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Title:

3 A comparative GWAS of eye colour in light and dark eye genetic backgrounds defined by

4 *HERC2* rs12913832 polymorphism

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23 **Short title:** *HERC2* comparative eye colour GWAS

24 **Keywords:** rs12913832, eye colour, genome-wide association study (GWAS), *HERC2*, *OCA2*,

25 quantitative eye colour measures

Abstract

rs12913832, a polymorphism located in an enhancer within the *HERC2* gene, which is known to regulate *OCA2* transcription, is heavily relied upon as a predictor of light versus dark eyes. Individuals with the GG genotype are projected to have blue eyes, while individuals with the AA or AG genotypes are projected to have darker eye colours (primarily brown). However, eye colour is a polygenic trait, and previous studies have revealed that a significant proportion of individuals self-report an eye colour that is not concordant with their genotype at rs12913832. Herein, we address the question: What common markers are influencing eye colour in individual's whose self-reported phenotype does not correspond to the expected phenotype based on their rs12913832 genotype? Building upon our prior investigation of iris pigmentation genetics in individuals with an expected "blue eye" background (rs12913832:GG genotype) in a sample of the Canadian Partnership for Tomorrow's Health (CanPath) cohort, this study extends the analysis to include individuals with an expected "brown eye" background (rs12913832:AA+AG). We identified variants in *SLC45A2*, *TYRP1*, *TYR*, *SLC24A4* and *TSPAN10*, which may influence eye colour presentation in individuals with the rs12913832:GG genotype and variants in *IRF4*, *TYRP1* and *OCA2*, which may influence eye colour presentation in individuals with the rs12913832:AA+AG genotype. These markers include well-known pigmentation-associated single nucleotide polymorphisms, such as rs16891982 (*SLC45A2*), rs1126809 (*TYR*), rs12203592 (*IRF4*), rs1800407 (*OCA2*) and rs6420484 (*TSPAN10*). Several of these loci were replicated using independent quantitative eye colour measures, including heterochromia and CIELAB colour dimensions. This research highlights the importance of gene-gene interactions and the polygenic nature of pigmentation traits, emphasizing modifying effects that can sometimes counteract the

60   dominant influence of rs12913832, contributing to advancements in pigmentation genetics and

61   forensic applications.

Introduction

Human pigmentation traits, such as hair, skin, and eye colour, represent some of the most striking

examples of visible genetic variation. These traits are highly polygenic, with complex gene-gene

and gene-environment interactions contributing to their phenotypic expression. Understanding the

genetic architecture underlying these traits not only informs biological mechanisms but also

enhances applications such as predictive modeling, reconstruction of ancestral phenotypes and

human identification. Of particular interest is the genetic basis underlying iris pigmentation,

which remains incompletely understood despite significant advances in large-scale genome-wide

association studies (GWAS) (1).

A key genetic determinant of human eye colour is the single nucleotide polymorphism (SNP)

rs12913832, located in a conserved enhancer region within the *HERC2* gene that regulates

transcription of the neighbouring *OCA2* gene (2–5). *OCA2* plays a central role in melanin

biosynthesis, and its expression levels are strongly influenced by allelic variation at rs12913832.

The ancestral A allele facilitates transcription factor binding and promotes long-range chromatin

looping, enabling physical interaction between the *OCA2* promoter and the *HERC2* enhancer. This

configuration enhances *OCA2* expression, leading to increased melanin production and resulting

in darker eye colours, such as brown and hazel (4,6). Conversely, the derived G allele reduces

chromatin looping efficiency, thereby decreasing *OCA2* expression and melanin production. This

reduction is associated with lighter iris pigmentation, characteristic of blue eyes (4,6). As such,

rs12913832 has become a key marker in pigmentation research and forensic DNA phenotyping

(FDP), due to its strong predictive value for distinguishing between light and dark eye colours

(2,4,7). Individuals with the GG genotype are typically predicted to have blue eyes, while those

with AG or AA genotypes are generally expected to exhibit brown eyes (**Fig 1**). However, while

these patterns hold true for most individuals, particularly within European populations where eye

colour diversity is greatest, genotype-phenotype mismatches have been reported.

Fig 1. **Expected eye colour phenotype based on rs12913832 genotype.**

Specifically, previous research has illustrated that some rs12913832:GG individuals present with

eye colours other than blue, and some rs12913832:AA or AG individuals show eye colours other

than brown (8–10). A recent publication found a notable discrepancy in the Canadian Partnership

for Tomorrow's Health (CanPath) cohort where 33% of individuals with the rs12913832:GG

genotype self-reported eye colours other than blue (11). This finding challenges the notion that

the rs12913832:GG genotype exclusively corresponds to blue eyes and highlights the presence of

additional genetic or regulatory factors that may modulate melanin production, resulting in a

broader spectrum of iris pigmentation. In a previous study (12), we stratified participants of the

CanPath cohort based on rs12913832 genotypes and identified a number of SNPs that seem to

modify the effect of the rs12913832 GG genotype, contributing to an explanation as to why

individuals self-report "non-blue" eye colour in spite of having severely reduced expression of the

OCA2 gene, which usually results in blue eye colour.

The present study builds on this foundation by extending the 2023 study in two important ways:

1) Identifying in the same CanPath sample used in the 2023 study potential genetic modifiers that

may explain individuals with rs12913832:AA+AG genotypes reporting eye colours other than

brown and hazel, and 2) Replicating the same markers in another independent sample of

individuals of European ancestry, for whom detailed quantitative iris pigmentation measures are

available (including measures capturing central heterochromia, defined as the presence of different

eye colours in the inner and outer portions of the iris). By incorporating additional GWAS and

post-GWAS analyses, this work aims to address the following research question: **What common**

markers are influencing eye colour variation in individuals whose self-reported phenotype

does not correspond to the expected phenotype associated with their genotype at rs12913832?

Analyzing both self-reported data and detailed quantitative measures of iris colour, we describe

polymorphisms that act to counteract the strong influence of rs12913832 on iris colour.

Methods

Study Dataset and Design

This research was conducted under the approval of the University of Toronto Research Ethics

Board (REB), adhering to Human Research Protocol #36429. The study dataset was drawn from

the Canadian Partnership for Tomorrow's Health (CanPath) biobank, Canada's largest collection

of population health data, containing genomic and phenotypic information from over 300,000

Canadians of predominantly European ancestry (13,14). Access to the data was granted through

CanPath, under application number DAO-034431 (Research Project Number 329058, 2018-10-01

start date). All analyses were conducted using de-identified data. The authors did not participate

in data collection and had no access to personally identifiable information at any stage of the

research. All participants provided written informed consent at the time of data collection by

CanPath.

The study utilized a subset of 5,481 individuals for which self-reported eye colour was available

(N_{GG}=2,724 and N_{AA+AG}=2,757), encompassing various Canadian provinces. **Fig. 2** illustrates the

provincial cohorts represented in the biobank. Among this group, 56.29% were female, and the

average age was approximately 55 years (standard error ± 0.12).

CanPath participants underwent genotyping between 2012 and 2018. Two genotyping chips were

utilized: the Axiom 2.0 UK Biobank Array (Affymetrix) (UKBB) for (N=3,212) individuals and

the Global Screening Array (GSA) 24v1+MDP (N=2,429) participants, conducted by CanPath.

The genotyping arrays analyzed between 658,296 and 813,168 SNPs. Imputation was carried out

previously on the Sanger Imputation Server using the Haplotype Reference Consortium dataset as

a reference for imputation (11,12).

Fig 2. Study cohorts and provinces participating in the Canadian Partnership for Tomorrow's Health
(CanPath). Additional details about the study and participating cohorts can be found in (14) and at canpath.ca.

Among the 2,724 participants with rs12913832:AA or AG genotypes and available self-reported

eye colour data, 512 reported eye colours other than brown or hazel. This observation suggests

that the rs12913832:AA+AG genotype alone does not fully account for dark eye pigmentation and

points to the possibility that other genetic variants may influence eye colour in individuals with

the AA+AG genotype.

To investigate, we piggybacked off the 2023 rs12913832:GG study (12) and applied the same

methods for AA+AG individuals in the CanPath sample, as well as a separate replication dataset.

Fig 3 illustrates the step-by-step computational methodology.

Fig3. **Step-by-step computational methodology**. Flow chart illustrating the computational steps used to perform a
genome-wide association study (GWAS) to identify genetic variants associated with eye colour in rs12913832:AA+AG
individuals.

