Non-autonomy of age-related morphological changes in the *C. elegans* germline stem cell niche

*Running title: aging niche morphology*

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**ABSTRACT**

A decline in tissue renewal and repair due to changes in tissue stem cells is a hallmark of aging.

Many stem cell pools are maintained by interaction with morphologically complex local niches.

Using the *C. elegans* hermaphrodite germline stem cell system, we analyzed age-related

changes in the morphology of the niche, the distal tip cell (DTC), and identified a molecular

mechanism that promotes a subset of these changes. We found that a long-lived *daf-2* mutant

exhibits a *daf-16*-dependent decline in number and length of long DTC processes. Surprisingly,

the tissue requirement for *daf-16(+)* is non-autonomous and is independent of the longevity

requirement: *daf-16(+)* in body wall muscle is both necessary and sufficient. We also

determined that pre-formed DTC processes deteriorate prematurely when the underlying

germline differentiates. We propose a reciprocal DTC-germline interaction model and speculate

a mechanism by which reducing *daf-2* activity prevents stem cell exhaustion. These studies

establish the *C. elegans* DTC as a powerful *in vivo* model for understanding age-related

changes in cellular morphology and their consequences in stem cell systems.

**SUMMARY**

The *C. elegans* germline stem cell niche morphology is markedly altered with age and is

regulated non-autonomously from the muscle by insulin/IGF-like signaling. Results suggest

reciprocal niche-germline regulation.

**INTRODUCTION**

Age-related changes in cell morphology can alter cell-cell interactions and signaling

(Lopez-Otin et al., 2023). Such changes are particularly important to understand in stem cell

systems where niche cells can take on complex morphologies and associate intimately with the

stem cells they regulate (Brunet et al., 2023). Age-related changes in the cytoskeleton,

extracellular matrix, cell adhesion properties, and other factors influence cell morphology and

function (DiLoreto and Murphy, 2015).

Here, we investigate age-related morphological changes in a simple stem cell niche *in*

*vivo*. Like many mammalian stem cell systems (Brunet et al., 2023; Lopez-Otin et al., 2023), the

*C. elegans* hermaphrodite germline stem cell pool becomes depleted with age (Garigan et al.,

2002; Killian and Hubbard, 2005; Kocsisova et al., 2019; Luo et al., 2010; Qin and Hubbard,

2015). The niche in this system is a single cell, the distal tip cell (DTC), that caps the end of the

gonad (Fig. 1) and produces membrane-bound DSL family ligands, APX-1 and LAG-2, that

activate Notch pathway signaling in juxtaposed germline stem cells (Austin and Kimble, 1987;

Henderson et al., 1994; Kimble and White, 1981; Nadarajan et al., 2009). The adult DTC

morphology is complex, with variable numbers and lengths of long cytoplasmic processes that

extend over and between germline stem and progenitor cells (Byrd et al., 2014; Hall et al., 1999;

Tolkin et al., 2024). DTC processes are not essential for establishment of the stem cell pool that

occurs during larval stages, but they contribute to contact-dependent Notch signaling in adults

(Lee et al., 2016).

Here we present an aging time-course analysis of changes in DTC morphology and

identify molecular mechanisms that delay or hasten them. Our analysis reveals several striking

changes to DTC morphology with age. We confirm an age-related proximal shift in nuclear

position that occurs at high penetrance (as seen by others under different life history and age

conditions (Kocsisova et al., 2019; Urman et al., 2024)), and that the number and length of long

continuous DTC processes decline with age. We examined these phenotypes for their

dependence on the DAF-2 insulin-like signaling (IIS) pathway, which influences lifespan. IIS had

a relatively minor effect on the nuclear position, but the effects of age on DTC process number

and length were highly IIS-dependent. In the *daf-2* mutant, long DTC processes persist with

age, and this persistence requires DAF-16/FOXO. Surprisingly, the cellular requirement for *daf-*

*16* is neither the DTC itself, nor the intestine where it is implicated in longevity. Instead, *daf-*

*16(+)* activity in the body wall muscle is both necessary and sufficient to regulate age-related

changes to the length of the DTC processes. We also determined that pre-formed DTC

processes deteriorate prematurely when the underlying germline ceases cell division by

differentiating. We propose a reciprocal DTC-germline interaction feedback model whereby a

slow germ cell division cycle in *daf-2* mutants delays stem cell exhaustion, in part, by stabilizing

DTC processes which then signal to germline stem cells.

**RESULTS**

***Adult DTC morphology is altered with age***

Age-related changes to hermaphrodite DTC features have been reported previously on a

short aging timeline and/or in mated worms (to 5 days of adulthood). These include a proximally

displaced nucleus (Kocsisova et al., 2019; Urman et al., 2024) and an increase in the size,

though not number, of gaps seen using a membrane reporter (Urman et al., 2024). We sought

to further characterize changes to DTC morphology in intact live self-fertile worms over a more

extensive timeline from Day 1 to Day 10 of adulthood, and to define morphometric parameters

to describe and compare age-related changes under different genetic conditions.

To analyze DTC morphology over time, we selected three time points starting from the

mid-L4 as determined by vulval morphology (Mok et al., 2015): Day 1 (24 hours post mid-L4),

when DTC elaboration has peaked in the wild type; Day 6 post mid-L4, when the reproductive

phase of wild-type self-fertile hermaphrodites is ending; and Day 10 post mid-L4, when worms

are on the verge of the steep population decline consistent with their ~2-3 week lifespan. To

minimize variability due to subtle differences in rearing conditions, for each cohort analyzed,

worms were picked as mid-L4 larvae on the same day and harvested for live imaging on Days

1, 6, and 10.

