Genes govern metabolism—Enzymes define pathways and metabolic relationships

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Abstract

Gene centric pathway mapping tools, widely used to interpret untargeted LCMS metabolomics

data, may underperform because a single metabolite can generate multiple spectral features,

inflating false positive rates. Classic enzymology, which established metabolite flow long before

gene sequencing, offers experimentally validated precursor product relationships that could

overcome these ambiguities. We evaluated whether enzymology defined precursor product

correlations are consistently detectable in human plasma LCMS data and explored their potential

to enhance pathway analysis and metabolite identification. Using a high resolution LCMS

platform, we detected amino acids and carnitine related metabolites in one individual sampled

eight times over five years and in 50 adults sampled 6 to 8 times each. Spearman correlations

were calculated for longitudinal and cross sectional data. In the single participant repeated

measures, strong positive correlations were observed for every direct precursor product pair

except the branch point metabolite palmitoylcarnitine. The longitudinal analysis reproduced

these patterns, and the same relationships were retained when analysis was cross sectional.

Despite contributions from multiple organ systems, plasma thus preserved core enzymatic

relationships. Precursor product proportionality, a fundamental principle of enzymology, is

readily detectable in large scale LCMS datasets and remains robust across longitudinal and cross

sectional designs. Applying these correlations to metabolomics workflows can improve pathway

analysis, help metabolite identification, and reveal how genetic variations, diets, therapeutic

drugs, and environmental exposures jointly impact metabolic pathways.

Recently, five biochemical and analytical challenges have been identified that can cause gene-

centric tools to underperform when applied to metabolomics data[1]. Untargeted liquid

chromatography mass spectrometry (LC-MS) metabolomics-based pathway analysis often relies

on pathway-mapping tools[2,3], but ambiguities in mass spectral peak assignment – multiple peaks

arising from the same metabolite – can generate high false positive rates[4,5]. Most metabolic

pathways were established through enzymology before gene sequencing became available, and

these studies offer direct, experimentally validated maps of metabolite flow. Thus, enzymology

offers a simple but powerful way forward to enhance pathway analysis.

Enzymology established that metabolites flow from precursors to products and form dynamic

linear, cyclical, and branched systems with regulation occurring at committed steps and branch

points[6] (**Fig. 1a**). Biochemical analyses showed that under steady-state conditions, the

concentrations of metabolites often vary proportionately[6,7] (see legend, **Fig. 1**). This principle

was demonstrated in the pathway-level identification of xenobiotics and xenobiotic metabolites

in blood[9] and endogenous metabolites in plant tissue[7]. Here, we show examples of precursor-

product correlations observed in LC-MS of amino acids (glutamine, glutamate, ornithine,

citrulline, and arginine) and carnitine-related metabolites (butyrobetaine, carnitine,

propionylcarnitine, O-acetylcarnitine, palmitoylcarnitine) in one person's plasma sampled eight

times across five years (**Fig. 1b**)[9]. Among these examples, the lack of proportional change of

palmitoylcarnitine with carnitine illustrates that positive correlation is not always present,

especially for metabolites at branch points and regulated sites.

To test whether positive precursor-product correlations exist at a population level, we analyzed

LC-MS plasma data from 50 individuals, each with six to eight visits[9]. Consistent with the data

found in the individual (**Fig. 1b**), amino acids and carnitine-related metabolites exhibit positive

correlations across the 50 individuals (**Fig. 1c**), except for the lack of proportional variation of

palmitoylcarnitine with carnitine. Unlike these longitudinal comparisons, however, most human

metabolomics studies use a cross-sectional design with only single LC-MS snapshots of

individuals. To determine whether enzymology-defined precursor-product correlations are

present in a cross-sectional sampling, we restricted comparisons of the 50 individuals to include

only baseline samples. Results show that the same relationships are present (**Fig. 1d**).

As metabolomics moves beyond static pathway maps, enzymology-defined precursor-product

correlations provide a mechanistically grounded view of metabolic pathways. Even though most

metabolism occurs in organ systems other than blood[1], the plasma data (**Fig. 1b, 1c, 1d**) show

that precursor-product relationships are preserved despite variable contributions of organ systems

(**Fig. 1e**). Incorporating these principles into metabolomics can improve pathway analysis, help

metabolite identification, and reveal how genetic variations, diets, therapeutic drugs, and

environmental exposures jointly impact metabolic pathways. As with gene-centric tools, one

must be cautious about enzyme-based relationships[6]; some precursors and products do not show

proportional behavior, and some distantly related metabolites have strong correlations. Despite

these limitations, the results show that precursor-product relationships are highly preserved under

different conditions and provide a valuable resource for LC-MS-based metabolomics research.

Figure Legend

Figure 1. Precursor-product relationships in human plasma. (a). Metabolism occurs in linear (left,

top), cyclic (left, middle), and branched pathways (left, bottom). For sequential enzyme reactions in an

open system at steady state, rates of precursor (S) input, formation of intermediate products (A, B, C, D),

and formation and elimination of end product (P) are equivalent. With the same reaction rates for all steps,

steady-state concentrations of intermediates [A, B, C, D] are related by the respective pseudo-first order

rate constants (k_j): $k_1[S] = k_2[A] = k_3[B] = k_4[C] = k_5[D]$. Because regulation typically occurs only at

branch points or at sites committing a hydrocarbon to a specific fate, this means that proportional

relationships of precursors and products are expected. (**b**) Proportional concentrations for precursor and

product metabolites in different pathways are evident in eight repeated blood samplings from an

individual collected over five years (blue). Examples are provided for amino acids and carnitine-related

metabolites, measured by liquid chromatography mass spectrometry (LC-MS) of plasma extracts (See

Reference 10 for methodological details[10]). All the metabolites on the x-axis are the precursors, and all

the metabolites on the y-axis are the products. Glutamine, glutamate, butyrobetaine, and carnitine are

considered part of a linear pathway, ornithine, citrulline, and arginine are considered part of a cyclical

pathway (urea cycle), and carnitine, propionylcarnitine, O-acetylcarnitine, and palmitoylcarnitine are

considered part of a branched pathway (carnitine shuttle). Sample collection involved overnight fasting

for the first blood draw, but not for the others. Note that an example for a precursor-product pair without

proportional concentrations, palmitoylcarnitine and carnitine (bottom row), is provided. (**c**) Proportional

concentrations for the same precursor and product pairs as in Panel b for 50 individuals with multiple

repeated blood samplings across five years (red). (**d**) Cross-sectional analysis of baseline samples for the

50 individuals in Panel c (black) shows that precursor-product correlations are preserved for the same

precursor and product pairs. (**e**) In multicompartment models, most metabolism occurs in organ

compartments rather than the blood. Despite this, the data in Panels **b** to **d** show that proportional

relationships of precursors and products are observed in blood plasma samples.

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