Optimized iMAP CRISPR Perturbation Illuminates the Context-Specific Functions of RNA Modification Factors

Short title:

iMAP optimization and application to RNA modification

Ge Wei[1,7], Zhengyu Jing[1,7], Weikai Wang[2,8], Bo Liu[1,8], Shaoshuai Mao[1,3,8], Junjun Lv[1,8], Xiaoming Zhang[1], Dingyu Cao[1], Ruilin Sun[5], Aibing Liang[6], Tian Chi[1,4,9]

[1]School of Life Science and Technology,

ShanghaiTech University, Shanghai 201210, China

[2]BGI Research, Shenzhen 518083, China

[3]Current address: Epigenic Therapeutics, Inc., Shanghai 201315, China

[4]Department of Immunobiology, Yale University Medical School, New Haven, CT, USA

[5]Shanghai Model Organisms Center, Inc., Shanghai, China

[6]Department of Hematology, Tongji Hospital, Tongji University School of Medicine, 1239 Siping Road, Shanghai 200092, P. R. China

[7]Equal contributions

[8]Equal contributions

[9]Correspondence

Tian Chi

School of Life Science and Technology,

ShanghaiTech University,

393 Middle Huaxia Road, Pudong, 201210 Shanghai, China

chitian@shanghaitech.edu.cn

Abstract

iMAP is a large-scale CRISPR perturbation platform based on a germline-transmitted transgene comprising individually floxed, tandemly linked guides that can be inducibly expressed via Cre-mediated recombination. A key limitation was strong recombination bias favoring the proximal guides. Here, we effectively mitigated this bias by inactivating a recombination hotspot, yielding guide representation nearly as uniform as traditional lentiviral libraries. Applying this optimized platform, we systematically profiled 70 (52%) RNA modification factors across 46 mouse tissues, uncovering pervasive tissue-specific essentiality. Notably, iMAP displayed much higher essential gene recovery than prior lentiviral CRISPR screens, attributable to its stronger guide depletion and lower technical noise. Finally, in a tumor model, iMAP identified *Thg1l* as a repressor of NK cell TNFα production; its ortholog knockout enhanced human NK cell-mediated tumor killing, underscoring therapeutic potential. All data are available in an interactive iMAP database. This work advances the Perturbation Atlas, a foundational resource for functional genomics.

Teaser

A unique CRISPR perturbation tool has been upgraded and deployed to reveal functions of epigenetic regulators in mice.

Introduction

Decades after sequencing of the human genome, the context-specific functions of ~20000 protein-coding genes across hundreds of cell types in the human body in health and disease remain poorly defined. A pressing task in functional genomics research is to construct “Cell and Tissue Perturbation Atlas”, which involves systematic disruption of mammalian genome and deciphering of the resulting phenotypes at the cellular and tissue levels in a high-throughput manner in both human and animal models (*1*). Such an atlas can reveal causal relationship between genetic circuits and cellular phenotypes, thus complementing various observational atlases and omics studies, such as the Human Cell Atlas aimed at profiling gene expression patterns of all human cell types (*1*–*3*). Importantly, with increasing amounts of perturbation data, artificial intelligence and machine learning can be deployed to predict gene function, thus enabling a Perturbation Atlas to serve as a generative causal foundation model, which will greatly empower our understanding of human biology in health and disease (*1*).

Pooled CRISPR screening is a standard and powerful high-throughput method for exploring gene functions in cell culture (*4*) and in transplanted immune cells (*5*), but its direct application to the cells *in situ* in mice remains challenging due to inefficient/uneven delivery of viral sgRNA libraries to target cells in their complex native environment(*6–9*). As a result, *in situ* screening has only been reported sporadically for a few solid organs (*10–17*). To eradicate the delivery problem inherent in viral libraries, thus achieving systematic *in situ* perturbation of diverse tissues throughout the whole mice, we have combined CRISPR-Cas9 and Cre-LoxP technologies into a virus-free platform named inducible mosaic animal for perturbation (iMAP) (*18–20*). iMAP uses a novel germline-transmitted transgene comprising 100 individually floxed gRNA-coding units ("guides") linked together in tandem (Figure 1A). The array of guides is placed under the control of a modified U6 promoter with a LoxP variant (Lox71) embedded. In mice carrying the transgene and also ubiquitously expressing CreER and Cas9, treatment of mice with Tamoxifen (TAM), which activates Cre, would trigger recombination between Lox71 and one of the downstream LoxP variant sites preceding each guide (LoxTC9), leading to the expression of all the guides in the mice, but only one of them per cell, thus converting the mice into mosaic organisms ready for phenotypic characterization. iMAP is advantageous over conventional pooled *in situ* CRISPR screens, in that CRISPR perturbation can be readily induced via simple TAM administration, allowing parallel analyses of diverse cell types and tissues throughout the mice, with the mouse lines recurrently accessible as they can be maintained, propagated and shared just as conventional transgenic mice. Importantly, while pooled CRISPR screens typically use multiple gRNAs per target gene, the nonorthodox "one guide/target" design proves acceptable for iMAP (see Discussion) (*18*). With a single iMAP mouse line, and using gRNA representation as the readout, we have rapidly constructed a miniature version of the mammalian perturbation atlas cataloguing the effects of perturbations of 90 genes across 39 organs/tissues/cells ("tissues" hereafter), which is hardly achievable using other tools. However, a major weakness of iMAP is that the recombination between Lox71 and the series of downstream LoxTC9 sites is biased toward the proximal (mainly the first 11 or g1-11) sites, leading to the underrepresentation of the distal guides at P0 in the post-recombination library, which necessitates the use of more cells in order to cover the distal guides (Figure 1A) (*18*). The bias in recombination thus compromises assay sensitivity, hampering the profiling of rare cells in the body.

All known types of RNA, including mRNA, tRNA and rRNA, are subject to dynamic chemical modifications, which impacts RNA stability, subcellular localization and/or interactions with other biomolecules (*21–23*). The modifications are deposited and removed by the "writers" and

94 “erasers”, respectively, whereas the “readers” bind the modification marks and recruit effector
95 machinery to alter cell functions, thus translating the modification signals into functional outputs.
96 RNA modification has emerged as a crucial regulator of gene expression in health and disease,
97 and is attracting increasing attention. The latest version of the Modomics Database documents
98 335 RNA modifications in nature and 137 modification factors (writers, erasers and readers) in
99 humans (https://genesilico.pl/modomics/) (*24*). Traditionally, RNA modification is investigated
100 mostly in cell culture. KO mice have also been described for some of the modification factors
101 (see further), but the KO often proves lethal, precluding in-depth characterization. Thus, the
102 roles of the modification factors in mice, especially in various cell types in adults, remain largely
103 unexplored.

104 In this study, we substantially improved the uniformity of guide representation in iMAP, which is
105 now close to that in conventional lentiviral sgRNA libraries used in pooled CRISPR screens. We
106 then interrogated the functions of 70 RNA modification factors across 46 organs/tissues/cell
107 types (“tissues” hereafter) in adult mice. Furthermore, we combined iMAP and a tumor model to
108 identify a candidate gene for potentiating tumor-killing function of NK cells, which was
109 subsequently validated in human NK cells. Finally, we describe a public iMAP database.
110

111 Results

112 iMAP optimization via elimination of recombination hotspot

113 The Lox71-TC9 recombination in the first-generation iMAP transgene is biased toward roughly
114 the first 11 guides (g1-11), indicating that the region encompassing ~g1-11, about 1.8 kb in
115 length, is a recombination “hot spot” (Figure 1A). We reasoned that its replacement with a 1.8-
116 kb inert “stuffer” sequence lacking LoxP sites might force Lox71 to skip the “hotspot” to reach
117 out to and rearrange with the downstream TC9 sites, thus mitigating the bias in recombination.
118 To test this, we constructed a *91-guide* transgene with a 1.8-kb stuffer inserted between g0 and
119 g1 (Figure 1B; see further for guide composition). In the ensuing transgenic mice, the U6
120 promoter was active, unmethylated and the *91-guide* array intact (Figure S1A-C). We profiled
121 P0 guides in adult *91-guide; Cre* mice at various levels of TAM-induced recombination, and
122 compared the profiles with that in *100-guide; Cre* mice at similar levels of recombination. There
123 is indeed substantial improvement in recombination uniformity in the *91-guide* array at all levels
124 of recombination (Figure 2A), which is more obvious if the guides are analyzed as groups
125 (Figure 2B-C). Specifically, for the *100-guide* array, the group comprising g1-10 was
126 predominant, constituting up to 49-72% of the total recombined P0 guides (yellow bar, Figure

127 2C top), contrary to the distal groups especially g41-99 (1-5%; last six bars, Figure 2C top). In
128 contrast, in *91-guide; Cre* mice, g1-10 dropped to 26-34% while g41-90 rose to 6-15% (yellow
129 vs last five bars, Figure 2C bottom), indicating marked reduction in recombination bias.
130
131 iMAP sensitivity is limited by the least abundant P0 guide generated after recombination. We
132 found that in representative samples from the two mouse lines, each with 76% recombination (a
133 level typically achieved in our routine experiments), such guides constituted 0.1% and 0.35% of
134 the total P0 guides for the *100-* and *91-guide* transgene, respectively (Figure 2D). Thus, the
135 stuffer led to ~3.5x increase in sensitivity.

136 For iMAP analysis of rare cells, it can be challenging to isolate enough cells to cover all the
137 guides in the library, particularly the underrepresented guides. We thus used mathematical
138 modeling to explore the relationship between the numbers of cells sampled and the % of the
139 guides detected. We assume a detected guide is represented by at least 100 cells, a depth we
140 previously found sufficient for iMAP analysis, and we express cell number in relative terms, as
141 "fold over the total guide number in the array" (e.g., a value of 1 for the *100-guide* array means
142 100 cells sampled). For *100-guide*, the model predicts that the cell number must be at least
143 1000 times the guide number (namely 100,000 in absolute number) to detect ~100% of the
144 guides, and the value drops to 60% and 40% when the cell number drops 2x and 4x,
145 respectively (Figure 2E, line 1). In contrast, for *91-guide*, cells only 300 times the guide number
146 suffice to cover nearly all the guides, reinforcing the notion that the stuffer led to 3x increase in
147 iMAP sensitivity (Figure 2E, line 2). This sensitivity is still 3x lower than the theoretical limit
148 achieved if the recombination becomes absolutely even (Figure 2E, line 3), but already
149 surpasses that of conventional lentiviral library, in that full coverage of the viral library requires
150 cells 500 times the gRNA number, contrary to only 300 times for *91-guide* (Figure 2E, line 4).
151 However, for partial coverage, the viral library is more sensitive. For example, to cover 80% of
152 the libraries, pooled CRISPR screen and iMAP would need 170x and 240x cells, respectively
153 (Figure 2E, line 4 vs 2). Thus, on balance, the two platforms perform similarly. Of note, for
154 pooled CRISPR screen, Unique Molecular Identifier (UMI) can be used to improve the screen
155 sensitivity, but this strategy is not applicable to iMAP, which is a limitation of the latter platform.

156 We conclude that the stuffer led to 3.5x increase in iMAP sensitivity, making it comparable to
157 that of the classic lentiviral sgRNA library.
158
159 **Overview of iMAP-91 phenotype**

160 As depicted in Figure 1B, the *91-guide* transgene features 78 guides targeting 70 RNA
161 modification genes (writers/erasers/readers), each gene targeted by one guide except that five
162 genes (*Alkbh3, Fto, Mettl3, Ythdc1, Ythdf1*) are each covered by several guides (to verify the
163 robustness of the "one-guide/target design"; see Discussion). The array also carries a positive
164 control targeting *Polr2a*, an essential gene whose perturbation causes cell death, and nine non-
165 targeting guides (negative control, NC) spiked into the array to control for bias in recombination.
166 We bred *91-guide; Ubc-CreER; CAG-Cas9* mice ("iMAP-91"), treated them with TAM around
167 D10 and analyzed guide representation in diverse tissues 3-4 months thereafter in three
168 females and three males. The mice were grossly normal except for atrophic testes (Figure 3A).
169 We profiled P0 guides in 46 tissues sampled from six mice, yielding 4,002 data points (87
170 important guides x46 tissues) each averaged from at least two (mostly 5-6) mice, the replicates
171 being highly reproducible (Figure S2). Guide depletion events were predominant, constituting
172 49% of the 4,002 data points using fold ≥ 2 as the threshold (black lines) or 74% using gNC for
173 comparison (green lines), contrary to only 0.1%–3% for the enrichment and 23%–50% for the
174 unaltered (Figure 3B). Note that the gNCs formed a sharp peak, attesting to the low noise level
175 of the system. We used simple bar graphs to display the enriched guides (Figure 3C), and a
176 heatmap to display depletion where the enrichment is scored as "unaltered" to better visualize
177 the depletion (Figure 3D). Guide enrichment was marginal (mostly 1.5x) and highly restricted,
178 detected for only four guides (g63-*Rpusd4,* 15-*Trdmt1,* 48-*Snu13,* 62-*Trmu*) and in only two
179 organs (eyeball and cerebellum; Figure 3C); KO data for these genes are scarce but not in
180 conflict with the iMAP data (see Discussion). In contrast, guide depletion was pervasive,
181 encompassing many genes with established functions, thereby enabling rigorous assessment of
182 the depletion's biological validity, as detailed below.
183
184 **Guide depletion pattern is biologically relevant**
185 In the guide-depletion heatmap, genes clustered into five groups (I–V) and tissues into six (A–F;
186 Figure 3D). We previously identified four salient features in the iMAP-100 heatmap that strongly
187 support the biological relevance of this clustering (*18*). These features are all recapitulated in
188 iMAP-91, as detailed below.
189
190 First, related guides tended to co-segregate. Thus, the eight gNC tightly co-segregated
191 (gCluster I), as were guides targeting different essential subunits of the same protein
192 complexes, including the following complexes: *Mettl3-Mettl14* (g53-56), *Iscu-Nfs1* (g83-84) ,
193 *Qtrt1-Qtr2* (g27-28) and to some extents *Trmt6-Trmt61a* (g77,69) and *Mettl1-Wdr4* (g65-76)

(Figure 3D)(*25–30*). Besides, the guides targeting four genes required for tRNA-U34 modifications (*Elp1*, *Elp3*, *Ctu1*, *Urm1*) tightly co-segregated (g42-45), although the guides targeting two other genes in the pathway (g16-*Alkbh8* and g73-*Trmt112*) did not (Figure 3E; see Discussion). Finally, different guides designed to inactivate the same genes, namely *Mettl3* (g53-55), *Ythdc1* (g66-67), *Ythdf1*(g17-18) and *Alkbh3* (g24,26,31), all (tightly) clustered (Figure S3A). On the other hand, guides predicted to impact the same genes but to different extents did not co-segregate, as in the case of *Fto* (g21, 41) and *Gon7* (g29, 85), albeit with the caveat that g85-*Gon7* additionally targets two unannotated genes related to *Gon7* (Figure S3B-C).

