Harden KP, Chein JM, Steinberg L. The Development of Impulse Control and Sensation-Seeking in Adolescence: Independent or Interdependent Processes? J of Research on Adolesc. 2016 Mar;26(1):37–44.

33. Pan N, Wang S, Zhao Y, Lai H, Qin K, Li J, et al. Brain gray matter structures associated with trait impulsivity: A systematic review and voxel-based meta-analysis. Human Brain Mapping. 2021;42(7):2214–35.

34. Gell M, Langner R, Küppers V, Cieslik EC, Satterthwaite TD, Eickhoff SB, et al. Charting the brain networks of impulsivity: Meta-analytic synthesis, functional connectivity modelling, and neurotransmitter associations. Imaging Neuroscience. 2024 Sep 25;2:1–22.

35. Noda Y, Barr MS, ElSalhy M, Masuda F, Tarumi R, Ogyu K, et al. Neural correlates of delay discount alterations in addiction and psychiatric disorders: A systematic review of magnetic resonance imaging studies. Progress in Neuro-Psychopharmacology and Biological Psychiatry. 2020 Apr;99:109822.

36. Hur YM, Bouchard TJ. The Genetic Correlation Between Impulsivity and Sensation Seeking Traits. Behav Genet. 1997 Sep 1;27(5):455–63.

37. Seroczynski AD, Bergeman CS, Coccaro EF. Etiology of the impulsivity/aggression relationship: Genes or environment? Psychiatry Research. 1999 Apr 19;86(1):41–57.

38. Bezdjian S, Baker LA, Tuvblad C. Genetic and environmental influences on impulsivity: A meta-analysis of twin, family and adoption studies. Clinical Psychology Review. 2011 Nov 1;31(7):1209–23.

39. Gustavson DE, Miyake A, Hewitt JK, Friedman NP. Genetic Relations Among Procrastination, Impulsivity, and Goal-Management Ability: Implications for the Evolutionary Origin of Procrastination. Psychol Sci. 2014 Jun 1;25(6):1178–88.

40. Anokhin AP, Grant JD, Mulligan RC, Heath AC. The Genetics of Impulsivity: Evidence for the Heritability of Delay Discounting. Biological Psychiatry. 2015 May;77(10):887–94.

41. Beauchamp JP, Cesarini D, Johannesson M. The psychometric and empirical properties of measures of risk preferences. J Risk Uncertain. 2017 Jun;54(3):203–37.

42. Harden KP, Kretsch N, Mann FD, Herzhoff K, Tackett JL, Steinberg L, et al. Beyond dual systems: A genetically-informed, latent factor model of behavioral and self-report measures related to adolescent risk-taking. Developmental Cognitive Neuroscience. 2017 Jun;25:221–34.

43. Sanchez-Roige S, Fontanillas P, Elson SL, Gray JC, de Wit H, MacKillop J, et al. Genome-wide association studies of impulsive personality traits (BIS-11 and UPPSP) and drug experimentation in up to 22,861 adult research participants identify loci in the *CACNA1I* and *CADM2* genes. J Neurosci. 2019 Feb 4;2662–18.

44. Sanchez-Roige S, Fontanillas P, Elson SL, Pandit A, Schmidt EM, Foerster JR, et al. Genome-wide association study of delay discounting in 23,217 adult research participants of European ancestry. Nat Neurosci. 2018 Jan;21(1):16–8.

45. Sanchez-Roige S, Jennings MV, Thorpe HHA, Mallari JE, Van Der Werf LC, Bianchi SB, et al. CADM2 is implicated in impulsive personality and numerous other traits by genome- and phenome-wide association studies in humans and mice. Transl Psychiatry. 2023 May 12;13(1):167.

46. 23and Me Research Team, eQTLgen Consortium, International Cannabis Consortium, Social Science Genetic Association Consortium, Karlsson Linnér R, Biroli P, et al. Genome-wide association analyses of risk tolerance and risky behaviors in over 1 million individuals identify hundreds of loci and shared genetic influences. Nat Genet. 2019 Feb;51(2):245–57.

