Convergent life-history evolution in *Hordeum*: Phylogenomic insights into climatic niche variation and functional genetic differentiation among annual and perennial wild relatives of barley

Hellwig, Timo[1,*]; Nina; Döring[1]; Haraldsson, Einar Baldvin[1]; von Korff, Maria[1,2,*]

[1] Institute of Plant Genetics, Heinrich-Heine-University, 40225 Düsseldorf, Germany

[2] Cluster of Excellence on Plant Sciences ''From Complex Traits towards Synthetic Modules'', 40225 Düsseldorf, Germany

* Corresponding authors: timo.hellwig@hhu.de, maria.korff.schmising@hhu.de

Abstract

Annual and perennial life-history strategies have evolved repeatedly across angiosperms, yet the genomic and environmental underpinnings of these transitions remain poorly understood, particularly in grasses. We generated de novo transcriptomes from 82 accessions representing 22 *Hordeum* species, including barley, and identified 257 single-copy orthologs present in all accessions to infer a robust phylogeny of the genus. By combining phylogenetic network inference with ABBA-BABA tests, we detected four cases of interspecific hybridization, three coinciding with major long-distance dispersal events across continents. Comparative climatic niche analysis indicated annual *Hordeum* species inhabit environments with higher temperatures, greater interannual variability, and increased human disturbance, compared to perennials, although no consistent precipitation differences were observed. Using our phylogeny as a framework, we analyzed selection and gene expression to uncover genomic changes associated with life-history strategy while accounting for phylogenetic non-independence. Additionally, we analysed gene copy number variations associated with life-history strategy. These analyses yielded 174 candidate genes across diverse biological functions, suggesting the genetic architecture underlying life-history evolution is more complex than assumed. Candidate genes grouped into six major functional categories, the most prominent being signal transduction and development, including regulators of flowering, dormancy, and meristem activity, metabolic and biosynthetic processes related to carbon allocation and storage, and stress response and defense, reflecting the resilience of perennials compared to the accelerated growth strategies of annuals. Our study reconstructs the evolutionary history and climatic niche differentiation of *Hordeum* species and demonstrates that convergent life-history evolution is driven by multifaceted, functionally diverse genetic mechanisms.

Keywords

life-history strategy, perenniality, annuality, comparative phylogenetics, phylotranscriptomics, *Hordeum*, barley, crop wild relatives

Introduction

Plants exhibit remarkable diversity in life history strategies to maximize survival and reproduction in specific environments. A fundamental distinction is between annual and perennial plants. Annuals complete their life cycle within one growing season and are semelparous (monocarpic), reproducing once before dying. Perennials live multiple years, ranging from biennials, with one vegetative year plus one reproductive year, to woody perennials surviving hundreds of years, reproducing annually (Friedman 2020). Although perenniality is often considered the evolutionary default, transitions between annual and perennial life histories occur repeatedly and rapidly across angiosperms. A strong evolutionary signal of the distribution of annual and perennial species in the tree of angiosperms indicate convergent evolution driven by common underlying factors, whether environmental, genetic, or both (Hjertaas et al. 2023).

Understanding the ecological and evolutionary implications of annual versus perennial life history strategies, including how they influence species distributions, community composition, and plant responses to environmental changes, is crucial for predicting ecosystem dynamics and informing conservation and restoration efforts (De Kort et al. 2021). Annuals tend to grow quickly and produce many seeds rapidly, an adaptive bet-hedging strategy for unpredictable or harsh environments (Garnier 1992; Stearns 1992). Many annuals are ruderal, colonizing disturbed habitats rapidly to avoid competition (Grime 1977). Perennials emphasize long-term survival and resource efficiency, investing in structural defenses and resource storage to withstand stresses and persist in stable environments (Chapin et al. 1990; Boege and Marquis 2005). In agriculture, perennials offer benefits such as permanent soil cover reducing erosion (Vallebona et al. 2016) and deeper roots that minimize nutrient leaching (Crews and Rumsey 2017; Lundgren and Des Marais 2020). The advantages of perennial crops are increasingly recognized in sustainable farming (Crews et al. 2018; Zhao and Wang 2024). About two-thirds of global calorie intake comes from annual crops, with five staples (wheat, rice, maize, soybean, barley) dominating (D'Odorico et al. 2014). Barley (*Hordeum vulgare* L.) is a versatile grain, ranking fourth in cultivated area (www.fao.org/faostat). The *Hordeum* genus, in the tribe Triticeae (Poaceae), originated ~10 million years ago in the Eastern Mediterranean and spread into Asia and the Americas (Blattner 2006). Most of ~30 *Hordeum* species radiated in South America (~18 species). Eurasian species relationships are well resolved, but some uncertainties remain regarding the New World phylogeny (Jörgensen 1986; Doebley et al. 1992; Svitashev et al. 1994; Marillia and Scoles 1996; Petersen and Seberg 1998; El-Rabey et al. 2002; Petersen and Seberg 2003; Blattner 2006; Jakob and Blattner 2006; Brassac and Blattner 2015; Jin et al. 2024; Mason-Gamer and White 2024; Feng et al.2025).

Hordeum includes both annual and perennial species, with at least three independent transitions between strategies (Brassac and Blattner 2015). Perennial *Hordeum* species are iteroparous, maintaining above-ground vegetation over winter and initiating growth under long days. A notable exception is Mediterranean perennial *H. bulbosum*, which dies above ground after seed set, surviving as bulbs underground that sprout in autumn rains (Koller and Highkin 1960; Fuerst et al. 2023). This 'pseudo-annual' strategy relies on below-ground organs for clonal propagation (Li et al. 2022). We focus here on the iteroparous perennials maintaining above-ground material. The barley secondary gene pool includes only *H. bulbosum*; other species are tertiary due to crossing barriers (Bothmer et al. 1983; Fedak 1985). Despite such crossing barriers, modern biotechnological methods may enable the utilization of genetic diversity in wild *Hordeum* species, which present a valuable resource for barley improvement and the generation of a perennial barley crop. However, the development of high-yielding, disease-resistant perennial crops remains a significant challenge, requiring ongoing research into the physiological traits and genetic mechanisms that control key differences in annual and perennial growth habits (Chapman et al. 2022).

Studies of trait variation between annual and perennial species span diverse lineages (e.g. De Souza and Da Silva 1987; Herron et al. 2020; Gonzalez-Paleo et al. 2024; Anokye et al. 2025), but genetic insights come mainly from a few models. Brassicaceae is probably the most prominent of these groups, where epi-

genetic control of the genes FLC (annuals) and PEP1 (perennials) dictates seasonal versus perpetual flowering, and modulating their dosages plus two related MADS-box genes can shift plants along an annual–perennial continuum (Bastow et al. 2004; Sung and Amasino 2004; Wang et al. 2009; Amasino 2010; Zhai et al. 2024). Contrary to the intrinsic seasonality that seems to differentiate annual from perennial Brassicaceae, in the tree model hybrid aspen, seasonal growth is regulated by external environmental cues (Böhlenius et al. 2006). Monocot life-history genetics remain elusive: only two perennial crop varieties, PR23 rice (crossed with wild *Oryza longistaminata*) and Kernza wheatgrass, have been developed, with most studies focusing narrowly on rhizome formation (Hu et al. 2003; Hu et al. 2003; Hu et al. 2011; Gruner and Miedaner 2021; S. Zhang et al. 2023). Comparative phylogenetics leverages species or clade phylogenies to test evolutionary hypotheses and, with the explosion of sequence data and advanced methods over the past thirty years, has become a versatile framework for studying phenomena such as trait evolution without prior knowledge of specific genes (Cornwallis and Griffin 2024). However, poorly resolved trees - due to limited genomic data, incomplete lineage sorting, or hybridization - can mislead inferences about trait evolution (Soltis and Soltis 2003). Overcoming these issues requires sophisticated phylogenetic models and high-quality genomic resources. The recent publication of genomes for all 21 diploid and three tetraploid *Hordeum* species (Feng et al. 2025) enables comparative genetic studies of the evolution and genetic architecture of various traits, including life-history strategies in the *Hordeum* clade.

