1 **Long-fuse evolution of carnivoran skeletal phenomes through the Cenozoic**
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Keywords: Carnivora; ecomorphology; macroevolution; Ornstein-Uhlenbeck modeling;

phylogenetic comparative methods

Abstract

Climatic change is hypothesized to promote phenotypic diversification. While neontological

analyses are often used to test this hypothesis, extant data only captures time-averaged signals of

surviving lineages. More nuanced tests require paired and longitudinal climatic and organismal

data. Here, we developed the most comprehensive phenomic dataset to-date of pan-carnivorans

to test hypotheses that Cenozoic climatic change influenced the evolution of the cranial,

appendicular, and axial skeleton. We found support for the hypothesis that a hierarchical

progression of ecological diversification across the Cenozoic significantly influenced the

establishment of modern carnivorans. Specifically, extinctions during the Eocene-Oligocene

Transition released crown carnivorans from a constrained adaptive zone to interfamilial skeletal

diversification. Intrafamilial skeletal diversification did not occur for another 20 million years

until after the Mid-Miocene Climate Transition. Our work demonstrates the essential role of

macroevolutionary data from the fossil record for revealing how major global climatic events

steered the evolutionary trajectories of modern skeletal phenomes.

Introduction

Ecological opportunities arising from climatic and environmental changes are often

hypothesized to promote phenotypic diversification across macroevolutionary time scales [1–3].

After the Early Eocene Climate Optimum, trends towards decreased global temperatures [4] and

consequent environmental transitions from closed forest habitats to open grassland habitats

[5,6] have facilitated the increase in phenotypic diversity and evolution of novel phenotypes in

several mammalian clades. For example, rodents, lagomorphs, and ungulates all independently

evolved hypsodont dentition along with the lengthening of limbs for more efficient cursoriality

in association with open habitats [7–10]. Similarly, phenotypic diversity in carnivores increased

[11–13], enabling them to exploit diverse habitats through a range of hunting behaviors from

ambush to cursorial or subterranean pursuit specialization [14–16]. However, the question of

how these environmental changes influence the rate at which phenotypic disparity accumulates

(i.e., tempo) and the pattern of phenotypic change across the phylogeny (i.e., mode) remains

difficult to resolve due to the absence of time-sequence data (i.e., paleontological data)

considered under a phylogenetic framework. The few studies that have successfully integrated

the fossil record into neontological analyses demonstrated that fossils are essential for accurate

model selection in phenotypic macroevolution analyses [17–21], thus providing a more informed

understanding of phenotypic variation through phylogenetic time (e.g., [15,19,22–25]). To-date,

these studies have primarily focused only on single phenotypic traits or structures like body size

or the skull. No study has yet to simultaneously test how major environmental shifts may have

facilitated the evolution of the skull, appendicular skeleton, and axial skeleton in both extant and

extinct species.

In this study, we take advantage of the rich fossil record of pan-carnivorans (i.e.,

Hyaenodonta; Oxyaenodonta; and Carnivoramorpha, which includes Carnivora) to test

hypotheses that climatic and environmental changes during the past 66 million years influenced

the tempo and mode of the cranial, appendicular, and axial skeletal systems. Pan-carnivorans

represent one of the best characterized crown mammal groups from the Cenozoic, with a crown-

group fossil record going back more than 55 million years and stem taxa being traced back to the

early Paleocene. With the extinction of non-avian dinosaurs, early pan-carnivorans became the

first large carnivorous mammals [26]. Although crown carnivoran lineages co-existed with their

now-extinct sister clades during much of the Eocene, most early carnivorans were small,

generalized predators serving as secondary consumers. It was not until later in the Oligocene and

Miocene that they expanded to large, specialized top predator niches [11–13,27,28]. Since their

first appearance as small mongoose-like forms [11,13,17], carnivorans have diversified into the

numerous body forms we see today, ranging from small, elongate weasels to big robust bears to

fully aquatic, streamlined seals [29]. Here, we address the as-yet unanswered question of what

drove the exceptional diversification of carnivoran body plans from a single blueprint.

The two leading hypotheses for the major influences on carnivoran morphological

diversification are ecological opportunity facilitated by climate change and the extinction of

coeval stem pan-carnivorans. Global cooling, increased aridity, and habitat shift from forest to

grasslands have led to several rises and falls of carnivoran and stem pan-carnivoran clades [11].

Carnivorans survived during these environmental and climatic changes, as dental records suggest

that crown carnivorans maintained their morphological diversity even during periods of

extinction whereas morphological diversity in stem groups continued to decline [12].

Furthermore, stem pan-carnivorans exhibited steeper declines in species richness than

carnivorans, and the overall percentage of stem pan-carnivoran taxa declined whereas those of

carnivoran taxa increased [11–13]. These patterns have been attributed to competition between

carnivorans and stem pan-carnivorans in which declining stem pan-carnivorans are replaced and

excluded by phylogenetically distinct but functionally similar crown carnivorans [11–13,27].

Although analyses based on dental traits have partially supported this hypothesis [11,12], no

quantitative evidence has been explicitly tested for this hypothesized link between

paleoenvironmental changes and the diversification of the remaining skeletal phenome.

Here, we tested if increased carnivoran skeletal diversity was facilitated by rapid climate

change, character displacement from stem ancestors, or independently from these factors. We

predicted that the Eocene Oligocene Transition (EOT) and Mid-Miocene Climate Transition

(MMCT) facilitated skeletal diversification within carnivorans, enabling species to exploit new

resources during these periods of novel ecological opportunity. We leveraged the vast pan-

carnivoran fossil record to quantify skeletal phenomes by incorporating traits from the skull,

appendicular skeleton, and axial skeleton of extinct and extant species. We then used a

phylogenetic comparative framework to examine the tempo and mode of skeletal evolution

utilizing a total evidence phylogeny that contains almost all known extant and extinct

carnivorans and their stem relatives [21].

Results

Skeletal trait imputation

We quantified the skeletal phenome of 118 extant species and 81 extinct species using

103 linear measurements obtained from 854 specimens held at 17 natural history museums

(Table S1). Together, this 192 species dataset includes seven traits in the cranium; seven traits in

the mandible; 13 traits in the forelimb; 13 traits in the hindlimb; and seven traits each in third

cervical, fifth cervical, first thoracic, middle thoracic, diaphragmatic thoracic, last thoracic, first

lumbar, middle lumbar, and last lumbar vertebrae (Fig. S1). We imputed missing trait

measurements (25.2% of trait measurements in the full dataset; 0% in extant dataset; 61.9% in

extinct dataset) using multiple imputation with PCA (MIPCA) under a Bayesian framework

[30,31]. Our assessment of the quality of the trait imputations revealed one-to-one relationships

(i.e., slopes are not significantly different from one) between imputed and empirical values of

each trait regardless of the amount of missing data (Fig. S2A), indicating that our phenome

dataset can reliably estimate missing trait data. Comparisons between log shape ratios calculated

from imputed traits and empirical traits also show one-to-one relationships in most situations

with 10–60% of randomly removed trait measurements (Fig. S2B).