We examined several different transgenes expressing fluorescent proteins in the DTC,

settling on *qIs57* which encodes GFP under the control of the *lag-2* promoter (Siegfried et al.,

2004). Compared to the other transgenes we examined, it revealed a suitably detailed set of

morphological features, did not appreciably dim with age, and allowed tracking of the position of

the nucleus (Fig. S1). To ensure that the transgene was not itself modulating lifespan, we

performed lifespan analyses in parallel with the canonical wild-type strain N2 and found no

significant difference (Fig. 1A). However, a greater percentage of worms were censored in

strains bearing *qIs57* (see Materials and methods). Adult DTCs are morphologically variable;

two DTCs from each timepoint are shown in Figure 1B and additional examples are in Figure

S2.

Using our image analysis pipeline (Gupta et al., 2024), we found several striking
changes to DTC morphology with age. Day 1 to Day 10 provides the most informative
comparisons (Fig. 1B,C; see Fig. S3 for additional timepoints). First, recapitulating and
extending the findings of others (Kocsisova et al., 2019; Urman et al., 2024), we observed that
the DTC nucleus, which normally sits at the distal end of the DTC, shifts proximally with age.
The percentage of DTCs with the nucleus ≥5 µm from the distal end went from 13% at Day 1 to
77% at Day 10, and these nuclei averaged 3 µm and 9 µm from the distal end at Day 1 and Day
10, respectively. Second, the number and length of processes extending proximally from the
DTC (Byrd et al., 2014; Hall et al., 1999; Tolkin et al., 2024) provided additional parameters. We
focused on those DTC processes that appeared continuous with the cell body in maximum
projection micrographs ("continuous processes" (CPs); Gupta et al., 2024; Tolkin et al., 2024).
Our methods do not distinguish between previously-defined subclasses of DTC processes that
intercalate with or extend along the germ cells (Byrd et al., 2014), but they reflect only gross
morphological features including the proportion of DTCs with processes that extend beyond a
20 µm threshold from the distal end, and the numbers and lengths (mean and maximum) of
such processes. At Day 1 post mid-L4, all DTCs had at least one process extending beyond 20
µm. By Day 10, 14% of DTCs had no CPs beyond this threshold. Among those DTCs with one
or more CP over the threshold, we observed fewer and shorter CPs over time: the mean
number of CPs decreased from 11 to 4 from Day 1 and Day 10, as did their mean and
maximum lengths (mean 37 to 30 µm, and maximum 60 to 38 µm) (Fig. 1C).
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***Insulin/insulin-like growth factor-1 signaling (IIS) promotes age-related changes in DTC***
***process number and length***
The IIS pathway is highly conserved and modulates longevity in *C. elegans*, *Drosophila*,
and mice (Kenyon, 2010b; Murphy and Hu, 2013; Russell and Kahn, 2007). In addition to
longevity, IIS regulates various physiological processes in *C. elegans*, including development,
metabolism, and stress resistance (Murphy and Hu, 2013). DAF-2 is the sole insulin/IGF-1
receptor in *C. elegans* (Kimura et al., 1997), and reducing *daf-2* activity (e.g. by the reduction-of-
function *(rf) daf-2(e1370)* allele) extends lifespan (Kenyon et al., 1993). To determine whether a
reduction in *daf-2* activity would affect age-related changes in DTC morphology, we examined
DTCs in *daf-2(rf)* worms. First, we tested whether the *qIs57* marker affected the *daf-2(rf)*
longevity phenotype and found that, if anything, the percentage of worms alive between 40-60
days was elevated, but the maximum lifespan was similar (Fig. 1A). We aged cohorts of wild-
type and *daf-2(rf)* worms in parallel and measured the parameters as described above.

First, we found that *daf-2(rf)* had a minimal and variable effect on age-related nuclear

position displacement (Figs 1C, S3). Relative to the wild type, *daf-2(rf)* displayed a reduced

proportion of DTCs that exhibit nuclear displacement of ≥5 μm at Day 10. However, a large

percentage (47%) of *daf-2(rf)* worms nevertheless displayed the nuclear displacement

phenotype (versus 77% in the wild type), and the average degree of displacement in microns

from the distal end was similar to the wild type in this cohort. The latter finding is consistent with

those of Kocsisova et al. (Kocsisova et al., 2022 Fig. S1). In summary, nuclear displacement

occurs at a high penetrance in both wild type and *daf-2(rf)* with an equal distance of

displacement, suggesting that IIS plays a minor role in regulating this aspect of age-related

changes to DTC morphology.

In contrast to the minor effects on nuclear position, reducing *daf-2* activity virtually halted

age-related changes in CP number and length from Day 1 to Day 10: all *daf-2(rf)* worms

possessed at least one CP ≥20 μm at Day 10, and there was no decrease in the average

number of CPs ≥20 μm, nor their mean or maximum lengths, as was seen in the wild type (Figs

1C, S4). We therefore focused on the DTC phenotypes related to number and length of CPs.

***The persistence of DTC process number and length in daf-2(rf) does not correlate***

***temporally with extended lifespan or reproductive span***

In addition to lifespan extension in *daf-2(rf)*, the reproductive span of self-fertile

hermaphrodites averages several days longer (Dillin et al., 2002), and in mated hermaphrodites,

the *e1370* allele, in particular, delays reproductive aging (Kocsisova et al., 2022). We wondered

whether the rate of age-related changes in the number and length of DTC processes might

correlate with reproductive span and/or population lifespan. If correlated with reproductive span,

we would expect DTC processes to be shorter after an additional week in *daf-2(rf)*. If the rate of

decline were associated with average lifespan, we would expect to observe shorter processes

by 30 days. We therefore collected additional measurements in *daf-2(rf)* up to Day 30 (Fig. 1D;

additional timepoints in Fig. S3B). Referencing our lifespan measurements, Day 30 in *daf-2(rf)*

reflects the percent alive of wild type at Day 15. We found that, remarkably, while the number of

CPs declined modestly in this cohort by Day 30 (Fig. 1D), it still remained above that of the wild

type at Day 10 (Fig. 1C). In addition, there was no statistically significant decline in the mean

length of CPs ≥20 μm up to 30 days, and only a modest decline in the maximum length.

