TITLE

What uniparental genetic markers tell us about the prehistoric human

colonization of the Americas

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Running title

Uniparental markers and the American colonization

ABSTRACT

From the perspective of uniparental markers, the view of the human
prehistoric settlement of America is that it resulted from a single main
migration after the Last Glacial Maximum, following a period of prolonged
stasis in Beringia. These pioneers rapidly expanded across a continent
previously uninhabited by humans, forming isolated population groups that
became genetically distinct. Ancient DNA studies and whole genome analyses
have confirmed this view, revealing a remarkable complexity in the ancestries
of the involved populations.

In this work, I have reanalyzed all publicly available mitochondrial DNA and Y-chromosome haplogroups in the American population using simple phylogenetic and phylogeographic methodologies.

The results provide a genetic perspective opposed to the currently established model. The arrival of the American settlers occurred more than 30,000 years ago, preceding the LGM. As genomic studies have uncovered, at least two Asian populations contributed to the ancestry of the immigrant population. Low population density and climatic deterioration led to a long period of demographic eclipse, during which small bands of hunter-gatherers made long journeys in search of favorable niches. After the LGM, the climate improved, and demographic expansions occurred in multiple independent centers. The founding and expansion ages of the uniparental haplogroups indicate that these centers were in South America, particularly the Colombian isthmus, the Andean region, the Southern Cone, and the Amazon. Subsequent dispersals occurred in North America, one involving mitochondrial haplogroups A2, C1, and D1, but not B2, and another, of lesser magnitude, represented by the expansion of haplogroups C4c and X2a.

This work offers a previously unexplored model for the colonization of the Americas.

INTRODUCTION

The interpretation of the prehistoric peopling of the Americas has been a multidisciplinary enterprise (Becerra-Valdivia and Higham 2020; Fiedel 2000). From a molecular genetics perspective, the first foundations of these interpretations are based on the information provided by the uniparentally inherited mitochondrial DNA (mtDNA) and the non-recombining portion of the Y-chromosome (NRY) on extant populations (Schurr 2004; Karafet et al., 1999). As ancient DNA (aDNA) samples are a mixture of endogenous aDNA and contaminating DNA, results on human samples were first questioned. Fortunately, new sequencing technologies have overcome this problem (Palencia-Madrid and De Pancorbo 2015), and current aDNA-based analyses have added an invaluable timing perspective to these interpretations (Bisso-Machado and Fagundes 2021). Later, technological improvements have made it possible to sequence entire genomes in present-day and fossil individuals. Analyses of these genomes have revealed extinct ancestries and genetically well-differentiated ancestries from the past that, admixed in different proportions, persist in current Native American populations along the American continent. This new information has improved previous hypotheses and formulated new ones (Willerslev and Meltzer 2021).

Thus, some conclusions have achieved general acceptance: 1) The Asiatic origin of the Native American ancestors; 2) The first migration, in one or several waves, occurred as a rapid movement towards the south of the double continent; 3) The initial low population density led to strong genetic differentiation geographically structured; 4) Subsequent migrations reshaped the genetic landscape of North and South America. Necessary for the first was the entrance from Beringia of several migratory waves that originated the settlement of Paleo- and Neo-Eskimos; 5) Time comparisons of population genetic profiles since post-Neolithic to present-day have uncovered an essential genetic continuity (although see (Llamas et al. 2016)).

However, other propositions are still under debate, such as: 1) the precise Asian origin of the Native American ancestors; 2) the routes followed by these pioneers as they expanded throughout the Americas; and, perhaps, the most contentious question, 3) when these first migrations occurred? Although the most recent studies based on complete genomes favor a post-Glacial dispersal (Moreno-Mayar et al. 2018), with or without a prolonged standstill in Beringia (Faught 2017), the first works with mitochondrial DNA favored an early entry, around 30 kya before the Last Glacial Maximum (LGM), avoiding the problem of having to cross the ice sheets of North America (Bonatto and Salzano 1997; Torroni et al. 1994). These age discrepancies are due to the molecular evolutionary rates used in each case. Unfortunately, the rate of molecular evolution is not constant; it depends on several parameters, one of which is the adequate population size (Ne), which has changed dramatically from the Paleolithic to present-day (Cabrera 2020). Therefore, like radiocarbon dating, molecular rates must be calibrated to obtain more accurate ages. One way to do this is by using ancient genomes and/or mitogenomes, but in the case of the American colonization, all the extracted aDNA postdates the LGM (Bisso-Machado and Fagundes 2021). The option is to use evolutionary rates calibrated with Paleolithic fossils to date Paleolithic events (Cabrera 2024). Despite their utility, whole-genome studies barely mention uniparental markers (Gusareva et al. 2025). But see (Moreno-Mayar et al. 2018; Krettek et al. 2025). The underlying reason for this abandonment is that these markers only offer monogenic insights, and only the set of all genes provides congruent results. In my opinion, this assumption is flawed. The leading cause of genetic differentiation in estimating phylogenies and genetic distances between populations or species is isolation. Isolation affects whole genomes and does not discriminate by sex. Populations differentiate genetically, due to the loss of previous polymorphisms, by fixation or extinction, and the accumulation of new polymorphisms by mutation in a time-dependent way. Thus, any DNA fragment, including those coding for uniparental markers, should be valid to

realize genetic comparisons whenever they have accumulated enough genetic differentiation. The generalized use of DNA barcoding in molecular taxonomy (Hubert and Hanner 2015) validates my previous assertion. Isolation is broken by migration/introgression, affecting uniparental markers and autosomes differently. Whether uniparental markers persist only in their original populations or both endure in the original and recipient populations, their coalescence node—the distance to their most recent common ancestor—will remain unchanged. However, if the introgressed fragments are not thoroughly masked for mixed autosomes, they will influence the genetic distances obtained between the populations/species involved. A paradigmatic example is the controversial phylogenetic relationships obtained between modern humans and their closest relatives, Neanderthals and Denisovans. While mtDNA and NRY sequences showed modern humans and Neanderthals as sister lineages (Posth et al. 2017; Petr et al. 2020), autosomes joined Denisovan and Neanderthal as the closest pair, leaving modern humans as an out-group (Posth et al. 2017). Contrary to the widespread acceptance that the proper relationship between hominids is that obtained with the autosomes, I have suggested that the valid phylogeny is deduced from the uniparental markers. In contrast, the one obtained from the autosomes results from later introgressions between the three groups of hominins (Cabrera 2021b). Taking into account the above-mentioned particularities of the uniparental markers, in this work, I perform a thorough reanalysis of all published mitogenomes and the most recently published Y-chromosome results, using phylogenetic and phylogeographic tools, to reconsider and shed new light on the hypotheses formulated to explain the prehistoric colonization of the New World by Asian peoples. The main conclusions obtained are: 1) that the first migration involved at least two Asian populations with most probable central and eastern Asian origins; 2) that this wave(s) occurred before 30 Kya; 3) that those pioneer colonizers did not stop in Beringia but kept trekking across the Americas leaving few genetic traces in past and present-day native American populations; 4) That after the LGM, small and relatively isolated resilient groups radiated from different areas to occupy new exploitable territories both in North and South America; 5) that these secondary expansive waves occurred first in southern- an meso-America than in northern North America; and 6) that gene flow existed among geographically adjacent groups.

RESULTS

All the phylogenetic and phylogeographic mtDNA results described below have been obtained from the trees presented in the supplementary figures

(Supplemental_Fig_S1.xls (HgA2), Fig_S2.xls (HGB2), Fig_S3.xls (HgC1), Fig_S4.xls (HgD), and Fig_S5.xls (HgX2), and all the NRY results described below have been obtained from the trees already published by other authors (Wei et al. 2018; Grugni et al. 2019; Pinotti et al. 2019; Villaescusa et al. 2021; Colombo et al. 2022; Paz Sepúlveda et al. 2022).

The most probable origin of the American Asian ancestors
The main Native American mtDNA haplogroup founders can be divided into two groups: 1) those belonging to macro-Haplogroup M (C1, C4c, D1, D2a1, D4e1b, and D4h3a), and 2) those that belong to macro-Haplogroup N (A2, B2, and X2a). The same grouping occurs when we look at the number of mutations accumulated in their basal stems that link them to the common nodes with Asian lineages. While C1, C4c, D1, D2a1 and D4h3a have two mutations, and D4e1b only one; A2 and B2 have five mutations and X2a four mutations (Table 1).

Table 1. Mutations on American basal branches to their common nodes with Asia								
American haplogroup	Mutations	Coalescent Age (Kya)	Asian haplogroup	Asian region				
A2	5	20,2	A-a1	NW China (Xinjiang)				
A10a	1	4,04	A10	Western Siberia				
B2	5	20,2	B4b1a	Altai				

C1	2	8,08	C4	Eastern Asia				
C4c	2	8,08	C4a'b'c	Eastern Asia				
D1	2	8,08	D4	Eastern Asia				
D4e1b	1	4,04	D4e1	Eastern Asia				
D2a1	2	8,08	D2a'b	Northeastern Siberia				
D4h3a	2	8,08	D4h3	NW China (Xinjiang)				
X2a	4	16,16	X2a'j	Tajikistan				

An unpaired t-test shows that the difference between groups is highly significant (t = 7.2164; df = 5; two-tailed P value = 0.0008). Furthermore, it is well known that macro-Haplogroup M is dominant in southern and eastern Asia (Marrero et al. 2016), while macro-Haplogroup N is more frequent in western Eurasia (Fregel et al. 2015). Consequently, the same occurs with the Asian ancestral sequences/haplogroups that show the shortest distances with the Native American haplogroups. It has already been demonstrated that the closest relatives for the macro-Haplogroup M American lineages are found in eastern Asia/southeastern Siberia (Starikovskaya et al. 1998; Derenko et al. 2010). Some further commentaries are pertinent for these lineages: Hg C1 comprises four common and well spread sub lineages, three of them (C1b, C1c, C1d) practicably exclusive of the Americas and one (C1a) with a wide Asian range, and several rare clades/isolates, the most ancient of which is represented by two identical sequences from Brazil (Simão et al. 2021) and

Paraguay (Avila et al. 2019) that carries twelve mutations in its basal stem (Supplementary_Fig_S3.xls). Curiously, the other isolates have been detected in northwestern Eurasia instead of East Asia: C1e in Iceland (Ebenesersdóttir et al. 2011), western Russia (Järve et al. 2019), and western China (Wang et al. 2021); C1f in Tajikistan (Peng et al. 2018) and C1g in Mesolithic remains from western Russia (Der Sarkissian et al. 2014). As the oldest clades are found in the New World, the most parsimonious conclusion is that all the C1 branches in Eurasia resulted from retro migrations from America. The oldest Native American samples harboring C1b lineages are USR1 from Alaska (Tackney et al. 2015) and I11974 from Chile (Posth et al. 2018), both around 12,000 years old; for C1c it is MW291667 from Argentina (Roca-Rada et al. 2021), approximately 8,500 years old; and for C1d it is Lapa01, 9,600 years old, from Brazil (Posth et al. 2018). Haplogroup D1 has the oldest samples in Yucatan, Mexico, represented by a Late Pleistocene skeleton (HN5/48) around 12 ky old (Chatters et al. 2014), in Spirit Cave, Nevada, around 11 kya (Moreno-Mayar et al. 2018), an approximately 10 ky old LLP.S2.E1 sample from Argentina (Roca-Rada et al. 2021), and the Sumidouro4, 9,645 years old, sample from Brazil (Moreno-Mayar et al. 2018). Until recently, the American haplogroup D4h3a was separated from its Asian root by six mutations, but recent aDNA studies (Mao et al. 2021) have rescued a Chinese sequence that shares four of the six basal substitutions with the American clade (Supplementary_Fig_S4.xls). The remains, from which this sequence was extracted, were unearthed at the Amur region, in northeastern China, and are around 14,000 years old. The oldest samples harboring D4h3a sequences in the New World are the 12,600-year-old Anzick1 from Montana (Rasmussen et al. 2014), the 10,300-year-old Shuká Káa from Alaska (Lindo et al. 2017), and the 10,400-year-old specimens from Lagoa Santa in Brazil (Moreno-Mayar et al. 2018).

About the macro-Haplogroup N, for the New World haplogroup A2, the closest Asian sequences are from haplogroup A-a1 (BZL_M16A, BZL_M16B) and belong to historic samples from Xinjiang, western China (Wang et al. 2021). For haplogroup B2, it is AY519494, included in the Asian haplogroup B4b (Supplementary_Fig_S2.xls), with the Altai region as its geographic origin (Starikovskaya et al. 2005). Finally, for the American haplogroup X2a, the closest Asian relatives are into haplogroup X2j, that groups samples from Egypt and Europe (Supplementary_Fig_S5.xls) but, for geographic proximity, I consider as the closest Asian sequence a partial one (sample 40) from Tajikistan (Ovchinnikov et al. 2014), as it carries 16179 and 16357 substitutions that are terminal mutations in the Egyptian X2j (Kujanová et al. 2009) mitogenomes (Supplementary_Fig_S5.xls). For haplogroups A2 and B2 the most ancient sequences, around 14,000 years old, were extracted from human

coprolites found in Paisley Caves, Oregon (Gilbert et al. 2008), next oldest for A2 is Lapa15 (around 8,700 years old) from Lapa do Santo, Brazil (Posth et al. 2018), and for B2 is USR2 (approximately 12,000 years old) from Upward Sun River site in Alaska (Tackney et al. 2015). The most ancient Native American sample with an X2a haplotype is the Kennewick Man from Washington, which is around 9,000 years old (Rasmussen et al. 2015).

A way to reconcile the close arrivals of the Asian ancestors carrying the Native American M and N lineages with their significant differences in the number of mutations accumulated on their basal stems is to assume that the route followed across Asia of the N carriers was longer and slower than that of the M carriers.

Finally, if we compare the number of lineages carried by the M and N groups to their respective population sizes upon arrival in America, we could estimate that the contribution of the eastern Asian contingent was 62.5% and that of western Eurasia 37.5%.

The NRY does not show American colonization patterns as clear as those of mtDNA, but using its phylogeny and phylogeography in the Americas and Asia I have considered the ancestors of the American clades C-MPB373, that has its closest haplotypes in the eastern Asian outgroup clade C-L1373 (Pinotti et al. 2019; Sun et al. 2021), QF746 that includes the Q-YP1475 subclade represented by the Saqqaq ancient sample (Rasmussen et al. 2010) and has as sister subclade Q-B143 represented by northeastern Siberian Koryaks (Karmin et al. 2015), and Q-FT9713, that has its closest sequences in Mongolia and northeastern China (https://www.yfull.com/tree/Q) as of eastern Asian origin, while C-P39/Z30536, having as outgroup the Central Asian subclade C-F1756 (Wei et al. 2018), Q-Z780, that includes the Native American ancient sample Anzick1 (Rasmussen et al. 2014) and has Q-L330 of Central Asia ascription (Dulik et al. 2012; Flegontov et al. 2019) as sister clade, Q-M3, that has as out group Q-L804 with a northern European geographic range (Hallast et al. 2015), and the rare Q-Y1150 and Q-M378 (Grugni et al. 2019) as of western Eurasian origin. Unlike mtDNA, most American male lineages appeared to have their ancestors in western Eurasia. It is interesting to note that the C-MPB373 South American lineage is phylogenetically an outgroup of C-F1699 that comprises numerous western Eurasian lineages, including also the Canadian C-P39/Z30536 branch (Pinotti et al. 2019). Also of interest is that the most abundant American clade, Q-M3 (Colombo et al. 2022), has as sister branch Q-L804, which is specific to northern Europe (Grugni et al. 2019). The oldest sample belonging to the eastern group of lineages is Lapa01, from Brazil, which is approximately 8,840 years old (Posth et al. 2018). For the western group, Anzick1 from Montana, with an approximate age of 12,600 years, represents

the oldest North American sample (Grugni et al. 2019). In contrast, the oldest sample (I11974) in South America was found in Los Rieles, Chile, and is approximately 10,500 years old (Posth et al. 2018).

The global ages of the first arrival and expansions of the Native American parental ancestors

A global average of the different mean mutational data, calculated for each American mitochondrial haplogroup using various approaches (Table 2), give mean mutational values of 7.11 (95% CI: 6.34 – 7.88) substitutions at the arrival in America of the founding lineages, and a mean of 4.42 (95%: 3.98 – 4.85) substitutions for their initial expansions into the American continent, which, in turn, produce mean ages of 28.72 (95%: 25.61 – 31.84) kya and of 17.86 (95% CI: 16.08 – 19.59) kya for the founding and expansion events respectively.

Table 2. Mutational means and coalescent mean ages for the main American mtDNA haplogroups									
HG	Phylogeny	Mean (Age Kya)	Type	N	95% CI: Mutation/Age(Kya)				
A2	Founding	7.55 (30,50)	Normal	1119 (Indiv)	7.37-7.73 / 29,77-31,23				
	Expansion	4.63 (18,70)	"	1083 (Indiv)	4.46-4.79 / 18,02-19,35				

	Foundi ng	7.25 (29,29)	Weight ed	118 (Clades)	6.62-7.88 / 26,74-31,83			
	Expansi on	4.35 (17,57)	"	118 (Clades)	4.04-4.66 / 16,32-18,83			
	Coalesc ent	9.13 (36,88)	Max (19 nt)	16 (Branche s)	8.46-9.79 / 34,18-39,55			
B2	Foundi ng	6.68 (26,99)	Normal	1081 (Indiv)	6.51-6.85 / 25,30-27,68			
	Expansi on	4.68 (18.90)	"	1070 (Indiv)	4.51-4.85 / 18,22-19,00			
	Foundi ng	6.62 (26,75)	Weight ed	80 (Clades)	6.21-7.03 / 25,09-28,40			
	Expansi on	4.69 (18,95)	"	80 (Clades)	4.28-5.11 / 17,29-20,64			

	Coalescent	6.42 (25,93)	Max (16 nt)	14 (Branches)	5.12-7.72 / 20,68-31,17			
C1	Founding	6.50 (26,26)	Normal	887 (Indiv)	6.33-6.67 / 25.57-26,95			
	Expansion	4.36 (17,61)	"	873 (indiv)	4.17-4.55 / 16,84-18,38			
	Founding	6.33 (25,57)	Weighted	138 Clades)	6.05-6.61 / 24,44-26,71			
	Expansion	4.01 (16,20)	"	138 (Clades)	3.71-4.31 / 14,99-17,41			
	Coalescent	6.22 (25,13)	Max (16 nt)	42(Branches)	5.54-6.90 /22,38-27,88			
C4c	Founding	6.32 (25,53)	Normal	34 (Indiv)	5.74-6.91 /23,19-27,92			

	Expansion	4.32 (17,45)	"	34 (Indiv)	3.74-4.91 / 15,11-19,84
	Founding	3.48 (14,06)	Weighted	8 (Clades)	2.85-4.11 /11,51-16,60
	Expansion	1.30 (5,25)	"	8 (Clades)	0.67-1.93 /2,71-7,80
	Coalescent	4.71 (19,03)	Max (9 nt)	3 (Branches)	2.47-6.96 /9,98-28,12
D1	Founding	7.00 (28,28)	Normal	496 (Indiv)	6.73-7.27 / 27,19-29,37
	Expansion	4.97 (20,08)	"	463 (Indiv)	4.68-5.27 / 18,91-21,29
	Founding	7.02 (28,36)	Weighted	58 (Clades)	6.53-7.51/26,38-30,34

	Expansi on	4.97 (20,08)	"	58 (Clades)	4.48-5.46 / 18,10-22,06			
	Coalesc ent	7.14 (28,86)	Max (16 nt)	28(Branc hes)	6.67-7.62 / 26,93-30,79			
D4h 3a	Foundi ng	6.20 (25,05)	Normal	80 (Indiv)	5.57-6.83 / 22,50-27,59			
	Expansi on	4.24 (17,13)	"	64 (Indiv)	3.63-4.85 /14,67-19,59			
	Foundi ng	6.27 (25,33)	weight ed	13 (Clades)	5.73-6.81 / 23,15-27,51			
	Expansi on	4.24 (17,13)	"	13 (Clades)	3.80-4.68 / 15,35-18,91			
	Coalesc ent	7.77 (31.39)	Max (12 nt)	12(Branc hes)	6.15-9.39 / 24,85-37,93			
X2a	Foundi	7.83	Normal	41 (Indiv)	7.11-8.55 / 28,72-			

ng	(31,63)			34,54				
Expansion	3.83 (15,47)	"	40 (indiv)	3.11-4.55 / 12,56-18,38				
Founding	6.83 (27,59)	Weighted	2 (Clades)	5.37-8.29 /21,69-33,49				
Expansion	2.83 (11,43)	"	2 (Clades)	1.37-4.29 / 5,53-17,33				
Coalescent	11.00 (44,44)	Max (15 nt)	2(Branches)	-39.82-61.82/-160,87-249,75				

The founding ages for the mtDNA Native American haplogroups, exposed above, coincide with the approximately 30 kya proposed in the first mtDNA studies (Bonatto and Salzano 1997; Torroni et al. 1994; Starikovskaya et al. 1998), and with the calibrated ages calculated by (Rieux et al. 2014), based on ancient mitogenomes. However, recent extant and ancient mitogenome studies (Bisso-Machado and Fagundes 2021; Llamas et al. 2016) and whole genome-based studies (Moreno-Mayar et al. 2018; Willerslev and Meltzer 2021) strongly support post-LGM ages for the settlement of the Americas,

although this is in part inspired by the most conservative archaeological ages

proposed for the American settlement.

The age for the first population expansions calculated here, 17.86 kya, after the rigors of the LGM had lessened on the continent, does not differ much from those proposed by most authors. The difference is that this expansion coincides with most authors' arrival in America proper. In contrast, here, the arrival in America occurred much earlier, around 30 kya, before the glacial period had its worst effects.

Concerning the NRY, it deserves mention that, assuming a first colonization of the Americas about 15 kya (Poznik et al. 2013), estimated a substitution rate for the Y-chromosome of 8.2 x 10-10 (95% CI: 7.2 x 10-10 – 9.2 x 10-10) per site, year. However, doubling the time to 30 kya, as proposed here based on mtDNA, the substitution rate would be 4.1 x 10-10 per site, year which is very similar to that obtained by (Francalacci et al. 2013) of 5.3 x 10-10 (95% CI: 4.2 X10-10- 7.0 x 10-10) per site, year and both estimates falling within the limits of the mean 4.7 x 10-10 (95% CI: 2.1 X 10-10 – 9.5 X 10-10) per site, year proposed by myself based on past population sizes obtained under phylogenetic considerations (Cabrera 2024). As before, these results contrast with the majority of the NRY younger coalescence ages estimated by other authors (The Genographic Consortium et al. 2016; Pinotti et al. 2019; Colombo et al. 2022; Paz Sepúlveda et al. 2022).

As YFull Tree v13.03.00 (YTree) has the most actualized NRY phylogeny and the most extensive phylogeography, it is increasingly used as a data source in scientific articles, as in the present work. The YFull substitution rate (Adamov et al. 2015) is identical to the one calculated by (Poznik et al. 2013) under the premise that the human American settlement occurred around 15 kya, because of this, I have halved their substitution rate values when I used them. Under these requirements, NRY haplogroup C clades would arrive in the Americas in separate waves, C-MPB373 first at about 32.2 kya (95% CI: 28.8 – 35.6 kya), and C-P39/Z30536 second at about 26.2 kya (95% CI: 22.8 – 29.8 kya). The three main NRY haplogroup Q clades have very similar ages around 30 kya: Q-F746, 31.8 kya (95% CI: 28.4 – 35.2); Q-Z780, 31.0 kya (95% CI: 29.0 – 33.0); Q-M3, 29.8 (27.0 – 32.8). The age of arrival to the Americas of the rare clades Q-M1150 (30.0 kya (95% CI:26.6 – 33.4 kya) and Q-M378, 21.4 kya (95% CI: 17.6 – 25.4 kya), that include European and Asian lineages, is only tentative (https://www.yfull.com/tree/Q).

Regional heterogeneity for founding and expansion events

Initially, I compared the founding and expansion ages of native haplogroups between North and South American subcontinents (Table 3). Except for haplogroup C1, the results show significantly older ages in South America for both founding and expansion events.

Table 3. Differences in Founding(F) and expansion(E) ages between the North America and South America subcontinents					
Haplogroup	t-value	df	p-value	oldest region	
A2 - F	3.0397	1119	0.0024	South America	
A2 -E	4.8326	1105	<0.0001	South America	
B2 - F	1.0604	1099	0.2892	ns	
B2 - E	3.1395	1083	0.0017	South America	
C1 - F	1.4777	499	0.1401	ns	
C1- E	0.4722	482	0.6370	ns	

D1 - F	5.7700	498	<0.0001	South America								
D1 - E	2.2473	466	0.0251	South America								
D4h3a - F	3.0245	69	0.0035	South America								
D4h3a - E	1.0115	59	0.3159	ns								

More detailed comparison between geographic regions, within and among subcontinents, using mean molecular age comparisons, pairwise genetic distances and Pearson correlation tests are described in supplementary tables (Supplemental Table S1.1 to S1.7 for A2; Supplemental Table S2.1 to S2.7 for B2; Supplemental Table S3.14, .7, .10, .13, .16 for C1b; Supplemental Table S3.2, .5, .8, .11, .14, .17 for C1c; Supplemental Table S.3.3, .6, .9, .12, .15, .18 for C1d; Supplemental Table S3.19 for C4c; Supplemental Table S4.1 to S4.6 for D1; Supplemental Table S4.7 to S4.9 for D4h3, and Supplemental Table S5.1 to S5.3 for X2a).