Second, relevant tissues tended to co-segregate just as relevant guides, including neural tissues (T2–9), skin-related tissues (T16-17, 20, 23, 25), intestine-related tissues (T26–29), lymphocyte subsets (T30–41), and male germ cell subsets (T42–46). Of note, different tissues apparently have different sensitivities to Cas9, with post-mitotic tissues relatively resistant, which poses a challenge for Perturb Atlas mapping (see Discussion).

Third, the genes generally required for cell survival in iMAP-91 were enriched with the ‘‘core essential genes’’ (CEGs) generally required for human tumor cell survival/proliferation (*31*). Specifically, of the 71 mouse genes targeted in iMAP-91, 12 were orthologous to human CEGs, all distributed among the last 15 mouse genes (73-87) in the heatmap except three (34-*Trmt5*, 48-*Snu13*, 58-*Dimt1* (Figure 3D, pink text); the exceptions have medical implications (see Discussion).

Fourth, guide depletion patterns in iMAP mice mirrored the phenotypes of conventional global single-gene KO mice. Thus, ablation of gCluster IV-V genes is all embryonic lethal (15/15), gClustre III mostly so (12/17) whereas gCluster I-II all viable except for one gene (1/12; Figure 3 F; Table S3).

The following analyses further affirm the heatmap's biological relevance while highlighting iMAP's superior performance relative to conventional CRISPR screens.

First, we measured editing efficiencies for 16 guides—sampled to represent all clusters in the heatmap—in a mouse tumor cell line. All were active (60–90% indel rates), with no correlation between indel efficiency and cluster assignment, confirming that clustering reflects gene functions rather than guide potency (Figure 3G).

228 Second, we computed the receiver operating characteristic area under the curve (ROC-AUC),
229 the standard metric for negative-selection CRISPR screens that measures how well validated
230 essential genes rank as more depleted than non-essential genes (*32*, *33*). Lacking gold-
231 standard lists for primary mouse cells, we used organismal viability as a proxy. To enhance
232 statistical power, we pooled data from iMAP-91 (71 genes; Figure 3D heatmap) and iMAP-100
233 (90 genes; Figure 4D heatmap) (*18*), yielding 161 genes total. MGI single-gene knockout data
234 classified 110 as embryonic essential (EE; lethal before E21), including 23 early embryonic
235 essential (EEE; lethal between E4.5–E8, more likely cell-essential), and 20 as non-essential (no
236 lifespan shortening; Table S3). Using non-essential genes as negatives and averaging $\log_2$ fold
237 changes across all surveyed tissues, we obtained ROC-AUC of 0.80 (95% CI: 0.69–0.92) or
238 0.89 (95% CI: 0.78–0.99) with EE or EEE as positives, respectively (Figure 3H, top plot,
239 columns 1–2). Strikingly, ROC-AUC rose to 0.98–1.00 when non-targeting guides defined the
240 negative class, indicating minimal technical noise in iMAP (Figure 3H, top plot, columns 3–4).
241 Tissue-specific analyses (cerebral cortex, liver, and spermatogonia, representing low,
242 intermediate, and high Cas9 sensitivity, respectively) confirmed that ROC-AUC increased with
243 sensitivity (e.g., 0.71, 0.80, 0.86 using EEE positives and non-essential negatives; Figure 3H,
244 column 2). Notably, these values surpass those from a lentiviral screen (see Discussion).
245
246 Finally, comparisons with genome-wide lentiviral screens in brain(*16*) and liver (*11*) revealed
247 that iMAP-91 uncovered far more essential genes: 12 core essentials (gCluster V) and 16 liver
248 essentials (gCluster IV), versus only one of these in brain and six in liver from the viral screens
249 (marked atop the heatmap; Figure 3D).

250 Collectively, these data underscore iMAP's outstanding performance. We next mined the
251 heatmap as described below.
252
253 Tissue Cluster A: differential effects of CEGs and uniqueness of cerebellum
254 In this group of tissues (brain, heart, and eyeball), guide depletion was undetectable/weak
255 except for gCluster V (g76-87), which was clearly depleted in all tissues, suggesting that g76-87
256 had targeted CEGs. Guide alteration patterns were similar among various brain regions, but
257 comparison of cerebral cortex vs cerebellar central part (Tissue 05 vs 02) revealed multiple
258 guides with differential representations, which occurred due to either quantitative (g78, 81, 82,
259 85, 87) or qualitative (g15, 48, 62, 70) differences in guide abundance between the two tissues
260 (Figure 4A, top). Specifically, in the “quantitative” group, the guides were depleted in both
261 tissues but to different extents, with g82 preferentially depleted in the cerebrum while the

262 remaining (g78, 81, 85, 87) just the opposite. Of note, these five guides (g78, 81, 82, 85, 87) all
263 target CEGs, suggesting that although CEGs are essential for cell survival, the kinetics of cell
264 death following CEG KO could be cell-type dependent. This said, for the most part, gCEGs are
265 depleted to similar extents in closely related tissue pairs and thus are useful controls for Cas9
266 cutting efficiencies (see, e.g, g82-*Polr2a* in Figure 4B-E). Regarding the "qualitative" group,
267 three guides (g48, 15 and 62) were selectively enriched in the cerebellum but no other tissue,
268 whereas g70 generally depleted in various tissues except cerebellum, thus exposing a unique
269 aspect of the cerebellum (Figure 4A and 3D).
270
271 Tissue Cluster B: brown vs white fat
272 Brown fat (bFAT) burns calories and is located primarily in the interscapular region, whereas
273 white fat stores excess energy and is distributed widely, with perigonadal fat (pFAT) and
274 mesenteric fat (mFAT) among the major white adipose tissues. Three guides (g58, 59, 72) were
275 selectively depleted in pFAT and mFAT but not bFAT, suggesting that their target genes (*Dimt1*,
276 *Mrm2* and *Txn1*) selectively promote white adipocyte survival/proliferation/development without
277 impacting bFAT (Figure 4B).
278
279 Tissue Cluster D: small vs large intestine
280 Five guides were differentially represented in the epithelium of small vs large intestine, which
281 remarkably include the "tetra-guides" targeting tRNA-U34 modification (g42-*Elp1*, 43-*Elp3*, 44-
282 *Ctu1*, 45-*Urm1*; Figure 4C). Specifically, all the four guides were depleted clearly (2-4x) in the
283 epithelium of large intestine, whereas in the small intestine epithelium, the depletion was
284 marginal (~1.5x) and restricted to g*Ctu1/Urm1*, suggesting preferential requirement of tRNA-
285 U34 modification for the large intestine.
286
287 Tissue Cluster E: Regulators of T cell development and activation
288 gCluster III–V guides were generally depleted across various immune cells but occasionally to
289 different extents in pairs of closely related T cell subsets:
290
291 First, over a dozen guides were preferentially depleted in naïve CD4 T cells relative to Tregs,
292 with the tetra-guides that target tRNA-U34 modification among the top candidates (Figure 4D,
293 panel 1). The two T cell subsets develop from the same precursor (DP thymocytes), suggesting
294 that this lineage bifurcation is impacted by tRNA-U34 modification.

295 Second, the tetra-guides targeting tRNA-U34 modification were also the top preferentially
296 depleted candidates in naïve relative to effector/memory(E/M) CD4 T cells (Figure 4D, panel 2).
297 Thus, tRNA-U34 modification seems a prominent repressor of both Treg development and CD4
298 T cell activation/differentiation. This function is specific, being absent from CD8 T cells; instead,
299 FTO, which erases N^6-methyladenosine (m^6A), emerged as a prominent repressor of CD8
300 differentiation into both E/M and central memory (CM) CD8 T cells (Figure 4D, panel 3-4). Of
301 note, multiple other guides were also enriched in CM relative to naïve CD8 cells (Figure 4D,
302 panel 4, red dots in the top plot), but these guides were severely depleted even in CM CD8
303 cells, making their relative enrichment in CM CD8 cells less interesting (not shown).

304 Tissue cluster F: Spermatogonia vulnerability

305 Our previous analysis of iMAP-100 mice reveals striking vulnerability of spermatogonia, which is
306 recapitulated in iMAP-91:

307 First, spermatogonia were more vulnerable than their progeny (Figure 4E): out of the 79
308 targeting guides, only seven failed to impact spermatogenesis; the remaining 72 guides were
309 depleted at least 2x in spermatogonia, and no further change was seen in the progeny except
310 for g*Nsun2, Cmtr2* and *Elp1* (and to some extent g*Elp3*) whose depletion emerged in
311 spermatogonia and exacerbated in P/D (for g*Nsun2*) or RS (for g*Elp1/Elp3/Cmtr2*; Figure 4E). In
312 other words, 72 guides blocked spermatogonia survival/proliferation/renewal, but only three of
313 them additionally impacted its progeny. Of note, for the four guides targeting tRNA-U34
314 modification, depletion in spermatogonia was moderate (2-4x) for g*Elp1/3* but dramatic (17-38x)
315 for g*Urm1/Ctu1*, highlighting their functional dichotomy (see Discussion).
316
317 Second, spermatogonia were more vulnerable than somatic cells, in that gCluster II was
318 preferentially depleted in spermatogonia relative to somatic cells (Figure 3D). In particular, four
319 guides (27-*Qtrt1*, 28-*Qtrt2*, 29-*Gon7-2* and 30-*Alkbh5*) were depleted marginally in somatic cells
320 but severely (6-27x) in the germline (Figure 4F). QTRT1 and QTRT2 form the heterodimeric
321 tRNA guanine transglycosylase (TGT) that catalyzes the base-exchange of a wobble guanine
322 (G34) with queuine (Q) in some tRNAs, with QTRT1 being the catalytic subunit(*34*). The co-
323 depletion of both *Qtrt* guides strongly argues for the essential and selective role of TGT in
324 spermatogenesis. Regarding *Gon7*, it encodes a subunit of the KEOPS complex that introduces
325 tRNA N6-threonylcarbamoyladenosine (t6A) modification(*35*). g29-*Gon7-2* is predicted to
326 eliminate the C-terminal 13 residues of GON7, thus revealing the selective and essential role of
327 this tail in spermatogonia (Figure S3C). Finally, *Alkbh5* encodes an m6A eraser, and its global

328 KO induces germ cell apoptosis and male sterility, but the mice seem otherwise normal, in
329 general agreement with the iMAP data (*36*).
330
331 In summary, these data highlight the vulnerability of spermatogonia, which has medical
332 implications (see Discussion).