We conclude that reducing *daf-2* markedly delays age-related changes in the number

and length of long DTC processes, and that these changes in DTC morphology do not directly

correlate with temporal extension of reproductive span or of population lifespan.

***Insulin/insulin-like growth factor-1 signaling (IIS) acts via DAF-16/FOXO to promote age-related changes in DTC process length***

Many phenotypes of *daf-2(rf)* mutants, including age-related phenotypes such as lifespan, depend on the DAF-16/FOXO transcription factor (Kenyon, 2010a; Murphy and Hu, 2013).To determine whether the *daf-2*-dependent age-related changes in DTC process number and length are also dependent on *daf-16*, we examined DTC morphology, as visualized with the *qIs57* marker, in double mutant strains bearing a *daf-16* null mutation *daf-16(mu86)* and *daf-2(e1370)* [hereafter referred to as *daf-16(0); daf-2(rf)*]. Although the vast majority of DTCs had one or more CP that exceeded 20 μm, the number of processes and their mean and maximum length decreased similar to the wild type (Fig. 2).

To assess the loss of *daf-16* alone, we measured the same parameters in parallel cohorts with the wild type, *daf-2(rf)*, and *daf-16(0); daf-2(rf)* (Figs 2, S5). In these trials, a small but significant percentage of CPs of Day 10 DTCs in *daf-16(0)* worms did not exceed 20 μm, though not as many as the wild type. CP number and length also decreased, as in the wild type. Taken together, we conclude that the maintenance of DTC process number and length over time in *daf-2(rf)* is highly dependent on *daf-16* activity.

***Non-autonomous daf-16 activity influences age-related decline of DTC process length***

Prior studies indicated that *daf-16* activity in specific tissues can influence distinct phenotypes while also contributing to systemic effects. For example, intestinal *daf-16* is associated with enhanced stress resistance and increased longevity (Libina et al., 2003; Zhang et al., 2022), and *daf-16* in the germ line and muscle promotes expansion of the progenitor pool during germline development (Michaelson et al., 2010). Additionally, *daf-16* activity in *fos-1a* expressing cells in the proximal somatic gonad (PSG) prevents the age-dependent decline in number of germline progenitors (Qin and Hubbard, 2015). Therefore, we sought to determine in which tissues *daf-16* most influences the maintenance of DTC processes with age that we observed in *daf-2(rf)*. For this analysis, we focused on the DTC length parameters (mean and maximum).

Our first approach took advantage of available extrachromosomal arrays (Libina et al., 2003; Qin and Hubbard, 2015). Each includes a tissue-restricted promoter driving a GFP::DAF-16a translational fusion. We performed the aging time course in the *daf-16(0); daf-2(rf)* background and compared DTC processes length parameters from control and array-bearing (phenotypically Rol) worms, and non-array bearing (phenotypically non-Rol) progeny from the

same array-bearing mothers. If *daf-16(+)* activity were important in a particular tissue location,

we expected that its expression would preserve DTC process length relative to *daf-16(0); daf-*

*2(rf)*.

To assess the approach, we first examined CPs in worms bearing an array that

expresses *daf-16a* from the *daf-16* promoter. We observed that all parameters (the proportion of

worms with DTC CPs over 20 μm, and the CP number and length – both mean and maximum)

were markedly stable relative to *daf-16(0); daf-2(rf)* (Fig. 3) and to non-array bearing siblings

(Fig. S6). The phenotypic similarity of *daf-16(0); daf-2(rf)* to non-array bearing progeny from

array-bearing mothers also suggested that the array did not confer a maternal effect.

We next tested for DTC-autonomy, expressing *daf-16a* from the *lag-2* promoter

(Michaelson et al., 2010). Surprisingly, ~15% of ~140 DTCs exhibited an unusual morphology at

Days 6 and 10, half displaying uncharacteristically long DTC processes, while others displayed

one thick CP containing a displaced nucleus (Fig. S7). None of these features were observed

under any other condition. Among the remaining ~85% of DTCs that exhibited normal

morphology, the CP length parameters were similar to *daf-16(0); daf-2(rf),* even though ~10% of

the DTCs in array-bearing worms at Day 10 had no CPs ≥ 20 μm (Fig. 3, S6). Thus, apart from

the low-penetrance alterations in gross morphology, *lag-2*-driven *daf-16a(+)* had no appreciable

effect on the persistence of DTC process length with age. We conclude that non-autonomous

*daf-16a(+)* activity is sufficient to prevent the age-related decline in DTC length when *daf-2* is

reduced.

***Intestinal activity of daf-16a does not promote age-related DTC length in daf-2(rf)***

Both heterologous expression approaches (Libina et al., 2003) and protein degradation

approaches (Zhang et al., 2022) indicate that the intestine is the major contributor of *daf-16*

activity that confers extended lifespan in *daf-2(rf)*. We speculated that changes in DTC

morphology might be a function of overall organismal aging. If so, we would expect that

intestinal *daf-16(+)* in the *daf-16(0); daf-2(rf)* mutant background would stabilize DTC

processes, similar to *daf-16(+)* driven by its own promoter.

