Article title: The roles of divergent and parallel selection in Amazonian and Andean bird

responses to glacial cycles

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are archived on the National Center for Biotechnology Information Sequence Read Archive

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Abstract

Both divergent and parallel selection can contribute to evolutionary change, yet their

relative contributions to adaptation and speciation are poorly understood, especially during

environmental change. We quantified selective sweep signatures and divergence times in

Amazonian avian sister species pairs that either differ in elevation and are expected to have

experienced strong divergent selection; or occur in similar lowland habitats and are expected to

have experienced strong parallel selection. We found that elevational differentiation resulted in a

greater accumulation of selective sweep signatures over the five-million-year timespan covered

by our dataset, supporting the importance of divergent selection. Nevertheless, lowland-restricted

species pairs accumulated more selective sweeps over the most recent two million years,

suggesting that parallel selection, driven by major Pleistocene glacial cycles, produced faster

evolutionary change than divergent selection in elevationally differentiated species pairs. Our

results highlight parallel selection as an important driver of adaptation and evolution, especially

during environmental instability.

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Introduction

Natural selection leading to adaptation is a fundamental force shaping genomic

differentiation and biological diversity [1–5]. This genomic change may ultimately lead to the

formation of new species due to the build-up of genetic incompatibilities that arise as closely

related species differentially adapt [6,7]. Divergent natural selection, driven by environmental

differences, can fast-track the formation of reproductive barriers between populations – a process

often referred to as ecological speciation [8–10]. Populations can also differentiate under parallel

selective forces when exposed to non-divergent environmental changes [11]. In this case, each

population adapts to the same environmental changes but in different ways. This results in

genomic differentiation that may lead to speciation, especially if adaptations are disrupted in

hybrid individuals, in a process often referred to as mutation order speciation [12,13]. Despite

divergent selection being a more studied and commonly proposed driver of trait evolution, recent

comparative analyses across thousands of terrestrial vertebrate species suggested divergent

selection drove differentiation in key ecological traits like body size and bill shape in only 10%

of species [14]. These inconsistent findings demonstrate fundamental uncertainties about the

roles of divergent versus parallel selection in adaptation and speciation.

The Amazon basin provides an ideal laboratory to investigate the roles of both divergent

and parallel selection in evolution. Spanning more than six million square kilometers with an

elevation range from near sea level to over six thousand meters along the eastern slope of the

Andes, the Amazon basin harbors an extensive array of habitat types. Lowland rainforests are

followed by the epiphyte encrusted subtropical montane forest zone at 1,300 to 2,400 meters,

above which the temperate montane forest zone occurs to treeline (about 3,300 m), with paramo

and puna grasslands above treeline [15] (Fig. 1). Many environmental factors vary with

elevation, including both climatic variables (e.g., precipitation, temperature, solar radiation, wind

speed, oxygen pressure, atmospheric CO_2, etc.) and biotic factors (e.g., niche breadth, forest

height, foraging strata, species diversity, competition strength, etc.) [16]. Moreover, glacial

cycles influenced temperature and possibly rainfall across the Amazon basin. Elevational zones

of the Andes shifted downslope during glacial periods. This environmental change over the past

several million years was most heavily influenced by the mid-to-late Pleistocene glacial cycles

[17–19]. However, unlike in northern latitudes, only the highest elevations of the Amazon basin

in the high Andes were directly glaciated. While most Andean species may have simply tracked

their preferred environmental conditions by moving downslope and upslope during and after

glacial periods, lowland species, often surrounded by river barriers on multiple sides, had

nowhere to migrate. Lowland populations may have been forced to adapt *in situ* to changing

conditions.

These historical patterns of tracking elevation or adapting *in situ* may be evident in the

population-level genomic variation of recently-diverged sister species pairs. Specifically, species

pairs from different elevational zones would likely experience very different environmental

pressures over time as species would first have had to adapt to different environments, then the

lowland member of a pair would have continued to adapt to changing conditions over time while

the highland member of a pair may have tracked its preferred habitat by migrating up- and down-

slope during periods of warming or cooling. Conversely, species pairs with both members from

the same elevational zone would have been lore likely to have experienced parallel

environmental pressures during glacial cycles resulting in parallel selection as both species in a

pair adapted to the same changes in climate [11]. Either circumstance of shared or different

elevational zones between members of a sister species pair would have led to an accumulation of

selective sweeps after divergence [20], but the selective forces underlying such sweeps could be

either divergent or parallel. Therefore, we used species pairs with both members from the

lowlands to study parallel selection (acknowledging that some divergent selection may also

occur), and species pairs with members from the lowlands and highlands to study divergent

selection.

Here, we harness the power of genome-wide data to assess adaptation in closely related

pairs of avian species. Positive selection may produce selective sweeps in the genome where the

target of selection, and linked nearby regions, become fixed or highly differentiated between

populations (high fixation index, F_{ST}), while genetic diversity within populations (nucleotide

polymorphism, π) in these regions is reduced [21–23]. Genomic signatures of strong selective

sweeps often stand out as 'islands of differentiation' in F_{ST} and 'valleys of genetic diversity' in

π, relative to the genomic background signature of these metrics [20]. We calculated the number

of such regions in the genomes for a series of closely related species or genetically differentiated

populations within a species ("species pair" hereafter) (Fig. 2). To test the effect of elevational

differentiation on the accumulation of selective sweeps through time, we calculated divergence

time using site frequency spectrum-based demographic modelling [24]. Species pairs were

divided into two categories: lowland-restricted pairs, in which both species in a pair occur in the

lowlands of the Amazon (i.e., generally below 1000 m; n=9 pairs); and elevationally-

differentiated pairs, where the two species in each pair are differentiated in the elevational zones

they occupy (n=8 pairs). Bayesian linear models were used to assess differences in the

accumulation of selective sweeps with respect to divergence time, between the lowland-restricted

and elevationally-differentiated species pairs [25].

In light of evolutionary theory and natural history of the Amazon basin, we hypothesized

that glacial cycles in the mid- to late-Pleistocene drove selective sweeps in birds of the

Amazonian lowlands and highlands. More specifically, we hypothesized that the accumulation of

selective sweeps between sister species pairs over time would depend on their status as

elevationally differentiated or elevationally similar species. We predict that both divergent and

parallel selection would lead to an accumulation of selective sweeps over time between members

of a species pair. However, divergent selection would lead to a greater rate of selective sweep

accumulation than parallel selection as members of a newly diverged species pair adapted to

different environments.