333 Uncovering a potential target for boosting human NK cancer therapy

334 Finally, we sought to enhance human cancer immune therapy, focusing on natural killer (NK)
335 cells, a pivotal first-line tumor killer (*37*, *38*). Release of TNFα is a key mechanism of NK-
336 mediated killing (*39*). Our strategy was to first use iMAP to identify potential candidate genes
337 whose KO could elevate TNFα expression in mouse NK cells, and then use the information to
338 engineer human NK cells, which simultaneously serves to validate the candidates; a similar
339 strategy has been taken to potentiate human CAR T cells in our previous study (*18*).
340 Specifically, we inoculated iMAP-91 mice with mouse colon cancer MC38 cells, administered
341 TAM to induce recombination, isolated and compared TNFα^{hi} vs TNFα$^{low/-}$ (Figure 5A). A single
342 guide (g86-*Thg1l*) was markedly enriched in the TNFα^{hi} relative to the TNFα$^{low/-}$ subset (Figure
343 5A), implicating *Thg1l*, a probable tRNA^{His} guanylyltransferase, as a repressor of TNFα
344 expression in mouse NK cells. To validate and translate this finding, we generated *THG1L*
345 knockout human NK cells. Upon co-culture with K562 leukemia targets, *THG1L* KO NK cells
346 secreted more TNFα (481 vs. 337 pg/mL) and exhibited enhanced target-cell lysis (65% vs.
347 39%) (Figure 5 B-C). Notably, expression of perforin, granzyme B, and the degranulation marker
348 CD107a remained unchanged (Figure 5C), indicating that increased TNFα drives the improved
349 killing. The phenotype was reproducible across biological replicates and with independent guide
350 RNAs (Figure S4). This proof-of-concept mirrors our prior success in engineering CAR T cells
351 (Liu et al., 2022) and establishes iMAP as a rapid platform for discovering immunomodulatory
352 targets translatable to human cell therapy.
353

354 iMAP website

355 We have constructed a public database for browsing, searching and analyzing iMAP data
356 (http://49.235.152.12/; Figure S5A). Four key modules are featured: “Heatmap”, which displays
357 interactive *Guide x Tissue* atlases (Figure S5B); “Gene Search”, which enables query for the
358 abundance of guides of interest across tissues (Figure S5C); “Network”, which presents an
359 interactive network of the guides surrounding a guide of interest, with relevant information
360 indicated, such as fold-changes of the guides, the number of tissues impacted by a guide, the
361 distances among the guides and the total number of guides connected with a particular guide

362 (Figure S5D); and "Tissue comparison", which identifies the guides differentially represented
363 between a pair of selected tissues (Figure S5E). Future efforts should culminate in the creation
364 of the panoramic Perturbation Atlas covering all the genes across all the cell types at tissue and
365 single-cell levels.
366
367 **Discussion**
368 Perturbation Atlas: challenges and countermeasures
369 A central goal in functional genomics is to build a comprehensive Perturbation Atlas that maps
370 the *in vivo* phenotypic consequences of gene knockout across diverse tissues. This tissue
371 diversity poses a challenge to iMAP (and classic CRISPR screens), because Cas9 cleavage
372 efficiency varies markedly between tissues. Specifically, post-mitotic tissues (especially the
373 brain) are relatively refractory to Cas9 probably due to limited chromatin accessibility; a potential
374 solution here is to use Cas13 (which acts on mRNA) for perturbation. On the other hand, the
375 male germline was highly sensitive to Cas9, with wide-spread guide depletion not only in
376 embryonic essential genes but also non-essential genes. This extraordinary depletion pattern
377 unlikely reflects cell death resulting from Cas9 induced DNA breaks. First, non-essential gene
378 depletion was much weaker, enabling their clear separation from early essential genes (ROC-
379 AUC = 0.86). More importantly, in an iMAP line targeting 6 marker genes each with 10 guides
380 (iMAP-61), testicular guide abundance perfectly reflects known biology: depletion of cell-
381 essential controls (*Polr2a, Kpnb1*), enrichment of tumor suppressors (*Pten, Trp53*), and no
382 change in lineage markers not expressed in testis (*Cd19, Cd45*) (*18*). This, the widespread
383 depletion in the male germline reflects genuine biological requirements. Anyway, given the
384 variability in Cas9 cleavage, direct comparisons of guide abundance across distantly related
385 tissues (e.g., brain vs. testis) are not meaningful. However, comparisons within closely matched
386 tissue pairs of similar cellular state and Cas9 sensitivity—such as cerebellum vs. cerebrum or
387 male germ cells at different developmental stages —remain valid and informative (Figure 4).
388
389 iMAP robustness despite one guide per target
390 Although conventional CRISPR libraries employ multiple guides per gene to reduce false hits,
391 iMAP deliberately uses only one guide to maximize library capacity, which fortunately does not
392 markedly sacrifice robustness thanks to several design features. First, iMAP guides were
393 selected using the state-of-the-art Rule Set 2 algorithm—the same algorithm that powers the
394 Brunello library. In tumor cell viability screens, a single Brunello guide per gene already
395 achieves excellent performance (ROC-AUC = 0.93), with additional guides providing only minor

396 improvement (up to 0.98 with 4 guides/target)—in sharp contrast to the earlier GeCKOv2 library,
397 which requires 6 guides to increase ROC-AUC from 0.73 to 0.91 (*32*). The iMAP libraries were
398 designed according to the same principles as Brunello and therefore deliver comparable
399 reliability with just 1 guide/target. Second, iMAP founder lines were pre-screened by RT-qPCR
400 for strong gRNA expression. Third, all guides are integrated at a single genomic locus in every
401 cell, minimizing cell-to-cell variability. Finally, iMAP exhibits exceptionally low technical noise
402 (see further). These features collectively deliver impressive performance across a series of
403 benchmarks: false hits are rare (1/10 false negatives and 1/590 false positives in the iMAP-61
404 control line targeting marker genes) (*18*); iMAP phenotypes consistently recapitulate known
405 biology; all 16 guides randomly selected from iMAP-91 proved active (Fig. 3G); and finally, as
406 elaborated in the next three sections, distinct guides targeting the same genes elicit similar
407 phenotypes, seeming inconsistencies with the literature are reconcilable, and iMAP
408 performance appears to rival that of classic CRISPR screens.
409
410 Consistencies among orthogonal guides targeting the same genes
411 As a metric for iMAP performance, we designed 2-3 guides per gene for four of the 70 RNA
412 modification genes, totaling 10 guides for 4 genes (Figure S3A). These guides are predicted to
413 produce null alleles, and the orthogonal guides indeed behaved similarly across all 46 tissues
414 surveyed, albeit with a minor discrepancy: for the two guides targeting *Ythdf1* (g17-18), g18 was
415 not altered in the germ line (T42-46), while g17 mildly (<1.5x) enriched (Figure S3A). Thus,
416 among 460 data points (10 guides x 46 tissues), only 5 (1 guide x 5 germ cell types) are
417 inconsistent among the orthogonal guides, comprising only ~1% of total data points. Of note, for
418 2 other genes (*Fto* and *Gon7*), two guides are designed for each, but the guides are predicted to
419 produce distinct effects, which was indeed fulfilled (Figure S3B-C).
420
421 Seeming inconsistencies with the literature
422 The iMAP data agree well with the literature, but with three exceptions. In each case, guide
423 abundance was altered in iMAP mice despite little defects in the KO mice, but each discrepancy
424 can be reconciled by considering cell competition unique to the mosaic iMAP mice.
425
426 First, g42-*Elp1* was depleted in the epithelium of large intestine whereas conditional *Elp3* KO in
427 the intestinal epithelium fails to produce overt phenotypes (*40*). Perhaps the KO cells are
428 intrinsically defective, but the defect can only be exposed in the mosaic iMAP mice where the
429 KO cells tend to be surrounded by, and thus potentially in competition with, normal cells in the

430 same niche. Indeed, due to cell competition, some mutant phenotypes can only be exposed in
431 mosaic systems (*41*, *42*).
432
433 Second, whereas g15-*Trdmt1* was marginally enriched in the cerebellum, *Trdmt1* KO mice are
434 rather healthy (MGI:1274787), suggesting that this expansion might occur only in the mosaic
435 iMAP mice, where the *Trdmt1* gRNA might enable the cells to become a “super competitor” to
436 outcompete the normal neighboring cells; such an advantage does not exist in KO mice where
437 all the cells are equally competitive (*43*). Alternatively, the putative cerebellum expansion might
438 indeed occur in KO mice, but is well tolerated. Of note, g62-*Trmu* was also enriched in the
439 cerebellum, but its KO mice die between implantation and somite formation, making it
440 impossible to infer its role in the brain (*44*). Importantly, the overall mild and severe phenotypes
441 in *Trdmt1* and *Trmu* KO mice, respectively, align well with the severe depletion across diverse
442 tissues of g62-*Trmu* but not g15-*Trdmt1* (Figure 3D). Furthermore, conditional *Trmu* KO in the
443 liver results in spotty liver necrosis, consistent with g62-*Trmu* being depleted in the liver in iMAP
444 mice (Figure 3D). Thus, the literature on *Trdmt1* and *Trmu* KO mice tends to support the iMAP
445 data.
446
447 Finally, g27-*Qtrt1* was depleted in spermatogonia (Figure 4F), but a gene-trap cassette that
448 eliminates a 48-aa region required for QTRT1 catalytic activity fails to compromise
449 spermatogenesis, which again can be explained by invoking the mosaic nature of iMAP tissues,
450 especially because the trapped allele still expresses the remaining 90% of QTRT1 (aa1–104
451 and 151-403) and presumably retains most of its non-enzymatic functions (*45*).
452
453 iMAP apparently outperformed lentiviral CRISPR screens *in vivo*
454 Genome-wide lentiviral CRISPR screens in the brain (*16*) and liver (*11*) provide a rare
455 opportunity to benchmark the classic platform against iMAP. iMAP-91 showed much higher
456 recovery rate of essential genes than the viral screens (Figure 3D), with a similar trend in iMAP-
457 100 (*18*)—the first evidence of iMAP's superior performance. To investigate further, we
458 compared ROC-AUC values in the liver (brain lentiviral data unavailable). Using non-essential
459 genes as negatives, iMAP achieved an AUC of 0.80, surpassing the lentiviral screen at 1
460 guide/target (AUC 0.73) and matching it at 5 guides/target (AUC 0.78; Figure S6A, white bars).
461 Strikingly, with non-targeting guides as negatives, iMAP substantially outperformed the lentiviral
462 screen even at 5 guides/target (AUC 0.92 vs. 0.82), likely reflecting its tighter null distribution
463 (Figure S6A, gray bars). Multiple guides conferred only marginal benefits for the lentiviral

464 screen—particularly with non-targeting negatives (ΔAUC = +0.03 from 1 to 5 guides/target)—as
465 anticipated given its high-quality library powered by Rule Set 2 (Figure S6A; compare 1 vs. 5
466 guides). To dissect the basis for the AUC differences, we examined $\log_2$ fold changes for guides
467 targeting shared early embryonic-essential genes (n=25 guides) and non-essential genes (n=23
468 guides), plus non-targeting guides (n=16 for iMAP; n=1,813 for lentiviral). Both platforms
469 correctly ranked depletion by essentiality (essential > non-essential > non-targeting controls;
470 Figure S6B), but iMAP showed ~4-fold stronger essential-gene depletion (median $|\log_2 FC| \approx$
471 1.37 vs. 0.32; Figure S6B, left panel). This indicates superior knockout efficiency in iMAP, likely
472 because its gRNA expression is more robust and uniform than the classic screens where the
473 viral vectors are randomly integrated and high-expressers not enriched. Regarding the non-
474 targeting guides, although both platforms exhibited near-zero median $\log_2$ fold changes (0.02),
475 the adjacent values range—which captures the bulk of non-outlier data—was ~3.7-fold
476 narrower in iMAP (width 0.23; −0.19 to 0.04) than in the lentiviral screen (width 0.84; −0.43 to
477 0.41; Figure S6B, right panel), signifying lower guide-to-guide variability in iMAP. The above fold
478 changes were averaged across biological replicates, precluding assessment of inter-replicate
479 variability. To address this, we computed the maximum-to-minimum fold-change ratio for each
480 non-targeting guide across replicate mice (ratio of 1 indicates identical values). Although
481 medians were comparable (1.29 for iMAP vs. 1.40 for lentiviral), the adjacent values range was
482 2.5-fold narrower in iMAP (width 0.49; 1.13 to 1.62) than in the lentiviral screen (width 1.23; 1.03
483 to 2.26), revealing greater mouse-to-mouse variability in the latter (Figure S6C)—likely
484 attributable to its per-mouse viral library delivery. Collectively, these findings indicate that iMAP
485 generates stronger on-target signals (perhaps via robust gRNA expression) and lower noise
486 (perhaps from avoiding viral delivery), underpinning its superior performance despite using one
487 guide per target. Side-by-side comparisons with identical guides would be required to confirm
488 these advantages.
489
490 Complexity of RNA modification factors
491 Our study sheds light on context-dependent gene functions and also reveals their complexity.
492 Two issues are particularly noteworthy, each concerning tRNA modifications, as detailed below.
493
494 The first issue involves tRNA wobble uridine (U34), whose modification (mcm^5s^2) is important for
495 accurate translation of a subset of tRNAs(*30*). mcm^5s^2 comprises a methoxycarbonylmethyl
496 group at C5 position (mcm^5) introduced by ELP1-6 in conjunction with ALKBH8 and TRMT112,
497 and a thiol group at C2 position (s^2) subsequently introduced by URM1 in conjunction with

498 CTU1/2, with mcm^5 being a prerequisite for s^2 (Figure 3F)(*46*). Of note, ELP1-6 forms the
499 Elongator Complex, whose main function is U34 modification rather than RNA polymerase
500 elongation (*47*). iMAP-91 targets six of the 11 factors in the modification pathway, namely *Elp1,*
501 *Elp3, Ctu1*, *Urm1, Alkbh8* and *Trmt112*. *Elp1/3* KO would eliminate all four modifications (cm^5,
502 mcm^5, mcm^5s^2, ncm^5) present on 11 tRNAs, whereas *Urm1/Ctu1* KO only mcm^5s^2 present on
503 three (Figure 3F). Thus, one might expect stronger phenotypes from *Elp1/3* KO. However,
504 *Urm1/Ctu1* KO is each lethal (MGI:1915455, 2385277), just like *Elp1/3* KO (MGI: 1914544,
505 1921445). In agreement with this, g*Ctu1*/*Urm1* depletion was as severe as g*Elp1/3* across
506 diverse tissues and even far more severe in spermatogonia, suggesting that *Ctu1*/*Urm1* have
507 functions beyond mcm^5s^2 writing. It is also unusual that although *Alkbh8* is crucial for mcm^5s^2
508 just as *Urm1/Ctu1*, g16-*Alkbh8* was only weakly depleted, and *Alkbh8* KO mice are alive with at
509 most weak defects despite the complete loss of mcm^5s^2 (*48*, *49*). This is perhaps because in
510 *Alkbh8* KO mice, (n)cm5 becomes highly abundant, which might compensate for the loss of
511 mcm^5s^2 (*49*). In apparent conflict with this speculation, g73-*Trmt112* was severely depleted
512 contrary to g*Alkbh8*, even though *Trmt112* acts in concert with *Alkbh8* in writing mcm^5. The data
513 suggest that *Trmt112* has functions beyond mcm^5 writing and predict that *Trmt112* KO, which
514 has not been reported, would be lethal.
515
516 The second issue regards the KEOPS complex mediating the N6-threonylcarbamoyladenosine
517 (t6A) modification at A37 of a subset of tRNAs(*35*). KEOPS consists of five subunits (LAGE3,
518 TPRKB, OSGEP, TRP53RK and GON7) each necessary for efficient t6A modification, with
519 mutations in human orthologs each lead to Galloway-Mowat syndrome (GAMOS) characterized
520 by organ anomalies and early death (*50*). However, although *Tprkb* and *Osgep* KO lead to
521 preweaning lethality reminiscent of GAMOS (MGI:1917036,1913496), *Lage3* KO mice are
522 viable with only a mild defect (neutrophil number change) unrelated to GAMOS (MGI:6257568),
523 dovetailing with g38-*Lage3* depletion being much less conspicuous than g70-*Tprkb*/g79-*Osgep*
524 depletion. Thus, in contrast to man, murine *Lage3* might be dispensable for t6A, or t6A
525 dispensable for survival, which underscores species-specificity of KEOPS function. Of note, our
526 study also bears on *Trp53rk* and *Gon7*, whose KO have not been reported. Specifically, murine
527 TRP53RK is encoded by two genes, *Trp53rka* and *Trp53rkb*. g47-*Trp53rkb* (but not g20-
528 *Trp53rka*) clusters with the guides whose target gene KO is lethal, predicting that *Trp53rkb* (but
529 not *Trp53rka*) KO would likely be lethal. For *Gon7*, two guides were designed: g85-*Gon7-1*
530 inactivates *Gon7* (together with two other genes), whereas g29-*Gon7-2* deletes GON7 C-
531 terminal tail (Figure S3C). Guide depletion was widespread for g85-*Gon7*-1 but restricted to

