FusionProt: Fusing Sequence and Structural Information for Unified Protein Representation Learning

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ABSTRACT

Proteins play a crucial role in biological processes and understanding the complex mechanisms within living organisms. Accurate protein representation is vital for advancements in drug development, synthetic biology, and understanding disease mechanisms. Current methodologies primarily focus on one-dimensional (1D) structures, emphasizing amino acid sequences. However, this approach neglects the essential three-dimensional (3D) structural details that determine protein functionality and interactions. Initial attempts to leverage both the 1D and 3D modalities have shown limited benefits compared to single modality representations. In this work, we introduce FusionProt, a novel protein representation learning framework designed to learn unified representations of both the 1D and 3D structures of proteins simultaneously. FusionProt incorporates an innovative learnable fusion token that serves as an adaptive bridge, enabling an iterative exchange of information between a protein language model and the protein's 3D structure graph. This token is integrated into the training process of both modalities, enabling seamless propagation of information and facilitating comprehensive representation through iterative learning cycles. Empirical evaluations across various biological tasks demonstrate that FusionProt achieves state-of-the-art performance, offering a significant advancement in protein representation learning.

1 INTRODUCTION

Proteins are essential for biological processes and for understanding complex mechanisms in living organisms. They are comprised of linear chains of amino acids that fold into a specific three-dimensional (3D) structure, which underscores their functional diversity and dynamic behaviors [58]. An effective understanding of proteins is essential for understanding disease mechanisms and synthetic biology and for advancing drug development [23].

Current methodologies for protein representation primarily emphasize the exploration of proteins' one-dimensional (1D) structures, specifically the relationships between amino acids. These approaches, such as ProteinBERT [1] and ESM [22, 33], often utilize text-based techniques and transformer architectures [49], which are trained on extensive protein sequence datasets. These models take an amino acid sequence as input and typically produce protein representations by averaging the representations of individual amino acids. However, this narrow focus on amino acid sequences neglects crucial protein structure details, thereby limiting the effectiveness of these methods.

The intricate 3D structure of proteins is crucial, as the conformation plays a pivotal role in determining their activities [57]. Proteins possess specific active sites for interactions with other molecules, which are defined by the 3D arrangement of amino acids. This 3D structure determines the specificity and affinity of binding interactions, aspects that a 1D representation cannot adequately capture [15, 55]. Furthermore, drug design relies on an understanding of 3D structures to identify binding sites and find molecules that modulate protein function [24, 26]. One of the state-of-the-art (SOTA) techniques today for 3D protein representation is GearNet [58]. This approach converts the 3D structure of a protein into a graph that captures its biological characteristics. Subsequently, graph neural network techniques [16, 37] are applied to this graph, facilitating the creation of comprehensive protein representations.

Recent research emphasizes the importance of comprehensive protein representation that includes both 1D and 3D structures to capture the protein's functional and interactional properties accurately. ESM-GearNet [57] was one of the first approaches to integrate these modalities. Although the study explored various fusion strategies, empirical results showed that the most effective method is using a large protein language model (PLM) such as ESM [22] to generate representations, which were then used as context for a graph encoder like GearNet [58]. Other approaches, such as SaProt [41], leverage an AlphaFold-based model [46] to reduce the 3D structure to tokens and train them along with amino acid tokens using a traditional PLM. However, these approaches are limited as they reduce one modality into context for another model, which potentially leads to the loss of critical structure information.

In this study, we introduce FusionProt (See Figure 1), a novel approach designed to learn a unified representation of the 1D and 3D structures of proteins simultaneously. Despite the vast number of proteins in nature, the number of known 3D structures remains limited [48]. To address this, we leverage an AlphaFold model [14] for accurate protein structure predictions. We introduce an innovative learnable fusion token that serves as an adaptive bridge, enabling an iterative exchange of information between a PLM and the protein's 3D structure graph. This token is integrated into the training process of both modalities, enabling seamless propagation of information and facilitating comprehensive representation through iterative learning cycles. In practice, this token is concatenated to the sequence, allowing attention mechanisms to query the unique fusion token alongside the amino acids. This process extracts and integrates valuable information, enhancing the learning of amino acid representations. Then, the fusion token is incorporated as an additional node in the graph representing the protein's 3D structure, connected to all nodes. A graph encoder (i.e., a structure model) processes this graph, generating a new representation for the fusion token, which is subsequently used in the PLM training over the amino acids. Through this iterative process, the model representations are combined to form a refined protein representation.

Unlike prior models such as ESM-GearNet [57], which perform static or one-shot fusion, such as simple concatenation after independent encoding, FusionProt introduces a dynamic mechanism where the fusion token continuously evolves through repeated interaction across layers. This design allows sequence and structure modalities to co-adapt throughout training, resulting in richer, functionally informed representations that better capture the complexity of protein behavior.

We perform an empirical evaluation over several protein tasks, spanning a broad spectrum of biological domains, comparing numerous methods of protein representations and achieving SOTA performance across various benchmarks, with statistical significance improvements. We present ablation tests to better study the performance of the algorithm.

The contributions of this study are threefold: (1) We introduce FusionProt, a novel approach that learns a unified representation for both the 1D and 3D structures of proteins simultaneously. Our main focus is on the fusion of 1D and 3D models in an effective manner. Our method enhances the accuracy of capturing functional and interactional properties of proteins, addressing limitations of previous methods that treated these structures separately; (2) We propose a novel fusion architecture that utilizes a specialized learnable fusion token, enabling an iterative exchange of information between a PLM and the protein's 3D structure graph. This token is integrated into the training process of both modalities, enabling seamless propagation of information and facilitating comprehensive representation through iterative learning cycles. The iterative process facilitates the exchange of contextually relevant structure and sequential features, improving the model's ability to capture both 1D and 3D protein characteristics; (3) We conduct an empirical evaluation of our work over several protein tasks establishing SOTA results and presenting a biological case study that further demonstrates the model's strengths. Also, we contribute our code to the community [1].

