Size-dependent invasion and therapeutic phenotype of 42MGBA glioblastoma spheroids

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ABSTRACT

Glioblastoma (GBM) is one of the most common malignant brain tumors, with patient mortality driven by invasion into the surround brain microenvironment and drug resistance. Multicellular spheroids are increasingly a common model to study GBM invasion and drug response in engineered biomaterials. However, a key design feature of tumor spheroid studies is the size of each spheroid (number of cells, diameter). Given the heterogenous growth of GBM cells at the surgical margin, spheroids of different sizes may also have clinical relevance. Here, we define shifts in behavior and drug response of wild type and temozolomide (TMZ) resistant GBM spheroids as a function of initial spheroid size. GBM spheroids ranging from 100 to 12,000 cells in size were embedded into a methacrylamide-functionalized gelatin (GelMA) hydrogel. GBM spheroid size had an inverse relationship with the number of apoptotic cells. We observed significant spheroid size dependent effects on TMZ efficacy for both TMZ resistant and wild type cells. Interestingly, high single doses of TMZ were more effective in reducing three-dimensional migration from smaller spheroids than metronomic dosing while high single dose and metronomic dosing were equally effective in reducing invasion for large TMZ-resistant spheroids. Our study highlights the importance of considering and reporting spheroid size for cancer tissue engineering studies considering invasion and drug resistance. It also informs future studies of residual GBM cohorts at the tumor margins most responsible for patient relapse and mortality.

Keywords: Glioblastoma; tumor spheroid; drug resistance; hydrogel; temozolomide

Running Head: Size-dependent phenotypes of GBM spheroids

1. Introduction

Glioblastoma (GBM) is the most common, aggressive, and lethal form of primary brain cancer [1,2]. While advances in radio- and chemotherapy have extended median survival from 12 to ~15mo, overall survival (<5% after 5 yrs) remains poor and largely unchanged over the past two decades [3-10]. Current standard-of-care includes maximal surgical resection followed by radiation and chemotherapy with the alkylating agent temozolomide (TMZ) [11]. However, mortality is driven by rapid diffusive spreading of GBM cells beyond the surgical margin [12-15]. GBM tumors recur rapidly (6.9mo post debulking) and in close proximity (>90% within 2cm) of the original tumor [16,17]. It is therefore important to develop strategies to target GBM cells that spread diffusely into tissue microenvironment beyond the surgical margin [18]. While studies of GBM progression and therapeutic response are commonly performed using *in vivo* assays [19-22], such models are less well-suited to examine both the kinetics of cell invasion as well as for nuanced studies of the role of the tumor microenvironment on progression.

In vitro experiments using both two-dimensional cell cultures [23,24] as well as three-dimensional biomaterial models of the brain tumor microenvironment [15,25-27] offer a lens to elucidate the instructive nature of the composition, mechanical properties, and multicellular nature of the GBM tumor microenvironment. Recent efforts have identified key features of the GBM microenvironment such as chemical and mechanical gradients [28-30] as well as fluid flow [31-33]. Our group has developed a methacrylamide-functionalized gelatin (GelMA) hydrogel to study the role of matrix stiffness, hypoxia, and hyaluronic acid content on GBM progression using both immortalized cell lines and patient-derived xenografts [34-36]. In these studies, metrics of GBM response can be gathered from low-density cell populations encapsulated in, or high-density cell spheroids embedded into, the hydrogel network. Both modes of study are important because single cells and cohorts of GBM cells may exist in the tumor margins after debulking [37-40]. We used these hydrogel models to show angiocrine signals influence GBM invasion and TMZ

resistance [41-44] and more recently to study the emergence of TMZ resistance [45] in response to physiologically-relevant single and metronomic [46,47] TMZ doses using isogenic pairs of TMZ-resistant and responsive GBM cell lines developed by *Tiek et al.* [48]. Metronomic dosing offers an alternative to a higher single dosage [46], has been suggested to offer better penetration and potentially circumvent resistance mechanisms seen with high single drug doses [49], and is more consistent with the clinical use of TMZ. Our previously used GBM spheroids to profile patterns of collective cell invasion in response to changes in matrix composition, hypoxia, and the presence of GBM stem cell subpopulations [36,50-53]. However, previous studies of the role of confinement and biophysical signals on liver development [54-56] and other cancer stem cell phenotypes [57] suggest spheroid size may be an essential design principal. Larger cohorts have been hypothesized to affect the emergence of hypoxia and necrosis as well as to induce changes in gene expression that may underlie invasion and chemoresistance [58]. However, given the likelihood that multiple scales of GBM cohorts are present in the tumor margins it is essential to understand the degree spheroid size influences metrics of GBM phenotype.

The goal of this project is to define the influence of GBM spheroid size on cell activity, invasion, and drug response within a model GelMA hydrogel. We fabricated spheroids from both TMZ-resistant or wild type (WT) GBM cell lines, with sizes ranging between 100 and 12,000 cells per spheroid. We report the effect of spheroid size on GBM cell growth and gene expression, apoptosis, and invasion. We define the effect of single vs. metronomic TMZ dosing on GBM spheroids as a function of size and TMZ-resistance status. Together, these establish design principles regarding spheroid sizes necessary to evaluate therapeutic compounds as well as to establish biophysical thresholds (e.g., spheroid surface area to volume) that alter GBM phenotypes *in vitro*.

2. Materials and Methods

2.1. Cell culture

Wild type (42MGBA-WT) and Temozolomide resistant (42MGBA-TMZres) 42MGBA cell pairs were acquired from the Riggins lab at Georgetown University [48]. 42MGBA cells were cultured using Dulbecco's Modified Eagle Medium (DMEM) with 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin; media was changed every 48 hours. Cells were maintained in incubators at 37°C and 5% CO_2 and used within 10 passages.

2.2. Spheroid formation

GBM spheroids were generated using two well established protocols. Ultra-low attachment plate method: Wild type (42MGBA-WT) and TMZ resistant (42MGBA-TMZres) GBM cells were formed into spheroids using the nutation method [44,59]. Briefly, GBM cells were seeded in Corning® spheroid microplates, incubated at 37°C on a rotating incubator shaker (ThermoFisher, Canoga Park, CA) at 60 rpm for 48 hours. Aggrewell™ microwell method: Wild type (42MGBA-WT) and TMZ resistant (42MGBA-TMZres) GBM spheroids were also generated at a lager quantity required for flow cytometry analysis. Briefly, AggreWell 400 or 800 microplates (STEMCELL Technologies, Vancouver, Canada) were pre-treated with anti-adherence rising solution (STEMCELL Technologies, Vancouver, Canada) for 5 minutes. The anti-adherence rinsing solution was then removed and replaced with 2 mL cell culture media seeded with the desired number of cells to form each spheroid.

2.3. Flow cytometry

All flow cytometry measurements were performed using a FACSymphony A1 configuration (BD Bioscience, Franklin Lake, NJ). Briefly, spheroids were recovered from biomaterial culture, dissociated using 0.5% Trypsin, then resuspended in 500 μL 5% FBS for analysis in biological triplicate. Live cells were gated by forward scatter (FSC) and side scatter (SSC). Fluorescence

data was collected for GFP (excitation laser: 488nm; bandpass filter: 530/30nm) and PE-Texas Red (excitation laser: 561nm; bandpass filter: 610/20nm) with a minimum of 50,000 singlet events were collected for each sample. Flow cytometry data was analyzed using FACSDiva™ Software (BD Bioscience, Franklin Lake, NJ) then graphed using Prism 8 software (GraphPad). Apoptotic cells were gated as PE-Texas Red (Bobo-3 iodide, Thermo Fisher Scientific, Waltham, MA)) and FITC (CalcienAM; Thermo Fisher Scientific, Waltham, MA) positive. Necrotic cells were gated as PE-Texas Red (Bobo-3 iodide, Thermo Fisher Scientific, Waltham, MA) positive (Figure S2).

2.4. GFP transduction of 42MGBA Cell lines

42MGBA cells were transfected using the CMV-rFLuc-T2A-GFP-mPGK-Puro lenti-labeler™ plasmid (Systems Bioscience, Palo Alto, CA) per manufacturer's instructions. Briefly, 7,500 42MGBA cells were seeded in 48 well plates with both TransDux Max and Max enhancers diluted in culture media at a final concentration of 1x. The multiplicity of infection (MOI) used was calculated based on the infectious forming units (IFUs) provided by the manufacturer, with lenti-labeler added to the cells at an expected MOI of 50. Cells were incubated at 37°C and 5% CO_2 for 72 hours before sorting (BigFoot Invitrogen Thermofisher Spectral Cell Sorter, ThermoFisher, Canoga Park, CA) for GFP positive cells. Isolated GFP expressing cells were expanded for 2 passages before experiments.

2.5. GelMA synthesis

GelMA macromers for all hydrogel culture were synthesized as previously described [60]. Briefly, porcine gelatin type A, 300 bloom (Sigma Aldrich, St. Louis, MO) was dissolved in carbon-bicarbonate buffer (pH 9.4) at 65 °C. 15 µL methacrylic anhydride (Sigma Aldrich, St. Louis, MO) was added dropwise per gram of gelatin, and the reaction proceeded for 2 hrs with continues stirring (250 RPM). The reaction was quenched with pre-heated deionized water and

dialyzed for seven days against deionized water with daily exchange. The product was then frozen at -20°C and lyophilized for 7 days. HNMR was used to determine the degree of functionalization (DOF). GelMA with DOF of 85% (data not shown) was used in this study.

2.6. Hydrogel gel encapsulation and Invasion assay

GFP expressing 42MGBA-WT or 42MGBA-TMZres spheroids was suspended in a 5wt% GelMA precursor solution and then pipetted into 20 μL volume (cylindrical) Teflon molds (1 spheroid per hydrogel construct). GelMA hydrogels were polymerized in the presence of lithium acylphosphinate (LAP, 0.1% w/v) and exposed to UV light for 35s using a UV lamp (λ = 365 nm, 5.79 mW/cm^2) [61]. GBM cell invasion assay was performed using a DMi8 Yokogawa W1 spinning disk confocal microscope with a Hamamatsu EM-CCD digital camera (Leica Microsystems, Buffalo Grove, IL) per previously described methods [44]. Overall GBM cell outgrowth area was quantified by using ImageJ (NIH, Bethesda, MD). Briefly, sequential slices (5 μm thick) were imaged along the hydrogels' depth. Maximum intensity projections were assembled from approximately 50 slices, with the composite image then binarized and analyzed with the analyze particle function on ImageJ (NIH, Bethesda, MD). Six independent spheroid outgrowth measurements were gathered per experimental group as a biological replicate.