Haplogroup A2
Globally, A2 is the most abundant haplogroup, representing approximately 44% of the Native American population, but it has an uneven geographic distribution. The oldest founding ages for A2 (Supplemental_Table_S1.5) are in Brazil and the Colombia-Isthmus area (≈ 34 kya), expanding first in Colombia around 23 kya. From the Colombian Isthmus region, significant latitudinal clines exist to Siberia (Slope -13.37; R^2 0.8396; P-value 0.0102), and to Tierra del Fuego (Slope 23.49; R^2 0.8091; P-value 0.0377). In addition, notice that in the most ancient samples of southern South America, mtDNA haplogroup A and B lineages were totally absent or very rare (Lalueza 1997; Moraga et al. 2000; Nakatsuka et al. 2020; Roca-Rada et al. 2021; Postillone et al. 2020) while they were detected in the most ancient samples recovered in northwestern North America more than 14 kya (Gilbert et al. 2008), and also were prominent in ancient samples from the Isthmus-Colombian area

(Capodiferro et al. 2021). Another geographic peculiarity of haplogroup A2 is its significant abundance in the eastern regions compared to the western ones (Kemp et al. 2007; Raff et al. 2011; Rothhammer et al. 2017; Simão et al. 2021). Notably, the oldest A2 haplotypes are found in North America and Brazil (Table S1.4). Furthermore, a secondary southward migration of some North American sub-lineages is also detectable. One of these lineages is A2f, which is significantly older in North America (≈35,000 ya) than in Mexico (≈23,000 ya) (t=3.6172; df 22; p-value 0.0015).

For haplogroup A2, the shortest molecular genetic distances between regions (Table S1.3) were found for the pairs CSAm–SSAm (0.657), NWSAm–NESAm (0.682), and NCAm–NNAm (0.681). When these distances are shown in a PCoA (Fig. 1), the first principal coordinate places NESAm in a central position, with the remaining regions of North- and South-America positioned to its left and right, respectively. The second principal coordinate separates the areas of each subcontinent as latitudinal clines, reflecting the Pearson correlation results.

Haplogroup B2

Haplogroup B2 is the second-largest Native American haplogroup, comprising approximately 24.0% of the global distribution. The oldest founding (approximately 30,000 years ago) and expansion (approximately 21,000 years ago) regional values for this haplogroup are found in South and Central America. As one moves northward from these southern regions, the values decrease in a clinal trend, following a northwest-southeast orientation to reach Central and North America. A significant negative correlation exists between

the molecular ages of haplogroup B2 and latitude (Slope: -13.57; R^2: 0.5455; P-value: 0.0061).

Unlike haplogroup A2, haplogroup B2 is either absent or rare in northern North America, which mirrors its low frequency in northeastern Siberia (Starikovskaya et al. 1998). Moreover, in contrast to haplogroups A2 and D1, haplogroup B2 is more prevalent in western continental regions than in eastern ones (Bisso-Machado et al. 2012; Brandini et al. 2018). The oldest B2 haplotypes have been identified in the Andean region.

The shortest genetic molecular distances were observed between the South South American (SSAm) and Central South American (CSAm) groups (0.492), as well as between the SSAm and Northwest South American (NWSAm) pairs (0.500). A Principal Coordinates Analysis (PCoA) represented these distances graphically, showing NWSAm in a central position, with SSAm and CSAm on the left and Northeastern South American (NESAm) and all northern American regions on the right. This arrangement follows a latitudinal cline, as confirmed by the Pearson correlation (Fig. 2). (Supplementary_Table_ S2.3). A PCoA bidimensional graphic representation of these distances shows NWSAm having a central position, with SSAm and CSAm at its left and NESAm and all northern American regions at its right, following a latitudinal cline as observed with the Pearson correlation (Fig. 2).

Haplogroup C1

Haplogroup C1 was initially considered a homogenous clade; however, its three main native branches reflect distinct phylogeographic movements that warrant particular attention.

For haplogroup C1b, the oldest founding and expansion regional ages (see Supplementary Table S3.13) are found in Northeast South America

(approximately 29,000 to 17,000 years ago), with younger ages extending down to Tierra del Fuego and up to Canada. The data shows a significant trend (Slope -12.20; R^2 0.8876; P-value 0.0166). These phylogeographic characteristics are somewhat similar to those of haplogroup A2. However, the longitudinal differentiation seen in the A2 clade is not present in C1b. The oldest haplotypes for C1b have been identified in northern South America, particularly in Ecuador and Colombia, and extend into Mexico (see Supplementary Table S3.10). Additionally, the shortest genetic distance between regions (refer to Supplementary Table S3.7) is observed between Northwest South America (NWSAm) and Northeast South America (NESAm) with a distance of 0.522, followed by NWSAm and Southern South America (SSAm) at 0.605, and SSAm to Central South America (CSAm) at 0.654.

In this context, the PCoA (Principal Coordinates Analysis) graphic representation emphasizes the close relationship between NWSAm–Mexico and the USA–Canada regions, positioning NESAm as the central point from which movements occurred both southward across Brazil and northward into Central America (see Fig. 3).

For Hg C1c, the earliest regional ages associated with founding and expansion (Supplementary_Table_ S3.14) are centered in NWSAm (≈ 34 kya / 26 kya). From this center, populations with younger ages follow southern and northern geographic transects ending in SSAm and Canada, respectively. The latter shows a significant correlation with latitude for both founding (Slope -14.79; R^2

0.8055; P-value 0.0152) and expansion ages (Slope -11.71; R^2 0.9629; P-value

0.0005). These results partially resemble those obtained for Hg B2 in its North

American subcontinent range. Congruent with its Andean center, the oldest

haplotypes for C1c are found in NWSAm and Mexico (Supplementary_Table_

S3.11). The shortest genetic distances within Hg C1c

(Supplementary_Table_S3.8) indicate the existence of three geographic

centers: the first represented by the CSAm-CAm pair (0.571), the second by the

SSAm-NWSam pair (0.630), and the third by the NWSAm-Mexico pair (0.644).

This pattern could be explained by supposing NWSAm as the leading center

that spread downward to Argentina and Chile and upward to Mexico, and a

later center that, from Brazil, reached Central America, erasing the signal of

the previous route from NWSAm to Mexico in this region. In this respect, it is

worth mentioning that the Y-chromosome Q-M925, mainly Mesoamerican, has

close phylogenetic ties with the Brazilian Q-Y26547 subclade (Colombo et al.

2022). This complex pattern is partially reflected in the PCoA plot (Fig. 4),

which shows a latitudinal cline from NWSA to Canada passing through NESAm,

with the unusual inclusion of SSAm in it, and an independent close relationship

between Central America and Brazil.

For Hg C1c, the oldest regional ages for founding and expansion

(Supplementary_Table_S3.14) are centered in NWSAm (≈ 34 kya / 26 kya).

From this center, populations with younger ages extend along southern and

northern geographic transects ending in SSAm and Canada, respectively. The

latter shows a significant correlation with latitude for both founding (Slope -

14.79; R^2 0.8055; P-value 0.0152) and expansion ages (Slope -11.71; R^2 0.9629;

P-value 0.0005). These findings somewhat resemble those seen for Hg B2

across its North American range.

Congruent with its Andean origin, the oldest haplotypes for C1c are found in

NWSAm and Mexico (Supplementary_Table_S3.11).

The shortest genetic distances within Hg C1c (Supplementary_Table_S3.8_

S3.8) indicate the existence of three geographic centers: the first represented

by the CSAm-CAm pair (0.571), the second by the SSAm-NWSam pair (0.630),

and the third by the NWSAm-Mexico pair (0.644). This pattern could be explained by assuming NWSAm as the primary center that spread downward to Argentina and Chile and upward to Mexico, with a later center originating from Brazil that reached Central America, obscuring the previous route from NWSAm to Mexico. Notably, the Y-chromosome Q-M925, mainly Mesoamerican, has close phylogenetic ties with the Brazilian Q-Y26547 subclade (Colombo et al. 2022). This complex pattern

The oldest ages for the founding and expansion of haplogroup C1c (Hg C1c), as shown in Supplementary Table S3.14, are centered in Northwest South America (NWSAm), estimated around 34,000 to 26,000 years ago. From this central point, younger populations form geographic transects extending south to Southern South America (SSAm) and north to Canada. The data show a significant correlation with latitude for both founding (slope: -14.79; R^2: 0.8055; P-value: 0.0152) and expansion ages (slope: -11.71; R^2: 0.9629; P-value: 0.0005). These results resemble those observed for haplogroup B2 in its North American distribution. Consistent with its Andean origin, the oldest haplotypes of C1c are found in NWSAm and Mexico (as shown in Supplementary Table S3.11). Analysis of genetic distances within Hg C1c, detailed in Supplementary Table S3.8, reveals three geographic centers: the first is represented by the Central South America-Central America (CSAm-CAm)

pair (distance: 0.571), the second by the Southern South America-Northwest South America (SSAm-NWSAm) pair (distance: 0.630), and the third by the Northwest South America-Mexico (NWSAm-Mexico) pair (distance: 0.644). This pattern suggests NWSAm may be the original center of expansion, spreading southward to Argentina and Chile, and northward to Mexico. A later center, likely originating from Brazil, may have reached Central America, obscuring evidence of the earlier route that linked NWSAm to Mexico. Notably, the Y-chromosome Q-M925, primarily found in Mesoamerica, shows close phylogenetic relationships with the Brazilian Q-Y26547 subclade (Colombo et al. 2022). This complex pattern is partly demonstrated in the Principal Coordinates Analysis (PCoA) plot (Fig. 4), which displays a latitudinal gradient from NWSAm to Canada, passing through Northeast South America (NESAm), while also revealing the unexpected inclusion of SSAm. Additionally, there is a clear close relationship between Central America and Brazil.

Hg C1d, with the oldest founding and expansion ages

(Supplemental_Table_S3.15) in CSAm and SSAm (approximately 38 kya / 33

kya), is the oldest C1 branch. In fact, it is the oldest Native American clade.

Although not reaching statistical significance, C1d shows a geographic trend of

decreasing population ages from North America to Central America

(Supplemental_Table_S3.15). This pattern resembles the one observed for Hg

A2. Additionally, it exhibits a significant latitudinal correlation from SSAm to

Central America, with the oldest founding (Slope -16.65; R² 0.7649; P-value

0.0415) and expansion ages (Slope -13.29; R² 0.6985; P-value 0.0382) in

southern South America. This geographic trend within the South American

subcontinent mirrors the pattern seen in Hg B2. The oldest C1d haplotypes are

found in CSAm and SSAm (Supplemental_Table_S3.12), supporting the idea of

an ancestral southern South American center of expansion for this clade. For

C1d, the closest genetic distances (Supplemental_Table_S3.9) are between the

SSAm-CSAm pair (0.591) and the NWSAm-NESAm pair (0.592). The latter

reflects the close genetic relationship between these regions, also seen in the

C1b clade. The PCoA bi-dimensional representation of these distances (Fig. 5)

shows a central group linking SSAm and CSAm with Central America, and two

distant groups: one on the left joining NWSAm and NESAm, and another on the

right grouping the northern North American populations and Mexico. The

independent and close genetic relationship between the South American

regions and Central America resembles what was found for C1c, which was

explained by a later expansion from Brazil to Central America, while the close

genetic affinity between NWSAm and NESAm mirrors what was observed in

C1b.

Haplogroup C4c

The founding and expansion ages for C4c are approximately 25 and 17,000

years ago, respectively (Table 2), but ages for its main branches vary

significantly (Supplemental_Table_S3.19). The most developed branch, C4c1,

has founding and expansion ages of about 16 and 12,000 years ago, both

following the LGM. In contrast, the less common C4c2 sister branch has a

founding age of 31 years ago, aligning more closely with other main Native

American haplogroups. Still, its expansion age is very recent, at roughly 3,000

years ago. C4c is a minor haplogroup with a limited geographic distribution that, aside from a sporadic find in Colombia and an ancient sample from Brazil, Loca Do Suin_9100 years old (Ferraz et al. 2023), is confined to NNAm (Supplemental_Fig_S3). Therefore, the unusual results observed for this lineage may result from limited sampling and/or strong drift effects.

Haplogroup D1

For D1, the oldest founding ages (around 31,000 years ago) are found in SSAm and CSAm (Supplementary_Table_S4.5). The earliest expansion ages are also identified in these same regions but at different times (CSAm approximately 23,000 years ago; SSAm approximately 18,000 years ago). Regarding regional D1 founding ages, there is a significant latitudinal correlation from SSAm to Central America (slope -16.65; R^2 0.5431; P-value 0.0370), while a non-significant decreasing trend is observed from Canada to Central America (Supplementary_Table_S4.5). The oldest D1 haplotypes are primarily located in NWSAm and later in SSAm (Supplementary Table S4.4). The shortest genetic distances in D1 are between SSAm-CSAm (0.618), USA-Canada (0.667), and those involving NWSAm with SSAm (0.768) and with NESAm (0.759). A PCoA plot (Fig. 6) visually illustrates these genetic relationships, placing all Mesoamerican and northern South American populations in a central position, distinctly separated from two groups—one consisting of northern North

American populations and the other of southern South American populations, which are very distant from each other.

The D1 phylogeographic data described above closely resemble the patterns observed for haplogroups B2 and C1d.

Haplogroup D4h3a

For D4h3a, the oldest founding age (approximately 30,000 years ago) and expansion age (roughly 22,000 years ago) occurred in CSAm (Supplementary_Table_S4.8). Age comparisons between regions are only significant when the oldest expansion ages of SSAm and CSAm are compared to those of NNAm and NWSAm (Supplementary Table S4.9). When living separately, one can observe that NESAm is isolated; the oldest D4h3a haplotypes are found in NWSAm, Mexico, and consistently in Chile (Supplementary_Table_S4.7). It has been suggested that D4h3a indicates a

Pacific southward expansion (Perego et al. 2009). However, these results do not support that idea. It appears that D4h3a was more common in the past and was most frequent in eastern regions compared to today.

Haplogroup X2

Haplogroup X2a is regarded as a minor Native American haplogroup with a limited geographic range confined to northern North America (Perego et al. 2009). The estimated ages for the origin and spread of haplogroup X2a are approximately 29 years ago and 16 years ago, respectively (Supplementary_Table_S5.1), with the oldest haplotypes found in the X2a1 branch in samples from the North American Great Plains. These ages are notably older in North America than in Canada (Supplementary Table S5.3). However, the discovery of a divergent X2 haplotype in Canada (Perego et al. 2009), which may define a new Native American haplogroup called X2g (Supplemental_Fig_S5), is noteworthy. Although the current distribution of X2a, both ancient and modern, is concentrated in the northeastern part of North America (Perego et al. 2009), with sporadic cases in Mexico (Peñaloza-Espinosa et al. 2007; Bodner et al. 2021), its oldest known sample, the Kennewick Man- approximately 9000 years old and carrying a basal X2a haplotype- was excavated in northwestern Washington state (Rasmussen et al.

2015). This finding raises the possibility that the X2a clade may have originated in the northern Pacific region (Perego et al. 2009).

NRY haplogroups

On a global scale, two major parental lineages with Pan-American distribution have been identified (Q-CTS1780/Z780 and Q-M3), both within haplogroup Q, with the first occurring at lower frequencies everywhere compared to the second. Additionally, two minor lineages belonging to haplogroup C, with a subcontinental range (C-MPB373 and C-P39/Z30536), are considered Native American founder lineages. The southern C-MPB373 has been consistently reported in present-day Colombians and Equatorians (Karafet et al. 1999; Roewer et al. 2013; Mezzavilla et al. 2015; Villaescusa et al. 2021). The northern C-P39/Z30536 has been found in Canada and northern North America, reaching as far as Mexico (Perez-Benedico et al. 2016), showing a northern to southern gradient of decreasing diversity (Dulik et al. 2012). Although phylogenetically Q-CTS1780/Z780 is the oldest clade, the demographic expansion following the LGM mainly occurred among males within the Q-M3 lineage that branched off simultaneously, mainly in Mesoamerica (Q-M925) and South America (Q-M848), forming well-differentiated regional subclades (Bortolini et al. 2003; Battaglia et al. 2013; Roewer et al. 2013; Geppert et al. 2015; The Genographic Consortium et al.

2016; Grugni et al. 2019; Pinotti et al. 2019; Gómez et al. 2021; Villaescusa et

al. 2021; Colombo et al. 2022; Kemp et al. 2007; Paz Sepúlveda et al. 2022).

Some of these minor Q subclades are also noteworthy due to their geographic

distributions and STR diversity. One such lineage is Q-MEH2, identified in

northeast Siberian Koryaks. Its close relationship to the extinct Paleo-Eskimo

Saqqaq (Rasmussen et al. 2010) suggests it could trace a recent migration from

Siberia to America across the Bering Strait (Malyarchuk et al. 2011). However,

Q-MEH2 has recently been found in Nahua populations from southern Mexico,

and based on its STR variability, it appears to have arrived much earlier in

Mexico than in northern America (Gómez et al. 2021). Another notable

example is the Mexican Q-PV3 lineage, initially considered a minor branch of

Q-L54, but its quite divergent haplotypic variability suggests it might represent

another founding Native American lineage (Gómez et al. 2021). This

subcontinental differentiation of male ancestries supports diverse

demographic histories in North and South America (Bortolini et al. 2003).

However, before drawing conclusions, it is necessary to examine what NRY

ancient DNA analyses indicate, especially since post-Columbian admixture

heavily impacted the Native American male genetic pool in both North (Malhi

et al. 2007) and South America (Resque et al. 2016). The picture was similar.

Although Q-CTS1780 might have been more common in the past across both

subcontinents (Posth et al. 2018; Grugni et al. 2019; Ferraz et al. 2023), Q-M3 was also the dominant NRY lineage in all ancient DNA data examined (Bisso-Machado and Fagundes 2021). Nonetheless, the detection of today's extinct lineages in the past (Posth et al. 2018; Colombo et al. 2022) supports the idea that strong founding and drift effects occurred in NRY Native American lineages. These findings do not align well with the hypothesis of a rapid and recent migratory wave from the north to explain the prehistoric colonization of South America.

DISCUSSION

The early arrival of Native uniparental lineages in the Americas, over 30 kya, as proposed here, renders the hypotheses of a prolonged (Hoffecker et al. 1993; Tamm et al. 2007) or brief (Pinotti et al. 2019) standstill in Beringia, a previous expansion of their ancestors into Asia (Wei et al. 2018; Sun et al. 2021; Ning et al. 2020), or the existence of a Pacific Coastal Route around 20 kya (Scheib et al. 2018; Davis and Madsen 2020; Li et al. 2023), unnecessary.

The archaeological evidence for humans in the Siberian Arctic around 30 kya (Pitulko et al. 2004) and possibly since 45 kya (Pitulko et al. 2017) supports the idea of an early entrance into America by its native ancestors. More challenging is gaining acceptance from archaeologists for human occupation in

the American continent before and around the LGM, but it looks pretty likely that this occurred around 16-18 kya at Monte Verde, Chile (Dillehay et al. 2015), at the Santa Elina site in Central Brazil during the LGM, at Chiquihuite Cave in Central Mexico about 30 kya (Ardelean et al. 2020), in the Colorado Plateau more than 36 kya (Rowe et al. 2022), at White Sands, New Mexico, during a stratigraphic record spanning from 24 kya to 17 kya (Holliday et al. 2025), and at Bluefish Caves, Canada, approximately 24 kya (Bourgeon et al. 2017).

Ancient genome studies have characterized the ancestral Native American population as a mixture of northern Eurasians and northern East Asians (Raghavan et al. 2014). The same dual contribution was previously suggested using NRY data (Karafet et al. 1999; Lell et al. 2002; Roewer et al. 2013), and a similar conclusion has been reached here based on mtDNA data. Genomic research indicates that the ancestral Native Americans arrived in the New World as an already admixed group (Moreno-Mayar et al. 2018). However, although coalescence ages do not show significant differences among haplogroups, the NRY phylogeny is more consistent with two successive waves: the first carrying C-MPB373 and Q-Z780 lineages, and the second comprising carriers of C-P39/Z30536 and Q-M3 lineages. Evidence for mtDNA is less conclusive, but the fact that the basal branches connecting haplogroups C and

D to their respective Asian roots are significantly shorter than those of A and B

could support the idea that C and D arrived earlier into America. Another clue

might be found in the Southern Cone of South America, the region farthest

from the continent's entry point. Ancient and modern samples from this area

show that haplogroups C and D were the first and predominant lineages

reaching the region—still the most common today (Lalueza 1997; García-Bour

et al. 2000; Moraga et al. 2000; Demarchi et al. 2001; Nakatsuka et al. 2020;

Postillone et al. 2020). The dominance of haplogroups C and D has also been

observed in Amazonian populations from Brazil (Merriwether and Ferrell

1996), suggesting an early southward migration. Furthermore, mtDNA diversity

for haplogroups C1 and D1 increases from north to south, while North

American populations exhibit higher diversity levels for haplogroups A2 and B2

(Lee and Merriwether 2015). In North America, haplogroups A, B, and X have

expanded across populations in a clinal pattern, whereas there is no evidence

of such spread for haplogroups C and D (Malhi et al. 2001). These patterns are

consistent with two successive migration waves. However, they may also be

explained by genetic drift, as samples from other regions show the absence of

some founder mtDNA haplogroups. For example, the Kuna from Panama lack C

and D (Batista et al. 1995), while the Ayoreo from Bolivia show no A or B

haplogroups (Dornelles et al. 2004). Based on the distribution of two rare

mtDNA haplogroups, D4h3 and X2a, with the first spreading along the Pacific coast and the second entering via the ice-free corridor, two migratory waves from Beringia have been proposed (Perego et al. 2009). Nevertheless, for D4h3, the authors noted that its rarity in North America and higher frequency and variation in South America require further explanation (Perego et al. 2009).

Another hypothesis, based on genome studies, suggests that after arriving in North America, the native ancestral population split into two distinct branches known as South Native Americans (SNA) and North Native Americans (NNA) (Moreno-Mayar et al. 2018), or their approximate equivalents, Ancestry A (ANC-A) and Ancestry B (ANC-B), respectively (Scheib et al. 2018). While the northern ancestry is mostly limited to Canada and eastern North American regions, the southern ancestry is detectable in the oldest sampled genomic sequence, Anzick-1, which is approximately 12700 years old from Montana (Rasmussen et al. 2014), and in the Spirit Cave sample from Nevada, around 10700 years old (Moreno-Mayar et al. 2018), extending southward to Mesoamerica and South America (Moreno-Mayar et al. 2018). There are single-parental markers that share the geographic range of NNA genomic ancestry, such as mtDNA haplogroups X2a (Brown et al. 1998) and C4c (Kashani et al. 2012), as well as NRY lineages C-P39 (Pinotti et al. 2019), Q-Y4276 (Grugni

et al. 2019). The situation becomes much more complex when explaining the transcontinental distribution of the southern component of the Native American genome. The initial dual explanation involving southward Pacific coastal expansion—where successive populations could be seen as mixtures of SNA and NNA components in varying proportions (Scheib et al. 2018)—was proven insufficient (Posth et al. 2018). First, the uniqueness of the northernmost SNA component, represented by the Clovis-associated Anzick-1 sample (SNA1), could not account for the genomic makeup of many South American populations, leading to the need to add a new, dominant non-Clovis SNA2 component, partly represented by Spirit Cave ancestry, to better explain the genomes of many southern populations. Approaching Mesoamerica, the Mexican Mixe population required gene flow from an unsampled population for accurate representation (Moreno-Mayar et al. 2018). Furthermore, several undiscovered genomic ancestries needed to be included to model the structure of various South American regional populations properly. An interesting case is the presence of a divergent ancestry related to Australasian people found in Amazonian tribes such as Surui, Karitiana, and Xavante (Raghavan et al. 2015; Skoglund et al. 2015). This component, called "population Y," was confirmed in ancient Brazilian samples like Sumidouro-5, approximately 10000 years old (Moreno-Mayar et al. 2018). At least one

mtDNA haplotype, likely from haplogroup B—whose southernmost lineage aligns well with the Australasian ancestry—was necessary to accompany this component. However, identifying the ancestral Asian population from which it entered the New World remains challenging. Basic lineages of haplogroup B appeared in eastern Asia around 40,000 years ago, as shown by the Chinese Tianyuan sample (Fu et al. 2013). The Australasian ancestry shared by ancient South American samples and Tianyuan is also notable (Yang et al. 2017), suggesting it could have arrived with populations deeply rooted in eastern Asia with ancient Australasian ties, or with carriers of mtDNA haplogroup B from central Eurasia. The Isthmus-Colombian region also showed genomic complexity, with a Pleistocene ancestry reaching Panamanians and remaining confined to the Isthmus (Capodiferro et al. 2021). Hunter-gatherers from the Colombian Altiplano also carried an unknown basal lineage from the initial South American genomic radiation (Krettek et al. 2025). In northwest South America, a previously unrecognized ancestry was distinctive of northern Ecuador and Colombian populations (Barbieri et al. 2019), while Native Peruvians from the Lake Titicaca area showed a Siberian component also found in ancient North Americans like Anzick-1, Kennewick, and Saqqaq (Lindo et al. 2018). The complex ancestries uncovered in South America have led some scholars to caution against oversimplifying the rapid post-glacial colonization

model, suggesting that a northward expansion from South America to explain the Pacific migration wave would require a more intricate history (Posth et al. 2018). Others propose that the dispersal pattern of SNA south of the continent's ice sheets involved complex admixture events among earlier populations (Moreno-Mayar et al. 2018). The uniparental markers analyzed in this study offer an alternative hypothesis, potentially integrating all genomic data from a different perspective.