47. Harden KP, Tucker-Drob EM. Individual differences in the development of sensation seeking and impulsivity during adolescence: Further evidence for a dual systems model. Developmental Psychology. 2011;47(3):739–46.

48. Wasserman AM, Mathias CW, Hill-Kapturczak N, Karns-Wright TE, Dougherty DM. The Development of Impulsivity and Sensation Seeking: Associations with Substance Use among At-Risk Adolescents. J of Research on Adolesc. 2020 Dec;30(4):1051–66.

49. Arnett J. Reckless behavior in adolescence: A developmental perspective. Developmental Review. 1992 Dec;12(4):339–73.

50. Burstein M. Use and Abuse of Alcohol and Illicit Drugs in US Adolescents: Results of the National Comorbidity Survey–Adolescent Supplement. Arch Gen Psychiatry. 2012 Apr 1;69(4):390.

51. Khurana A, Romer D, Betancourt LM, Brodsky NL, Giannetta JM, Hurt H. Experimentation versus progression in adolescent drug use: A test of an emerging neurobehavioral imbalance model. Dev Psychopathol. 2015 Aug;27(3):901–13.

52. Green R, Wolf BJ, Chen A, Kirkland AE, Ferguson PL, Browning BD, et al. Predictors of Substance Use Initiation by Early Adolescence. AJP. 2024 May 1;181(5):423–33.

53. Becker J, Burik CAP, Goldman G, Wang N, Jayashankar H, Bennett M, et al. Resource profile and user guide of the Polygenic Index Repository. Nat Hum Behav. 2021 Dec;5(12):1744–58.

54. Liu M, Jiang Y, Wedow R, Li Y, Brazel DM, Chen F, et al. Association studies of up to 1.2 million individuals yield new insights into the genetic etiology of tobacco and alcohol use. Nat Genet. 2019 Feb;51(2):237–44.

55. Gustavson DE, Friedman NP, Fontanillas P, Elson SL, the 23andMe Research Team, Palmer AA, et al. The Latent Genetic Structure of Impulsivity and Its Relation to Internalizing Psychopathology. Psychol Sci. 2020 Aug;31(8):1025–35.

56. Mallard TT, Tubbs JD, Jennings M, Zhang Y, Gustavson DE, Grotzinger AD, et al. The pleiotropic architecture of human impulsivity across biological scales. medRxiv. 2023. doi:10.1101/2023.11.28.23299133.

57. Gustavson DE, Morrison CL, Mallard TT, Jennings MV, Fontanillas P, Elson SL, et al. Executive Function and Impulsivity Predict Distinct Genetic Variance in Internalizing Problems, Externalizing Problems, Thought Disorders, and Compulsive Disorders: A Genomic Structural Equation Modeling Study. Clinical Psychological Science. 2024 Sep;12(5):865–81.

58. Cardoso Melo D, Trindade Pons V, Mallard TT, Sanchez-Roige S, Palmer AA, Xie T, et al. Genomic structural equation modeling of reward-related traits: exploring the genetic factor structure and its relationship with psychopathology. Psychiatry Research. 2025 Feb 1;344:116335.

59. Tunney RJ, James RJE. Individual differences in decision-making: evidence for the scarcity hypothesis from the English Longitudinal Study of Ageing. R Soc open sci. 2022 Oct;9(10):220102.

60. Tunney RJ. Economic and social deprivation predicts impulsive choice in children. Sci Rep. 2022 May 27;12(1):8942.

61. Noble KG, Houston SM, Brito NH, Bartsch H, Kan E, Kuperman JM, et al. Family income, parental education and brain structure in children and adolescents. Nat Neurosci. 2015 May;18(5):773–8.

62. Dennis E, Manza P, Volkow ND. Socioeconomic status, BMI, and brain development in children. Transl Psychiatry. 2022 Jan 24;12(1):1–10.