We performed a principal component analysis (PCA) incorporating all trait

measurements with size removed. Seven PC axes were found meaningful following procedures

by [32]; these were subsequently used as our proxy of the skeletal phenome (76.3% of explained

variance; Fig. 1; see Table S2 for trait loadings). The geometric mean of all trait measurements

was used as our proxy for skeletal size. We similarly performed PCAs to reduce the dimensions

of each skeletal component (i.e., cranium, mandible, forelimb, hindlimb, and each of the nine

vertebrae).

Evolution of carnivoran skeletal phenome and size

Phylogenetic signal in carnivoran skeletal phenome was moderate (Pagel's $\lambda = 0.60$, p =

0.001; Blomberg's $K = 0.58$, p = 0.001), suggesting that disparity in skeletal phenome primarily

occurs within clades as opposed to among clades. Although disparity through time (DTT)

analyses indicated that subclade disparity in skeletal phenome does not differ from what is

expected under Brownian motion (BM; morphological disparity index [MDI] = 0.018, P =

0.632), further inspection revealed higher subclade disparity than expected under BM during two

time intervals. The first occurs from ~39 to ~37 Ma (million years ago; MDI = 0.534, P = 0.012),

right between the Middle Eocene Climatic Optimum 40 Ma and the Eocene-Oligocene

Transition 34 Ma (Fig. 2A,B). The second occurs from ~14 to ~1 Ma, coinciding with the

conclusion of the Middle Miocene Climatic Optimum and the Mid-Miocene Climate Transition

~14 Ma (MDI = 0.238, P = 0.018) (Fig. 2C).

We tested 16 alternative hypotheses that could explain the disparity of carnivoran skeletal

phenome by fitting a series of macroevolutionary models (Fig. 3). We found nearly equal support

for the two release and radiate models (Table 1), where the evolution of skeletal phenomes

transitions from an Ornstein–Uhlenbeck (OU) process to BM at the EOT 34 Ma (OUBMi$_{EOT}$:

AICcW = 0.66) and at the MMCT 14 Ma (OUBMi$_{MMCT}$: ΔAICc = 1.33, AICcW = 0.34). These

models indicated that evolutionary rates slowed after both the EOT (mean $\sigma^2_{pre-EOT}$ = 0.148;

mean $\sigma^2_{post-EOT}$ = 0.013) and the MMCT (mean $\sigma^2_{pre-MMCT}$ = 0.048; mean $\sigma^2_{post-MMCT}$ = 0.013).

Mean stationary variances ($\sigma^2/2\alpha$) for the OU portion of the OUBMi$_{EOT}$ and OUBMi$_{MMCT}$ models

were 0.148 (α = 0.50) and 0.285 (α = 0.08), respectively. In contrast, models with distinct optima

among pan-carnivoran groups (i.e., sOU and cOU models) were poorer fits (ΔAICc = 89.80–

107.95), even compared to single regime models based on BM or OU. Simulations under the

parameters of the OUBMi$_{EOT}$ model indicated that there was substantial power to distinguish this

model from BM and alterative models (AICcW > 0.99; Table S3; Fig. 4A). Simulations under

the parameters of the OUBMi$_{MMCT}$ model indicated that there was substantial power to

distinguish this model from all alterative models (ΔAICc = 1.96; AICcW = 0.27; Table S3; Fig.

4B) except the BMBM$_{MMCT}$ (AICcW = 0.73; Table S3). These simulations thus suggest that the

EOT rather than the MMCT may have a stronger effect on shifts of evolutionary mode of

skeletal phenome.

In contrast, high phylogenetic signal (Pagel's λ = 0.91, p = 0.001; Blomberg's K = 0.56,

p = 0.001) and DTT analyses were united in showing that disparity in skeletal size did not

significantly deviate from BM and tends to occur within clades (Fig. 2D–F). The OUBMi$_{MMCT}$

model was the best supported model (AICcW = 0.58) for skeletal size, suggesting that the

evolution of skeletal size transitions from an OU process (α = 0.05; $\sigma^2_{pre-MMCT}$ = 0.019) to BM

($\sigma^2_{post-MMCT}$ = 0.008) during the MMCT (Table 1). However, simulations could not distinguish

this best supported model from OU1 (AICcW = 0.21), cOU3 (AICcW = 0.16), OUOU$_{EOT}$

(AICcW 0.14), sOU2 (AICcW = 0.09) and the OUOU$_{MMCT}$ (AICcW = 0.09) models (Table S3).

Thus, climatic changes may have had less of an impact on the evolution of skeletal size than

suggested by macroevolutionary models.

Evolution of carnivoran skeletal components

Phylogenetic signal of the skeletal components ranged from moderate to high (Pagel's λ

= 0.44–0.94, all p = 0.001; Blomberg's K = 0.38–0.45, all p = 0.001; Table S4), suggesting that

variance in skeletal components tended to follow BM and primarily occurred within clades as

opposed to among clades. DTT analyses revealed three general patterns in the evolution of these

skeletal components (Fig. S3). First, all vertebrae largely followed BM expectations except

during last 10–20 million years (My). Second, the cranium and limbs exhibited elevated subclade

disparity compared to BM during the Middle Eocene Climatic Optimum 40 Ma and from the

Early Miocene to present. Third, the mandible exhibited elevated subclade disparity throughout

the majority of pan-carnivoran history.

Climate models with shifts in evolutionary rate, mode, and/or optima were the best fitting

models for all skeletal components (Table S5; see Table S6 for parameter estimates from best

model). The OUBMi$_{MMCT}$ model was best supported for the hindlimb and most vertebrae

(AICcW = 0.51–1.00; Table S5). Simulations under the parameters of this model indicated that

there was substantial power to distinguish from alterative models for these components except

for the last thoracic and first and middle lumbar vertebrae (Table S7). For the cranium, the

BMBM$_{EOT}$ (AICcW = 0.37), OUBMi$_{MMCT}$ (ΔAICc = 0.59, AICcW = 0.28), and OUBM$_{EOT}$

(ΔAICc = 1.43, AICcW = 0.18) models were equally supported; however, simulations could not

distinguish these models from BM (Table S7). The OUOU$_{EOT}$ (AICcW = 0.54) and

OUBMi$_{MMCT}$ (ΔAICc = 1.32; AICcW = 0.28) models were equally supported for the mandible,

and simulations indicated there was substantial power to distinguish them from BM and

alterative models (Table S7). The OUOU$_{MMCT}$ model was best supported for the forelimb

(AICcW = 0.84), and the OUBM$_{EOT}$ model was best supported for the third cervical vertebrae

(AICcW = 0.69); simulations indicated substantial power to distinguish these models from BM

in both scenarios. Lastly, the BMBM$_{EOT}$ and OUBMi$_{EOT}$ models were equally supported for the

first thoracic and middle thoracic vertebrae (AICcW = 0.30–0.81), but simulations could not

distinguish these models from BM and EB models (Table S7).

Discussion

The global cooling, increased aridity, and habitat shift from forests to grasslands during

the past 56 My has spurred phenotypic diversification in many mammalian clades [7–10,14,15].

Here, we found that climatic and environmental changes from the Eocene-Oligocene Transition

(EOT) and Mid-Miocene Climate Transition (MMCT) significantly facilitated the diversification

of crown carnivoran skeletal phenomes across two distinct phases.