In this study, we generated and utilised transcriptomic data from 1-10 accessions of all diploid *Hordeum* species, the 24 recently published *Hordeum* genomes (Feng et al. 2025), and publicly available observational data to address three research aims using comparative phylogenomic analyses: (i) increase the resolution of the *Hordeum* phylogeny and investigate ancient inter-species hybridisation events, (ii) uncover the distribution of annual and perennial *Hordeum* species within climatic space, and (iii) explore genetic mechanisms (gene copy number variation, differential expression, divergent signatures of selection) that differentiate between annual and perennial *Hordeum* species.

Results

Hordeum phylogeny and interspecies hybridization

We constructed three phylogenetic trees using distinct datasets and methodologies to assess their topological congruence and investigate various evolutionary phenomena. First, we generated a maximum-likelihood accession-based tree using the combined RNA-seq and reference genome dataset. In this framework, each tip represented an individual sample (accession), allowing us to capture intraspecific variation and evaluate the robustness of species boundaries. This tree was built using 257 single-copy orthologs (scHOGs) shared across all *Hordeum* and outgroup accessions, encompassing a total sequence length of 440,610 bp and 52,026 parsimony-informative sites (Tab. 1). Second, we constructed a Bayesian species tree employing the multi-species coalescent model while simultaneously estimating divergence times. This approach enhances the accuracy of both tree topology and branch length estimation by mitigating biases associated with unequal sampling, intraspecific and gene tree variation, and confounding signals from incomplete lineage sorting and/or hybridization. The species tree was also inferred from the combined RNA-seq and reference genome dataset. However, due to the high computational demands of this approach, the analysis was limited to 90 single-copy orthologs (scHOGs), which had a total alignment length of 225,942 bp and 22,571 parsimony-informative sites (Tab. 1). Third, we constructed a maximum-likelihood tree from the reference genomes of (Feng et al. 2025) only (genome accession tree). This tree was mainly a side-product of the analysis of gene CNV and was used for visualization. Note that the data contained only a single accession per species, so while it appears like a species tree, it is an accession-based tree that lacks information about intraspecies variation. The tree

130   included several distant outgroup species and was based on 380 scHOGs, a total alignment length of
131   558,769 bp and 115,640 parsimony-informative sites (Tab. 1).

132   *Table 1*: Summary statistics of single copy hierarchical orthogroups used for inference of the three phylo-
133   genetic trees (for a comprehensive list of statistics see supplementary Tab. S5).

	alignment length [bp]	alignment length without missing data [bp]	variable sites	proportion of variable sites [%]	parsimony informative sites
Accession based tree (257 scHOG; RNA-seq + reference genomes; Fig. 1)	440,610	355,954	86,670	0.2	52,026
Species tree (90 scHOG; RNA-seq + reference genomes; Fig. 2)	225,942	189,900	36,153	0.16	22,571
Genome accession tree (380 scHOG; reference genomes; Fig. 4A)	558,769	453,715	294,500	0.53	115,640

The topographies of the three trees were almost identical (Fig. 1, 2, 4a, S7). Around 10.3 million years
ago the *Hordeum* clade split into two large monophyletic groups (95% HPD interval: 8.2, 15.8; for a
complete list of exact divergence times, see Fig. S8). One group consisted of the sections Hordeum and
Trichostachys with three species growing in Europe, the Mediterranean and central Asia. All species in
these two sections are annuals, except for the pseudo-annual *H. bulbosum* (Fig. 1, 2). The other large
monophyletic groups included sections Marina and Stenostachys. Section Marina contained only two
annual species from Europe and the Mediterranean. Section Stenostachys was the largest section with 16
diploid and two tetraploid species separated into the Asian series Sibirica and the American section
Critesion. Only the three youngest of these 18 species (*H. pussilum*, *H. euclaston*, *H. intercedens*) are
annual species. Support for interspecies nodes was overall high. In the species tree, only three posterior
probabilities fell below 1, with values of 0.99, 0.94, and 0.86 - all within the series Stenostachys. In the
accession-based tree, the support values of 88.6/96 UFB/SH-aLRT, the split of *H. comosum* from the
other Critesion species was the only interspecies node supported with < 90% bootstrap values (Fig. 1). In
the genome accession tree, the node following the split to *H. comosum* had very low support with 34.8/65
(Fig. 4a, S7). This also highlighted the only incongruence of the three trees, which depicted *H. comosum*
as the youngest South American species in the species tree (Fig. 2), but as the second youngest lineage,
after the clade of *H. muticum* and *H. cordobense,* in the two trees based on individual accessions (Fig 1,
4a, S7). The overall high topographic congruence and node supported of the three trees suggested a solid
phylogenetic inference. However, the case of *H. comosum* indicated that evolutionary processes in the
closely related Patagonian species complex might be challenging to discern with the utilized data and/or
methods.

Interspecies hybridization is one such process that often limits phylogenetic analyses when using strictly
bifurcating trees. We expanded our phylogenetic analysis using a two-step approach to investigate
reticulations within the *Hordeum* tree. First, we searched for candidate hybridization events by creating
ten repetitions of phylogenetic networks with the number of reticulations ranging from 0-10. Eight
candidate networks were selected as they showed the best statistical support (Fig. S5, S6). Reticulations
that occurred in at least two of the candidate networks were considered potentially true hybridization

events. This analysis yielded five supported interspecies reticulations (R1–R5; Fig. 2). Notably, three of
these events (R1–R3) were linked to long-distance dispersal. Specifically, we detected a reticulation
between *H. bulbosum* (Eurasia) and the most recent common ancestor (MRCA) of extant American
species (R1). Additionally, two reticulations were identified involving *H. californicum* (North America):
one with the MRCA of South American species (excluding *H. comosum*; R2) and another with the South
American MRCA of *H. muticum* and *H. cordobense* (R3). Surprisingly, we identified only a single
reticulation among the youngest, less diverged species (*H. pussilum* → *H. stenostachys*; R5). This event
also involved long-distance dispersal, however, it was ultimately refuted in the second step of our
procedure, where we calculated D-statistics with ABBA-BABA tests to validate R1-R5. Each candidate
reticulation was assessed using multiple ABBA-BABA tests, for which we calculated summary statistics
(table inlay Fig. 2; Tab. S6). The single test for R5 was not statistically significant ($p > 0.05$). In contrast,
the remaining reticulations were strongly supported, with more than ~90% of their respective tests
yielding p-values < 0.05. Variation in substitution rates among species can generate homoplasies that
mimic ABBA patterns, potentially leading to false-positive D-statistics.

Figure 1: Accession based maximum likelihood tree. Node labels depict UFB/SH-aLRT values. Node labels are
only shown if they were not the maximum of 100/100. Infrageneric groupings (sections and series) were defined
sensu Blattner (2009).