If we accept that Asian populations gave rise to Native Americans before 30 kya, then the story of human settlement in America can be viewed differently. It is known that since their arrival, the climate gradually worsened, covering Arctic and Antarctic regions with ice (Clark et al. 2009), clearing the Amazon and Central American rainforests (Häggi et al. 2017), and transforming most of the land into forests and dry steppes (University of Geneva, Switzerland et al. 2001). During this tough period, humans had to survive in small, isolated groups and adapt to different glacial refugia, leaving few archaeological traces. When the climate improved, these groups grew in number and expanded into new, now habitable areas. Unlike the rapid southern wave of colonization, the American continent was not empty of humans. The post-LGM colonization was not a single or multiple waves spreading from northern North America to settle Mesoamerica and then South America. Instead, it involved more or less

simultaneous radiations from various regions, most of which, based on

uniparental markers, are centered in South America. One major expansion

center for mtDNA is the northwest Colombian-Isthmus area, where younger

haplogroup A2 and C1b clines are seen stretching north and south. A similar

distribution of A2 frequencies has also been observed in this region (Keyeux et

al. 2002). Another potential core is in the Andean region, where mtDNA

haplogroup C1c shows its oldest origins and expansion ages, with younger

clines extending toward Canada and down to SSAm. This pattern mirrors that

of haplogroup B, which expanded from Chile and the Andean region into

Mesoamerica. However, it appears that at least part of the B2 cline results

from displacement by other mtDNA lineages in North America, as B was

present early on in northeast Asia as an ancestral B lineage (Fu et al. 2013) and

in Alaska as a basal B2 lineage (Tackney et al. 2015), before nearly disappearing

from both areas. The highest frequencies and genetic diversity of haplogroup

B2 are found in the Andean Altiplano (Gayà-Vidal et al. 2011; Taboada-Echalar

et al. 2013), the Bolivian Piedmont (Corella et al. 2007), and the Argentinian

Gran Chaco (Demarchi et al. 2001). Although the classic explanation for the B2

cline suggests descent from California (Taboada-Echalar et al. 2013), South

American origins have also been proposed, from the Peruvian Andes (Arencibia

et al. 2025) or the Chilean Southern Cone (Bravi et al. 2024). Similarly, the C1d

haplogroup, like B2, shows its most ancient origins and expansion in the southern Cone, spreading northward with younger ages toward Central America, indicating a significant geographic pattern. In North America, some lineages of C1d show an expansion history that ends in Central America, paralleling trends observed in haplogroup A2 sub-lineages. C1a (Tamm et al. 2007), a northeast Asiatic branch of C1, along with minor European sub-clades C1e (Ebenesersdóttir et al. 2011) and C1f (Der Sarkissian et al. 2014), suggest a later reverse migration from America (Tamm et al. 2007). Using the evolutionary rate proposed here, the founding age of C1a in Asia is about 26.02 kya (95% CI: 18.58-33.49 kya), signaling a return to Asia during worsening weather conditions. Its spread across Asia around 13.9 kya (95% CI: 6.46-21.37 kya) points to a Mesolithic migration, similar to the pattern seen in the northeast European C1f clade (Der Sarkissian et al. 2014). This early American retro-migration might also be supported by the phylogeny and phylogeography of the Y-chromosome haplogroups C-L1373 (Pinotti et al. 2019; Colombo et al. 2022) and Q-L54 (Grugni et al. 2019; Pinotti et al. 2019).

Another mtDNA haplogroup indicating the southern Cone region as a key center of expansion is D1, which shows a significant negative correlation from this area to Central America. Its mean coalescent ages are younger as you move upward, although there is a longitudinal bias leaning toward central and

eastern South American regions. D1 is most common in the eastern parts of the Andean region, with ancient regional clades in Central Argentina (García et al. 2012) and the southern Cone (Bodner et al. 2012). Similarly, for A2 and C1d, a southward expansion from Canada to Central America is observed for D1. These expansion patterns might explain the B2 retraction seen in North America. The idea that mtDNA haplogroup D4h3a supports a Pacific coastal route for peopling of the Americas (Perego et al. 2009), later backed by genomic studies (Moreno-Mayar et al. 2018), is not supported by the evidence here. The earliest ages for the founding and expansion of D4h3a are found in SSAm and CSAm, and are significantly older than those in NWSAm and NNAm (Supplementary_Table_S4.9_ S4.9). Additionally, although D4h3a is absent today, it has been found in ancient samples from Alaska, Shuká-Káa, 10.3 kya (Lindo et al. 2017), from British Columbia, Canada, 939, 6.5 kya (Lindo et al. 2017), and Montana, Anzick 1, 12.6 kya (Rasmussen et al. 2014). It was also detected in ancient remains from the Brazilian Amazon, Lagoa Santa, 10 kya (Moreno-Mayar et al. 2018). This pattern closely resembles that of B2. Therefore, rather than modeling a southward Pacific Coastal expansion, both markers seem to be remnants of a broader past occupation that was displaced by more recent human migrations.

A specific area in the Amazon Basin (CSAm) deserves special mention. It participated in all the mtDNA radiation centers discussed earlier and is known as a major reservoir of deep mtDNA lineages (De Souza et al. 2025; Di Corcia et al. 2017). It stands out for the ancient presence of very deep-rooted Y-chromosome lineages such as C-L1373* (Colombo et al. 2022) and for the detection of lineages that were previously thought to be unique to the northern subcontinent, like Q-Y4303* (Grugni et al. 2019). This genetic diversity is also supported by whole-genome studies on ancient specimens (Moreno-Mayar et al. 2018) and current populations (Ribeiro-Dos-Santos et al. 2020; Capodiferro et al. 2023). This genetic landscape aligns with the region's linguistic richness. Notably, out of 71 identified linguistic isolates across the continent, over 45% are in Brazil, compared to 15% in North and Mesoamerica combined (Blench and Cb). Future archaeological findings will likely uncover human settlements much older than currently recognized. Furthermore, the differences in genetic and linguistic diversity between the northern (NNA) and southern (SNA) American branches make the idea of simultaneous colonization of both subcontinents from a northward expansion unlikely.

MATERIAL AND METHODS:

Material

For the mtDNA phylogenetic and phylogeographic analyses, I searched for

partial and complete mitogenomes in the NCBI GenBank,

www.ncbi.nlm.nih.gov/genbank/ ; Mitomap, www.mitomap.org/MITOMAP ;

EMPOP, https://empop.online ; Ian Logan 2023,

www.ianlogan.co.uk/sequences_by_group/haplogroup_select.htm ; Genome

Warehouse (GWH) public repository, https://bigd.big.ac.cn/gwh ; and AmtDB,

www.http://amtdb.org . Additionally, for phylogeographic analyses only, I also

searched YFull and MTree. The NRY phylogenetic and phylogeographic

analyses were based on the ISOGG database, http://www.isogg.org/tree/ ;

YFull YTree v13.03.00; YTree; and data compiled by other authors (Wei et al.

2018; Grugni et al. 2019; Pinotti et al. 2019; Gómez et al. 2021; Colombo et al.

2022; Paz Sepúlveda et al. 2022). For brevity, the Y-chromosome lineages in

the text are referred to by the letter of their basic haplogroup and their

terminal mutation.

Phylogeny

Maximum parsimony intraspecific phylogenetic trees (Bandelt et al. 1999)

were constructed manually, and haplogroup nomenclature followed the

PhyloTree database Building 17, http://www.phylotree.org. Haplogroup

classification was confirmed using HaploGrep 3 3.2.1, https://haplogrep.i-med.ac.at (Weissensteiner et al. 2016). Newly identified subgroups not classified in PhyloTree were provisionally named by the first letter and digit of their primary haplogroup, followed by an asterisk and consecutive numbers. Both labels are shown on the respective tree when a more formal nomenclature is available in YFullMTree.

Evolutionary rates

It is well known that the mtDNA evolutionary rate experiences a time-dependent effect, slowing down as we go back in time (Henn et al. 2008). The main reason for this effect has been linked to differences in population size, with the smallest populations during Paleolithic times and exponential growth in recent times (Cabrera 2021a). To account for this anomaly, I used an estimated Paleolithic mitogenome substitution rate of 1.5×10^{-8} (95% CI: $0.42 - 9.07 \times 10^{-8}$) per site per year (assuming a mtDNA genome length of 16,500 base pairs), meaning one substitution occurs roughly every 4,040 years (Cabrera 2024), to estimate haplogroup coalescent ages for the early prehistoric settlement of the Americas. Using a similar approach, a proposed mean evolutionary rate of 0.47×10^{-9} (95% CI: $0.21 - 0.95 \times 10^{-9}$) per site per year was used to calculate coalescent ages of the Y-chromosome (Cabrera 2024).

Ages

I have used three approaches to estimate haplogroup coalescent ages: 1)

applying statistics rho (Forster et al. 1996) and Sigma (Saillard et al. 2000) on

individuals within haplogroups, along with the evolutionary rates mentioned

above; 2) utilizing clades within main haplogroups to calculate a pooled mean,

then applying normal averaging and standard error statistics for mutation rates

across haplogroups; 3) since the sample sizes of these clades vary significantly,

I calculated weighted averages and weighted standard errors to apply with the

same normal statistics; 4) a common discussion in genetics is that haplotypes

at low frequency might become much more common within a population,

although this is generally an exception; typically, rare haplotypes—those at the

tails of a distribution—are likely to go extinct due to genetic drift, while

haplotypes near the mean tend to increase in frequency. Given that the Native

American founding haplogroups have experienced genetic drift over time, I

used the most divergent lineages within them as proxies for their past means

to estimate coalescence ages (Cabrera 2022; Cabrera 2021a).

Phylogeography

For phylogeographic purposes, the American continent has been divided into

seven geographic regions: 1) North Northern America (NNAm), including

Alaska and Canada; 2) North America, covering the entire United States (Nam);

3) Mesoamerica, comprising Mexico, Belize, Guatemala, El Salvador, Honduras,

and Nicaragua (Mam); 4) Northeastern South America or Isthmus-Colombian

area, including Costa Rica, Panama, and Colombia; 5) Northwestern South

America (NWSAm), also known as the Andean region, including Ecuador, Peru,

and Bolivia; 6) Central South America (CSAm), including Brazil, Paraguay, and

Uruguay; 7) Southern South America (SSAm), including Argentina and Brazil.

Additionally, within these regions, distinctions are sometimes made between

inland and coastal Pacific and Atlantic areas, as well as between Highland

Andes and Amazonian zones. To support the phylogenetic and

phylogeographic analyses, I have implemented some straightforward

algorithms described below.

Genetic distances

In addition to the molecular distance based on the number of mutations

between two homologous DNA sequences (FST), I have implemented a

phylogeographic distance (DXY) based on genetic affinity (IXY) between regions

X and Y. This distance is measured by the frequency with which they form

sister branches within a haplogroup tree (nXY) relative to how often each

region participates as a sister branch within the measured haplogroup (nX and

nY).

$$I_{xy} = \frac{2n_{xy}}{nx + ny}$$

Then D_{XY} is calculated as $1 - I_{XY}$.

Additionally, when searching for sister branches between X and Y regions within a haplogroup, I also observed cases where one sequence from one of the involved regions served as a direct outgroup to the pair, indicating it as ancestral relative to the other region, which was considered derived. The significance of this relationship was tested using observed-expected chi-square statistics.

Additionally, for phylogeographic purposes, I have examined each major Native American founder haplogroup to identify the regions where their oldest sub-haplogroups are found and the areas from which they initially expanded.

Finally, to identify the Asian centers where the ancestors of the major American founder clades may have originated, I searched for the most ancient Asian sequences (less mutated) that are from the Asian-American shared phylogenetic nodes and are found in these regions.

Other statistics

Analysis of molecular variance (AMOVA), correlation/linear regression, and graphical Principal Coordinates Analysis (PCoA) were performed using the

GenAlEx 6.5 software (Peakall and Smouse 2012). Descriptive normal statistics, 95% confidence intervals for means, and two-tailed t-tests were conducted with GraphPad https://www.graphpad.com/quickcalcs/. Weighted means and standard errors were calculated using the Weighted calculator (https://calculatodo.com).

False positive correction

Since I performed multiple statistical tests for each founder haplogroup, I applied a Bonferroni p-value adjustment. However, because the Bonferroni correction is very conservative, I also used the Benjamin and Hochberg 1995 (Benjamini and Hochberg 1995) false discovery rate (FDR) method as a less restrictive alternative.

DATA ACCESS

All the sequences and haplotypes used in this study are available in the public domain. Databases and references are detailed in the text.

COMPETING INTEREST STATEMENT

The author declares no conflicts of interest regarding this work.

REFERENCES

Adamov D, Guryanov V, Karzhavin S, Tagankin V, Urasin V. 2015. Defining a New Rate Constant for Y-Chromosome SNPs based on Full Sequencing Data.

Ardelean CF, Becerra-Valdivia L, Pedersen MW, Schwenninger J-L, Oviatt CG, Macías-Quintero JI, Arroyo-Cabrales J, Sikora M, Ocampo-Díaz YZE, Rubio-Cisneros II, et al. 2020. Evidence of human occupation in Mexico around the Last Glacial Maximum. *Nature* **584**: 87–92.

Arencibia V, Muñoz M, Crespo CM, Russo MG, Vera P, Lia VV, García Guraieb S, Goñi RA, Avena S, Puebla A, et al. 2025. Novel B2 mitogenomes from Continental southern Patagonia's Late Holocene: New insights into the peopling of the Southern Cone. *American Journal of Biological Anthropology* **186**: e24822.

Avila E, Graebin P, Chemale G, Freitas J, Kahmann A, Alho CS. 2019. Full mtDNA genome sequencing of Brazilian admixed populations: A forensic-focused evaluation of an MPS application as an alternative to Sanger sequencing methods. *Forensic Science International: Genetics* **42**: 154–164.

Bandelt HJ, Forster P, Rohl A. 1999. Median-joining networks for inferring intraspecific phylogenies. *Molecular Biology and Evolution* **16**: 37–48.

Barbieri C, Barquera R, Arias L, Sandoval JR, Acosta O, Zurita C, Aguilar-Campos A, Tito-Álvarez AM, Serrano-Osuna R, Gray RD, et al. 2019. The Current

Genomic Landscape of Western South America: Andes, Amazonia, and Pacific Coast. *Molecular Biology and Evolution* **36**: 2698–2713.

Batista O, Kolman CJ, Bermingham E. 1995. Mitochondrial DNA diversity in the Kuna Amerinds of Panama. *Human Molecular Genetics* **4**: 921–929.

Battaglia V, Grugni V, Perego UA, Angerhofer N, Gomez-Palmieri JE, Woodward SR, Achilli A, Myres N, Torroni A, Semino O. 2013. The First Peopling of South America: New Evidence from Y-Chromosome Haplogroup Q, ed. D. O'Rourke. *PLoS ONE* **8**: e71390.

Becerra-Valdivia L, Higham T. 2020. The timing and effect of the earliest human arrivals in North America. *Nature* **584**: 93–97.

Benjamini Y, Hochberg Y. 1995. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. *Journal of the Royal Statistical Society Series B: Statistical Methodology* **57**: 289–300.

Bisso-Machado R, Bortolini MC, Salzano FM. 2012. Uniparental genetic markers in South Amerindians. *Genet Mol Biol* **35**: 365–387.

Bisso-Machado R, Fagundes NJR. 2021. Uniparental genetic markers in Native Americans: A summary of all available data from ancient and contemporary populations. *American J Phys Anthropol* **176**: 445–458.

Blench R, CB C. [DRAFT CIRCULATED FOR COMMENT].

Bodner M, Perego UA, Gomez JE, Cerda-Flores RM, Rambaldi Migliore N, Woodward SR, Parson W, Achilli A. 2021. The Mitochondrial DNA Landscape of Modern Mexico. *Genes* **12**: 1453.

Bodner M, Perego UA, Huber G, Fendt L, Röck AW, Zimmermann B, Olivieri A, Gómez-Carballa A, Lancioni H, Angerhofer N, et al. 2012. Rapid coastal spread of First Americans: Novel insights from South America's Southern Cone mitochondrial genomes. *Genome Res* **22**: 811–820.

Bonatto SL, Salzano FM. 1997. Diversity and Age of the Four Major mtDNA Haplogroups, and Their Implications for the Peopling of the New World. *The American Journal of Human Genetics* **61**: 1413–1423.

Bortolini M-C, Salzano FM, Thomas MG, Stuart S, Nasanen SPK, Bau CHD, Hutz MH, Layrisse Z, Petzl-Erler ML, Tsuneto LT, et al. 2003. Y-Chromosome Evidence for Differing Ancient Demographic Histories in the Americas. *The American Journal of Human Genetics* **73**: 524–539.

Bourgeon L, Burke A, Higham T. 2017. Earliest Human Presence in North America Dated to the Last Glacial Maximum: New Radiocarbon Dates from Bluefish Caves, Canada ed. J.P. Hart. *PLoS ONE* **12**: e0169486.

Brandini S, Bergamaschi P, Cerna MF, Gandini F, Bastaroli F, Bertolini E, Cereda C, Ferretti L, Gómez-Carballa A, Battaglia V, et al. 2018. The Paleo-Indian Entry into South America According to Mitogenomes. *Molecular Biology and Evolution* **35**: 299–311.

Bravi CM, Motti JMB, García A. 2024. Letter to the editor: A Southern Cone origin rather than Peruvian affinities for ancient Patagonian B2 mitogenomes. *American Journal of Biological Anthropology* **184**: e24934.

Brown MD, Hosseini SH, Torroni A, Bandelt H-J, Allen JC, Schurr TG, Scozzari R, Cruciani F, Wallace DC. 1998. mtDNA Haplogroup X: An Ancient Link between Europe/Western Asia and North America? *The American Journal of Human Genetics* **63**: 1852–1861.

Cabrera VM. 2020. Counterbalancing the time-dependent effect on the human mitochondrial DNA molecular clock. *BMC Evol Biol* **20**: 78.

Cabrera VM. 2024. Early male and female footprints of modern humans across Eurasia and Australasia. *Medical Research Archives* **12**.

Cabrera VM. 2021a. Human molecular evolutionary rate, time dependency, and transient polymorphism effects viewed through ancient and modern mitochondrial DNA genomes. *Sci Rep* **11**: 5036.

Cabrera VM. 2021b. Revisiting the Hominin Phylogeny: An Alternative Introgression Scenario to Reconcile the Uniparental and Autosomal DNA Topologies. *Jpeb* **9**.

Capodiferro MR, Aram B, Raveane A, Rambaldi Migliore N, Colombo G, Ongaro L, Rivera J, Mendizábal T, Hernández-Mora I, Tribaldos M, et al. 2021. Archaeogenomic distinctiveness of the Isthmo-Colombian area. *Cell* **184**: 1706-1723.e24.

Capodiferro MR, Chero Osorio AM, Rambaldi Migliore N, Tineo Tineo DH, Raveane A, Xavier C, Bodner M, Simão F, Ongaro L, Montinaro F, et al. 2023. The multifaceted genomic history of Ashaninka from Amazonian Peru. *Current Biology* **33**: 1573-1581.e5.

Chatters JC, Kennett DJ, Asmerom Y, Kemp BM, Polyak V, Blank AN, Beddows PA, Reinhardt E, Arroyo-Cabrales J, Bolnick DA, et al. 2014. Late Pleistocene Human Skeleton and mtDNA Link Paleoamericans and Modern Native Americans. *Science* **344**: 750–754.

Clark PU, Dyke AS, Shakun JD, Carlson AE, Clark J, Wohlfarth B, Mitrovica JX, Hostetler SW, McCabe AM. 2009. The Last Glacial Maximum. *Science* **325**: 710–714.

Colombo G, Traverso L, Mazzocchi L, Grugni V, Rambaldi Migliore N, Capodiferro MR, Lombardo G, Flores R, Karmin M, Rootsi S, et al. 2022. Overview of the Americas' First Peopling from a Patrilineal Perspective: New Evidence from the Southern Continent. *Genes* **13**: 220.

Corella A, Bert F, Pérez-Pérez A, Gené M, Turbón D. 2007. Mitochondrial DNA diversity of the Amerindian populations living in the Andean Piedmont of Bolivia: Chimane, Moseten, Aymara, and Quechua. *Annals of Human Biology* **34**: 34–55.

Davis LG, Madsen DB. 2020. The coastal migration theory: Formulation and testable hypotheses. *Quaternary Science Reviews* **249**: 106605.

De Souza FG, Matos GB, Sena Santos C, Souza TP, Gobbo AR, Da Costa PF, Salgado CG, Lopes Eufraseo G, Vitor De Souza Fernandes A, Santos-Lobato BL, et al. 2025. Mitochondrial ancestry from complete mitogenomes highlights a lack of characterization of indigenous haplogroups in the Brazilian Amazon population. *Commun Biol* **8**: 835.

Demarchi DA, Panzetta-Dutari GM, Motran CC, De Los Angeles López De Basualdo M, Marcellino AJ. 2001. Mitochondrial DNA haplogroups in Amerindian populations from the Gran Chaco. *American J Phys Anthropol* **115**: 199–203.

Der Sarkissian C, Brotherton P, Balanovsky O, Templeton JEL, Llamas B, Soubrier J, Moiseyev V, Khartanovich V, Cooper A, Haak W, et al. 2014. Mitochondrial Genome Sequencing in Mesolithic North East Europe Unearths a New Sub-Clade within the Broadly Distributed Human Haplogroup C1, ed. A. Achilli. *PLoS ONE* **9**: e87612.

Derenko M, Malyarchuk B, Grzybowski T, Denisova G, Rogalla U, Perkova M, Dambueva I, Zakharov I. 2010. Origin and Post-Glacial Dispersal of Mitochondrial DNA Haplogroups C and D in Northern Asia, ed. M.T.P. Gilbert. *PLoS ONE* **5**: e15214.

Di Corcia T, Sanchez Mellado C, Davila Francia TJ, Ferri G, Sarno S, Luiselli D, Rickards O. 2017. East of the Andes: The genetic profile of the Peruvian Amazon populations. *American J Phys Anthropol* **163**: 328–338.

Dillehay TD, Ocampo C, Saavedra J, Sawakuchi AO, Vega RM, Pino M, Collins MB, Scott Cummings L, Arregui I, Villagran XS, et al. 2015. New Archaeological Evidence for an Early Human Presence at Monte Verde, Chile, ed. J.P. Hart. *PLoS ONE* **10**: e0141923.

Dornelles CL, Battilana J, Fagundes NJR, Freitas LB, Bonatto SL, Salzano FM. 2004. Mitochondrial DNA and *Alu* insertions in a genetically peculiar

population: The Ayoreo Indians of Bolivia and Paraguay. *American J Hum Biol* **16**: 479–488.

Dulik MC, Zhadanov SI, Osipova LP, Askapuli A, Gau L, Gokcumen O, Rubinstein S, Schurr TG. 2012. Mitochondrial DNA and Y Chromosome Variation Provide Evidence for a Recent Common Ancestry between Native Americans and Indigenous Altaians. *The American Journal of Human Genetics* **90**: 229–246.

Ebenesersdóttir SS, Sigurðsson Á, Sánchez-Quinto F, Lalueza-Fox C, Stefánsson K, Helgason A. 2011. A new subclade of mtDNA haplogroup C1 found in Icelanders: Evidence of pre-Columbian contact? *American J Phys Anthropol* **144**: 92–99.

Faught MK. 2017. Where was the PaleoAmerind standstill? *Quaternary International* **444**: 10–18.