2 RELATED WORK

2.1 Sequence-based Representation Learning

Proteins are comprised of sequences of amino acids, that establish a natural analogy to tokens in natural language processing. Recently, the adoption of unsupervised deep learning techniques has become prevalent in modeling protein sequence data.

The advent of Transformers [49] has led to the development of numerous PLMs such as MSA Transformer [32], ProteinBERT [1], ProteinLM [54], ProtBERT-BFD [5], ProtTrans [6], ESM-1b [33], and ESM-2 [22] which is considered as the SOTA PLM. These models were trained on data from UniRef [42] which contains hundreds of billions of protein sequences, via masked language modeling [3].

A protein's biological function hinges on its 3D native structure [4]. However, many PLMs do not explicitly encode protein 3D structures, which are pivotal in understanding protein functions.

In this work, we aim to overcome this limitation by enhancing a PLM through novel fusion algorithms that integrate protein 3D structure models into the protein embedding. This approach seeks to capture both sequential and structure attributes of proteins, thereby advancing the capabilities of existing PLMs in biological research and applications.

2.2 Structure-based Representation Learning

The rising success of AlphaFold [14, 38] in predicting the 3D structure of proteins has led to deeper insights into their functional roles. Moreover, the release of more than 200 million protein structures in AlphaFoldDB [47, 48] has significantly advanced the development of large-scale protein structure models [45].

Protein structures are commonly represented as graphs, where amino acids serve as the nodes. Therefore, utilizing protein structure models on these graphs is a common practice. Models such as GVP [13], CDConv [7] and GearNet [58] have shown promising results, with different learning techniques. GearNet, which incorporates a Multiview Contrastive pre-training algorithm, is considered SOTA [58] and has outperformed the IEConv model [11], which proposed to apply a learnable kernel function on edge features. Furthermore, CDConv [7] has outperformed HoloProt [39] and ProNet [50] in a recent study [25].

Foldseek [46] suggested a different approach, optimized for protein structural search, which utilizes a VQ-VAE [44] to encode protein structures into informative tokens. Then, SaProt [41] integrates residue and structure tokens during training, derived from encoding 3D protein structures using Foldseek.

In this study, we enhance the structure-based representation learning approach by integrating sequential information, thus capturing both sequential and structure attributes of proteins.

2.3 Joint Representation Learning

The integration of protein sequence-based models with protein structure models has gained popularity in recent years [17–19, 31, 53]. Early efforts, such as LM-GVP [51] or MIF-ST [56], aimed to combine PLMs with Graph Neural Networks (GNNs) [36]. More recently, ESM-GearNet [57] proposed to incorporated sequential information into distinct residue-level models such as GearNet, GVP, and CDConv [57]. SaProt-GearNet [41] also integrates GearNet and achieves similar results. However, these approaches are limited by reducing one modality into context for another model, potentially losing critical structure information.

Unlike earlier approaches, we introduce a novel fusion architecture that simultaneously learns a unified representation for both the 1D and 3D structures of proteins. To the best of our knowledge, no existing PLMs are based on an iterative fusion of structure and sequential models simultaneously.

3 METHODS

We introduce the **FusionProt** model (See Figure 1). Given a protein $p = (S, R)$, which combines its amino acid sequence S and its 3D structure R, FusionProt simultaneously learns a unified representation for both the 1D and 3D structures of the protein. We propose a specialized learnable fusion token designed to enable an iterative exchange of information between a PLM and the protein's 3D structure graph.

In this section, we formally define sequential (1D) and structure (3D) protein layers, which are used in the model as part of the protein representation learning, and present the FusionProt