Preliminary Statistics and Quality Control (QC)

Participants with rs12913832:AA+AG genotypes were extracted from CanPath arrays (UKBB and

GSA). We retained only those individuals who self-reported as having European-related ancestry

and for which self-reported eye colour data were available. Individuals who self-reported amber

eyes were excluded due to the low frequency of individuals who reported this colour category.

Pre-imputation QC was performed as described in Lona-Durazo et al (2022) (11) . Post-imputation

QC began with the removal of all variants other than bi-allelic SNPs (e.g., insertion/deletions,

multi-allelic markers, and other types of polymorphisms) using bcftools (version 1.13) (15,16).

Additionally, bcftools was also used to ensure that only SNPs were represented in the imputed

genotype files, no insertion/deletions (indels) or other types of polymorphisms, as well as to

remove markers with low imputation accuracy (`INFO>0.8`) and duplicated markers. Variants

with strong deviations from Hardy-Weinberg equilibrium (HWE) (`--hwe 1E-6`) were filtered

from the imputed files using `vcftools` (version 0.1.16.) (17). Pre-imputation and post-

imputation QCs were performed separately on each array.

An initial principal component analysis (PCA) was performed using a pruned (`--`

`indeppairwise 50 1 0.1`) set of unimputed variants from both CanPath arrays projected

onto the 1000s Genomes Project (1KG) Phase 3 samples using PLINK (version 1.9) (18,19) to

identify any outliers that did not cluster within the European superpopulation of the 1KGP based

on the first three principal components. After removal of outliers, 1,412 samples were retained

from the GSA array and 1,312 from the UKBB array (for a total of 2,724 AA/AG individuals). A

second PCA was then performed for each array using PLINK (version 1.9) (18,19) and the first 10

principal components were retained as covariates and added to the phenotype files corresponding

to each array using R studio (version 4.2.0) (20).

Genome-wide Association Study (GWAS)

A genome-wide association study (GWAS) targeting eye colour was independently conducted on

individuals with AA+AG genotypes from each array, employing both linear and logistic mixed

models via SAIGE (version 1.0.5) (21). In the linear model, eye colour is coded as follows: blue=1,

green=2, hazel=3, and brown=4. The logistic model was coded as follows: blue=1 versus non-

blue=0 (brown and hazel). The linear model used for categorical eye colours followed an additive

framework, wherein the estimated effect size corresponds linearly to the count of effect alleles.

The GWAS model included covariates such as age, sex, and the first ten principal components as

fixed effects and a Genetic Relationship Matrix (GRM) as a random effect, alongside imputed

genotype data.

Meta-analysis

A meta-analysis was carried out using the beta coefficients and standard errors from the linear

GWAS results corresponding to the two distinct CanPath arrays, along with data from each logistic

GWAS using the software Metasoft (version 2.0.1) (22). Metasoft assesses input GWAS using two

types of models, 1) fixed effects (FE), most appropriate when effect sizes are assumed to be

consistent across studies, and 2) an enhanced random effects model (RE2), which is more suitable

when there is variability or heterogeneity between datasets (23).

Metasoft computes two key indicators of heterogeneity: Cochran's Q and I^2. The Q statistic reflects

the weighted squared differences between individual study effects and the combined effect

estimate; a P-value below 0.10 typically indicates significant heterogeneity. The I^2 metric (24,25)

estimates the proportion of variation across studies that exceeds what would be expected by

chance, calculated as $I^2 = 100\% \times (Q - df)/Q$.

When I^2 exceeds 50%, this may suggest that a random effects model is more appropriate than a

fixed effects model (26,27). In the absence of heterogeneity, both models are expected to produce

similar outcomes. Metasoft also generates Bayesian posterior probabilities indicating the

likelihood that an association exists in each study included in the meta-analysis (23).

Conditional and Joint Analysis (COJO) Using GWAS Summary Statistics and Fine-Mapping

The combined summary statistics from the linear GWAS meta-analysis were subsequently

processed using the COJO method included in the GCTA suite (28). COJO is used to determine

whether genome-wide significant signals (P-value<5E-8) identified in a GWAS (or meta-analysis)

are attributable to a single variant or represent multiple independent signals. The analysis was

based on fixed-effects (FE) model P-values from the meta-analysis and incorporated weighted

average effect allele frequencies derived from both contributing datasets. A linkage disequilibrium

(LD) reference was generated from the GSA array as it had the larger sample size of the two arrays

for AA and AG individuals.

Statistical fine-mapping was performed using LDSTORE2 (v2.0) and FINEMAP (v1.4) (29),

focusing on regions highlighted by the COJO analysis. This approach helps pinpoint variants most

likely to causally influence eye colour. FINEMAP uses a Bayesian model to integrate GWAS or

meta-analysis summary statistics with SNP LD data, producing posterior probabilities for putative

causal variants.

In the first step, LDSTORE2 was used to generate LD correlation matrices from the GSA

24v1MDP array data within 1 Mb windows (±500 kb from lead SNPs). Next, these LD matrices

were input into FINEMAP along with the defined genomic windows. The analysis assumed up to

10 causal variants per region, generating 95% credible sets – groups of SNPs with the highest

posterior probability of being causal. Each set is summarized by its posterior inclusion

probabilities and a k-value indicating the most probable number of causal variants.

SNP Annotation

To annotate the genome-wide significant variants (P<5E-08) identified through meta-analysis, we

employed SNPnexus, a web-based annotation tool (https://www.snp-nexus.org/v4/)   (30–34).

Gene and variant classification were based on data from the UCSC Genome Browser and Ensembl

databases using the hg19 human genome build. If, through SNPNexus, we were able to determine

if a variant was listed in ClinVar (35), then the clinical annotation was also documented. The

potential functional impact of non-synonymous coding variants was evaluated using SIFT (36)

and PolyPhen (37) prediction scores, which provide qualitative outputs such as "probably

damaging", "possibly damaging/deleterious-low confidence", or "benign/tolerated". For non-

coding variants, we applied the CADD scoring system (38), which ranks variant deleteriousness

across the genome. A CADD score of 20 or higher suggests the variant is among the top 1% of the

most deleterious substitutions (34). Furthermore, we investigated protein-level expression of

implicated genes using STRING (39) and searched for eQTLs (expression Quantitative Trait Loci)

in GTEx (https://www.gtexportal.org/home/) (40).

Replication in Independent Sample of Individuals of European Ancestry with Quantitative

Measures of Iris Colour

The SNPs identified in the CanPath GWAS were replicated in a sample of individuals of European ancestry living in Canada, for whom detailed eye colour quantitative measures based on high-resolution iris pictures are available. The sample and the methods used to measure iris pigmentation have been described in a previous paper (41). The sample includes 549 individuals with ancestry from different regions in Europe. High-resolution iris pictures were taken using a Miles Professional Iris Camera and quantitative measures of iris colour using the CIE 1976 L*a*b* (CIELAB) colour space were obtained using a web-based application (41). The L* coordinate represents the lightness dimension and ranges from 0 to 100, with 0 being black and 100 being white. The a* and b* coordinates represent variation in colour, with negative values of a* indicating green and positive values of a* indicating red, and negative values of b* indicating blue and positive values of b* indicating yellow. Individuals with blue eye colour are characterized by negative a* and b* values, and higher L* values than those observed for individuals with non-blue eyes. However, there is considerably variation in L*, a* and b* dimensions within individuals reporting "blue" or "brown" eye colour (41). In addition to L*, a* and b* values, the amount of central heterochromia (i.e., differences in iris colour between the inner and outer portions of the iris) was measured by a colour metric (ΔE) that represents differences in the average colour of the ciliary zone (outer iris region) and the average colour of the pupillary zone Inner iris region). Two different cameras were used during the study, and camera was used as a covariate in the statistical analysis. Participants in the study also self-reported eye colour in 5 categories: 0=Light blue, Light gray or light green; 1=Blue, gray or green, 2=Hazel or light brown, 3=Dark brown, 4=Brownish black.