2.7. TMZ treatment

We performed TMZ studies in GelMA hydrogels as previously described [45]. Briefly, temozolomide (TMZ, Calbiochem via Millipore Sigma, Burlington, MA) was dissolved in dimethyl sulfoxide (DMSO) at a concentration of 200 mM. DMSO (□ 0.5% v/v); DMSO alone was used as a vehicle control for all experimental studies. Hydrogels containing GBM spheroids were cultured for 24 hours prior to the administration of TMZ. Hydrogels were subsequently treated with one of the following TMZ dose regimens: a single dose of 500 μM or five daily doses of

100μM (a cumulative dose of 500 μM). Media was replaced for both groups daily with either the appropriate concentration of TMZ or DMSO control.

2.8. Statistics

Statistics were performed using SPSS. Normality of data was determined using the Shapiro–Wilk test, and equality of variance was determined using Levene's test. For normal data, comparisons between two groups were performed using a *t*-test, while comparisons between multiple groups were performed using a one-way ANOVA followed by Tukey's post-hoc when assumptions were met. In the case where data was not normal or groups had unequal variance, comparisons between two groups were performed using a Mann–Whitney test, while comparisons between multiple groups were performed using a Kruskal–Wallis test with Dunn's post-hoc. Significance was determined as $p < 0.05$. All quantitative analyses were performed on hydrogels or microfluidic assays set up across at least three independent experiments.

3. Results

3.1. Library of GBM spheroids as a function of cell size

We first created spheroids libraries for both wild type (42MGBA-WT) and TMZ-resistant (42MGBA-TMZres) GBM cell lines ranging in size from 100 cells to 12,000 cells (Figure 2A). We quantified the resultant size and morphology of the spheroids via overall projected area and circularity metrics from bright-field microscopy images. Spheroid projected area scale with numbers of cells and broadly, all spheroids displayed consistent circularity indices of greater than 60%. Spheroids generated from TMZ-resistant GBM cells were significantly larger than those generated from the same numbers of wild-type GBM cells (Figure 2B). And while not statistically significant, TMZ-resistant spheroids trended towards increased circularity than GBM wild type spheroids for all sizes up to up to 2500 cells; smaller ($\leq$2500 cells) TMZ-resistant

spheroids also trended toward higher circularity than TMZ-resistant spheroids of larger (≥5000 cells) size (Figure 2C). We characterized the packing of GBM cells using fluorescent images obtained from CMV-rFLuc-T2A-GFP-mPGK-Puro lenti-labeler™ labeled wildtype and TMZ-resistant cells formed into 10,000 cell spheroids (Figure S1). TMZ-resistant spheroids appear to be more densely packed with a brighter GFP core, whereas the fluorescent signal for the wild type spheroids display a more diffuse organization.

3.2. Patterns of GBM proliferation and apoptosis is influenced by spheroid size and TMZ-resistant status.

We subsequently examined patterns of cell proliferation and death within GBM spheroids as a function of spheroid size and TMZ resistant status (42MGBA-WT vs. 42MGBA-TMZres). Immunostaining was performed 48 hours after encapsulation spheroids into GelMA hydrogels. Broadly, we observed Ki-67+ proliferative cells in all 42MGBA-WT spheroids regardless of spheroid size (Figure 3A). Interestingly, we observed higher levels of staining in 42MGBA-TMZres spheroids increasingly at the spheroid periphery, with the effect most pronounced in larger spheroids (Figure 3B). We also interrogated patterns of apoptosis and cell death in GBM spheroids as a function of spheroid size and TMZ-resistance status 48 hours after formation (Figure 3 C, D). Broadly, bobo-3 iodide positive (dead) GBM cells were observed in spheroids of all sizes for both 42MGBA-WT and42MGBA-TMZres cells (Figure 3 C, D)

To better assess whether spheroid size influenced the emergence of an apoptotic or necrotic GBM population, large quantities of spheroids or each size were generated using an AggreWell™ microwell plate process. These spheroids were pooled by size then dissociated for flow cytometry analysis of the cellular content (Figure S2). Dissociation with 0.5% trypsin showed no effects on the GBM cell viability (data not shown). Disaggregated cells were analyzed for expression of bobo-3 iodide (PE-TexasRedA) and CalceinAM (FITC-A), with cells

characterized as apoptotic (bobo-3+/CalceinAM+) vs. necrotic (bobo-3+/CalceinAM-). We observed few (<5%) overall necrotic cells and no significant difference in the fraction of necrotic cells in spheroids as a function of spheroid size or TMZ-resistance phenotype (Figure 4B). Interestingly, we observed significant effects of both spheroid size and TMZ-resistance on the emergence of apoptotic GBM cells (Figure 4A). We observed significantly higher fractions of apoptotic cells in 42MGBA-WT vs. 42MGBA-TMZres spheroids for almost all spheroid sizes with higher fractions of apoptotic cells in smaller ($\leq$ 5000 cells) than larger (10,000, 12,000 cells) spheroids regardless of TMZ-resistance status. For all subsequent studies we chose two spheroid condition (1,000 vs. 10,000 cells) that displayed different levels of proliferative capacity and apoptosis.

3.3 Single high dose and metronomic TMZ reduces GBM invasion for WT, but not TMZ-resistant spheroids

Spheroids initiated with 1,000 and 10,000 cells were encapsulated in GelMA hydrogel and subjected to treatment via TMZ. TMZ exposure was either a single exposure to 500 μM TMZ, replaced after 24 hours with complete medium containing equivalent concentration of DMSO, or metronomic dosing via 100 μM TMZ each day for 5 consecutive days, with all treatments compared to a DMSO control. Small (1,000 cells/spheroid) and large (10,000 cells/spheroid) 42MGBA-WT spheroids displayed significant invasion into the surrounding GelMA hydrogel, marked by increased in projected area with time. While trends emerged within 3 days of treatment, the effect of a single high dose or metronomic dosing on small 42MGBA-WT spheroids did not become significant until 5 days after treatment, with significantly reduced invasion in response to metronomic doing (vs. control) and a single high TMZ dose (vs. metronomic, control; Figure 5A, C). The effects of TMZ treatment on larger (10,000 cells/spheroid) GBM spheroids followed similar trends, though with a single high TMZ dose significantly reducing cell invasion as early as 1 day post treatment, and with metronomic dosing

or a single high dose both significantly reducing invasion by day 5 (Figure 5B, D).

We observed limited invasion of 42MGBA-TMZres GBM cells for both small and large spheroids. Smaller spheroids (initiated with 1,000 cells) did not show any changes in cell invasion in response to either a single high dose or metronomic dosing (Figure 6A, C). While both TMZ treatment groups significantly reduced projected surface area on day 3 and 5 for large spheroids (Figure 6B, D), there was no different between TMZ treatments and overall invasion was small compared to wild type GBM cells.

3.4. Difference in gene expression across spheroids of different sizes

We performed qRT-qPCR on gene targets related to cell death (*CASP3*, *TGF-β*), cellular growth (*PTEN*, *TIMP-1*), invasion (*MMP2*), and drug resistance (*CD44*, *HIF1α*) (Figure 7A). Wild type GBM cells in larger spheroids showed increased expression of *PTEN*, *STAT3*, *CASP3*, *TGF-β*, *TIMP-1* and *MMP2*, but reduced CD44 expression. Interestingly, a different expression pattern was observed for TMZ resistant cells, where larger spheroids demonstrated increased expression of *TIMP-1* and *CD44*, but reduced expression of *TGF-β*. Interestingly, quantitative analysis of *TIMP-1* (ΔCT normalized to actin expression) showed trends towards higher *TIMP-1* expression in larger vs. smaller spheroids for both wild type and TMZ resistant cells, but with TMZ resistant cells displaying significantly higher *TIMP-1* expression vs. wild type cells (Figure 7B).

3.5. Change in gene expression on Day 5 after TMZ treatments

We subsequently examined changes in gene expression of GBM spheroids of different sizes after 5 days of metronomic or single high TMZ treatment. In general, wild type cells showed an increased expression (vs. DMSO control) of *CASP3*, *TIMP-1*, *HIF1α*, and *TGF-β* in response to

either TMZ treatment (Figure 7C). *TIMP-1* expression was significantly higher in the wild type cell line compared to the TMZ resistance cell line after TMZ treatment (Figure 7D); while less pronounced, similar trends were also observed for increased *CASP3* expression in wildtype GBM cells after TMZ treatment (Figure 7E). Interestingly, small wild type spheroids showed higher expression of *PTEN*, *STAT3*, *MMP2*, *TGF-β* and *CD44* compared to the larger spheroids when treated with either metronomic or single high TMZ doses. Even though there was no statistical significance, several trends were observed in changes in gene expression for the TMZ resistant cells. Notably, metronomic dosing increased expression of *CASP3*, *PTEN*, and *STAT3* compared to a single high TMZ dose, while larger spheroids trended towards increased expression of *MMP2*, *TIMP-1*, *TGF-β*, and *CD44* in response to single high TMZ doses (Figure 7C, S2).