532 spermatogonia for g*Gon7*-2. Thus, *Gon7*/related genes are essential for diverse tissues, but the
533 GON7 C-terminus is largely dispensable except for spermatogonia, highlighting the unusual
534 vulnerability of this cell type.
535
536 Therapeutic targets
537 First, we demonstrated the potential of iMAP in identifying targets for improving human NK-
538 based therapy. Although the actual candidates emerging from the current screen of 70 RNA
539 modification factors are limited in both quantity (only one gene) and quality (moderate
540 potentiation of NK-mediated killing), larger-scale screens might uncover better candidates,
541 which in turn necessitates the improvement in iMAP throughput. Our preliminary data suggest
542 that a 10x-increase in the throughput (from 100 to 1000 guides/mouse line) is feasible (not
543 shown).
544
545 Second, our study, together with a previous report that global *Alkbh5* KO selectively impacts
546 spermatogenesis (*36*), highlights *Qtrt1/2* and *Alkbh5* as appealing male contraceptive targets.
547
548 Finally, our study bears on targeted cancer therapy. Guides targeting mouse orthologs of human
549 CEGs were in general severely and ubiquitously depleted as expected, but g34-*Trmt5* was a
550 striking exception, being hardly affected in most tissues despite normal editing rate (~70% indel)
551 (Fig. 3D, 3G). This could reflect an evolutionary divergence in *Trmt5* function (i.e., essential for
552 human but not mice), or more interestingly, a dichotomy between cancer and normal cells (i.e.,
553 cancer but not normal cells is addicted to *Trmt5*). In the latter scenario, *Trmt5* inhibitors could
554 become cancer drugs that are broad-spectrum yet safe. Such drugs would be transformative;
555 indeed, eight CEGs have been FDA-approved as cancer therapy targets, but all of them are
556 indispensable for normal cells, and their inhibitors are all highly toxic and of limited applications
557 (*51*).

558 Limitations of the study
559 First, the false negative rates are acceptable but still much higher than false positive (1/10 vs
560 1/590 in iMAP-61), which can be minimized by guide pre-validation via high-throughput
561 screening (*52*). Second, some tissues might be sensitive to Cas9-induced double-stranded
562 breaks, leading to false positives. Guides targeting safe regions can be included to control for
563 such genome toxicity in the future. Finally, the *91-guide* array was constructed years ago,
564 covering all the mouse RNA modification factors known at the time, but the number has almost
565 doubled since then, necessitating the creation of new iMAP lines to catch up with the rapid

566 progress in the field.

567

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577 **Author contributions** T.C. conceived of and supervised the project. B.L. and Z.J. generated
578 the *91-guide* line, Z.J. created the iMAP database, G.W., Z.J. and W. W. did the experiments
579 and/or bioinformatics analysis with technical help from the remaining authors.

580

581 **Declaration of interests** The authors declare no competing interests.

582 **Data and code availability**

583 All original code has been deposited as an R package at GitHub and can be accessed using the
584 following link https://github.com/Jing-Zhengyu/iMAP. Any additional information required to
585 reanalyze the data reported in this paper is available from the lead contact upon request. The
586 source data and the codes for their plotting, together with the plasmid maps, are available at
587 https:// github.com/Jing-Zhengyu/Data-of-iMAP. The raw data for NGS is available at
588 http://www.ncbi.nlm.nih.gov/bioproject/1328907 , with the accession ID being PRJNA1328907.

589

590 **Methods and Protocols**

591 **Mice**

592 The *91-guide* array, the ensuing transgenic line and the iMAP-91 mice were generated
593 essentially as described (*18*). Briefly, we designed candidate guides using CRISPick and
594 selected those likely to induce out-of-frame indels using inDelphi. Guide sequences are listed in
595 Table S1. The transgene was assembled using Golden-Gate cloning (the sequence available at
596 https://github.com/Jing-Zhengyu/Data-of-iMAP) and randomly integrated into the genome via
597 *piggyBac* transposition following microinjection of fertilized eggs. PCR primers for determining
598 transgene integration sites, transgene integrity, U6 promoter methylation, genotypes and P0

599 guide expression are listed in Table S1. The founder line bearing the transgene inserted at
600 Chr6:69964770(+) (coordinates based on Mouse mm39) was used for further studies. Animal
601 experiments were approved by the Animal Ethics Committee at ShanghaiTech, and performed
602 in accordance with the institutional guidelines.
603
604 **Perturbation Atlas and comparison with lentiviral screen in the liver**
605 The atlas was constructed essentially as described (*18*). Briefly, postnatal day 10 (D10) iMAP
606 pups (*91-guide; Cre; Cas9*) were given TAM at 0.2 g/kg daily for four days and sacrificed 3-4
607 months later. Diverse solid organs/tissues were collected and genomic DNA extracted before
608 PCR-NGS profiling of P0 guide representation. Immune and germ cell subsets were isolated by
609 electronic sorting, and genomic DNA equivalent to ~30,000 cells were used for PCR-NGS; PCR
610 primers are listed in Table S1. The reads were normalized, fold-change over control (*91-guide*;
611 *Cre*) calculated and data visualized in heatmap using a home-made script package
612 (https://github.com/Jing-Zhengyu/iMAP). Briefly, the reads counts of targeting guides in
613 experimental and control mice are first normalized to the non-targeting guides in the respective
614 groups before deriving the ratios (fold-changes) of the reads counts. To compare iMAP with the
615 lentiviral screen in the liver reported by Keys and Knouse, we first performed blast search for
616 ~6500 control gRNAs listed in Table S2 of the Keys and Knouse paper against the mouse
617 genome, which led to the identification of 1813 “non-targeting guides” that do not match the
618 mouse genome. We then obtained gRNA reads counts from Table S3 of that paper for these
619 non-targeting guides and also for all the targeting guides, and calculated their fold-changes
620 relative to the plasmid library using the same script as for iMAP. The data were presented in
621 Figure S6.
622
623 **Analysis of gRNA efficiency in N2a cells**
624 We co-transfected vectors expressing gRNA (150 ng) or Cas9 (350 ng) into the mouse
625 neuroblastoma N2a cells cultured on 48-well plates. 72 h post-transfection, we extracted
626 genomic DNA, amplified 500-900-bp regions encompassing the sites targeted by gRNAs using
627 50 ng genomic DNA as templates. The amplicons were then sequenced using the Sanger
628 method and analyzed using ICE (https://www.synthego.com). PCR primers are listed in Table
629 S1.
630
631 **Screens for potential therapeutic targets in tumor-infiltrating NK cells**
632 MC38 mouse colon cancer cells, gift from Yong Cang (ShanghaiTech University), maintained in

633 DMEM supplemented with 10% FBS and 1% penicillin-streptomycin, were detached using
634 0.25% trypsin, washed with PBS, resuspended at 1×10^6 cells per 100 µl PBS and injected the
635 100ul subcutaneously into the dorsal flank of 5-8 weeks-old iMAP-91 mice. Tamoxifen was
636 administered via oral gavage (0.6 g/kg) on the day of tumor inoculation. Tumor growth was
637 monitored every 2 days using calipers, and volumes calculated as $(L \times W^2)/2$,
638 where *L* and *W* represent the longest and perpendicular diameters, respectively. Tumors were
639 excised when reaching 1–1.5 cm^3, which took 18-25 days, minced and digested in RPMI-1640
640 containing 1 mg/ml collagenase IV (Sigma-Aldrich) and 0.1 mg/ml DNase I (Sigma-Aldrich) for
641 20–30 minutes at 37°C with gentle agitation. The digested samples were filtered through a 70-
642 µm strainer and cells pelleted by centrifugation at 400 ×g for 5 minutes (4°C). To identify TNFa
643 regulators, cells were first stained for CD45.2-BB700, CD3-PE-Cy7 and NK1.1-APC, fixed with
644 BD Cytofix/Cytoperm™ for 20 minutes at 4°C, washed with BD Perm/Wash™ and stained with
645 TNFa-PE. $CD45.2^+CD3^-NK1.1^+$ NK cells were sorted into $TNFa^{high}$ (top 30%) and $TNFa^{low/-}$
646 (bottom 30%) populations using a BD FACS Aria III and analyzed by PCR-NGS.

647 **Target validation in human NK cells**

648 Human PBMCs (Rubei Biotechnology) were cultured in Corning ® Lymphocyte Serum-free
649 Medium supplemented with 10% FBS, 1% penicillin-streptomycin and 200 U/ml recombinant IL-
650 2 (Novoprotein). NK cells were expanded using an IL-21-based NK Expansion Kit (Zhongwin
651 Biotech) with stimulation on days 0 and 7. NK cell purity (>90% $CD3^-CD56^+$) was confirmed by
652 flow cytometry on day 10. K562 leukemia cells stably expressing firefly luciferase (gift from
653 Guang Yang, ShanghaiTech) (*11*, *53*) were maintained in RPMI-1640 with 10% FBS and 1%
654 penicillin-streptomycin under standard conditions. RNP was assembled by incubating 40 pm
655 each of recombinant Cas9 (Sino Biological) and sgRNA or non-targeting control (Table S1) at
656 37°C for 15 min. RNP was mixed with 3×10^6 NK cells resuspended in 20 µl P3 Primary Cell
657 Solution (Lonza) before electroporation using a 4D-Nucleofector™ (program CM-137). 24 h
658 post-transfection, successful editing was confirmed by PCR-Sanger sequencing, and NK
659 (0.2×10^6/well) were co-incubated with K562 (0.4×10^6/well) cells for 4-6 h in 96-well plates in
660 PBMC culture media. To assess K562 lysis, cells were washed with PBS, resuspended in media
661 containing cell-permeable D-Luciferin potassium salt (150 µg/ml, Beyotime) and incubated at
662 37℃ for 5 min before measuring luminescence using an Infinite M200 Pro plate reader (Tecan)
663 (*53*). The lysis rate was calculated as (1-X/N) %, where X is the luminescence intensity of K562
664 cells co-cultured with electroporated NK cells, and N is that of K562 cells never exposed to NK
665 cells. CD107a, perforin and Granzyme B were detected by FACS, whereas TNFa secreted to

the media by ELISA.

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854 **Figure legends**
855
856 **Figure 1. Strategy for minimizing recombination bias**

857 (A) First-generation iMAP transgene. Lox71 and TC9 are LoxP variants embedded in the U6
858 promoter and precede each gRNA-coding gene ("guide"), respectively. The guide at Position 0
859 (P0) is g*Cd19* (not shown). The last guide (g*Cd47*) facilitates PCR-NGS quantification of
860 recombined products. Following recombination between Lox71 and a LoxTC9 preceding a
861 guide, the intervening sequence is deleted, re-locating the guide to P0 for transcription. The
862 recombination between Lox71 and the series of LoxTC9 sites is biased toward the proximal
863 LoxTC9 sites except the first site, perhaps due to steric hindrance created by the transcription
864 machinery at U6 promoter.

865 (B) Optimized configuration. The "stuffer" harbors an expression cassette for monitoring U6
866 promoter activity, which proved unsuccessful (Figure S1D). The guide at Position 0 (P0) is
867 g*Cd19* (not shown). The target genes and gRNAs are listed in Table S1.

868

869 **Figure 2. Improved guide representation in the optimized iMAP transgene**

870 (A) P0 guide profiles as a function of recombination extents. *100-guide; Cre* (top) or *91-guide;*
871 *Cre* (bottom) mice were treated with increasing doses of TAM before analysis of various organs
872 (Figure S1E), and samples with progressive recombination, as measured by g*Cd19* deletion,
873 are selected for display. Shown is the relative abundance of individual guide among all P0
874 guides, with each line representing a sample. The guides are arranged according to their
875 positions on the array. An ID number is appended to each guide if several guides are designed
876 for the same genes. Two identical non-targeting guides (g50, 51) were inadvertently built into
877 the *91-guide* array. The colored bar at the bottom specifies the groups of guides whose
878 abundance is analyzed in B. Source data in Table S2.