Genotyping was carried out with Illumina's Infinium Multi-Ethnic Global Array (MEGA) at the

Clinical Genomics Centre (Mount Sinai Hospital, Toronto, Ontario, Canada) using standard

protocols. QC steps were implemented to remove samples and markers, according to the following

criteria, Sample QC: 1) Removal of samples with missing call rates <0.9, 2) Removal of samples

that were outliers in Principal Component Analysis (PCA) plots, 3) Removal of samples with sex

discrepancies, 4) Removal of samples that were outliers for heterozygosity, and 5) Removal of

related individuals (pi-hat> 0.2). Marker QC: 1) Removal of markers with genotype call rate <0.95,

2/removal of markers with Hardy-Weinberg Es <10−6, 3) Removal of Insertion/Deletion (Indel)

markers, 4) Removal of markers with allele frequencies <0.01, 5) Removal of markers not present

in the 1000 Genomes reference panel, or that do not match on chromosome, position and alleles,

6) Removal of A/T or G/C SNPs with MAF >40% in the 1000 Genomes European reference

samples, and 7) Removal of SNPs with allele frequency differences >20% between the study

sample and the 1000 Genomes European reference sample. After performing the QC steps

described above, the samples were phased using the program SHAPEIT2 and imputed at the

Sanger Imputation Service, using the Positional Burrows-Wheeler Transform (PBWT) algorithm

(42), and the samples of the 1000 Genomes as reference haplotypes. All the markers included in

this analysis have imputation INFO scores higher than 0.9, indicating good imputation confidence.

Following the same approach used for the CanPath sample, individuals were stratified in two

categories based on *HERC2* rs12913832 genotype (GG and AA+AG). Association of the selected

markers with the quantitative traits were tested using linear regression with sex, camera, and 4

principal components as covariates. Age was not included as a covariate because there was a

narrow range in participant's age (18-35 years) and age did not have a significant effect on the

traits. Given that some quantitative traits (a*, b* and ΔE) showed non-normal distributions, an

inverse normal transformation of the residuals from a regression of a*, b* and ΔE on sex, camera

and 4 principal components was carried out, and these normalized values were used as dependent

variables in the linear regression. A linear model was also used to evaluate association of the

markers with the five self-reported eye colour categories (0=Light blue, Light gray or light green;

1=Blue, gray or green, 2=Hazel or light brown, 3=Dark brown, 4=Brownish black).

Results

Preliminary Statistics and Quality Control (QC)

After applying all QC filters, genotypes and self-reported eye colour phenotypes were retained

from a total of 2,724 AA+AG individuals (1,412 from the GSA array and 1,312 from the UKBB

array). Eye colour categories included blue, green, hazel and brown.   Of the 2,724

rs12913832:AA+AG participants, 512 did not self-report brown or hazel eyes, representing

41.16%. As was reported in 2023, of the 2,757 rs12913832:GG participants for which self-

reported eye colour data were available, 904 did not report blue eyes. **Fig 4** illustrates eye colour

frequency stratified by genotype at the rs12913832 locus. In total, 5,481 CanPath participants are

represented when considering all rs12913832 genotypes combined.

Fig 4. **Eye colour distribution among 2,757 rs12913832:GG and 2,724 rs12913832:AA+AG Canadian**
Partnership for Tomorrow's Health (CanPath) participants genotyped with the UKBB array and GSA array.
Self-reported eye colour categories retained for investigation were described as blue, green, hazel, and brown.
Individuals with missing phenotype information or who self-reported their eye colour as amber were excluded.

After QC and imputation 8,822,293 variants were retained from AA+AG individuals genotyped

with the GSA array for further analysis and 7,104,980 variants were retained from AA+AG

individuals genotyped with the UKBB array.

Genome-Wide Association Study (GWAS) and Meta-Analysis

As was performed on the GG individuals in the 2023 study, two types of GWAS were performed

on AA+AG individuals from each array using SAIGE (v1.0.5) (21). A linear mixed model was

applied where eye colour was coded as blue=1, green=2, hazel=3, and brown=4 in accordance

with an additive genetic model, as well as a logistic model comparing blue versus non-blue (hazel

and brown) eye colour. The first ten genetic principal components generated during the QC stage

were included as covariates, along with age and sex.

The summary statistics (P-values, betas, and SE) from the linear and logistic models conducted in

each CanPath array GWAS were then integrated in a meta-analysis using Metasoft (v2.0.1) (22).

The resulting Manhattan plot from the linear model is illustrated in **Fig 5** (Supplementary **Figs S1**

and **S2** present the QQ plots for each meta-analysis and the Manhattan plot from the logistic

model). Significant peaks are observed in chromosomes 6 (*IRF4*), 9 (*TYRP1*) and 15 (*OCA2*).

Fig 5. **Manhattan plot of eye colour meta-analysis based on linear model GWAS in rs12913832:AA+AG**
individuals (*N*=2,724) from the Canadian Partnership for Tomorrow's Health (CanPath) project genotyped
with the Axiom UKBB and GSA arrays. GWAS were performed with SAIGE (21). Categorical eye colours coded
as blue=1, green=2, hazel=3, and brown=4. The meta-analysis was conducted with Metasoft (22). The black horizontal
line indicates the genome-wide threshold (P<5E-8). Significant peaks are observed in chromosomes 6 (*IRF4*), 9
(*TYRP1*) and 15 (*OCA2*).

Comparing GWAS Lead SNPs from GG and AA+AG Individuals.

Table 1 illustrates the top SNPs from the meta-analysis according to the fixed effects (FE) linear

model in each significant genomic region. Several SNPs had evidence of heterogeneity among the

two studies (indicated by I^2 and Cochran's Q values in the meta-analysis), however, they all

retained genome-wide significant P-values in the random effects (RE2) model too, which considers

heterogeneity among studies. Given that heterogeneity between studies did not impact SNP

significance, the fixed effects (FE) model was prioritized for downstream analyses. Please note

that that in the 2023 publication (12) the effect alleles in the tables were incorrect, and a correction

was submitted.

Table 1. Top SNP from each significant genomic region in meta-analysis of GG stratified individuals and
AA+AG stratified individuals using a linear model (N_{GG}=2,747, N_{AA+AG}=2,724).

Marker ID	Chr	A1	A2	A1 Frequency	Gene	P-value (FE)	Beta	SE
GG Stratified Individuals								
rs35389	5	G	A	0.046	*SLC45A2*	1.18E-12	0.352	0.049
rs2733836	9	T	G	0.438	*TYRP1*	1.37E-13	0.133	0.018
rs1126809	11	A	G	0.272	*TYR*	4.58E-13	-0.179	0.019
rs12896399	14	T	G	0.401	*SLC24A4*	2.05E-35	-0.221	0.018
rs62075722	17	A	G	0.364	*TSPAN10*	6.36E-10	-0.116	0.019
AA+AG Stratified Individuals								
rs12203592	6	T	C	0.164	*IRF4*	3.13E-26	-0.318	0.030
rs7853779	9	A	G	0.389	*TYRP1*	3.12E-8	0.129	0.023
rs1800407	15	T	C	0.128	*OCA2*	4.10E-15	-0.292	0.037

Based on the categorical coding of the model (1=blue, 2=green, 3=hazel, 4=brown), a positive beta can be interpreted
as an indication of eye colour "darkening" and a negative beta can be interpreted as an indication of eye colour
"lightening" for the effect allele (A1). We highlight alleles increasing pigmentation in the rs12913832 GG (e.g. blue-
eye) background with red font and alleles decreasing pigmentation in the rs12913832 AA+AG (e.g. non blue-eye)
background in blue font.
Marker ID, reference SNP cluster ID;
Chr, chromosome;
A1, effect allele: the allele whose effects in relation to eye colour are being studied;
A2, the alternative allele;
A1 frequency, fraction of chromosomes in population that carry the effect allele;
Gene, the gene the SNP is located in, or the gene the SNP is nearest to;
Beta, per unit increase (if positive) or decrease (if negative) in the outcome;
P-value (FE), parameter of statistical significance used to determine the certainty of an association, based off
the fixed effects meta-analysis model;
SE, standard error.
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Conditional and Joint Analysis (COJO)

Results from the COJO analysis using GCTA (28) for the GG individuals were previously

described in 2023 (12), but they are summarized in **Table 2** to easily compare the different signals

coming from each genetic background.   The COJO analysis of the linear model conducted on

AA+AG individuals identified one significant genome-wide SNP driving the signals on

chromosomes 6 and 9, while two genome-wide significant SNPs were observed driving the signal

on chromosome 15. Among these, rs12203592 in *IRF4* is a well-established causal pigmentation

variant, previously associated with both skin and hair pigmentation, as well as melanoma

susceptibility (6,43,44). The SNP rs7853779 on chromosome 9, located near *TYRP1*, had prior

associations with pigmentation traits and melanoma susceptibility (1,45). Two SNPs on

chromosome 15, rs1800407 and rs4778138, both situated in or near the *OCA2* gene, have been

linked to variation in eye colour in earlier research (46–48).