To address this, we evaluated the degree of clustering of ABBA patterns, distinguishing between those
arising from true hybridization events (clustered) and those resulting from homoplasies (randomly distrib-
uted). This analysis reinforced the validity of reticulations R1–R4 as genuine hybridization events as the
majority of their tests supported a strong clustering tendency of their ABBA patterns (table inlay Fig. 2;
Tab. S6). Our analysis initially identified a large number of potential hybridization events (Fig. S6). How-
ever, given the statistical challenges associated with these methods, we applied stringent filtering to prior-

itize the sensitivity of our approach. This refinement yielded four well-supported reticulations (Fig. 2).
Notably, the majority of these hybridization events -three out of four- were linked to long-distance dis-
persal, rather than occurring between species growing in proximity. This may indicate an important role
of dispersal-driven hybridization in shaping the evolutionary history of *Hordeum*.Transitions between life
history strategies appear to have occurred independently on at least three occasions: from annual to peren-
nial along the *H. bulbosum* lineage, again from annual to perennial between ~7.5 and 5.5 million years
ago during the expansion into Central Asia, and ~1.5 million years ago along the branch leading to the
American annuals (Fig. 2). Notably, similar to the first transition, the second shift was also linked to a
dispersal event, this time from South to North America. It should be noted, however, that the exact
branches along which the true transitions occurred cannot be inferred from the tree.

195
196 *Figure 2:* Bayesian species tree. Numbers at the nodes are posterior probabilities. Species names are coloured ac-
197 cording to their geographic distribution. Tip colours represent life cycle strategy. Lines R1-5 represent candidate re-
198 ticulations. The table summarize the results of the ABBA-BABA test.

Biogeography of annual and perennial *Hordeum* species

Divergent life cycle strategies are widely regarded as evolutionary adaptations shaped by natural selection
in response to species-specific environmental pressures, because these strategies influence a suite of
functional traits critical to plant fitness, including growth dynamics, reproductive investment, and survival
mechanisms. Herein, we investigated macroclimatic variables of *Hordeum* species' publicly available
observations to assess whether large-scale patterns of temperature and precipitation are associated with
these contrasting life cycle strategies within the clade.

Annual species occupied a substantially smaller space in the bioclimatic PCA than perennials (Figure
3A). The space occupied by perennials covered most of the annuals' and additional areas along the axis of
precipitation seasonality (bio15) and temperature annual range (bio7). Accordingly, the climate hyper
volumes representing realized niche spaces were larger in perennials (0.93) than in annuals (0.73).
Perennials covered 92.3% of the annual niche space and annuals 58.1% of the perennial space. It is
important to note, however, that the analysis included a greater number of perennial (13) than annual (7)
species, with the broader climatic niche of perennials largely driven by two specific clades. The first
comprised the central and East Asian species *H. bogdanii*, *H. brevisubulatum*, and *H. roshevitzii* (series

Sibirica), which inhabit regions characterized by a pronounced continental climate, exhibiting warm summers (bio10), extremely low winter temperatures (bio6), and consequently, an exceptionally large annual temperature range (bio7, 3A). The second group includes *H. comosum*, *H. patagonicum*, and *H. pubiflorum*, native to Patagonia and the Andes, where high precipitation seasonality (bio15) expands the climatic niche of perennials. These two clades consist of closely related species (Fig. 1, 2, 4A), suggesting that the broader niche space observed in perennial species may primarily reflect clade-specific adaptations rather than an intrinsic consequence of their perennial life history strategy.

Figure 3: Habitat description of *Hordeum* species. A: Principal component analysis of species by bioclimatic variables. B: Boxplots of percentage landcover type of annual and perennial species with p-values from phylogenetic non-parametric ANOVA. C-F: Boxplots of monthly mean temperature (C), precipitation (D) and interannual variation of temperature (E) and precipitation (F) of annual and perennial species with posterior propabilities that annuals show higher values than perennials.

Consistent with this perspective, the climatic niche spaces of individual species did not differ significantly between annual and perennial *Hordeum* species (p ~ 0.36; Fig. S9). No consistent differences in mean precipitation across entire seasons were apparent between the habitats of annual and perennial species; the strengh of evidence was month-specific. Posterior probabilities indicate moderate support for higher precipitation in annual habitats from late autumn to early-spring months ($P_{annual > perennial}$ ~ 0.76 - 0.82; Fig. 3D). By contrast, summer months show moderate evidence for greater precipitation in perennial habitats ($P_{annual > perennial}$ ~ 0.18 - 0.25, i.e. perennial > annual).

Temperature, however, tended to be higher in the habitats of annual species throughout the year. Posterior probabilities provide moderate to strong evidence that annuals experience higher temperatures in essen-

tially all months ($P_{annual > perennial} \sim 0.72$–$0.93$; Fig. 3C), with the strongest support from late summer through
the winter months ($P_{annual > perennial} \geq 0.88$).

Analyses of interannual climatic variation produced a broadly similar picture for temperature variability:
there was moderate to strong evidence that annuals occupy regions with greater interannual temperature
variability with $P_{annual > perennial} \sim 0.78 - 0.96$ throughout the year (Fig. 3E). Interannual variation in precipit-
ation shows weaker and more heterogeneous evidence: annual habitats show moderate evidence for
greater interannual precipitation variability from late-autumn to early spring months ($P_{annual > perennial} \sim 0.74$
$- 0.86$; Fig. 3F), while the summer months exhibit no evidence for consistent differences between life his-
tories.

In terms of land-use associations, annual species exhibited a tendency to occur more frequently near crop-
lands and less frequently in grasslands compared to perennials. However, the only statistically significant
difference in occurrence frequencies was observed for built-up areas. Built-up areas encompass all forms
of human-made infrastructure, including residential or industrial buildings, roads, bridges, and other con-
structed features of the urban or semi-urban landscape. On average, that type of landscape accounted for
$\sim 15\%$ and $\sim 8\%$ of land cover in the 100×100 m grid cells surrounding observations of annual and peren-
nial species, respectively (Fig. 3B).

Taken together, differences in climatic niche size between annual and perennial *Hordeum* species are
likely shaped by clade-specific adaptations. Meanwhile, climatic distinctions between their habitats were
not characterized by a consistent year-round separation in precipitation, instead, posterior probabilities in-
dicate a seasonally opposing pattern, with annual species occupying wetter habitats in winter, while per-
ennial species tend to occur in wetter habitats during summer. These seasonal shifts contrast with the
much clearer temperature signal. Annual habitats were consistently warmer throughout the year, with
moderate to strong evidence for higher temperatures in all months, and they also showed markedly
greater interannual temperature variability, particularly from late autumn through early spring. In line
with the reduced temperature predictability of their environments, annual species also occurred more fre-
quently in human-disturbed landscapes, which are known for lower ecological stability.

Divergent genetic determinants in annual and perennial *Hordeum* species

Understanding the genetic differences that distinguish annual from perennial Hordeum species can clarify
the mechanisms underlying repeated transitions between these life-history strategies. We focused on three
genomic features divergent between annual and perennial *Hordeum* species to uncover the genetic archi-
tecture (complexity) underlying the different life histories: (i) signatures of divergent selection in protein-
coding sequences, (ii) differential gene expression, and (iii) gene copy number variation.