In the initial phase, EOT-induced climatic and environmental changes along with the

decline of coeval stem pan-carnivorans released crown carnivorans from a constrained adaptive

zone to radiation under Brownian motion, albeit at a slower evolutionary rate (Fig. 4A). With the

onset of the EOT, global temperatures plummeted and the first Antarctic ice sheets appeared,

transitioning Earth's climate from a warm, equable "greenhouse" to a cooler, temperate

"icehouse" with increased seasonality [33–35]. This period of climate change led to habitat

transitions from warm-humid forest to dry-temperate forest-steppe and grasslands, which in turn

led to massive faunal turnover [34–39]. Among pan-carnivorans, the EOT led to tremendous

taxonomic turnover, resulting in the decline of Hyaenodonta and Oxyaenodonta and giving

opportunities for carnivoran to radiate [11,40] under a BM pattern (Fig. 4A). Our analyses

suggesting EOT-induced interfamilial diversification is consistent with paleontological records

indicating that early carnivorans exhibited generalized, small-bodied forms and did not exhibit

major lineage and phenotypic diversification until after the EOT [11,12,27]. Niche competition

from stem pan-carnivorans may have initially constrained early carnivoran diversification prior

to the EOT [11–13,27]. However, the EOT marked the decline of stem pan-carnivorans [11,40],

possibly because they had become too specialized (i.e., macroevolutionary ratchet) to adapt to

the climatic and environmental changes brought by the EOT [11,12]. Thus, although both stem

pan-carnivorans and early carnivorans exhibited a macroevolutionary ratchet towards large body

size and hypercarnivory during this time, only carnivorans retained and increased their

phenotypic diversity, giving them the opportunity to exploit habitats of the Oligocene [12,13].

This may have further excluded stem pan-carnivorans from novel ecological opportunities [11–

13,27,28], facilitating stem pan-carnivoran decline but carnivoran diversification and the

appearance of all modern carnivoran families from the Early Oligocene to the Mid-Miocene [11].

Poor AICc support for all the optima shift models supports this hypothesis (Table 1). The lack of

distinct optima indicates pan-carnivoran groups exhibit similar skeletal phenomes and

ecomorphologies, further supporting that carnivorans displaced and replaced stem pan-

carnivorans.

Further intrafamilial diversification of carnivoran skeletal phenomes did not occur for

another 20 My until after the MMCT. During this second phase, another major period of rapid

temperature decline, increased aridity, and enhanced seasonality facilitated further trends

towards grasslands from forest habitats [6,41–43]. Our results suggested that these rapid climatic

and environmental changes along with few competing clades of carnivorous mammals enabled

crown carnivorans to exploit the new ecological niches from the late Miocene to Pleistocene

[11–13,28]. Specifically, intrafamilial diversification of skeletal phenomes may have facilitated

resource partitioning among ecologically similar taxa by increasing ecomorphological diversity

or innovations in locomotor mode, hunting behavior, and dietary ecology [15,44–46].

Consistently, the appendicular skeleton and posterior region of the vertebrae column, two

skeletal components that exhibit strong relationships with locomotor and hunting behavior

evolution [45], were characterized by models that incorporate shifts in evolutionary rate and/or

mode at the MMCT (Table S5). This suggests that evolutionary shifts or radiation of novel traits

in the appendicular and axial skeleton coincided with environmental changes of the Mid-

Miocene. For example, elbow-joint morphologies indicative of pounce-pursuit behaviors evolved

by the late Miocene within canids [15], presumably in response to the Mid-Miocene

diversification of grazing ungulates in open habitats [47,48]. Similarly, small, elongate body

plans with short legs evolved within mustelids [16,46], presumably to exploit the Mid-Miocene

diversification of rodents in open habitats, both above and subterranean [7,49]. More recent work

with extant carnivorans indicated that specific skeletal components exhibited strong selection

towards these distinct ecological peaks, and functional trade-offs and covariance among and

within skeletal components further explain their distribution across the adaptive landscape

[45,50,51]. Although multi-optima OU models with distinct ecological peaks may better capture

the evolutionary mode of these skeletal components, we are unable to test these models without

ecological data from extinct species. Unfortunately, ecological classifications are typically

inferred using the same data (i.e., skeletal traits) used in our analyses, which may lead to biased

results if the inferred ecologies of extinct species are used to test for ecological peaks.

Multi-phasic evolution driven by climatic and environmental changes and coupled with

taxonomic turnover appears to characterize the evolution of placental mammals. The delayed

intrafamilial diversification in carnivorans is reminiscent of the long fuse model used to describe

the diversification of placental mammals, which posits that the initial interordinal diversification

of placental mammals occurred in the Cretaceous whereas the majority of intraordinal

diversification took place after the K-Pg mass extinction event in response to newly available

niche [52–54]. The Paleocene–Eocene Thermal Maximum (PETM) may have also facilitated

distinct phases of mammalian evolution, albeit this has yet to be quantitatively tested. The rise of

forested environments and warmer climates after the PETM resulted in the replacement of stem

lineages of the early Paleocene such as multituberculates, pantodonts, and condylarths by crown

mammals such as primates, perissodactyls, and artiodactyls [55–57]. This turnover led to the

evolution of crown mammalian orders that dominated the Cenozoic and today's ecosystems [56].

Together, this body of work suggests that sequential evolutionary phases triggered by climatic

and environment changes were largely hierarchical. For carnivorans, the hierarchical progression

of ecological diversification across the Cenozoic led to the establishment of modern species.

Specifically, climatic and environmental changes from the K-Pg, PETM, EOT, and MMCT

facilitated diversification of infraclass Placentalia, mirorder Pan-Carnivora, families within

Carnivora, and genera within carnivoran families, respectively. Thus, our results suggest that the

long-fuse EOT/MMCT pattern we report here is as consequential for establishing modern-day

carnivore guilds as the PETM were for establishing the modern orders of mammals. Overall, this

work highlights the importance of analyzing data from extant and extinct species under a

phylogenetic comparative framework to disentangle the multiple climatic events that contributed

to the tiered evolution and diversity of carnivoran skeletal phenomes. Genetic analyses that

improve our assessment methods of skeletal variation will bring even more insight [58–60]. We

posit that quantification of skeletal phenomes in other crown mammals and their extinct relatives

will similarly reveal the effect of Cenozoic climatic change on the multi-phasal, hierarchical

establishment of modern-day mammalian biodiversity.