Again, using an available transgene expressing *daf-16(+)* that alters lifespan (Libina et

al., 2003), we tested the sufficiency of *daf-16a* expressed from an intestine-specific promoter.

We found that although all worms bore DTCs with CPs ≥ 20 μm, CP lengths still declined

significantly with age, comparable to that observed in *daf-16(0); daf-2(rf)* controls (Fig. 3, S6). In

conclusion, although intestinal *daf-16(+)* contributes the majority of the longevity phenotype in

*daf-2(rf)* (Libina et al., 2003; Zhang et al., 2022), it does not regulate the length of DTC

processes with age.

**Neuronal *daf-16a(+)* activity mildly influences the age-related changes in DTC length**

Although the intestine is the major contributor of *daf-16* on lifespan, there is neuronal

contribution as well (Uno et al., 2021; Zhang et al., 2022). We tested an array expressing *daf-*

*16a(+)* from a pan-neuronal promoter and observed that while all DTCs in the worms bearing

the neuronally-expressed *daf-16a(+)* array contained CPs that crossed the 20 μm threshold, the

CPs showed a modest but significant length decline with age. We conclude that neuronal

activity of *daf-16(+)* makes a minor contribution to maintaining the length of DTC processes with

age in *daf-2(rf)*.

**Proximal somatic gonad (PSG) *daf-16a(+)* activity does not prevent the age-related loss**

**of DTC process length**

Previous results indicate that *daf-16a* expressed in *fos-1a*-positive cells of the proximal

somatic gonad (PSG) in a *daf-16(0); daf-2(rf)* double mutant delays age-dependent loss of

germline progenitors (Qin and Hubbard, 2015), and *daf-16*-dependent DOS-3, a non-canonical

ligand for the GLP-1/Notch receptor, was identified as a relevant PSG signal (Zhang et al.,

2024). We hypothesized that a PSG signal might also contribute to DTC morphology. Although

prior experiments used *naEx239[*pGC629*(fos1p::gfp::daf-16a)* + pRF4*]* in *daf-16(m26); daf-*

*2(e1370)* (Qin and Hubbard, 2015), to keep the *daf-16* background consistent, we moved the

array into the *daf-16(mu86); daf-2(rf)* background, and measured CP number and length.

In short, *daf-16a(+)* activity in the PSG did not prevent the age-dependent loss of DTC

process length. However, these worms were markedly less healthy than other strains bearing

*daf-16(+)* arrays: only ~50% of worms survived to Day 4, a defect not observed with other *daf-*

*16(+)* arrays. Moreover, nearly a third (29%) of worms bearing the array had no DTC process

≥20 μm. Of those that did, the mean and maximum lengths were lower than the control on Day

1 and nevertheless continued to decrease with age (Figs 3A, S6).

Collectively, these findings demonstrate that the expression of *daf-16a(+)* via *fos-1a*

promoter is insufficient to prevent the age-dependent DTC process decline in a *daf-16(0); daf-*

*2(rf)* background. This result raises the possibility that while *daf-16a(+)* in the PSG promotes

maintenance of the progenitor pool with age (Qin and Hubbard, 2015; Zhang et al., 2024), it

does not delay the decline of DTC process length with age.

**Germline *daf-16* exerts a minor effect on DTC process length**

The germ line is the major focus of activity for *daf-16* in modulating expansion of the progenitor zone (PZ) in larval stages (Michaelson et al., 2010), and the development of adult DTC morphology is partially dependent on the germ line (Tolkin et al., 2024). To determine whether germline *daf-16* is required for long continuous DTC processes to persist in *daf-2(rf)*, we used RNAi feeding and the *rrf-1* mutant in which RNAi is less effective in the soma but is retained in the germ line (Kumsta and Hansen, 2012; Sijen et al., 2001). We note that in these experiments (as in Fig. 2) *daf-2(rf)* displays an increase in mean (Fig. 3B) and maximum (Fig. S8) CP length from Day 1 to Day 10*.* We speculate that this may be related to a delay in the PZ reaching its maximum size in *daf-2(rf)* worms (see Discussion). Reducing *daf-16* by RNAi in *rrf-1(+)* eliminated this increase. In the absence of *rrf-1*, the mean CP length increased in both the L4440 control and with *daf-16(RNAi)*, suggesting that the contribution of *daf-16* from the soma is more relevant for this phenotype. For maximum length, however (Fig. S8), a minor effect of *daf-16(RNAi)* is seen in the absence of *rrf-1*. We conclude that the germline *daf-16* plays a minor role in regulating the length of the longest DTC process in *daf-2(rf)* with age.

**Body wall muscle-specific *daf-16a(+)* is sufficient and necessary for the DTC process length reduction with age in *daf-2(rf)***

We measured DTC parameters in *daf-16(0); daf-2(rf)* worms bearing an array expressing *daf-16a(+)* in body wall muscle (*myo-3p::GFP::daf-16a*; Libina et al., 2003). All the worms carrying the array had CPs ≥20 µm (Fig. 3). Furthermore, CP length was maintained in *daf-16(0); daf-2(rf)* relative to controls without the array, and was similar to worms carrying the *daf-16p::daf-16* array (Figs 3A, S6). Together, these findings indicate that array-borne expression of *daf-16a* in body wall muscle is sufficient to maintain the length of DTC processes with age in *daf-2(rf)*.

To assess the necessity of body wall muscle DAF-16, we used an auxin-mediated degron approach (Zhang et al., 2015). We generated a strain bearing *daf-2(rf),* a *TIR1::mRub*y fusion expressed under the *myo-3* promoter (Sabatella et al., 2021) together with degron-tagged DAF-16::GFP (Zhang et al., 2022), and the DTC marker. Following the red and green fluorescent protein tags, we confirmed the tissue-specific expression of TIR1 and auxin-dependent loss of DAF-16. In all of the vehicle controls (*daf-2(rf)* and with TIR1 or the DAF-16::degron alone), the DTCs in *daf-2(rf)* strains not only maintained but increased in length over the 10 day interval, while mean length of DTC processes in the strain bearing both *myo-3p::TIR1* and the degron-tagged DAF-16 decreased (Figs 3C, S6).