Methods

Study species

We examined genomic divergence of 17 closely related allopatric species pairs: 9

lowland-restricted pairs, in which both species occur at low elevations in the Amazon basin; and

8 pairs at different elevations, in which one species belongs lower in the Amazonian lowlands

than the other in the Andean highlands. In the latter category, both species of the *Chlorospingus*

pair are Andean, but one occurring at higher elevations, *C. parvirostris*, than its sister species, *C.*

flavigularis (Supplementary Fig. S1). To explore the differences in elevation, we collated

elevational information for each species from either a book specializing in Amazonian birds [26]

when available, or the Birds of the World online resource if not [27–34]. At least four individuals

per species were analyzed (mean = 9.5; range = 4 to 24). Divergence time of species pairs spans

from 0.7 Ma to about 6 Ma, representing different stages of the speciation continuum.

Blood or tissue samples of most bird species from the tropical Andes were collected in

the Manu Biosphere Reserve in the Department of Cusco or at a variety of locations in the

Department of San Martín, Peru. Samples of lowland-restricted species were mostly obtained

from Matto Grosso, Brazil. Complementary tissue samples were obtained from Museu Paraense

Emilio Goeldi (MPEG), the Louisiana State University Museum of Natural Science (LSUMNS),

and the Field Museum of Natural History (FMNH).

Library preparation and sequence data

A total of 323 individuals were analyzed among 34 species (17 species pairs). DNA was

extracted from blood or tissue samples using E.N.Z.A tissue or blood extraction kits (Omega

Bio-Tek). Genotyping-By-Sequencing (GBS) libraries [35] were generated for all samples. The

Cornell Institute of Genomic Diversity generated libraries for several lowland species in the

dataset and these were sequenced on the Illumina HiSeq 2000 platform using 100 bp single-end

sequencing. An almost identical GBS method (following the protocol modifications in

Luzuriaga-Aveiga *et al.* 2021) was used to generate libraries for the remaining species. The latter

were sequenced on the Illumina HiSeq 4000 platform by the Centre for Applied Genomics at the

Hospital for Sick Children (SickKids), using 100 bp paired-end sequencing. Sequences of some

individuals have already been published by the Weir Lab. Details of individuals used, accession

numbers, and sequencing method can be found in the Supporting Online Materials.

Sequences were demultiplexed and filtered using the function *process_radtags* of the

STACKS 2.3 pipeline [37], with default parameters. Reads were trimmed for adapters and low-

quality sequences using Trimmomatric-0.38 [38], with the following parameters: TRAILING:3

SLIDINGWINDOW:4:10 MINLEN:30. Sequence reads of individuals of each species were

aligned to the reference genome of its closest relative available (Supporting Online Materials)

using Bowtie2: --phred33 --very-sensitive --mp 5,1 --score-min L, -0.6, -0.6. The *mpileup* and

call functions of BCFtools 1.12 [39] was used to call genotypes using default settings. Individual

genotype calls with < 5x depth and with a genotype likelihood of < 20 were set to missing.

For each species pair we generated three different filtered datasets, each used for different

purposes. Dataset 1 was used to calculate d_{XY} and π, for which variable and non-variable sites

are needed. We excluded sites with more than 90% missing genotype calls and SNPs with high

heterozygosity (>0.75). Dataset 2 was used to calculate F_{ST}, for which only biallelic sites are

required. SNPs with a minimum of 4 genotype calls per population were retained. We excluded

sites with minor allele frequency <0.05, with mean sequencing depth greater than 95th

percentile, and with observed heterozygosity exceeding 0.75. Dataset 3 was used to generate Site

Frequency Spectra (SFS). We used identical filtering parameters as in dataset 1, except we

removed genotype calls with < 10x depth, and to reduce linkage disequilibrium, we thinned

dataset 3 to retain only one SNP per 10 kb.

Detection of lineage-specific selective sweeps

We first ran a relatedness analysis of individuals within species, using *ngsRelate* [40].

For this analysis, we used unfiltered VCF files of all individuals per species and kept default

parameters for the estimation of Jacquard coefficients and relatedness summary statistics.

Individuals with co-ancestry coefficient (theta) > 0.1, equivalent to half sibling relationship or

higher [41] one of the two related individuals was removed.

Next, we examined signatures of selective sweeps in each of the 17 species pairs, using

summary statistics. We used Pixy 1.2.7 [42] to calculate d_{XY}, using Dataset 1, and F_{ST} and π,

using Dataset 2, across 250 kb non-overlapping windows. Only windows with a minimum of 10

SNPs were retained. The female *W*-chromosome was excluded because many individuals were

male and we did not have enough female individuals in many comparisons to include the *W*. F_{ST},

d_{XY} and π for each window were transformed into *z*-scores by subtracting the mean across

windows and then dividing it by the standard deviation across windows. Mean and standard

deviations were calculated separately for each macrochromosome (chromosomes > 2.5-6 μm

long [43]; chromosomes 1-10, 1A, 4A and Z) and jointly for the microchromosomes

(chromosomes < 2.5 μm long; chromosomes 11- 29). One tailed *P*-values were calculated from

z-scores using the *pnorm* function in R [44], with lower.tail = FALSE to detect significant peaks

of F_{ST}, and lower.tail = TRUE to detect significant drops of π and d_{XY}. Peaks or drops were

considered significant if their *p*-value was ≤ 0.05.

Windows were considered to exhibit signatures consistent with lineage-specific selective

sweeps if they possessed significantly elevated peaks in F_{ST} and significantly reduced drops in π

in one (lineage-specific sweeps) or both (sweeps targeting the same window in both species)

species of a pair, and with no significant drop in d_{XY}. In contrast, a simultaneous drop in d_{XY}

(Fig. 2D) together with elevated in F_{ST} and π in both species of a pair would be consistent with

either a selective sweep that occurred in the common ancestor or of a selective sweep that

crossed species boundaries through introgression [45]. Selective sweep signatures may span

more than one 250 Kb window. To avoid counting these as multiple sweeps, we combined

windows of selection less than 1 Mb apart as a single sweep. We standardized the number of

detected selective sweeps across species pairs by dividing the number of windows of selection

observed by the total number of windows assessed per species pair.