Methods

Morphological data

We quantified the skeletal phenome of 118 extant species and 81 extinct species using

103 linear trait measurements obtained from 208 extant specimens and 646 fossils (see Table S1

for list of specimens and museums). Together, this dataset includes seven traits in the cranium;

seven traits in the mandible; 13 traits in the forelimb (scapula, humerus, ulna, radius, third

metacarpal); 13 traits in the hindlimb (pelvis, femur, tibia, fibula, calcarean, and third

metatarsal); and seven traits in third cervical, fifth cervical, first thoracic, middle thoracic (the

sixth thoracic if the species has 13 or 14 thoracic vertebrae or the seventh thoracic if the species

has 15 or 16 thoracic vertebrae), diaphragmatic thoracic, last thoracic, first lumbar, middle

lumbar (the third lumbar if the species has four, five, or six lumbar vertebrae or the fourth

lumbar if the species has seven lumbar vertebrae), and last lumbar vertebrae (Fig. S1). These

measurements were chosen based on previous investigations of distinct ecomorphology among

carnivorans [23,61–64]. All measurements were taken using Mitutoyo digital calipers or a tape

measure. All specimens were fully mature, determined by the closure of exoccipital-basioccipital

and basisphenoid-basioccipital sutures on the cranium, full tooth eruption, or closure of sutures

on the limb bones or vertebrae. In our extant dataset, we obtained representatives from 14 of the

16 carnivoran families. We were unable to collect data from Prionodontidae, which contains two

extant species of Asiatic linsangs, and Odobenidae, which contains a single extant species, the

walrus. We only used male specimens to limit the differing degrees of sexual dimorphism that

carnivorans exhibit [65,66]. In our extinct dataset, we obtained representatives from seven extant

carnivoran families, seven extinct carnivoran families, stem carnivoramorphs, and stem pan-

carnivorans (Hyaenodontidae and Oxyaenidae). We were limited by the availability of fossils

that were formally described, placed in a phylogenetic context, and free from taphonomic or

other deformation.

Due to the fragmented nature of the fossil record, 73 extinct species were missing trait

measurements, resulting in 25.2% missing data in the full dataset (61.9% in extinct dataset). We,

therefore, imputed missing trait measurements using MIPCA (multiple imputation with PCA) in

the R package missMDA v1.19 [30]. MIPCA predicts missing trait measurements using an

iterative PCA algorithm with a predefined number of dimensions. The number of dimensions

was estimated by generalized cross-validation criterion, and the iterative algorithm was

performed under a Bayesian framework that alternates imputation of the data set and draw of the

PCA parameters in a posterior distribution [31].

We assessed the quality of the trait imputations by comparing imputed and empirical data

using a subset of species (n = 124) that contain all 103 measured traits. Specifically, we

generated four datasets with different percentages (10%, 30%, 60%, and 80%) of randomly

removed trait measurements and used MIPCA to impute these removed traits. We then assessed

the accuracy of MIPCA by plotting the imputed traits against the empirical average traits and

conducting reduced major axis regressions to test if the slope significantly differs from one. We

found strong relationships between imputed traits and empirical traits where the slope is not

significantly different from one for all datasets with different percentages of removed traits (see

Fig. S2 for two trait examples).

Using the imputed dataset, we removed the effects of size by dividing each skeletal trait

by the geometric mean of all 103 traits (i.e., log shape ratio: ln(trait/geometric mean) [67,68]. We

then conducted a principal component analysis (PCA) on the entire dataset and retained the first

seven PC axes (76.3% of explained variance) as our proxy of skeletal phenome. The number of

PC axes to retain was determined using the getMeaningfulPCs function in the R package Morpho

v2.11 [69]. We also used individual PCAs to reduce the dimension of each skeletal component

(i.e., log shape ratios from the cranium, mandible, forelimb, hindlimb, and each of the nine

vertebrae) and retained meaningful PC axes for subsequent analyses.

Phylogenetic data

We used a modified species-level phylogeny containing almost all known extant and

extinct carnivorans and stem relatives for our phylogenetic comparative methods [21]. After

pruning, the phylogeny contained all 118 extant species and 73 of the 81 extinct species in our

dataset. We manually added the following extinct species: *Crassidia intermedia* was placed as

basal caniform and sister to the genus *Daphoenus* [70]; *Corumictis wolsani* was placed as stem

to Mustelidae and sister to the genus *Promartes* [71]; *Cyonasua brevirostris* was placed as sister

to genus *Nasua* [72]; *Eoarctos vorax* was placed as sister to Ursidae [73]; *Puma pardoides* was

placed as sister to *Puma concolor* [74]; *Satherium ingens* was placed as sister to sister to genus

Pteronura [75]; *Neovulpavus mccarrolli* was placed as a stem carnivoramorphan and sister to

Miacis washakius [76]; and *Promartes olcotti* was placed as a stem to Mustelidae and sister to

the genus *Corumictis* [77] (Table S8).

Phylogenetic Comparative Methods

We first measured phylogenetic signal in skeletal phenome, skeletal size, and each

skeletal component using the physignal.z function in R package geomorph v4.0.6 [78]. We then

examined subclade disparity in each dataset of retained PC axes by performing disparity through

time (DTT) analyses [79] using fdtt [22], a modified version of the dtt function in the R package

geiger v2.0.11 [80]. We compared carnivoran DTT to a null, constant-rates distribution derived

from 10,000 Brownian motion simulations, and computed the morphological disparity index

(MDI) as the area between the empirical DTT curve and the median of these simulations.

We then tested 16 alternative hypotheses that could explain carnivoran skeletal evolution

by fitting a series of macroevolutionary models (Fig. 3). Models 1–4 are based on single rate

Brownian motion or single optimum Ornstein-Uhlenbeck process:

1. Brownian motion (BM): Suggests that disparity in skeletal phenome accumulates

proportionally to evolutionary time under a random walk.

2. Trend: Suggests that disparity in skeletal phenome accumulates under BM with a

directional drift.

3. Early burst (EB): Suggests that disparity in skeletal phenome initially accumulates

rapidly but declines over time.

4. Single optimum Ornstein-Uhlenbeck process (OU1): Suggests that disparity in skeletal

phenome is constrained or under selection to a single optimum over time.

Models 5–12 are climate shift models to test if evolutionary rates, skeletal optima, and/or

evolutionary mode differs before and after the Eocene Oligocene Transition (EOT) and Mid

Miocene Climate Transition (MMCT):

5. Rate shift after the EOT ($BMBM_{EOT}$). Suggests that skeletal phenome exhibits two

distinct evolutionary rates before and after the EOT.

6. Rate shift after the MMCT ($BMBM_{MMCT}$). Similar to the $BMBM_{EOT}$ model, but the shift

occurs at the MMCT.

7. Optima shift after the EOT ($OUOU_{EOT}$). Suggests that skeletal phenome exhibits two

distinct evolutionary optima before and after the EOT with the same evolutionary rate.

8. Optima shift after the MMCT ($OUOU_{MMCT}$). Similar to the $OUOU_{EOT}$ model, but the

shift occurs at the MMCT.

9. Ecological release after the EOT ($OUBM_{EOT}$). Suggests that skeletal phenomes are

constricted in ancestral carnivorans via an OU model due to extrinsic factors associated

with the EOT, followed by the increased skeletal disparity after the release from these

factors via BM.

10. Ecological release after the MMCT ($OUBMi_{MMCT}$). Similar to the $OUBM_{EOT}$ model, but

the shift occurs at the MMCT.

11. Release and radiate after the EOT ($OUBMi_{EOT}$). Similar to the $OUBM_{EOT}$ model, except

the evolutionary rate also shifts after the EOT.

12. Release and radiate after the MMCT (OUBMi$_{MMCT}$). Similar to the OUBMi$_{MMCT}$ model,

except the evolutionary rate also shifts after the MMCT.