63. Garcia-Garcia I, Neseliler S, Morys F, Dadar M, Yau YHC, Scala SG, et al. Relationship between impulsivity, uncontrolled eating and body mass index: a hierarchical model. Int J Obes. 2022 Jan;46(1):129–36.

64. Lozano JH, Gordillo F, Pérez MA. Impulsivity, intelligence, and academic performance: Testing the interaction hypothesis. Personality and Individual Differences. 2014 Apr 1;61–62:63–8.

65. Vainik U, Dagher A, Dubé L, Fellows LK. Neurobehavioural correlates of body mass index and eating behaviours in adults: A systematic review. Neuroscience & Biobehavioral Reviews. 2013 Mar 1;37(3):279–99.

66. Duckworth AL, Taxer JL, Eskreis-Winkler L, Galla BM, Gross JJ. Self-Control and Academic Achievement. Annual Review of Psychology. 2019 Jan 4;70(Volume 70, 2019):373–99.

67. Amlung M, Petker T, Jackson J, Balodis I, MacKillop J. Steep discounting of delayed monetary and food rewards in obesity: a meta-analysis. Psychol Med. 2016 Aug;46(11):2423–34.

68. Morys F, Yu E, Shishikura M, Paquola C, Vainik U, Nave G, et al. Neuroanatomical correlates of genetic risk for obesity in children. Transl Psychiatry. 2023 Jan 3;13(1):1.

69. Yengo L, Sidorenko J, Kemper KE, Zheng Z, Wood AR, Weedon MN, et al. Meta-analysis of genome-wide association studies for height and body mass index in ~700000 individuals of European ancestry. Human Molecular Genetics. 2018 Oct 15;27(20):3641–9.

70. Okbay A, Wu Y, Wang N, Jayashankar H, Bennett M, Nehzati SM, et al. Polygenic prediction of educational attainment within and between families from genome-wide association analyses in 3 million individuals. Nat Genet. 2022 Apr;54(4):437–49.

71. Grotzinger AD, Rhemtulla M, de Vlaming R, Ritchie SJ, Mallard TT, Hill WD, et al. Genomic structural equation modelling provides insights into the multivariate genetic architecture of complex traits. Nat Hum Behav. 2019 May;3(5):513–25.

72. Vilar-Ribó L, Hatoum AS, Grotzinger AD, Mallard TT, Team 23andMe Research, Elson S, et al. Impulsivity facets and substance use involvement: insights from genomic structural equation modeling. Psychological Medicine. 2025 Jan;55:e51.

73. Grotzinger AD, Mallard TT, Akingbuwa WA, Ip HF, Adams MJ, Lewis CM, et al. Genetic architecture of 11 major psychiatric disorders at biobehavioral, functional genomic and molecular genetic levels of analysis. Nat Genet. 2022 May;54(5):548–59.

74. Mallard TT, Linnér RK, Grotzinger AD, Sanchez-Roige S, Seidlitz J, Okbay A, et al. Multivariate GWAS of psychiatric disorders and their cardinal symptoms reveal two dimensions of cross-cutting genetic liabilities. Cell Genom. 2022. Jun 8; 2(6):100140.

75. Watanabe K, Taskesen E, van Bochoven A, Posthuma D. Functional mapping and annotation of genetic associations with FUMA. Nat Commun. 2017 Nov 28;8(1):1826.

76. de Leeuw CA, Mooij JM, Heskes T, Posthuma D. MAGMA: Generalized Gene-Set Analysis of GWAS Data. PLoS Comput Biol. 2015 Apr 17;11(4):e1004219.

77. Zorina-Lichtenwalter K, Bango CI, Van Oudenhove L, Čeko M, Lindquist MA, Grotzinger AD, et al. Genetic risk shared across 24 chronic pain conditions: identification and characterization with genomic structural equation modeling. Pain. 2023 Oct;164(10):2239–52.

78. Casey BJ, Cannonier T, Conley MI, Cohen AO, Barch DM, Heitzeg MM, et al. The Adolescent Brain Cognitive Development (ABCD) study: Imaging acquisition across 21 sites. Dev Cogn Neurosci. 2018 Aug;32:43–54.