4. Discussion

Glioblastoma is marked by significant invasion into the surrounding brain parenchyma. The tumor margins are marked by significant transition in matrix and cellular environment as well as the presence of large and small cohorts of GBM cells (e.g., pseudopalisades) that are linked to progression, therapeutic resistance, and patient mortality. Cancer tissue engineering platforms offer the opportunity to dissect the role of multiple components of the tumor microenvironment on metrics of progression and therapeutic response. Many first generation studies have used GBM cells distributed within a hydrogel biomaterial, simulating low-density cancer cell populations in order to assess how matrix composition [50,62,63], stiffness [50,64-67], and structure [68] influence invasion and drug response. While important, the multicellular nature of GBM cohorts may also significantly influence cell activity. Yet while multicellular tumor spheroids are increasingly used in a wide range of cancer tissue engineering studies, spheroids sizes can differ drastically among studies [69,70]. Larger cohorts are often hypothesized to lead to greater

diffusional gradients for nutrients and oxygen that may contribute to the formation of necrotic core, regional changes in cell proliferative activity that may promote chemoresistance, and shifts in gene expression associated with a more aggressive phenotype [71,72]. The poroelastic nature of the brain also complicates surgical resection of GBM tumors; debulking requires a precise surgical margin be placed across a diffuse invasion front, resulting in the possibility of GBM cell cohorts of different scales may remain in the margin.

The goal of this study was to define the role of spheroid scale on metrics of GBM progression (invasion, drug response) traditionally used in cancer engineering models. The frontline therapy for GBM is the alkylating agent TMZ, which can induce apoptosis in dividing GBM cells [73]. However, the primary cytotoxic lesion can be repaired by a suicide DNA repair enzyme, methylguanine methyltransferase (MGMT) [74]. Because TMZ response can be tied to the presence of proliferative cells, tissue engineering models of TMZ response have often relied on supraphysiological TMZ doses. We recently validated a tissue engineering workflow, using an isogenically-matched pair of TMZ resistant and responsive cells with disparate MGMT expression levels generated by Tiek et al. [48], to assess TMZ response at physiological and metronomic dosing levels. While primarily for cells suspended in a GelMA hydrogel matrix, these studies confirmed that 42MGBA-TMZres cells showed reduced TMZ sensitivity in GelMA hydrogels. Our current study adds to this narrative, showing spheroid size is an important design characteristic that influence GBM cell behavior.

We created GBM spheroids ranging in size from <130 μm (wild type 100 cells/spheroid) to >600 μm (TMZ-resistant 12,000 cells/spheroid) in diameter using both 42MGBA-WT and 42MGBA-TMZres cells. While spheroid size increased with number of cells per spheroid and showed a high degree of circularity, spheroids created from TMZ-resistant cells were significantly larger, more compact, and trended towards a higher degree of circularity than wild type GBM cells.

While qualitative fluorescent images suggested that ki-67+ (proliferative) and bobo3+ (dead) cells were both more likely to be found towards the periphery of WT and TMZ-resistant spheroids, flow cytometry analysis of spheroids showed that a higher fraction of apoptotic cells were present in wild type (vs. TMZ-resistant) cells, particularly in smaller (<5000 cells/spheroid) spheroids. While previous work using mammary carcinoma cells [75] has suggested the formation of a necrotic core in tumor spheroids, we observed only a small fraction of necrotic cells. However, the increased fraction of apoptotic cells we observed in small GBM spheroids suggests that a relationship between spheroid size and apoptosis and necrosis may be more variable between cell types. There is an opportunity to explore extended time of spheroid culture to observe whether increased fractions of necrotic GBM cells can be observed. However, the increase in Ki-67 staining observed in GBM spheroids, particularly in larger spheroids containing TMZ-resistant GBM cells, was also consistent with increased 42MGBA-TMZres proliferation seen previously in 2D culture [48].

GBM invasion vs. proliferation (go vs. grow) plasticity suggests a process by which GBM cells can navigate complex selection pressures in the tumor margins [76,77]. Interestingly, we observed that while 42MGBA-TMZres proliferation was higher in spheroid culture, both large and small spheroids formed from 42MGBA-WT cells displayed significantly increased invasion into GelMA hydrogels. This finding was also consistent with our prior work that showed for 5000 cell spheroids 42MGBA-WT cells were significantly more invasive than TMZ-resistant cells [45]. Importantly, our work here extends the range over which this observation is consistent (1000 – 10,000 cells/spheroid) and suggesting an invasive phenotype is conserved for 42MGBA-WT cells across a wide range of biophysical environments. More interestingly, invasion of 42MGBA-WT cells was significantly reduced by both metronomic and a single high dose of TMZ, though the effect took 3 – 5 days to emerge. Interestingly, 42MGBA-TMZres$_r$ invasion was largely insensitive to TMZ treatment except for large (10,000 cells/spheroid) spheroid cultures.

Combined with prior work that showed 2D cultures of 42MGBA-TMZres cells (versus 42MGBA-WT) displayed reduced G2/M and increased G1 phase [48], but no change in metabolic activity in 3D GelMA culture [45], in response to 100 μm TMZ, these studies suggest that while 42MGBA-WT cells show significant reduction in invasion in response to TMZ, spatial patterns of cell proliferation vs. invasion may emerge in large 42MGBA-TMZres spheroids that are consistent with go-vs. grow plasticity. Our findings are separate from prior work with ovarian cancer cells that showed larger (5000 cells) spheroids were more resistant to taxol and cisplatin compared to smaller (500, 200, 100 or 50 cells) spheroids [78]. As larger spheroids have also been suggested to exhibited poor drug biotransport [71], our finding suggest the need for future study to more closely examine the spatial dependent shifts in GBM cell behavior and drug response.

Finally, our analysis of shifts in GBM gene expression suggested the largest changes were due to differences between TMZ-resistant and wild type cells, rather an as an effect of spheroid size. However, interestingly, TMZ-resistant (vs. wild type) cells displayed significantly upregulated expression of the MMP inhibitor TIMP-1. TIMP-1 has a multitude of functions, and has been implicated in suppressing cell proliferation and metabolic activity in mesenchymal stem cells [79], but increasing cell proliferation and invasion via the TIMP1/FAK/Akt pathway in colorectal cancer cells [80]. Here, we observed increased TIMP-1 expression in large (10,000 cells) vs. small (1,000 cells) spheroids for both wild type and TMZ-resistant lines, but increased TIMP-1 expression in TMZ-resistant vs. wild type spheroids. We also observed phenotypic shifts in expression of Caspase-3 (*CASP3*), a protease implicated in programmed cell death. *CASP3* expression was elevated in wild type spheroids after both metronomic and single high TMZ doses, especially in small spheroids. Interestingly, while we did not see a difference in metronomic vs. a single high TMZ dose on invasion in TMZ resistant cells, the combination of metronomic dosing and small spheroids induced the greatest increase in *CASP3* expression in TMZ resistant cells (Figure 7E). We observed a minor inverse effect of spheroid size on *MMP-2*

expression in 42MGBA-WT specimens as well as a mild increase with spheroid size for 42MGBA-TMZres specimens. These finding, while consistent broadly with prior studies using drug resistant (vs. wild type) glioblastoma cell lines but not considering spheroid size [48,81], now suggests that smaller GBM spheroids may display outsized responses to TMZ treatment in tissue engineering models.

5. Conclusions

We report the adaptation of a well-characterized GelMA hydrogel system to evaluate the combined effect of TMZ dosing strategy (metronomic vs. single high TMZ dose), TMZ-resistance status, and the size of GBM spheroids on metrics of GBM invasion and drug response. While spheroids embedded into a hydrogel matrix are increasingly common in studies of GBM cell behavior, an explicit consideration of the role of spheroid scale is essential to interpret these findings. We report that TMZ-resistant and wild type cells maintain discrete TMZ response phenotypes, with increased invasion but increased sensitivity to TMZ by wild type GBM specimens. Analysis of spatial pattern of cell proliferation as well as underlying gene expression data suggests that smaller GBM spheroids may exhibit increased fraction of apoptotic cells as well as a more robust response to TMZ therapeutics. These findings suggest that spheroid size should be rigorously reported and that future investigation must also consider the role of spheroid size on matrix remodeling (TIMP1) and apoptosis-associated (*CASP3*) proteases.

Acknowledgements

Funding sources include the National Cancer Institutes of the National Institutes of Health under Awards Number R01 CA256481 (BACH), R01 CA279195 (R.B.R.), and T32 CA009686 (fellowship funding for A.A-K.; T32 principal investigator (PI): Dr. Chunling Yi). The content is solely the responsibility of the authors and does not necessarily represent the official views of

the NIH. The authors are also grateful for additional funding provided by the Department of Chemical & Biomolecular Engineering and the Cancer Center at Illinois at the University of Illinois Urbana-Champaign. The authors also wish to acknowledge assistance from the Core Facilities at the Carl R. Woese Institute for Genomic Biology. and the Cytometry and Microscopy to Omics Facilities at the Roy J. Carver Biotechnology center. Specifically, Dr. Marcin Wozniak for training and FLOW assistance.

Contributions (CRediT: Contributor Roles Taxonomy[82,83])

S. Fok: Conceptualization, Data curation, Formal Analysis, Visualization, Investigation, Methodology, Writing – original draft, Writing – review & editing. **A. Appiah-Kubi**: Resources, Writing – review & editing. **R.B. Riggins:** Resources, Funding acquisition, Writing – review & editing. **B.A.C. Harley:** Conceptualization, Resources, Project administration, Funding acquisition, Supervision, Writing – review & editing.