Models 13–16 are optima shift models to test if carnivorans and stem relatives exhibit

separate optima. Because ecological factors influence several aspects of the carnivoran

skeleton [44,45,64,81], distinct skeletal phenomes may potentially indicate a reduction of

ecological competition through character displacement:

13. Stepwise optimal shift (sOU2): Suggests that carnivorans and its stem relatives exhibit

separate skeletal phenome optima via a two-peak OU model.

14. Stepwise optimal shift (sOU3): Suggests that carnivorans, stem carnivoramorph relatives,

and stem pan-carnivoran relatives exhibit separate skeletal phenome optima via a three-

peak OU model.

15. Clade optimal shift (cOU3): Suggests that feliforms and caniforms exhibit separate

skeletal phenome optima after an initial optimal shift from stem relatives via a three-peak

OU model.

16. Clade optimal shift (cOU4): Suggests that feliforms and caniforms exhibit separate

skeletal phenome optima after initial optimal shifts from stem carnivoramorph relatives

and stem pan-carnivoran via a four-peak OU model.

All models were fit to skeletal phenome, skeletal size, and each skeletal component datasets with

measurement error incorporated using the R package mvMORPH v1.1.7 [82]. We did not have

complete intraspecific data to calculate standard errors for each species. Instead, we estimated

standard errors of each species using the following procedure. First, we estimated the standard

deviation of each PC axis of each species by multiplying the range of each PC axis with 5%.

This approximation assumes that on average each species would have an estimated standard

deviation of 5% of the maximum range of the full species range on that axis. We then estimated

the standard error by dividing the estimated standard deviation with the square root of the total

number of species. Optima shifts in the sOU and cOU models were specified on the phylogeny

using the paintSubTree function in the R package phytools v2.4.4 [83]. Relative support for each

of the 16 models was assessed through computation of small sample-corrected Akaike (AICc).

Models with ΔAICc < 2 were considered as equally well supported. We assessed whether we had

adequate power to accurately distinguish between complex models from Brownian motion using

simulations [84]. We generated 500 simulations for all datasets using the parameter estimates of

the best-supported model(s) from the empirical datasets. We then ran each set of simulated data

through all 16 models to determine whether the best model(s) could be accurately recovered.

As a sensitivity analysis, we also fit all 16 models to the first seven axes from a

phylogenetic PCA of skeletal phenome (56.6% of explained variance). We found nearly equal

support for the two release and radiate models (OUBMi$_{EOT}$: AICcW = 0.74; OUBMi$_{MMCT}$:

ΔAICc = 2.06, AICcW = 0.26; Table S9), a result similar to model selection using a standard

PCA. We chose not to continue with the phylogenetic PCA because pPCA is more difficult to

interpret as it is a mixture of major axes that describe non-phylogenetic variation and scores that

contain phylogenetic components of variation [85]. Furthermore, pPCA axes are not orthogonal

to each other, so the first few axes we used in this study may include less variance explained than

PCA by containing correlated variance components rather than independent ones.

Acknowledgments

Declaration of interests

The authors declare no competing interests

Data accessibility.

Dataset and R script are uploaded on Dryad Digital Repository:

doi.org/10.5061/dryad.z8w9ghxrz.

References

1. Simpson GG. 1944 *Tempo and Mode in Evolution*. New York, NY: Columbia University
Press.

2. Simpson GG. 1955 *Major features of evolution*. New York, NY: Columbia University Press.

3. Schluter D. 2000 *The Ecology of Adaptive Radiation*. Oxford University Press.

4. Zachos J, Pagani M, Sloan L, Thomas E, Billups K. 2001 Trends, Rhythms, and Aberrations
in Global Climate 65 Ma to Present. *Science* **292**, 686–693. (doi:10.1126/science.1059412)

5. Retallack GJ. 2001 Cenozoic expansion of grasslands and climatic cooling. *J. Geol.* **109**, 407–
426. (doi:10.1086/320791)

6. Strömberg CAE. 2011 Evolution of Grasses and Grassland Ecosystems. *Annu. Rev. Earth
Planet. Sci.* **39**, 517–544. (doi:10.1146/annurev-earth-040809-152402)

7. Samuels JX, Hopkins SSB. 2017 The impacts of Cenozoic climate and habitat changes on
small mammal diversity of North America. *Glob. Planet. Change* **149**, 36–52.
(doi:10.1016/j.gloplacha.2016.12.014)

8. Damuth J, Janis CM. 2011 On the relationship between hypsodonty and feeding ecology in
ungulate mammals, and its utility in palaeoecology. *Biol Rev* **86**, 733–758. (doi:10.1111/j.1469-
185x.2011.00176.x)

9. Janis CM, Damuth J, Theodor JM. 2002 The origins and evolution of the North American
grassland biome: the story from the hoofed mammals. *Palaeogeogr Palaeoclim Palaeoecol* **177**,
183–198. (doi:10.1016/s0031-0182(01)00359-5)

10. Levering D, Hopkins S, Davis E. 2017 Increasing locomotor efficiency among North
American ungulates across the Oligocene-Miocene boundary. *Palaeogeogr. Palaeoclimatol.*
Palaeoecol. **466**, 279–286. (doi:10.1016/j.palaeo.2016.11.036)

11. Van Valkenburgh B. 1999 Major patterns in the history of carnivorous mammals. *Annu. Rev.*
Earth Planet. Sci. **27**, 463–493.

12. Friscia AR, Valkenburgh BV. 2010 Ecomorphology of North American Eocene carnivores:
evidence for competition between Carnivorans and Creodonts. In *Carnivoran Evoution*, pp. 311–
341. Cambridge, United Kingdom: Cambridge University Press.
(doi:10.1017/cbo9781139193436.012)

13. Wesley-Hunt GD. 2005 The morphological diversification of carnivores in North America.
Paleobiology **31**, 35–55. (doi:10.1666/0094-8373(2005)031<0035:tmdoci>2.0.co;2)

14. Andersson K, Werdelin L. 2003 The evolution of cursorial carnivores in the Tertiary:
implications of elbow-joint morphology. *Proc R Soc B* **270**, S163–S165.
(doi:10.1098/rsbl.2003.0070)

15. Figueirido B, Martín-Serra A, Tseng ZJ, Janis CM. 2015 Habitat changes and changing
predatory habits in North American fossil canids. *Nature Communications* **6**, 1–11.
(doi:10.1038/ncomms8976)

16. Law CJ. 2019 Evolutionary shifts in extant mustelid (Mustelidae: Carnivora) cranial shape,
body size and body shape coincide with the Mid-Miocene Climate Transition. *Biol. Lett.* **15**,
20190155–6. (doi:10.1098/rsbl.2019.0155)

17. Finarelli JA, Flynn JJ. 2006 Ancestral state reconstruction of body size in the Caniformia
(Carnivora, Mammalia): the effects of incorporating data from the fossil record. *Syst. Biol.* **55**,
301–313. (doi:10.1080/10635150500541698)

18. Slater GJ, Harmon LJ, Alfaro ME, Alfaro ME. 2012 Integrating fossils with molecular
phylogenies improves inference of trait evolution. *Evolution* **66**, 3931–3944.
(doi:10.1111/j.1558-5646.2012.01723.x)