79. Garavan H, Bartsch H, Conway K, Decastro A, Goldstein RZ, Heeringa S, et al. Recruiting the ABCD sample: Design considerations and procedures. Developmental Cognitive Neuroscience. 2018 Aug;32:16–22.

80. Barch DM, Albaugh MD, Avenevoli S, Chang L, Clark DB, Glantz MD, et al. Demographic, physical and mental health assessments in the adolescent brain and cognitive development study: Rationale and description. Developmental Cognitive Neuroscience. 2018 Aug 1;32:55–66.

81. Hagler DJ, Hatton SeanN, Cornejo MD, Makowski C, Fair DA, Dick AS, et al. Image processing and analysis methods for the Adolescent Brain Cognitive Development Study. NeuroImage. 2019 Nov 15;202:116091.

82. Desikan RS, Ségonne F, Fischl B, Quinn BT, Dickerson BC, Blacker D, et al. An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. NeuroImage. 2006 Jul;31(3):968–80.

83. Hagler DJ, Ahmadi ME, Kuperman J, Holland D, McDonald CR, Halgren E, et al. Automated white-matter tractography using a probabilistic diffusion tensor atlas: Application to temporal lobe epilepsy. Human Brain Mapping. 2009 May;30(5):1535–47.

84. White NS, Leergaard TB, D'Arceuil H, Bjaalie JG, Dale AM. Probing tissue microstructure with restriction spectrum imaging: Histological and theoretical validation. Human Brain Mapping. 2013;34(2):327–46.

85. White NS, McDonald CR, Farid N, Kuperman JM, Kesari S, Dale AM. Improved Conspicuity and Delineation of High-Grade Primary and Metastatic Brain Tumors Using "Restriction Spectrum Imaging": Quantitative Comparison with High B-Value DWI and ADC. American Journal of Neuroradiology. AJNR Am J Neuroradiol. 2013 May;34(5):958-64, S1.

86. Beer JC, Tustison NJ, Cook PA, Davatzikos C, Sheline YI, Shinohara RT, et al. Longitudinal ComBat: A method for harmonizing longitudinal multi-scanner imaging data. NeuroImage. 2020 Oct 15;220:117129.

87. Baurley JW, Edlund CK, Pardamean CI, Conti DV, Bergen AW. Smokescreen: a targeted genotyping array for addiction research. BMC Genomics. 2016 Feb 27;17(1):145.

88. Uban KA, Horton MK, Jacobus J, Heyser C, Thompson WK, Tapert SF, et al. Biospecimens and the ABCD study: Rationale, methods of collection, measurement and early data. Developmental Cognitive Neuroscience. 2018 Aug 1;32:97–106.

89. Luciana M, Bjork JM, Nagel BJ, Barch DM, Gonzalez R, Nixon SJ, et al. Adolescent neurocognitive development and impacts of substance use: Overview of the adolescent brain cognitive development (ABCD) baseline neurocognition battery. Dev Cogn Neurosci. 2018 Feb 21;32:67–79.

90. Sloan ME, Sanches M, Tanabe J, Gowin JL. Delay discounting and family history of psychopathology in children ages 9–11. Sci Rep. 2023 Dec 11;13(1):21977.

91. Kohler RJ, Lichenstein SD, Yip SW. Hyperbolic discounting rates and risk for problematic alcohol use in youth enrolled in the Adolescent Brain and Cognitive Development study. Addiction Biology. 2022 Mar;27(2).

92. Pagliaccio D, Luking KR, Anokhin AP, Gotlib IH, Hayden EP, Olino TM, et al. Revising the BIS/BAS Scale to study development: Measurement invariance and normative effects of age and sex from childhood through adulthood. Psychological Assessment. 2016;28:429–42.

93. Gonzalez R, Thompson EL, Sanchez M, Morris A, Gonzalez MR, Feldstein Ewing SW, et al. An update on the assessment of culture and environment in the ABCD Study®: Emerging literature and protocol updates over three measurement waves. Developmental Cognitive Neuroscience. 2021 Dec;52:101021.