879 (B) Same as A, but displays the guides as groups defined by the colored bar in A. The values
880 above the pies indicate the extent of g*Cd19* deletion; e.g., the third pie shows 70% of g*Cd19*
881 survived recombination, translating to 30% deletion. The upper and lower pie charts are paired.

882 (C) Same as B but displays guide abundance relative to all the re-arranged P0 guides (hence
883 with g*Cd19* excluded), which better exposes the bias in recombination. The dotted lines denote
884 the theoretical guide abundance if the recombination is absolutely even.

885 (D) Same as A, but compares a pair of samples with 76% recombination from the two mouse
886 lines. The inset displays the distal and hence low-abundance guides.

887 (E) Monte Carlo simulation of guide detectability as a function of number of the cells sampled.
888 The simulation is based on actual guide representation after recombination (#1-2) or viral
889 transduction (#4; data averaged from two lentiviral libraries reported in (*54*).)

890

891 **Figure 3. iMAP-91 characterization**

892 (A) Atrophic testes.

893 (B) Frequency distribution of the fold-changes for the 79 targeting (pink) and eight NC (cyan)
894 guides across 46 tissues. The 4002 (87x46) datapoints were binned according to the fold-
895 change magnitudes, and the frequencies of each bin displayed for each guide, with the highest
896 frequency in each histogram set as 100%. Black lines mark 2-fold changes, whereas the green
897 lines indicate the 5th and 95th percentiles of gNC values.

898 (C) Guide enrichment. Tissue code 01, 02, 03 refers to eyeball, cerebellar central part and
899 cerebellar flocculus, respectively, as specified in (D). Values are mean ± SD (n=6). *$p < 0.05$, **p
900 < 0.01, ***$p< 0.001$, Student's t-test.

901 (D) Cluster analysis of guide depletion. Displayed are 87 guides comprising 78 guides targeting
902 RNA modification factors, one targeting *Polr2a* (g82), and eight gNC (g1-8). Gene essentiality
903 per literature is indicated at the top of the heatmap, where shRNA-brain and CRISPR-brain refer
904 to, respectively, the shRNA- and sgRNA-based genomewide *in situ* screens for neuronal
905 essential genes in the mouse striatum (*16*), CRISPR-liver is a sub-genomewide *in situ* screen
906 for liver essential genes (*11*), and CEG2 (pink) the core essential gene set identified in human
907 tumor cell lines(*31*); three genes targeted by iMAP-91 are not covered in the CRISPR-liver
908 screen (gray box). Each data point (guide-tissue combination) is averaged from 2 to 6 (mostly 5-
909 6) biological replicates. Source data in Table S3.

910 (E) Cascade reaction leading to tRNA-U34 modifications (*46*).

911 (F) Guide clustering coincides with the phenotypes of global single-gene KO mice. Analyzed are
912 the genes whose KO phenotypes are available from MGI. The number of genes in each
913 category is indicated. *Alkbh3* fits both Cluster II and III because it has three guides, two (g24,
914 26) residing in Cluster II and the slightly distant third (g31) at the very beginning of Cluster III.
915 Source data in Table S3.

916 (G) Editing rates of representative gRNAs. gRNAs and Cas9 were co-expressed in N2a cells in
917 duplicates before indel quantification.

918 (H) ROC-AUC analysis. The data are pooled from iMAP-91 and iMAP-100. The numbers of
919 targeted genes and non-targeting guides are bracketed. Error bars represent 95% confidence
920 intervals (CI, Delong's method). The analysis was performed using pROC in R. The KO
921 phenotypes are compiled from MGI (https://www.informatics.jax.org/). Source data in Table S3.
922

923 **Figure 4. Mining the heatmap data**

924 (A-D) Comparing tissue pairs. Shown are differentially represented guides (top) and their
925 absolute abundance normalized to *91-guide; Cre* control mice (bottom). Red and green dots
926 within the top plots indicate significant enrichment and depletion, respectively (fold-change> 2,
927 p<0.05).

928 (E) Guide depletion during spermatogenesis. Same as A-D, but focusing on the male germline.

929 (F) Germline-specific guide depletion. Values are mean ± SD, n = 15–227, each data point
930 being a guide-tissue-replicate combination. ****p < 0.0001.

931

932 **Figure 5. Screening and validation of NK regulators**

933 (A) Genetic screens for NK regulators in tumor-bearing iMAP-91 mice. Tumor-infiltrating cells
934 were stained for CD45.2, CD3 and NK1.1 together with intracellular TNFα before indicated NK
935 subsets were isolated by flow cytometry and analyzed by NGS. Each data point in the volcano
936 plot is averaged from 3 biological replicates. The bar graph displays the raw data (fold-changes)
937 for g*Thg1l*, where the three biological replicates are indicated by dots with different colors. *p <
938 0.05, Student's t-test. Source data in Table S3.

939 (B) Workflow for target validation in human NK cells *in vitro.*

940 (C) Successful editing and its effects on NK function for *THG1L*. Data are from one
941 representative experiment, with a biological replicate shown in Figure S4A. *THG1L* KO human
942 NK cells were incubated with K562 tumor cells for 4-6 hr before FACS analysis. The data show
943 that *THG1L* KO increased TNFα secretion in NK cells and enhanced NK cells mediated lysis,
944 but did not affect human NK degranulation, perforin or granzyme B. Dots represent technical

945 repeats (n=3). *$p < 0.05$, **$p < 0.01$, ***$p < 0.001$, ****$p < 0.0001$, Student's t-test.

946

Figure 1

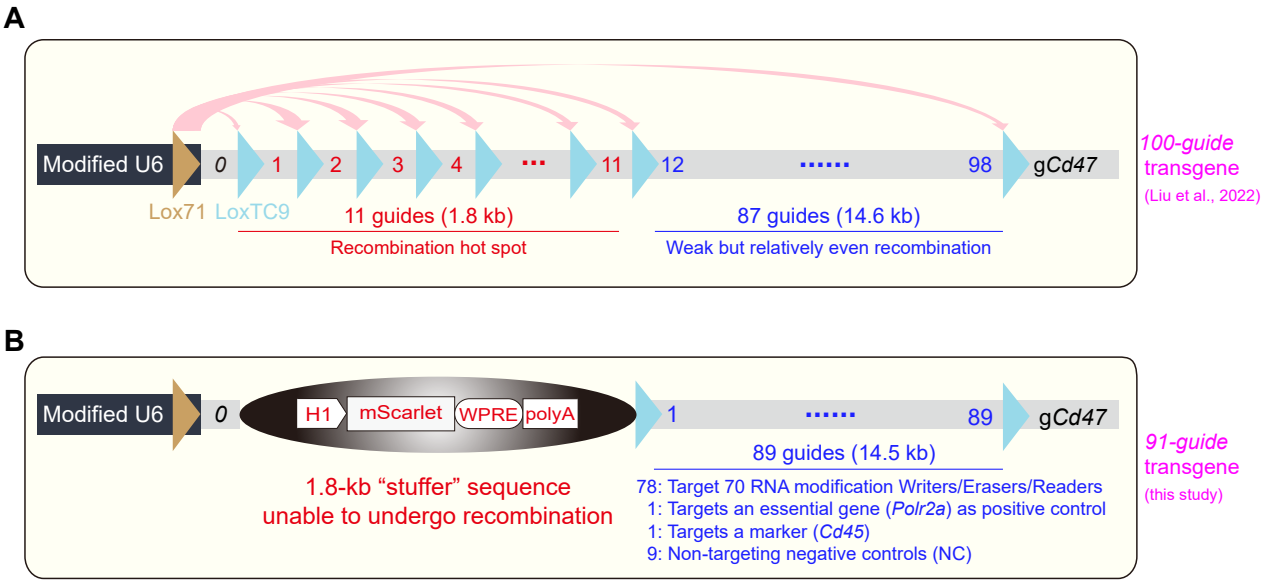
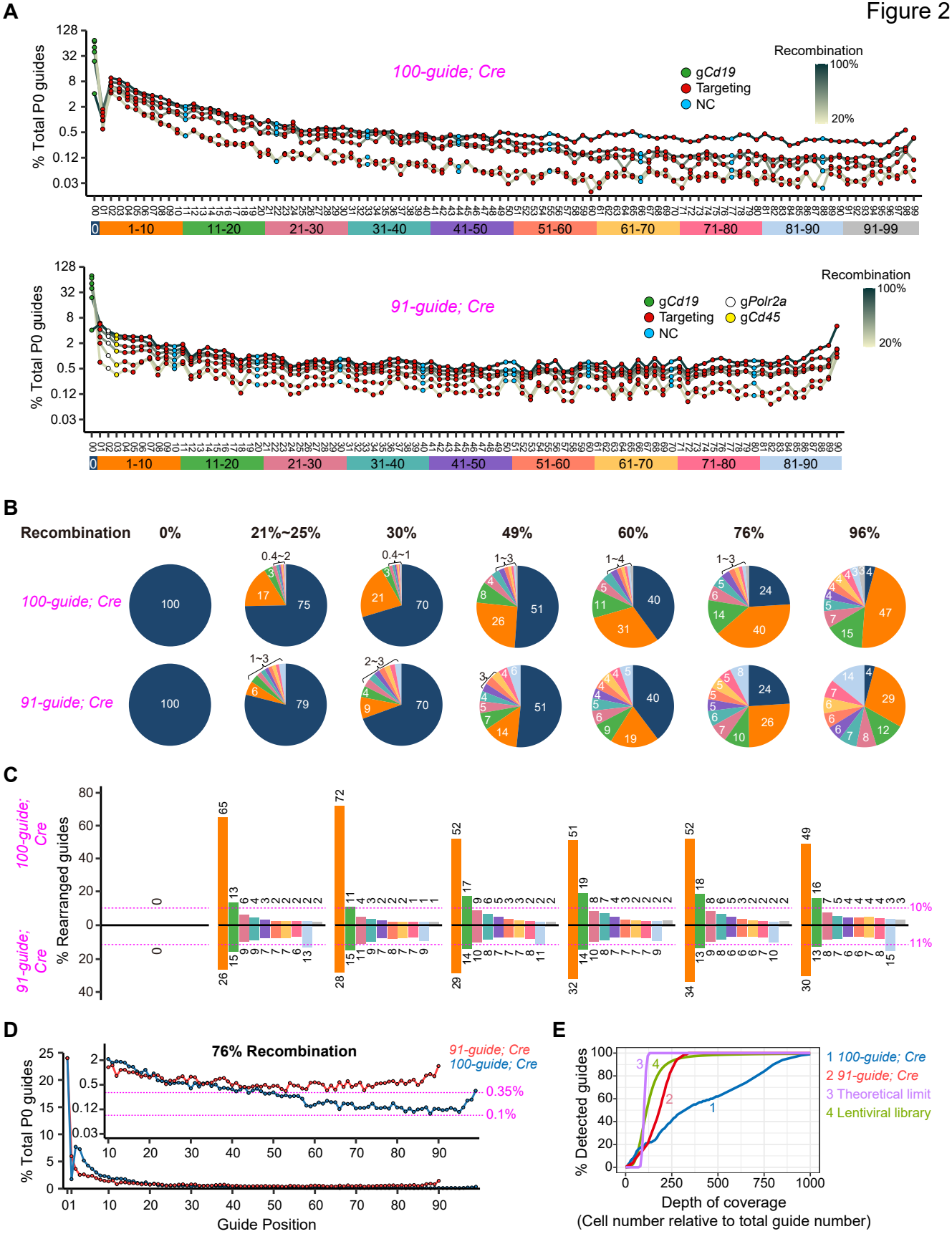

A
Modified U6
0
1
2
3
4
11
12
98
gCd47
Lox71
LoxTC9
11 guides (1.8 kb)
Recombination hot spot
87 guides (14.6 kb)
Weak but relatively even recombination
100-guide transgene
(Liu et al., 2022)
B
Modified U6
0
H1
mScarlet
WPRE
polyA
1
89
gCd47
89 guides (14.5 kb)
1.8-kb "stuffer" sequence
unable to undergo recombination
78: Target 70 RNA modification Writers/Erasers/Readers
1: Targets an essential gene (Polr2a) as positive control
1: Targets a marker (Cd45)
9: Non-targeting negative controls (NC)
91-guide transgene
(this study)

Figure 2


A
100-guide; Cre
% Total P0 guides
Recombination
100%
20%
gCd19
Targeting
NC
1-10
11-20
21-30
31-40
41-50
51-60
61-70
71-80
81-90
91-99
91-guide; Cre
gPolr2a
gCd45
B
Recombination
0%
21%~25%
30%
49%
60%
76%
96%
100-guide; Cre
91-guide; Cre
C
% Rearranged guides
10%
11%
D
76% Recombination
% Total P0 guides
Guide Position
0.35%
0.1%
E
% Detected guides
Depth of coverage
(Cell number relative to total guide number)
1 100-guide; Cre
2 91-guide; Cre
3 Theoretical limit
4 Lentiviral library