[1] https://github.com/kalifadan/FusionProt

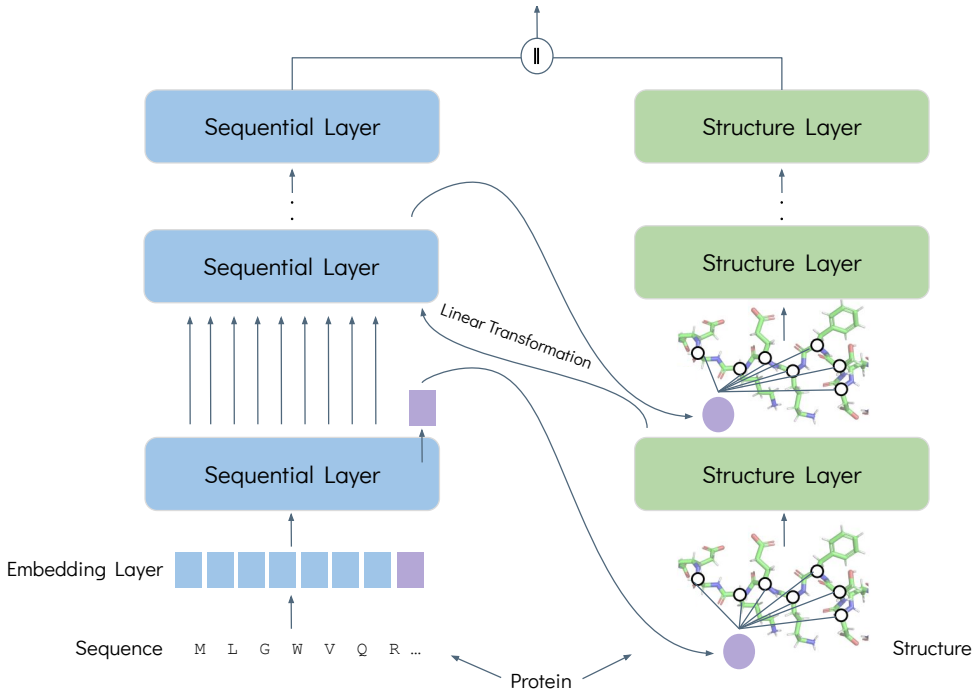

Figure 1: The FusionProt pre-training architecture. The model is trained on amino acid sequences and their corresponding 3D structures, utilizing the SOTA AlphaFold2 [14] for accurate protein structure predictions. We introduce an innovative learnable fusion token that serves as an adaptive bridge, enabling an iterative exchange of information between a PLM and the protein's 3D structure graph. This token is concatenated to the protein sequence, allowing attention mechanisms to query the unique fusion token alongside the amino acids. Then, the fusion token is incorporated as an additional node in the graph representing the protein's 3D structure, connected to all nodes. A graph encoder (i.e., a structure model) processes this graph, generating a new representation for the fusion token. This representation is subsequently used in the PLM's sequential layers. A learnable linear transformation is applied between each sequential and structure layer pair to align and adapt their distinct modality spaces. Through this iterative process, the model representations are combined to form a refined protein representation.

algorithm. Information from both layers is fused simultaneously during learning throughout the proposed specialized fusion token.

3.1 Protein Sequential Layer

3.1.1 Protein Sequence. The simplest level of protein structure, known as the primary structure, is a 1D structure, consisting of a linear sequence of amino acids, referred to as residues. The protein sequence exhibits similarities with natural language sequences, making the application of language models a common practice in this domain [1, 54].

Given a protein sequence $S = [s_1, s_2, ..., s_n]$, where n denotes the number of residues, the sequential model aims to capture the essential features of the protein sequence and outputs a protein representation denoted by $z = [z_1, z_2, ..., z_n] \in \mathbb{R}^{n \times D}$, where D is the embedding dimension.

3.1.2 Sequential Layer Definition. We utilize ESM-2 [22] as our sequential model, leveraging its superior performances as a PLM. Alternatively, other advanced sequential models (i.e., PLMs) can be used.

Within the context of a given sequential layer denoted by l, we denote $z^{(l)}$ as the output representation of this layer, initialized with $z_i^{(0)} = \text{Embedding}(s_i) \in \mathbb{R}^D$, where D denotes the embedding representation dimension. The layer performs the following update:

$$z^{(l)} = Attention\left(z_1^{(l-1)}, z_2^{(l-1)}, ..., z_n^{(l-1)}\right)$$

where n denotes the number of residues in the protein, and *Attention* is a multi-head self-attention layer [49].

3.2 Protein Structure Layer

3.2.1 Protein 3D Structure. A protein 3D structure is uniquely determined by its primary structure (amino acid sequences) [4]. There are only 20 standard residue (amino acid) types, each containing multiple components connected to a central carbon atom known as alpha carbon. Following GearNet [58], we use only alpha carbons to represent the main backbone structure of each protein. Therefore, we define a protein 3D structure as $R = [r_1, r_2 ... r_n] \in \mathbb{R}^{n \times 3}$, where r_i represents the Cartesian coordinates of the $i - th$ alpha carbon atom of each amino acid, and n denotes the number of residues, which is the sequence length.

3.2.2 Protein Structure Graph. We represent proteins using a multi-relational residue graph $G = (V, E, T)$, where V is the set of residues,

E is the set of edges, and T is the edge types. The set of edges E consists of three directed edge types, namely sequential edges, radius edges, and KNN edges:

$$E_{\text{seq}} = \{(i,j) \mid i,j \in V, |j-i| < d_{\text{seq}}\}$$
$$E_{\text{radius}} = \{(i,j) \mid i,j \in V, |r_j - r_i| < d_{\text{radius}}\}$$
$$E_{\text{KNN}} = \{(i,j) \mid i,j \in V, j \in \text{KNN}(i)\}$$
$$E = E_{\text{seq}} \cup E_{\text{radius}} \cup E_{\text{KNN}}$$

where $d_{\text{seq}} = 3$ defines the sequential distance threshold, $d_{\text{radius}} = 10\text{Å} = 0.1$ [nm] defines the spatial distance threshold, and $\text{KNN}(i)$ indicates the K-nearest neighbors [30] of node i with $k = 10$ (all parameters were set as reported in GearNet [58]).

3.2.3 Structure Layer Definition. We utilize GearNet [58] as the structure model, leveraging its contextual understanding of protein structures. Alternatively, other advanced structure models can be used in the algorithm (See Section 6.2).

Given a protein's 3D structure, R, we construct its corresponding protein structure graph $G = (V, E, T)$ (See Section 3.2.2). Then, the structure layer employs a relational message passing procedure, based on a relational graph convolutional neural network [37].

Within the context of a given structure layer denoted by l, we denote $u^{(l)}$ as the output representations of this layer, initialized with $u_i^{(0)} = \text{Embedding}(v_i) \in \mathbb{R}^D$, where $v_i \in V$, and D denotes the embedding representation dimension. The layer performs the following update:

$$u_i^{(l)} = u_i^{(l-1)} + \sigma\left(\sum_{t \in T} W_t \sum_{j \in \mathcal{N}_t(i)} u_j^{(l-1)}\right)$$

where $\mathcal{N}_t(i)$ is the set of neighbors of i with edge type t, $\sigma(\cdot)$ is a ReLU activation function, and the weight matrix W_t is learned per edge type t. This approach allows the model to incorporate various types of relational information, thereby enhancing its ability to learn comprehensive protein representations.