Across analyses of divergent selection, differential gene expression, and gene copy-number variation, we
identified 174 candidate genes. Of these, 150 could be assigned to five broad functional categories - de-
velopment, metabolism, cellular organisation, stress response, and transcriptional regulation - while 24
encoded proteins of unknown function or retrotransposon-related proteins (Tab. 2; Fig. 4A; Tab. S7). Be-
cause we derived functional assignments from conserved domains and annotations in *Arabidopsis thali-
ana* and other model plants, these categories should be considered general heuristics rather than precise
descriptions of gene function in grasses or *Hordeum*. Candidate genes were distributed across all func-
tional categories without significant enrichment (χ^2 tests, $p < 0.05$; Fig. 4A), indicating that divergence
between annual and perennial life histories involves these functional categories equally. In line with this
pattern, GO enrichment analyses identified terms largely corresponding to these five functional categor-
ies, including abiotic and biotic stress responses (oxidoreductase and hydrolase activities, nucleoside
phosphate binding), metabolism (carbohydrate, hexose, small-molecule, vitamin, and protein metabolic
processes), cellular organisation (microtubule-based movement, cellular component assembly), and tran-
scriptional regulation (ribonucleotide and nucleotide binding; Tab. S12, S13).These results indicate that a
complex genetic architecture covaries with repeated transitions between annuality and perenniality in

Hordeum. The following sections summarise the results from the selection, expression and gCNV ana-
lyses.

Table 2: Summary of the categorization of candidate genes across analyses of divergent genetic mechan-
isms; selection, differential expression and gene copy number variation (for a comprehensive list of can-
didate genes, their descriptions and categorizations see Tab. S7).

Category	Explanation	Total number of genes per category	Number of genes from selection analysis	Number of genes from differential expression analysis	Number of genes from gCNV analysis
Signal Transduction & Development	Includes receptors, two-component response regulators, and other proteins involved in perceiving signals and controlling developmental programs.	45	25	11	9
Metabolic & Biosynthetic Processes	Contains proteins that participate in the construction and breakdown of cellular components, covering core metabolic pathways and biosynthetic activities.	29	20	4	5
Membrane Transport & Cellular Organization	Groups proteins involved in moving molecules across membranes and those contributing to cellular trafficking and structural organization.	28	16	8	4
Stress Response & Defense Mechanisms	Encompasses proteins that mediate responses to environmental stress, aid in cellular protection, or are implicated in defence responses.	27	12	9	6
Transcription & Translation	Encompasses proteins that together coordinate transcriptional control, RNA maturation and stability, and the assembly and function of the ribosome.	21	13	4	4
Mobile Genetic Elements & Uncharacterised Functions	Groups retrotransposon-related proteins, reverse transcriptase domains, and genes with unknown functions.	24	6	5	13
	Total number of genes per analysis	174	92	41	41

To detect signatures of divergent selection between annual and perennial Hordeum species, we analyzed
2,908 scHOGs represented by at least two accessions per species using two complementary approaches:
(i) D_n/D_s ratio modelling, and (ii) correlation between gene evolutionary rates and life-history strategy. A
total of 576 genes showed significant Dn/Ds differences, and 442 showed significant trait-rate correla-
tions. Applying a conservative criterion that required significance in both analyses, we identified 92 can-
didate scHOGs (Tab. S8). Most candidates (62) exhibited negative trait-rate correlations, indicating lower
relative evolutionary rates in annuals and correspondingly higher D_n/D_s ratios in perennials. These genes
thus appear to be under stronger selective constraints in annuals than in perennials. Because our analyses
capture contemporary evolutionary rates rather than events associated with the original life-history trans-
itions, the direction of historical transitions (~8–4.5 Myr from annual to perennial; ~1.5 Myr for the re-
verse; Fig. 2) is unlikely to influence these patterns. Nine genes showed the opposite pattern, with higher

evolutionary rates and higher Dn/Ds ratios in annuals. The remaining 20 genes displayed conflicting signals between the two methods, typically cases where one statistic was significant but effect sizes were small (e.g., RNA.HOG0025477). These genes may represent true candidates, but their support is weaker relative to the core set.

To quantify expression divergence between annual and perennial accessions, we analyzed pooled RNA-seq data from leaves, roots, stems, and inflorescences, which provided 7,942 scHOGs with adequate read counts. A generalized linear mixed model incorporating phylogenetic relatedness identified 41 DEGs at $p_{adj} \leq 0.01$ and $|log_2FC| \geq 1$ (Fig. 4C; Tab. S11; Fig. S10). Of these, 25 DEGs were on average more highly expressed in annuals, and 16 were more highly expressed in perennials. Variation analyses showed that 20 genes had >10% higher expression coefficient of variation (CV) in annuals, 8 genes had higher CV in perennials, and 13 showed similar variability in both life-history groups The pattern of higher mean expression and greater variability in annuals appears consistent with their fast-paced, resource-intensive life strategy, which may require both elevated and flexible gene regulation, in contrast to the more stable and regulated expression profiles of long-lived perennial.

To examine potential genomic dosage differences, we tested 29,565 gene families for consistent copy-number differences between life histories. We identified 41 gene families in which all perennial species across the different branches of the phylogeny possessed equal or higher copy numbers than annuals (Tab. S9). No families showed the reverse pattern. Among these, ten families had a single copy in annuals but between two and nine copies in perennials (e.g., GENO.HOG0004489, GENO.HOG0005530; Tab. S9). Another eight families were absent in annuals but present in one or more copies in perennials. More than half of these gene families fell into the "Mobile Genetic Elements & Unknown Functions" category, reflecting the high propensity of transposable elements to undergo copy-number expansion. To further investigate if these copies were also expressed, we examined gene expression for each gene copy within these families. In six families, perennials expressed more gene copies in our data than were present in annuals. Notable examples include GENO.HOG0004711, a senescence regulator, and GENO.HOG0007793, a TEOSINTE BRANCHED1-like gene, which function to integrate hormonal and environmental signals to regulate bud dormancy, suppress branching and flowering, and thus maintain perennial growth cycles (Kebrom et al. 2010; Dong et al. 2019; Zhao and Wang 2024). We note that these expression results are conservative, as multimapping reads were excluded. Some paralogs may also be expressed only under specific developmental or environmental contexts not represented in our sampling. Nonetheless, these findings demonstrate consistent gCNV differences between life histories and establish a basis for more detailed investigations into whether this variation affects expression, alters genomic dosage, or supplies raw material for evolutionary innovations such as neo-functionalization.

Figure 4: Divergent genetic mechanisms in annual and perennial Hordeum species without *H. bulbosum*. A: Radar plot of the number of genes per analysis assigned to categories. Numbers are relative to the size of the analyses. B: The genome accession tree excluding outgroup species (left). Node labels depict UFB/SH-aLRT values. Node labels are only shown if they were not 100/100. Gene copy number of a selection of candidate gene families (right). For the complete genome accession tree, see Fig. S7. For a comprehensive list of genes that are members of the selected gene families, see Tab. S9 and for the description of the gene families, Tab. S10. C: Ridgeplot of $\log_{10}$ transformed expression of HOGs which are differentially expressed in annual (red) and perennial (cyan) *Hordeum* species. HOGs are sorted according to the categories they were assigned to, and within categories, whether annuals (below the dotted line) or perennials (above the dotted line) showed higher expression.

Although no single gene was identified by more than one analytical approach, candidate genes from all analyses converged on broadly similar functional categories (Tab. S7).