94. Wills TA, Windle M, Cleary SD. Temperament and novelty seeking in adolescent substance use: Convergence of dimensions of temperament with constructs from

Cloninger's theory. Journal of Personality and Social Psychology. 1998;74(2):387–406.

95. Wills TA, Dishion TJ. Temperament and Adolescent Substance Use: A Transactional Analysis of Emerging Self-Control. Journal of Clinical Child & Adolescent Psychology. 2004 Feb;33(1):69–81.

96. Smolker HR, Wang K, Luciana M, Bjork JM, Gonzalez R, Barch DM, et al. The Emotional Word-Emotional Face Stroop task in the ABCD study: Psychometric validation and associations with measures of cognition and psychopathology. Developmental Cognitive Neuroscience. 2022 Feb 1;53:101054.

97. Akshoomoff N, Beaumont JL, Bauer PJ, Dikmen SS, Gershon RC, Mungas D, et al. Viii. Nih Toolbox Cognition Battery (cb): Composite Scores of Crystallized, Fluid, and Overall Cognition. Monographs of the Society for Research in Child Development. 2013;78(4):119–32.

98. Akshoomoff N, Newman E, Thompson WK, McCabe C, Bloss CS, Chang L, et al. The NIH Toolbox Cognition Battery: Results from a Large Normative Developmental Sample (PING). Neuropsychology. 2014 Jan;28(1):1–10.

99. Akshoomoff N, Brown TT, Bakeman R, Hagler DJ. Developmental Differentiation of Executive Functions on the NIH Toolbox Cognition Battery. Neuropsychology. 2018 Oct;32(7):777–83.

100. Ronan L, Alexander-Bloch A, Fletcher PC. Childhood Obesity, Cortical Structure, and Executive Function in Healthy Children. Cerebral Cortex. 2020 Apr 14;30(4):2519–28.

101. Singh GK. Area Deprivation and Widening Inequalities in US Mortality, 1969–1998. Am J Public Health. 2003 Jul;93(7):1137–43.

102. Kind AJH, Jencks S, Brock J, Yu M, Bartels C, Ehlenbach W, et al. Neighborhood Socioeconomic Disadvantage and 30-Day Rehospitalization. Ann Intern Med. 2014 Dec 2;161(11):765–74.

103. Ge T, Chen CY, Ni Y, Feng YCA, Smoller JW. Polygenic prediction via Bayesian regression and continuous shrinkage priors. Nat Commun. 2019 Apr 16;10(1):1776.

104. McArdle JJ. Dynamic but Structural Equation Modeling of Repeated Measures Data. In: Nesselroade JR, Cattell RB, editors. Handbook of Multivariate Experimental Psychology. Boston, MA: Springer US; 1988. p. 561–614.

105. Duncan TE, Duncan SC. An introduction to latent growth curve modeling. Behavior Therapy. 2004;35(2):333–63.

106. Rosseel Y. **lavaan**: An *R* Package for Structural Equation Modeling. J Stat Soft. 2012;48(2).

107. R Core Team. A Language and Environment for Statistical Computing. [Internet]. 2023. Available from: https://www.R-project.org/

108. McIntosh AR, Lobaugh NJ. Partial least squares analysis of neuroimaging data: applications and advances. NeuroImage. 2004 Jan 1;23:S250–63.

109. Krishnan A, Williams LJ, McIntosh AR, Abdi H. Partial Least Squares (PLS) methods for neuroimaging: A tutorial and review. NeuroImage. 2011 May 15;56(2):455–75.

110. McIntosh AR, Mišić B. Multivariate Statistical Analyses for Neuroimaging Data. Annual Review of Psychology. 2013 Jan 3;64(Volume 64, 2013):499–525.

111. Rahim M, Thirion B, Varoquaux G. Multi-output predictions from neuroimaging: assessing reduced-rank linear models. In: 2017 International Workshop on Pattern Recognition in Neuroimaging (PRNI). 2017. p. 1–4.