These results suggest that *daf-16* in body wall muscle is a major contributor to the

persistence of long DTC processes in *daf-2(rf)* with age.

**Body wall muscle DAF-16a attenuates the rate of germline progenitor zone decline with**

**age**

Prior studies implicated muscle-expressed *daf-16a* activity in a minor but significant role

downstream of *daf-2* in promoting larval expansion of the germline progenitor pool (Michaelson

et al., 2010), whereas it did not significantly influence the endpoint of the progenitor cell number

in aged worms (Qin and Hubbard, 2015).

Our DTC morphology analysis prompted us to re-examine the possibility of a role for

*myo-3p::daf-16a(+)* in limiting the PZ pool over an aging time-course in *daf-2(rf)*. We found that

the PZ pool started with fewer cells on Day 1, consistent with the results of Michaelson et al.

(2010). Relative to controls, *daf-16(0); daf-2(rf)* with *myo-3p::daf-16a(+)* does not affect the

terminal PZ cell count, consistent with the results of Qin and Hubbard (2015). However, the

presence of the *myo-3p::daf-16a(+)* array does attenuate the rate of decline of the PZ, similar to

*daf-2(rf)* (Figs 4A, S9). This result suggests that muscle-produced *daf-16(+)* prevents the loss of

both the PZ and DTC processes length over time.

**Germline proliferative status is correlated with stability of DTC processes**

A striking feature of the aged *daf-2(rf)* germ line is the relative stability of the PZ pool

compared to the wild type (Fig. 4). In mated worms, *daf-2(rf)* also displays a more proximal

border of SYGL-1-positive stem cells and a more constant position of meiotic entry relative to

the wild type (Kocsisova et al., 2022). We further observed that the mitotic index did not decline

significantly from Day 1 to Day 10 in *daf-2(rf)* (Fig. 4B). These results prompted us to consider

the relationship between DTC processes, mitotic progression and stem cell fate.

Out-growth of the DTC processes in early adulthood is dependent on an underlying

substrate of the undifferentiated germ cells (Byrd et al., 2014; Tolkin et al., 2024). We therefore

wondered whether adult DTC processes, once formed, would be stable in the absence of the

underlying PZ in early adulthood. We analyzed the number and length of long DTC processes in

worms bearing a temperature sensitive allele of *glp-1/* Notch in which the entire PZ differentiates

at the restrictive temperature (Austin and Kimble, 1987). We examined the nuclear morphology

of distal germ cells and measured DTC parameters from parallel cohorts of worms that

underwent the same temperature-shift regime (Fig. 5A). After 24 hours at the restrictive

temperature, although virtually all distal germ cells had entered meiotic prophase, the CP

number and length parameters were unaffected. By 48 hours, however, the CP length was
reduced relative to the wild type (Fig. 5A-C).
We conclude that premature differentiation of the PZ pool causes a premature decline in
the complexity of the DTC morphology, but with a time delay between differentiation and the
decline of DTC processes. Loss of the stem cells eventually leads to changes in DTC
morphology that resemble what is seen in older adults.
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355 **DISCUSSION**
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Here we define age-related changes in DTC niche morphology and assess their
dependence on IIS, a molecular mechanism that affects longevity. We found that an age-related
shift in DTC nucleus position is only modestly affected by IIS in self-fertile hermaphrodites, as
was previously shown in mated worms (Kocsisova et al., 2022), while the decline in the number
and length of long DTC processes is highly dependent on IIS. We identify body wall muscle as a
major source of a *daf-16(+)*-dependent activity that can support both the persistence of long
DTC processes in a *daf-2(rf)* mutant and the underlying germline progenitor pool. Finally, we
determine a dependency relationship between the germ cell proliferative state and maintenance
of pre-formed DTC processes.
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**A model for reciprocal feedback interactions between the aging germline and niche**
In a seeming paradox, *daf-2(+)* expands the larval germline progenitor zone (PZ) but
drives loss of the PZ with age, and both effects are dependent on *daf-16*. In larvae, the germline
cell cycle is slower in worms with reduced *daf-2* activity and this is dependent on *daf-16(+)* in
the germ line and body wall muscle (Michaelson et al., 2010; Roy et al., 2016). In aged adult
worms, a larger PZ pool is maintained with reduced *daf-2* activity (Qin and Hubbard, 2015; Fig.
4A). In this case, one *daf-16*-dependent signal, DOS-3, from the proximal somatic gonad likely
supports GLP-1/Notch activity in the germ line (Zhang et al., 2024). We found that, in addition,
both the PZ pool and long DTC processes are maintained in old *daf-2(rf)* adults in a *daf-16(+)*
muscle-autonomous manner. The exact *daf-16*-dependent mechanism – and whether it acts on
the directly on the DTC, the germline, or both – awaits further study. We speculate that such a
factor could be a *daf-16*-dependent secreted factor – e.g., a mitokine or myokine signaling
mechanism, or some other mechanism secondary to the IIS role in muscle function (Herndon et
al., 2002; Wang et al., 2019).

Regardless of the exact mechanism, these observations suggest a model for interplay

between the control of cell fate and cell cycle that may contribute to the dynamics of the

germline stem cell system over time with age. Several additional relevant features of the system

are noted below.