The largest group fell into Signal 'Transduction & Development', comprising 45 of the 92 candidates. These genes include regulators of meristem maintenance, organ boundary formation, bud dormancy, and flowering-time control, all prime regulators of life-history strategies. Examples include *SCARECROW-LIKE6* (RNA.HOG0015866), involved in radial patterning and stem cell maintenance; a LOB-domain protein (GENO.HOG0022843), associated with boundary specification; and several NAC-domain transcription factors (GENO.HOG0031918, GENO.HOG0020669, RNA.HOG0010144), which influence both meristem function and senescence regulation (Bolle 2004; Curaba et al. 2013; Xing et al. 2024; Christiansen & Gregersen 2014). Four MYB transcription factors were identified under divergent selection (RNA.HOG0029412), differential expression (GENO.HOG0012470), and genomic copy-number variation (GENO.HOG000448, GENO.HOG0005530). MYB proteins orchestrate transcriptional net-

works that control cell division, differentiation, and organ patterning in response to both intrinsic and ex-
trinsic signals (Dubos et al., 2010; Ramsay & Glover, 2005).
A second group of candidates belonged to 'Transcription & Translation', including chromatin remodeler
orthologs such as RNA.HOG0026351 (orthologous to *AtSKIP*), RNA.HOG0027556 (orthologous to *At-*
PIE1), and RNA.HOG0028859 (orthologous to *AtSWP73*). These genes are known to regulate the key
floral repressor FLC in *Arabidopsis* (Carter et al., 2018; K. Choi et al., 2007), and their presence among
our candidate genes suggests that chromatin remodelers may play a role in shaping life history strategies
in *Hordeum* as well.
Another 29 candidate genes fell into the 'Metabolic & Biosynthetic Processes' category, encompassing
carbohydrate metabolism, carbon allocation, and cell-wall biosynthesis and remodeling. Several genes en-
code glycosyltransferases (GENO.HOG0007840, RNA.HOG0026057, RNA.HOG0010428), which cata-
lyze sugar-nucleotide–dependent reactions important for carbohydrate modification. Others, including in-
vertases (GENO.HOG0002011, GENO.HOG0008038) and pyrophosphate–fructose-6-phosphate phos-
photransferase (RNA.HOG0011119), are involved in carbon partitioning between growth and storage tis-
sues (van der Merwe et al. 2010; Duan et al. 2016). Additional candidates, such as 4-coumarate-CoA lig-
ase, glycerophosphodiester phosphodiesterase, glycosyl hydrolases, pectate lyases, and glucan endo-β-
glucosidases, participate in cell-wall formation and remodeling, which can affect structural and metabolic
properties of tissues. While the precise roles of these genes in annual–perennial differences remain to be
determined, their functions point to potential variation in carbohydrate processing, carbon allocation, and
cell-wall biosynthesis, processes that may underlie differences in growth strategy and resource manage-
ment between life-histories strategies.
The 'Stress Response & Defense' category (27 candidates) included AP2/ERF transcription factors
strongly upregulated in annuals (GENO.HOG0021020, GENO.HOG0021873), orthologous to *AtERF105*
and *AtERF53*, which mediate cold tolerance, ABA signaling, and heat responses (Hsieh et al. 2013; Illgen
et al. 2020; Ma et al. 2024). Perennials, by contrast, showed higher expression of genes involved in oxid-
ative stress mitigation, such as flavin-containing monooxygenases (GENO.HOG0016866, GEN-
O.HOG0007274) and chalcone synthase (GENO.HOG0019416), which contributes to flavonoid biosyn-
thesis and antioxidant defense (Dao et al. 2011). These contrasting signatures suggest that annuals invest
more heavily in acute stress-responsive transcriptional activation, whereas perennials rely on constitutive
or longer-term protective pathways consistent with multi-year survival.

Taken together, our investigation into divergent genetic mechanisms in annual and perennial *Hordeum*
species revealed candidate genes across all three genomic features analyzed (signatures of selection,
gCNV, differential expression) spanning a wide range of biological functions. This functional diversity
suggests that the life-history divergence between annuals and perennials is underpinned by multiple, in-
terconnected genetic processes, rather than a single dominant pathway.

Discussion

***Hordeum* phylogeny and interspecies hybridization**

The phylogenetic relationships among the Old World *Hordeum* species (sect. Hordeum, Trichostachys,
Marina, and ser. Sibirica; Fig. 1) have been consistently resolved over decades of research (Jörgensen
1986; Doebley et al. 1992; Svitashev et al. 1994; Marillia and Scoles 1996; Petersen and Seberg 1998; El-
Rabey et al. 2002; Petersen and Seberg 2003; Blattner 2006; Jakob and Blattner 2006; Brassac and
Blattner 2015; Jin et al. 2024; Mason-Gamer and White 2024; Feng et al. 2025). In contrast, phylogenetic
relationships within ser. Critesion, particularly among the more recently diversified South American
Hordeum species, have remained poorly resolved, frequently characterized by polytomies likely resulting
from the limited number and type of genetic markers used in earlier studies (Svitashev et al. 1994;
Petersen and Seberg 1998; Petersen and Seberg 2003; Blattner 2004; Jakob and Blattner 2006). A notable
advance was made by Brassac and Blattner (2015), whose use of 13 genomic loci substantially improved

phylogenetic resolution within the genus. While our species tree broadly corroborates their topology, there are some differences, most notably in the placement of *H. pusillum*, which they recovered as sister to *H. stenostachys* and *H. erectifolium*. This topological difference has important implications for reconstructing the evolution of life-history strategies in *Hordeum*, as their scenario would require at least one additional transition between strategies relative to our results. Importantly, node support values in our species tree are markedly higher at all points of topological disagreement with (Brassac & Blattner, 2015) and also exceed those reported by Jin et al. (2024), whose species tree is otherwise fully congruent with ours. This is likely due to the substantially larger data set that we used, with 22,571 parsimony informative sites (Tab. 1) compared to 2,852 and 7,064 in the data set of Brassac & Blattner (2015) and Jin et al. (2024), respectively. Given the larger dataset and consistently higher node support in our analyses, we consider our phylogenetic reconstruction to represent a more robust and reliable representation of species relationships in *Hordeum* compared to previous studies. As such, it provides a more suitable framework for inferring the evolution of life-history strategies within the genus. More broadly, these results underscore the critical importance of accurate phylogenetic relationships in comparative phylogenomic analyses, as incorrect assumptions about species' evolutionary histories can generate misleading signals, obscure true patterns of trait evolution, and ultimately lead to erroneous inferences about the genetic architecture underlying phenotypic diversity.

As increasingly large genomic datasets and more sophisticated analytical approaches are applied to phylogenetic inference, our understanding of species relationships continues to improve. However, it has also become evident that while additional data can strengthen support for many relationships, certain phylogenetic conflicts persist despite these advances (Mason-Gamer and White 2024). We observed a similar pattern in *Hordeum*: although the placement of most species was consistent across all our reconstructed trees, the position of *H. comosum* remained unstable, varying among alternative topologies. This phylogenetic instability is also reflected in other recent studies, where *H. comosum* was either recovered as sister to *H. patagonicum* and *H. pubiflorum*, forming the youngest independent South American lineage, as in our species tree, or as the second youngest independent South American lineage - following the divergence of the *H. muticum* and *H. cordobense* clade - as observed in both of our accession-based trees (Brassac and Blattner 2015; Jin et al. 2024; Feng et al. 2025). Since *H. comosum* is nested within the large clade of perennials, this instability is not relevant for our comparative phylogenomic analyses of life-history strategies. However, it may present challenges for studies focusing on other traits where the early South American lineages play a more important role.