Table 2. Independent signals identified by COJO (28) for genome-wide significant peaks illustrated in the meta-
analysis.

CHR	Marker ID	POS	A1	A2	Beta	SE	P-value (FE)	Joint beta	Joint SE	Joint P-value	LD (*r²*)	Nearest gene	MAF
					GG individuals								
5	rs35389	33954880	G	A	0.352	0.049	1.180E-12	0.352	0.051	4.550E-12	0	*SLC45A2*	0.046
9	rs2733836	12692252	T	G	0.133	0.018	1.370E-13	0.133	0.018	6.410E-13	0	*TYRP1*	0.438
11	rs1126809	89017961	A	G	-0.179	0.02	1.200E-19	-0.179	0.021	3.950E-18	0	*TYR*	0.272
14	rs12896399	92773663	T	G	-0.221	0.018	2.050E-35	-0.221	0.019	4.020E-30	0	*SLC24A4*	0.401
17	rs62075722	79611271	A	G	-0.116	0.019	6.360E-10	-0.116	0.019	1.390E-09	0	*TSPAN10*	0.365
					AA+AG individuals								
6	rs12203592	396321	T	C	-0.318	0.03	3.130E-26	-0.318	0.031	2.170E-24	0	*IRF4*	0.164
9	rs7853779	12612893	A	G	0.129	0.023	3.120E-08	0.129	0.024	4.340E-08	0	*TYRP1*	0.389
15	rs1800407	28230318	T	C	-0.292	0.037	4.100E-15	-0.274	0.038	8.150E-13	0.082	*OCA2*	0.128
15	rs4778138	28335820	G	A	0.179	0.027	3.040E-11	0.164	0.028	2.710E-09	0	*OCA2*	0.273

Alleles increasing pigmentation in individuals with rs12913832 GG background are highlighted in red, and alleles
decreasing pigmentation in individuals with rs12913832 AA+AG background are highlighted in blue.
Chr, chromosome;
Marker ID, Reference SNP cluster ID;
POS, Physical position (hg19/GRCh37);
A1, effect allele: the allele whose effects in relation to eye colour are being studied;
A2, alternative allele;
Beta, per unit increase (if positive) or decrease (if negative) in the outcome;
SE, Standard error;
P-value (FE), P-value, parameter of statistical significance used to determine the certainty of an association
based off the fixed effects meta-analysis model;

 Joint beta, SE and **P-value**, effect size, standard error and P-value from a joint analysis of all the selected
SNPs;
LD(r²), Linkage disequilibrium correlation between the SNP i and SNP i + 1 for the SNPs on the list;
Nearest gene, the gene the SNP is located in, or the gene the SNP is nearest to;
MAF, minor allele frequency, fraction of chromosomes in population that carry the minor allele (frequency
<0.5).

Fine-mapping

Fine-mapping results are represented in **Table 3**. Only SNPs with $\log_{10}$ of Bayes Factor ($\log_{10}BF$)

greater than 2 have been included in the table. For some regions (*TYRP1, TYR, SLC24A4* and

TSPAN10 for the GG stratified individuals), the *k* value (most likely number of causal SNPs) is

greater than one, however, only SNPs within one credible set surpassed a $\log_{10}BF>2$. Thus,

although FINEMAP suggests the presence of multiple signals in some instances, the markers in

the additional credible sets did not always have sufficiently high $\log_{10}BF$, so we focused on the

markers that surpassed the thresholds for "considerable evidence" as recommended by the

FINEMAP documentation (29).

For the GG stratified individuals, the markers with the top PIP in the FINEMAP analysis coincide

with the lead SNPs identified in the COJO analysis: *SLC45A2* rs35389, *TYRP1* rs2733836, *TYR*

rs1126809, *SLC24A4* rs12896399 (there is another marker with the same PIP in this region,

rs12896471), and *TSPAN10* rs62075722. The non-synonymous variant rs1126809 is highlighted

as the causal variant in the *TYR* region, with PIP=0.960. All the other variants have PIP values

lower than 0.5.

For the AA+AG stratified individuals, the FINEMAP analysis identifies a single credible set in

the *IRF4* and *TYRP1* regions. The marker rs12203592 is highlighted as the putative causal SNP

within the *IRF4* region, with PIP=1.00. In contrast, there are several markers in high LD in the

credible set of the *TYRP1* region, with PIP values<0.10. The top SNPs identified by FINEMAP

coincide with the lead signals in the COJO analysis. Finally, the FINEMAP analysis identifies 3

independent credible sets in the *OCA2/HERC2* region. In the first credible set, FINEMAP

identifies rs1800407 as the causal variant, with PIP=0.997. In the second credible set, FINEMAP

highlights two SNPs: rs7164220 (PIP=0.472) and rs4778138 (PIP=0.341). The latter was the lead

SNP identified in the COJO analysis. The variant with the top PIP in the third credible set is

rs79380392 (PIP=0.135), which is located within the *HERC2* gene. In contrast to FINEMAP,

which points to three independent causal sets, COJO only identified two independent regions in

the *OCA2/HERC2* region. **Fig S3** illustrates LD matrices for SNPs in the *TYRP1* region and

HERC2/OCA2 region. **Fig S4** illustrates examples of regional plots for signals represented in

Table 3.

Table 3. Fine-mapping results. Fine-mapping was performed using 1Mb windows (500 kb on each side of
significant SNP).

Marker ID	Chr	A1	A2	Nearest gene	Beta	SE	P-value	MAF	PIP	*k*
GG individuals										
rs35389	5	**G**	A	*SLC45A2*	0.352	0.049	1.180E-12	0.046	0.408	
rs35395	5	T	**C**	*SLC45A2*	-0.357	0.050	1.475E-12	0.034	0.335	
rs16891982	5	C	**G**	*SLC45A2*	-0.328	0.047	3.481E-12	0.038	0.155	1
rs35407	5	A	**G**	*SLC45A2*	-0.345	0.051	9.106E-12	0.034	0.066	
rs2733836	9	**T**	G	*TYRP1*	0.133	0.018	1.368E-13	0.438	0.068	2
rs1126809	11	A	**G**	*TYR*	-0.179	0.020	1.196E-19	0.272	0.960	2
rs12896399	14	T	**G**	*SLC24A4*	-0.221	0.018	2.052E-35	0.401	0.154	
rs12896471	14	C	**G**	*SLC24A4*	-0.221	0.018	2.059E-35	0.425	0.154	
rs56137766	14	T	**A**	*SLC24A4*	-0.222	0.018	2.089E-35	0.430	0.136	
rs35983729	14	T	**G**	*SLC24A4*	-0.223	0.018	2.205E-35	0.432	0.128	2
rs4144266	14	A	**G**	*SLC24A4*	-0.223	0.018	2.367E-35	0.432	0.119	
rs4904866	14	C	**T**	*SLC24A4*	-0.221	0.018	3.456E-35	0.425	0.089	
rs746586	14	C	**T**	*SLC24A4*	-0.221	0.018	3.994E-35	0.426	0.075	
rs12883151	14	A	**G**	*SLC24A4*	-0.222	0.018	3.960E-35	0.432	0.070	
rs62075722	17	A	**G**	*TSPAN10*	-0.116	0.019	6.357E-10	0.365	0.174	
rs6420484	17	A	**G**	*TSPAN10*	-0.116	0.019	6.602E-10	0.366	0.169	2
rs34635363	17	**G**	A	*NPLOC4*	0.114	0.019	1.528E-09	0.364	0.079	
AA+AG individuals										

rs12203592	6	T	C	*IRF4*	-0.318	0.030	3.134E-26	0.164	1.000	1
rs7853779	9	A	G	*TYRP1*	0.129	0.023	3.123E-08	0.389	0.074	
rs10960723	9	C	A	*TYRP1*	-0.128	0.023	3.674E-08	0.391	0.064	
rs713596	9	A	G	*TYRP1*	-0.128	0.023	3.749E-08	0.391	0.063	1
rs2762457	9	G	T	*TYRP1*	-0.126	0.023	4.855E-08	0.402	0.050	
rs2762460	9	T	C	*TYRP1*	-0.125	0.023	4.975E-08	0.409	0.049	
rs2733831	9	A	G	*TYRP1*	-0.125	0.023	5.343E-08	0.408	0.046	
rs1800407	15	T	C	*OCA2*	-0.292	0.037	4.100E-15	0.128	0.997	
rs7164220	15	C	A	*OCA2*	-0.179	0.030	1.545E-09	0.220	0.472	3
rs4778138	15	G	A	*OCA2*	0.179	0.027	3.040E-11	0.273	0.341	
rs79380392	15	C	T	*HERC2*	0.249	0.055	5.354E-06	0.049	0.135	