3.3 FusionProt

3.3.1 Fusion Design. We introduce a specialized learnable fusion token, designed to enable an iterative exchange of information between a PLM and the protein's 3D structure graph. This fusion token serves as a dynamic bridge, enabling iterative information exchange between the two modalities. During training (See Figure 1), the specialized fusion token, holds information from both models, the sequential and the structure. In practice, the amino acids query the associated unique fusion token to extract and integrate valuable information for learning amino acid representations via attention mechanisms. Additionally, the fusion token is incorporated as an additional node in the graph representing the protein's 3D structure. We connect it to all nodes, with a new edge type, thus enabling the structure model to learn important features from the sequential model. Hence, the token enables a novel fusion between both models, leading to an enhanced protein representation.

The choice to connect the fusion token to all nodes in the 3D graph is informed by principles from GNNs [36], where nodes exchange information to capture global and local dependencies. The fusion token is not merely an addition to the sequence or structure; rather, it mediates the dynamic interaction between the two modalities, ensuring iterative refinement of the protein representation. This approach contrasts with previous methods that simply concatenate or independently process the sequence and structure information. This novel fusion mechanism leverages multi-modal learning and graph-based information propagation. By iteratively combining the 1D protein sequence and 3D structure features, FusionProt captures both local and global dependencies, enabling the model to learn richer and more holistic representations of proteins.

Algorithm 1 FusionProt Algorithm

Input: Protein $p = (S, R)$ with sequence $S = [s_1, ..., s_n]$, and 3D structure $R = [r_1, ..., r_n] \in \mathbb{R}^{n\times 3}$, Sequential encoder with L_1 layers, Structure encoder L_2 layers.
Output: Unified protein representation h

Create a learnable fusion token $s_{n+1} = f$
Augment sequence: $S' = [s_1, ..., s_n, s_{n+1}]$
Initialize sequential embeddings: $z_i^{(0)} = \text{Embedding}(s_i)\ \forall i \in [1, ..., n+1]$
Construct graph $G = (V, E, T)$ from R
Add node v_{n+1} to G, connect $v_{n+1} \leftrightarrow v_i$ with edge type $t_f\ \forall i \in [1, ..., n]$
Set $u_i^{(0)} = \text{Embedding}(v_i)$ for all $v_i \in V$
for $l = 1$ to L_2 **do**
  for $j = 1$ to $L_1 \div L_2$ **do**
    $l_l = j + (l-1) \cdot (L_1 \div L_2)$
    $z^{(l_j)} = Attention(z_1^{(l_j-1)}, ..., z_n^{(l_j-1)}, z_{n+1}^{(l_j-1)})$
  end for
  $u_{n+1}^{(l-1)} = \text{Linear}_l(z_{n+1}^{(l_j)})$
  Update node v_{n+1} in graph G with $u_{n+1}^{(l-1)}$
  $u_i^{(l)} = u_i^{(l-1)} + \sigma\left(\sum_{t\in T} W_t \sum_{j\in\mathcal{N}_t(i)} u_j^{(l-1)}\right)\ \forall u_i \in V$
  $z_{n+1}^{(l_j)} = \text{Linear}'_l(u_{n+1}^{(l)})$
end for
Concatenate final embeddings: $h = [z_{1:n}^{(L_1)}; u_{1:n}^{(L_2)}]$
return h

3.3.2 Fusion Algorithm. We provide a formal description of the FusionProt algorithm in Algorithm 1.

Given a protein $p = (S, R)$, which combines its amino acid sequence S and its 3D structure R, FusionProt concatenates the learnable fusion token to the sequence, resulting in a new sequence $S' = [s_1, s_2, ..., s_n, f]$ with a length of $n + 1$. Then, it is used to initialize the sequential embedding layer:

$$\forall s_i \in S' : z_i^{(0)} = \text{Embedding}(s_i)$$

Within the context of a given sequential layer denoted by l, the algorithm yields a sequence representation:

$$z^{(l)} = Attention\left(z_1^{(l-1)}, z_2^{(l-1)}, ..., z_n^{(l-1)}, z_{n+1}^{(l-1)}\right)$$

where $z_{n+1}^{(l-1)}$ is the intermediate representation of the fusion token. Then, the fusion token representation is passed into the corresponding structure layer l, represented as node $n + 1$, while connecting to all nodes in the 3D structure graph, yielding a structure representation:

$$u_i^{(l)} = u_i^{(l-1)} + \sigma\left(\sum_{t\in T'} W_t \sum_{j\in\mathcal{N}_t(i)} u_j^{(l-1)}\right)$$

where $u_{n+1}^{(l-1)} = \text{Linear}_l\left(z_{n+1}^{(l-1)}\right)$, $T' = T \cup \{t_f\}$, $\mathcal{N}_{t_f}(n+1) = V$, and $\forall i \in [1, ..., n] : \mathcal{N}_{t_f}(i) = \{n+1\}$, as we connected the fusion token to all residues in the protein structure graph with a new type of edge t_f (in both directions). To bridge the gap between the sequential and structure representation space, we utilize an affine transformation Linear_l (a linear layer) per each structure layer l to project these representations into the same space. Subsequently, we set the fusion token representation $u_{n+1}^{(l)}$ as the input for the following sequential layer $l+1$, resulting in:

$$z_{n+1}^{(l)} = \text{Linear}_{l'}\left(u_{n+1}^{(l)}\right)$$

where $\text{Linear}_{l'}$ is the affine transformation in the opposite direction (i.e., structure to sequential space or vice versa).

The protein representation undergoes transformations across the structure and sequential layers, where L_1 and L_2 are the number of sequential and structure layers, respectively, and $L_1 \geq L_2$ (note that L_1 need not be equal to L_2). Our aim is to uniformly integrate 3D structure information across the L_1 sequential and L_2 structure layers. Therefore, the fusion token is passed after $L_1 \div L_2$ sequential layers into the structure model. Then, the fusion token is passed back into the sequential model after every structure layer.