Figures

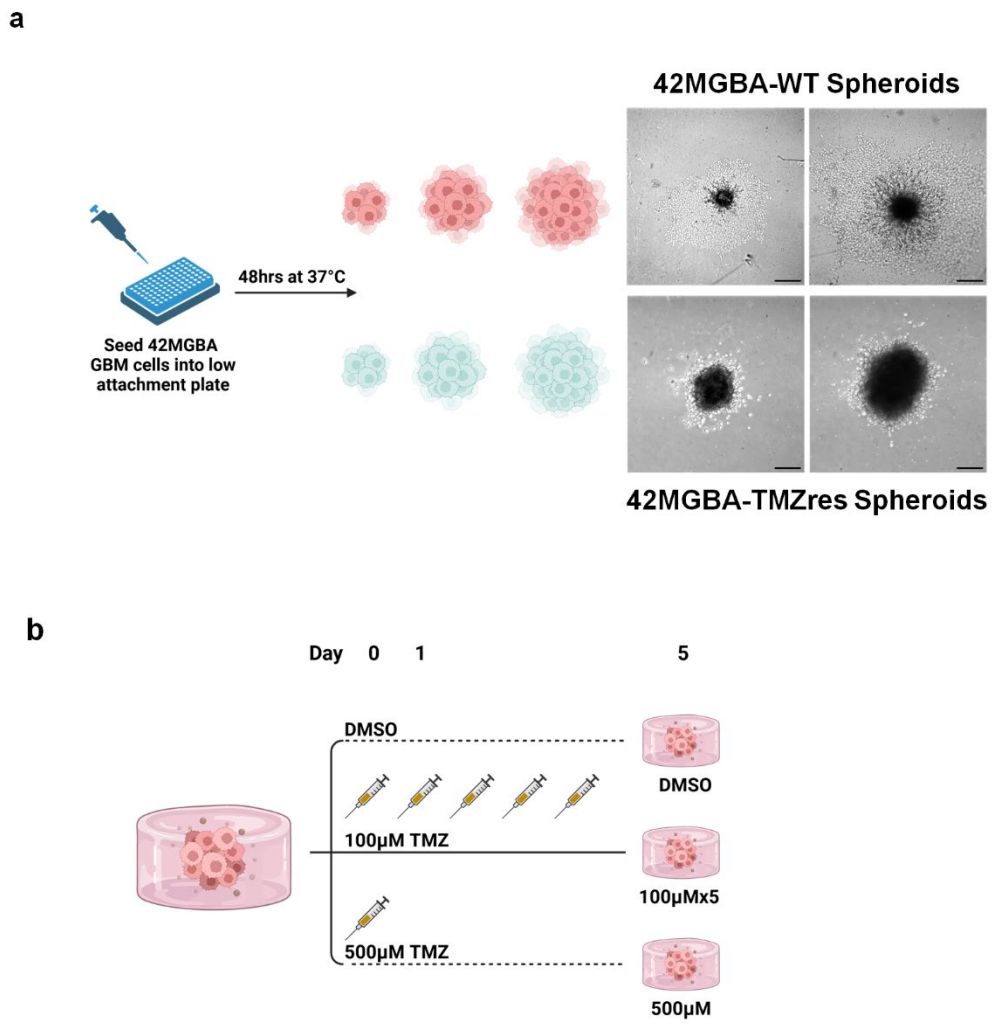

Figure 1. Conceptual illustration of experimental plan. (a) GBM spheroids were formed form isogenically-matched pairs of wild type (42MGBA-WT) or TMZ-resistant (42MGBA-TMZres) cells across a range of spheroid sizes (100 – 12,000 cells/spheroid). (b) Schematic depiction of the TMZ treatments. GBM spheroids encapsulated in GelMA hydrogel are subjected to a DMSO control, metronomic treatment (100μM per day for 5 days) via TMNZ, or single high TMZ dose (500μM at day 1). Cell phenotype, invasion, and gene expression patterns were traced for up to 5 days after initial TMZ exposure.

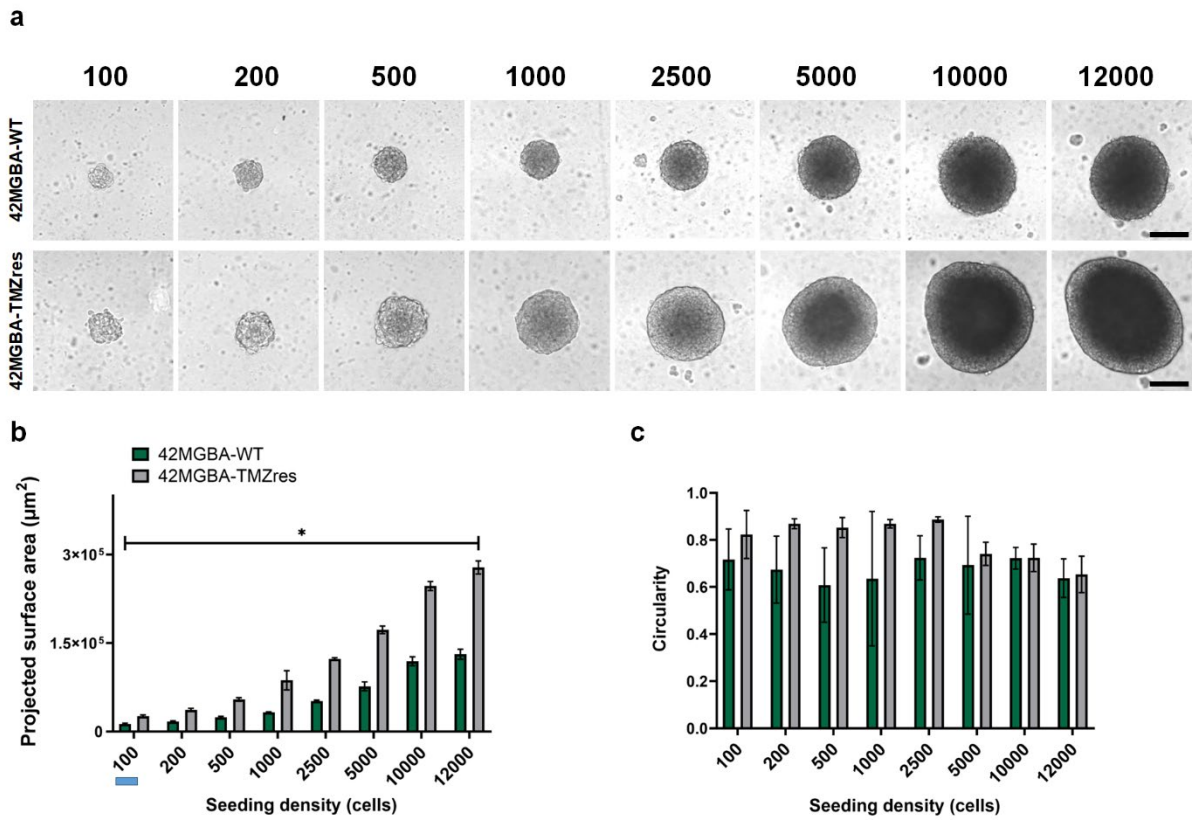

Figure 2. A library of GBM spheroids as a function of cell size (a) Bright field image of 42-MGBA spheroids initiated at different seeding densities different number of cells (n=8/group). (b) Projected surface area of GBM spheroids initiated as a function of size (n = 8/group). (c) Circularity of 42MGBA spheroids initiated at different seeding densities (n= 8/group). Statistical analysis was done by Kruskal Wallis test followed by Mann Whitney U Test. Statistical significance is represented as * $p < 0.05$. Scale bar: 200μm.

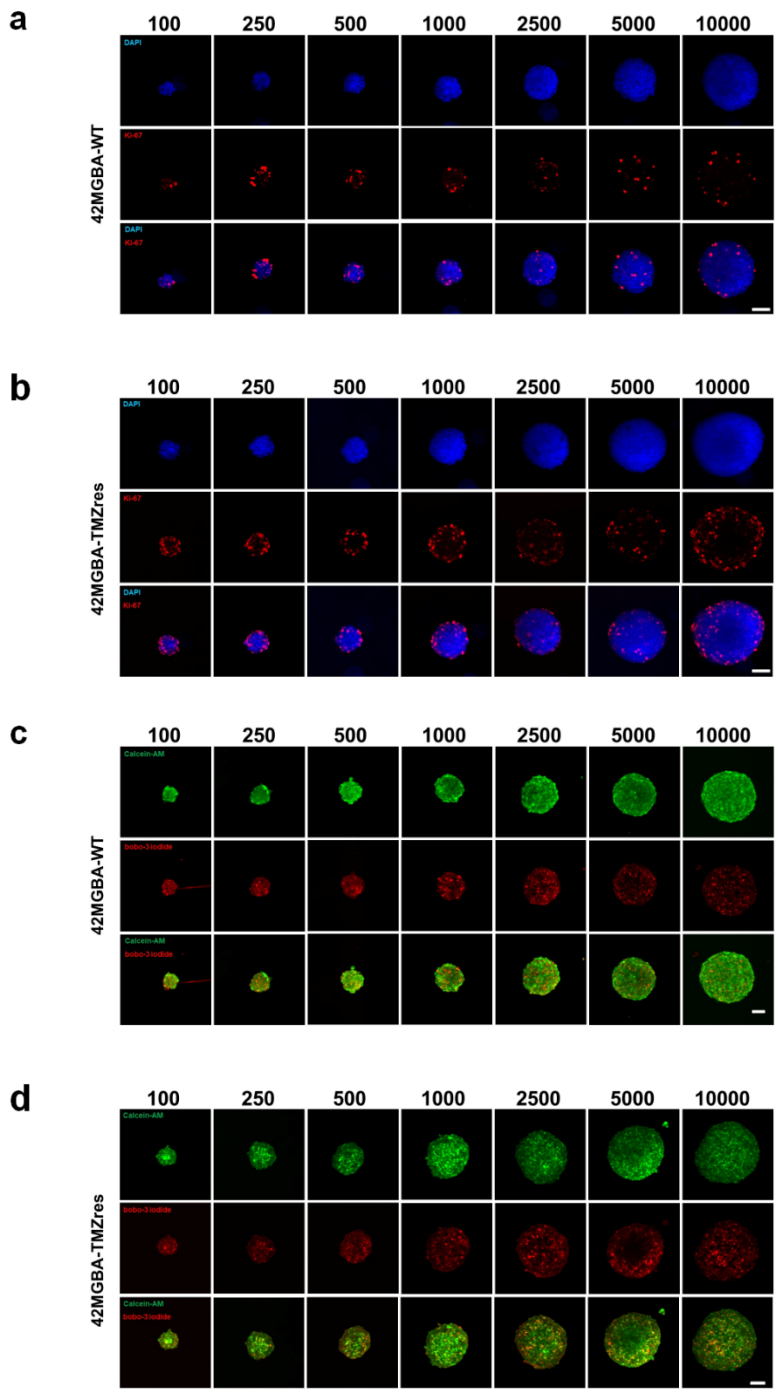

Figure3. Patterns of GBM cell proliferation and apoptosis. Representative fluorescent images of (a) wild type vs. (b) TMZ-resistant spheroids labeled with DAPI (nuclei) and Ki-67 (G1/S/G2/M phases of the cell cycle) as a function of spheroid size (n=8/group). Representative fluorescent images of (c) wild type vs. (d) TMZ-resistant spheroids labeled with Calcein-AM (live cells) and bobo-3 iodide (dead cells) as a function of spheroid size (n= 8/group). Scale bar: 200μm.