Estimation of divergence times

We used FASTSIMCOAL2 [24] to estimate the effective population size (N_e) of each

member of a species pair, as well as divergence time from a common ancestor. We used

Arlequin v3.5 [46] to generate a multidimensional site frequency spectrum (SFS) of species by

re-filtering SNPs from down-samples of at least 4 (diploid) individuals per species. We used a

minimum site depth cutoff of 10 per individual and manually entered the number of non-variable

sites into the SFS. We used family-specific generation times found in the literature to calibrate

both per generation mutation rates, calculated from whole-genome datasets in Feng *et al.* (2020)

and time of divergence in Myr (Supplementary Table S1). These mutation rates were used to

calibrate our demographic models. To build an understanding of the data, we explored a linear

model of $\log_{10}N_e$ and mean π in a species, with the expectation that the two values should be

correlated.

Patterns of adaptive genomic evolution through time

Using linear regression, we compared how the number of detected selective sweeps

accumulated through time for species pairs restricted to the lowlands versus different elevations.

If elevational differentiation promotes divergent selection, we would expect to find a higher

number of selective sweeps and *y*-intercept (or slope) values in species pairs that occur at

different elevations versus lowland-restricted pairs. Potential phylogenetic non-independence of

sister pairs was corrected using the following approach. We pruned a time-calibrated phylogeny

of all birds of the world [48] to retain only one representative species for each pair in the dataset,

using the *keep.tip* function in the R package APE [49]. A phylogenetic multilevel model with the

number of detected sweeps dependent on the interaction of divergence time of each pair and

category (lowland versus different elevations), and the phylogenetic variance-covariance matrix

as a varying intercept was fit in the R package brms [25,50,51]. Specifically, we used a normal

distribution to fit the model, with normally distributed priors of mean 0 and standard deviation

0.01 for the model intercept and coefficients, and Student t-distributed priors with 3 degrees of

freedom for overall standard deviation and sigma. The model was run over 4 independent

Hamiltonian Monte Carlo chains, using 5,000 burn-in and 15,000 sampling iterations each

(60,000 iterations total). Adapt delta was raised to 0.999, and maximum tree depth was raised to

12. Trace lots, potential scale reduction factors $\hat{R}$ of 1, and posterior predictive checks over 100

draws were used to assess model fit. To assess differences between species pairs in the lowlands

versus those from different elevations, we used the *hypothesis* function in brms. Specifically, we

tested for three differences in brms:

1. To assess whether elevationally differentiated or lowland-restricted species pairs

240   had a greater rate of accumulation of selective sweeps over time, we tested

241   whether the slope of elevationally differentiated species pairs was greater than

242   the slope of lowland-restricted species pairs.

243   2. To assess the number of selective sweeps at different times due to species pairs

244   being elevationally differentiated or lowland-restricted, we tested whether the

245   number of observed selective sweeps was higher in the birds from different

246   elevations than lowland-restricted birds at 0.7, 2, 3, 4, and 5 Myr. The time at 0.7

247   Myr was approximately at the start of the mid- to late-Pleistocene glacial cycles.

248   3. To assess whether there were more selective sweeps in the lowlands-restricted

249   species pairs than the elevationally differentiated species pairs extrapolated to the

250   present day, we tested whether the intercept of the lowland-restricted species

251   pairs was higher than the intercept of the elevationally-differentiated species

252   pairs.

In addition, we used linear modeling to assess the possibility that other patterns may have

driven the observed selective sweeps, as opposed to ecologically-driven parallel or divergent

selection. Using linear models in R, we tested models of the number of detected sweeps

dependent on an interaction of either mean π or $\log_{10}N_e$ and elevational category at a species-

level, highland or lowland. Another linear model was used to assess whether the accumulation of

selective sweeps in highland-lowland pairs was driven by suboscine or oscine species. This was

done in a manner consistent with the overall model of selective sweeps and divergence time in

brms, except only lowland-highland species pairs were used, and a categorical variable of

suboscine or oscine was tested in its interaction with divergence time, rather than elevational

category, as previously done. Here, a directional hypothesis test was used to assess whether the

suboscine and oscine species each accumulated sweeps over time at a slope greater than 0, as we

predicted for the species pairs representing different elevations. We also visualized the

proportion of overlap in elevational ranges between lowlands-restricted species pairs and those

from different elevations, and plotted those values against the number of observed selective

sweeps.

Results

The species categorized as from highland elevations had generally greater elevational

ranges than lowland species (Supplementary Fig. S2). Two species pairs in genera

Chlorospingus and *Ramphocelus* had substantial overlap in elevational ranges between the low

and high elevation species in these pairs, but the populations studied nevertheless appear to be

elevational replacements where they co-occur on the same mountain slopes [26].

The standardized number of detected selective sweeps varied between species pairs and

elevational categories in a model with overall R^2 of 0.65 (95% credible interval (CI): [0.34,

0.88]) (Fig. 3). Lowland-restricted pairs had a flat relationship between detected selective sweeps

and age of the species pair (slope = $8.20\text{x}10^{-5}$, 95% CI: [$-1.62\text{x}10^{-3}$, $1.52\text{x}10^{-3}$]) while the pairs

from differing elevations had a strong positive increase (slope = $3.14\text{x}10^{-3}$, 95% CI: [$7.69\text{x}10^{-4}$,

$5.31\text{x}10^{-3}$]). An evidence ratio of 155.66 from a directional hypothesis test corresponded to a

99.4% posterior probability that the pairs from differing elevations had a higher slope than the

lowland-restricted pairs. In a test for whether the slope for lowland-restricted birds was > 0, the

evidence ratio was 0.83 and posterior probability was 45.5%, thus we could not distinguish this

slope from 0. The third and final use of the *hypothesis* function in brms showed that lowland-

restricted species pairs had more observed selective sweeps until approximately 2 Myr (evidence

ratio 12.09, posterior probability 92.4%), where the number of selective sweeps was

indistinguishable between the lowland-restricted and elevationally-differentiated pairs (evidence

ratio 1.02, posterior probability 50.4%), while beyond 2 Myr in the past, the elevationally-

differentiated pairs had more observed selective sweeps (evidence ratios of 19.48, 91.31, and

143.93 and posterior probabilities of 95.1%, 98.9%, and 99.3% at 3, 4, and 5 Myr respectively).