First, stem cell fate and cell cycle rate are independently controlled. The PZ consists of a

distal-most SYGL-1(+) germline stem cells (GSCs) that are actively responding to GLP-1/Notch

(Kershner et al., 2013), as well as their more proximal SYGL-1(-) progeny that complete a cell

division before differentiating (Fox and Schedl, 2015). Reducing *glp-1* activity moves the border

of the PZ distally and reduces the number of GSCs but, importantly, does not alter their rate of

cell cycle progression (Fox and Schedl, 2015; Michaelson et al., 2010; Roy et al., 2016). By

contrast, reducing *daf-2* slows the cell cycle (e.g., lowers M-phase and S-phase indices, a rough

proxy) in larvae but does not influence the stem versus non-stem cell fate decision governed by

GLP-1/Notch (Michaelson et al., 2010; Roy et al., 2016). Worms with reduced *daf-2* activity

therefore reach adulthood with fewer cells PZ cells. This pool declines with age, but at a rate far

slower than the wild type (Kocsisova et al., 2022; Qin and Hubbard, 2015; Fig. 4A). Second, as

in many mammalian stem cell systems (Brunet et al., 2023), *C. elegans* GSCs become depleted

over time (Kocsisova et al., 2019; Qin and Hubbard, 2015), and, importantly, the PZ pool is

effectively "used up" over time. This is accelerated in the presence of replete sperm that

encourage germline flux (Kocsisova et al., 2019; Qin and Hubbard, 2015). In addition, with age,

the mitotic index slows, even in the presence of replete sperm (Kocsisova et al., 2019). Finally,

in early adulthood, the outgrowth of DTC processes partially depends on adhesion to underlying

dividing germ cells (Tolkin et al., 2024).

Taking these observations together, we propose a simple two-component feedback

system that supports homeostasis of the progenitor pool in early adulthood, the loss of which

contributes to the collapse of both niche morphology and the stem cell pool with age. In this

model, the early adult PZ steady state is maintained by a combination of DTC-to-GSC signaling

via GLP-1/Notch to designate the stem cell fate in responding germ cells, as well as DAF-2-

dependent inputs that promote cell cycle progression and stability of DTC processes. Over time,

however, as cells cycle and then leave the PZ through differentiation, the pool is depleted,

overlying DTC processes become less stable, and shortened DTC processes further deplete the

stem cell pool.

In the *daf-2(rf)* scenario, we propose that the persistence of slow-cycling yet

undifferentiated germ cells promotes the stability of DTC processes, perhaps by adhesion

mechanisms similar to those that act in the early adult (Tolkin et al., 2024). This stability could

then support DTC-to-GSC interactions along the DTC processes to promote stem cell fate (Fig.

5D). This additional GLP-1/Notch signaling afforded by stable DTC processes may, in turn,

support the pool of cells in an undifferentiated, proliferation-competent (though slow-cycling)

state. Reducing *daf-2* would thereby delay the age-related declines in both the progenitor pool

and niche morphology.

Temporally, the decline of the stem cell pool and PZ precedes that of the DTC

processes (Figs 1,4). Similarly, when germ cells are forced to differentiate, a time delay occurs

before the DTC declines (Fig.5). This suggests that the germline-to-DTC stabilization of the

processes is important for the behavior of the system. Our observation that loss of *glp-1* leads to

premature collapse of already-formed DTC processes also implicates the germline-to-DTC

branch of the feedback to maintain DTC processes.

While this model provides a framework for reciprocal signaling between DTC processes

and the germ line, additional influences act on the system. For example, the regulation of GLP-

1/Notch signaling with age (Kocsisova et al., 2019) is complex: in addition to DOS-3-mediated

signaling from the proximal somatic gonad (Zhang et al., 2024), the shift in the position of the

DTC also alters the patterning of signaling (Urman et al., 2024), and there may be germline-

autonomous aspects of reduced GLP-1/Notch signaling since, at least to Day 4, *sygl-1* RNA

levels do not change (Urman et al., 2024). Our model also predicts that the effects of DAF-16

from the proximal somatic gonad that act on the germ line should affect the DTC too, since they

preserve the PZ. However, we did not observe the predicted stabilization of DTC processes

when *daf-16(+)* was expressed in the proximal somatic gonad in *daf-16(0); daf-2(rf)*.  Finally,

many aspects of the germ cell cycle are unusual, including a highly abbreviated G1,

constitutively high levels of cyclin E/CDK2 throughout the cell cycle, and transcriptional

regulation of CDK-2 through inhibition of DPL-1 (Fox et al., 2011; Furuta et al., 2018). How such

cell cycle features may be modified with age is unresolved.

**General implications for aging stem cell systems and regenerative medicine**

Our analysis suggests an unexpected mechanism whereby "hypertrophic" cell cycle

promoting activity of a growth factor may lead to age-related stem cell exhaustion that includes

interdependence between the stem cells and the niche. Importantly, our results demonstrate

that niche morphology is a key aspect of aging stem cell systems. Non-autonomous control of

niche morphology, by adjacent stem cells and/or other tissues, may be important in other stem

cell systems as well. Our work further suggests that deeper knowledge of *in vivo* interactions

that are influenced by cellular morphology and cell cycle as well as cell-cell signaling, are

required to promote stem cell regeneration, especially in aging systems.

**MATERIALS AND METHODS**

*Nematode strains and maintenance*

The *C. elegans* strains utilized in this study were raised on standard nematode growth media

(NGM) agar and *E. coli* OP50 at 20ºC, unless otherwise specified. All strains were derived from

Bristol N2 (Brenner, 1974). All strains, including their complete genotypes and sources, are

provided in Table S1. The strains generated in this study were obtained through genetic crosses

and homozygosity was confirmed by progeny testing – visually for strains bearing fluorescent

markers or by DNA sequence analysis based on information in WormBase (Davis et al., 2022).