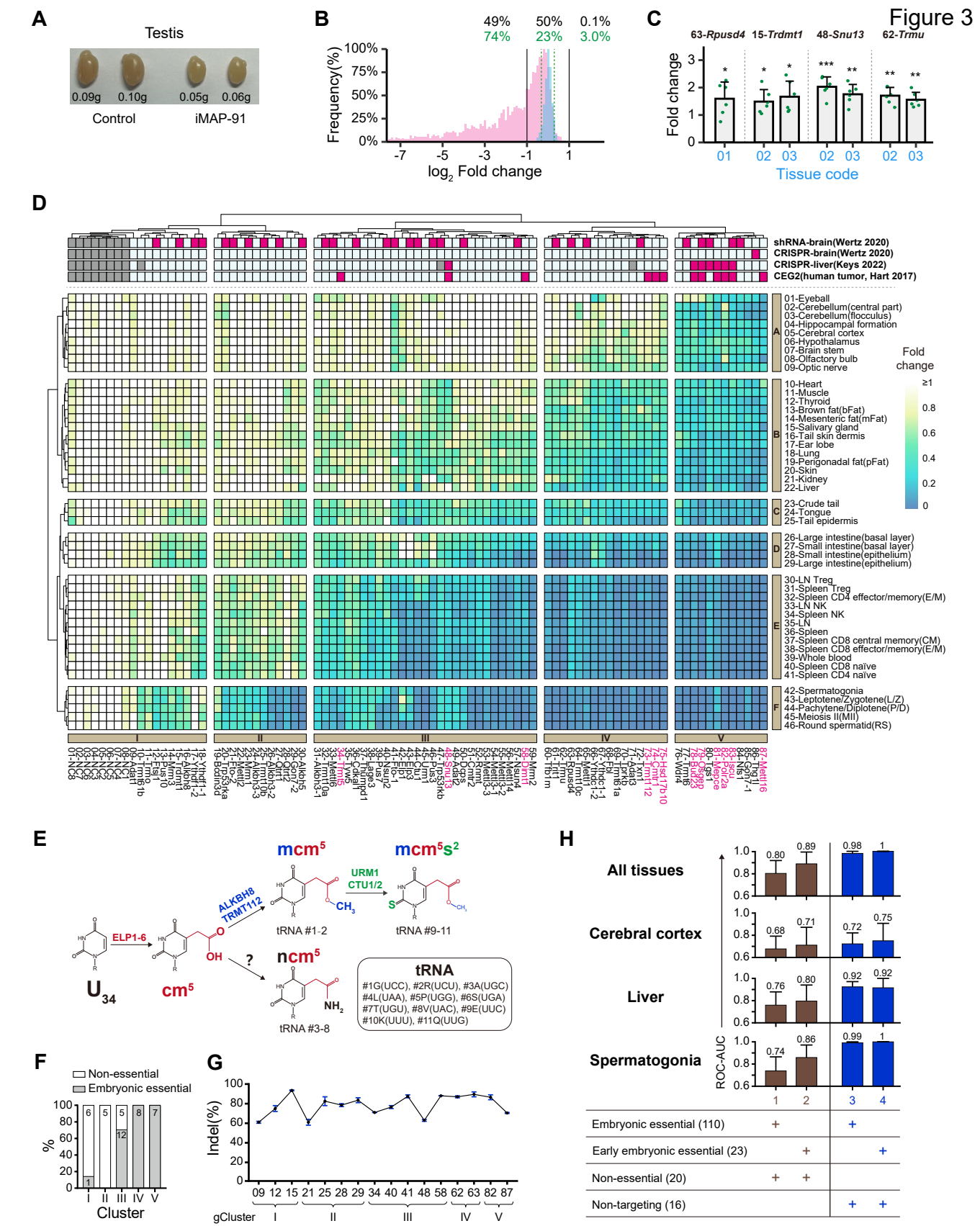


Figure 3
A
Testis
0.09g
0.10g
0.05g
0.06g
Control
iMAP-91
B
49%
50%
0.1%
74%
23%
3.0%
Frequency(%)
100%
75%
50%
25%
0%
-7
-5
-3
-1
1
log2 Fold change
C
63-Rpusd4
15-Trdmt1
48-Snu13
62-Trmu
Fold change
01
02
03
02
03
02
03
Tissue code
D
shRNA-brain(Wertz 2020)
CRISPR-brain(Wertz 2020)
CRISPR-liver(Keys 2022)
CEG2(human tumor, Hart 2017)
01-Eyeball
02-Cerebellum(central part)
03-Cerebellum(flocculus)
04-Hippocampal formation
05-Cerebral cortex
06-Hypothalamus
07-Brain stem
08-Olfactory bulb
09-Optic nerve
10-Heart
11-Muscle
12-Thyroid
13-Brown fat(bFat)
14-Mesenteric fat(mFat)
15-Salivary gland
16-Tail skin dermis
17-Ear lobe
18-Lung
19-Perigonadal fat(pFat)
20-Skin
21-Kidney
22-Liver
23-Crude tail
24-Tongue
25-Tail epidermis
26-Large intestine(basal layer)
27-Small intestine(basal layer)
28-Small intestine(epithelium)
29-Large intestine(epithelium)
30-LN Treg
31-Spleen Treg
32-Spleen CD4 effector/memory(E/M)
33-LN NK
34-Spleen NK
35-LN
36-Spleen
37-Spleen CD8 central memory(CM)
38-Spleen CD8 effector/memory(E/M)
39-Whole blood
40-Spleen CD8 naïve
41-Spleen CD4 naïve
42-Spermatogonia
43-Leptotene/Zygotene(L/Z)
44-Pachytene/Diplotene(P/D)
45-Meiosis II(MII)
46-Round spermatid(RS)
Fold change
≥1
0.8
0.6
0.4
0.2
0
I
II
III
IV
V
E
mcm5
mcm5s2
URM1
CTU1/2
ALKBH8
TRMT112
ELP1-6
U34
cm5
ncm5
tRNA #1-2
tRNA #9-11
tRNA #3-8
tRNA
#1G(UCC), #2R(UCU), #3A(UGC)
#4L(UAA), #5P(UGG), #6S(UGA)
#7T(UGU), #8V(UAC), #9E(UUC)
#10K(UUU), #11Q(UUG)
F
Non-essential
Embryonic essential
%
Cluster
G
Indel(%)
gCluster
H
All tissues
Cerebral cortex
Liver
Spermatogonia
ROC-AUC
Embryonic essential (110)
Early embryonic essential (23)
Non-essential (20)
Non-targeting (16)

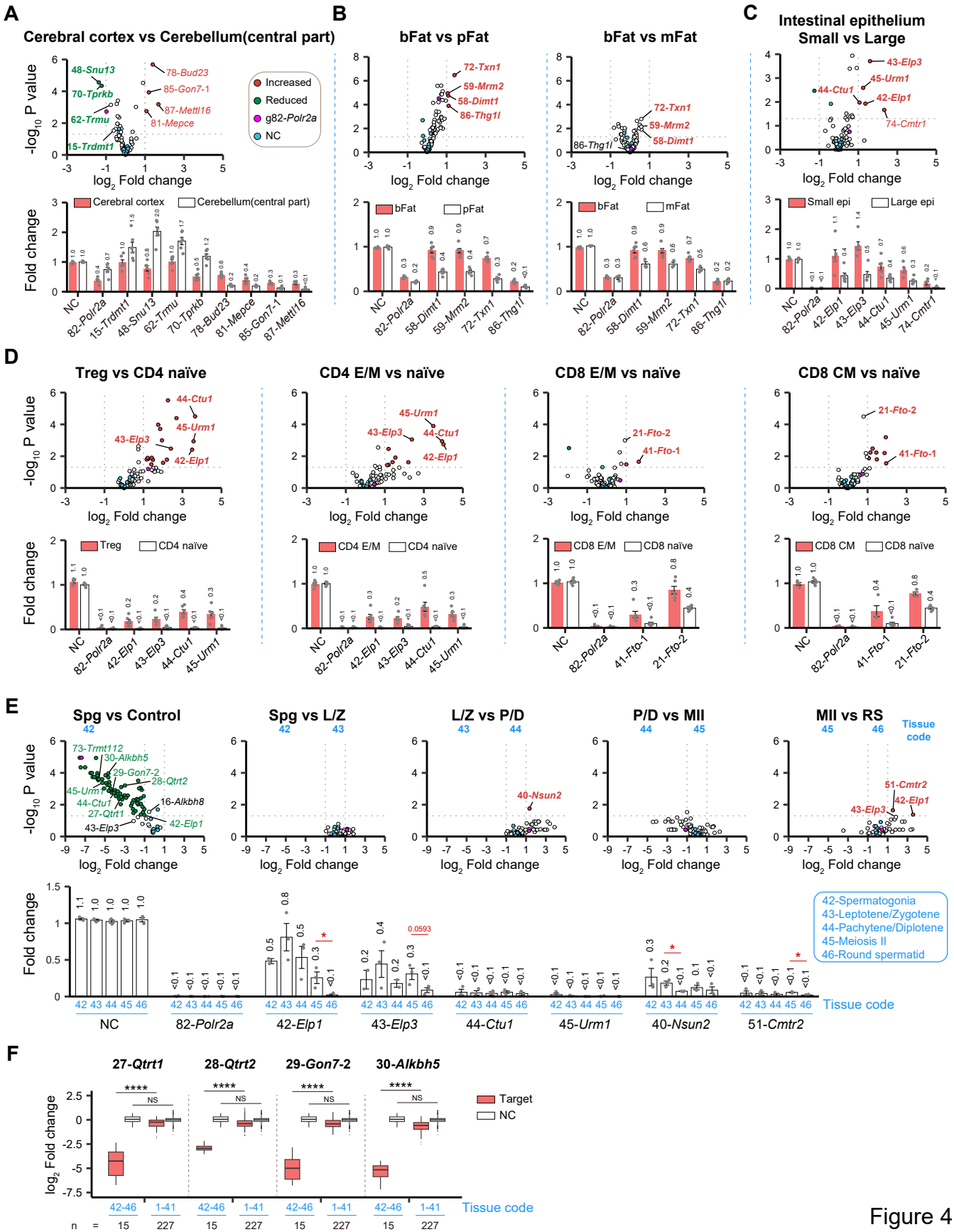
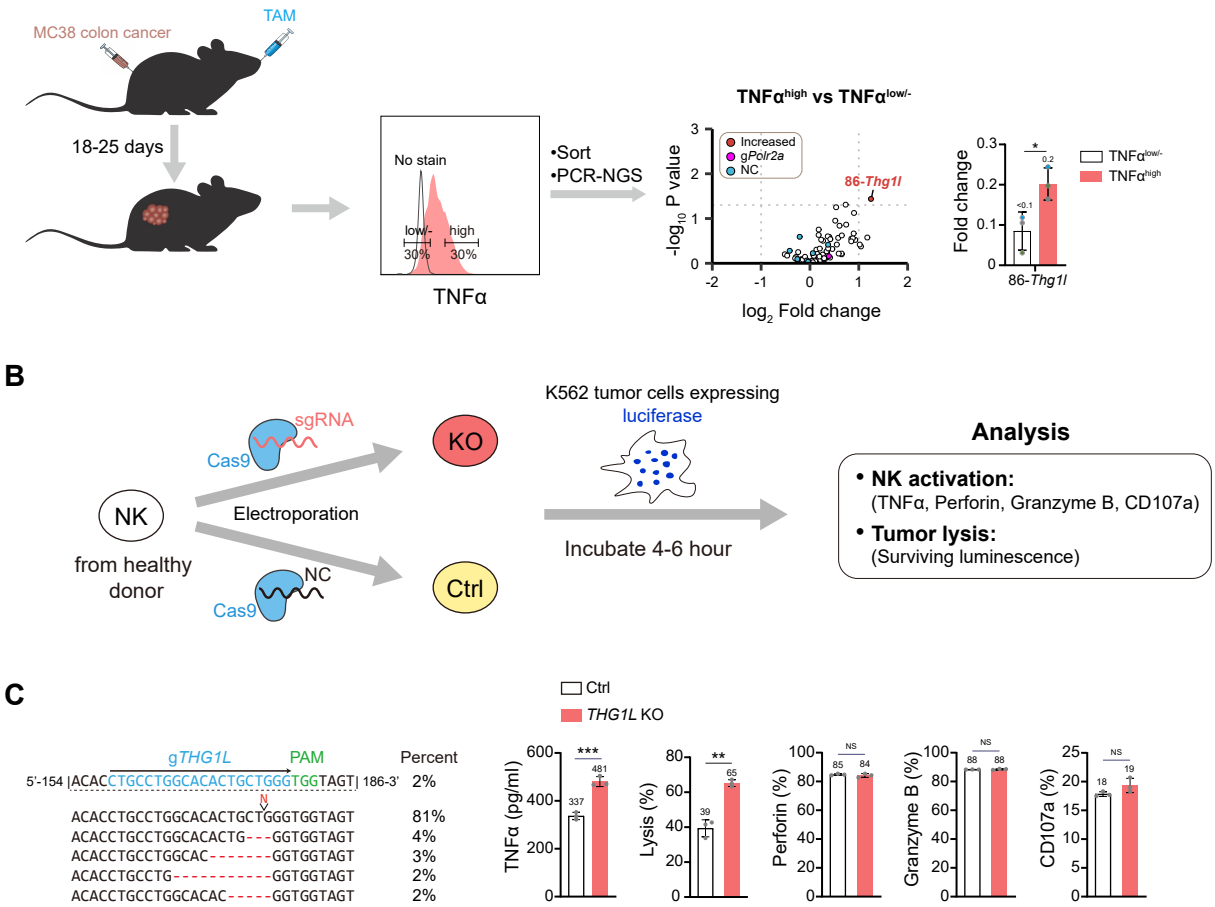
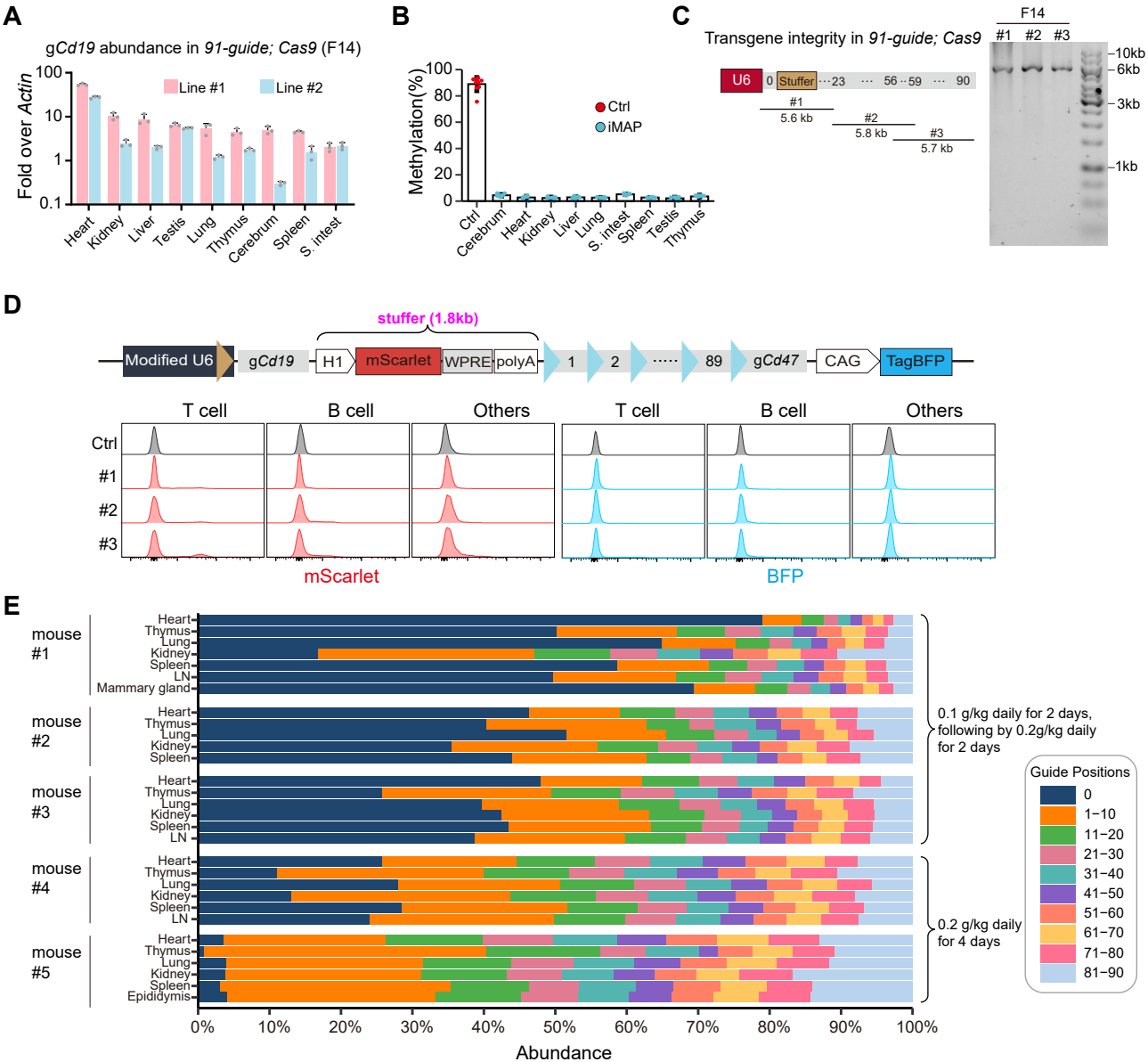
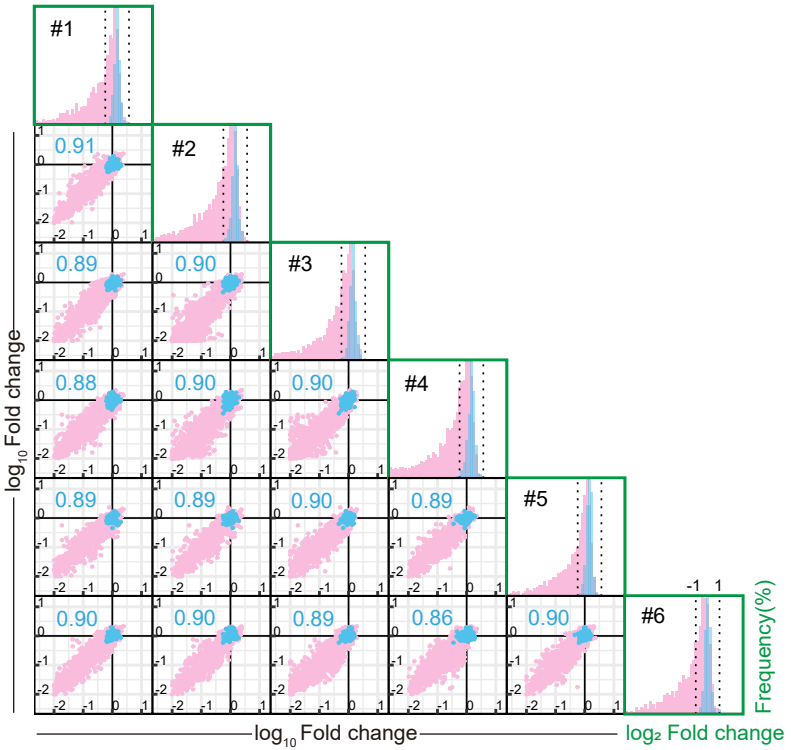