The lowland-restricted species pairs had a higher intercept than the elevationally differentiated

species pairs (evidence ratio 23.77, posterior probability 96.0%).

Observed π or N_e were correlated (r=0.72, adjusted R^2=0.50, p=$2.01\text{x}10^{-6}$; Supplementary

Fig. S3), and neither drove observations of selective sweeps, (p=0.17 and 0.40, respectively)

(Supplementary Figs. S4; S5). A paired t-test of pairs at different elevations shows that highland

species have consistently lower π but not $\log_{10}N_e$ relative to their lowland close relative (paired t-

test: $p = 0.025$ (π), $p = 0.28$ (N_e); Fig. 4).

A limitation of our study design is the taxonomic mismatch between species categories.

All species pairs from the Amazonian lowlands are suboscines, whereas species pairs at different

elevations are phylogenetically more diverse, including two non-passerine pairs (motmots and

woodpeckers), three oscines (tanagers and icterids) and three suboscines (woodcreepers,

manakins, and flycatchers). However, we found that non-suboscine species in elevationally

differentiated pairs accumulated selective sweeps at a similar pace through time as suboscine

species, and both exhibited signatures of divergent selection as slopes were greater than 0 (oscine

evidence ratio 44.6, 97.8% posterior probability; suboscine evidence ratio 381.2, 99.7% posterior

probability) (Fig. S5). Moreover, all linear models were corrected for phylogenetic structure. As

such, we do not believe that the differences in the patterns found here between lowland-restricted

species pairs and pairs with elevation differentiation were driven by phylogenetic composition.

In addition, no relationship was evident between the proportion of elevational range overlap and

the number of selective sweeps in either category of lowlands-restricted species pairs or those

from different elevations (Supplementary Fig. S7).

Discussion

Divergent selection is celebrated as a key driver of rapid evolutionary change, while the

contribution of parallel selection is less well understood. Both have the potential to drive a rapid

accumulation of selective sweeps. Here we quantified the number of selective sweep signatures

accumulated through evolutionary time in related pairs of Amazonian species that differ greatly

in elevation and therefore likely experienced stronger divergent selection. We compared these

with pairs restricted to similar habitats at low elevations, which likely experienced weaker

divergent selection but are expected to have been strongly influenced by parallel selection

pressures. We found a substantially steeper increase in the number of selective sweeps through

time in elevationally differentiated pairs of species while species pairs at the same elevation

showed a flat relationship between sister species age and the number of observed selective

sweeps. This difference in slope was robust to phylogenetic structure, independent of genome-

wide π or N_e, and is consistent with the long-term effect of ecologically driven divergent

selection accelerating evolution over the course of millions of years in elevationally

differentiated pairs compared to lowland restricted pairs. Nevertheless, lowland restricted pairs

accumulated more selective sweeps over the glacial cycles of the past two million years, which

we suggest reflects the power of parallel selection to generate bursts of adaptation and genomic

evolution during periods of intense climate change.

For species pairs restricted to lowland regions, selective sweeps did not accumulate as a

function of time like they did in elevationally differentiated pairs. This observation would make

sense if the major climatic cycles of the past 700 kyr were overwhelmingly responsible for

selection compared to earlier periods. Under this scenario all species pairs from the same

lowland elevations older than 700 kyr (i.e., all species pairs in our dataset) should have

experienced roughly the same amount of selection, regardless of their differing time of

divergence from a common ancestor. These climatic cycles are believed to have caused a general

cooling and possibly decreased rainfall in lowland Amazonia [52,53]. Unlike highland species

which can track their preferred environments downslope during glacial periods, low elevation

species have nowhere to go, and are thus forced to adapt in parallel. Mid- to late-Pleistocene

climatic cycles would have thus intensified parallel selection pressures to adapt in lowland

species while the much more stable environment prior to these cycles would exert much weaker

selection. The flat relationship of our lowland restricted pairs between ~700 kyr and 5 Myr is

thus consistent with almost no detectable effect of selection prior to the glacial cycles, but a

substantial effect during them from ~700 kyr to the present. This contrasts with elevationally

differentiated pairs of species where the accumulation of selective sweeps through time is

strongly positive, suggesting ongoing divergent selection up to 5 Myr was driven by elevational

differentiation rather than just the effect of more recent Pleistocene glacial cycles.

In addition to the rate of selective sweep accumulation, the absolute number of observed

sweeps at different times was also informative for elucidating the roles for different types of

selection. There were more observed selective sweeps due to parallel than divergent selection

from the present up until approximately 2 Myr, after which there are more selective sweeps

attributable to divergent selection (Fig. 3). Therefore, while divergent selection had a consistent

effect over time and led to more selective sweeps in species pairs from different elevations,

parallel selection had a more prominent role in evolution during glacial cycles. This

interpretation relies on our assumption that species pairs at the same elevations experience

mostly parallel selection given the similar environments they are in. While environment is much

less variable across lowland Amazonia than it is along the elevational transects of the Andes,

some divergent selection driven by fine-scale spatial heterogeneity may have contributed to

selective sweeps in lowland-restricted species pairs [54] and our estimates cannot untangle the

contribution of these from the effect of parallel selection. Nevertheless, this finding challenges a

widespread belief that ecological and environmental differences accelerate speciation (i.e.,

ecological speciation) and are its key drivers, while parallel selection plays a less important role

in speciation [10]. Instead, our results suggest that parallel selection may be capable of driving

even faster evolutionary change when environments are unstable and species are forced to adapt

in situ.