Lastly, the final hidden states of both the sequential and structure layers are concatenated to form the final protein output representation, which is:

$$h = \left[z^{(L_1)}, u^{(L_2)}\right]$$

3.4 Pre-training Objective

Following GearNet [58] and ESM-GearNet [57], we use Multiview Contrastive learning as our pre-training objective, for a fair comparison. Other pre-training algorithms could also be applied. We compare different pre-training algorithms in Section 6.3.

The Multiview Contrastive objective is to preserve the similarity between correlated protein subcomponents when mapped to a lower-dimensional latent space. Therefore, for a protein structure graph, we randomly select consecutive subsequences and apply random edge masking to hide 15% of edges in the protein graph, generating diverse views. Then, we align their representations in the latent space with an InfoNCE loss [43].

4 EMPIRICAL EVALUATION

In this section, we outline our empirical setting. We performed analysis on several downstream tasks, demonstrating the value of the representation we provide in real-world bioinformatics tasks.

4.1 Protein Structure Dataset

To reduce reliance on the availability and quality of external structural data, we leverage AlphaFold2 [14], a well-known SOTA model, which enables us to generate 3D structures and eliminate the need for external sources. Furthermore, many recent high-performing methods, such as SaProt [41] and ESM-GearNet [57], also used it for structure generation, ensuring consistency in comparisons. Therefore, we use the AlphaFold protein structure database [47, 48] for pre-training, which contains 805K protein structures predicted by AlphaFold2 [14]. Alternatively, other models, such as ESMFold [21], can be applied. Since AlphaFold predictions may include local inaccuracies, we assess FusionProt's robustness to structure noise in Section 6.4.

4.2 Tasks

We evaluated our proposed method on several common downstream tasks, selected based on SOTA works of GearNet [57] and SaProt [41], to assess its effectiveness for protein representation learning. These tasks include protein annotations such as Enzyme Commission (EC) prediction, Gene Ontology (GO) prediction, and Mutation Stability Prediction. The GO prediction task includes three sub-tasks: predicting a protein's biological processes (BP), molecular functions (MF), and cellular components (CC).

To ensure the validity of our results, we validate all downstream task splits by ensuring that sequences in one set share no more than 30% Needleman–Wunsch sequence identity with any sequence in the other sets.

4.2.1 Enzyme Commission Number Prediction. Annotation of enzyme function has a wide range of real-world applications, including metagenomics, diagnosis of enzyme-deficiency-related diseases [20], and cellular metabolism [34]. This task focuses on determining enzyme function by predicting EC numbers, which characterize a protein's catalytic activity in biochemical reactions. It involves 538 binary classification problems derived from the third and fourth levels of the EC classification tree [52]. We used dataset splits from DeepFRI [9], and the evaluation metric is the $F_{\max}$ score.

4.2.2 Gene Ontology Term Prediction. The GO knowledgebase [10] provides a set of structured and controlled terms describing gene products and their molecular properties. Many real-world biological applications, such as predictions of protein-protein interactions, rely on GO term-protein annotations [12, 29]. This benchmark includes three tasks: predicting a protein's biological processes, molecular functions, and cellular components. Each task involves multiple binary classification problems based on GO term annotations. We used dataset splits from DeepFRI [9], and the evaluation metric is the $F_{\max}$ score.

4.2.3 Mutation Stability Prediction. Mutation stability prediction (MSP) is crucial in bioinformatics for understanding how genetic mutations affect protein stability, which plays a key role in disease mechanisms and drug development. This task aims to predict whether a mutation enhances a protein complex's stability. We utilize the datasets and hyperparameters from ESM-GearNet [57]. Evaluation is based on AUROC.

4.3 Baselines

We compare with numerous baseline models, including PLMs and structure models. We used ProtBERT-BFD [5], DeepFRI [9], ESM-1b [33], and ESM-2 [22] as SOTA PLM sequential models. GearNet [58], GVP [13], and CDConv [7] are used as SOTA structure models. Furthermore, we include MIF-ST [56], ESM-GearNet [57], SaProt [41] and SaProt-GearNet [41] as SOTA in joint learning of structure models with sequential models. For consistency, when using ESM-2 we used the ESM-2-650M variant [22]. In addition,

Table 1: Evaluation results on EC and GO prediction under various pre-trained baseline models. "PLM" and "Structure Info." indicate the usage of protein language models and structure information in the model, respectively. The $F_{\max}$ score is the evaluation metric. Statistically significant results with $p < 0.05$ using a t-test are marked with an asterisk (*). For FusionProt, we report the mean and standard deviation over 3 independent fine-tuning runs (with different seeds). The best result is highlighted in bold.

Method	PLM	Structure Info.	EC	GO-BP	GO-MF	GO-CC
			$F_{\max}$	$F_{\max}$	$F_{\max}$	$F_{\max}$
ProtBERT-BFD [5]	✓	✗	0.838	0.279	0.456	0.408
DeepFRI [9]	✓	✗	0.631	0.399	0.465	0.460
ESM-1b [33]	✓	✗	0.859	0.320	0.661	0.392
ESM-2 [22]	✓	✗	0.877	0.345	0.668	0.411
GVP [13]	✗	✓	0.886	0.495	0.672	0.420
CDConv [7]	✗	✓	0.820	0.453	0.654	0.479
GearNet [58]	✗	✓	0.871	0.481	0.650	0.476
MIF-ST [56]	✓	✓	0.803	0.239	0.627	0.322
ESM-GearNet [57]	✓	✓	0.886	0.512	0.670	0.495
SaProt [41]	✓	✓	0.884	0.486	0.678	0.479
SaProt-GearNet [41]	✓	✓	0.886	0.512	0.672	0.504
FusionProt	✓	✓	$\mathbf{0.904}^{*}\pm0.002$	$\mathbf{0.524}^{*}\pm0.004$	$\mathbf{0.689}^{*}\pm0.003$	$\mathbf{0.518}^{*}\pm0.003$

GearNet [58], ESM-GearNet [57], and FusionProt are pre-trained with the same objective, which is the Multiview Contrast [58] objective, as prior work showed its superior performance.