Ferraz T, Suarez Villagran X, Nägele K, Radzevičiūtė R, Barbosa Lemes R, Salazar-García DC, Wesolowski V, Lopes Alves M, Bastos M, Rapp Py-Daniel A, et al. 2023. Genomic history of coastal societies from eastern South America. *Nat Ecol Evol* **7**: 1315–1330.

Fiedel SJ. 2000. The Peopling of the New World: Present Evidence, New Theories, and Future Directions. *Journal of Archaeological Research* **8**: 39–103.

Flegontov P, Altınışık NE, Changmai P, Rohland N, Mallick S, Adamski N, Bolnick DA, Broomandkhoshbacht N, Candilio F, Culleton BJ, et al. 2019. Palaeo-Eskimo genetic ancestry and the peopling of Chukotka and North America. *Nature* **570**: 236–240.

Forster P, Harding R, Torroni A, Bandelt H-J. 1996. Origin and Evolution of Native American mtDNA Variation: A Reappraisal. *Am J Hum Genet.*

Francalacci P, Morelli L, Angius A, Berutti R, Reinier F, Atzeni R, Pilu R, Busonero F, Maschio A, Zara I, et al. 2013. Low-Pass DNA Sequencing of 1200 Sardinians Reconstructs European Y-Chromosome Phylogeny. *Science* **341**: 565–569.

Fregel R, Cabrera V, Larruga JM, Abu-Amero KK, González AM. 2015. Carriers of Mitochondrial DNA Macrohaplogroup N Lineages Reached Australia around 50,000 Years Ago following a Northern Asian Route, ed. F. Calafell. *PLoS ONE* **10**: e0129839.

Fu Q, Meyer M, Gao X, Stenzel U, Burbano HA, Kelso J, Pääbo S. 2013. DNA analysis of an early modern human from Tianyuan Cave, China. *Proc Natl Acad Sci USA* **110**: 2223–2227.

García A, Pauro M, Nores R, Bravi CM, Demarchi DA. 2012. Phylogeography of mitochondrial haplogroup D1: An early spread of subhaplogroup D1j from Central Argentina. *American J Phys Anthropol* **149**: 583–590.

García-Bour J, Pérez-Pérez A, Prats E, Turbón D. 2000. MOLECULAR APPROACH TO THE PEOPLING OF THE AMERICAS BY SEQUENCING mtDNA FROM EXTINCT FUEGUIANS AND PATAGONS. *Chungará (Arica)* **32**.

Gayà-Vidal M, Moral P, Saenz-Ruales N, Gerbault P, Tonasso L, Villena M, Vasquez R, Bravi CM, Dugoujon J. 2011. mtDNA and Y-chromosome diversity in Aymaras and Quechuas from Bolivia: Different stories and special genetic traits of the Andean Altiplano populations. *American J Phys Anthropol* **145**: 215–230.

Geppert M, Ayub Q, Xue Y, Santos S, Ribeiro-dos-Santos Â, Baeta M, Núñez C, Martínez-Jarreta B, Tyler-Smith C, Roewer L. 2015. Identification of new SNPs in native South American populations by resequencing the Y chromosome. *Forensic Science International: Genetics* **15**: 111–114.

Gilbert MTP, Jenkins DL, Götherstrom A, Naveran N, Sanchez JJ, Hofreiter M, Thomsen PF, Binladen J, Higham TFG, Yohe RM, et al. 2008. DNA from Pre-Clovis Human Coprolites in Oregon, North America. *Science* **320**: 786–789.

Gómez R, Vilar MG, Meraz-Ríos MA, Véliz D, Zúñiga G, Hernández-Tobías EA, Figueroa-Corona MDP, Owings AC, Gaieski JB, Schurr TG. 2021. Y chromosome

diversity in Aztlan descendants and its implications for the history of Central Mexico. *iScience* **24**: 102487.

Grugni V, Raveane A, Ongaro L, Battaglia V, Trombetta B, Colombo G, Capodiferro MR, Olivieri A, Achilli A, Perego UA, et al. 2019. Analysis of the human Y-chromosome haplogroup Q characterizes ancient population movements in Eurasia and the Americas. *BMC Biol* **17**: 3.

Gusareva ES, Ghosh AG, Kharkov VN, Khor S-S, Zarubin A, Moshkov N, Kalsi N, Ratan A, Heinle CE, Cooke N, et al. 2025. From North Asia to South America: Tracing the longest human migration through genomic sequencing. *Science* **388**: eadk5081.

Häggi C, Chiessi CM, Merkel U, Mulitza S, Prange M, Schulz M, Schefuß E. 2017. Response of the Amazon rainforest to late Pleistocene climate variability. *Earth and Planetary Science Letters* **479**: 50–59.

Hallast P, Batini C, Zadik D, Maisano Delser P, Wetton JH, Arroyo-Pardo E, Cavalleri GL, De Knijff P, Destro Bisol G, Dupuy BM, et al. 2015. The Y-Chromosome Tree Bursts into Leaf: 13,000 High-Confidence SNPs Covering the Majority of Known Clades. *Molecular Biology and Evolution* **32**: 661–673.

Henn BM, Gignoux CR, Feldman MW, Mountain JL. 2008. Characterizing the Time Dependency of Human Mitochondrial DNA Mutation Rate Estimates. *Molecular Biology and Evolution* **26**: 217–230.

Hoffecker JF, Powers WR, Goebel T. 1993. The Colonization of Beringia and the Peopling of the New World. *Science* **259**: 46–53.

Holliday VT, Windingstad JD, Bright J, Phillips BG, Butler JB, Breslawski R, Bowman JE. 2025. Paleolake geochronology supports a Last Glacial Maximum (LGM) age for human tracks at White Sands, New Mexico. *Science Advances*.

Hubert N, Hanner R. 2015. DNA Barcoding, species delineation and taxonomy: a historical perspective. *DNA Barcodes* **3**. http://access.portico.org/stable?au=phx3c2h64c3 (Accessed July 29, 2025).

Järve M, Saag L, Scheib CL, Pathak AK, Montinaro F, Pagani L, Flores R, Guellil M, Saag L, Tambets K, et al. 2019. Shifts in the Genetic Landscape of the Western Eurasian Steppe Associated with the Beginning and End of the Scythian Dominance. *Current Biology* **29**: 2430-2441.e10.

Karafet TM, Zegura SL, Posukh O, Osipova L, Bergen A, Long J, Goldman D, Klitz W, Harihara S, De Knijff P, et al. 1999. Ancestral Asian Source(s) of New World Y-Chromosome Founder Haplotypes. *The American Journal of Human Genetics* **64**: 817–831.

Karmin M, Saag L, Vicente M, Sayres MAW, Järve M, Talas UG, Rootsi S, Ilumäe A-M, Mägi R, Mitt M, et al. 2015. A recent bottleneck of Y chromosome diversity coincides with a global cultural change. *Genome Res* **25**: 459–466.

Kashani BH, Perego UA, Olivieri A, Angerhofer N, Gandini F, Carossa V, Lancioni H, Semino O, Woodward SR, Achilli A, et al. 2012. Mitochondrial haplogroup C4c: A rare lineage entering America through the ice-free corridor? *American J Phys Anthropol* **147**: 35–39.

Kemp BM, Malhi RS, McDonough J, Bolnick DA, Eshleman JA, Rickards O, Martinez-Labarga C, Johnson JR, Lorenz JG, Dixon EJ, et al. 2007. Genetic analysis of early Holocene skeletal remains from Alaska and its implications for the settlement of the Americas. *American J Phys Anthropol* **132**: 605–621.

Keyeux G, Rodas C, Gelvez N, Carter D. 2002. Possible Migration Routes into South America Deduced from Mitochondrial DNA Studies in Colombian Amerindian Populations. *Human Biology* **74**: 211–233.

Krettek K-L, Casas-Vargas A, Mas-Sandoval A, Alvis LA, Reiter E, Madero JM, Urban M, Vargas JCN, Usaquén W, Cuenca JVR, et al. 2025. A 6000-year-long genomic transect from the Bogotá Altiplano reveals multiple genetic shifts in the demographic history of Colombia. *Science Advances*.

Kujanová M, Pereira L, Fernandes V, Pereira JB, Černý V. 2009. Near Eastern Neolithic genetic input in a small oasis of the Egyptian Western Desert. *American J Phys Anthropol* **140**: 336–346.

Lalueza C. 1997. Lack of founding Amerindian mitochondrial DNA lineages in extinct aborigines from Tierra del Fuego-Patagonia. *Human Molecular Genetics* **6**: 41–46.

Lee, Merriwether. 2015. Identification of Whole Mitochondrial Genomes from Venezuela and Implications on Regional Phylogenies in South America. *Human Biology* **87**: 29.

Lell JT, Sukernik RI, Starikovskaya YB, Su B, Jin L, Schurr TG, Underhill PA, Wallace DC. 2002. The Dual Origin and Siberian Affinities of Native American Y Chromosomes. *The American Journal of Human Genetics* **70**: 192–206.

Li Y-C, Gao Z-L, Liu K-J, Tian J-Y, Yang B-Y, Rahman ZU, Yang L-Q, Zhang S-H, Li C-T, Achilli A, et al. 2023. Mitogenome evidence shows two radiation events and dispersals of matrilineal ancestry from northern coastal China to the Americas and Japan. *Cell Reports* **42**: 112413.

Lindo J, Achilli A, Perego UA, Archer D, Valdiosera C, Petzelt B, Mitchell J, Worl R, Dixon EJ, Fifield TE, et al. 2017. Ancient individuals from the North American

Northwest Coast reveal 10,000 years of regional genetic continuity. *Proc Natl Acad Sci USA* **114**: 4093–4098.

Lindo J, Haas R, Hofman C, Apata M, Moraga M, Verdugo RA, Watson JT, Llave CV, Witonsky D, Beall C, et al. 2018. The genetic prehistory of the Andean highlands 7000 years BP through European contact. *SCIENCE ADVANCES*.

Llamas B, Fehren-Schmitz L, Valverde G, Soubrier J, Mallick S, Rohland N, Nordenfelt S, Valdiosera C, Richards SM, Rohrlach A, et al. 2016. Ancient mitochondrial DNA provides a high-resolution time scale of the peopling of the Americas. *Sci Adv* **2**: e1501385.

Malhi RS, Kemp BM, Eshleman JA, Cybulski J, Smith DG, Cousins S, Harry H. 2007. Mitochondrial haplogroup M was discovered in prehistoric North Americans. *Journal of Archaeological Science* **34**: 642–648.

Malhi RS, Schultz BA, Smith DG. 2001. Distribution of Mitochondrial DNA Lineages among Native American Tribes of Northeastern North America. *Human Biology* **73**: 17–55.

Malyarchuk B, Derenko M, Denisova G, Maksimov A, Wozniak M, Grzybowski T, Dambueva I, Zakharov I. 2011. Ancient links between Siberians and Native Americans revealed by subtyping the Y chromosome haplogroup Q1a. *J Hum Genet* **56**: 583–588.

Mao X, Zhang H, Qiao S, Liu Y, Chang F, Xie P, Zhang M, Wang T, Li M, Cao P, et al. 2021. The deep population history of northern East Asia from the Late Pleistocene to the Holocene. *Cell* **184**: 3256-3266.e13.

Marrero P, Abu-Amero KK, Larruga JM, Cabrera VM. 2016. Carriers of human mitochondrial DNA macrohaplogroup M colonized India from southeastern Asia. *BMC Evol Biol* **16**: 246.

Merriwether DA, Ferrell RE. 1996. The Four Founding Lineage Hypothesis for the New World: A Critical Reevaluation. *Molecular Phylogenetics and Evolution* **5**: 241–246.

Mezzavilla M, Geppert M, Tyler-Smith C, Roewer L, Xue Y. 2015. Insights into the origin of rare haplogroup C3* Y chromosomes in South America from high-density autosomal SNP genotyping. *Forensic Science International: Genetics* **15**: 115–120.

Moraga ML, Rocco P, Miquel JF, Nervi F, Llop E, Chakraborty R, Rothhammer F, Carvallo P. 2000. Mitochondrial DNA polymorphisms in Chilean aboriginal populations: Implications for the peopling of the southern cone of the continent. *Am J Phys Anthropol* **113**: 19–29.

Moreno-Mayar JV, Vinner L, De Barros Damgaard P, De La Fuente C, Chan J, Spence JP, Allentoft ME, Vimala T, Racimo F, Pinotti T, et al. 2018. Early human dispersals within the Americas. *Science* **362**: eaav2621.

Nakatsuka N, Luisi P, Motti JMB, Salemme M, Santiago F, D'Angelo Del Campo MD, Vecchi RJ, Espinosa-Parrilla Y, Prieto A, Adamski N, et al. 2020. Ancient genomes in South Patagonia reveal population movements associated with technological shifts and geography. *Nat Commun* **11**: 3868.

Ning C, Fernandes D, Changmai P, Flegontova O, Yüncü E, Maier R, Altınışık NE, Kassian AS, Krause J, Lalueza-Fox C, et al. 2020. The genomic formation of First American ancestors in East and Northeast Asia. http://biorxiv.org/lookup/doi/10.1101/2020.10.12.336628 (Accessed July 29, 2025).

Ovchinnikov IV, Malek MJ, Drees K, Kholina OI. 2014. Mitochondrial DNA variation in Tajiks living in Tajikistan. *Legal Medicine* **16**: 390–395.

Palencia-Madrid L, De Pancorbo MM. 2015. Next-generation sequencing as a suitable analysis for the authentication of degraded and ancient DNA. *Forensic Science International: Genetics Supplement Series* **5**: e338–e340.

Paz Sepúlveda PB, Mayordomo AC, Sala C, Sosa EJ, Zaiat JJ, Cuello M, Schwab M, Rodríguez Golpe D, Aquilano E, Santos MR, et al. 2022. Human Y

chromosome sequences from Q Haplogroup reveal a South American settlement pre-18,000 years ago and a profound genomic impact during the Younger Dryas ed. F. Calafell. *PLoS ONE* **17**: e0271971.

Peakall R, Smouse PE. 2012. GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research—an update. *Bioinformatics* **28**: 2537–2539.

Peñaloza-Espinosa RI, Arenas-Aranda D, Cerda-Flores RM, Buentello-Malo L, González-Valencia G, Torres J, Álvarez B, Mendoza I, Flores M, Sandoval L, et al. 2007. Characterization of mtDNA Haplogroups in 14 Mexican Indigenous Populations. *Hub* **79**: 313–320.

Peng M-S, Xu W, Song J-J, Chen X, Sulaiman X, Cai L, Liu H-Q, Wu S-F, Gao Y, Abdulloevich NT, et al. 2018. Mitochondrial genomes uncover the maternal history of the Pamir populations. *Eur J Hum Genet* **26**: 124–136.

Perego UA, Achilli A, Angerhofer N, Accetturo M, Pala M, Olivieri A, Kashani BH, Ritchie KH, Scozzari R, Kong Q-P, et al. 2009. Distinctive Paleo-Indian Migration Routes from Beringia Marked by Two Rare mtDNA Haplogroups. *Current Biology* **19**: 1–8.

Perez-Benedico D, La Salvia J, Zeng Z, Herrera GA, Garcia-Bertrand R, Herrera RJ. 2016. Mayans: a Y chromosome perspective. *Eur J Hum Genet* **24**: 1352–1358.

Petr M, Hajdinjak M, Fu Q, Essel E, Rougier H, Crevecoeur I, Semal P, Golovanova LV, Doronichev VB, Lalueza-Fox C, et al. 2020. The evolutionary history of Neanderthal and Denisovan Y chromosomes. *Science* **369**: 1653–1656.

Pinotti T, Bergström A, Geppert M, Bawn M, Ohasi D, Shi W, Lacerda DR, Solli A, Norstedt J, Reed K, et al. 2019. Y Chromosome Sequences Reveal a Short Beringian Standstill, Rapid Expansion, and early Population structure of Native American Founders. *Current Biology* **29**: 149-157.e3.

Pitulko V, Pavlova E, Nikolskiy P. 2017. Revising the archaeological record of the Upper Pleistocene Arctic Siberia: Human dispersal and adaptations in MIS 3 and 2. *Quaternary Science Reviews* **165**: 127–148.

Pitulko VV, Nikolsky PA, Girya EYu, Basilyan AE, Tumskoy VE, Koulakov SA, Astakhov SN, Pavlova EYu, Anisimov MA. 2004. The Yana RHS Site: Humans in the Arctic Before the Last Glacial Maximum. *Science* **303**: 52–56.

Posth C, Nakatsuka N, Lazaridis I, Skoglund P, Mallick S, Lamnidis TC, Rohland N, Nägele K, Adamski N, Bertolini E, et al. 2018. Reconstructing the Deep Population History of Central and South America. *Cell* **175**: 1185-1197.e22.

Posth C, Wißing C, Kitagawa K, Pagani L, Van Holstein L, Racimo F, Wehrberger K, Conard NJ, Kind CJ, Bocherens H, et al. 2017. Deeply divergent archaic mitochondrial genome provides a lower time boundary for African gene flow into Neanderthals. *Nat Commun* **8**: 16046.

Postillone M, Cobos V, Urrutia C, Dejean C, Gonzalez P, Perez S, Bernal V. 2020. Mitochondrial DNA Diversity and Evolutionary History of Native Human Populations of Argentinean Northwest Patagonia. *Hub*. https://muse.jhu.edu/article/751584 (Accessed July 29, 2025).

Poznik GD, Henn BM, Yee M-C, Sliwerska E, Euskirchen GM, Lin AA, Snyder M, Quintana-Murci L, Kidd JM, Underhill PA, et al. 2013. Sequencing Y Chromosomes Resolves Discrepancy in Time to Common Ancestor of Males Versus Females. *Science* **341**: 562–565.

Raff JA, Bolnick DA, Tackney J, O'Rourke DH. 2011. Ancient DNA perspectives on American colonization and population history. *Am J Phys Anthropol* **146**: 503–514.

Raghavan M, Skoglund P, Graf KE, Metspalu M, Albrechtsen A, Moltke I, Rasmussen S, Stafford Jr TW, Orlando L, Metspalu E, et al. 2014. Upper Palaeolithic Siberian genome reveals dual ancestry of Native Americans. *Nature* **505**: 87–91.

Raghavan M, Steinrücken M, Harris K, Schiffels S, Rasmussen S, DeGiorgio M, Albrechtsen A, Valdiosera C, Ávila-Arcos MC, Malaspinas A-S, et al. 2015. Genomic evidence for the Pleistocene and recent population history of Native Americans. *Science* **349**: aab3884.

Rasmussen M, Anzick SL, Waters MR, Skoglund P, DeGiorgio M, Stafford TW, Rasmussen S, Moltke I, Albrechtsen A, Doyle SM, et al. 2014. The genome of a Late Pleistocene human from a Clovis burial site in western Montana. *Nature* **506**: 225–229.

Rasmussen M, Li Y, Lindgreen S, Pedersen JS, Albrechtsen A, Moltke I, Metspalu M, Metspalu E, Kivisild T, Gupta R, et al. 2010. Ancient human genome sequence of an extinct Palaeo-Eskimo. *Nature* **463**: 757–762.

Rasmussen M, Sikora M, Albrechtsen A, Korneliussen TS, Moreno-Mayar JV, Poznik GD, Zollikofer CPE, Ponce De León MS, Allentoft ME, Moltke I, et al. 2015. The ancestry and affiliations of Kennewick Man. *Nature* **523**: 455–458.

Resque R, Gusmão L, Geppert M, Roewer L, Palha T, Alvarez L, Ribeiro-dos-Santos Â, Santos S. 2016. Male Lineages in Brazil: Intercontinental Admixture and Stratification of the European Background, ed. F. Calafell. *PLoS ONE* **11**: e0152573.

Ribeiro-Dos-Santos AKC, Santos SEB, Luciamach A. Heterogeneity of Mitochondrial DNA Haplotypes in Pre-Columbian Natives of the Amazon Region.

Rieux A, Eriksson A, Li M, Sobkowiak B, Weinert LA, Warmuth V, Ruiz-Linares A, Manica A, Balloux F. 2014. Improved Calibration of the Human Mitochondrial Clock Using Ancient Genomes. *Molecular Biology and Evolution* **31**: 2780–2792.

Roca-Rada X, Politis G, Messineo PG, Scheifler N, Scabuzzo C, González M, Harkins KM, Reich D, Souilmi Y, Teixeira JC, et al. 2021. Ancient mitochondrial genomes from the Argentinian Pampas inform the early peopling of the Southern Cone of South America. *iScience* **24**: 102553.

Roewer L, Nothnagel M, Gusmão L, Gomes V, González M, Corach D, Sala A, Alechine E, Palha T, Santos N, et al. 2013. Continent-Wide Decoupling of Y-Chromosomal Genetic Variation from Language and Geography in Native South Americans, ed. S.M. Williams. *PLoS Genet* **9**: e1003460.

Rothhammer F, Fehren-Schmitz L, Puddu G, Capriles J. 2017. Mitochondrial

DNA haplogroup variation of contemporary mixed South Americans reveals

prehistoric displacements linked to archaeologically-derived culture history.

American J Hum Biol **29**: e23029.

Rowe TB, Stafford TW, Fisher DC, Enghild JJ, Quigg JM, Ketcham RA, Sagebiel

JC, Hanna R, Colbert MW. 2022. Human Occupation of the North American

Colorado Plateau ~37,000 Years Ago. *Front Ecol Evol* **10**: 903795.

Saillard J, Forster P, Lynnerup N, Bandelt H-J, Nørby S. 2000. mtDNA Variation

among Greenland Eskimos: The Edge of the Beringian Expansion. *The American

Journal of Human Genetics* **67**: 718–726.

Scheib CL, Li H, Desai T, Link V, Kendall C, Dewar G, Griffith PW, Mörseburg A,

Johnson JR, Potter A, et al. 2018. Ancient human parallel lineages within North

America contributed to a coastal expansion. *Science* **360**: 1024–1027.

Schurr TG. 2004. The Peopling of the New World: Perspectives from Molecular

Anthropology. *Annu Rev Anthropol* **33**: 551–583.

Simão F, Ribeiro J, Vullo C, Catelli L, Gomes V, Xavier C, Huber G, Bodner M,

Quiroz A, Ferreira AP, et al. 2021. The Ancestry of Eastern Paraguay: A Typical

South American Profile with a Unique Pattern of Admixture. *Genes* **12**: 1788.

Skoglund P, Mallick S, Bortolini MC, Chennagiri N, Hünemeier T, Petzl-Erler ML, Salzano FM, Patterson N, Reich D. 2015. Genetic evidence for two founding populations of the Americas. *Nature* **525**: 104–108.

Starikovskaya EB, Sukernik RI, Derbeneva OA, Volodko NV, Ruiz-Pesini E, Torroni A, Brown MD, Lott MT, Hosseini SH, Huoponen K, et al. 2005. Mitochondrial DNA Diversity in Indigenous Populations of the Southern Extent of Siberia, and the Origins of Native American Haplogroups. *Annals of Human Genetics* **69**: 67–89.

Starikovskaya YB, Sukernik RI, Schurr TG, Kogelnik AM, Wallace DC. 1998. mtDNA Diversity in Chukchi and Siberian Eskimos: Implications for the Genetic History of Ancient Beringia and the Peopling of the New World. *The American Journal of Human Genetics* **63**: 1473–1491.

Sun J, Ma P, Cheng H, Wang C, Li Y, Cui Y, Yao H, Wen S, Wei L. 2021. Post-last glacial maximum expansion of Y-chromosome haplogroup C2a-L1373 in northern Asia and its implications for the origin of Native Americans. *American J Phys Anthropol* **174**: 363–374.

Taboada-Echalar P, Álvarez-Iglesias V, Heinz T, Vidal-Bralo L, Gómez-Carballa A, Catelli L, Pardo-Seco J, Pastoriza A, Carracedo Á, Torres-Balanza A, et al. 2013.

The Genetic Legacy of the Pre-Colonial Period in Contemporary Bolivians, ed. C. Lalueza-Fox. *PLoS ONE* **8**: e58980.

Tackney JC, Potter BA, Raff J, Powers M, Watkins WS, Warner D, Reuther JD, Irish JD, O'Rourke DH. 2015. Two contemporaneous mitogenomes from terminal Pleistocene burials in eastern Beringia. *Proc Natl Acad Sci USA* **112**: 13833–13838.

Tamm E, Kivisild T, Reidla M, Metspalu M, Smith DG, Mulligan CJ, Bravi CM, Rickards O, Martinez-Labarga C, Khusnutdinova EK, et al. 2007. Beringian Standstill and Spread of Native American Founders, ed. D. Carter. *PLoS ONE* **2**: e829.

The Genographic Consortium, Jota MS, Lacerda DR, Sandoval JR, Vieira PPR, Ohasi D, Santos-Júnior JE, Acosta O, Cuellar C, Revollo S, et al. 2016. New native South American Y chromosome lineages. *J Hum Genet* **61**: 593–603.

Torroni A, Neel JV, Barrantes R, Schurr TG, Wallace DC. 1994. Mitochondrial DNA "clock" for the Amerinds and its implications for timing their entry into North America. *Proc Natl Acad Sci USA* **91**: 1158–1162.