367   Our methods are unlikely to capture all selective sweeps. First, adaptive alleles do not

always generate F_{ST} peaks when traits under selection are highly polygenic, because polygenic

traits may evolve with incomplete sweeps at many loci. In such cases, the influence of selection

on individual loci is likely too small to be detected [23]. Second, soft sweeps acting on standing

variation may generate too weak of a signature for our methods to detect, because with standing

variation the target of selection will have recombined with a variety of surrounding genetic

variants, which then fail to materialize as a sweep signature when the target is fixed (Fig. 2). It

thus remains possible that lowland-restricted species did experience selection before the mid- to

late-Pleistocene glacial cycles in primarily polygenic traits or soft sweeps. The stronger parallel

selection forces induced by the glacial cycles would then have driven an increase in hard sweep

signatures more recently which is the primary signal we detect. This scenario is also plausible

because greater nucleotide diversity observed in the lowland species (Supplementary Fig. S2)

means there is a larger pool of standing genetic variants present that soft selection could have

acted upon. Given these considerations, we can only conclude that our different patterns

uncovered between species pairs that do and do not have elevational differentiation primarily

reflect the effect of hard selective sweeps while the effect of soft sweeps and selection on highly

polygenic traits remains unknown.

384   Another limitation of this work is that genomic architecture may have influenced the

results. We analyzed F_{ST} and π in windows of at least 250 kb, thus, smaller sweeps may go

undetected and their detection will require whole genome sequencing. The larger sweeps we

were able to detect may be associated with architecture such as chromosomal inversions, which

are understood to support local adaptation and selective sweeps in a range of species [55,56].

Also, centromeres and telomeres possess low recombination rates that facilitate sweeps, which

lower π and elevate F_{ST} over broad genomic regions, thereby generating strong sweep signatures

that are easily detected (see Cutter & Payseur 2013). However, there are only a limited number

of these genomic features, and if they were a major contributor to our detections of selective

sweeps, the number of such selective sweep signatures we detected might accumulate rapidly

over short time scales and then remain constant with increasing time, as sweeps in older species

pairs would be overwritten by subsequent sweeps. In such a scenario, the elevationally-

differentiated pairs should also exhibit a flat relationship between species pair age and the

number of selective sweeps, like lowland-restricted pairs. Instead, we uncovered a strong

positive slope for elevationally-differentiated pairs (Fig. 3), indicating that avian genomes are

capable of accumulating detectable selective sweep signatures through time. Therefore, the

approach we adopted for detecting divergent selection might be influenced by, but was not

invalidated by, conserved genomic architecture.

In conclusion, despite the limitations of our approach, we found clear differences

between elevationally differentiated and lowland restricted species pairs in terms of selective

sweep accumulation through time. Over the five-million-year timescale of our dataset, strong

divergent selection associated with elevational differentiation resulted in a much greater number

of selective sweeps than seen in lowland restricted pairs, which confirms the importance of

divergent selection as has previously been emphasized (e.g., Schluter 2009). Nevertheless, our

data suggests lowland restricted species pairs accumulated more selective sweeps over the major

Pleistocene glacial cycles consistent with climate change forcing them to adapt *in situ*, while

highland species could track their preferred environment up and down slope with less need for

adaptation. Parallel selection may thus result in limited adaptation when environments are highly

constant through time (i.e., prior to major Pleistocene glacial cycles) but may elevate the rate of

accumulation of selective sweeps during periods of environmental instability (i.e., during major

Pleistocene glacial cycles) when parallel environmental changes force species to adapt. Parallel

selection is likely to target different genes in different populations, leading to genomic

differentiation and the accumulation of genetic incompatibilities that ultimately are believed to

play a key driving force in Amazonian speciation [57,58]. Our findings are consistent with a

recent meta-analysis of birds, mammals, reptiles, and other groups which found preliminary

evidence that parallel selection was a generally stronger force in shaping trait evolution than

divergent natural selection [14]. Together, these results suggest parallel selection may drive

adaptation and speciation during periods of intense environmental change in the most species

rich region of the planet. While the role of divergent natural selection has been well documented,

there remains a great need for studies to address the contribution of parallel selection to

adaptation and speciation.

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Literature cited

1. Darwin C, Wallace A. 1858 On the Tendency of Species to form Varieties; and on the Perpetuation of Varieties and Species by Natural Means of Selection. *Journal of the Proceedings of the Linnean Society of London. Zoology* **3**, 45–62. (doi:10.1111/j.1096-3642.1858.tb02500.x)

2. Maan ME, Seehausen O. 2011 Ecology, sexual selection and speciation: Ecology, sexual selection and speciation. *Ecology Letters* **14**, 591–602. (doi:10.1111/j.1461-0248.2011.01606.x)

3. Ogden R, Thorpe RS. 2002 Molecular evidence for ecological speciation in tropical habitats. *Proceedings of the National Academy of Sciences* **99**, 13612–13615. (doi:10.1073/pnas.212248499)

4. Price T. 2008 *Speciation in birds*. Greenwood Village, Colo: Roberts and Co.

5. Schluter D. 2000 *The ecology of adaptive radiation*. Oxford: Oxford University Press.

6. Campagna L, Toews DPL. 2022 The genomics of adaptation in birds. *Current Biology* **32**, R1173–R1186. (doi:10.1016/j.cub.2022.07.076)

7. Nosil P, Funk DJ, Ortiz-Barrientos D. 2009 Divergent selection and heterogeneous genomic divergence. *Molecular Ecology* **18**, 375–402. (doi:10.1111/j.1365-294X.2008.03946.x)

8. Nosil P. 2012 *Ecological speciation*. Oxford ; New York: Oxford University Press.

9. Rundle HD, Nosil P. 2005 Ecological speciation: Ecological speciation. *Ecology Letters* **8**, 336–352. (doi:10.1111/j.1461-0248.2004.00715.x)

10. Schluter D. 2009 Evidence for Ecological Speciation and Its Alternative. *Science* **323**, 737–741. (doi:10.1126/science.1160006)

11. Cerca J. 2023 Understanding natural selection and similarity: Convergent, parallel and repeated evolution. *Molecular Ecology* **32**, 5451–5462. (doi:10.1111/mec.17132)

12. Mani GS, Clarke BC. 1990 Mutational Order: A Major Stochastic Process in Evolution. *Proceedings of the Royal Society of London. Series B, Biological Sciences* **240**, 29–37.