112. Mirchi N, Betzel RF, Bernhardt BC, Dagher A, Mišić B. Tracking mood fluctuations with functional network patterns. Social Cognitive and Affective Neuroscience. 2019 Jan 4;14(1):47–57.

113. Budescu DV. Dominance analysis: A new approach to the problem of relative importance of predictors in multiple regression. Psychological Bulletin. 1993;114(3):542–51.

114. Azen R, Budescu DV. The dominance analysis approach for comparing predictors in multiple regression. Psychological Methods. 2003;8(2):129–48.

115. Wang J, Gu R, Kong X, Luan S, Luo YLL. Genome-wide association studies (GWAS) and post-GWAS analyses of impulsivity: A systematic review. Progress in Neuro-Psychopharmacology and Biological Psychiatry. 2024 Jun;132:110986.

116. ADHD Working Group of the Psychiatric Genomics Consortium (PGC), Early Lifecourse & Genetic Epidemiology (EAGLE) Consortium, 23andMe Research Team, Demontis D, Walters RK, Martin J, et al. Discovery of the first genome-wide significant risk loci for attention deficit/hyperactivity disorder. Nat Genet. 2019 Jan;51(1):63–75.

117. Palmer CE, Pecheva D, Iversen JR, Hagler DJ, Sugrue L, Nedelec P, et al. Microstructural development from 9 to 14 years: Evidence from the ABCD Study. Developmental Cognitive Neuroscience. 2022 Feb;53:101044.

118. Rapuano KM, Laurent JS, Hagler DJ, Hatton SN, Thompson WK, Jernigan TL, et al. Nucleus accumbens cytoarchitecture predicts weight gain in children. PNAS. 2020 Oct 27;117(43):26977–84.

119. Gullo MJ, Loxton NJ, Dawe S. Impulsivity: Four ways five factors are not basic to addiction. Addictive Behaviors. 2014 Nov;39(11):1547–56.

120. Boog M, Goudriaan AE, Van De Wetering BJM, Deuss H, Franken IHA. The Concepts of Rash Impulsiveness and Reward Sensitivity in Substance Use Disorders. Eur Addict Res. 2013;19(5):261–8.

121. Porter BE. Neurogenesis and epilepsy in the developing brain. Epilepsia. 2008 Jun;49(s5):50–4.

122. Stiles J, Jernigan TL. The Basics of Brain Development. Neuropsychol Rev. 2010 Dec;20(4):327–48.

123. Huttenlocher PR, Dabholkar AS. Regional differences in synaptogenesis in human cerebral cortex. Journal of Comparative Neurology. 1997;387(2):167–78.

124. Peter R. H. Synaptic density in human frontal cortex — Developmental changes and effects of aging. Brain Research. 1979 Mar 16;163(2):195–205.

125. Tebbenkamp ATN, Willsey AJ, State MW, Šestan N. The developmental transcriptome of the human brain: implications for neurodevelopmental disorders. Current Opinion in Neurology. 2014 Apr;27(2):149–56.

126. Tierney AL, Nelson CA. Brain Development and the Role of Experience in the Early Years. Zero Three. 2009 Nov 1;30(2):9–13.

127. Chen Y, Ide JS, Li CS, Chaudhary S, Le TM, Wang W, et al. Gray matter volumetric correlates of dimensional impulsivity traits in children: Sex differences and heritability. Human Brain Mapping. 2022 Jun;43(8):2634–52.

128. Kubera KM, Schmitgen MM, Maier-Hein KH, Thomann PA, Hirjak D, Wolf RC. Differential contributions of cortical thickness and surface area to trait impulsivity in healthy young adults. Behavioural Brain Research. 2018 Sep;350:65–71.

129. Holmes AJ, Hollinshead MO, Roffman JL, Smoller JW, Buckner RL. Individual Differences in Cognitive Control Circuit Anatomy Link Sensation Seeking, Impulsivity, and Substance Use. J Neurosci. 2016 Apr 6;36(14):4038–49.