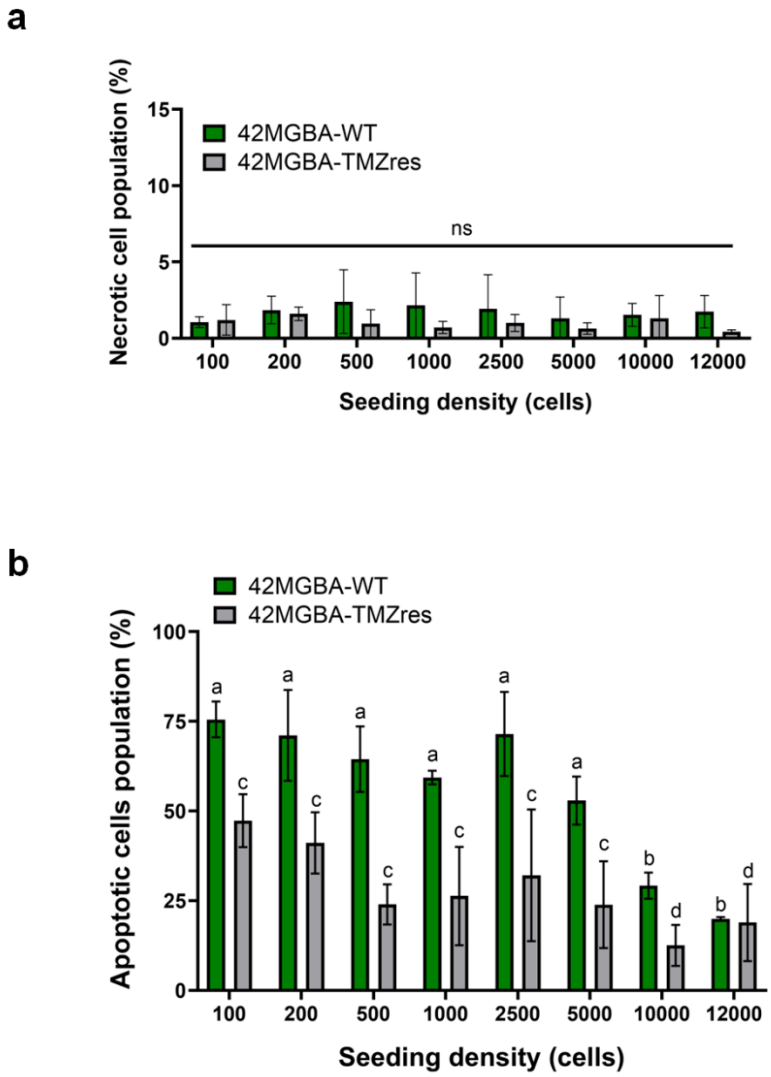

Figure 4. Percentage of apoptotic vs. necrotic GBM cells as function of spheroid size and TMZ-resistance. (a) The percentage of necrotic cells in spheroids as a function of spheroid size and TMZ resistance status (n=3). (b) The percentage of apoptotic cell population in spheroids as a function of spheroid size and TMZ resistance status (n=3). Statistical analysis was done by Kruskal Wallis test followed by Mann Whitney U Test. Different letters denoted significant level at $p < 0.05$.

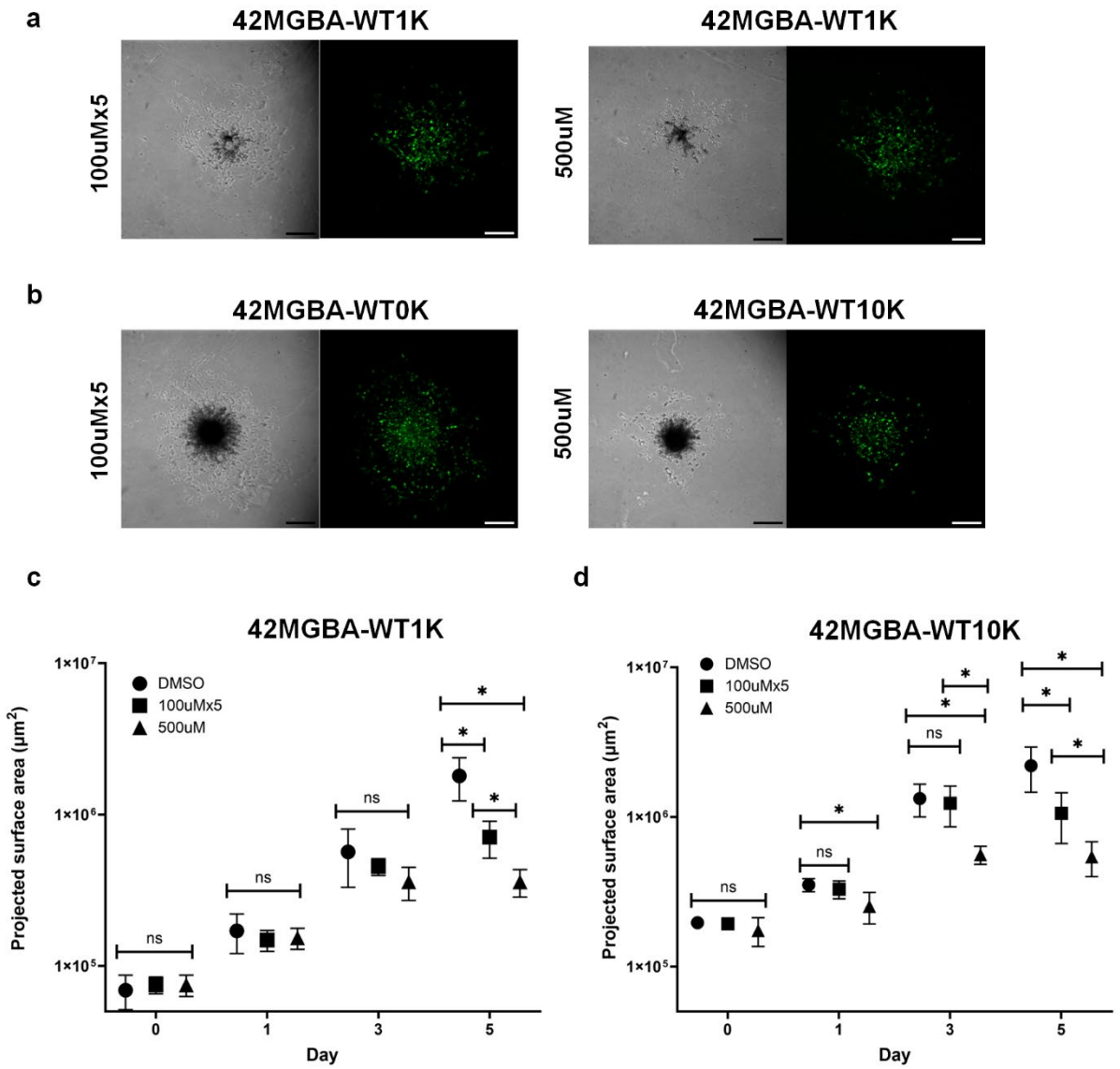

Figure 5. The effect of a single high TMZ dose or metronomic dosing on invasion of WT spheroids. (a) Representation image of small (1,000 cells) 42MGBA-WT spheroids treated with metronomic (left) vs. a single high TMZ dose (right). (b) Representation image of large (10,000 cells) 42MGBA-WT spheroids initiated with 10,000 cells treated with metronomic (left) vs. a single high TMZ dose (right). (c) Quantifying cell outgrowth for small (1,000 cell) spheroids over the course of 5 days after TMZ treatment (n = 3/group). (d) Quantifying cell outgrowth for large (10,000 cell) spheroids over the course of 5 days after TMZ treatment (n = 3/group). Statistical significance is represented as * $p < 0.05$.