The causes of this phylogenetic uncertainty remain speculative. A limited number of accessions per species - as used for *H. comosum* herein, in Jin et al. (2024) and Feng et al. (2025) - may constrain the ability accurately determine species relationships. A limited number of genetic markers - as used by Brassac and Blattner (2015) - can increase the risk of stochastic error and reduce the power to resolve complex evolutionary relationships. Additionally, it is well recognized that evolutionary histories are not always adequately represented by strictly bifurcating tree models (e.g. Bapteste et al., 2013; DeSalle & Riley, 2020; Morrison, 2005). We identified a large number of candidate inter-species introgressions in *Hordeum* using a phylogenetic network approach (Fig. S5, S6). However, the robust detection of introgression remains inherently challenging, as signals of gene flow are often subtle, restricted to specific genomic regions, and can be confounded by incomplete lineage sorting (Hibbins and Hahn 2022). To mitigate these challenges and minimize false positives, we focused on introgression events that were consistently recovered across multiple networks and further validated them using D-statistics. This conservative approach yielded four statistically well-supported cases of inter-species introgression. Notably, three of these events coincided with long-distance dispersal events - from Europe/West Asia to East Asia/North America (R1) and from South to North America (R2, R3; Fig. 2). None of our detected introgressions involved *H. comosum*, and therefore, reticulations fail to account for the inconsistent placement of this species in both our and earlier reported trees. Previous studies investigating inter-species introgression in

Hordeum have identified a variety of putative gene flow events; however, none of the introgression
events reported, including those detected in our study, have been independently corroborated (Jin et al.
2024; Feng et al. 2025). This apparent discrepancy is most likely attributable to the fact that different
studies targeted distinct genomic regions, applied varying statistical frameworks, or differed in sampling
strategies. This observation highlights a fundamental challenge in studying introgression in large-genome
taxa like *Hordeum* (~3–5 Gbp in diploid genomes; Feng et al. 2025) and underscores the necessity for
continued and integrative research efforts. A single study alone is unlikely to capture the full complexity
of introgression dynamics in a genus of this size and evolutionary history. Despite the methodological
challenges and limited congruence among studies, the accumulating evidence suggests that inter-species
hybridization may have occurred repeatedly and across broad geographic and phylogenetic scales
throughout the evolutionary history of *Hordeum*. The combination of previous reports on putative intro-
gression events, our results herein, and the presence of numerous polyploid *Hordeum* species - many of
which originated from hybridization between distantly related lineages across the genus (see results
herein: Fig. S7; Brassac and Blattner 2015; Jin et al. 2024; Feng et al. 2025) - collectively support a scen-
ario where inter-species hybridization may have been a recurrent and pervasive evolutionary process in
Hordeum.

Biogeography and evolution of annual and perennial *Hordeum* species

While annuality in plants is often considered to be the result of adaptations to unpredictable environ-
ments, high temperatures or seasonal droughts (Stearns 1992), empirical evidence shows that not all plant
genera consistently exhibit clear associations between life-history strategies and such climatic character-
istics of their native habitats (Boyko et al. 2023; Poppenwimer et al. 2023).

Annual and perennial *Hordeum* species exhibited distinct seasonal climatic patterns in their habitats. In
winter, habitats of annual species were both warmer and wetter than those of perennials, likely promoting
vegetative growth and seedling establishment. In contrast, during summer, annual habitats were hotter
and drier, possibly imposing terminal stress that favours senescence and survival in the seed stage, a typ-
ical annual-perennial ecological pattern (Stearns 1992). This seasonal contrast in temperature and precip-
itation is consistent with classical life-history theory for grasses: accelerated development and early mat-
uration are common escape strategies of grasses facing potentially terminal stress (Loka et al., 2019;
Jagadish, 2020; Bahrami et al., 2021).

We did not observe significant differences in niche space between annual and perennial *Hordeum* species
when analyzed individually (Supplementary Fig. S9). However, when annuals or perennials were con-
sidered jointly, perennial species occupied a substantially larger niche space (Fig. 3A). This pattern was
primarily driven by the series Sibirica, which expanded the niche breadth of perennials due to their asso-
ciation with environments characterized by a high annual temperature range (bio7) and low minimum
winter temperatures (bio6). Notably, the correlation between low winter temperatures and perenniality
has been identified as a general pattern not across all flowering plants, but within the Pooideae subfamily
(Boyko et al., 2023). According to life-history theory, perennial species possess an adaptive advantage in
environments where adult mortality is low but seedling mortality is high (Cole 1954; Charnov and Schaf-
fer 1973). This theoretical framework may help explain the observed transition from annuality to perenni-
ality during *Hordeum*'s expansion from Western Asia, characterized by a Mediterranean climate, into
Siberia's continental climate (Fig. 3). In this context, harsh Siberian winters may have acted as an envir-
onmental filter, selecting against annuality in the winter type *Hordeum* species of Europe and South-West
Asia due to elevated seedling mortality. Conversely, regions where annual *Hordeum* species are most pre-
valent, such as Mediterranean climates, are characterized by high summer temperatures that compromise
adult plant survival, while mild winters facilitate seedling establishment and survival, thereby favouring
annuality.

Low levels of ecological predictability have been linked to the prevalence of annual species, as their rapid life cycles may enable them to complete reproduction before the onset of unfavourable conditions (Stearns 1992; Poppenwimer et al. 2023). In our study, annual *Hordeum* species were found to occur more frequently in areas near human-made infrastructure (built-up areas; Fig. 3D). Although anthropogenic disturbance has not directly driven transitions between life history strategies in *Hordeum*, such disturbances are typically more intense near human settlements and may exert selective pressure favouring an annual growth habit. Furthermore, we observed greater interannual variability in temperatures and winter precipitation in regions inhabited by annual *Hordeum* species (Fig. 3C). Together, these findings suggest that annuality in *Hordeum* may confer an adaptive advantage under conditions of environmental unpredictability, consistent with life-history theory and previous findings in other plant taxa.

Divergent genetic mechanisms in annual and perennial *Hordeum* species

We used three complementary genome-wide approaches, scans for divergent selection, differential gene expression, and gene copy-number variation, to identify orthologous genes that consistently distinguish annual from perennial *Hordeum* species. By integrating phylogenetic relationships from our species tree and accounting for at least three independent life-history transitions within *Hordeum*, we controlled for shared ancestry and pinpointed genetic signatures genuinely linked to life-history divergence. This led to 174 high-confidence candidate genes across diverse functional categories, suggesting a convergent genetic basis for annual and perennial strategies in *Hordeum*. Genetic convergence mirrors known phenotypic patterns: across angiosperms, annuals typically exhibit faster growth, earlier flowering, and higher reproductive allocation, whereas perennials favor conservative resource use (Garnier 1992; Lundgren and Des Marais 2020; Hjertaas et al. 2023). Similar trends, such as earlier flowering and greater early biomass in annual *Hordeum*, have been observed, though leaf-economic traits are less consistent (Anokye et al. 2025). Thus, our discovery of convergent genetic changes parallels broader ecological syndromes. Most previous genetic studies of annual-perennial divergence lacked phylogenetic replication, making it hard to separate true convergence from shared ancestry. Exceptions in *Arabis* and *Draba* used independent annual-perennial sister lineages to uncover convergent mechanisms (Heidel et al. 2016; Kiefer et al. 2017). For instance, Heidel et al. (2016) found five gene families retained in Arabis and Draba perennials but lost in annuals. Notably, our *Hordeum* gCNV analysis recovered families with matching predicted functions for three of these: oxidoreductases (GENO.HOG0003613, GENO.HOG0005718, GENO.HOG0007274), an F-box protein family (GENO.HOG0002661), and zinc-finger proteins (GENO.HOG0001510, GENO.HOG0009714, GENO.HOG0026903; Tab. S9). Despite such examples, phylogenetically replicated studies of genetic convergence of life-history strategies remain scarce, and to the best of our knowledge, none have hitherto been reported in grasses. In this context, our work contributes to a growing but still small literature that applies replicated, phylogenetically informed comparisons to pinpoint genetic changes specifically linked to life-history transitions, rather than unrelated lineage-specific evolution.