19. Slater GJ. 2013 Phylogenetic evidence for a shift in the mode of mammalian body size
evolution at the Cretaceous-Palaeogene boundary. *Methods Ecol Evol* **4**, 734–744.
(doi:10.1111/2041-210x.12084)

20. Mitchell JS. 2015 Extant-only comparative methods fail to recover the disparity preserved in
the bird fossil record. *Evolution* **69**, 2414–2424. (doi:10.1111/evo.12738)

21. Faurby S, Silvestro D, Werdelin L, Antonelli A. 2024 Reliable biogeography requires fossils:
insights from a new species-level phylogeny of extinct and living carnivores. *Proc. R. Soc. B*
291, 20240473. (doi:10.1098/rspb.2024.0473)

22. Slater GJ, Goldbogen JA, Pyenson ND. 2017 Independent evolution of baleen whale
gigantism linked to Plio-Pleistocene ocean dynamics. *Proc R Soc B* **284**, 20170546–8.
(doi:10.1098/rspb.2017.0546)

23. Jones KE, Angielczyk KD, Polly PD, Head JJ, Fernandez V, Lungmus JK, Tulga S, Pierce
SE. 2018 Fossils reveal the complex evolutionary history of the mammalian regionalized spine.
Science **361**, 1249–1252. (doi:10.1126/science.aar3126)

24. Jones KE, Angielczyk KD, Pierce SE. 2019 Stepwise shifts underlie evolutionary trends in
morphological complexity of the mammalian vertebral column. *Nat Commun* **10**, 5071.
(doi:10.1038/s41467-019-13026-3)

25. Goswami A *et al.* 2022 Attenuated evolution of mammals through the Cenozoic. *Science*
378, 377–383. (doi:10.1126/science.abm7525)

26. Janis CM, Scott KM, Jacobs LL. 1998 *Evolution of Tertiary Mammals of North America:*
Terrestrial carnivores, ungulates, and ungulatelike mammals. Cambridge, United Kingdom:
Cambridge University Press.

27. Flynn JJ. 1998 Early Cenozoic Carnivora ('Miacoidea'). In *Evolution of Tertiary Mammals*
of North America: Terrestrial carnivores, ungulates, and ungulatelike mammals (eds CM Janis,
KM Scott, LL Jacobs), pp. 110–123. Cambridge, United Kingdom: Cambridge University Press.

28. Werdelin L. 1996 Carnivoran Ecomorphology: A Phylogenetic Perspective. In *Carnivore*
Behavior, Ecology, and Evolution. Volume II, pp. 582–624. Cornell, NY: Cornell University
Press. (doi:10.7591/9781501745829-024)

29. Law CJ. 2021 Evolutionary and morphological patterns underlying carnivoran body shape
diversity. *Evolution* **75**, 365–375. (doi:10.1111/evo.14143)

30. Josse J, Husson F. 2016 missMDA: a package for handling missing values in multivariate
data analysis. *J Stat Softw* **70**, 1–31. (doi:10.18637/jss.v070.i01)

31. Audigier V, Husson F, Josse J. 2016 Multiple imputation for continuous variables using a
Bayesian principal component analysis†. *J. Stat. Comput. Simul.* **86**, 2140–2156.
(doi:10.1080/00949655.2015.1104683)

32. Bookstein FL. 2014 *Measuring and Reasoning: Numerical Inference in the Sciences*.
Cambridge University Press.

33. Liu Z, Pagani M, Zinniker D, DeConto R, Huber M, Brinkhuis H, Shah SR, Leckie RM,
Pearson A. 2009 Global Cooling During the Eocene-Oligocene Climate Transition. *Science* **323**,
1187–1190. (doi:10.1126/science.1166368)

34. Prothero DR, Berggren WA. 1992 *Eocene-Oligocene Climatic and Biotic Evolution*.
Princeton, NJ: Princeton University Press.

35. Hutchinson DK *et al.* 2020 The Eocene-Oligocene transition: a review of marine and
terrestrial proxy data, models and model-data comparisons. *Clim. Past Discuss.* **2020**, 1–71.
(doi:10.5194/cp-2020-68)

36. Vries D de, Heritage S, Borths MR, Sallam HM, Seiffert ER. 2021 Widespread loss of
mammalian lineage and dietary diversity in the early Oligocene of Afro-Arabia. *Commun. Biol.*
4, 1172. (doi:10.1038/s42003-021-02707-9)

37. Meng J, McKenna MC. 1998 Faunal turnovers of Palaeogene mammals from the Mongolian
Plateau. *Nature* **394**, 364–367. (doi:10.1038/28603)

38. Sun J, Ni X, Bi S, Wu W, Ye J, Meng J, Windley BF. 2014 Synchronous turnover of flora,
fauna and climate at the Eocene–Oligocene Boundary in Asia. *Sci. Rep.* **4**, 7463.
(doi:10.1038/srep07463)

39. Prothero DR. 1985 Mid-Oligocene Extinction Event in North American Land Mammals.
Science **229**, 550–551. (doi:10.1126/science.229.4713.550)

40. Solé F, Fischer V, Verger KL, Mennecart B, Speijer RP, Peigné S, Smith T. 2022 Evolution
of European carnivorous mammal assemblages through the Palaeogene. *Biol. J. Linn. Soc.* **135**,
734–753. (doi:10.1093/biolinnean/blac002)

41. Herbert TD, Lawrence KT, Tzanova A, Peterson LC, Caballero-Gill R, Kelly CS. 2016 Late
Miocene global cooling and the rise of modern ecosystems. *Nat. Geosci.* **9**, 843–847.
(doi:10.1038/ngeo2813)

42. Flower BP, Kennett JP. 1994 The middle Miocene climatic transition: East Antarctic ice
sheet development, deep ocean circulation and global carbon cycling. *Palaeogeogr.,*
Palaeoclim., Palaeoecol. **108**, 537–555. (doi:10.1016/0031-0182(94)90251-8)

43. Cerling TE, Harris JM, MacFadden BJ, Leakey MG, Quade J, Eisenmann V, Ehleringer JR.
1997 Global vegetation change through the Miocene/Pliocene boundary. *Nature* **389**, 153–158.
(doi:10.1038/38229)

44. Slater GJ. 2022 Topographically distinct adaptive landscapes for teeth, skeletons, and size
explain the adaptive radiation of Carnivora (Mammalia). *Evolution* (doi:10.1111/evo.14577)

45. Law CJ, Hlusko LJ, Tseng ZJ. 2024 Uncovering the mosaic evolution of the carnivoran
skeletal system. *Biol. Lett.* **20**, 20230526. (doi:10.1098/rsbl.2023.0526)

46. Law CJ, Slater GJ, Mehta RS. 2019 Shared extremes by ectotherms and endotherms: Body
elongation in mustelids is associated with small size and reduced limbs. *Evolution* **73**, 735–749.
(doi:10.1111/evo.13702)

47. Janis CM, Damuth J, Theodor JM. 2000 Miocene ungulates and terrestrial primary
productivity: Where have all the browsers gone? *Proc. Natl. Acad. Sci.* **97**, 7899–7904.
(doi:10.1073/pnas.97.14.7899)