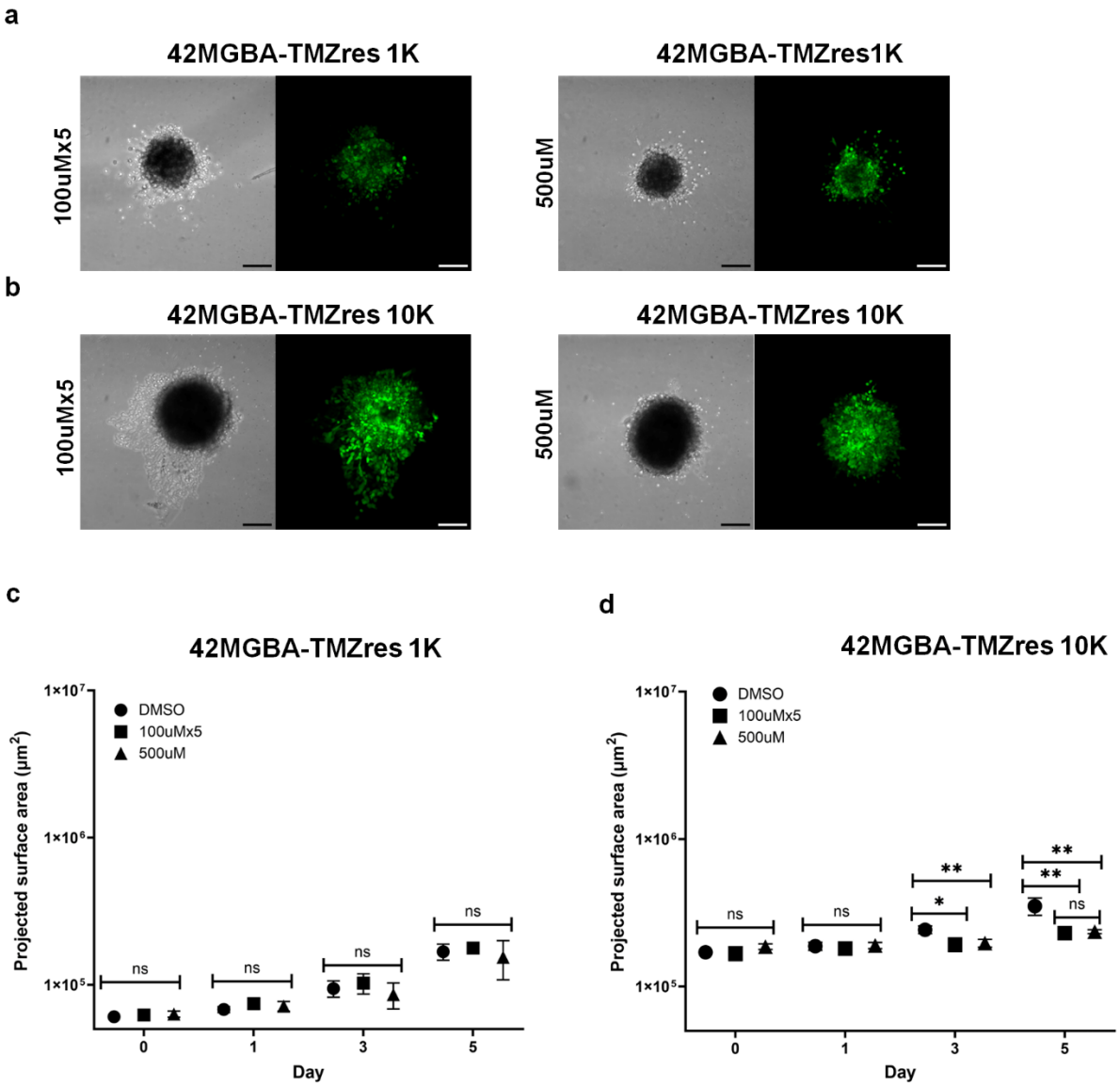

Figure 6. The effect of a single high TMZ dose or metronomic dosing on invasion of TMZ-resistant spheroids. (a) Representation image of small (1,000 cells) 42MGBA-TMZres spheroids treated with metronomic (left) vs. a single high TMZ dose (right). (b) Representation image of large (10,000 cells) 42MGBA-TMZres spheroids initiated with 10,000 cells treated with metronomic (left) vs. a single high TMZ dose (right). (c) Quantifying cell outgrowth for small (1,000 cell) TMZ-resistant spheroids over the course of 5 days after TMZ treatment (n = 3/group). (d) Quantifying cell outgrowth for large (10,000 cell) TMZ-resistant spheroids over the course of 5 days after TMZ treatment (n = 3/group). Statistical significance is represented as * $p < 0.05$.

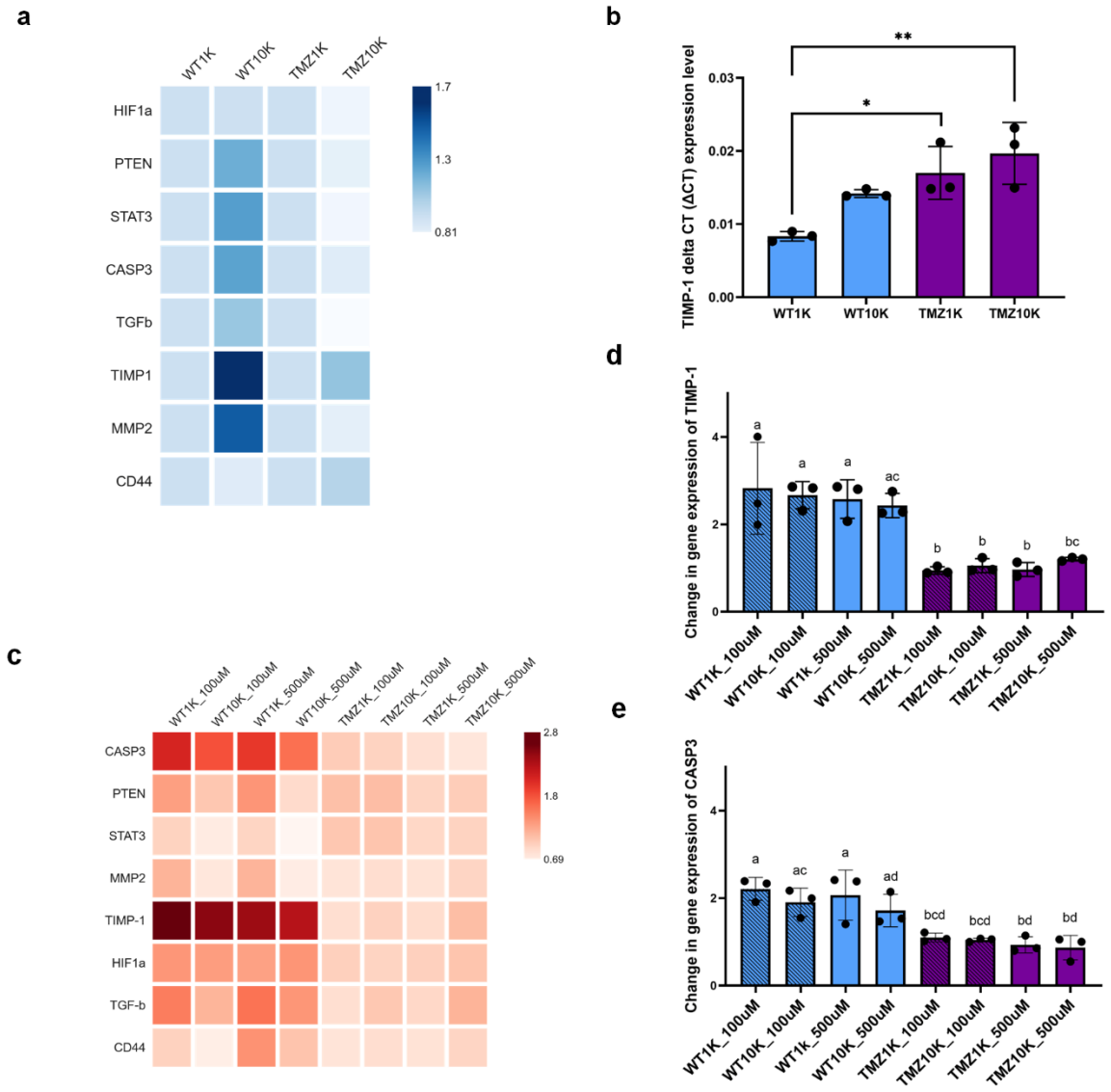

Figure 7. Shift in gene expression as a function of GBM spheroid size, TMZ treatment, and TMZ-resistance status. (a) Heatmap of relative gene expression of wild type vs. TMZ resistance spheroids of different sizes (1000 or 10000 cells) in conventional culture. (b) ΔCT value of *TIMP-1* of wildtype and TMZ-resistant spheroids of different sizes (1,000 vs. 10,000 cells). All data were normalized to ΔCT value of house-keeping gene Actin (n = 3/group). (c) Heatmap of relative gene expression of wild type vs. TMZ resistant spheroids of different sizes (1,000 vs. 10,000 cells) and as a function of TMZ treatment (single 500 μM dose vs. 5 days of 100 μM metronomic treatment). (d) *TIMP-1* expression for GBM spheroids as a function of size, TMZ resistance status and TMZ dose, relative to the DMSO control (n = 3/group). (e) *CASP3* gene expression for GBM spheroids as a function of size, TMZ resistance status and TMZ dose, relative to DMSO control (n = 3/group). Statistical analysis was completed by One- way ANOVA followed by Tukey HSD Test. Different letters denoted significance level at $p < 0.05$.

References

1 Amaral, R. L. F., Miranda, M., Marcato, P. D. & Swiech, K. Comparative Analysis of 3D Bladder Tumor Spheroids Obtained by Forced Floating and Hanging Drop Methods for Drug Screening. *Front Physiol* **8**, 605, doi:10.3389/fphys.2017.00605 (2017).

2 Wu, W. *et al.* Glioblastoma multiforme (GBM): An overview of current therapies and mechanisms of resistance. *Pharmacological Research* **171**, 105780, doi:https://doi.org/10.1016/j.phrs.2021.105780 (2021).

3 Mohammed, S., Dinesan, M. & Ajayakumar, T. Survival and quality of life analysis in glioblastoma multiforme with adjuvant chemoradiotherapy: a retrospective study. *Rep Pract Oncol Radiother* **27**, 1026-1036, doi:10.5603/RPOR.a2022.0113 (2022).

4 Mondragon-Soto, M. *et al.* Clinical, Therapeutic, and Prognostic Experience in Patients With Glioblastoma. *Cureus* **14**, e29856, doi:10.7759/cureus.29856 (2022).

5 Furnari, F. B. *et al.* Malignant astrocytic glioma: genetics, biology, and paths to treatment. *Genes Dev* **21**, 2683-2710, doi:10.1101/gad.1596707 (2007).

6 Johnson, D. R. & O'Neill, B. P. Glioblastoma survival in the United States before and during the temozolomide era. *J Neurooncol* **107**, 359-364, doi:10.1007/s11060-011-0749-4 (2012).

7 Charles, N. A., Holland, E. C., Gilbertson, R., Glass, R. & Kettenmann, H. The brain tumor microenvironment. *Glia* **59**, 1169-1180, doi:10.1002/glia.21136 (2011).

8 Jackson, C., Ruzevick, J., Phallen, J., Belcaid, Z. & Lim, M. Challenges in immunotherapy presented by the glioblastoma multiforme microenvironment. *Clin Dev Immunol* **2011**, 20, doi:10.1155/2011/732413 (2011).

9 Stupp, R. *et al.* Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med* **352**, 987-996, (2005).

10 Okada, M. *et al.* Tumor-associated macrophage/microglia infiltration in human gliomas is correlated with MCP-3, but not MCP-1. *Int J Oncol* **34**, 1621-1627, doi:10.3892/ijo_00000292 (2009).