University of Geneva, Switzerland, Ray N, Adams JM. 2001. A GIS-based Vegetation Map of the World at the Last Glacial Maximum (25,000-15,000 BP).

IA. http://intarch.ac.uk/journal/issue11/rayadams_index.html (Accessed July 29, 2025).

Villaescusa P, Seidel M, Nothnagel M, Pinotti T, González-Andrade F, Alvarez-Gila O, M. De Pancorbo M, Roewer L. 2021. A Y-chromosomal survey of Ecuador's multi-ethnic population reveals new insights into the tri-partite population structure and supports an early Holocene age of the rare Native American founder lineage C3-MPB373. *Forensic Science International: Genetics* **51**: 102427.

Wang W, Ding M, Gardner JD, Wang Y, Miao B, Guo W, Wu X, Ruan Q, Yu J, Hu X, et al. 2021. Ancient Xinjiang mitogenomes reveal intense admixture with high genetic diversity. *Sci Adv* **7**: eabd6690.

Wei L-H, Wang L-X, Wen S-Q, Yan S, Canada R, Gurianov V, Huang Y-Z, Mallick S, Biondo A, O'Leary A, et al. 2018. Paternal origin of Paleo-Indians in Siberia: insights from Y-chromosome sequences. *Eur J Hum Genet* **26**: 1687–1696.

Weissensteiner H, Pacher D, Kloss-Brandstätter A, Forer L, Specht G, Bandelt H-J, Kronenberg F, Salas A, Schönherr S. 2016. HaploGrep 2: mitochondrial haplogroup classification in the era of high-throughput sequencing. *Nucleic Acids Res* **44**: W58–W63.

Willerslev E, Meltzer DJ. 2021. Peopling of the Americas as inferred from ancient genomics. *Nature* **594**: 356–364.

Yang MA, Gao X, Theunert C, Tong H, Aximu-Petri A, Nickel B, Slatkin M, Meyer M, Pääbo S, Kelso J, et al. 2017. 40,000-Year-Old Individual from Asia Provides Insight into Early Population Structure in Eurasia. *Current Biology* **27**: 3202-3208.e9.

TITLE:

What uniparental genetic markers tell us about the prehistoric human

colonization of the Americas

INTRODUCTION:

The interpretation of the prehistoric peopling of the Americas has been a

multidisciplinary enterprise (Becerra-Valdivia and Higham 2020; Fiedel 2000).

From a molecular genetics perspective, the first foundations of these

interpretations were based on the information provided by the uniparentally

inherited mitochondrial DNA (mtDNA) and the non-recombining portion of the

Y-chromosome (NRY) on extant populations (Schurr 2004; Karafet et al. 1999).

As ancient DNA (aDNA) samples are a mixture of endogenous aDNA and

contaminating DNA, at first, its results on human samples were questioned.

Fortunately, new sequencing technologies have managed to overcome this

problem (Palencia-Madrid and De Pancorbo 2015), and current aDNA-based

analyses have added an invaluable timing perspective to these interpretations

(Bisso-Machado and Fagundes 2021). Later, technological improvements have

made it possible to sequence entire genomes in both present-day and fossil

individuals. Analyses of these genomes have revealed extinct ancestries and

genetically well-differentiated ancestries from the past that, admixed in

different proportions, persist in current Native American populations along the

American continent. This new information has served to improve previous

hypotheses and to formulate new ones (Willerslev and Meltzer 2021).

Thus, some conclusions have achieved general acceptance: 1) The Asiatic origin

of the Native American ancestors; 2) The first migration, in one or several

waves, occurred as a rapid movement towards the south of the double

continent; 3) The initial low population density led to strong genetic

differentiation geographically structured; 4) Subsequent migrations reshaped

the genetic landscape of North and South America. Important for the first was

the entrance from Beringia of several migratory waves that originated the

settlement of Paleo- and Neo-Eskimos; 5) Time comparisons of population

genetic profiles since post-Neolithic to present-day have uncovered an important genetic continuity (although see (Llamas et al. 2016)).

However, other propositions are still under debate as: 1) the precise Asian origin of the Native American ancestors; 2) the routes followed by these pioneers as they expanded throughout the Americas; and, perhaps, the most contentious question, 3) when these first spreads occurred? Although the most recent studies based on complete genomes favor a post-Glacial dispersal (Moreno-Mayar et al. 2018), with or without a prolonged standstill in Beringia (Faught 2017), the first works with mitochondrial DNA favored an early entry, around 30 kya before the Last Glacial Maximum (LGM), avoiding the problem of having to cross the ice sheets of North America (Bonatto and Salzano 1997; Torroni et al. 1994).

These age discrepancies are due to the molecular evolutionary rates used in each case. Unfortunately, the rate of molecular evolution is not a constant; it depends on several parameters, one of which, the effective population size (Ne), has changed dramatically since the Paleolithic to present-day (Cabrera 2020). Therefore, similar to radiocarbon dating, molecular rates need to be calibrated to obtain more accurate ages. One way to do this is by using ancient genomes and/or mitogenomes, but in the case of the American colonization, all the extracted aDNA postdates the LGM (Bisso-Machado and Fagundes

2021). The option chosen here is to use evolutionary rates calibrated with Paleolithic fossils to date Paleolithic events (Cabrera 2024).

Despite their unquestionable utility, uniparental markers are barely mentioned in whole-genome studies (Gusareva et al. 2025). But see (Moreno-Mayar et al. 2018; Krettek et al. 2025). The underlying reason for this abandonment is that these markers only offer monogenic insights, and that only the set of all genes provides congruent results. In my opinion, this assumption is flawed. The main cause of genetic differentiation used to estimate phylogenies and genetic distances between populations or species is isolation. Isolation affects whole genomes and does not discriminate by sex. Populations differentiate genetically, due to the loss of previous polymorphisms, by fixation or extinction, and the accumulation of new polymorphisms by mutation in a time-dependent way. Thus, any DNA fragment, including those coding for uniparental markers, should be valid to realize genetic comparisons whenever they have accumulated enough genetic differentiation. The generalized use of DNA barcoding in molecular taxonomy (Hubert and Hanner 2015) validates my previous assertion. Isolation is broken by migration/introgression, affecting uniparental markers and autosomes differently. Regardless of whether uniparental markers persist only in their original populations or whether both endure in the recipient population, their coalescence node—that is, the

distance to their most recent common ancestor—will remain unchanged. However, for mixed autosomes, if the introgressed fragments are not thoroughly masked, they will influence the genetic distances obtained between the populations/species involved. A paradigmatic example of that is the case of the controversial phylogenetic relationships obtained between modern humans and their closest relatives, Neanderthals and Denisovans. While mtDNA and NRY sequences showed modern humans and Neanderthals as sister lineages (Posth et al. 2017; Petr et al. 2020), autosomes joined Denisovan and Neanderthal as the closest pair, leaving modern humans as an out-group (Posth et al. 2017). Contrary to the widespread acceptance that the true relationship between hominids is that obtained with the autosomes, I have suggested that the valid phylogeny is the one deduced from the uniparental markers, while the one obtained from the autosomes is the result of later introgressions between the three groups of hominins (Cabrera 2021b).

Taking into account the above-mentioned particularities of the uniparental markers, in this work, I perform a thorough reanalysis of all published mitogenomes and the most recently published Y-chromosome results, using phylogenetic and phylogeographic tools, to reconsider and shed new light on the hypotheses formulated to explain the prehistoric colonization of the New World by Asian peoples. The main conclusions obtained are: 1) that the first

migration involved at least two Asian populations with most probable central and eastern Asian origins; 2) that this wave(s) occurred before 30 Kya; 3) that those pioneer colonizers did not stop in Beringia but kept trekking across the Americas leaving few genetic traces in past and present-day native American populations; 4) That after the LGM, small and relatively isolated resilient groups radiated from different areas to occupy new exploitable territories both in North and South America; 5) that these secondary expansive waves occurred first in southern- an meso-America than in northern North America; and 6) that gene flow existed among geographically adjacent groups.

RESULTS

All the phylogenetic and phylogeographic mtDNA results described below have been obtained from the trees presented in the supplementary figures (Suplemental_Fig_S1.xls (HgA2), Fig_S2.xls (HGB2), Fig_S3.xls (HgC1), Fig_S4.xls (HgD), and Fig_S5.xls (HgX2), and all the NRY results described below have been obtained from the trees already published by other authors (Wei et al. 2018; Grugni et al. 2019; Pinotti et al. 2019; Villaescusa et al. 2021; Colombo et al. 2022; Paz Sepúlveda et al. 2022).

The most probable origin of the American Asian ancestors

The main Native American mtDNA haplogroup founders can be divided into two groups: 1) those belonging to macro-Haplogroup M (C1, C4c, D1, D2a1, D4e1b, and D4h3a), and 2) those that belong to macro-Haplogroup N (A2, B2, and X2a). Curiously, the same grouping occurs when we look at the number of mutations accumulated in their basal stems that link them to the common nodes with Asian lineages. While C1, C4c, D1, D2a1 and D4h3a have two mutations, and D4e1b only one; A2 and B2 have five mutations and X2a four mutations (Table 1).

Table 1. Mutations on American basal branches to their common nodes with Asia							
American haplogroup	Mutations	Coalescent Age (Kya)	Asian haplogroup	Asian region			
A2	5	20,2	A-a1	NW China (Xinjiang)			
A10a	1	4,04	A10	Western Siberia			

B2	5	20,2	B4b1a	Altai					
C1	2	8,08	C4	Eastern Asia					
C4c	2	8,08	C4a'b'c	Eastern Asia					
D1	2	8,08	D4	Eastern Asia					
D4e1b	1	4,04	D4e1	Eastern Asia					
D2a1	2	8,08	D2a'b	Northeastern Siberia					
D4h3a	2	8,08	D4h3	NW China (Xinjiang)					
X2a	4	16,16	X2a'j	Tajikistan					

An unpaired t-test shows that the difference between groups is highly significant (t = 7.2164; df = 5; two-tailed P value = 0.0008). Furthermore, it is well known that macro-Haplogroup M is dominant in southern and eastern

Asia (Marrero et al. 2016), while macro-Haplogroup N is more frequent in western Eurasia (Fregel et al. 2015). Consequently, the same occurs with the Asian ancestral sequences/haplogroups that show the shortest distances with the Native American haplogroups. It has already been demonstrated that the closest relatives for the macro-Haplogroup M American lineages are found in eastern Asia/southeastern Siberia (Starikovskaya et al. 1998; Derenko et al. 2010). Some further commentaries are pertinent for these lineages: Hg C1 comprises four common and well spread sub lineages, three of them (C1b, C1c, C1d) practicably exclusive of the Americas and one (C1a) with a wide Asian range, and several rare clades/isolates, the most ancient of which is represented by two identical sequences from Brazil (Simão et al. 2021) and Paraguay (Avila et al. 2019) that carries twelve mutations in its basal stem (Supplementary_Fig_S3.xls). Curiously, the other isolates have been detected in northwestern Eurasia instead of East Asia: C1e in Iceland (Ebenesersdóttir et al. 2011), western Russia (Järve et al. 2019), and western China (Wang et al. 2021); C1f in Tajikistan (Peng et al. 2018) and C1g in Mesolithic remains from western Russia (Der Sarkissian et al. 2014). As the oldest clades are found in the New World, the most parsimonious conclusion is that all the C1 branches found in Eurasia were the result of retro migrations from America. The oldest Native American samples harboring C1b lineages are USR1 from Alaska

(Tackney et al. 2015) and I11974 from Chile (Posth et al. 2018), both around 12,000 years old; for C1c it is MW291667 from Argentina (Roca-Rada et al. 2021), approximately 8,500 years old; and for C1d it is Lapa01, 9,600 years old, from Brazil (Posth et al. 2018). Haplogroup D1 has the oldest samples in Yucatan, Mexico, represented by a Late Pleistocene skeleton (HN5/48) around 12 ky old (Chatters et al. 2014), in Spirit Cave, Nevada, around 11 kya (Moreno-Mayar et al. 2018), an approximately 10 ky old LLP.S2.E1 sample from Argentina (Roca-Rada et al. 2021), and the Sumidouro4, 9,645 years old, sample from Brazil (Moreno-Mayar et al. 2018). Until recently, the American haplogroup D4h3a was separated from its Asian root by six mutations, but recent aDNA studies (Mao et al. 2021) have rescued a Chinese sequence that shares four of the six basal substitutions with the American clade (Supplementary_Fig_S4.xls). The remains, from which this sequence was extracted, were unearthed at the Amur region, in northeastern China, and are around 14,000 years old. The oldest samples harboring D4h3a sequences in the New World are the 12,600-year-old Anzick1 from Montana (Rasmussen et al. 2014), the 10,300-year-old Shuká Káa from Alaska (Lindo et al. 2017), and the 10,400-year-old specimens from Lagoa Santa in Brazil (Moreno-Mayar et al. 2018).

In relation to the macro-Haplogroup N, for the New World haplogroup A2, the closest Asian sequences are from haplogroup A-a1 (BZL_M16A, BZL_M16B) and belong to historic samples from Xinjiang, western China (Wang et al. 2021). For haplogroup B2, it is AY519494, included in the Asian haplogroup B4b (Supplementary_Fig_S2.xls), being the Altai region as its geographic origin (Starikovskaya et al. 2005). Finally, for the American haplogroup X2a, the closest Asian relatives are into haplogroup X2j, that groups samples from Egypt and Europe (Supplementary_Fig_S5.xls) but, for geographic proximity, I consider as the closest Asian sequence a partial one (sample 40) from Tajikistan (Ovchinnikov et al. 2014), as it carries 16179 and 16357 substitutions that are terminal mutations in the Egyptian X2j (Kujanová et al. 2009) mitogenomes (Supplementary_Fig_S5.xls). For haplogroups A2 and B2 the most ancient sequences, around 14,000 years old, were extracted from human coprolites found in Paisley Caves, Oregon (Gilbert et al. 2008), next oldest for A2 is Lapa15 (around 8,700 years old) from Lapa do Santo, Brazil (Posth et al. 2018), and for B2 is USR2 (approximately 12,000 years old) from Upward Sun River site in Alaska (Tackney et al. 2015). The most ancient Native American sample with an X2a haplotype is the Kennewick Man from Washington, which is around 9,000 years old (Rasmussen et al. 2015).

A way to reconcile the close arrivals of the Asian ancestors carrying the Native

American M and N lineages with their significant differences in the number of

mutations accumulated on their basal stems is to assume that the route

followed across Asia of the N carriers was longer and slower than that of the M

carriers.

Finally, if we compare the number of lineages carried by the M and N groups to

their respective population sizes upon arrival in America, we could estimate

that the contribution of the eastern Asian contingent was 62.5% and that of

western Eurasia 37.5%.

The NRY does not show American colonization patterns as clear as those of

mtDNA, but using its phylogeny and phylogeography in the Americas and Asia I

have considered the ancestors of the American clades C-MPB373, that has its

closest haplotypes in the eastern Asian outgroup clade C-L1373 (Pinotti et al.

2019; Sun et al. 2021), QF746 that includes the Q-YP1475 subclade

represented by the Saqqaq ancient sample (Rasmussen et al. 2010) and has as

sister subclade Q-B143 represented by northeastern Siberian Koryaks (Karmin

et al. 2015), and Q-FT9713, that has its closest sequences in Mongolia and

northeastern China (https://www.yfull.com/tree/Q) as of eastern Asian

origin, while C-P39/Z30536, having as outgroup the Central Asian subclade C-

F1756 (Wei et al. 2018), Q-Z780, that includes the Native American ancient

sample Anzick1 (Rasmussen et al. 2014) and has Q-L330 of Central Asia ascription (Dulik et al. 2012; Flegontov et al. 2019) as sister clade, Q-M3, that has as out group Q-L804 with a northern European geographic range (Hallast et al. 2015), and the rare Q-Y1150 and Q-M378 (Grugni et al. 2019) as of western Eurasian origin. Unlike mtDNA, it appears that the vast majority of the American male lineages had their ancestors in western Eurasia. It is interesting to note that the C-MPB373 South American lineage is phylogenetically an out-group of C-F1699 that comprises numerous western Eurasian lineages, including also the Canadian C-P39/Z30536 branch (Pinotti et al. 2019). Also of interest is that the most abundant American clade, Q-M3 (Colombo et al. 2022), has as sister branch Q-L804, which is specific to northern Europe (Grugni et al. 2019). The oldest sample belonging to the eastern group of lineages is Lapa01, from Brazil, which is approximately 8,840 years old (Posth et al. 2018). For the western group, Anzick1 from Montana with an approximate age of 12,600 years represents the oldest North American sample (Grugni et al. 2019), whereas in South America, the oldest sample (I11974) was found in Los Rieles, Chile, being approximately 10,500 years old (Posth et al. 2018).

The global ages of the first arrival and expansions of the Native American parental ancestors

A global average of the different mean mutational data, calculated for each

American mitochondrial haplogroup using different approaches (Table 2), give

mean mutational values of 7.11 (95% CI: 6.34 – 7.88) substitutions at the

arrival in America of the founding lineages, and a mean of 4.42 (95%: 3.98 –

4.85) substitutions for their initial expansions into the American continent,

which, in turn, produce mean ages of 28.72 (95%: 25.61 – 31.84) kya and of

17.86 (95% CI: 16.08 – 19.59) kya for the founding and expansion events

respectively.

Table 2. Mutational means and coalescent mean ages for the main American mtDNA haplogroups								
HG	Phylogeny	Mean (Age Kya)	Type	N	95% CI: Mutation/Age(Kya)			
A2	Founding	7.55 (30,50)	Normal	1119 (Indiv)	7.37-7.73 / 29,77-31,23			
	Expansi	4.63	"	1083	4.46-4.79 / 18,02-			

	on	(18,70)		(Indiv)	19,35		
	Founding	7.25 (29,29)	Weighted	118 (Clades)	6.62-7.88 / 26,74-31,83		
	Expansion	4.35 (17,57)	"	118 (Clades)	4.04-4.66 / 16,32-18,83		
	Coalescent	9.13 (36,88)	Max (19 nt)	16 (Branches)	8.46-9.79 / 34,18-39,55		
B2	Founding	6.68 (26,99)	Normal	1081 (Indiv)	6.51-6.85 / 25,30-27,68		
	Expansion	4.68 (18.90)	"	1070 (Indiv)	4.51-4.85 / 18,22-19,00		
	Founding	6.62 (26,75)	Weighted	80 (Clades)	6.21-7.03 / 25,09-28,40		
	Expansion	4.69	"	80	4.28-5.11 / 17,29-		

	on	(18,95)		(Clades)	20,64			
	Coalescent	6.42 (25,93)	Max (16 nt)	14 (Branches)	5.12-7.72 / 20,68-31,17			
C1	Founding	6.50 (26,26)	Normal	887 (Indiv)	6.33-6.67 / 25.57-26,95			
	Expansion	4.36 (17,61)	"	873 (indiv)	4.17-4.55 / 16,84-18,38			
	Founding	6.33 (25,57)	Weighted	138 Clades)	6.05-6.61 / 24,44-26,71			
	Expansion	4.01 (16,20)	"	138 (Clades)	3.71-4.31 / 14,99-17,41			
	Coalescent	6.22 (25,13)	Max (16 nt)	42(Branches)	5.54-6.90 /22,38-27,88			
C4c	Founding	6.32	Normal	34 (Indiv)	5.74-6.91 /23,19-			

	ng	(25,53)			27,92		
	Expansion	4.32 (17,45)	"	34 (Indiv)	3.74-4.91 / 15,11-19,84		
	Founding	3.48 (14,06)	Weighted	8 (Clades)	2.85-4.11 /11,51-16,60		
	Expansion	1.30 (5,25)	"	8 (Clades)	0.67-1.93 /2,71-7,80		
	Coalescent	4.71 (19,03)	Max (9 nt)	3 (Branches)	2.47-6.96 /9,98-28,12		
D1	Founding	7.00 (28,28)	Normal	496 (Indiv)	6.73-7.27 / 27,19-29,37		
	Expansion	4.97 (20,08)	"	463 (Indiv)	4.68-5.27 / 18,91-21,29		
	Founding	7.02	Weight	58	6.53-7.51/26,38-		

	ng	(28,36)	ed	(Clades)	30,34
	Expansion	4.97 (20,08)	"	58 (Clades)	4.48-5.46 / 18,10-22,06
	Coalescent	7.14 (28,86)	Max (16 nt)	28(Branches)	6.67-7.62 / 26,93-30,79
D4h3a	Founding	6.20 (25,05)	Normal	80 (Indiv)	5.57-6.83 / 22,50-27,59
	Expansion	4.24 (17,13)	"	64 (Indiv)	3.63-4.85 /14,67-19,59
	Founding	6.27 (25,33)	weighted	13 (Clades)	5.73-6.81 / 23,15-27,51
	Expansion	4.24 (17,13)	"	13 (Clades)	3.80-4.68 / 15,35-18,91
	Coalescent	7.77 (31.39)	Max (12 nt)	12(Branches)	6.15-9.39 / 24,85-37,93

X2a	Foundi ng	7.83 (31,63)	Normal	41 (Indiv)	7.11-8.55 / 28,72-34,54			
	Expansi on	3.83 (15,47)	"	40 (indiv)	3.11-4.55 / 12,56-18,38			
	Foundi ng	6.83 (27,59)	Weight ed	2 (Clades)	5.37-8.29 /21,69-33,49			
	Expansi on	2.83 (11,43)	"	2 (Clades)	1.37-4.29 / 5,53-17,33			
	Coalesc ent	11.00 (44,44)	Max (15 nt)	2(Branch es)	-39.82-61.82/-160,87-249,75			

The founding ages for the mtDNA Native American haplogroups, exposed

above, coincide with the approximately 30 kya proposed in the first mtDNA

studies (Bonatto and Salzano 1997; Torroni et al. 1994; Starikovskaya et al.

1998), and with the calibrated ages calculated by (Rieux et al. 2014), based on

ancient mitogenomes. However, recent extant and ancient mitogenome

studies (Bisso-Machado and Fagundes 2021; Llamas et al. 2016) and whole

genome-based studies (Moreno-Mayar et al. 2018; Willerslev and Meltzer 2021) strongly support post-LGM ages for the settlement of the Americas, although this is in part inspired by the most conservative archaeological ages proposed for the American settlement.

The age for the first population expansions calculated here, 17.86 kya, after the rigors of the LGM had lessened on the continent, does not differ much from those proposed by most authors. The difference is that for most authors, this expansion coincides with its arrival in America proper, while here, the arrival in America occurred much earlier, around 30 kya, before the glacial period had its worst effects.

Concerning the NRY, it deserves mention that, assuming a first colonization of the Americas about 15 kya, (Poznik et al. 2013) estimated a substitution rate for the Y-chromosome of 8.2×10^{-10} (95% CI: $7.2 \times 10^{-10} - 9.2 \times 10^{-10}$) per site, year. However, doubling the time to 30 kya, as proposed here based on mtDNA, the substitution rate would be 4.1×10^{-10} per site, year which is very similar to that obtained by (Francalacci et al. 2013) of 5.3×10^{-10} (95% CI: 4.2×10^{-10}- 7.0×10^{-10}) per site, year and both estimates falling within the limits of the mean 4.7×10^{-10} (95% CI: $2.1 \times 10^{-10} - 9.5 \times 10^{-10}$) per site, year proposed by myself based on past population sizes obtained under phylogenetic considerations (Cabrera 2024). As before, these results contrast with the

majority of the NRY younger coalescence ages estimated by other authors (The Genographic Consortium et al. 2016; Pinotti et al. 2019; Colombo et al. 2022; Paz Sepúlveda et al. 2022).

As YFull Tree v13.03.00 (YTree) has the most actualized NRY phylogeny and the most extensive phylogeography, it is increasingly used as a source of data in scientific articles, as in the present work. The YFull substitution rate (Adamov et al. 2015) is identical to the one calculated by (Poznik et al. 2013) under the premise that the human American settlement occurred around 15 kya, because of this, I have halved their substitution rate values when I used them. Under these requirements, NRY haplogroup C clades would arrive in the Americas in separate waves, C-MPB373 first at about 32.2 kya (95% CI: 28.8 – 35.6 kya), and C-P39/Z30536 second at about 26.2 kya (95% CI: 22.8 – 29.8 kya). The three main NRY haplogroup Q clades have very similar ages around 30 kya: Q-F746, 31.8 kya (95% CI: 28.4 – 35.2); Q-Z780, 31.0 kya (95% CI: 29.0 – 33.0); Q-M3, 29.8 (27.0 – 32.8). The age of arrival to the Americas of the rare clades Q-M1150 (30.0 kya (95% CI:26.6 – 33.4 kya) and Q-M378, 21.4 kya (95% CI: 17.6 – 25.4 kya), that include European and Asian lineages, is only tentative (https://www.yfull.com/tree/Q).