A
Cerebral cortex vs Cerebellum(central part)
-log10 P value
log2 Fold change
48-Snu13
70-Tprkb
62-Trmu
15-Trdmt1
78-Bud23
85-Gon7-1
87-Mettl16
81-Mepce
Increased
Reduced
g82-Polr2a
NC
Fold change
Cerebral cortex
Cerebellum(central part)
NC
82-Polr2a
15-Trdmt1
48-Snu13
62-Trmu
70-Tprkb
78-Bud23
81-Mepce
85-Gon7-1
87-Mettl16
B
bFat vs pFat
72-Txn1
59-Mrm2
58-Dimt1
86-Thg1l
bFat
pFat
82-Polr2a
58-Dimt1
59-Mrm2
72-Txn1
86-Thg1l
bFat vs mFat
mFat
C
Intestinal epithelium
Small vs Large
43-Elp3
45-Urm1
44-Ctu1
42-Elp1
74-Cmtr1
Small epi
Large epi
42-Elp1
43-Elp3
44-Ctu1
45-Urm1
74-Cmtr1
D
Treg vs CD4 naïve
44-Ctu1
45-Urm1
43-Elp3
42-Elp1
Treg
CD4 naïve
CD4 E/M vs naïve
CD4 E/M
CD8 E/M vs naïve
21-Fto-2
41-Fto-1
CD8 E/M
CD8 naïve
CD8 CM vs naïve
CD8 CM
E
Spg vs Control
Spg vs L/Z
L/Z vs P/D
P/D vs MII
MII vs RS
Tissue code
73-Trmt112
30-Alkbh5
29-Gon7-2
28-Qtrt2
45-Urm1
44-Ctu1
27-Qtrt1
16-Alkbh8
43-Elp3
42-Elp1
40-Nsun2
51-Cmtr2
42-Spermatogonia
43-Leptotene/Zygotene
44-Pachytene/Diplotene
45-Meiosis II
46-Round spermatid
0.0593
NC
82-Polr2a
42-Elp1
43-Elp3
44-Ctu1
45-Urm1
40-Nsun2
51-Cmtr2
F
27-Qtrt1
28-Qtrt2
29-Gon7-2
30-Alkbh5
****
NS
Target
NC
Tissue code
n =
15
227

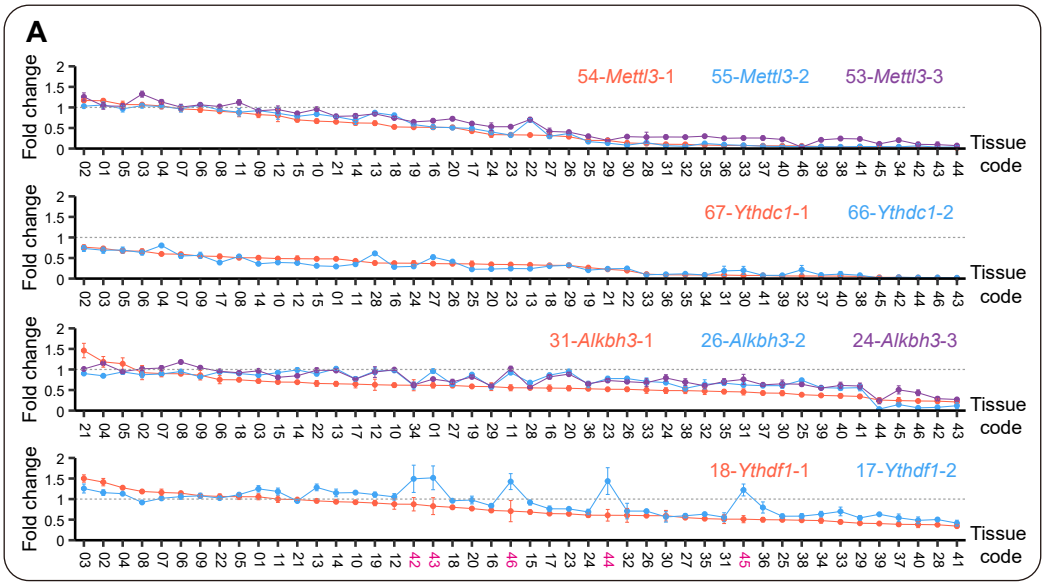
Figure 4

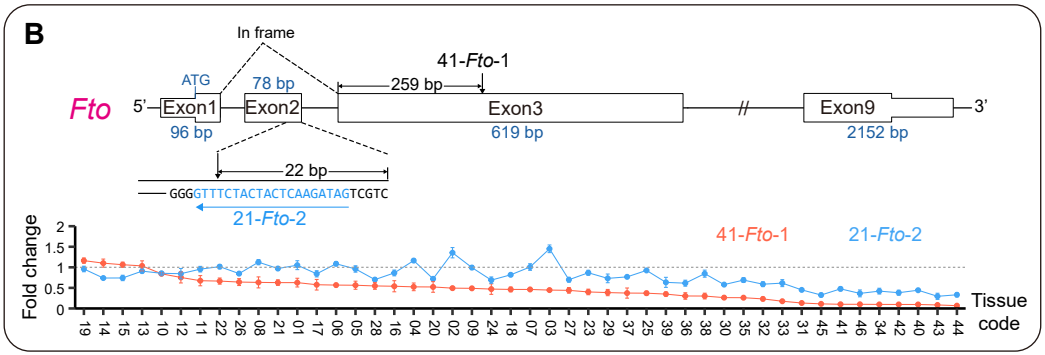
Figure 5

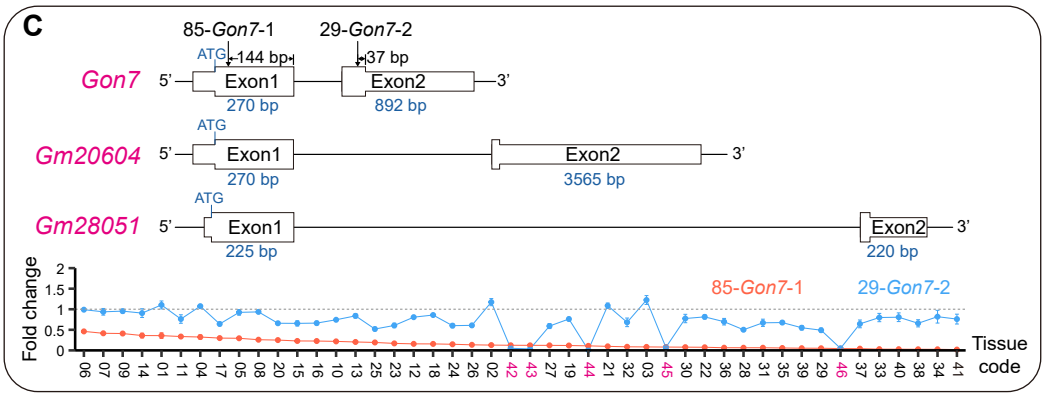

A
MC38 colon cancer
TAM
18-25 days
No stain
low/-
high
30%
30%
TNFα
•Sort
•PCR-NGS
TNFα^high vs TNFα^low/-
Increased
gPolr2a
NC
86-Thg1l
-log10 P value
log2 Fold change
Fold change
TNFα^low/-
TNFα^high
86-Thg1l
B
sgRNA
Cas9
KO
NK
from healthy donor
Electroporation
NC
Cas9
Ctrl
K562 tumor cells expressing
luciferase
Incubate 4-6 hour
Analysis
• NK activation:
(TNFα, Perforin, Granzyme B, CD107a)
• Tumor lysis:
(Surviving luminescence)
C
gTHG1L
PAM
Percent
5'-154 |ACACCTGCCTGGCACACTGCTGGGTGGTAGT| 186-3' 2%
N
ACACCTGCCTGGCACACTGCTGGGTGGTAGT 81%
ACACCTGCCTGGCACACTG---GGTGGTAGT 4%
ACACCTGCCTGGCAC-------GGTGGTAGT 3%
ACACCTGCCTG----------GGTGGTAGT 2%
ACACCTGCCTGGCACAC-----GGTGGTAGT 2%
Ctrl
THG1L KO
TNFα (pg/ml)
Lysis (%)
Perforin (%)
Granzyme B (%)
CD107a (%)
***
**
NS