Strains generated for this study are GC1593, GC1607, GC1616, GC1676, GC1682, GC1683,

GC1691, GC1692, GC1696, GC1778, GC1817, GC1819, and GC1912.

*Aging and lifespan assays*

Multiple plates of approximately 20 L4 hermaphrodites per plate were prepared, and worms

were transferred daily to fresh plates to prevent overcrowding and mixing with their progeny.

Worms were collected for imaging or PZ counting at intervals indicated. Controls were run in

parallel within each cohort; cohorts share a color in each figure that reports DTC morphometrics

or PZ nuclei counts, or mitotic index. Lifespan assays (Fig. 1A) started with 100 worms per

genotype. For N2 and CB1370, ~15% were censored due to bagging, desiccating on the petri

dish wall, or were otherwise missing. For the two strains that contain *qIs57* (JK2869 and

GC1607) the rate of censoring for the same reasons was higher, at 40-50%.

*Imaging*

With the exception of images of GC1412 in Fig. S1, DTC images were acquired using a Apo

60X oil immersion lens on a Nikon W1 spinning disk confocal microscope and subsequently

processed, quantified, and analyzed as described in Gupta et al., 2024 (Gupta et al., 2024). The

DTC was visualized using the *qIs57* GFP marker, with a 488 nm excitation laser set at ≤10%

power.  The exposure time was set to 100 ms or 1 frame, the EM Gain Multiplier was set to 50,

the conversion gain was set to 1, and the readout mode was set to 'EM Gain 10 MHz at 16-bit'.

The images were acquired as Z-stacks at 0.5 μm step size and saved in .nd2 format. The

excitation laser used for DAPI was 405 nm at 20-30% power, and for RFP was 561 nm at 20-

30% power range.

***DAPI staining and progenitor zone (PZ) counts***

DAPI staining (using Vectashield from Vector Laboratories™ H-1200-10) was performed as

described in Pepper et al., 2003, except that fixation and staining were done in a 9-well glass

depression plates (Corning™ 7220-85) to minimize the loss of older worms. Images were

acquired using a 60X oil immersion lens on a Nikon W1 spinning disk confocal microscope. The

progenitor zone (PZ) is defined as the region before the occurrence of more than one crescent-

shaped nucleus within the same transverse section of the gonad (Crittenden et al., 2006; Killian

and Hubbard, 2005). After visualizing the entire stack, the PZ region of interest was identified,

and the nuclei were counted manually using the multi-point tool in ImageJ™.

***Auxin-mediated degradation***

For auxin-inducible protein degradation (AID) experiments, 3 mM of Indole-3-acetic acid (IAA,

auxin) (Thermo Scientific™ A10556.14) or an equivalent volume of carrier EtOH for controls

was added to NGM agar prior to pouring plates. OP50 bacteria were concentrated

approximately 2x prior to seeding the auxin-containing plates to compensate for auxin inhibition

of bacterial growth (Zhang et al., 2022). Worms were cultured on both the auxin-containing

plates and the control plates for multiple generations prior to initiating the aging protocol.

**Analysis of *glp-1(e2141)***

Worms were collected at the mid-L4 stage and cultured at 15°C for 48 hours to allow for DTC

elaboration comparable to that seen in Day 1 adults raised at 20°C. A group of worms were

separated from the cohort and imaged (T0), while the rest were shifted to 25°C and imaged 24

and 48 hours later. Cohorts of worms that were grown under the same conditions were split and

analyzed live for DTC features or for cell counts after DAPI staining, as described above.

**RNAi bacterial feeding**

RNAi feeding was carried out as previously described (Timmons et al., 2001) with minor

modifications. Single colonies of *E. coli* strain HT115 carrying either an empty vector (L4440) or

a vector containing *daf-16* cDNA sequences were obtained from frozen stocks on solid LB

plates supplemented with 100 μg/ml ampicillin and 10 μg/ml tetracycline, grown overnight at

37°C and used to inoculate liquid LB cultures supplemented with 100 µg/ml ampicillin. Cultures

were grown at 37°C with shaking for 6-8 hours and used to seed NGM plates supplemented

with 100 µg/ml ampicillin and 0.5% β-lactose. Seeded RNAi plates were kept at room

temperature for 24 hours prior to adding worms. Complete penetrance of the *bli-3* Bli phenotype

was used as a positive control to confirm the efficacy of RNAi reagents in each experimental

replicate. Fresh plates were seeded twice a week over the course of the experiment. L4

hermaphrodites were cultured on either L4440- or *daf-16* RNAi-seeded plates for 1 generation

prior to starting the aging assays.

***Statistics***

For lifespan assays, the Mantel-Cox log-rank test was used. Fisher's exact test was used for

pair-wise comparisons of proportion plots (DTCs with nuclear displacement ≥5 µm and DTCs

with any CPs ≥20 µm). For determining significance between nuclear displacement, number,

mean length, and maximum length of CPs ≥20 µm, the number of PZ nuclei, or mitotic index:

two-tailed t-test for pairwise comparisons (main figures), and two-tailed t-test with Bonferroni

correction for multiple comparisons (supplement). When reporting the mean and maximum CP

length, we did not include DTCs that with no CPs >20 µm. All sample numbers (n) are reported

in figures.