SNPs with a log10 of Bayes Factor greater than 2 are recorded in the table. Alleles increasing pigmentation in
individuals with rs12913832 GG background are highlighted in red, and alleles decreasing pigmentation in individuals
with rs12913832 AA+AG background are highlighted in blue. A thick border indicates separation of chromosomal
fine-mapped regions, a thin border indicates separation of credible sets (the collection of variants that together have a
95% probability of containing the true causal SNP).
Marker ID, reference SNP cluster ID;
Chr, chromosome;
A1, effect allele: the allele whose effects in relation to eye colour are being studied;
A2, alternative allele;
Nearest gene, the gene the SNP is located in, or the gene the SNP is nearest to;
Beta, per unit increase (if positive) or decrease (if negative) in the outcome
SE, standard error;
P-value, parameter of statistical significance used to determine the certainty of an association;
MAF, minor allele frequency;
PIP, posterior inclusion probability;
k, most likely number of causal SNPs.

SNP Annotations

Beyond the rs12913832 genotype, acting as a genetic background in this study, three other SNPs

within *OCA2* and one within *HERC2* were nominated as candidate causal loci in AA+AG stratified

individuals. One of them is a non-synonymous SNP (rs1800407) and three are intronic variants

(rs4778138, rs7164220, rs79380392). These results are similar to the conditional analysis with

GCTA-COJO, where the program picked up multiple independent signals coming from this region.

We annotated the putative regulatory function of the fine-mapped SNPs with a $\log_{10}$ of Bayes

Factor greater than 2 using diverse databases (e.g. ENCODE, Roadmap Epigenomics Project)

encompassed within SNPnexus (30–34). A summary of the output can be found in Supplementary

Table S1. All SNPs in the table were identified as either overlapping with a protein-coding gene

or were located near a protein coding gene by SNPNexus (30–34). Annotation details for the lead

SNPs identified by COJO are described in **Table 4** below.

Table 4. Annotation summary of lead SNPs.

GG Stratified				
SNP	**Overlapped or nearest gene**	**Protein Consequence**	**Eye Colour Darkening Allele**	**Annotation**
rs35389	*SLC45A2*, overlapped	Protein coding; Intronic	**G**	eQTL in Thyroid (P=8.3E-5) (GTEx database). Low CADD PHRED score of 0.530. Confirmed associations with skin pigmentation, eye colour and hair colour according to GAD. Classified as Benign by ClinVar.
rs2733836	*TYRP1*, overlapped	Protein coding; Intronic	**T**	eQTL for many brain tissues (GTEx database). Low CADD score (2.549). No confirmed associations (unconfirmed association with Type 2 Diabetes).
rs1126809	*TYR*, overlapped	Protein coding; Non-synonymous	**G**	eQTL in Skin – Sun Exposed (Lower leg) (P=2.35E-5) and Testis (P=1.5E-4) (GTEx database). Reported as "Deleterious" by SIFT (Score=0.30) and "Probably Damaging" by PolyPhen (Score=0.994). Confirmed associations with multiple phenotypes including many pigmentation related phenotypes such as type IB (yellow) oculocutaneous albinism and type I-A (tyrosinase negative) oculocutaneous albinism according to GAD. Previously reported as significant in GWAS of melanoma, squamous cell carcinoma, optic disc size, basal cell carcinoma, hair colour, eye colour, sunburn, tanning response and vitiligo based on SNPnexus' GWAS catalogue. This SNP has conflicting interpretations of pathogenicity in ClinVar. Many reports have associated the variant with conditions like albinism and general abnormalities in pigmentation-related metabolic processes (49–51).
rs12896399	*SLC24A4*, nearest downstream gene	Protein coding	**G**	eQTL in Skin – Not Sun Exposed (Suprapubic) (P=1.1E-8), Skin – Sun Exposed (Lower leg) (P=5.0E-6), Whole blood (P=2.1E-5) and Muscle – Skeletal (P=4.1E-5) (GTEx database). CADD PHRED score of 5.795. Previously reported as significant in GWAS of hair colour, hair morphology traits and eye colour (blue vs. green eyes) based on SNPnexus' GWAS catalogue. The G allele increases expression and is associated with darker skin. This means that increased expression of this gene may counteract the effects of lower *OCA2* expression in GG individuals.
rs62075722	*TSPAN10*, overlapped	Non-coding, intronic	**G**	eQTL in Skin – Not Sun Exposed (Suprapubic) (P=4.1E-18) and Skin – Sun Exposed (Lower leg) (P=6.9E-16) (GTEx database). CADD PHRED score of 6.286. Confirmed associations with coronary artery disease, Parkinson's disease, and eye colour. Previously reported as significant in GWAS of brown vs. black hair and corneal astigmatism based on SNPnexus' GWAS catalogue. A

SNP	Overlapped or nearest gene	Protein Consequence	Eye Colour Lightening Allele	Annotation
				G allele is associated with increased expression and darker iris colour.
AA+AG Stratified				
rs12203592	*IRF4*, overlapped	Protein coding; Intronic	T	eQTL in Skin – Sun exposed (Lower leg) (P=6.5E-7) and many other tissues in GTEx. CADD PHRED score of 11.780. Confirmed associations with hair colour and eye colour in GAD. Previously reported as significant in GWAS of facial pigmentation, squamous cell carcinoma, tanning, sunburns, non-melanoma skin cancer, hir colour, eye colour and freckling based on SNPnexus' GWAS catalogue. Marked as "Affects" by ClinVar, reaffirming its influence on pigmentation phenotypes. The T allele decreases expression of *IRF4* and is associated with lighter iris colour. This means that lower expression of *IRF4* counteracts the effect of AA+AG, promoting lighter eye colours.
rs7853779	*TYRP1*, downstream	Protein coding	G	CADD PHRED score of 7.814. No confirmed associations in GAD, SNPnexus' GWAS catalogue or ClinVar.
rs1800407	*OCA2*, overlapped	Protein coding; Non-synonymous	T	Reported as "Deleterious" by SIFT (Score=0.20) and "Probably Damaging" by PolyPhen (Score=0.986). Confirmed associations with hair colour, eye colour Parkinson's disease and socioeconomic factors in GAD. Previously reported as significant in GWAS of squamous cell carcinoma, cataracts, skin pigmentation, eye colour, macular thickness and tanning response based on SNPnexus' GWAS catalogue. Classified as benign or likely benign in ClinVar, it is related to both oculocutaneous albinism, pigmentation variation and melanoma susceptibility. Variant has a haploinsufficiency (HI) score of 30, and homozygous mutations lead to oculocutaneous albinism type 2 (4,52,53).
rs4778138	*OCA2*, overlapped	Protein coding; Intronic	A	eQTL in Whole Blood (P=5.8E-20), Nerve-Tibial (P=2.9E-7) and Artery-Aorta (P=8.2E-5) (GTEx database). CADD PHRED score of 1.982. Confirmed associations with hair colour, eye colour, Parkinson's disease and socioeconomic factors in GAD. Previously reported as significant in GAS of squamous cell carcinoma and melanoma.

SNPs were annotated using SNPnexus (https://www.snp-nexus.org/v4/) (30–34) and GTEx (https://gtexportal.org/home/) (40).
Note that GTEx does not have iridial tissue information, so focus was placed on skin tissue information (sun exposed and not-
exposed). Integrated in SNPnexus are additional annotation tools. We utilized CADD PHRED scores (38), SIFT (36) and
PolyPhen (37) to assess the deleteriousness of a variant, GAD (53) and ClinVar (35) reports to learn about associated diseases
and other phenotypes.
GTEx, Genotype-Tissue Expression Project;
eQTL, expression Quantitative Trait Locus;
sQTL, splicing Quantitative Trait Locus;
CADD PHRED, Combined Annotation Dependent Depletion scaled score (represents the probability of a variant
being deleterious relative to all other possible variants);
SIFT, Sorting Intolerant from Tolerant, predicts whether an amino acid substitution affects protein function based on
sequence homology (applied to naturally occurring nonsynonymous polymorphisms and laboratory induced missence
mutations);

PolyPhen, Polymorphism Phenotyping prediction tool used to asses the potential impact of an amino acid substitution
on a protein's structure and function
GAD, Genetic Association Database;
ClinVar, public archive of reports of human variations classified for diseases and drug responses, with supporting
evidence.