Regional heterogeneity for founding and expansion events

In a first approach, I made comparisons of the native haplogroups' founding and expansion ages between the North and South American subcontinents (Table 3). Except for haplogroup C1, the results reflect significantly older ages in South America, both in founding and expansion events.

Table 3. Differences in Founding(F) and expansion(E) ages between the North America and South America subcontinents										
Haplogroup	t-value	df	p-value	oldest region						
A2 - F	3.0397	1119	0.0024	South America						
A2 -E	4.8326	1105	<0.0001	South America						
B2 - F	1.0604	1099	0.2892	ns						
B2 - E	3.1395	1083	0.0017	South America						
C1 - F	1.4777	499	0.1401	ns						

C1- E	0.4722	482	0.6370	ns
D1 - F	5.7700	498	<0.0001	South America
D1 - E	2.2473	466	0.0251	South America
D4h3a - F	3.0245	69	0.0035	South America
D4h3a - E	1.0115	59	0.3159	ns

More detailed comparison between geographic regions, within and among

subcontinents, using mean molecular age comparisons, pairwise genetic

distances and Pearson correlation tests are described in supplementary tables

(Supplemental Table S1.1 to S1.7 for A2; Supplemental Table S2.1 to S2.7 for

B2; Supplemental Table S3.14, .7, .10, .13, .16 for C1b; Supplemental Table

S3.2, .5, .8, .11, .14, .17 for C1c; Supplemental Table S.3.3, .6, .9, .12, .15, .18

for C1d; Supplemental Table S3.19 for C4c; Supplemental Table S4.1 to S4.6 for

D1; Supplemental Table S4.7 to S4.9 for D4h3, and Supplemental Table S5.1 to

S5.3 for X2a).

Haplogroup A2

Globally, A2 is the most abundant haplogroup, representing approximately 44% of the Native American population, but it has an uneven geographic distribution. The oldest founding ages for A2 (Supplemental_Table_S1.5) are found in Brazil and the Colombia-Isthmus area (≈ 34 kya), expanding first in Colombia around 23 kya. From the Colombian Isthmus region, significant latitudinal clines exist to Siberia (Slope -13.37; R^2 0.8396; P-value 0.0102), and to Tierra del Fuego (Slope 23.49; R^2 0.8091; P-value 0.0377). In addition, notice that in the most ancient samples of southern South America, mtDNA haplogroup A and B lineages were totally absent or very rare (Lalueza 1997; Moraga et al. 2000; Nakatsuka et al. 2020; Roca-Rada et al. 2021; Postillone et al. 2020) while they were detected in the most ancient samples recovered in northwestern North America more than 14 kya (Gilbert et al. 2008), and also were prominent in ancient samples from the Isthmus-Colombian area (Capodiferro et al. 2021). Another geographic peculiarity of haplogroup A2 is its significant abundance in the eastern areas compared to the western ones (Kemp et al. 2007; Raff et al. 2011; Rothhammer et al. 2017; Simão et al. 2021). It is also noteworthy that the oldest A2 haplotypes are found in North America and Brazil (Table S1.4). Furthermore, a secondary southward migration of some North American sub-lineages is also detectable. One of these lineages is

A2f, which is significantly older in North America (≈35,000 ya) than in Mexico (≈23,000 ya) (t=3.6172; df 22; p-value 0.0015).

For haplogroup A2, the shortest molecular genetic distances between regions (Table S1.3) were obtained for the pairs CSAm–SSAm (0.657), NWSAm–NESAm (0.682), and NCAm–NNAm (0.681). When these distances are represented in a PCoA (Fig. 1), it shows that the first principal coordinate places NESAm in a central position, leaving respectively at its left and right the remaining regions of North- and South-America. The second principal coordinate separates the regions of each sub-continent as latitudinal clines, mimicking the Pearson correlation results.

Haplogroup B2

With a global value of 24.0 %, hg B2 is the second-largest Native American haplogroup. The oldest founding (≈ 30 kya) and expansion (≈ 21 kya) regional values for this haplogroup (Supplementary_Table_ S2.6) are found in SSAm and CSAm. Going up from these southern areas, values descend in a clinal trend following a NWSAm orientation to reach Central and North America. This negative correlation between B2 molecular ages and latitude (Supplementary_Table_ S2.5) is highly significant (Slope: -13.57; R^2 0.5455; P-

values 0.0061). Unlike A2, Hg B2 is currently absent or very rare in northern North America, mimicking its practical absence in northeastern Siberia (Starikovskaya et al. 1998). Also, unlike A2 and D1, Hg B2 is more frequent in western continental regions than in the eastern ones (Bisso-Machado et al. 2012; Brandini et al. 2018). The oldest B2 haplotypes were found in the Andean region (Supplementary_Table_ S2.4).

The shortest genetic molecular distances were between SSAm-CSAm (0.492) and SSAm-NWSAm (0.500) pairs (Supplementary_Table_ S2.3). A PCoA bidimensional graphic representation of these distances shows NWSAm having a central position, with SSAm and CSAm at its left and NESAm and all northern American regions at its right, following a latitudinal cline as observed with the Pearson correlation (Fig. 2).

Haplogroup C1

Although in the beginning Hg C1 was considered as a homogenous clade, its three main native branches reflect different phylogeographic movements that need particular attention.

For Hg C1b, the oldest founding/expansion regional ages (Supplementary_Table_ S3.13) are in NESAm (≈ 29 kya/17 kya), with younger

ages going clinally down to Tierra del Fuego and up to Canada. The latter

showing a significant trend (Slope -12.20; RSquare 0.8876; P-value 0.0166).

These phylogeographic characteristics are rather similar to those found for

HgA2. However, the longitudinal differentiation observed for the A2 clade is

not seen in C1b. The oldest haplotypes for C1b are found in northern South

America, including both Ecuador and Colombia, and reaching Mexico

(Supplementary_Table_ S3.10). Furthermore, the shortest genetic distance

between regions (Supplementary_Table_ S3.7) is that of the pair NWSAm-

NESAm (0.522), followed by NWSAm-SSAm (0.605) and SSAm-CSAm (0.654). In

this case, the PCoA graphic representation highlights the close relationship

between NWSAm-Mexico and USA-Canada, leaving NESAm as the center from

which southward movements across Brazil and northward to Central America

occurred (Fig. 3).

For Hg C1c, the oldest founding/expansion regional ages

(Supplementary_Table_ S3.14) have their center in NWSAm (≈ 34 kya / 26 kya).

From this center, populations with younger ages conform southern and

northern geographic transects ending in SSAm and Canada, respectively, the

latter showing a significant correlation with latitude both for founding (Slope -

14.79; R^2 0.8055; P-value 0.0152) and expansion ages (Slope -11.71; R^2 0.9629;

P-value 0.0005). These results partially resemble those obtained for Hg B2 in its North American subcontinent range.

Congruent with its Andean center, the oldest haplotypes for C1c are found in NWSAm and Mexico (Supplementary_Table_ S3.11).

The shortest genetic distances within Hg C1c (Supplementary_Table_S3.8) point to the existence of three geographic centers, the first represented by the CSAm-CAm pair (0.571), the second by the SSAm-NWSam pair (0.630), and the third by the NWSAm-Mexico pair (0.644). This pattern could be explained supposing NWSAm as the main center that spread downward to Argentina and Chile and upward to Mexico, and a later center that, from Brazil, reached Central America, deleting in this region the signal of the previous route that from NWSAm had reached Mexico. In this respect, it is worth mentioning that the Y-chromosome Q-M925, mainly Mesoamerican, has close phylogenetic ties with the Brazilian Q-Y26547 subclade (Colombo et al. 2022). This complex pattern is partially reflected in the PCoA plot (Fig. 4), which shows a latitudinal cline from NWSA to Canada passing through NESAm, with the anomalous inclusion of SSAm in it, and an independent close relationship between Central America and Brazil.

Hg C1d, with the oldest founding/expansion ages (Supplemental_Table_ S3.15) in CSAm and SSAm (≈ 38 kya / 33 kya), is the oldest C1 branch. In fact, it is the oldest Native American clade. Although without reaching significance, C1d shows a geographic descendant trend in population ages from North America to Central America (Supplemental_Table_S3.15). This pattern resembles the one found for Hg A2. In addition, it presents a significant latitudinal correlation from SSAm to Central America with the oldest founding (Slope -16.65; R^2 0.7649; P-value 0.0415) and expansion ages (Slope -13.29; R^2 0.6985; P-value 0.0382) in southern South America. This geographic trend in the South American subcontinent parallels the one described for Hg B2. The oldest haplotypes for C1d are found in CSAm and SSAm (Supplemental_Table_S3.12), reinforcing the existence of an ancestral southern South American center of expansion for this clade.

For C1d, the closest genetic distances (Supplemental_Table_S3.9) are between the SSAm-CSAm pair (0.591) and the NWSAm-NESAm pair (0.592). The latter one resembles the close genetic relationship between these regions, also observed for the C1b clade. The PCoA bidimensional representation of these distances (Fig. 5) shows a central group joining SSAm and CSAm with Central America, and two distant groups, one at the left joining NWSAm and NESAm, and the other at the right, grouping the northern North American populations

and Mexico. The independent and close genetic relationship between the South American regions and Central America is similar to the one found for C1c, which was explained supposing a later expansion from Brazil to Central America, while the close genetic affinity between NWSAm-NESAm is a replica of what was found for C1b.

Haplogroup C4c

Founding and expansion ages for C4c are around 25 kya and 17 kya, respectively (Table 2), but ages for its main branches are very different (Supplemental_Table_S3.19). The most developed branch, C4c1, has founding and expansion ages of approximately 16 kya and 12 kya, respectively, both after the LGM. However, the less frequent C4c2 sister branch has a founding age of 31 kya, more in line with the rest of the main Native American haplogroups, but a very recent expansion age of only about 3000 ya. C4c is a minoritarian haplogroup with a limited geographic range that, except for a sporadic occurrence in Colombia and in an ancient sample from Brazil, Loca Do Suin_9100 years old (Ferraz et al. 2023), is restricted to NNAm (Supplemental_Fig_S3). Thus, the anomalous results obtained for this lineage may be due to poor sampling and/or strong drift effects.

Haplogroup D1

For D1, the oldest founding ages (≈ 31 kya) are found in SSAm and CSAm (Supplementary_Table_S4.5). The earliest expansion ages are also found in the same regions but at different times (CSAm ≈ 23 kya; SSAm ≈ 18 kya). For D1 regional founding ages, a significant latitudinal correlation exists from SSAm to Central America (Slope -16.65; R^2 0.5431; P-value 0.0370), and a non-significant decreasing trend is observed from Canada to Central America (Supplementary_Table_S4.5). The oldest D1 haplotypes are mainly in NWSAm and then in SSAm (Supplementary_Table_S4.4).

The D1 shortest genetic distances are for the pairs SSAm-CSAm (0.618), USA-Canada (0.667), and those involving NWSAm with SSAm (0.768) and with NESAm (0.759). A PCoA plot (Fig. 6) graphically reflects these genetic relationships, grouping in a central position all the Mesoamerican and northern South American populations, well separated from two, in turn, very distant groups, composed of northern North American and southern South American populations, respectively.

The D1 phylogeographic data described above show a great similarity with the patterns obtained for haplogroups B2 and C1d.

Haplogroup D4h3a

For D4h3a, the oldest founding (≈ 30 kya) and expansion ages (≈ 22 kya)

occurred in CSAm (Supplementary_Table_S4.8). Age comparisons between

regions only reach significance when the oldest expansion ages of SSAm and

CSAm are compared to those of NNAm and NWSAm (Supplementary Table

S4.9). Living apart, one isolates from NESAm, the oldest D4h3a haplotypes are

in NWSAm, Mexico, and consistently, in Chile (Supplementary_Table_S4.7). It

has been proposed that D4h3a is a signal of a Pacific southward expansion

(Perego et al. 2009). However, these results do not support that suggestion. It

seems that D4h3a was more common in the past and most frequent in eastern

areas than it is today.

Haplogroup X2

Haplogroup X2a is considered a minor Native American haplogroup with a

limited spatial range confined to northern North America (Perego et al. 2009).

The founding and expansion ages for haplogroup X2a are approximately 29 kya

and 16 kya, respectively (Supplementary_Table_S5.1), and the oldest

haplotypes are found in the X2a1 branch in samples from the North American

Great Plains. Ages are significantly older in North America than in Canada

(Supplementary Table S5.3). However, the detection of a divergent X2

haplotype in Canada (Perego et al. 2009), that defines a possible new Native

American haplogroup, named X2g (Supplemental_Fig_S5), deserves mention.

Although the modern and ancient geographic distribution of X2a is biased to the northeast of North America (Perego et al. 2009), with sporadic incidences in Mexico (Peñaloza-Espinosa et al. 2007; Bodner et al. 2021), its oldest sample, the Kennewick Man, about 9000 years old, harboring a basal X2a haplotype, was exhumed in the northwestern Washington state (Rasmussen et al. 2015), raising the possibility that the X2a clade could be originated in the northern Pacific area (Perego et al. 2009).

NRY haplogroups

At a global scale, two major parental lineages with Pan-American distribution have been defined (Q-CTS1780/Z780 and Q-M3), both within haplogroup Q, the first in lower frequencies everywhere compared to the second. In addition, two minor lineages, belonging to haplogroup C, with a subcontinental range (C-MPB373 and C-P39/Z30536) have also been considered Native American founder lineages. The southern C-MPB373 has been reported in extant Colombians and Equatorians consistently (Karafet et al. 1999; Roewer et al. 2013; Mezzavilla et al. 2015; Villaescusa et al. 2021). The northern C-P39/Z30536 has been found in Canada and northern North America, reaching also Mexico (Perez-Benedico et al. 2016), and drawing a northern to southern gradient of decreasing diversity (Dulik et al. 2012). Although phylogenetically Q-CTS1780/Z780 is the oldest clade, the demographic expansion observed

after the LGM occurred, in males, almost exclusively within the Q-M3 lineage that simultaneously branched, mainly in Mesoamerica (Q-M925) and in South America (Q-M848), producing well differentiated regional subclades (Bortolini et al. 2003; Battaglia et al. 2013; Roewer et al. 2013; Geppert et al. 2015; The Genographic Consortium et al. 2016; Grugni et al. 2019; Pinotti et al. 2019; Gómez et al. 2021; Villaescusa et al. 2021; Colombo et al. 2022; Kemp et al. 2007; Paz Sepúlveda et al. 2022). Some of these minor Q subclades also deserve mention due to their geographic distributions and STR diversity. One of them is Q-MEH2, this lineage was detected in northeast Siberian Koryaks and its very close relationship to the extinct Paleo-Eskimo Saqqaq (Rasmussen et al. 2010) suggested the possibility that it could trace a recent migration from Siberia to America across the Bering Strait (Malyarchuk et al. 2011). However, Q-MEH2 has been recently detected in Nahua from southern Mexico, and from its STR variability, it was proposed that it arrived much earlier in Mexico than in northern America (Gómez et al. 2021). Another striking example is the Mexican Q-PV3 lineage, in principle, it was considered a minor branch of Q-L54, but its quite divergent haplotypic variability suggested the possibility that it might represent another founding Native American lineage (Gómez et al. 2021). This subcontinental differentiation of male ancestries is in support of diverse demographic histories in North and South America (Bortolini et al. 2003).

However, before to reach to any conclusion, because post-Columbian admixture heavily affected the Native American male genetic pool, in both North (Malhi et al. 2007) and South America (Resque et al. 2016), a consultation on what NRY ancient DNA analyses say is necessary. The picture was similar. Although Q-CTS1780 could be more abundant in the past in both subcontinents (Posth et al. 2018; Grugni et al. 2019; Ferraz et al. 2023), Q-M3 was also the predominant NRY lineage in all ancient DNA data (Bisso-Machado and Fagundes 2021). However, the detection in the past of today's extinct lineages (Posth et al. 2018; Colombo et al. 2022) supports the view that strong founding and drift effects occurred in NRY Native American lineages. These data do not fit well with the hypothesis of a rapid and recent migratory wave from the North to explain the prehistoric colonization of South America.

DISCUSSION

The early entry to the Americas of the founder Native uniparental lineages, more than 30 kya, proposed here, makes the hypotheses of a prolonged (Hoffecker et al. 1993; Tamm et al. 2007) or short (Pinotti et al. 2019) standstill in Beringia, a previous expansion of its ancestors into Asia (Wei et al. 2018; Sun et al. 2021; Ning et al. 2020), or the existence of a Pacific Coastal Route around 20 kya (Scheib et al. 2018; Davis and Madsen 2020; Li et al. 2023), unnecessaries.

The archaeological evidence for the presence of humans in the Siberian Arctic around 30 kya (Pitulko et al. 2004) and, possibly, since 45 kya (Pitulko et al. 2017) is in support of a possible early entrance in America of its Native ancestors. More difficult is to obtain a general acceptance, from archaeologists, of human occupation in the American continent before and around the LGM, but it seems very probable that this occurred around 16-18 kya in Monte Verde, Chile (Dillehay et al. 2015), in Santa Elina site, Central Brazil during the LGM, at Chiquihuite Cave in Central Mexico about 30 kya (Ardelean et al. 2020), in the Colorado Plateau more than 36 kya (Rowe et al. 2022), in White Sands, New Mexico, during a stratigraphic record that spans from 24 kya to 17 kya (Holliday et al. 2025), and in Bluefish Caves, Canada, approximately 24 kya (Bourgeon et al. 2017).

Ancient genome-based studies have described the ancestral Native American population as an admixture of northern Eurasians and northern East Asian populations (Raghavan et al. 2014). The same dual contribution was proposed previously using NRY information (Karafet et al. 1999; Lell et al. 2002; Roewer et al. 2013), and an identical conclusion has been reached here based on mtDNA data. Genomic research proposed that the ancestral Native Americans arrived on the New Continent as an already admixed population (Moreno-Mayar et al. 2018). However, although coalescence ages do not show

significant differences between haplogroups, the NRY phylogeny is more compatible with the arrival of two consecutive waves, the first carrying C-MPB373 and Q-Z780 lineages, and the second being the carriers of C-P39/Z30536 and Q-M3 lineages. The evidence is weaker for mtDNA, but, as mentioned before, the fact that the basal branches joining haplogroups C and D to their respective Asian nodes are significantly shorter than those of A and B could be taken as an argument to favor a first arrival into America of the former pair. Another clue could lie in the Southern Cone of South America, the region furthest from the entrance to the continent. Ancient and current sampling of the region discovered that haplogroups C and D were the first and unique lineages reaching the area being even today the most prevalent (Lalueza 1997; García-Bour et al. 2000; Moraga et al. 2000; Demarchi et al. 2001; Nakatsuka et al. 2020; Postillone et al. 2020). The prevalence of haplogroups C and D has also been observed in Amazonian populations from Brazil (Merriwether and Ferrell 1996), which could be the origin of an early southward migration. In addition, it has been demonstrated that mtDNA diversity increases from north to south for haplogroups C1 and D1, whereas North American populations show higher diversity levels for haplogroups A2 and B2 (Lee and Merriwether 2015). Moreover, in North America, haplogroups A, B, and X have undergone population clinal expansions, while there is no

evidence of this spread in haplogroups C and D (Malhi et al. 2001). These

patterns are all compatible with two consecutive waves of migration. However,

these results could also be attributed to drift effects, as samples in other

regions also show the total absence of some founder mtDNA haplogroups, as is

the case of the Kuna from Panama in which the absence of C and D has been

documented (Batista et al. 1995) and, contrarily, the Ayoreo from Bolivia

showed a total absence of A and B (Dornelles et al. 2004). Based on the

distribution of two rare mtDNA haplogroups, D4h3 and X2a, the first spreading

into the Americas along the Pacific coast and the second entering through the

ice-free corridor, two migratory waves from Beringia were proposed (Perego et

al. 2009). However, for the case of D4h3, the authors admitted that the fact

that it is very rare in North America and more frequent and with greater

variation in South America, needs some additional explanation (Perego et al.

2009).

Another proposition, based on genome studies, is that after it arrives in

North America, the native ancestral population splits into two distinct

branches known as South Native Americans (SNA) and North Native Americans

(NNA) (Moreno-Mayar et al. 2018) or their approximate counterparts, Ancestry

A (ANC-A) and Ancestry B (ANC-B), respectively (Scheib et al. 2018). While the

northern ancestry is geographically rather limited to Canada and eastern North

American regions, the southern ancestry is detectable in the oldest genomic

sequence sampled, Anzick1_12700 years old from Montana (Rasmussen et al.

2014), and in the Spirit Cave sample from Nevada, 10700 years old (Moreno-

Mayar et al. 2018) extending southwards to Mesoamerica and southern

America (Moreno-Mayar et al. 2018). There are mono-parental markers that

share the geographic range of the NNA genomic ancestry as the mtDNA

haplogroups X2a (Brown et al. 1998) and C4c (Kashani et al. 2012) and the NRY

lineages C-P39 (Pinotti et al. 2019), Q-Y4276 (Grugni et al. 2019). The situation

is much more complex when the transcontinental distribution of the southern

component of the Native American genome has to be explained. The initial

dual explanation of southward Pacific coastal expansion in which the

successive populations could be represented as admixtures, in different

proportions, of the SNA and NNA components (Scheib et al. 2018) was proved

to be insufficient (Posth et al. 2018). First of all, the uniqueness of the

northernmost SNA component, represented by the Clovis associated Anzick1

sample (SNA1), could not explain the genomic compositions of many south

American populations and an additional, new and dominant non-Clovis SNA2,

represented in part by the Spirit Cave ancestry, had to be added to properly

explain the genome composition of many southern populations. Entering

Mesoamerica, the Mexican Mixe needed the addition of gene flow from an

unsampled population to be fully represented (Moreno-Mayar et al. 2018).

Moreover, several yet undiscovered genomic ancestries had to be added to

satisfactorily model the genomic structure of many South American well-

differentiated regional populations. An enigmatic case was the presence in

Amazonian Surui, Karitiana, and Xavante tribes of a divergent ancestry related

to Australasian people (Raghavan et al. 2015; Skoglund et al. 2015). This

component, named "population Y," was confirmed in ancient Brazilian samples

as Sumidouro5, approximately 10000 years old (Moreno-Mayar et al. 2018). At

least one mtDNA haplotype had to accompany the Australasian ancestry, and,

due to its southernmost distribution, a lineage from haplogroup B seems to be

the best candidate. However, with which ancestral Asian population it entered

the New World is difficult to determine since, on the one hand, basic lineages

of B were present in eastern Asia around 40 kya, as attested by the Chinese

Tianyuan sample mtDNA analysis (Fu et al. 2013). Moreover, the Australasian

ancestry detected in the South American Amazonians is also shared by the

Tianyuan sample (Yang et al. 2017). Therefore, the Australasian component

could enter America with the descendants of a population deeply settled in

eastern Asia, which had ancient Australasian ties, but, likewise, it could enter

with the carriers of mtDNA haplogroup B from central Eurasia. The Isthmus-

Colombian area was also a center of genome complexity, and a Pleistocene

ancestry was brought to the Panamanians, and it remained restricted to the Isthmus area (Capodiferro et al. 2021), and hunter-gatherers from the Colombian Altiplano also harbored an unknown basal lineage that seems derived from the initial South American genomic radiation (Krettek et al. 2025). In the northwest side of South America, a previous undetected ancestry was characteristic of northern Ecuador and Colombian populations (Barbieri et al. 2019), and Native Peruvians from the Lake Titicaca region showed a Siberian component also present in ancient North Americans as Anzick1, Kennewick, or Saqqaq (Lindo et al. 2018). The complex ancestries found in South America forced some followers of the rapid post-glacial colonization of America to make some cautionary comments, as that a possible northward radiation from South America to explain the Pacific migratory wave would require a more complex history (Posth et al. 2018), or that the SNA dispersal pattern south of the Continent's ice sheets involved complex admixture events between earlier established populations (Moreno-Mayar et al. 2018). The analysis of the uniparental markers realized in this study offers an alternative hypothesis that could integrate all the genomic information under a different perspective.

If we accept that the entry of Asian populations that gave rise to Native Americans occurred before 30 kya, the history of human settlement in America can be told differently. It is known that since its arrival, the weather gradually

worsened, covering with ice the regions near the Arctic and Antarctic (Clark et

al. 2009), clearing the Amazon and Central American rainforests (Häggi et al.

2017), and transforming most of the territory into forest and dry steppes

(University of Geneva, Switzerland et al. 2001). During this harsh phase,

humans had to live in small isolated bands, and adapt to different glacial

refugia that would ensure their survival, therefore, leaving few archaeological

traces of their existence. When the weather improved, these isolated bands

grew demographically and spread to new, now habitable territories. Thus,

differently from the fast southern wave colonization, the American continent

was not void of previous human occupation, and the post-LGM colonization of

the New World was not accomplished by one or several consecutive spreads

from northern North America to settle first Mesoamerica and after South

America, but from more or less contemporaneous radiations in multiple

geographic centers the majority of which, uniparental markers, situate in South

America. One of the main expansion centers for mtDNA is the northwest

Colombian-Isthmus area, where double latitudinal younger clines northwards

and southwards are detectable for haplogroups A2 and C1b. A similar trend in

A2 relative frequencies has also been detected in the same area (Keyeux et al.