4.4 Implementation Details

4.4.1 Pre-Training Phase. Following ESM-GearNet [57], we adopt exactly the same hyperparameters, training objectives, and model selection strategies. Specifically, we choose a pre-trained ESM-2-650M [22] as the base PLM, with 33 layers (i.e., $L_1 = 33$), and GearNet [58] with 6 layers ($L_2 = 6$) and 512 hidden dimensions as the structure model. The embedding dimension D is set to 1280. In addition, we apply the Multiview Contrast [58] as the pre-training algorithm. During pre-training, FusionProt is trained for 50 epochs using a learning rate of 2e-4, and a global batch size of 256 proteins. To accommodate long protein sequences, inputs are truncated to a maximum of 1,024 tokens. Both models are implemented using the TorchDrug library.

4.4.2 Fine-Tuning Phase. During inference, we incorporate task-specific classification heads to enable predictions for these tasks. Furthermore, following the latest SOTA model, SaProt [41], we conducted evaluations of our model and all baselines using the same set of hyperparameters to ensure fair comparisons. Specifically, we used AdamW optimizer with parameters $\beta_1 = 0.9$ and $\beta_2 = 0.98$, and we applied a weight decay of $L_2 = 0.01$. We consistently used a batch size of 64 and set the learning rate to 2e-5.

4.4.3 Computational Complexity. FusionProt was trained using 4× NVIDIA A100 80GB GPUs over 48 hours, totaling 192 GPU hours. It retains the underlying architecture of the base sequence and structure encoders (e.g., ESM-2 and GearNet), applying each layer only once, similar to prior baselines such as ESM-GearNet [57]. The proposed fusion mechanism introduces only lightweight additions and arithmetic operations via a token exchange module between encoders, contributing a bounded $O(1)$ overhead per layer. At inference time, FusionProt incurs a marginal runtime increase of approximately 2–5% relative to SOTA baselines, with latency rising from approximately 0.012 seconds to 0.014 seconds per 1,000 residues. Consequently, the overall computational complexity of FusionProt remains comparable to existing methods in both theoretical and empirical terms.

5 EMPIRICAL RESULTS

In this section, we present our empirical results. We report the mean and standard deviation of FusionProt's performance over three independent fine-tuning runs (with different random seeds) to ensure reproducibility. Additionally, we validate the statistical significance of performance differences, using a two-tailed paired t-test with a 95% confidence level ($p < 0.05$), comparing observations from the tested models. Significant results are marked with an asterisk (*) in the tables.

5.1 Task 1: EC Number Prediction

In Table 1, we compared the performance of FusionProt with eleven baseline methods on the EC number prediction task. In particular, FusionProt significantly outperformed all baseline methods, achieving the highest $F_{\max}$ score.

PLMs such as ProtBERT-BFD [5] or ESM-2 [22], which rely solely on sequence data, produced substantially lower $F_{\max}$ scores, compared to FusionProt. This highlights the benefit of incorporating 3D structure information into representation learning.

Similarly, FusionProt, which leverages both structure and sequential information, performed better than SOTA structure models such as GearNet [58] or CDConv [7], which trained only on 3D structure

data. This indicates that while structure models are strong, they are not sufficient alone, as they require PLMs for optimal performance, probably due to their effective self-attention layers [49].

While models such as MIF-ST [56], ESM-GearNet [57], and SaProt-GearNet [41] also attempt to utilize both types of information, they reduce one modality into a context for another model, leading to a loss of critical structure information. In addition, the importance of the fusion technique is emphasized by the significant underperformance of MIF-ST compared to other fusion models. Similarly to ESM-GearNet, MIF-ST uses outputs from a pre-trained sequence-only PLM as input to a structure model.

By integrating 1D and 3D protein structure information synergistically, FusionProt attempts to capture subtle structure features that influence enzyme specificity and activity, which are crucial for EC prediction. This performance improvement is essential for real-world applications, such as the diagnosis of enzyme deficiency-related diseases [20].

5.2 Task 2: GO Term Prediction

Table 1 presents the results of the GO term prediction tasks compared to eleven baseline methods, including PLMs, structure models, and ensemble models. FusionProt showed strong performance across the board, achieving the highest $F_{\max}$ scores in all tasks, with a statistical significance.

The results demonstrate that sequential or structure information alone is insufficient, similarly to the EC prediction task (See Section 5.1). This indicates that the novel fusion of FusionProt is critical for achieving superior performance.

In the GO-BP task, which involves predicting a protein's role in biological processes, FusionProt achieves an $F_{\max}$ score of 0.524, significantly outperforming the next-best method. Biological processes often rely on long-range structural interactions and cooperative protein functions, which makes this task particularly sensitive to accurate 3D structure representations. The substantial performance gap between SaProt and SaProt-GearNet further emphasizes the critical role of structure in this setting, a factor that FusionProt leverages more effectively through early integration.

In the GO-CC task, which predicts the cellular component of proteins, FusionProt achieves an $F_{\max}$ of 0.518, outperforming all baselines including ESM-GearNet [57], probably due to the more direct connection between 3D structure characteristics and subcellular localization [8]. The prediction of cellular components is strongly based on the spatial organization of proteins within the cell, which is well represented by detailed 3D structures [40].