A
gCd19 abundance in 91-guide; Cas9 (F14)
Fold over Actin
Line #1
Line #2
Heart
Kidney
Liver
Testis
Lung
Thymus
Cerebrum
Spleen
S. intest
B
Methylation(%)
Ctrl
iMAP
C
Transgene integrity in 91-guide; Cas9
F14
#1
#2
#3
U6
Stuffer
5.6 kb
5.8 kb
5.7 kb
10kb
6kb
3kb
1kb
D
stuffer (1.8kb)
Modified U6
gCd19
H1
mScarlet
WPRE
polyA
gCd47
CAG
TagBFP
T cell
B cell
Others
Ctrl
mScarlet
BFP
E
mouse #1
mouse #2
mouse #3
mouse #4
mouse #5
Mammary gland
Epididymis
0.1 g/kg daily for 2 days, following by 0.2g/kg daily for 2 days
0.2 g/kg daily for 4 days
Guide Positions
0
1−10
11−20
21−30
31−40
41−50
51−60
61−70
71−80
81−90
Abundance


#1
#2
#3
#4
#5
#6
0.91
0.89
0.90
0.88
0.90
0.90
0.89
0.89
0.90
0.89
0.90
0.90
0.89
0.86
0.90
log10 Fold change
log10 Fold change
log2 Fold change
Frequency(%)
-1
1



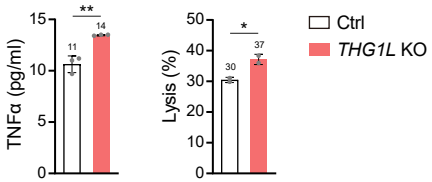




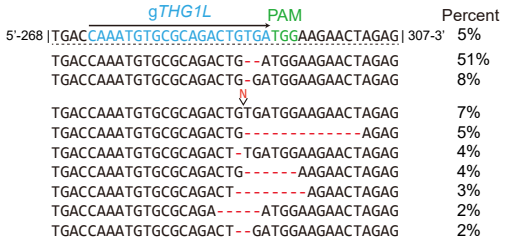
Tissue Code

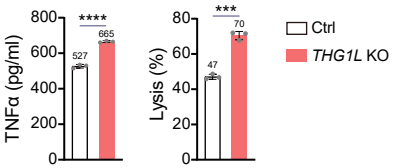
01-Eyeball
02-Cerebellum(central part)
03-Cerebellum(flocculus)
04-Hippocampal formation
05-Cerebral cortex
06-Hypothalamus
07-Brain stem
08-Olfactory bulb
09-Optic nerve
10-Heart
11-Muscle
12-Thyroid
13-Brown fat(bFat)
14-Mesenteric fat(mFat)
15-Salivary gland
16-Tail skin dermis
17-Ear lobe
18-Lung
19-Perigonadal fat(pFat)
20-Skin
21-Kidney
22-Liver
23-Crude tail
24-Tongue
25-Tail epidermis
26-Large intestine(basal layer)
27-Small intestine(basal layer)
28-Small intestine(epithelium)
29-Large intestine(epithelium)
30-LN Treg
31-Spleen Treg
32-Spleen CD4 effector/memory(E/M)
33-LN NK
34-Spleen NK
36-Spleen
37-Spleen CD8 central memory(CM)
38-Spleen CD8 effector/memory(E/M)
39-Whole blood
40-Spleen CD8 naive
41-Spleen CD4 naive
42-Spermatogonia
43-Leptotene/Zygotene(L/Z)
44-Pachytene/Diplotene(P/D)
45-Meiosis II(MII)
46-Round spermatid(RS)

A

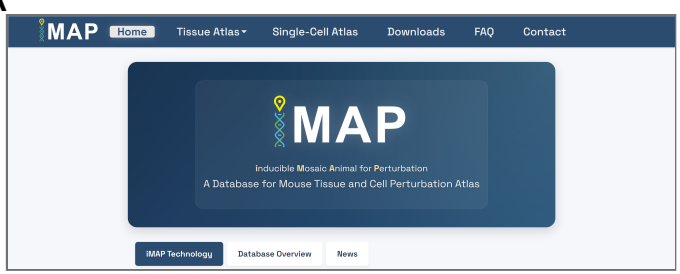

TNFα (pg/ml)
15
10
5
0
**
11
14
Lysis (%)
50
40
30
20
10
0
*
30
37
Ctrl
THG1L KO

B

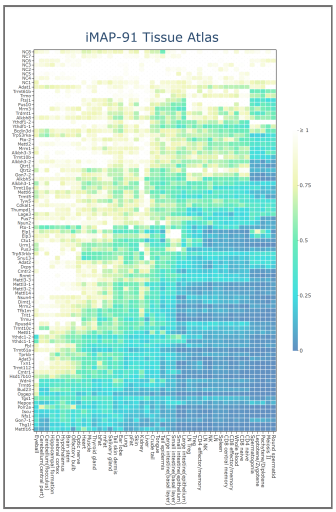

gTHG1L
PAM
Percent
5'-268 |TGACCAAATGTGCGCAGACTGTGATGGAAGAACTAGAG| 307-3' 5%
TGACCAAATGTGCGCAGACTG--ATGGAAGAACTAGAG 51%
TGACCAAATGTGCGCAGACTG-GATGGAAGAACTAGAG 8%
N
TGACCAAATGTGCGCAGACTGTGATGGAAGAACTAGAG 7%
TGACCAAATGTGCGCAGACTG----------AGAG 5%
TGACCAAATGTGCGCAGACT-TGATGGAAGAACTAGAG 4%
TGACCAAATGTGCGCAGACTG------AAGAACTAGAG 4%
TGACCAAATGTGCGCAGACT--------AGAACTAGAG 3%
TGACCAAATGTGCGCAGA-----ATGGAAGAACTAGAG 2%
TGACCAAATGTGCGCAGACT--GATGGAAGAACTAGAG 2%


TNFα (pg/ml)
800
600
400
200
0
****
527
665
Lysis (%)
80
60
40
20
0
***
47
70
Ctrl
THG1L KO

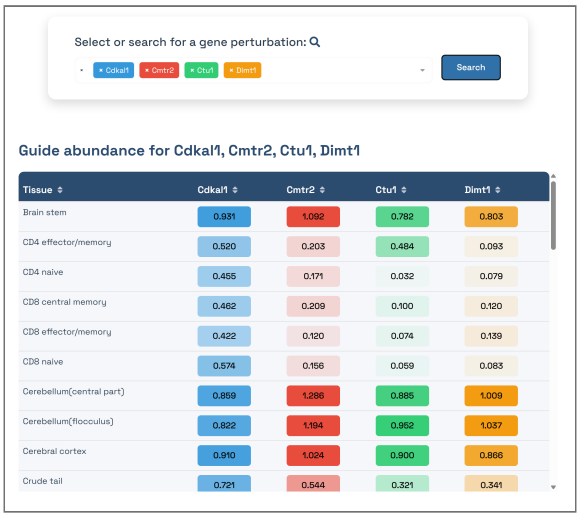
A



B

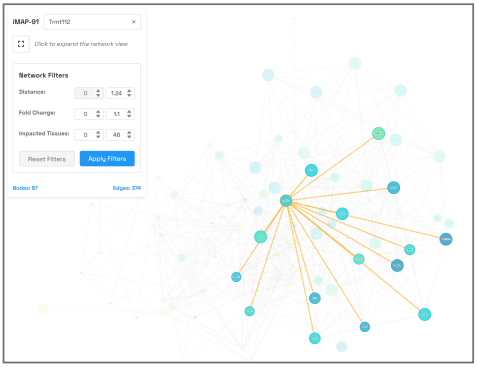


C

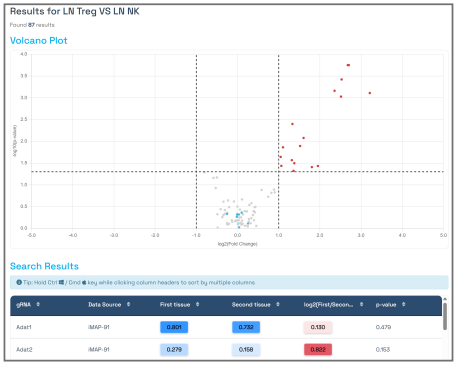


Tissue	Cdkal1	Cmtr2	Ctu1	Dimt1
Brain stem	0.931	1.092	0.782	0.803
CD4 effector/memory	0.520	0.203	0.484	0.093
CD4 naive	0.455	0.171	0.032	0.079
CD8 central memory	0.462	0.209	0.100	0.120
CD8 effector/memory	0.422	0.120	0.074	0.139
CD8 naive	0.574	0.156	0.059	0.083
Cerebellum(central part)	0.859	1.286	0.885	1.009
Cerebellum(flocculus)	0.822	1.194	0.952	1.037
Cerebral cortex	0.910	1.024	0.900	0.866
Crude tail	0.721	0.544	0.321	0.341

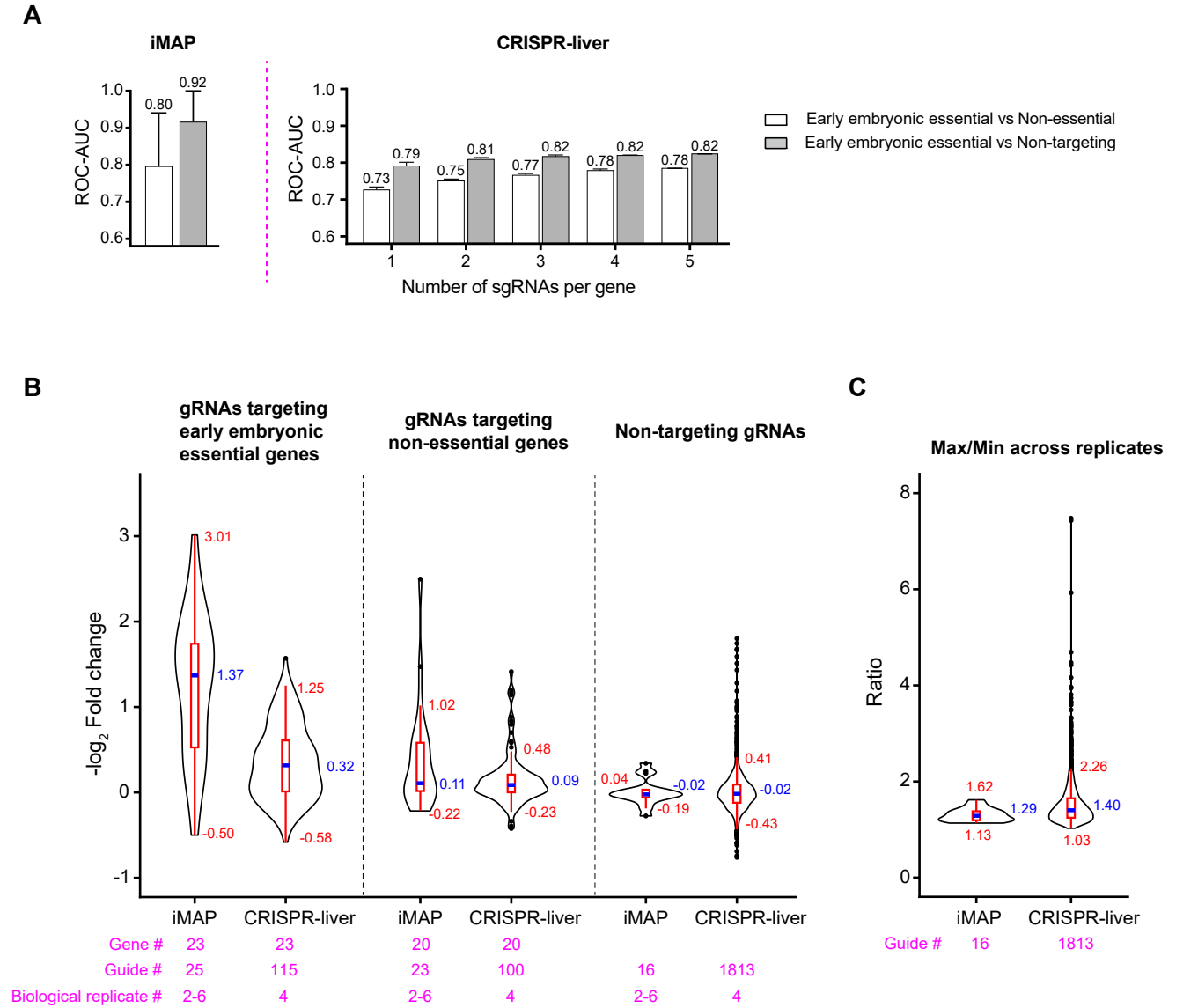
D



E



gRNA	Data Source	First tissue	Second tissue	log2(First/Secon...	p-value
Adat1	iMAP-91	0.801	0.732	0.130	0.479
Adat2	iMAP-91	0.279	0.158	0.823	0.153


A
iMAP
CRISPR-liver
ROC-AUC
1.0
0.9
0.8
0.7
0.6
0.80
0.92
0.73
0.79
0.75
0.81
0.77
0.82
0.78
0.82
0.78
0.82
1
2
3
4
5
Number of sgRNAs per gene
Early embryonic essential vs Non-essential
Early embryonic essential vs Non-targeting
B
gRNAs targeting early embryonic essential genes
gRNAs targeting non-essential genes
Non-targeting gRNAs
-log2 Fold change
3.01
1.37
-0.50
1.25
0.32
-0.58
1.02
0.11
-0.22
0.48
0.09
-0.23
0.04
-0.02
-0.19
0.41
-0.02
-0.43
iMAP
CRISPR-liver
iMAP
CRISPR-liver
iMAP
CRISPR-liver
Gene # 23 23 20 20
Guide # 25 115 23 100 16 1813
Biological replicate # 2-6 4 2-6 4 2-6 4
C
Max/Min across replicates
Ratio
1.62
1.29
1.13
2.26
1.40
1.03
iMAP
CRISPR-liver
Guide # 16 1813