2002). Another apparent center is located in the Andean region where mtDNA

haplogroup C1c shows its oldest foundation and expansion ages, with younger

geographic clines running up to Canada and down to SSAm. This pattern

parallels the one found for haplogroup B that expanded from Chile and the

Andean region, entering into Mesoamerica. However, it seems that, at least in

part, the B2 cline is the result of its displacement in North America by other

mtDNA lineages since it was present very early in northeast Asia as an

ancestral B lineage (Fu et al. 2013) and in Alaska as a basal B2 lineage (Tackney

et al. 2015) to practically disappear later in both areas. The highest frequencies

and genetic diversity values for haplogroup B2 have been found in the Andean

Altiplano (Gayà-Vidal et al. 2011; Taboada-Echalar et al. 2013), in the Bolivian

Piedmont (Corella et al. 2007), and in the Argentinian Gran Chaco (Demarchi et

al. 2001). Although the classic route to explain the B2 cline is to assume a

descent from California (Taboada-Echalar et al. 2013), South American origins,

as proposed here, have also been previously formulated from the Peruvian

Andes (Arencibia et al. 2025) or the Chilean Southern Cone (Bravi et al. 2024).

Also, as B2, C1d is another mtDNA clade that shows its most ancient founding

and expansion ages in the southern Cone, going up, with younger values, to

Central America, presenting a significant geographic correlation. In North

America, some lineages of this clade present an expansion age descendant

trend that ends in Central America; likewise, the observed trend for

haplogroup A2 sub-lineages. C1a (Tamm et al. 2007) is a northeast Asiatic

branch of C1 that, together with the minor European sub-clades C1e

(Ebenesersdóttir et al. 2011) and C1f (Der Sarkissian et al. 2014), suggests a

subsequent retro-migration from America (Tamm et al. 2007). Applying the

evolutionary rate proposed here, the founding age in Asia of C1a is 26.02 kya

(95% CI: 18.58-33.49 kya), which would signal a return to the Asian continent

with worsening weather conditions. Its expansion, in the Asian continent,

around 13.9 kya (95% CI: 6.46-21.37 kya) would indicate a Mesolithic spread

like the one found for the northeast European C1f clade (Der Sarkissian et al.

2014). This possible early American reflux can also be deduced from the

phylogeny and phylogeography of the Y-chromosome haplogroups C-L1373

(Pinotti et al. 2019; Colombo et al. 2022) and Q-L54 (Grugni et al. 2019; Pinotti

et al. 2019).

Another mtDNA haplogroup that signals to the southern Cone region as an

important center of expansion is D1, showing a significant negative correlation

from this area to Central America, with mean coalescent ages younger going

upwards, although with a longitudinal bias leaning towards central and eastern

South American regions. D1 has the greatest frequencies at the eastern areas

of the Andean region, with regional ancient clades in Central Argentina (García

et al. 2012) and the southern Cone (Bodner et al. 2012). Likewise, for A2 and

C1d, a southward expansion from Canada to Central America is detected for

D1. These expansive movements could be the cause of the B2 retraction

observed in North America. The suggestion that mtDNA haplogroup D4h3a

would confirm the hypothesis of a Pacific coastal route for the peopling of

America (Perego et al. 2009), subsequently reinforced by genomic studies

(Moreno-Mayar et al. 2018), is not supported by the evidence presented here.

The oldest founding and expansion ages for D4h3a are found in SSAm and

CSAm, being significantly older than those observed in NWSAm and NNAm

(Supplementary Table S4.9). Furthermore, although today absent, D4h3a has

been detected in ancient samples from Alaska, Shuká-Káa, 10.3 kya (Lindo et

al. 2017), British Columbia, Canada, 939, 6.5 kya (Lindo et al. 2017), and

Montana, Anzick 1, 12.6 kya (Rasmussen et al. 2014). Furthermore, it was

observed in ancient remains from the Brazilian Amazon, Lagoa Santa, 10 kya

(Moreno-Mayar et al. 2018). This pattern is very similar to the one observed

for B2. Thus, better than modelling a southward Pacific Coastal expansion,

both markers seem to represent the remnants of a more extensive occupation

in the past, which was displaced by more recent human migrations.

A particular commentary deserves the Amazonian area (CSAm). It participated

in all mtDNA centers of radiation commented above, and it is described as a

large reservoir of mtDNA deep lineages (De Souza et al. 2025; Di Corcia et al.

2017), and stands out for the ancient presence of very basal Y-chromosome

lineages as C-L1373*(Colombo et al. 2022), or the detection there of lineages that were considered exclusive to the northern subcontinent as Q-Y4303*(Grugni et al. 2019). Moreover, this genetic complexity has been corroborated by whole-genome-based studies in ancient specimens (Moreno-Mayar et al. 2018) and in extant populations (Ribeiro-Dos-Santos et al. 2020; Capodiferro et al. 2023). This genetic landscape has its correlate in the linguistic richness of the area. It is enough to point out that of the 71 linguistic isolates described for the double continent, more than 45% have been found in Brazil compared to 15% observed in North and Mesoamerica as a whole (Blench and Cb). It is expected that future archaeological discoveries in the area will unequivocally confirm human settlements much older than those currently accepted. Again, the differences found in genetic and linguistic complexity between the northern (NNA) and the southern (SNA) American branches make the simultaneous colonization of both subcontinents from a North American southward spread very improbable.

MATERIAL AND METHODS:

Material

For the mtDNA phylogenetic and phylogeographic analyses, I searched partial

and complete mitogenomes at the NCBI GenBank,

www.ncbi.nlm.nih.gov/genbank/; Mitomap,www.mitomap.org/MITOMAP;

EMPOP, https://empop.online; Ian Logan 2023,

www.ianlogan.co.uk/sequences_by_group/haplogroup_select.htm; Genome

Warehouse (GWH) public repository, https://bigd.big.ac.cn/gwh, and AmtDB,

www.http://amtdb.org; in addition, for phylogeographic analyses only, I also

searched YFull, MTree. The NRY phylogenetic and phylogeographic analyses

were based on the ISOGG database, http://www.isogg.org/tree/, YFull YTree

v13.03.00, YTree, and the data compiled by other authors (Wei et al. 2018;

Grugni et al. 2019; Pinotti et al. 2019; Gómez et al. 2021; Colombo et al. 2022;

Paz Sepúlveda et al. 2022). For brevity, the Y-chromosome lineages in the text

are referred to by the letter of their basic haplogroup and their terminal

mutation.

Phylogeny

Maximum parsimony intraspecific phylogenetic trees (Bandelt et al. 1999)

were constructed by hand, and haplogroup nomenclature was according to the

PhyloTree database Building 17, http://www.phylotree.org. Haplogroup

classification was confirmed using HaploGrep 3 3.2.1, https://haplogrep.i-

med.ac.at. (Weissensteiner et al. 2016). New subgroups not classified in

PhyloTree were provisionally named by the first letter and digit of the primary haplogroup to which they belong, followed by an asterisk and consecutive numbers. When a more formal nomenclature exists in YFullMTree, both labels are shown on the corresponding tree.

Evolutionary rates

It is well known that the mtDNA evolutionary rate suffers a time-dependent effect, slowing down going backwards (Henn et al. 2008). The main cause of this effect has been attributed to differences in population size, with the smallest sizes in Paleolithic times and an exponential growth in present times (Cabrera 2021a). To compensate this anomaly, I have used an estimated mitogenome Paleolithic substitution rate of 1.5×10^{-8} (95% CI: 0.42 - 9.07 x 10-8) per site per year (assuming a mtDNA genomic length of 16,500 base pairs) that is, one substitution in every 4,040 years (Cabrera 2024), to calculate haplogroup coalescent ages for the initial prehistoric settlement of the Americas. Following a similar reasoning, a putative mean evolutionary rate of 0.47×10^{-9} (95% CI: 0.21 – 0.95 x 10-9) per site per year was proposed to calculate coalescent ages of the Y-chromosome (Cabrera 2024).

Ages

I have performed three approaches to calculate haplogroup coalescent ages: 1) using statistics rho (Forster et al. 1996) and Sigma (Saillard et al. 2000) on individuals within haplogroups, and the evolutionary rates proposed above; 2) using clades within the main haplogroups to calculate a pooled mean to use normal average and standard error statistics to calculate mutation mean and standard errors for haplogroups ; 3) As the sample sizes of the clades have large differences, I calculated weighted averages and weighted standard errors to use them with normal statistics as above; 4) One of the most commented genetic drift effects is the possibility that haplotypes in low frequency become much more frequent in a population. However, it should be noted that this possibility is an exception more than a rule; it is normal for rare haplotypes, located at the tails of a distribution, to become extinct after drift, whereas those around the mean to augment in frequency. As it is evident that the Native American founding haplogroups suffered genetic drift fluctuations in the past, I have used the most divergent lineages left within haplogroups as possible past means to calculate their coalescence ages (Cabrera 2022)(Cabrera 2021a).

Phylogeography

For phylogeographic purposes the American continent has been divided into the following seven geographic regions: 1) North northern America (NNAm)

including Alaska and Canada; 2) North America, comprising the entire United States (Nam); 3) Mesoamerica, including Mexico, Belize, Guatemala, El Salvador, Honduras and Nicaragua (Mam); 4) Northeastern south America or Isthmus-Colombian area, comprising Costa Rica, Panama and Colombia; 5) Northwestern South America (NWSAm) or Andean region, including Ecuador, Peru and Bolivia; 6) Central South America (CSAm) including Brazil, Paraguay and Uruguay; 7) South southern America (SSAm) joining Argentina and Brazil. In addition, within regions, when necessary, it has been distinguished between inland vs coastal Pacific and coastal Atlantic areas and between Highland Andes vs Amazonian areas.

To reinforce the phylogenetic and phylogeographic analyses, I have implemented some simple algorithms, which I describe below.

Genetic distances

Besides the molecular distance based on the number of mutations between two homologous DNA sequences (F_{ST}), I have implemented a phylogeographic distance (D_{XY}) based on the genetic affinity (I_{XY}) between regions X and Y, measured as the number of times they form sister branches within a haplogroup tree (n_{XY}) versus the number of times each one participates as a sister branch within the haplogroup being measured (n_X and n_Y):

Then D_{XY} is calculated as $1 - I_{XY}$.

In addition, when searching for sister branches between X and Y regions within a haplogroup, I also recorded the cases in which one sequence, belonging to one of the regions involved, was a direct outgroup of the pair, signaling it as ancestral with respect to the other region considered derived. The significance of this relation was checked by observed-expected chi-square statistics.

Also, for phylogeographic purposes, I have searched, within each major Native American founder haplogroup, the regions where their oldest founder sub-haplogroups are found and the regions from which they first expanded.

Finally, to find the Asian centers from which the ancestors of the major American founder clades might have departed, I have looked for the most ancient Asian sequences (mutationally less evolved) that come from the Asian-American shared phylogenetic nodes, and which are the regions where these sequences are found.

Other statistics

Analysis of molecular variance (AMOVA), correlation/linear regression, and graphical Principal coordinates analysis (PCoA) have been implemented with the GenAlEx 6.5 software (Peakall and Smouse 2012). Descriptive normal statistics, 95% confidence intervals of means, and two-tailed t-tests were

implemented with GraphPad https://www.graphpad.com/quickcalcs/.

Weighted mean and standard errors were obtained with the Weighted calculator (https://calculatodo.com).

False positive correction

As I have performed multiple statistical tests for each founder haplogroup, I have applied a Bonferroni p-value adjustment. However, as the Bonferroni correction is very conservative, I have also used the Benjamin and Hochberg 1995(Benjamini and Hochberg 1995) false discovery rate (FDR) algorithm as a less restrictive alternative.

REFERENCES

Adamov D, Guryanov V, Karzhavin S, Tagankin V, Urasin V. 2015. Defining a New Rate Constant for Y-Chromosome SNPs based on Full Sequencing Data.

Ardelean CF, Becerra-Valdivia L, Pedersen MW, Schwenninger J-L, Oviatt CG, Macías-Quintero JI, Arroyo-Cabrales J, Sikora M, Ocampo-Díaz YZE, Rubio-Cisneros II, et al. 2020. Evidence of human occupation in Mexico around the Last Glacial Maximum. *Nature* **584**: 87–92.

Arencibia V, Muñoz M, Crespo CM, Russo MG, Vera P, Lia VV, García Guraieb S, Goñi RA, Avena S, Puebla A, et al. 2025. Novel B2 mitogenomes from

Continental southern Patagonia's Late Holocene: New insights into the peopling of the Southern Cone. *American Journal of Biological Anthropology* **186**: e24822.

Avila E, Graebin P, Chemale G, Freitas J, Kahmann A, Alho CS. 2019. Full mtDNA genome sequencing of Brazilian admixed populations: A forensic-focused evaluation of a MPS application as an alternative to Sanger sequencing methods. *Forensic Science International: Genetics* **42**: 154–164.

Bandelt HJ, Forster P, Rohl A. 1999. Median-joining networks for inferring intraspecific phylogenies. *Molecular Biology and Evolution* **16**: 37–48.

Barbieri C, Barquera R, Arias L, Sandoval JR, Acosta O, Zurita C, Aguilar-Campos A, Tito-Álvarez AM, Serrano-Osuna R, Gray RD, et al. 2019. The Current Genomic Landscape of Western South America: Andes, Amazonia, and Pacific Coast. *Molecular Biology and Evolution* **36**: 2698–2713.

Batista O, Kolman CJ, Bermingham E. 1995. Mitochondrial DNA diversity in the Kuna Amerinds of Panama. *Human Molecular Genetics* **4**: 921–929.

Battaglia V, Grugni V, Perego UA, Angerhofer N, Gomez-Palmieri JE, Woodward SR, Achilli A, Myres N, Torroni A, Semino O. 2013. The First Peopling of South America: New Evidence from Y-Chromosome Haplogroup Q ed. D. O'Rourke. *PLoS ONE* **8**: e71390.

Becerra-Valdivia L, Higham T. 2020. The timing and effect of the earliest human arrivals in North America. *Nature* **584**: 93–97.

Benjamini Y, Hochberg Y. 1995. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. *Journal of the Royal Statistical Society Series B: Statistical Methodology* **57**: 289–300.

Bisso-Machado R, Bortolini MC, Salzano FM. 2012. Uniparental genetic markers in South Amerindians. *Genet Mol Biol* **35**: 365–387.

Bisso-Machado R, Fagundes NJR. 2021. Uniparental genetic markers in Native Americans: A summary of all available data from ancient and contemporary populations. *American J Phys Anthropol* **176**: 445–458.

Blench R, Cb C. [DRAFT CIRCULATED FOR COMMENT].

Bodner M, Perego UA, Gomez JE, Cerda-Flores RM, Rambaldi Migliore N, Woodward SR, Parson W, Achilli A. 2021. The Mitochondrial DNA Landscape of Modern Mexico. *Genes* **12**: 1453.

Bodner M, Perego UA, Huber G, Fendt L, Röck AW, Zimmermann B, Olivieri A, Gómez-Carballa A, Lancioni H, Angerhofer N, et al. 2012. Rapid coastal spread of First Americans: Novel insights from South America's Southern Cone mitochondrial genomes. *Genome Res* **22**: 811–820.

Bonatto SL, Salzano FM. 1997. Diversity and Age of the Four Major mtDNA Haplogroups, and Their Implications for the Peopling of the New World. *The American Journal of Human Genetics* **61**: 1413–1423.

Bortolini M-C, Salzano FM, Thomas MG, Stuart S, Nasanen SPK, Bau CHD, Hutz MH, Layrisse Z, Petzl-Erler ML, Tsuneto LT, et al. 2003. Y-Chromosome Evidence for Differing Ancient Demographic Histories in the Americas. *The American Journal of Human Genetics* **73**: 524–539.

Bourgeon L, Burke A, Higham T. 2017. Earliest Human Presence in North America Dated to the Last Glacial Maximum: New Radiocarbon Dates from Bluefish Caves, Canada ed. J.P. Hart. *PLoS ONE* **12**: e0169486.

Brandini S, Bergamaschi P, Cerna MF, Gandini F, Bastaroli F, Bertolini E, Cereda C, Ferretti L, Gómez-Carballa A, Battaglia V, et al. 2018. The Paleo-Indian Entry into South America According to Mitogenomes. *Molecular Biology and Evolution* **35**: 299–311.

Bravi CM, Motti JMB, García A. 2024. Letter to the editor: A Southern Cone origin rather than Peruvian affinities for ancient Patagonian B2 mitogenomes. *American Journal of Biological Anthropology* **184**: e24934.

Brown MD, Hosseini SH, Torroni A, Bandelt H-J, Allen JC, Schurr TG, Scozzari R, Cruciani F, Wallace DC. 1998. mtDNA Haplogroup X: An Ancient Link between

Europe/Western Asia and North America? *The American Journal of Human Genetics* **63**: 1852–1861.

Cabrera VM. 2020. Counterbalancing the time-dependent effect on the human mitochondrial DNA molecular clock. *BMC Evol Biol* **20**: 78.

Cabrera VM. 2024. Early male and female footprints of modern humans across Eurasia and Australasia. *Medical Research Archives* **12**.

Cabrera VM. 2021a. Human molecular evolutionary rate, time dependency and transient polymorphism effects viewed through ancient and modern mitochondrial DNA genomes. *Sci Rep* **11**: 5036.

Cabrera VM. 2021b. Revisiting the Hominin Phylogeny: An Alternative Introgression Scenario to Reconcile the Uniparental and autosomal DNA Topologies. *Jpeb* **9**.

Capodiferro MR, Aram B, Raveane A, Rambaldi Migliore N, Colombo G, Ongaro L, Rivera J, Mendizábal T, Hernández-Mora I, Tribaldos M, et al. 2021. Archaeogenomic distinctiveness of the Isthmo-Colombian area. *Cell* **184**: 1706-1723.e24.

Capodiferro MR, Chero Osorio AM, Rambaldi Migliore N, Tineo Tineo DH, Raveane A, Xavier C, Bodner M, Simão F, Ongaro L, Montinaro F, et al. 2023.

The multifaceted genomic history of Ashaninka from Amazonian Peru. *Current Biology* **33**: 1573-1581.e5.

Chatters JC, Kennett DJ, Asmerom Y, Kemp BM, Polyak V, Blank AN, Beddows PA, Reinhardt E, Arroyo-Cabrales J, Bolnick DA, et al. 2014. Late Pleistocene Human Skeleton and mtDNA Link Paleoamericans and Modern Native Americans. *Science* **344**: 750–754.

Clark PU, Dyke AS, Shakun JD, Carlson AE, Clark J, Wohlfarth B, Mitrovica JX, Hostetler SW, McCabe AM. 2009. The Last Glacial Maximum. *Science* **325**: 710–714.

Colombo G, Traverso L, Mazzocchi L, Grugni V, Rambaldi Migliore N, Capodiferro MR, Lombardo G, Flores R, Karmin M, Rootsi S, et al. 2022. Overview of the Americas' First Peopling from a Patrilineal Perspective: New Evidence from the Southern Continent. *Genes* **13**: 220.

Corella A, Bert F, Pérez-Pérez A, Gené M, Turbón D. 2007. Mitochondrial DNA diversity of the Amerindian populations living in the Andean Piedmont of Bolivia: Chimane, Moseten, Aymara and Quechua. *Annals of Human Biology* **34**: 34–55.

Davis LG, Madsen DB. 2020. The coastal migration theory: Formulation and testable hypotheses. *Quaternary Science Reviews* **249**: 106605.

De Souza FG, Matos GB, Sena Santos C, Souza TP, Gobbo AR, Da Costa PF, Salgado CG, Lopes Eufraseo G, Vitor De Souza Fernandes A, Santos-Lobato BL, et al. 2025. Mitochondrial ancestry from complete mitogenomes highlights a lack of characterization of indigenous haplogroups in Brazilian Amazon population. *Commun Biol* **8**: 835.

Demarchi DA, Panzetta-Dutari GM, Motran CC, De Los Angeles López De Basualdo M, Marcellino AJ. 2001. Mitochondrial DNA haplogroups in Amerindian populations from the Gran Chaco. *American J Phys Anthropol* **115**: 199–203.

Der Sarkissian C, Brotherton P, Balanovsky O, Templeton JEL, Llamas B, Soubrier J, Moiseyev V, Khartanovich V, Cooper A, Haak W, et al. 2014. Mitochondrial Genome Sequencing in Mesolithic North East Europe Unearths a New Sub-Clade within the Broadly Distributed Human Haplogroup C1 ed. A. Achilli. *PLoS ONE* **9**: e87612.

Derenko M, Malyarchuk B, Grzybowski T, Denisova G, Rogalla U, Perkova M, Dambueva I, Zakharov I. 2010. Origin and Post-Glacial Dispersal of Mitochondrial DNA Haplogroups C and D in Northern Asia ed. M.T.P. Gilbert. *PLoS ONE* **5**: e15214.

Di Corcia T, Sanchez Mellado C, Davila Francia TJ, Ferri G, Sarno S, Luiselli D, Rickards O. 2017. East of the Andes: The genetic profile of the Peruvian Amazon populations. *American J Phys Anthropol* **163**: 328–338.

Dillehay TD, Ocampo C, Saavedra J, Sawakuchi AO, Vega RM, Pino M, Collins MB, Scott Cummings L, Arregui I, Villagran XS, et al. 2015. New Archaeological Evidence for an Early Human Presence at Monte Verde, Chile ed. J.P. Hart. *PLoS ONE* **10**: e0141923.

Dornelles CL, Battilana J, Fagundes NJR, Freitas LB, Bonatto SL, Salzano FM. 2004. Mitochondrial DNA and *Alu* insertions in a genetically peculiar population: The Ayoreo Indians of Bolivia and Paraguay. *American J Hum Biol* **16**: 479–488.

Dulik MC, Zhadanov SI, Osipova LP, Askapuli A, Gau L, Gokcumen O, Rubinstein S, Schurr TG. 2012. Mitochondrial DNA and Y Chromosome Variation Provides Evidence for a Recent Common Ancestry between Native Americans and Indigenous Altaians. *The American Journal of Human Genetics* **90**: 229–246.

Ebenesersdóttir SS, Sigurðsson Á, Sánchez-Quinto F, Lalueza-Fox C, Stefánsson K, Helgason A. 2011. A new subclade of mtDNA haplogroup C1 found in icelanders: Evidence of pre-columbian contact? *American J Phys Anthropol* **144**: 92–99.

Faught MK. 2017. Where was the PaleoAmerind standstill? *Quaternary International* **444**: 10–18.

Ferraz T, Suarez Villagran X, Nägele K, Radzevičiūtė R, Barbosa Lemes R, Salazar-García DC, Wesolowski V, Lopes Alves M, Bastos M, Rapp Py-Daniel A, et al. 2023. Genomic history of coastal societies from eastern South America. *Nat Ecol Evol* **7**: 1315–1330.

Fiedel SJ. 2000. The Peopling of the New World: Present Evidence, New Theories, and Future Directions. *Journal of Archaeological Research* **8**: 39–103.

Flegontov P, Altınışık NE, Changmai P, Rohland N, Mallick S, Adamski N, Bolnick DA, Broomandkhoshbacht N, Candilio F, Culleton BJ, et al. 2019. Palaeo-Eskimo genetic ancestry and the peopling of Chukotka and North America. *Nature* **570**: 236–240.

Forster P, Harding R, Torroni A, Bandelt H-J. 1996. Origin and Evolution of Native American mtDNA Variation: A Reappraisal. *Am J Hum Genet*.

Francalacci P, Morelli L, Angius A, Berutti R, Reinier F, Atzeni R, Pilu R, Busonero F, Maschio A, Zara I, et al. 2013. Low-Pass DNA Sequencing of 1200 Sardinians Reconstructs European Y-Chromosome Phylogeny. *Science* **341**: 565–569.

Fregel R, Cabrera V, Larruga JM, Abu-Amero KK, González AM. 2015. Carriers of Mitochondrial DNA Macrohaplogroup N Lineages Reached Australia around 50,000 Years Ago following a Northern Asian Route ed. F. Calafell. *PLoS ONE* **10**: e0129839.

Fu Q, Meyer M, Gao X, Stenzel U, Burbano HA, Kelso J, Pääbo S. 2013. DNA analysis of an early modern human from Tianyuan Cave, China. *Proc Natl Acad Sci USA* **110**: 2223–2227.

García A, Pauro M, Nores R, Bravi CM, Demarchi DA. 2012. Phylogeography of mitochondrial haplogroup D1: An early spread of subhaplogroup D1j from Central Argentina. *American J Phys Anthropol* **149**: 583–590.

García-Bour J, Pérez-Pérez A, Prats E, Turbón D. 2000. MOLECULAR APPROACH TO THE PEOPLING OF THE AMERICAS BY SEQUENCING mtDNA FROM EXTINCT FUEGUIANS AND PATAGONS. *Chungará (Arica)* **32**.