48. Janis C. 2008 The Ecology of Browsing and Grazing. *Ecol. Stud.* , 21–45. (doi:10.1007/978-
3-540-72422-3_2)

49. Calede JJM, Hopkins SSB, Davis EB. 2011 Turnover in burrowing rodents: The roles of
competition and habitat change. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **311**, 242–255.
(doi:10.1016/j.palaeo.2011.09.002)

50. Law CJ, Blackwell EA, Curtis AA, Dickinson E, Hartstone☐Rose A, Santana SE. 2022
Decoupled evolution of the cranium and mandible in carnivoran mammals. *Evolution* **76**, 2959–
2974. (doi:10.1111/evo.14578)

51. Law CJ, Hlusko LJ, Tseng ZJ. 2025 The Carnivoran Adaptive Landscape Reveals Trade-offs
among Functional Traits in the Skull, Appendicular, and Axial Skeleton. *Integr. Org. Biol.* **7**,
obaf001. (doi:10.1093/iob/obaf001)

52. Meredith RW *et al.* 2011 Impacts of the Cretaceous Terrestrial Revolution and KPg
Extinction on Mammal Diversification. *Science* **334**, 521–524. (doi:10.1126/science.1211028)

53. Springer MS, Foley NM, Brady PL, Gatesy J, Murphy WJ. 2019 Evolutionary Models for the
Diversification of Placental Mammals Across the KPg Boundary. *Front. Genet.* **10**, 1241.
(doi:10.3389/fgene.2019.01241)

54. Archibald JD, Deutschman DH. 2001 Quantitative Analysis of the Timing of the Origin and
Diversification of Extant Placental Orders. *J. Mamm. Evol.* **8**, 107–124.
(doi:10.1023/a:1011317930838)

55. Rose K. 2006 *The Beginning of the Age of Mammals*. Baltimore, MD: John Hopkins
University Press. (doi:10.56021/9780801884726)

56. Novacek MJ. 1999 100 Million Years of Land Vertebrate Evolution: The Cretaceous-Early
Tertiary Transition. *Annals of the Missouri Botanical Garden* **86**, 230–258.

57. Gingerich PD. 2006 Environment and evolution through the Paleocene–Eocene thermal
maximum. *Trends Ecol. Evol.* **21**, 246–253. (doi:10.1016/j.tree.2006.03.006)

58. Hlusko LJ, Schmitt CA, Monson TA, Brasil MF, Mahaney MC. 2016 The integration of
quantitative genetics, paleontology, and neontology reveals genetic underpinnings of primate
dental evolution. *Proc Natl Acad Sci USA* **113**, 9262–9267. (doi:10.1073/pnas.1605901113)

59. Sadier A *et al.* 2023 Bat teeth illuminate the diversi fi cation of mammalian tooth classes.
Nature Communications **14**, 4687.

60. Couzens AMC, Sears KE, Rücklin M. 2021 Developmental influence on evolutionary rates
and the origin of placental mammal tooth complexity. *Proc. Natl. Acad. Sci.* **118**, e2019294118.
(doi:10.1073/pnas.2019294118)

61. Radinsky LB. 1981 Evolution of skull shape in Carnivores .1. Representative modern
Carnivores. *Biol J Linn Soc* **15**, 369–388. (doi:10.1111/j.1095-8312.1981.tb00770.x)

62. Friscia AR, Van Valkenburgh B, Biknevicius AR. 2007 An ecomorphological analysis of
extant small carnivorans. *J Zool* **272**, 82–100. (doi:10.1111/j.1469-7998.2006.00246.x)

63. Pierce SE, Clack JA, Hutchinson JR. 2011 Comparative axial morphology in pinnipeds and
its correlation with aquatic locomotory behaviour. *J Anat* **219**, 502–514. (doi:10.1111/j.1469-
7580.2011.01406.x)

64. Samuels JX, Meachen JA, Sakai SA. 2013 Postcranial morphology and the locomotor habits
of living and extinct carnivorans. *J Morpho* **274**, 121–146. (doi:10.1002/jmor.20077)

65. Law CJ. 2019 Solitary meat-eaters: solitary, carnivorous carnivorans exhibit the highest
degree of sexual size dimorphism. *Sci Rep-uk* **9**, 15344. (doi:10.1038/s41598-019-51943-x)

66. Morris JS, Carrier DR. 2016 Sexual selection on skeletal shape in Carnivora. *Evolution* **70**,
767–780. (doi:10.1111/evo.12904)

67. Mosimann JE. 1970 Size Allometry: Size and Shape Variables with Characterizations of the
Lognormal and Generalized Gamma Distributions. *J. Am. Stat. Assoc.* **65**, 930–945.
(doi:10.1080/01621459.1970.10481136)

68. Claude J. 2013 Log-shape ratios, Procrustes superimposition, elliptic Fourier analysis: three
worked examples in R. *Hystrix, the Italian Journal of Mammalogy* **24**, 94–102.

69. Schlager S. 2016 *Morpho: Calculations and visualisations related to Geometric*
Morphometrics. R-package version 2.4. See
ftp://xyz.csail.mit.edu/pub/CRAN/web/packages/Morpho/Morpho.pdf.

70. Tomiya S, Tseng ZJ. 2016 Whence the beardogs? Reappraisal of the Middle to Late Eocene
"Miacis" from Texas, USA, and the origin of Amphicyonidae (Mammalia, Carnivora). *Royal*
Society Open Science **3**, 160518. (doi:10.1098/rsos.160518)

71. Paterson R, Samuels JX, Rybczynski N, Ryan MJ, Maddin HC. 2019 The earliest mustelid in
North America. *Zool J Linn Soc-lond* (doi:10.1093/zoolinnean/zlz091)

72. Forasiepi AM, Soibelzon LH, Gomez CS, Sánchez R, Quiroz LI, Jaramillo C, Sánchez-
Villagra MR. 2014 Carnivorans at the Great American Biotic Interchange: new discoveries from
the northern neotropics. *Naturwissenschaften* **101**, 965–974. (doi:10.1007/s00114-014-1237-4)

73. Wang X, Emry RJ, Boyd CA, Person JJ, White SC, Tedford RH. 2022 An exquisitely
preserved skeleton of Eoarctos vorax (nov. gen. et sp.) from Fitterer Ranch, North Dakota (early
Oligocene) and systematics and phylogeny of North American early arctoids (Carnivora,
Caniformia). *J. Vertebr. Paléontol.* **42**, 1–123. (doi:10.1080/02724634.2022.2145900)

74. Cherin M, Iurino DA, Sardella R. 2013 Earliest occurrence of Puma pardoides (Owen, 1846)
(Carnivora, Felidae) at the Plio/Pleistocene transition in western Europe: New evidence from the
Middle Villafranchian assemblage of Montopoli, Italy. *C. R. Palevol* **12**, 165–171.
(doi:10.1016/j.crpv.2013.01.002)

75. DeNeve Weeks DL. 2020 Resilience and Sensitivity to Changing Environments in North
American river otters (Lontra canadensis). UC Merced Electronic Theses and Dissertations.