Supplementary **Fig S5** and **Table S2** illustrate the STRING network and summarize the functional

enrichments within the networks. There are slight differences in the biological processes enriched for

genes which were associated with GG individuals versus AA+AG individuals. In GG individuals,

genes can be seen enriched for melanin biosynthetic processes and developmental pigmentation, while

in AA+AG individuals, genes were enriched in melanosome membrane. Interestingly, under the

"Human Phenotype (Monarch)" STRING domain, the genes associated with the AA+AG, or brown

eye background, the phenotype with the strongest signal (FDR=1.96E-5) was blue irides, or a markedly

blue colouration of the iris.

Replication in European ancestry cohort with quantitative iris pigmentation measures

Replication results for the six different analyses are reported in Supplementary **Table S3**

(Supplementary **Fig S6** illustrates the distributions of rs12913832 genotype across self-reported

eye colour category for this replication cohort). Of the five independent signals from GG

individuals, three markers survived Bonferroni correction in at least one quantitative measure (*TYR*

rs1126809 in the L and b* dimensions, *SLC24A4* rs12896399 in the b* dimension, and *TSPAN10*

rs62075722 in the b* dimension). Of the four independent signals from AA+AG individuals, three

markers passed Bonferroni correction in multiple dimensions (*IRF4* rs12203592 in the L, a*, b*

and ΔE dimensions, *OCA2* rs1800407 in the L and a* dimensions, and *OCA2* rs4778138 in the L

and a* dimensions). Two markers demonstrated nominal significance in other quantitative

measurements (*SLC24A4* rs12896399 for the GG stratified sample and *TYRP1* rs7853779 for the

AA+AG stratified sample,). The effects appear strongest in the b* dimension (Blue [-] to yellow

[+]) for GG stratified individuals and in the L dimension (Lightness dimension: 0 (black) to 100

(white)) for AA+AG stratified individuals. We also conducted a linear regression using the self-

reported eye colour categories in the independent replication dataset, and the results were

concordant with the quantitative measures. In GG stratified individuals, *TYR* rs1126809 was

nominally significant, and in AA+AG stratified individuals, all markers, with the exception of

TYRP1 rs7853779, survived Bonferroni correction. Importantly, rs12913832 was also assessed in

the replication dataset in AA+AG stratified individuals. This marker illustrated significance in the

L, a* and ΔE dimensions, as well as the linear model – indicative that eye colour is dosage

dependent with respect to genotype at rs12913832. In particular, rs12913832 is very strongly

associated with ΔE (p=2.97E-07), suggesting that having one copy of the G allele increases the

likelihood of central heterochromia (e.g. differences in iris colour between the inner and outer

rings of the pupil).

Discussion

This study expands upon previous work investigating genetic modifiers of iris pigmentation by

analyzing individuals whose self-reported eye colour does not align with the expected phenotype

based on their *HERC2* rs12913832 genotype. Specifically, we focused on those with a

homozygous derived genotype (rs12913832:GG), which typically predicts blue eyes, and

heterozygous or homozygous ancestral genotypes (rs12913832:AG or AA), which are more often

associated with brown eyes. Despite the heavy reliance on this single SNP in forensic and

anthropological models, exceptions in phenotype suggest additional modifiers. Our genome-wide

approach successfully identified several SNPs that modify eye colour in these "non-concordant"

individuals and replicated key findings using high-resolution quantitative iris colour data in an

independent European sample.

Through our GWAS meta-analyses and fine-mapping analyses we identified loci in known

pigmentation-associated regions including *SLC45A2*, *TYRP1*, *TYR*, *SLC24A4*, and *TSPAN10* in

the GG group (where blue eyes is expected) and *IRF4*, *TYRP1*, and *OCA2* in the AA+AG group

(where brown eyes is expected).

Genotype-stratified Associations with Eye Colour in Recent Studies

In 2020, Meyer and colleagues (10) conducted a candidate gene study investigating the association

between brown eye colour in rs12913832:GG individuals and SNPs in previously known

pigmentation genes (*TYR*, *TYRP1* and *SLC24A4*). The group identified four variants to be the most

promising candidates for the explanation of the genotype-phenotype discordances observed in GG

individuals: *TYRP1* rs35866166:C, *TYRP1* rs62538956:C, *SLC24A4* rs1289469:C, and *TYR*

rs1126809:G. Of these four markers, three were replicated in our fine-map analyses: *TYRP1*

rs35866166:C, *TYRP1* rs62538956:C and *TYR* rs1126809:G. The *TYR* rs1126809 SNP is

represented in **Table 3** and has a high PIP of 0.96, strongly suggesting it is the causal variant, with

the G allele strongly associated with darker iris pigmentation in rs12913832:GG individuals. The

TYRP1 rs35866166 and *TYRP1* rs62538956 markers are present in our 95% credible set, however,

they each had very low PIP values (0.001 and 0.003 respectively) and our fine-mapping analysis

prioritize a different variant within this region (rs2733836). That being said, we did find that the

effect direction was the same in our study, with the rs35866166:C and rs62538956:C alleles

corresponding to positive beta values (in the direction of eye colour darkening with our linear

model). The authors did note that none of the variants identified in the candidate gene study

showed statistically significant associations with eye colour after traditional multiple testing

correction, but in our GWAS we found rs1126809 and rs62538956 to surpass genome-wide

significance (P=1.200E-19 and P=5.846E-9 respectively), and rs35866166 to be nominally

significant (P=2.950E-6). Three years later, in 2023, Salvo and colleagues (54) performed an

additional candidate gene study investigating the association between variants in the

HERC2/OCA2 region and blue eye colour in rs12913832:AA+AG individuals. They identified

five novel variants to be the most promising candidates for explanation of blue eye colour in

individuals with a brown-eye background: rs74409036:A (*OCA2*), rs78544415:T (*OCA2*),

rs72714116:T (*OCA2*), rs191109490:C (*HERC2*) and rs551217952:C (*HERC2*). None of these

SNPs are genome-wide significant in our analysis, and our analysis points to different markers

within this region. We describe in more detail the main regions and polymorphisms identified in

our stratified analysis below.

Eye Colour Darkening in GG Stratified Individuals

SLC24A4 and **SLC45A2**: *SLC24A4* encodes a sodium/calcium/potassium exchange protein

(GeneCards, 2025). Variants in this gene have been previously implicated in skin, hair and eye

pigmentation, although its precise function in melanogenesis remains unclear (55,56). Our lead

SNP in this region is rs12896399, which is situated approximately 15,000 base pairs upstream of

the *SLC24A4* transcription start site and has been included in established eye colour prediction

tools, including IrisPlex (57–59) and Snipper 3.5 (60). The T allele of rs12896399 is associated

with blue eye colour (59), a relationship also observed in this study. In our preliminary 2023

investigation of rs12913832:GG individuals (12), we observed strong linkage disequilibrium

among SNPs within the *SLC24A4* credible set, with pairwise comparisons yielding r^2 values

greater than 0.9. eQTL data in GTEx indicates that a G allele at rs12896399 is associated with

increased *SLC24A4* expression, suggesting that higher expression of this transporter is able to

compensate for the reduced expression of *OCA2* in rs12913832:GG individuals. Our results point

to rs12896399:G, or closely linked variants, as strong candidates for darker eye colours in GG

individuals.

SLC45A2 encodes another transmembrane transporter involved in melanin biosynthesis. This gene

is expressed in melanoma cell lines and is implicated in oculocutaneous albinism type 4. Its

polymorphisms are most commonly associated with variation in skin and hair pigmentation (61).

The *SLC45A2* variant rs16891982 is included in both the IrisPlex (57–59) and The Snipper 3.5

(60) models, and appeared in our COJO and fine-mapping analyses of rs12913832:GG individuals.