130. Friedman NP, Robbins TW. The role of prefrontal cortex in cognitive control and executive function. Neuropsychopharmacol. 2022 Jan;47(1):72–89.

131. Langner R, Leiberg S, Hoffstaedter F, Eickhoff SB. Towards a human self-regulation system: Common and distinct neural signatures of emotional and behavioural control. Neuroscience & Biobehavioral Reviews. 2018 Jul;90:400–10.

132. Jones DT, Graff-Radford J. Executive Dysfunction and the Prefrontal Cortex. CONTINUUM: Lifelong Learning in Neurology. 2021 Dec;27(6):1586–601.

133. Schultz W. Reward functions of the basal ganglia. J Neural Transm. 2016 Jul;123(7):679–93.

134. Wise RA, Rompre PP. Brain Dopamine and Reward. Annu Rev Psychol. 1989 Jan;40(1):191–225.

135. Urbanik A, Kozub J, Karcz P, Ostrogórska M. Changes in the brain directly following alcohol consumption—a study of healthy male individuals, with the use of proton magnetic resonance spectroscopy (1HMRS) and diffusion (DWI). Alcohol and Alcoholism. 2021 Jun 29;56(4):415–24.

136. Volkow ND, Michaelides M, Baler R. The Neuroscience of Drug Reward and Addiction. Physiological Reviews. 2019 Oct 1;99(4):2115–40.

137. Aydogan G, Daviet R, Karlsson Linnér R, Hare TA, Kable JW, Kranzler HR, et al. Genetic underpinnings of risky behaviour relate to altered neuroanatomy. Nat Hum Behav. 2021 Jun;5(6):787–94.

138. Escelsior A, Inuggi A, Amadeo MB, Engel-Yeger B, Trabucco A, Esposito D, et al. Sensation seeking correlates with increased white matter integrity of structures associated with visuospatial processing in healthy adults. Front Neurosci. 2023 Oct 26;17:1267700.

139. Owens MM, Hyatt CS, Gray JC, Miller JD, Lynam DR, Hahn S, et al. Neuroanatomical correlates of impulsive traits in children aged 9 to 10. J Abnorm Psychol. 2020 Nov;129(8):831–44.

140. Cafiero R, Brauer J, Anwander A, Friederici AD. The Concurrence of Cortical Surface Area Expansion and White Matter Myelination in Human Brain Development. Cerebral Cortex. 2019 Feb 1;29(2):827–37.

141. Lebel C, Deoni S. The development of brain white matter microstructure. NeuroImage. 2018 Nov;182:207–18.

142. Miller AP, Gizer IR. Dual-systems models of the genetic architecture of impulsive personality traits: neurogenetic evidence of distinct but related factors. Psychol Med. 2024 Jun;54(8):1533–43.

143. Martin AR, Kanai M, Kamatani Y, Okada Y, Neale BM, Daly MJ. Clinical use of current polygenic risk scores may exacerbate health disparities. Nat Genet. 2019 Apr;51(4):584–91.

A | *lack of self-control* PGS   **B** | *reward drive* PGS   **C** | *sensation seeking* PGS

lack of self-control PGS

Surface Area

Thickness

Volume

caudate
accumbens
thalamus
pallidum
hippocampus
putamen
amygdala

HNI

RNI

White Matter

	inferior-fronto-occipital fasiculus	uncinate	cingulate cingulum	parahippocampal cingulum	superior longitudinal fasiculus	inferior longitudinal fasiculus	inferior frontal	superior frontal cortex	anterior thalamic radiations	fornix	superior corticostriate	striatal inferior frontal cortex	corticospinal/pyramidal	forceps major	forceps minor	corpus callosum
lh-FA	*	*	*		*										*	*
rh-FA	*		*		*	*	*	*					*			
lh-MD																
rh-MD																

Bootstrap Ratio

|4.00| |4.00|

smaller change   bigger change