This reliance on structural information benefits models such as CDConv [7], although it exhibits weaker performance in EC prediction. In contrast, predicting molecular functions involves complex interactions and dynamic changes that are less directly captured by 3D structures [35], resulting in a comparatively smaller performance gap than in the other tasks.

5.3 Task 3: Mutation Stability Prediction

Table 2 reports the results for the MSP task, which evaluates a model's ability to assess the effect of amino acid substitutions on protein stability, a critical problem in protein engineering, variant effect prediction, and drug design.

Following the approach in ESM-GearNet [57], we demonstrate the robustness of our proposed method by comparing its performance with the SOTA method, specifically GVP [13] for the MSP task. Due to space limitations, we present a comparison to the SOTA and refer the reader to additional baselines in [57].

FusionProt achieves the highest AUROC among all evaluated methods, reaching 0.745 with statistical significance ($p < 0.05$), outperforming both structure-aware (GVP) and sequence-structure fused (ESM-GearNet) baselines. Notably, FusionProt improves upon ESM-GearNet by approximately 24.37%, and surpasses the current SOTA GVP by 5.1%. This highlights the effectiveness of our iterative fusion mechanism in preserving and integrating long-range sequence and structural dependencies, which are especially important for modeling stability changes that may arise from distal or context-dependent mutations.

Compared to GVP, which is limited to local spatial interactions, and ESM-GearNet, which performs shallow one-shot fusion, FusionProt allows multi-layer cross-modal refinement via a learnable token and bidirectional interaction path. This enables a more expressive and biologically grounded representation, particularly for capturing distributed compensatory effects in allosteric regions, commonly overlooked in residue-centric models.

Table 2: Evaluation results on MSP prediction across different models. "PLM" and "Structure" indicate the usage of protein language models and structure information in the model, respectively. The AUROC score is used as the evaluation metric. Statistically significant results ($p < 0.05$, t-test) are marked with an asterisk (*). For FusionProt, we report the mean and standard deviation over 3 independent fine-tuning runs (with different seeds). The best result is in bold.

Method	PLM	Structure	MSP (AUROC)
ESM-GearNet [57]	✓	✓	0.599
GVP [13] (SOTA)	✗	✓	0.709
FusionProt	✓	✓	$\mathbf{0.745}^{*}\pm0.008$

6 ABLATION AND ANALYSIS

We conduct a series of ablation and diagnostic studies to evaluate the key design decisions behind FusionProt. These include varying the fusion-injection frequency, comparing different structure encoders, and testing alternative pre-training objectives. We also assess the model's robustness to noise in predicted 3D structures. Finally, we present biological case studies that highlight FusionProt's practical utility in real-world tasks such as drug discovery and disease research.

6.1 Ablation on Fusion-Injection Frequency

We aimed to determine the optimal number of fusion injections in our FusionProt model, which fused information between the structure and sequential models using a specialized learnable fusion token. In our standard method, given a sequential model with L_1 layers and a structure model with L_2 layers, where $L_1 \geq L_2$, the fusion token is passed after $L_1 \div L_2$ sequential layers into the structure

model (See Section 3.3.2). Then, the fusion token is passed back into the sequential model after every structure layer. In this ablation, we experimented with different injection frequencies: half the regular number (every $2 \cdot (L_1 \div L_2)$ sequential layers and two structure layers) and one-third the regular number (every $3 \cdot (L_1 \div L_2)$ sequential layers and three structure layers). The results in Table 3 indicate that the standard fusion injection frequency consistently provides the best performance across tasks. Reducing fusion injections negatively impacted performance, highlighting the importance of the fusion token. By decreasing the frequency of injections, the model's ability to integrate these two types of information is compromised, leading to suboptimal predictions.

Table 3: Ablation test for the number of fusion injections in FusionProt. Statistically significant results with $p < 0.05$ using a t-test are marked with an asterisk (*). The best result is highlighted in bold.

Number of Fusion	EC	GO-BP	GO-MF	GO-CC
	$F_{\max}$	$F_{\max}$	$F_{\max}$	$F_{\max}$
Few (One-Third)	0.876	0.511	0.659	0.498
Medium (Half)	0.882	0.514	0.669	0.504
Full (Standard)	**0.904***	**0.524***	**0.689***	**0.518***

6.2 3D Structure Models Comparison

FusionProt is designed to be flexible and modular, allowing the structure encoder component to be replaced with alternative architectures. The ablation study for various SOTA 3D structure models is detailed in Table 4. We evaluated several structure models, including GearNet [58], GVP [13], and CDConv [7]. For each structure model, we applied our fusion technique and compared their performance with FusionProt, which utilizes the GearNet [58] model (See Section 3.2.3). The GVP model replaces standard MLPs [28] in GNN [36] layers with generalized vector perceptrons, which handle scalar and geometric features as vectors that adapt to spatial rotations. In contrast, CDConv utilizes GearNet's multi-type message passing to capture sequential and spatial interactions among residues. For consistency, we adopted the hyperparameters of ESM-GearNet [57]. Specifically, we selected 8 layers of CDConv [7] with an initial hidden dimension of 256, and 5 layers of GVP [13] with scalar features of 256 and vector features of 16 dimensions. Our results demonstrate that the FusionProt model, utilizing the GearNet structure, consistently achieves superior performance compared to other models. This aligns with previous studies [57], which have also established GearNet as a SOTA approach for joint representation learning.