Gayà-Vidal M, Moral P, Saenz-Ruales N, Gerbault P, Tonasso L, Villena M, Vasquez R, Bravi CM, Dugoujon J. 2011. mtDNA and Y-chromosome diversity in Aymaras and Quechuas from Bolivia: Different stories and special genetic traits of the Andean Altiplano populations. *American J Phys Anthropol* **145**: 215–230.

Geppert M, Ayub Q, Xue Y, Santos S, Ribeiro-dos-Santos Â, Baeta M, Núñez C, Martínez-Jarreta B, Tyler-Smith C, Roewer L. 2015. Identification of new SNPs

in native South American populations by resequencing the Y chromosome. *Forensic Science International: Genetics* **15**: 111–114.

Gilbert MTP, Jenkins DL, Götherstrom A, Naveran N, Sanchez JJ, Hofreiter M, Thomsen PF, Binladen J, Higham TFG, Yohe RM, et al. 2008. DNA from Pre-Clovis Human Coprolites in Oregon, North America. *Science* **320**: 786–789.

Gómez R, Vilar MG, Meraz-Ríos MA, Véliz D, Zúñiga G, Hernández-Tobías EA, Figueroa-Corona MDP, Owings AC, Gaieski JB, Schurr TG. 2021. Y chromosome diversity in Aztlan descendants and its implications for the history of Central Mexico. *iScience* **24**: 102487.

Grugni V, Raveane A, Ongaro L, Battaglia V, Trombetta B, Colombo G, Capodiferro MR, Olivieri A, Achilli A, Perego UA, et al. 2019. Analysis of the human Y-chromosome haplogroup Q characterizes ancient population movements in Eurasia and the Americas. *BMC Biol* **17**: 3.

Gusareva ES, Ghosh AG, Kharkov VN, Khor S-S, Zarubin A, Moshkov N, Kalsi N, Ratan A, Heinle CE, Cooke N, et al. 2025. From North Asia to South America: Tracing the longest human migration through genomic sequencing. *Science* **388**: eadk5081.

Häggi C, Chiessi CM, Merkel U, Mulitza S, Prange M, Schulz M, Schefuß E. 2017. Response of the Amazon rainforest to late Pleistocene climate variability. *Earth and Planetary Science Letters* **479**: 50–59.

Hallast P, Batini C, Zadik D, Maisano Delser P, Wetton JH, Arroyo-Pardo E, Cavalleri GL, De Knijff P, Destro Bisol G, Dupuy BM, et al. 2015. The Y-Chromosome Tree Bursts into Leaf: 13,000 High-Confidence SNPs Covering the Majority of Known Clades. *Molecular Biology and Evolution* **32**: 661–673.

Henn BM, Gignoux CR, Feldman MW, Mountain JL. 2008. Characterizing the Time Dependency of Human Mitochondrial DNA Mutation Rate Estimates. *Molecular Biology and Evolution* **26**: 217–230.

Hoffecker JF, Powers WR, Goebel T. 1993. The Colonization of Beringia and the Peopling of the New World. *Science* **259**: 46–53.

Holliday VT, Windingstad JD, Bright J, Phillips BG, Butler JB, Breslawski R, Bowman JE. 2025. Paleolake geochronology supports Last Glacial Maximum (LGM) age for human tracks at White Sands, New Mexico. *Science AdvAnceS*.

Hubert N, Hanner R. 2015. DNA Barcoding, species delineation and taxonomy: a historical perspective. *DNA Barcodes* **3**. http://access.portico.org/stable?au=phx3c2h64c3 (Accessed July 29, 2025).

Järve M, Saag L, Scheib CL, Pathak AK, Montinaro F, Pagani L, Flores R, Guellil M, Saag L, Tambets K, et al. 2019. Shifts in the Genetic Landscape of the Western Eurasian Steppe Associated with the Beginning and End of the Scythian Dominance. *Current Biology* **29**: 2430-2441.e10.

Karafet TM, Zegura SL, Posukh O, Osipova L, Bergen A, Long J, Goldman D, Klitz W, Harihara S, De Knijff P, et al. 1999. Ancestral Asian Source(s) of New World Y-Chromosome Founder Haplotypes. *The American Journal of Human Genetics* **64**: 817–831.

Karmin M, Saag L, Vicente M, Sayres MAW, Järve M, Talas UG, Rootsi S, Ilumäe A-M, Mägi R, Mitt M, et al. 2015. A recent bottleneck of Y chromosome diversity coincides with a global change in culture. *Genome Res* **25**: 459–466.

Kashani BH, Perego UA, Olivieri A, Angerhofer N, Gandini F, Carossa V, Lancioni H, Semino O, Woodward SR, Achilli A, et al. 2012. Mitochondrial haplogroup C4c: A rare lineage entering America through the ice-free corridor? *American J Phys Anthropol* **147**: 35–39.

Kemp BM, Malhi RS, McDonough J, Bolnick DA, Eshleman JA, Rickards O, Martinez-Labarga C, Johnson JR, Lorenz JG, Dixon EJ, et al. 2007. Genetic analysis of early holocene skeletal remains from Alaska and its implications for the settlement of the Americas. *American J Phys Anthropol* **132**: 605–621.

Keyeux G, Rodas C, Gelvez N, Carter D. 2002. Possible Migration Routes into South America Deduced from Mitochondrial DNA Studies in Colombian Amerindian Populations. *Human Biology* **74**: 211–233.

Krettek K-L, Casas-Vargas A, Mas-Sandoval A, Alvis LA, Reiter E, Madero JM, Urban M, Vargas JCN, Usaquén W, Cuenca JVR, et al. 2025. A 6000-year-long genomic transect from the Bogotá Altiplano reveals multiple genetic shifts in the demographic history of Colombia. *Science AdvAnceS*.

Kujanová M, Pereira L, Fernandes V, Pereira JB, Černý V. 2009. Near Eastern Neolithic genetic input in a small oasis of the Egyptian Western Desert. *American J Phys Anthropol* **140**: 336–346.

Lalueza C. 1997. Lack of founding Amerindian mitochondrial DNA lineages in extinct aborigines from Tierra del Fuego-Patagonia. *Human Molecular Genetics* **6**: 41–46.

Lee, Merriwether. 2015. Identification of Whole Mitochondrial Genomes from Venezuela and Implications on Regional Phylogenies in South America. *Human Biology* **87**: 29.

Lell JT, Sukernik RI, Starikovskaya YB, Su B, Jin L, Schurr TG, Underhill PA, Wallace DC. 2002. The Dual Origin and Siberian Affinities of Native American Y Chromosomes. *The American Journal of Human Genetics* **70**: 192–206.

Li Y-C, Gao Z-L, Liu K-J, Tian J-Y, Yang B-Y, Rahman ZU, Yang L-Q, Zhang S-H, Li C-T, Achilli A, et al. 2023. Mitogenome evidence shows two radiation events and dispersals of matrilineal ancestry from northern coastal China to the Americas and Japan. *Cell Reports* **42**: 112413.

Lindo J, Achilli A, Perego UA, Archer D, Valdiosera C, Petzelt B, Mitchell J, Worl R, Dixon EJ, Fifield TE, et al. 2017. Ancient individuals from the North American Northwest Coast reveal 10,000 years of regional genetic continuity. *Proc Natl Acad Sci USA* **114**: 4093–4098.

Lindo J, Haas R, Hofman C, Apata M, Moraga M, Verdugo RA, Watson JT, Llave CV, Witonsky D, Beall C, et al. 2018. The genetic prehistory of the Andean highlands 7000 years BP though European contact. *SCIENCE ADVANCES*.

Llamas B, Fehren-Schmitz L, Valverde G, Soubrier J, Mallick S, Rohland N, Nordenfelt S, Valdiosera C, Richards SM, Rohrlach A, et al. 2016. Ancient mitochondrial DNA provides high-resolution time scale of the peopling of the Americas. *Sci Adv* **2**: e1501385.

Malhi RS, Kemp BM, Eshleman JA, Cybulski J, Smith DG, Cousins S, Harry H. 2007. Mitochondrial haplogroup M discovered in prehistoric North Americans. *Journal of Archaeological Science* **34**: 642–648.

Malhi RS, Schultz BA, Smith DG. 2001. Distribution of Mitochondrial DNA Lineages among Native American Tribes of Northeastern North America. *Human Biology* **73**: 17–55.

Malyarchuk B, Derenko M, Denisova G, Maksimov A, Wozniak M, Grzybowski T, Dambueva I, Zakharov I. 2011. Ancient links between Siberians and Native Americans revealed by subtyping the Y chromosome haplogroup Q1a. *J Hum Genet* **56**: 583–588.

Mao X, Zhang H, Qiao S, Liu Y, Chang F, Xie P, Zhang M, Wang T, Li M, Cao P, et al. 2021. The deep population history of northern East Asia from the Late Pleistocene to the Holocene. *Cell* **184**: 3256-3266.e13.

Marrero P, Abu-Amero KK, Larruga JM, Cabrera VM. 2016. Carriers of human mitochondrial DNA macrohaplogroup M colonized India from southeastern Asia. *BMC Evol Biol* **16**: 246.

Merriwether DA, Ferrell RE. 1996. The Four Founding Lineage Hypothesis for the New World: A Critical Reevaluation. *Molecular Phylogenetics and Evolution* **5**: 241–246.

Mezzavilla M, Geppert M, Tyler-Smith C, Roewer L, Xue Y. 2015. Insights into the origin of rare haplogroup C3* Y chromosomes in South America from high-

density autosomal SNP genotyping. *Forensic Science International: Genetics* **15**: 115–120.

Moraga ML, Rocco P, Miquel JF, Nervi F, Llop E, Chakraborty R, Rothhammer F, Carvallo P. 2000. Mitochondrial DNA polymorphisms in Chilean aboriginal populations: Implications for the peopling of the southern cone of the continent. *Am J Phys Anthropol* **113**: 19–29.

Moreno-Mayar JV, Vinner L, De Barros Damgaard P, De La Fuente C, Chan J, Spence JP, Allentoft ME, Vimala T, Racimo F, Pinotti T, et al. 2018. Early human dispersals within the Americas. *Science* **362**: eaav2621.

Nakatsuka N, Luisi P, Motti JMB, Salemme M, Santiago F, D'Angelo Del Campo MD, Vecchi RJ, Espinosa-Parrilla Y, Prieto A, Adamski N, et al. 2020. Ancient genomes in South Patagonia reveal population movements associated with technological shifts and geography. *Nat Commun* **11**: 3868.

Ning C, Fernandes D, Changmai P, Flegontova O, Yüncü E, Maier R, Altınışık NE, Kassian AS, Krause J, Lalueza-Fox C, et al. 2020. The genomic formation of First American ancestors in East and Northeast Asia.
http://biorxiv.org/lookup/doi/10.1101/2020.10.12.336628 (Accessed July 29, 2025).

Ovchinnikov IV, Malek MJ, Drees K, Kholina OI. 2014. Mitochondrial DNA variation in Tajiks living in Tajikistan. *Legal Medicine* **16**: 390–395.

Palencia-Madrid L, De Pancorbo MM. 2015. Next generation sequencing as a suitable analysis for authentication of degraded and ancient DNA. *Forensic Science International: Genetics Supplement Series* **5**: e338–e340.

Paz Sepúlveda PB, Mayordomo AC, Sala C, Sosa EJ, Zaiat JJ, Cuello M, Schwab M, Rodríguez Golpe D, Aquilano E, Santos MR, et al. 2022. Human Y chromosome sequences from Q Haplogroup reveal a South American settlement pre-18,000 years ago and a profound genomic impact during the Younger Dryas ed. F. Calafell. *PLoS ONE* **17**: e0271971.

Peakall R, Smouse PE. 2012. GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research—an update. *Bioinformatics* **28**: 2537–2539.

Peñaloza-Espinosa RI, Arenas-Aranda D, Cerda-Flores RM, Buentello-Malo L, González-Valencia G, Torres J, Álvarez B, Mendoza I, Flores M, Sandoval L, et al. 2007. Characterization of mtDNA Haplogroups in 14 Mexican Indigenous Populations. *hub* **79**: 313–320.

Peng M-S, Xu W, Song J-J, Chen X, Sulaiman X, Cai L, Liu H-Q, Wu S-F, Gao Y, Abdulloevich NT, et al. 2018. Mitochondrial genomes uncover the maternal history of the Pamir populations. *Eur J Hum Genet* **26**: 124–136.

Perego UA, Achilli A, Angerhofer N, Accetturo M, Pala M, Olivieri A, Kashani BH, Ritchie KH, Scozzari R, Kong Q-P, et al. 2009. Distinctive Paleo-Indian Migration Routes from Beringia Marked by Two Rare mtDNA Haplogroups. *Current Biology* **19**: 1–8.

Perez-Benedico D, La Salvia J, Zeng Z, Herrera GA, Garcia-Bertrand R, Herrera RJ. 2016. Mayans: a Y chromosome perspective. *Eur J Hum Genet* **24**: 1352–1358.

Petr M, Hajdinjak M, Fu Q, Essel E, Rougier H, Crevecoeur I, Semal P, Golovanova LV, Doronichev VB, Lalueza-Fox C, et al. 2020. The evolutionary history of Neanderthal and Denisovan Y chromosomes. *Science* **369**: 1653–1656.

Pinotti T, Bergström A, Geppert M, Bawn M, Ohasi D, Shi W, Lacerda DR, Solli A, Norstedt J, Reed K, et al. 2019. Y Chromosome Sequences Reveal a Short Beringian Standstill, Rapid Expansion, and early Population structure of Native American Founders. *Current Biology* **29**: 149-157.e3.

Pitulko V, Pavlova E, Nikolskiy P. 2017. Revising the archaeological record of the Upper Pleistocene Arctic Siberia: Human dispersal and adaptations in MIS 3 and 2. *Quaternary Science Reviews* **165**: 127–148.

Pitulko VV, Nikolsky PA, Girya EYu, Basilyan AE, Tumskoy VE, Koulakov SA, Astakhov SN, Pavlova EYu, Anisimov MA. 2004. The Yana RHS Site: Humans in the Arctic Before the Last Glacial Maximum. *Science* **303**: 52–56.

Posth C, Nakatsuka N, Lazaridis I, Skoglund P, Mallick S, Lamnidis TC, Rohland N, Nägele K, Adamski N, Bertolini E, et al. 2018. Reconstructing the Deep Population History of Central and South America. *Cell* **175**: 1185-1197.e22.

Posth C, Wißing C, Kitagawa K, Pagani L, Van Holstein L, Racimo F, Wehrberger K, Conard NJ, Kind CJ, Bocherens H, et al. 2017. Deeply divergent archaic mitochondrial genome provides lower time boundary for African gene flow into Neanderthals. *Nat Commun* **8**: 16046.

Postillone M, Cobos V, Urrutia C, Dejean C, Gonzalez P, Perez S, Bernal V. 2020. Mitochondrial DNA Diversity and Evolutionary History of Native Human Populations of Argentinean Northwest Patagonia. *hub*. https://muse.jhu.edu/article/751584 (Accessed July 29, 2025).

Poznik GD, Henn BM, Yee M-C, Sliwerska E, Euskirchen GM, Lin AA, Snyder M, Quintana-Murci L, Kidd JM, Underhill PA, et al. 2013. Sequencing Y

Chromosomes Resolves Discrepancy in Time to Common Ancestor of Males Versus Females. *Science* **341**: 562–565.

Raff JA, Bolnick DA, Tackney J, O'Rourke DH. 2011. Ancient DNA perspectives on American colonization and population history. *Am J Phys Anthropol* **146**: 503–514.

Raghavan M, Skoglund P, Graf KE, Metspalu M, Albrechtsen A, Moltke I, Rasmussen S, Stafford Jr TW, Orlando L, Metspalu E, et al. 2014. Upper Palaeolithic Siberian genome reveals dual ancestry of Native Americans. *Nature* **505**: 87–91.

Raghavan M, Steinrücken M, Harris K, Schiffels S, Rasmussen S, DeGiorgio M, Albrechtsen A, Valdiosera C, Ávila-Arcos MC, Malaspinas A-S, et al. 2015. Genomic evidence for the Pleistocene and recent population history of Native Americans. *Science* **349**: aab3884.

Rasmussen M, Anzick SL, Waters MR, Skoglund P, DeGiorgio M, Stafford TW, Rasmussen S, Moltke I, Albrechtsen A, Doyle SM, et al. 2014. The genome of a Late Pleistocene human from a Clovis burial site in western Montana. *Nature* **506**: 225–229.

Rasmussen M, Li Y, Lindgreen S, Pedersen JS, Albrechtsen A, Moltke I, Metspalu M, Metspalu E, Kivisild T, Gupta R, et al. 2010. Ancient human genome sequence of an extinct Palaeo-Eskimo. *Nature* **463**: 757–762.

Rasmussen M, Sikora M, Albrechtsen A, Korneliussen TS, Moreno-Mayar JV, Poznik GD, Zollikofer CPE, Ponce De León MS, Allentoft ME, Moltke I, et al. 2015. The ancestry and affiliations of Kennewick Man. *Nature* **523**: 455–458.

Resque R, Gusmão L, Geppert M, Roewer L, Palha T, Alvarez L, Ribeiro-dos-Santos Â, Santos S. 2016. Male Lineages in Brazil: Intercontinental Admixture and Stratification of the European Background ed. F. Calafell. *PLoS ONE* **11**: e0152573.

Ribeiro-Dos-Santos AKC, Santos SEB, Luciamach A. Heterogeneity of Mitochondrial DNA Haplotypes in Pre-Columbian Natives of the Amazon Region.

Rieux A, Eriksson A, Li M, Sobkowiak B, Weinert LA, Warmuth V, Ruiz-Linares A, Manica A, Balloux F. 2014. Improved Calibration of the Human Mitochondrial Clock Using Ancient Genomes. *Molecular Biology and Evolution* **31**: 2780–2792.

Roca-Rada X, Politis G, Messineo PG, Scheifler N, Scabuzzo C, González M, Harkins KM, Reich D, Souilmi Y, Teixeira JC, et al. 2021. Ancient mitochondrial

genomes from the Argentinian Pampas inform the early peopling of the Southern Cone of South America. *iScience* **24**: 102553.

Roewer L, Nothnagel M, Gusmão L, Gomes V, González M, Corach D, Sala A, Alechine E, Palha T, Santos N, et al. 2013. Continent-Wide Decoupling of Y-Chromosomal Genetic Variation from Language and Geography in Native South Americans ed. S.M. Williams. *PLoS Genet* **9**: e1003460.

Rothhammer F, Fehren-Schmitz L, Puddu G, Capriles J. 2017. Mitochondrial DNA haplogroup variation of contemporary mixed South Americans reveals prehistoric displacements linked to archaeologically-derived culture history. *American J Hum Biol* **29**: e23029.

Rowe TB, Stafford TW, Fisher DC, Enghild JJ, Quigg JM, Ketcham RA, Sagebiel JC, Hanna R, Colbert MW. 2022. Human Occupation of the North American Colorado Plateau ~37,000 Years Ago. *Front Ecol Evol* **10**: 903795.

Saillard J, Forster P, Lynnerup N, Bandelt H-J, Nørby S. 2000. mtDNA Variation among Greenland Eskimos: The Edge of the Beringian Expansion. *The American Journal of Human Genetics* **67**: 718–726.

Scheib CL, Li H, Desai T, Link V, Kendall C, Dewar G, Griffith PW, Mörseburg A, Johnson JR, Potter A, et al. 2018. Ancient human parallel lineages within North America contributed to a coastal expansion. *Science* **360**: 1024–1027.

Schurr TG. 2004. The Peopling of the New World: Perspectives from Molecular Anthropology. *Annu Rev Anthropol* **33**: 551–583.

Simão F, Ribeiro J, Vullo C, Catelli L, Gomes V, Xavier C, Huber G, Bodner M, Quiroz A, Ferreira AP, et al. 2021. The Ancestry of Eastern Paraguay: A Typical South American Profile with a Unique Pattern of Admixture. *Genes* **12**: 1788.

Skoglund P, Mallick S, Bortolini MC, Chennagiri N, Hünemeier T, Petzl-Erler ML, Salzano FM, Patterson N, Reich D. 2015. Genetic evidence for two founding populations of the Americas. *Nature* **525**: 104–108.

Starikovskaya EB, Sukernik RI, Derbeneva OA, Volodko NV, Ruiz-Pesini E, Torroni A, Brown MD, Lott MT, Hosseini SH, Huoponen K, et al. 2005. Mitochondrial DNA Diversity in Indigenous Populations of the Southern Extent of Siberia, and the Origins of Native American Haplogroups. *Annals of Human Genetics* **69**: 67–89.

Starikovskaya YB, Sukernik RI, Schurr TG, Kogelnik AM, Wallace DC. 1998. mtDNA Diversity in Chukchi and Siberian Eskimos: Implications for the Genetic History of Ancient Beringia and the Peopling of the New World. *The American Journal of Human Genetics* **63**: 1473–1491.

Sun J, Ma P, Cheng H, Wang C, Li Y, Cui Y, Yao H, Wen S, Wei L. 2021. Post-last glacial maximum expansion of Y-chromosome haplogroup C2A-L1373 in

northern Asia and its implications for the origin of Native Americans. *American J Phys Anthropol* **174**: 363–374.

Taboada-Echalar P, Álvarez-Iglesias V, Heinz T, Vidal-Bralo L, Gómez-Carballa A, Catelli L, Pardo-Seco J, Pastoriza A, Carracedo Á, Torres-Balanza A, et al. 2013. The Genetic Legacy of the Pre-Colonial Period in Contemporary Bolivians ed. C. Lalueza-Fox. *PLoS ONE* **8**: e58980.

Tackney JC, Potter BA, Raff J, Powers M, Watkins WS, Warner D, Reuther JD, Irish JD, O'Rourke DH. 2015. Two contemporaneous mitogenomes from terminal Pleistocene burials in eastern Beringia. *Proc Natl Acad Sci USA* **112**: 13833–13838.

Tamm E, Kivisild T, Reidla M, Metspalu M, Smith DG, Mulligan CJ, Bravi CM, Rickards O, Martinez-Labarga C, Khusnutdinova EK, et al. 2007. Beringian Standstill and Spread of Native American Founders ed. D. Carter. *PLoS ONE* **2**: e829.

The Genographic Consortium, Jota MS, Lacerda DR, Sandoval JR, Vieira PPR, Ohasi D, Santos-Júnior JE, Acosta O, Cuellar C, Revollo S, et al. 2016. New native South American Y chromosome lineages. *J Hum Genet* **61**: 593–603.

Torroni A, Neel JV, Barrantes R, Schurr TG, Wallace DC. 1994. Mitochondrial DNA "clock" for the Amerinds and its implications for timing their entry into North America. *Proc Natl Acad Sci USA* **91**: 1158–1162.

University of Geneva, Switzerland, Ray N, Adams JM. 2001. A GIS-based Vegetation Map of the World at the Last Glacial Maximum (25,000-15,000 BP). *IA*. http://intarch.ac.uk/journal/issue11/rayadams_index.html (Accessed July 29, 2025).

Villaescusa P, Seidel M, Nothnagel M, Pinotti T, González-Andrade F, Alvarez-Gila O, M. De Pancorbo M, Roewer L. 2021. A Y-chromosomal survey of Ecuador's multi-ethnic population reveals new insights into the tri-partite population structure and supports an early Holocene age of the rare Native American founder lineage C3-MPB373. *Forensic Science International: Genetics* **51**: 102427.

Wang W, Ding M, Gardner JD, Wang Y, Miao B, Guo W, Wu X, Ruan Q, Yu J, Hu X, et al. 2021. Ancient Xinjiang mitogenomes reveal intense admixture with high genetic diversity. *Sci Adv* **7**: eabd6690.

Wei L-H, Wang L-X, Wen S-Q, Yan S, Canada R, Gurianov V, Huang Y-Z, Mallick S, Biondo A, O'Leary A, et al. 2018. Paternal origin of Paleo-Indians in Siberia: insights from Y-chromosome sequences. *Eur J Hum Genet* **26**: 1687–1696.

Weissensteiner H, Pacher D, Kloss-Brandstätter A, Forer L, Specht G, Bandelt H-J, Kronenberg F, Salas A, Schönherr S. 2016. HaploGrep 2: mitochondrial haplogroup classification in the era of high-throughput sequencing. *Nucleic Acids Res* **44**: W58–W63.

Willerslev E, Meltzer DJ. 2021. Peopling of the Americas as inferred from ancient genomics. *Nature* **594**: 356–364.

Yang MA, Gao X, Theunert C, Tong H, Aximu-Petri A, Nickel B, Slatkin M, Meyer M, Pääbo S, Kelso J, et al. 2017. 40,000-Year-Old Individual from Asia Provides Insight into Early Population Structure in Eurasia. *Current Biology* **27**: 3202-3208.e9.