13. Nosil P, Flaxman SM. 2011 Conditions for mutation-order speciation. *Proc. R. Soc. B* **278**, 399–407. (doi:10.1098/rspb.2010.1215)

14. Anderson SAS, Weir JT. 2022 The role of divergent ecological adaptation during allopatric speciation in vertebrates. *Science* **378**, 1214–1218. (doi:10.1126/science.abo7719)

15. Ridgely RS, Greenfield PJ. 2001 *The birds of Ecuador*. Ithaca, NY: Comstock Pub.

16. Smith BT *et al.* 2014 The drivers of tropical speciation. *Nature* **515**, 406–409. (doi:10.1038/nature13687)

17. Hooghiemstra H. 1989 Quaternary and upper-pliocene glaciations and forest development in the tropical andes: Evidence from a long high-resolution pollen record from the sedimentary basin of Bogotá, Colombia. *Palaeogeography, Palaeoclimatology, Palaeoecology* **72**, 11–26. (doi:10.1016/0031-0182(89)90129-6)

18. Hooghiemstra H, Melice JL, Berger A, Shackleton NJ. 1993 Frequency spectra and paleoclimatic variability of the high-resolution 30–1450 ka Funza I pollen record (Eastern Cordillera, Colombia). *Quaternary Science Reviews* **12**, 141–156. (doi:10.1016/0277-3791(93)90013-C)

19. Hooghiemstra H, Van Der Hammen T. 2004 Quaternary Ice-Age dynamics in the Colombian Andes: developing an understanding of our legacy. *Phil. Trans. R. Soc. Lond. B* **359**, 173–181. (doi:10.1098/rstb.2003.1420)

20. Wolf JBW, Ellegren H. 2017 Making sense of genomic islands of differentiation in light of speciation. *Nat Rev Genet* **18**, 87–100. (doi:10.1038/nrg.2016.133)

21. Boitard S, Schlotterer C, Nolte V, Pandey RV, Futschik A. 2012 Detecting Selective Sweeps from Pooled Next-Generation Sequencing Samples. *Molecular Biology and Evolution* **29**, 2177–2186. (doi:10.1093/molbev/mss090)

22. Hejase HA, Salman-Minkov A, Campagna L, Hubisz MJ, Lovette IJ, Gronau I, Siepel A. 2020 Genomic islands of differentiation in a rapid avian radiation have been driven by recent selective sweeps. *Proc Natl Acad Sci USA* **117**, 30554–30565. (doi:10.1073/pnas.2015987117)

23. Stephan W. 2016 Signatures of positive selection: from selective sweeps at individual loci to subtle allele frequency changes in polygenic adaptation. *Mol Ecol* **25**, 79–88. (doi:10.1111/mec.13288)

24. Excoffier L, Foll M. 2011 fastsimcoal: a continuous-time coalescent simulator of genomic diversity under arbitrarily complex evolutionary scenarios. *Bioinformatics* **27**, 1332–1334. (doi:10.1093/bioinformatics/btr124)

25. Bürkner P-C. 2021 Bayesian Item Response Modeling in *R* with **brms** and *Stan*. *J. Stat. Soft.* **100**. (doi:10.18637/jss.v100.i05)

26. Schulenberg TS, Stotz DF, Lane DF, O'Neill JP, Parker III TA. 2007 *Birds of Peru*. 1st edn. Princeton, New Jersey: Princeton University Press.

27. Del Hoyo J, Collar N, Kirwan GM, Sharpe C. 2020 Rondonia Warbling-Antbird (Hypocnemis ochrogyna). In *Birds of the World* (eds SM Billerman, BK Keeney, PG Rodewald, TS Schulenberg), Cornell Lab of Ornithology. (doi:10.2173/bow.ronwaa1.01)

28. Del Hoyo J, Collar N, Kirwan GM. 2020 Spix's Warbling-Antbird (Hypocnemis striata). In *Birds of the World* (eds SM Billerman, BK Keeney, PG Rodewald, TS Schulenberg), Cornell Lab of Ornithology. (doi:10.2173/bow.spiwaa1.01)

29. Marantz CA, Aleixo A, Bevier LR, Patten MA. 2020 Spix's Woodcreeper (Xiphorhynchus spixii). In *Birds of the World* (eds SM Billerman, BK Keeney, PG Rodewald, TS Schulenberg), Cornell Lab of Ornithology. (doi:10.2173/bow.spiwoo1.01)

30. Snow D. 2020 Red-headed Manakin (Ceratopipra rubrocapilla). In *Birds of the World* (eds SM Billerman, BK Keeney, PG Rodewald, TS Schulenberg), Cornell Lab of Ornithology. (doi:10.2173/bow.rehman1.01)

31. Snow D. 2020 Crimson-hooded Manakin (Pipra aureola). In *Birds of the World* (eds SM Billerman, BK Keeney, PG Rodewald, TS Schulenberg), Cornell Lab of Ornithology. (doi:10.2173/bow.crhman1.01)

32. Snow D, De Juana E. 2020 Snow-capped Manakin (Lepidothrix nattereri). In *Birds of the World* (eds SM Billerman, BK Keeney, PG Rodewald, TS Schulenberg), Cornell Lab of Ornithology. (doi:10.2173/bow.sncman1.01)

33. Snow D, Sharpe C. 2020 Opal-crowned Manakin (Lepidothrix iris). In *Birds of the World* (eds SM Billerman, BK Keeney, PG Rodewald, TS Schulenberg), Cornell Lab of Ornithology. (doi:10.2173/bow.opcman1.01)

34. Zimmer K, Isler ML, De Juana E. 2020 Silvered Antbird (Sclateria naevia). In *Birds of the World* (eds SM Billerman, BK Keeney, PG Rodewald, TS Schulenberg), Cornell Lab of Ornithology. (doi:10.2173/bow.silant1.01)

35. Elshire RJ, Glaubitz JC, Sun Q, Poland JA, Kawamoto K, Buckler ES, Mitchell SE. 2011 A Robust, Simple Genotyping-by-Sequencing (GBS) Approach for High Diversity Species. *PLoS ONE* **6**, e19379. (doi:10.1371/journal.pone.0019379)

36. Luzuriaga-Aveiga V, Ugarte M, Weir JT. 2021 Distinguishing genomic homogenization from parapatric speciation in an elevationally replacing pair of *Ramphocelus* tanagers. *Mol Ecol* **30**, 5517–5529. (doi:10.1111/mec.16128)