However, rs16891982 was not the top candidate within the credible set (PIP=0.155). Instead,

rs35389 emerged as the most likely causal variant, with a PIP of 0.408. These two SNPs are in

linkage disequilibrium, as confirmed by LDpair (D'=1.0, R^2=0.6964, χ^2=700.577, P<0.001 with

EUR reference) (Machiela & Chanock, 2015). While rs35389 has not been previously associated

with eye colour, it has been evaluated as an ancestry informative marker (62) and has shown

significant associations with methylation patterns in the context of population stratification (63).

TYRP1: Notably, *TYRP1* emerged as a consistent locus across both genetic backgrounds. *TYRP1*

encodes the tyrosinase-related protein 1, which is a member of the tyrosinase family and is

involved in the synthesis of eumelanin (64). Specifically, it is responsible for the stabilization of

tyrosinase, which is an important enzyme in the initiation of melanin synthesis (65) and contributes

to melanosomal structure and maturation (66). *TYRP1* is primarily expressed in melanocytes and

the retinal pigment epithelium (RPE) (67). Only one marker, rs2733836, illustrated association in

the GG background. There are no *TYRP1* variants currently included in eye colour prediction tools

specifically, although rs683 appears for the prediction of both hair and eye colour (57–59). In

2009, Liu et al. (55) compared predictive power of a six-SNP model (*HERC2* rs12913832, *OCA2*

rs1800407, *SLC24A4* rs12896399, *SLC45A2* rs16891982, *TYR* rs1393350, and *IRF4*

rs122203592) with a 15 SNP model where two *TYRP1* variants were included (rs1408799 and

rs683) and found that the inclusion of the additional SNPs provided very minimal additive effects

and did not significantly improve predictive accuracy. rs2733836 (associated in this study),

rs1408799 and rs683 are in linkage disequilibrium (D'>0.7) (Supplementary **Fig S3**), yet

rs2733836 has not previously been associated with eye colour. Not much is known of the variant's

consequence apart from the fact that it is located 2 kilobases upstream of *TYRP1*; the variant itself

is not a protein coding variant and is therefore not expressed, but the gene is, and can be found

most highly expressed in skin (30–34).

TYR: The primary *TYR* variant identified in our study was rs1126809, a coding SNP in exon 4 of

TYR, and most likely causal variant in this region (PIP=0.960). The alternative allele (A) induces

a missense mutation that renders the tyrosinase enzyme thermosensitive and less efficient,

reducing melanin synthesis (68,69). Given that *TYR* is the rate-limiting enzyme in melanogenesis

(70,71), individuals carrying the A allele are expected to exhibit lighter pigmentation. Our results

indicate that the presence of the rs1126809 G allele can oppose/counteract the effects of reduced

OCA2 expression driven by the rs12913832:GG genotype

There is currently only one *TYR* variant included in eye colour phenotyping models: rs1393350

(57–60,72,72–76). rs1393350 and rs1126809 are in near-perfect linkage disequilibrium (D'=1.0,

R²=0.9532, χ^2=958.8784, P<0.001, with EUR reference), as reported by LDpair (77). Our findings

echo that of Meyer et al., 2020 (10) and raise the possibility that rs1126809 may be a more

biologically informative marker than rs1393350. Both variants reached genome-wide significance

in our GWAS and were identified in fine-mapping analyses; however, rs1393350 demonstrated a

notably low posterior inclusion probability (PIP=0.017) despite its strong P-value (P=5.81E-18),

suggesting it may not be the causal variant but rather tags a signal in high LD. Importantly, while

rs1393350 is known to associate with iris colour (with the A allele enriched in green eyes and the

G allele in blue) (56), its moderating function remains poorly defined. In contrast, rs1126809 is a

functional missense variant that reduces tyrosinase enzymatic activity through thermosensitivity,

directly impacting melanin production.

TSPAN10: *TSPAN10*, or tetraspanin-10, has recently been associated with both quantitative and

categorical measurements of eye colour (1,63). It is known to be expressed in the eye, including

iris, ciliary body and RPE (78), however, its specific contribution to pigmentation variation has

yet to be fully identified. There are no *TSPAN10* (or *NPLOC4*) variants currently included in any

DNA phenotyping tools. In our analysis, rs62075722 illustrated the highest PIP (0.174) of the

credible set and is an eQTL in GTEx (40), although it should be noted that a PIP of 0.174 is low

and high LD among variants in the credible set makes it difficult to identify a singular putative

causal variant. The expression data indicates that a G allele at this locus is associated with

increased expression and a darker iris colour in GG stratified individuals. This is concordant with

the other two variants identified by FINEMAP, rs6420284 and rs34635363, which are also

significantly expressed in skin tissue in GTEx. Both the rs6420284:G allele and the rs34635363:G

allele are associated with increased expression and darker iris pigmentation.

Eye Colour Lightening in AA+AG Stratified Individuals

IRF4: The transcription factor *IRF4* has been shown to enhance *TYR* gene expression through

interaction with MITF, a key regulator in melanocyte development (6). One specific *IRF4* variant,

rs12203592, has been demonstrated to influence *IRF4* expression in melanocytes (79), and is

included in the IrisPlex prediction panel (57,58). In the present study, we detected a significant

association between eye colour and rs12203592 in individuals carrying the AA or AG genotypes

at rs12913832. Prior research also suggests that interactions between *IRF4* and *OCA2* may

modulate eye pigmentation. For instance, both Laino et al. (80) and Lona-Durazo et al. (11)

reported that individuals with rs12913832 AA or AG genotypes – generally linked to darker eye

colour – can still present with blue eyes if they carry one or two copies of the rs12203592:T allele,

which has been associated with lighter pigmentation (80). The T allele at rs12203592 leads to

decreased *IRF4* expression (40). Phenotypically, carriers of the T allele tend to exhibit lighter eye

colour (e.g., blue or green), particularly among those with the rs12913832:AA or AG genotypes,

where the effect of the T allele can partially override the darker eye phenotype typically associated

with these genotypes.

TYRP1: The observation of *TYRP1* variants (rs7853779, rs10960723, rs713596, rs276257,

rs2762460, rs2733831) in the AA+AG genetic background is both interesting, and more difficult

to interpret as there are no other studies, candidate gene or otherwise, which have investigated the

role of *TYRP1* SNPs in individuals with an rs12913832:AA+AG genetic background. Of the six

TYRP1 variants with a $\log10_{BF} > 2$, only three have published associations in scientific literature.

rs2762460 has been previously associated with breast cancer (81), and epidemiological studies

have proposed that melanoma and breast cancer may share genetic factors (82,83), positioning

TYRP1 as a pleiotropic candidate gene. rs2762457 was used as a tag SNP in a candidate gene study

of individuals with a blue eye genetic background, however, it was determined not to be associated

with eye colour, but rather in strong LD with other SNPs associated with eye colour (10). Of note,

our study indicates that variants within this gene, which is known to play an important role in

melanin synthesis, may be responsible for darker eye colours in rs12913832:GG individuals and

lighter eye colours in rs12913832:AA+AG individuals.

It is also interesting to note that the *TYRP1* variant identified in the GG genetic background is in

high LD with the *TYRP1* variants identified in the AA+AG genetic background (all pairwise

relationships have a D'>0.9 and R^2>0.69, as confirmed by LDmatrix (with EUR reference) (77)).

Variants in this region may make good candidates for SNP-SNP interaction tests in future studies,

however, it would be important to consider that formal tests of epistatic interactions often suffer

from statistical power and multiple hypothesis testing limitations.

HERC2/OCA2: *OCA2* encodes the P protein, a critical transmembrane component of the

melanosome that plays a vital role in melanin biosynthesis and pigmentation (84). In addition to

transporting melanosomal content, the P protein helps regulate melanosomal pH, which in turn

influences both the amount and quality of melanin deposited in melanocytes (5,7). Dysfunction or

loss of this protein, observed in both mice and humans, results in oculocutaneous albinism,

underscoring its essential function in pigmentation pathways (7,85). Regulation of *OCA2*

expression occurs via a long-range enhancer located within intron 86 of the neighbouring *HERC2*

gene. Reduced expression of *OCA2*, mediated by variation in this enhancer, disrupts melanosome

maturation and limits the effective processing and transport of melanin. This results in lighter

pigmentation (such as that observed in blue eyes) due to compromised P protein availability (5).