76. Tomiya S, Zack SP, Spaulding M, Flynn JJ. 2021 Carnivorous mammals from the middle
Eocene Washakie Formation, Wyoming, USA, and their diversity trajectory in a post-warming
world. *J. Paléontol.* **95**, 1–115. (doi:10.1017/jpa.2020.74)

77. Wang X, McKenna MC, Dashzeveg D. 2005 Amphicticeps and Amphicynodon (Arctoidea,
Carnivora) from Hsanda Gol Formation, central Mongolia and phylogeny of basal arctoids with
comments on zoogeography. **3483**, 1–60. (doi:10.1206/0003-
0082(2005)483[0001:aaaacf]2.0.co;2)

78. Baken EK, Collyer ML, Kaliontzopoulou A, Adams DC. 2021 geomorph v4.0 and gmShiny:
Enhanced analytics and a new graphical interface for a comprehensive morphometric experience.
Methods Ecol Evol **12**, 2355–2363. (doi:10.1111/2041-210x.13723)

79. Harmon LJ, Harmon LJ, Schulte JA, Larson A, Losos JB. 2003 Tempo and mode of
evolutionary radiation in iguanian lizards. *Science* **301**, 961–964. (doi:10.1126/science.1084786)

80. Pennell MW, Eastman JM, Slater GJ, Brown JW, Uyeda JC, FitzJohn RG, Alfaro ME,
Harmon LJ. 2014 geiger v2.0: an expanded suite of methods for fitting macroevolutionary
models to phylogenetic trees. *Bioinformatics* **30**, 2216–2218.
(doi:10.1093/bioinformatics/btu181)

81. Law CJ. 2021 Ecological drivers of carnivoran body shape evolution. *Am Nat* **198**, 406–420.
(doi:10.5061/dryad.pg4f4qrpm)

82. Clavel J, Escarguel G, Merceron G. 2015 mv morph: an R package for fitting multivariate
evolutionary models to morphometric data. *Methods Ecol Evol* **6**, 1311–1319.
(doi:10.1111/2041-210x.12420)

83. Revell LJ. 2024 phytools 2.0: an updated R ecosystem for phylogenetic comparative methods
(and other things). *PeerJ* **12**, e16505.

84. Boettiger C, Coop G, Ralph P. 2012 Is your phylogeny informative? Measuring the power of
comparative methods. *Evolution* **66**, 2240–2251. (doi:10.5061/dryad.m6r6hc12)

85. Polly PD, Lawing AM, Fabre A-C, Goswami A. 2013 Phylogenetic principal components analysis and geometric morphometrics. *Hystrix, the Italian Journal of Mammalogy* **24**, 33–41.

Fig. 1. Pruned pan-carnivoran phylogeny with ancestral reconstruction of PC 1 of skeletal phenome, calculated from all 103 size-corrected skeletal traits.

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Fig. 2. Disparity-through-time plots of carnivoran skeletal phenome and size. (A) Skeletal
phenome deviates from Brownian motion 37 Ma (blue arrow) near the beginning of the Eocene-
Oligocene Transition (EOT) and 14 Ma near the end of the Mid-Miocene Climate Transition
(MMCT). Blue solid line indicates the empirical subclade DTT, and the dashed black line
indicates the median DTT based on 10,000 simulations of trait evolution under Brownian
motion. The grey shaded area indicates the 95% DTT range for the simulated data. Above the
DTT is the global deep-sea oxygen isotope records as a proxy for global temperature change
(modified from [4]). DTT suggests increased subclade disparity at two time points: (B) just prior
to the EOT (~39-37 Ma; MDI = 0.534, P = 0.012) and (C) just after the MMCT (~14 Ma; MDI =
0.238, P = 0.018). For B. and C., histograms show the distribution of simulated subclade
disparities and red arrows signify the empirical subclade disparities at time 37 Ma and 14 Ma,
respectively. (D) Skeletal size does not deviate from Brownian motion across time and at (E) ~37
Ma and (F) ~14 Ma.
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Fig. 3. Alternative hypotheses for the evolution of carnivoran skeletal phenomes. (A) Brownian
motion. (B) Brownian motion with a directional drift. (C) Brownian motion with an evolutionary
rate that increases or decreases exponentially through time. (D) Single optimum under an
Ornstein-Uhlenbeck process. (E) Single rate shift after the Eocene-Oligocene Transition (EOT)
or Mid-Miocene Climate Transition (MMCT). (F) Single optimal shift after the EOT or MMCT.
(G) Constrained evolution followed by unconstrained evolution after the EOT or MMCT. (H)
Constrained evolution followed by unconstrained evolution after the EOT or MMCT with
distinct evolutionary rates. (I) Stepwise optimal shifts between carnivorans and its stem relatives
(sOU2) or among carnivorans, stem carnivoramorphs, and stem pan-carnivorans (sOU3). (J)
Stepwise optimal shifts followed by clade optimal shifts among caniforms, feliforms, and its
stem relatives (cOU3) or among caniforms, feliforms, stem carnivoramorphs, and stem pan-
carnivorans (cOU4).
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Fig. 4. Simulations of the parameter estimates of the best supported models for skeletal phenome. 100 simulations were used for visualization.

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702 **Table 1.** Comparisons of evolutionary models for skeletal phenome and skeletal size. Rows in
703 boldface type represent the best supported model(s) as indicated by the lowest ΔAICc score.
704 Models with ΔAICc < 2 were considered as equally well supported. AICcW = AICc weights.

Trait	Model	AICc	ΔAICc	AICcW
skeletal phenome	BM1	883.53	51.98	0.00
	trend	880.84	49.30	0.00
	EB	891.17	59.62	0.00
	OU1	907.73	76.18	0.00
	$BMBMi_{EOT}$	849.15	17.60	0.00
	$OUOU_{EOT}$	890.45	58.91	0.00
	$OUBM_{EOT}$	883.35	51.81	0.00
	$OUBMi_{EOT}$	**831.55**	**0.00**	**0.66**
	$BMBMi_{MMCT}$	859.67	28.12	0.00
	$OUOU_{MMCT}$	898.95	67.40	0.00
	$OUBM_{MMCT}$	879.37	47.82	0.00
	$OUBMi_{MMCT}$	**832.87**	**1.33**	**0.34**
	sOU2	921.34	89.80	0.00
	sOU3	931.93	100.38	0.00
	cOU3	939.50	107.95	0.00
	cOU4	926.50	94.95	0.00
skeletal size	BM1	168.80	6.09	0.03
	trend	167.63	4.93	0.05
	EB	170.86	8.15	0.01
	OU1	172.89	10.19	0.00
	$BMBMi_{EOT}$	168.31	5.61	0.04
	$OUOU_{EOT}$	168.48	5.78	0.03
	$OUBM_{EOT}$	165.88	3.18	0.12
	$OUBMi_{EOT}$	167.92	5.22	0.04
	$BMBMi_{MMCT}$	171.14	8.44	0.01
	$OUOU_{MMCT}$	171.89	9.19	0.01
	$OUBM_{MMCT}$	166.69	3.98	0.08
	$OUBMi_{MMCT}$	**162.70**	**0.00**	**0.58**
	sOU2	173.31	10.61	0.00
	sOU3	175.23	12.52	0.00
	cOU3	175.31	12.61	0.00
	cOU4	177.37	14.66	0.00