37. Catchen JM, Amores A, Hohenlohe P, Cresko W, Postlethwait JH. 2011 *Stacks* : Building and Genotyping Loci *De Novo* From Short-Read Sequences. *G3 Genes|Genomes|Genetics* **1**, 171–182. (doi:10.1534/g3.111.000240)

38. Bolger AM, Lohse M, Usadel B. 2014 Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* **30**, 2114–2120. (doi:10.1093/bioinformatics/btu170)

39. Li H. 2011 A statistical framework for SNP calling, mutation discovery, association mapping and population genetical parameter estimation from sequencing data. *Bioinformatics* **27**, 2987–2993. (doi:10.1093/bioinformatics/btr509)

40. Korneliussen TS, Moltke I. 2015 NgsRelate: a software tool for estimating pairwise relatedness from next-generation sequencing data. *Bioinformatics* , btv509. (doi:10.1093/bioinformatics/btv509)

41. Ackerman MS, Johri P, Spitze K, Xu S, Doak TG, Young K, Lynch M. 2017 Estimating Seven Coefficients of Pairwise Relatedness Using Population-Genomic Data. *Genetics* **206**, 105–118. (doi:10.1534/genetics.116.190660)

42. Korunes KL, Samuk K. 2021 PIXY : Unbiased estimation of nucleotide diversity and divergence in the presence of missing data. *Molecular Ecology Resources* **21**, 1359–1368. (doi:10.1111/1755-0998.13326)

43. Griffin DK, Robertson LBW, Tempest HG, Skinner BM. 2007 The evolution of the avian genome as revealed by comparative molecular cytogenetics. *Cytogenet Genome Res* **117**, 64–77. (doi:10.1159/000103166)

44. R Core Team. 2021 R: A language and environment for statistical computing.

45. Irwin DE, Alcaide M, Delmore KE, Irwin JH, Owens GL. 2016 Recurrent selection explains parallel evolution of genomic regions of high relative but low absolute differentiation in a ring species. *Mol Ecol* **25**, 4488–4507. (doi:10.1111/mec.13792)

46. Excoffier L, Lischer HEL. 2010 Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources* **10**, 564–567. (doi:10.1111/j.1755-0998.2010.02847.x)

47. Feng S *et al.* 2020 Dense sampling of bird diversity increases power of comparative genomics. *Nature* **587**, 252–257. (doi:10.1038/s41586-020-2873-9)

48. Pulido-Santacruz P, Weir JT. 2016 Extinction as a driver of avian latitudinal diversity gradients: EXTINCTION AS A DRIVER OF DIVERSITY GRADIENTS. *Evolution* **70**, 860–872. (doi:10.1111/evo.12899)

49. Paradis E, Schliep K. 2019 ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics* **35**, 526–528. (doi:10.1093/bioinformatics/bty633)

50. Bürkner P-C. 2017 **brms** : An *R* Package for Bayesian Multilevel Models Using *Stan*. *J. Stat. Soft.* **80**. (doi:10.18637/jss.v080.i01)

51. Bürkner P-C. 2018 Advanced Bayesian Multilevel Modeling with the R Package brms. *The R Journal* **10**, 395. (doi:10.32614/RJ-2018-017)

52. Ramírez-Barahona S, Eguiarte LE. 2013 The role of glacial cycles in promoting genetic diversity in the Neotropics: the case of cloud forests during the Last Glacial Maximum. *Ecol Evol* **3**, 725–738. (doi:10.1002/ece3.483)

53. Rull V. 2011 Neotropical biodiversity: timing and potential drivers. *Trends in Ecology & Evolution* **26**, 508–513. (doi:10.1016/j.tree.2011.05.011)

54. Michot V, Corpetti T, Ronchail J, Espinoza JC, Arvor D, Funatsu BM, Dubreuil V. 2024 Seasonal types in homogeneous rainfall regions of the Amazon basin. *Intl Journal of Climatology* **44**, 1224–1244. (doi:10.1002/joc.8380)

55. Cutter AD, Payseur BA. 2013 Genomic signatures of selection at linked sites: unifying the disparity among species. *Nat Rev Genet* **14**, 262–274. (doi:10.1038/nrg3425)

56. Wellenreuther M, Bernatchez L. 2018 Eco-Evolutionary Genomics of Chromosomal Inversions. *Trends in Ecology & Evolution* **33**, 427–440. (doi:10.1016/j.tree.2018.04.002)

57. Cronemberger ÁA, Aleixo A, Mikkelsen EK, Weir JT. 2020 Postzygotic isolation drives genomic speciation between highly cryptic *Hypocnemis* antbirds from Amazonia. *Evolution* **74**, 2512–2525. (doi:10.1111/evo.14103)

58. Pulido-Santacruz P, Aleixo A, Weir JT. 2018 Morphologically cryptic Amazonian bird species pairs exhibit strong postzygotic reproductive isolation. *Proc. R. Soc. B.* **285**, 20172081. (doi:10.1098/rspb.2017.2081)

Tables & Figures

Figure 1. Geographic ranges of two example species pairs overlaid on a topographical map of South America. In A) the foothill forest zone occurs from approximately 600 to 1,300 m (foothill), the epiphyte encrusted subtropical montane forest zone from approximately 1,300 to 2,400 m (subtropical), the temperate montane forest zone to the treeline at approximately 3,300 m (temperate), and the alpine zone exists above 3,300 m. One species pair has the Amazonian Motmot (*Momotus momota*), in red, distributed in lowland forest from 0 to 700 m and its sister species, the Andean Motmot (*M. aequatorialis*), in blue, distributed in Andean cloud forest from 1000 to 2300 m. The other species pair has both species distributed in low elevation rainforest in different regions of the Amazon basin, with the Opal-crowed Manakin *(Lepidothrix iris)* distributed east of the Tapajos River and the Snowy-crowed Manakin (*L. nattereri*) distributed west of the river. Insets A and B show 250 km-long transects running perpendicular to the contact zones between each species pair. Coloured bands show the elevational profiles of each species. Shaded relief topographical base map obtained from NASA (https://www.nasa.gov). Major rivers of the Amazon basin are shown in blue.