11 Stupp, R. *et al.* Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *New England Journal of Medicine* **352**, 987-996, doi:10.1056/NEJMoa043330 (2005).

12 Paw, I., Carpenter, R. C., Watabe, K., Debinski, W. & Lo, H. W. Mechanisms regulating glioma invasion. *Cancer Lett* **362**, 1-7, doi:10.1016/j.canlet.2015.03.015 (2015).

13 Watkins, S. *et al.* Disruption of astrocyte-vascular coupling and the blood-brain barrier by invading glioma cells. *Nature communications* **5**, 4196, doi:10.1038/ncomms5196 (2014).

14 Velpula, K. K. *et al.* Glioma stem cell invasion through regulation of the interconnected ERK, integrin alpha6 and N-cadherin signaling pathway. *Cellular signalling* **24**, 2076-2084, doi:10.1016/j.cellsig.2012.07.002 (2012).

15 Ananthanarayanan, B., Kim, Y. & Kumar, S. Elucidating the mechanobiology of malignant brain tumors using a brain matrix-mimetic hyaluronic acid hydrogel platform. *Biomaterials* **32**, 7913-7923, doi:S0142-9612(11)00776-9 [pii] 10.1016/j.biomaterials.2011.07.005 (2011).

16 Bagley, S. J. *et al.* Glioblastoma Clinical Trials: Current Landscape and Opportunities for Improvement. *Clin Cancer Res* **28**, 594-602, doi:10.1158/1078-0432.Ccr-21-2750 (2022).

17 Clavreul, A. *et al.* Glioblastoma-associated stromal cells (GASCs) from histologically normal surgical margins have a myofibroblast phenotype and angiogenic properties. *J Pathol* **233**, 74-88, doi:10.1002/path.4332 (2014).

18 Seker-Polat, F., Pinarbasi Degirmenci, N., Solaroglu, I. & Bagci-Onder, T. Tumor Cell Infiltration into the Brain in Glioblastoma: From Mechanisms to Clinical Perspectives.

Cancers (Basel) **14**, doi:10.3390/cancers14020443 (2022).
19 Carlson, B. L., Pokorny, J. L., Schroeder, M. A. & Sarkaria, J. N. Establishment, maintenance and in vitro and in vivo applications of primary human glioblastoma multiforme (GBM) xenograft models for translational biology studies and drug discovery. *Current protocols in pharmacology / editorial board, S.J. Enna* **Chapter 14**, Unit 14 16, doi:10.1002/0471141755.ph1416s52 (2011).
20 Kitange, G. J. *et al.* Evaluation of MGMT promoter methylation status and correlation with temozolomide response in orthotopic glioblastoma xenograft model. *J Neurooncol* **92**, 23-31, doi:10.1007/s11060-008-9737-8 (2009).
21 Sarkaria, J. N. *et al.* Use of an orthotopic xenograft model for assessing the effect of epidermal growth factor receptor amplification on glioblastoma radiation response. *Clin Cancer Res* **12**, 2264-2271, doi:10.1158/1078-0432.CCR-05-2510 (2006).
22 Carlson, B. L., Pokorny, J. L., Schroeder, M. A. & Sarkaria, J. N. in *Current protocols in pharmacology / editorial board, S.J. Enna* (John Wiley & Sons, Inc., 2001).
23 Birgersdotter, A., Sandberg, R. & Ernberg, I. Gene expression perturbation in vitro--a growing case for three-dimensional (3D) culture systems. *Semin Cancer Biol* **15**, 405-412, doi:10.1016/j.semcancer.2005.06.009 (2005).
24 Li, C. *et al.* Cell type and culture condition-dependent alternative splicing in human breast cancer cells revealed by splicing-sensitive microarrays. *Cancer Res* **66**, 1990-1999, doi:10.1158/0008-5472.Can-05-2593 (2006).
25 Sood, A., Kumar, A., Dev, A., Gupta, V. K. & Han, S. S. Advances in Hydrogel-Based Microfluidic Blood-Brain-Barrier Models in Oncology Research. *Pharmaceutics* **14**, doi:10.3390/pharmaceutics14050993 (2022).
26 Xiao, W., Ehsanipour, A., Sohrabi, A. & Seidlits, S. K. Hyaluronic-Acid Based Hydrogels for 3-Dimensional Culture of Patient-Derived Glioblastoma Cells. *J Vis Exp*, doi:10.3791/58176 (2018).
27 Thakor, J. *et al.* Engineered Hydrogels for Brain Tumor Culture and Therapy. *Biodes Manuf* **3**, 203-226, doi:10.1007/s42242-020-00084-6 (2020).
28 Wolf, K. J., Chen, J., Coombes, J. D., Aghi, M. K. & Kumar, S. Dissecting and rebuilding the glioblastoma microenvironment with engineered materials. *Nature Reviews Materials*, doi:10.1038/s41578-019-0135-y (2019).
29 Rape, A., Ananthanarayanan, B. & Kumar, S. Engineering strategies to mimic the glioblastoma microenvironment. *Adv Drug Deliv Rev*, doi:10.1016/j.addr.2014.08.012 (2014).
30 Kim, Y. & Kumar, S. CD44-mediated adhesion to hyaluronic acid contributes to mechanosensing and invasive motility. *Mol Cancer Res* **12**, 1416-1429, doi:10.1158/1541-7786.MCR-13-0629 (2014).
31 Hammel, J. H., Cook, S. R., Belanger, M. C., Munson, J. M. & Pompano, R. R. Modeling Immunity In Vitro: Slices, Chips, and Engineered Tissues. *Annu Rev Biomed Eng* **23**, 461-491, doi:10.1146/annurev-bioeng-082420-124920 (2021).
32 Munson, J. M., Bellamkonda, R. V. & Swartz, M. A. Interstitial flow in a 3D microenvironment increases glioma invasion by a CXCR4-dependent mechanism. *Cancer Res* **73**, 1536-1546, doi:10.1158/0008-5472.CAN-12-2838 (2013).
33 Atay, N., Yuan, J., Cornelison, C. & Munson, J. TAMI-15. THE EFFECT OF INTERSTITIAL FLUID FLOW AND ASTROCYTES / MICROGLIA ON INVASION, PROLIFERATION AND STEMNESS OF PATIENT-DERIVED GLIOMA STEM CELLS. *Neuro-Oncology* **22**, ii216-ii216, doi:10.1093/neuonc/noaa215.904 (2020).
34 Neves, E. R. *et al.* Targeting Glioblastoma Tumor Hyaluronan to Enhance Therapeutic Interventions that Regulate Metabolic Cell Properties. *Advanced Therapeutics* **7**, 2400041, doi:https://doi.org/10.1002/adtp.202400041 (2024).
35 Pedron, S., Becka, E. & Harley, B. A. C. Spatially-gradated hydrogel platform as a three-

dimensional engineered tumor microenvironment. *Adv Mater* **27**, 1567-1572, doi:10.1002/adma.201404896 (2015).
36 Chen, J.-W., Leary, S., Barnhouse, V., Sarkaria, J. & Harley, B. A. C. Matrix hyaluronic acid and hypoxia influence a CD133+ subset of patient-derived glioblastoma cells. *Tissue Eng Part A* **28**, 330-340, doi:10.1089/ten.tea.2021.0117 (2022).
37 Perche, F. & Torchilin, V. P. Cancer cell spheroids as a model to evaluate chemotherapy protocols. *Cancer Biol Ther* **13**, 1205-1213, doi:10.4161/cbt.21353 (2012).
38 Martínez-González, A., Calvo, G., Pérez Romasanta, L. & Pérez-García, V. Hypoxic cell waves around necrotic cores in glioblastoma: a biomathematical model and Its therapeutic implications. *Bull Math Biol* **74**, 2875-2896, doi:10.1007/s11538-012-9786-1 (2012).
39 Rong, Y., Durden, D. L., Van Meir, E. G. & Brat, D. J. 'Pseudopalisading' necrosis in glioblastoma: a familiar morphologic feature that links vascular pathology, hypoxia, and angiogenesis. *J Neuropathol Exp Neurol* **65**, 529-539 (2006).
40 Brat, D. J. *et al.* Pseudopalisades in glioblastoma are hypoxic, express extracellular matrix proteases, and are formed by an actively migrating cell population. *Cancer Res* **64**, 920-927, doi:10.1158/0008-5472.can-03-2073 (2004).
41 Ngo, M. T. & Harley, B. A. C. The influence of hyaluronic acid and glioblastoma cell co-culture on the formation of endothelial cell networks in gelatin hydrogels. *Advanced healthcare materials* **6**, 1700687, doi:10.1002/adhm.201700687 (2017).
42 Ngo, M. T. & Harley, B. A. C. Perivascular signals alter global gene expression profile of glioblastoma and response to temozolomide in a gelatin hydrogel. *Biomaterials* **198**, 122-134, doi:https://doi.org/10.1016/j.biomaterials.2018.06.013 (2019).
43 Ngo, M. T., Karvelis, E. & Harley, B. A. C. Multidimensional hydrogel models reveal endothelial network angiocrine signals increase glioblastoma cell number, invasion, and temozolomide resistance. *Integr Biol (Camb)* **12**, 139-149, doi:10.1093/intbio/zyaa010 (2020).
44 Ngo, M. T., Sarkaria, J. N. & Harley, B. A. C. Perivascular Stromal Cells Instruct Glioblastoma Invasion, Proliferation, and Therapeutic Response within an Engineered Brain Perivascular Niche Model. *Advanced Science* **9**, 2201888, doi:https://doi.org/10.1002/advs.202201888 (2022).
45 Kriuchkovskaia, V. A., Eames, E. K., Riggins, R. B. & Harley, B. A. C. Acquired Temozolomide Resistance Instructs Patterns of Glioblastoma Behavior in Gelatin Hydrogels. *Advanced healthcare materials*, e2400779, doi:10.1002/adhm.202400779 (2024).
46 Kong, D. S. *et al.* Phase II trial of low-dose continuous (metronomic) treatment of temozolomide for recurrent glioblastoma. *Neuro Oncol* **12**, 289-296, doi:10.1093/neuonc/nop030 (2010).
47 Bosio, A. *et al.* Metronomic Temozolomide in Heavily Pretreated Patients With Recurrent Isocitrate Dehydrogenase Wild-type Glioblastoma: A Large Real-Life Mono-Institutional Study. *Clinical Oncology* **35**, e319-e327, doi:https://doi.org/10.1016/j.clon.2023.01.012 (2023).
48 Tiek, D. M. *et al.* Alterations in Cell Motility, Proliferation, and Metabolism in Novel Models of Acquired Temozolomide Resistant Glioblastoma. *Sci Rep* **8**, 7222, doi:10.1038/s41598-018-25588-1 (2018).
49 Kim, J. T. *et al.* Metronomic treatment of temozolomide inhibits tumor cell growth through reduction of angiogenesis and augmentation of apoptosis in orthotopic models of gliomas. *Oncol Rep* **16**, 33-39 (2006).
50 Chen, J.-W., Pedron, S. & Harley, B. A. C. The combined influence of hydrogel stiffness and matrix-bound hyaluronic acid content on glioblastoma invasion. *Macromol Biosci* **17**, 1700018, doi:10.1002/mabi.201700018 (2017).