6.3 Pre-training Algorithm Ablation

Table 5 compares FusionProt, trained with Multiview Contrastive learning (Section 3.4), with other pre-training algorithms. First, we tested self-prediction methods [58], which aim to predict one part of the protein given the remaining context. Specifically, we utilized the Residue Type Prediction method, a self-supervised task that performs masked prediction on individual residues. Next, we evaluated diffusion-based methods [59], inspired by the success of diffusion models in capturing the relationship between sequences and structures. During training, noise levels are added to structures and sequences, with higher noise levels indicating more distortion. We tested the SiamDiff objective, which refines structures via torsional adjustments and noise reduction. The results show that the Multiview Contrastive approach outperformed both Residue Type Prediction (using only sequence data) and SiamDiff (which integrates sequence and structure separately). Unlike methods that treat sequence and structure separately, Multiview Contrast integrates both, aligning subsequence representations from the same protein to capture interrelated sequences and structure patterns.

Table 4: Ablation test for various structure models used by FusionProt. For all models, the PLM is the ESM-2 [22]. Statistically significant results with $p < 0.05$ using a t-test are marked with an asterisk (*). The best result is highlighted in bold.

Method	EC	GO-BP	GO-MF	GO-CC
	$F_{\max}$	$F_{\max}$	$F_{\max}$	$F_{\max}$
FusionProt (GVP)	0.889	0.507	0.677	0.462
FusionProt (CDConv)	0.837	0.478	0.665	0.497
FusionProt (GearNet)	**0.904***	**0.524***	**0.689***	**0.518***

Table 5: Ablation test for the FusionProt model with different pre-training algorithms. Statistically significant results with $p < 0.05$ using a t-test are marked with an asterisk (*). The best result is highlighted in bold.

Method	EC	GO-BP	GO-MF	GO-CC
	$F_{\max}$	$F_{\max}$	$F_{\max}$	$F_{\max}$
Residue Type	0.890	0.524	0.673	0.506
SiamDiff	0.874	0.509	0.645	0.512
Multiview Contrast	**0.904***	**0.524***	**0.689***	**0.518***

6.4 Robustness to Noise in 3D Structures

Robustness in protein modeling often refers to a model's ability to tolerate minor perturbations [2]—an important property given that predicted structures (e.g., from AlphaFold2) may contain local inaccuracies, especially in flexible regions. To evaluate FusionProt's robustness to such noise, we injected Gaussian perturbations into the alpha-carbon backbone coordinates of AlphaFold2-predicted structures. The noise was zero-mean, with standard deviations ranging from 0.1 Å to 1.5 Å, simulating local structure deviations commonly observed in predicted or low-confidence regions. FusionProt maintained stable performance up to a noise level of 0.9 Å, with only a modest drop of 3–5 $F_{\max}$ points observed in the EC and GO-BP tasks beyond this threshold. These results demonstrate that

(a) (b)

Figure 2: 3D structure of proteins: (a) DNA-directed RNA polymerase subunit omega and (b) D-alanine–D-alanine ligase.

FusionProt is resilient to realistic structure noise and remains effective even when relying on imperfect 3D structures, as commonly encountered in high-throughput or large-scale prediction settings.

6.5 Biological Case Study

To demonstrate the capabilities of FusionProt, we conducted an in-depth analysis of its outputs, focusing on the quality of protein representations it generates. Our primary objective was to evaluate the model's ability to predict EC numbers, as outlined in Section 4.2.1, using pre-trained representations without any fine-tuning. This approach allowed us to assess the inherent quality of the learned representations. For example, consider the DNA-directed RNA polymerase subunit omega protein. See Figure 2a for an illustration of its 3D structure. Despite being identified early in RNA polymerase research, this protein's function remained unclear and controversial for a long time. Recently, it has been found to play a structural role, aiding in the recruitment of the largest subunit to the enzyme assembly [27]. Due to its complexity, this protein posed a challenge for existing protein models; those models, including ESM-2 [22], failed to predict its EC number. However, our FusionProt model predicts it successfully with high confidence (0.90). As another example, D-alanine–D-alanine ligase has a relatively simple 3D structure, as illustrated in Figure 2b. Both our model and ESM-2 successfully predict this protein's EC number; this is likely due to the straightforward nature of its structure. The ability of FusionProt to accurately predict the EC number of this protein highlights its effectiveness in capturing complex structure and functional details, underscoring its potential for advancing protein function prediction and real-world biological applications. The unique fusion of the learnable fusion token in the learning process enables the model to incorporate detailed structure and sequential information, which is crucial for understanding the functional aspects of proteins.

7 CONCLUSIONS

In this paper, we introduced FusionProt, a novel architecture designed to learn a unified representation of the 1D and 3D structures of proteins simultaneously. FusionProt incorporates a specialized learnable fusion token that enables an iterative exchange of information between a PLM and the protein's 3D structure graph, facilitating a more comprehensive representation through iterative learning cycles between a PLM and a structure model.

We propose a novel fusion algorithm that enables effective propagation of information between a PLM and a structure model. This fusion technique outperforms previous methods, which often convert one modality into context for another model, potentially leading to the loss of crucial structural information.

We evaluated our proposed method on several protein-level tasks to assess its effectiveness in protein representation learning. These tasks include protein annotations, which are essential for real-world medical applications. FusionProt significantly outperformed all baseline methods, including SOTA models, across all tasks, with statistically significant improvements.

We perform ablation tests to examine the contribution of our novel approach of learning unified representations of both 1D and 3D structures of proteins simultaneously. The tests confirm that our approach significantly enhances FusionProt's ability to capture the intricate relationships between protein sequences and their 3D structures, leading to improved performance across various real-world tasks.

Although the proposed fusion token mechanism is tailored for bidirectional interaction between sequence and structure modalities, it is conceptually extensible to more than two information channels. For example, additional modalities such as ligand descriptors, protein dynamics, or expression context could, in principle, be integrated into the framework by assigning separate fusion tokens or adopting a shared token passed through each encoder block in sequence. We leave the systematic evaluation of such multi-modal extensions to future work.

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