The repeated transition between annual and perennial life history in *Hordeum* supports the idea that life-history transitions involve relatively simple genetic changes. This view originates from theoretical arguments suggesting that traits evolving repeatedly over short evolutionary timescales are unlikely to involve complex genetic architectures. The view was further reinforced by QTL studies that often find only a few major-effect loci and genes (Friedman and Willis 2013; Heidel et al. 2016 ; Kiefer et al. 2017; Gruner and Miedaner 2021) and by work in Brassicaceae linking annual-perennial shifts to altered expression of only a few MADS-box genes (Melzer et al. 2008; Li et al. 2024; Zhai et al. 2024). While studies in the Brassicaceae suggest that genetic changes in a limited number of developmental genes can drive transitions along life-history strategies (Zhai et al. 2024), candidate genes differentiating between annual and perennial *Hordeum* species spanned a broad array of functions. However, from our analysis we cannot distinguish between genes that are directly causal for differences in plant longevity and those that more likely covary with life history variation, such as biotic and abiotic stress resistance genes. Our data sug-

gests that the genetic basis of perennialism involves a suite of changes in plant developmental,
physiology, nutrient allocation and defense in *Hordeum*, suggesting that successful, stable transitions in
nature require coordinated changes across multiple genes and pathways that together produce the full
suite of associated traits. In the context of the increased interest in perennialization of annual crops
(Crews et al. 2018; Zhao and Wang 2024), this has significant implications: breeding perennial crops that
are agronomically competitive to their annual counterparts may demand genetic modifications in many
functions and pathways.

Signal-transduction and developmental regulators formed the largest functional category among our can-
didates. We detected a *Hordeum* SCARECROW-LIKE6 (SCL6) ortholog under divergent selection. In
Arabidopsis, SCL6 overexpression delays flowering and extends lifespan, sustaining meristem activity
(Xing et al. 2024). A TEOSINTE BRANCHED1 (TB1)-like gene family was also under divergent selec-
tion. TB1 functions as an integrator of hormonal and environmental signals, regulating bud dormancy,
suppressing branching and flowering, and thereby contributing to the maintenance of perennial growth
cycles (Dong et al., 2019; Kebrom et al., 2010; Zhao & Wang, 2024). Transcription factors in the NAC
and MYB families, key mediators of ABA- and JA-dependent stress, dormancy, and senescence path-
ways, also exhibited divergent signatures of selection. For instance, *Arabidopsis* ORE1 (a NAC) is a mas-
ter regulator of senescence (Chun et al., 2023), and barley NAC promoters often contain ABA- or GA-re-
sponsive elements (Arif et al., 2025), suggesting shifts in the timing of senescence and dormancy between
strategies. We further identified three chromatin-remodelling genes under divergent selection, orthologs
of Arabidopsis SKIP, PIE1, and SWP73, which participate in SWR1 and SWI/SNF complexes that regu-
late the floral repressor FLC in *Arabidopsis* (Aslam et al., 2019; Carter et al., 2018; Choi et al., 2017; Cre-
villén & Dean, 2011). Mutations in these genes in Arabidopsis reduce FLC expression and cause early
flowering (Bastow et al., 2004; Hepworth & Dean, 2015), suggesting epigenetic regulation of flowering
may also be involved in life-history divergence in *Hordeum*. Regulation of senescence is also closely
linked to source-sink dynamics. As discussed above, transcription factors like NAC and MYB influence
aging processes. Early activation of these genes in annuals may promote senescence to remobilize nutri-
ents into developing seeds, while perennials may delay their expression to prolong leaf function. Beyond
transcriptional control, we also identified several metabolic enzymes under divergent selection. Notably,
multiple glucosyltransferases showed such signatures and several studies highlight the role of glucosyl-
transferases in modulating source-sink dynamics (Stein & Granot, 2019). Similarly, UDP-glycosyltrans-
ferases (UGTs) fine-tune source–sink balance by regulating hormone levels and sugar availability (Ross
et al., 2001). Another enzyme that showed divergent signatures of selection was pyrophosphate–
fructose-6-phosphate 1-phosphotransferase (PFP). PFP has been shown to play a critical role in regulating
carbon partitioning between growth and storage in sink tissues in annual (rice; (Duan et al., 2016) and
perennial (sugarcane; van der Merwe et al., 2010) grasses. Our findings suggest that a key component of
the genetic differences between annual and perennial *Hordeum* species lies in carbon metabolism and
source-sink allocation. Annuals tend to prioritize rapid growth and seed development by channelling pho-
tosynthates quickly, while perennials allocate more resources toward maintenance, defense, and long-
term storage. Perennial species are expected to exhibit higher stress resilience since they need to maintain
their vegetative material over several years while annuals for example avoid late season stress by com-
pleting their life cycle early (Calone et al., 2022; Chapman et al., 2022; Lundgren & Des Marais, 2020).
Consequentially, we also found several candidate genes categories as ‚Stress Response & Defense Mech-
anisms‘. For example, we observed differential expression of two genes from the AP2/ERF transcription
factor family, which are known to regulate a wide range of abiotic and biotic stress responses (Ma et al.,
2024). Conversely, higher expression in perennials were observed for Chalcone synthase (CHS). CHS is a
key enzyme in flavonoid biosynthesis, catalyzing the first step toward production of anthocyanins and
other flavonoids, and it is strongly induced by stress (Dao et al., 2011). Divergent signatures of selection
were observed for a chloroplastic ClpB chaperone, which play crucial roles in cell adaptation to high-tem-
perature conditions (Mishra & Grover, 2016). Our candidate genes in these functional categories align

with classic annual - perennial life-history trade-offs. ‚Signal Transduction & Development' genes set the developmental program by controlling the timing of flowering, dormancy, and meristem activity. Genes related to „Metabolic & Biosynthetic Processes" execute resource-use strategies by modulating carbon flux, storage compound synthesis, and metabolic priorities. In addition, the enrichment of 'Stress Response & Defense Mechanism' genes supports the notion that perennials invest more in long-term tissue protection and stress resilience, enabling them to survive across multiple growing seasons. In contrast, annuals often evade late-season stress by accelerating their life cycle, prioritizing rapid growth and reproductive output.

Material and Methods

Plant Material and RNA sequencing

We compiled a collection of 82 genebank accessions, which included 19 *Hordeum* and two outgroup species (Supplementary Tab. S1). The seeds of these accessions were sown in freshly prepared soil mixture, 99.6 % (v/v) Mini tray soil (MIM800, Balster Einheitserdewerk, Fröndenberg, Germany) and 0.4 % (v/v) Osmocote exact standard 3-4M (Scotts Company LLC). Seeds were stratified in the dark for 14 days at 4 C° and subsequently germinated under long day conditions in walk-in phytochambers (day: 16h, 22 °C; night: 8h, 18 °C). Seedlings that developed 2-3 true leaves were moved to vernalization for 9-14 weeks (day: 8h, 4°C; night: 16h, 4°C). After vernalization, seedlings were moved back into walk-in phytochambers (day: 16h, 22 °C; night: 8h, 18 °C) and grown to maturity.