Figure 2. Hard and soft selective sweeps, with corresponding expectations of π, F_{ST} and d_{XY} under divergent and parallel selection. In hard sweeps (A–C, E), a positively selected de novo mutation (turquoise circle) arises in a single genomic background. Selection causes fixation of this mutation together with closely linked surrounding blocks of the genomic background that have not had time to experience recombination (coloured segments). The soft sweep (D) is similar, but the allele that is the target of selection is segregating in the population prior to becoming positively selected, resulting in all genomic backgrounds it is associated with increasing in frequency. The resulting patterns of π, F_{ST} and d_{XY} after selection can be used to detect potential selective sweeps. Hard sweeps at different loci (A) result in drops in π only in the species that is the target of selection for a given locus. The drop in π elevates F_{ST}. Hard sweeps at the same site (B–C) result in a drop in π in both species and an increase in F_{ST}. They produce almost identical patterns of F_{ST} and π under divergent (B) and parallel (C) selection but differ only in the value of F_{ST} at the site that is the target of selection, with the site having an F_{ST} of 1 under divergent selection (where different alleles at the site of selection are fixed), and an F_{ST} of 0 under parallel selection (where the same allele at the site of selection is fixed). Sweeps resulting from divergent or parallel selection cannot easily be discriminated. Soft sweeps at different loci (D) produce similar patterns in π and F_{ST} as hard sweeps, but the drops in π and increases in F_{ST} are less extreme, making them harder to detect. A sweep in the common ancestor of two populations (E) should result in both daughter populations inheriting low π around the selected site, but, unlike sweeps at the same site in both populations, an ancestral sweep results in a drop in F_{ST} and in d_{XY} between the daughter populations. Turquoise circles show the allele that is the target of selection. Coloured lines represent different haplotype blocks.

Figure 3. Number of observed selective sweeps accumulated between species pairs versus their divergence times, calculated from the nuclear genome using demographic modeling. Species pairs restricted to the Amazonian lowlands are shown with purple, and pairs occupying different elevational zones from the Amazonian lowlands towards the high Andes are shown in orange. Lines show model fits with phylogenetic relatedness accounted for. The lightest shading shows 95% credible intervals, while darker shading shows 10% credible intervals to emphasize posterior distributions underlying estimates of this linear model. Dashed lines reach from the most recent estimate of each line to the present day of 0 Myr, to emphasize selective sweep accumulation in the mid- to late-Pleistocene.

Figure 4. Estimated effective population size (log$_{10}N_{e}$, top panels A and B) and box and whisker plots of average genome-wide nucleotide diversity (mean π, bottom panels C and D) across 250 Kb genomic windows. Species delineated by category, between pairs from different elevations (A and C) and lowland-restricted pairs (B and D). Lowland species are in purple, while highland species are in orange.

Supporting Information

Supplementary Table S1: Generation times and mutation rates used in coalescent modelling for major families represented in the dataset.

Family	Genus	Generation time	Mutation rate
Icteridae	*Psarocolius*	Mean = 1.83	2.096×10^{-9}
Thraupidae	*Ramphocelus*	Mean = 2.86	3.98×10^{-9}
Passerellidae	*Chlorospingus*	Mean = 2.33	2.99×10^{-9}
Thamnophilidae	*Hypocnemoides* *Hypocnemis* *Sclateria*	Mean = 2.44	4.14×10^{-9}
Dendrocolaptidae	*Xiphorhynchus* *Dendrocincla*	Mean = 2.83	4.14×10^{-9}
Tyrannidae	*Mionectes*	Mean = 1.98	3.73×10^{-9}
Pipridae	*Ceratopipra* *Pipra* *Lepidothrix* *Manacus* *Chiroxiphia*	Mean = 3.34	6.06×10^{-9}
Picidae	*Dryobates*	Mean = 2.86	6.89×10^{-9}
Momotidae	*Momotus*	Mean = 2.85	5.84×10^{-9}

Figure S1. Phylogeny of the taxa included in the dataset. Tips of the tree represent sister species or subspecies pairs, colour-coded by their elevational category: purple for lowland-restricted pairs and orange for pairs at different elevations. *Dendrocincla fuliginosa* from the lowlands represents two subspecies, *D. f. atrirostris* and *D. f. rufoolivacea*. *Dendrocincla* at different elevations represents *D. merula* and *D. tyrannina.*

Figure S2. Elevational ranges for species in the present study. For species where both a core range and maximum and minimum observations were recorded, dashed lines were used to show the extreme ends of a range. However, this information was not available consistently. While genera *Chlorospingus* and *Ramphocelus* (bold) had largely overlapping species elevational ranges, they were allopatric and likely experienced different historical environments.

Figure S3. Linear regression between effective population size (N_e) and mean genome-wide nucleotide diversity (π).

Figure S4. Regression analyses between number of lineage-specific windows of selection and average genome-wide nucleotide diversity, π, for individual species (not species pairs). Here, mean π interacted with elevational category of lowland or highland. Species from the Andean highlands are shown in orange, and lowland Amazonian species are shown in purple. This model was not significant (*p*=0.17).

Figure S5. Regression analyses between the number of lineage-specific windows of selection and $\log_{10}$ effective population size ($\log_{10}N_e$) for individual species. Here, $\log_{10}N_e$ interacted with elevational category of lowland or highland. Species from the Andean highlands are shown in orange, and lowland Amazonian species are shown in purple. This model was not significant (p=0.40).

Figure S6. Number of observed selective sweeps accumulated between species pairs versus their divergence times, calculated from the nuclear genome using demographic modeling for only elevationally differentiated species pairs. Oscine species pairs are shown in purple, and suboscine pairs are shown in orange. Lines show model fits with phylogenetic relatedness accounted for. Shading shows the 95% credible intervals.

Figure S7. Proportion of elevational range overlap for each sister species pair, within lowlands-restricted or different elevation categories, against observed selective sweeps.

A

alpine

temperate

subtropical — Andean Motmot

foothill

lowland — Amazonian Motmot

B

Tapajos River

Snowy-crowned Manakin ← → Opal-crowned Manakin

lowland

Transect perpendicular to contact zone