51 Chen, J.-W., Blazek, A., Lumibao, J., Gaskins, H. R. & Harley, B. A. C. Hypoxia activates enhanced invasive potential and endogenous hyaluronic acid production by glioblastoma cells. *Biomater Sci* **6**, 854-862, doi:10.1039/C7BM01195D (2018).
52 Chen, J.-W. *et al.* Influence of hyaluronic acid transitions in tumor microenvironment on glioblastoma malignancy and invasive behavior. *Front Mater* (2018).
53 Chen, J.-W. E. *et al.* Crosstalk between microglia and patient-derived glioblastoma cells inhibit invasion in a three-dimensional gelatin hydrogel model. *Journal of neuroinflammation* **17**, 346, doi:10.1186/s12974-020-02026-6 (2020).
54 Kaylan, K. B. *et al.* Spatial patterning of liver progenitor cell differentiation mediated by cellular contractility and Notch signaling. *Elife* **7**, doi:10.7554/eLife.38536 (2018).
55 Underhill, G. H. & Khetani, S. R. Bioengineered Liver Models for Drug Testing and Cell Differentiation Studies. *Cell Mol Gastroenterol Hepatol* **5**, 426-439 e421, doi:10.1016/j.jcmgh.2017.11.012 (2018).
56 Kaylan, K. B., Ermilova, V., Yada, R. C. & Underhill, G. H. Combinatorial microenvironmental regulation of liver progenitor differentiation by Notch ligands, TGFbeta, and extracellular matrix. *Sci Rep* **6**, 23490, doi:10.1038/srep23490 (2016).
57 Lee, J., Abdeen, A. A., Wycislo, K. L., Fan, T. M. & Kilian, K. A. Interfacial geometry dictates cancer cell tumorigenicity. *Nat Mater* **15**, 856-862, doi:10.1038/nmat4610 (2016).
58 Bruns, J., Egan, T., Mercier, P. & Zustiak, S. P. Glioblastoma spheroid growth and chemotherapeutic responses in single and dual-stiffness hydrogels. *Acta Biomater* **163**, 400-414, doi:10.1016/j.actbio.2022.05.048 (2023).
59 Raghavan, S. *et al.* Comparative analysis of tumor spheroid generation techniques for differential in vitro drug toxicity. *Oncotarget* **7**, 16948-16961, doi:10.18632/oncotarget.7659 (2016).
60 Chen, J. E., Pedron, S. & Harley, B. A. C. The Combined Influence of Hydrogel Stiffness and Matrix-Bound Hyaluronic Acid Content on Glioblastoma Invasion. *Macromol Biosci* **17**, doi:10.1002/mabi.201700018 (2017).
61 Mahadik, B. P., Pedron Haba, S., Skertich, L. J. & Harley, B. A. The use of covalently immobilized stem cell factor to selectively affect hematopoietic stem cell activity within a gelatin hydrogel. *Biomaterials* **67**, 297-307, doi:10.1016/j.biomaterials.2015.07.042 (2015).
62 Pedron, S., Hanselman, J. S., Schroeder, M. A., Sarkaria, J. N. & Harley, B. A. C. Extracellular hyaluronic acid influences the efficacy of EGFR tyrosine kinase inhibitors in a biomaterial model of glioblastoma. *Advanced healthcare materials* **6**, 1700529, doi:10.1002/adhm.201700529 (2017).
63 Pedron, S., Becka, E. & Harley, B. A. Regulation of glioma cell phenotype in 3D matrices by hyaluronic acid. *Biomaterials* **34**, 7408-7417, doi:10.1016/j.biomaterials.2013.06.024 (2013).
64 Heffernan, J. M., Overstreet, D. J., Le, L. D., Vernon, B. L. & Sirianni, R. W. Bioengineered scaffolds for 3D analysis of glioblastoma proliferation and invasion. *Ann Biomed Eng* **43**, 1965-1977, doi:10.1007/s10439-014-1223-1 (2015).
65 Wong, S. Y. *et al.* Constitutive activation of myosin-dependent contractility sensitizes glioma tumor-initiating cells to mechanical inputs and reduces tissue invasion. *Cancer Res* **75**, 1113-1122, doi:10.1158/0008-5472.can-13-3426 (2015).
66 Rape, A. D. & Kumar, S. A composite hydrogel platform for the dissection of tumor cell migration at tissue interfaces. *Biomaterials* **35**, 8846-8853, doi:10.1016/j.biomaterials.2014.07.003 (2014).
67 Umesh, V., Rape, A. D., Ulrich, T. A. & Kumar, S. Microenvironmental stiffness enhances glioma cell proliferation by stimulating epidermal growth factor receptor signaling. *PloS one* **9**, e101771, doi:10.1371/journal.pone.0101771 (2014).

68 Neves, E. R., Harley, B. A. C. & Pedron, S. Microphysiological systems to study tumor-stroma interactions in brain cancer. *Brain Research Bulletin* **174**, 220-229, doi:https://doi.org/10.1016/j.brainresbull.2021.06.012 (2021).
69 Ho, W. Y., Yeap, S. K., Ho, C. L., Rahim, R. A. & Alitheen, N. B. Development of multicellular tumor spheroid (MCTS) culture from breast cancer cell and a high throughput screening method using the MTT assay. *PLoS One* **7**, e44640, doi:10.1371/journal.pone.0044640 (2012).
70 Ivanov, D. P. & Grabowska, A. M. Spheroid arrays for high-throughput single-cell analysis of spatial patterns and biomarker expression in 3D. *Sci Rep* **7**, 41160, doi:10.1038/srep41160 (2017).
71 Singh, S. K. *et al.* Critical Role of Three-Dimensional Tumorsphere Size on Experimental Outcome. *BioTechniques* **69**, 333-338, doi:10.2144/btn-2020-0081 (2020).
72 Li, X. F. *et al.* Visualization of hypoxia in microscopic tumors by immunofluorescent microscopy. *Cancer Res* **67**, 7646-7653, doi:10.1158/0008-5472.Can-06-4353 (2007).
73 Svec, R. L. *et al.* Tunable Stability of Imidazotetrazines Leads to a Potent Compound for Glioblastoma. *ACS Chem Biol* **13**, 3206-3216, doi:10.1021/acschembio.8b00864 (2018).
74 Svec, R. L. *et al.* Novel Imidazotetrazine Evades Known Resistance Mechanisms and Is Effective against Temozolomide-Resistant Brain Cancer in Cell Culture. *ACS Chemical Biology* **17**, 299-313, doi:10.1021/acschembio.2c00022 (2022).
75 Cheng, G., Tse, J., Jain, R. K. & Munn, L. L. Micro-Environmental Mechanical Stress Controls Tumor Spheroid Size and Morphology by Suppressing Proliferation and Inducing Apoptosis in Cancer Cells. *PloS one* **4**, e4632, doi:10.1371/journal.pone.0004632 (2009).
76 Castellan, M. *et al.* Single-cell analyses reveal YAP/TAZ as regulators of stemness and cell plasticity in glioblastoma. *Nature Cancer* **2**, 174-188, doi:10.1038/s43018-020-00150-z (2021).
77 Ruiz-Ontanon, P. *et al.* Cellular plasticity confers migratory and invasive advantages to a population of glioblastoma-initiating cells that infiltrate peritumoral tissue. *Stem Cells* **31**, 1075-1085, doi:10.1002/stem.1349 (2013).
78 Gunay, G. *et al.* The effects of size and shape of the ovarian cancer spheroids on the drug resistance and migration. *Gynecol Oncol* **159**, 563-572, doi:10.1016/j.ygyno.2020.09.002 (2020).
79 Egea, V. *et al.* Tissue inhibitor of metalloproteinase-1 (TIMP-1) regulates mesenchymal stem cells through let-7f microRNA and Wnt/β-catenin signaling. *Proceedings of the National Academy of Sciences* **109**, E309-E316, doi:doi:10.1073/pnas.1115083109 (2012).
80 Ma, B. *et al.* TIMP1 promotes cell proliferation and invasion capability of right-sided colon cancers via the FAK/Akt signaling pathway. *Cancer Sci* **113**, 4244-4257, doi:10.1111/cas.15567 (2022).
81 Nagane, M., Levitzki, A., Gazit, A., Cavenee, W. K. & Huang, H.-J. S. Drug resistance of human glioblastoma cells conferred by a tumor-specific mutant epidermal growth factor receptor through modulation of Bcl-X_L and caspase-3-like proteases. *Proceedings of the National Academy of Sciences* **95**, 5724-5729, doi:doi:10.1073/pnas.95.10.5724 (1998).
82 Brand, A., Allen, L., Altman, M., Hlava, M. & Scott, J. Beyond authorship: attribution, contribution, collaboration, and credit. *Learned Publishing* **28**, 151-155, doi:https://doi.org/10.1087/20150211 (2015).
83 Allen, L., Scott, J., Brand, A., Hlava, M. & Altman, M. Publishing: Credit where credit is due. *Nature* **508**, 312-313, doi:10.1038/508312a (2014).