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**FIGURE LEGENDS**

**Figure 1. The age-related shift in DTC nuclear position and decline in DTC processes**
**number and length have different levels of dependence on *daf-2*.**
**(A)** Survival curves for *daf-2(+)* and *daf-2(rf)* worms with and without the *qIs57*[*lag-2p::GFP*]
transgene. N2 is the standard wild type, and age is indicated in days post mid-L4 stage.
**(B)** Representative micrographs of DTCs in live worms of the indicated genotypes at adult Day
1, 6 and 10, post mid-L4. The 20 μm region and the area analyzed for number and length of
continuous processes (CPs; see Gupta et al., 2024; Tolkin et al., 2024) are indicated, as is the
position of the longest CP. Open arrowheads indicate non-DTC structures. Scale bars are 20
μm.
**(C)** Quantified age-related DTC features (from top left to bottom right): proportion of DTCs with
nuclear displacement ≥5 μm (black bar), position of nuclear displacement (μm) from the distal
end, proportion of DTCs with any CPs ≥20 μm (black bar), number of CPs ≥20 μm per DTC,
mean and maximum length of CPs ≥20 μm per DTC. N values for proportion plots are the same
as for the nuclear displacement superplot (top middle). For all superplots, each dot represents a
single DTC (n value shown), colors indicate parallel cohorts distinct from those in other main
figures, large circles in color are averages for each cohort, the black dot is the pooled average
for all cohorts shown, and black line connects pooled average values, indicating the direction
and extent of differences between Day 1 and Day 10. Day 10 n values for mean and maximum
CP length are lower since only DTCs with CPs >20 μm were included in these measurements.
**(D)** Number, mean and maximum length of CPs in *daf-2(rf)* at indicated ages in days post mid-
L4.
**For all panels**, *qIs57* carries *lag-2p::GFP* and *daf-2(rf)* is *daf-2(e1370).* See methods for
statistics details; comparisons are made between Day 1 and Day 10 within a given genotype; for
proportion plots, only p < 0.05 are indicated; otherwise, NS is "not significant", or p ≥ 0.05; * p <
0.05, *** p < 0.001.

**Figure 2. Loss of *daf-16* largely reverses the effects of *daf-2* on CP number and length.**
From top to bottom: proportion of DTCs with any CPs ≥20 μm (black bars), number of CPs ≥20
μm per DTC, mean and maximum length of CPs ≥20 μm per DTC in the indicated genotypes for
worms imaged on Day 1 and Day 10 post mid-L4. N values for proportion plots correspond to
the "Number of CPs" superplot to the right; n values for mean and maximum length plots reflect

only those DTCs with CPs ≥ 20 μm. For superplots, dots and circles are as described in Figure

1; colors indicate cohorts and replicates are distinct from those presented in other main figures.

See methods for statistics details; NS is "not significant", * p < 0.05, ** p < 0.01, *** p < 0.001,

and blue asterisks indicate a significant increase from Day 1 to Day 10.

**Figure 3. *daf-16(+)* in body wall muscle is sufficient and necessary for *daf-2(rf)***

**suppression of age-related changes in DTC process length.**

**(A)** Proportion of DTCs with CPs ≥20 μm, and mean length of CPs ≥20 μm per DTC for

indicated genotypes at Day 1 and Day 10 post mid-L4. N values for mean length plots reflect

only those DTCs with CPs ≥ 20 μm. **(B)** Superplots showing mean length of CPs ≥20 μm in

genotypes indicated. *L4440* indicates the empty vector control and *daf-16 RNAi* is the vector

carrying *daf-16* cDNA sequences. **(C)** Mean length of CPs ≥20 μm in worms raised on 3mM

auxin an in genotypes indicated, where *daf-16::degron* is *hq389[daf-16::GFP::degron]* and

*Muscle::TIR1* is *emcSi71[myo-3p::TIR-1::mRuby]*.  **For all panels,** *qIs57* carries *lag-2p::GFP*,

*daf-16(0)* is *daf-16(mu86),* and *daf-2(rf)* is *daf-2(e1370).* For all superplots, colors indicate

cohorts within panels; replicates shown here are distinct from those in other main figures. See

methods for statistics details; NS is "not significant", * p < 0.05, ** p < 0.01, *** p < 0.001, and

blue asterisks indicate a significant increase.

**Figure 4. The progenitor zone (PZ) and mitotic index are influenced by *daf-16*.**

**(A)** The number of PZ nuclei in (n) gonads in worms of the indicated genotypes at Day 1 and

Day 10 post mid-L4.  **(B)** The mitotic index (number of mitotic figures over total cells in the PZ)

in the indicated genotypes at Day 1 and Day 10 post mid-L4. **For all panels,** *qIs57* carries *lag-*

*2p::GFP*, *daf-16(0)* is *daf-16(mu86),* and *daf-2(rf)* is *daf-2(e1370).* For all superplots, colors

indicate cohorts within panels. See methods for statistics details; NS is "not significant", * p <

0.05, ** p < 0.01, *** p < 0.001.

**Figure 5. Premature decline of DTC processes upon germline stem cell differentiation.**

**(A)** Experimental design and images of DAPI-stained control and *glp-1(e2141)* worms at

indicated time points and temperatures. **(B)** Examples of DTC morphology in parallel to those

shown in panel **(A). (A, B)** Scale bars are 20 μm. **(C)** Quantification of number and length (mean

and maximum) of CPs ≥20 μm in control and *glp-1(e2141)* worms at indicated time points.

Colors indicate cohorts within panels. See methods for statistics details; NS is "not significant", *

p < 0.05, *** p < 0.001. **(D)** Cartoon representation of a two-component feedback model for

niche-stem cell interaction at early and late ("old") adult stages. Dotted box indicates area of

direct DTC-germline interaction that is lost with age. White circles represent undifferentiated

germ cells, and grey circles represent differentiated germ cells. See text for details.

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**Figure 1**

**Figure 2**

**Figure 3**

**Figure 4**

**Figure 5**