In our study, multiple *HERC2/OCA2* variants were observed to be associated with eye colour

variation in AA+AG individuals. By applying a Bayesian fine-map approach we found that the

most likely number of causal variants for this genomic region was three, for which three candidates

were located in *OCA2* (rs1800407, rs4778138/rs7164220) and one candidate in *HERC2*

(rs79380392). rs1800407 is a missense variant currently included in forensic phenotyping tools

(57–59). The rs1800407:T allele encodes the amino acid glutamate and having just one copy of

the T allele is associated with an increased probability of having green and hazel eye colours in

European populations (52,53). Given this, the presence of the T allele at rs1800407 in our study

may help explain the elevated frequency of self-reported green and hazel eye colours by

individuals with rs12913832:AA or AG genotypes (**Fig 4**). With a PIP of 0.997, and the only SNP

with a $\log_{10}BF>2$ in this credible set, rs1800407 appears to be the first of three causal SNPs in this

region. In the second credible set (rs7164220 and rs4778138), it is difficult to parse which variant

is the causal variant. rs4778138 was identified as an independent signal in the COJO analysis,

however, rs7164220 possesses the slightly higher PIP in the FINEMAP analysis (PIP=0.472).

rs4778138 has been previously identified as being involved in a three-SNP haplotype system

(rs4778138, rs4778241, and rs7495174) associated with blue eyes (86). In addition to rs4778138,

we also observed rs7495174 from the blue eye haplotype in our study; although it was only

nominally significant (PIP=004, P=1.87E-7), it may help explain the instances of blue eyes in the

AA+AG cohort. Finally, rs79380392 appears to be the causal SNP in the third credible set as the

only SNP with a $\log_{10}BF>2$. It is important to note that, although highlighted by FINEMAP, this

SNP did not reach genome-wide significance in our study (Table 3), so this putative causal variant

must be interpreted with caution.

Replication

The replication analyses provided strong confirmation for the majority of associations identified

in the discovery cohort. Among GG stratified individuals, three out of five independent signals

(*TYR* rs1126809, *SLC24A4* rs12896399 and *TSPAN10* rs62075722) replicated with Bonferroni-

corrected significance in at least one quantitative dimension. These associations were most

commonly observed in the b* dimension (blue-yellow). Among AA+AG individuals, three of four

loci (*IRF4* rs12203592, *OCA2* rs1800407, and *OCA2* rs4778138) survived Bonferroni correction

across multiple quantitative measures. These associations were most commonly observed in the

L* (lightness) dimension, consistent with the decrease in eumelanin concentration that

accompanies a transition from brown eye colours to intermediate and blue eye colours. The results

from the linear model using self-reported eye colour categories were also quite consistent with the

CanPath sample results, with the replicated variants showing the same effect directions as those

observed in linear CanPath analysis.

Importantly, rs12913832 was included in the AA+AG replication because there is variation in this

group (1 versus 2 copies of the ancestral allele). The results indicate that having one derived allele

(G) increases the likelihood of lighter eyes, in other words, the marker does not behave in an

entirely dominant fashion. In fact, the effect of this marker is particularly strong for ΔE, indicating

that heterozygous individuals are more likely to have central heterochromia. This observation has

been reported in previous studies, where each copy of the ancestral allele (A) decreased the colour

difference between the ciliary and pupillary zones (41). The same study also notes that rs12913832

may have both dominant/recessive and additive effects on iris colour variation, depending on

which dimensions of colour space are being measured (41).

Conclusion

A limitation of our study is the reliance on self-reported eye colour categories provided by

participants in the CanPath cohorts. While practical, such categorical data may not reflect the full

spectrum of iris pigmentation as accurately as quantitative measurement approaches (41,63,87).

Additionally, participants may report their eye colour inaccurately or may not know which

category to report (consider an individual with central heterochromia where the outer portion of

their eye appears blue but there is a brown ring around the pupil). In this regard, we were fortunate

to be able to replicate many of our signals in an independent European dataset for which

quantitative iris colour measurements and a quantitative measurement of heterochromia were

available. It is also important to note that the main goal of our project was to identify potential

modifiers of the effect of rs12913832 polymorphism on eye colour, and not to evaluate the

improvement on eye colour prediction based on the addition of these regions to current eye colour

prediction panels, which ideally would require having access to additional large datasets with well-

curated eye colour and genotype data. Finally, we fully acknowledge that solely focusing on

individuals with European genetic ancestry limits the conclusions that can be drawn. Future studies

would benefit from both increased size and diversity of study samples. Not only would it be

interesting to see replication of results in other genetic ancestry groups, but an expansion of sample

size and diversity may also draw attention to other genomic regions that are involved in the large

spectrum of observed eye colour variation. In particular, recently admixed populations would

provide an excellent opportunity to study in more detail gene-gene interactions shaping eye colour

variation. The presence of uncharacterized alleles and epistatic effects may explain in part why

forensic prediction tools for pigmentary traits have poorer performance in recently admixed

populations than in European groups (88–90).

Together, these findings emphasize that the genetic modifiers of eye colour differ substantially

between individuals with the rs12913832:GG and rs12913832:AA+AG backgrounds. When

combined with expression information from GTEx, a clear pattern emerges from our GWAS data.

For those with GG genotypes, who typically present with blue eyes, variants in core melanin

synthesis genes (*TYR*, *TYRP1*), transporter proteins located on melanosomes (*SLC24A4*,

SLC45A2), and potentially *TSPAN10* appear to contribute to darker-than-expected eye colour.

Specifically, we observe that rs1126809:G (*TYR*), rs12896399:G (*SLC24A4*), and rs62075722:G

(*TSPAN10*) all lead to increased expression of their respective genes (as reported in GTEx), and

demonstrated eye colour darkening effects in GG stratified individuals. Conversely, among

AA+AG individuals, lighter iris colours appear to result from a) other markers disrupting the

expression or function of the *OCA2* gene and its product, independently of the *HERC2* rs12913832

polymorphism, and b) modulation by *IRF4*, which interacts with MITF and TFAP2α (transcription

factor AP2α) to regulate *TYR* expression (91). For the two lead SNPs which had eQTLs in GTEx,

we see that the rs12203592:T (*IRF4*) and rs1800407:T allele (*OCA2*) decrease expression in their

respective genes, contributing to eye colour lightening in the AA+AG stratified individuals.

Interestingly, *TYRP1* emerges in both genotype stratified groups. It seems that *TYRP1* may work

both to counteract low expression *OCA2* levels in individuals with GG genotypes, and also

decrease melanin levels in individuals with AA+AG genotypes.

In summary, this study advances our understanding of eye colour variability by identifying genetic

variants that may modulate pigmentation in individuals with discordant *HERC2* rs12913832

genotypes. The identification of shared and background-specific loci underscores the polygenic

and regulatory complexity of iris pigmentation. By integrating categorical self-report data with

quantitative phenotype replication, our findings provide new insights into how moderating genetic

factors contribute to subtle (and sometimes more pronounced) deviations from expected eye

colour.

Acknowledgements

The Canadian Partnership for Tomorrow's Health (CanPath) research is only possible with the

commitment of its research participants, its staff and its funders. The data used in this research

were made available by CanPath with contributions from CARTaGENE, Alberta's Tomorrow

Project, the Ontario Health Study, the BC Generations Project, and Atlantic PATH. Computational

analyses were carried out using the high-performance computing and storage resources of the

Digital Research Alliance of Canada.

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1025 Authors' Roles

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1027 CLA: Formal Analysis, Writing – original draft, Writing – review and editing. FLD: Methodology,

1028 Writing – review and editing. ME: Data collection, Writing – review and editing. EJP:

1029 Conceptualization, Methodology, Writing – review and editing, Supervision.

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1031 Competing Interests

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1033 N/A

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Fig 1

Fig 2

Preliminary Statistics and Quality control (QC)
PLINK (v2.0) and bcftools (v1.13) used to create separate files for individuals with AA+AG genotypes.

Genome-wide association study (GWAS)
SAIGE (v1.0.5) used to run logistic and linear model GWAS.

Meta-analysis
Metasoft (v2.0.1) used to combine different arrays used in CanPath study.

Conditional and joint analysis (COJO) using GWAS summary statistics
GCTA used to conduct COJO.

Fine-mapping
FINEMAP (v1.4) used to explore genome-wide significant signals and zoom in on local regions.

SNP annotation
SNPs explored in SNPNexus provide genomic context and understanding of function.

Replication
Replication in independent European sample (quantitative eye colour measurements)

Fig 3

Fig 4

Fig 5