Tissue samples were taken from roots, leaves, stems and inflorescence. Root samples were taken during re-potting of plants into 7x7cm pots around 14 days after transfer out of vernalization. Leaf samples were taken from the second or third fully developed true leaf at Zeitgeber (ZT) 13.5-15.5 and 21.5-23.5. Inflorescence samples were taken in the booting stage (Zadok stage 60-69) sampling only the central part of the inflorescence (middle third of the inflorescence). Stem samples were taken at the same time from the same tiller as the inflorescence sample. We removed the leaves from the stem and sampled one random internode Note that not all plants flowered within one year, and therefore, we could not sample all tissues of all plants (Tab. S1).

RNA was extracted from all tissue samples using a RNeasy kit (Qiagen, Hilden, Germany). The extracted RNA of roots and leaves were pooled, and the RNA of stem and inflorescence were pooled for each accession with equal amounts of RNA per tissue sample. Both pools per accessions were sent to Biomarker Technologies (BMK) GmbH (Münster, Germany) for strand-specific poly-A enriched mRNA library preparation and sequencing of min. 6Gb/sample on an Illumina Novaseq 6000 with 150bp paired-end reads.

Transcriptome assemblies

Raw reads from the leaves-root and the stem-inflorescence pools were merged into a single fastq file per accession. After assessing the raw reads' quality using FastQC (v0.11.9; Andrews, 2010) and MultiQC (v1.7; Ewels et al., 2016), the raw reads were filtered with fastp (v0.23.2; Chen et al., 2018; Fig. S1) using the following parameter settings: -l 50, --detect_adapter_for_pe, --cut_right, --cut_right_mean_quality 20, -c.

Transcripts were assembled *denovo* using rnaSPADES (v3.9.1; Bushmanova et al., 2019; Prjibelski et al., 2020) with default settings. Additionally, a genome-guided assembly was done with Trinity (v2.2.0; Grabherr et al., 2011). We ran Trinity three times with kmer 20, 26 and 32 and the parameters --SS_lib_type RF and --genome_guided_max_intron 2500. The bam files required for genome-guided assembly were created by mapping the filtered reads against the reference genomes of the respective species (Tab. S1) except for the outgroup *Psathyrostachys juncea* (Fisch.), which due to the lack of a

reference genome, was mapped against the reference of the second outgroup *Dasypyrum villosum* (L.
Borbás; X. Zhang et al. 2023). Our collection contained two tetraploid species, *Hordeum bulbosum* (L.;
autotetraploid) and *Hordeum murinum* ssp. *murinum* (L.; allotetraploid). For autotetraploid *H. bulbosum*
we arbitrarily chose genome 1 from the reference genome and treated the two accessions as diploids.
Reads from *H. m.* ssp. *murinum* accessions were mapped against the allotetraploid genome. Reads that
mapped to genomes 1 and 2 (herein denoted as 'G' and 'M', respectively, *sensu* Rajhathy & Morrison,
1962) were extracted and separately mapped against the separated G or M genome reference. The
separated reads from the G and M genome of each *H. m.* ssp. *murinum* accessions were treated like two
diploid samples for further procedure. The mapping procedure was done using STAR (v2.7.10a; Dobin et
al., 2013) with default settings. In a subsequent filtering step with SAMtools (v1.3.1; Danecek et al.,
2021), only uniquely mapped reads were retained.

The four assemblies per accession were merged into a single fasta file and mapped against the same
reference genome used for read mapping using Minimap2 with the '-ax splice' option (v2.24; H. Li,
2018). The longest transcript was extracted from the resulting bam file with AGAT (v0.8.0; Dainat,
2020). We used BLAST+ (v2.2.26; Camacho et al., 2009) with the parameters -max_target_seqs 1, -
outfmt 6 and -evalue 1e-5 to create a database based on the concatenated proteins of the barley variety
Morex (v3; Mascher et al., 2021) and *D. villosum*. This database was input for Transdecoder (v5.5.0;
Haas, 2024) which we used with default parameters to predict open reading frames and collections of the
longest transcript, coding and protein sequences.

The sequence production and transcriptome assemblies' workflow was summarized in Fig. S2.

Detection and processing of orthologs

The predicted protein sequences from the 85 assemblies (77 accessions from diploid species + 2 *H.*
bulbosum + 3 *H. murinum* G-genomes + 3 *H. murinum* M-genomes) were complemented with protein
sequences from 13 accessions from the utilized *Hordeum* reference genomes (Feng et al. 2025). Thus, the
final collection consisted of 95 accessions plus 3 'artificial' diploid *H. m.* ssp. *murinum* samples from in
total 21 *Hordeum* and two outgroup species (Tab. S1, S2). The protein sequences of this collection were
used as input for OrthoFinder (v2.5.4; Emms & Kelly, 2019) with parameters -M msa and -T fasttree. The
resulting gene families will have the prefix "RNA." in the following to distinguish them from the second
OrthoFinder analysis described below.

After inspecting the resulting tree created by OrthoFinder, seven accessions did not cluster at their
expected positions in the tree (marked red in Fig. S3). Note, that these seven accessions could not
represent intra-species variation as they clustered far away from the approximate position of their species
in the tree. For example, *H. chilense* is a South American species, yet two of its samples clustered deep in
the clade of an Asian species (*H. roshevitzii*). We consider it to be likely, that all of these seven plants did
not belong to the taxa they were assigned to and, therefore, represented faulty phylogenetic signals.. Since
these accessions may have biased the detection of orthologs, we removed them and repeated the analysis
with the pruned set of 87 accessions plus three 'artificial' diploid *H. m.* ssp. *murinum* (Tab. S1) and
continued with this pruned set of accessions.

After running OrthoFinder again with the pruned set of accessions, we extracted single-copy hierarchical
orthogroups (scHOGs) which were present in all accessions and aligned their coding sequences to create a
multiple sequence alignment (MSA) for each scHOG. Alignment was done with MAFFT (v7.407; Katoh
et al., 2002). Each MSA was trimmed using Clipkit (v1.4.1; Steenwyk et al., 2020) with default
parameters to increase the signal-to-noise ratio and improve the MSAs' quality for subsequent
phylogenetic analyses.

Phylogenetic tree inference

We created partition schemes from scHOGs using PartitionFinder (v2.1.1; Lanfear et al., 2017) using raxml, a greedy search across all models and aicc as a decision criterion for model selection.

Fasta files were created that contained the scHOG sequences from the determined best fitting partitioning scheme and partition-trees of each partition were created using IQtree (v2.2.0; Minh et al., 2020) with 1000 ultrafast bootstrap values (UFB). Substitution models were determined with ModelFinder (Kalyaanamoorthy et al. 2017). The partition-trees and the MSAs were used to estimate a number of properties and sorting the partitions according to their usefulness for phylogenetic inferences following the method of genesortR (Koch 2021).

None of the partitions exhibited any properties that suggested they may introduce a bias (Fig. S4). Hence, all partitions were concatenated and used to create an accession-based phylogenetic tree using the maximum-likelihood approach implemented in IQtree (v2.2.0; Minh et al., 2020) with the best fitting models of nucleotide substitution determined by ModelFinder (Kalyaanamoorthy et al. 2017). We used 100,000 UFB and 100,000 bootstrap replicates of the SH-like approximate likelihood ratio test (SH-aLRT; Guindon et al., 2010) to test the robustness of the inference. Additional applied parameters were -m MFP+ and --sampling GENESITE.

A Bayesian analysis with parallel inference of the species tree and divergence times is computationally expensive and was not feasible with all scHOGs. Therefore, we only used coding sequences of the first 50 partitions (= 90 scHOGs), according to the order determined by the genesortR analysis, for Bayesian tree inference with the StarBeast3 model (v1.1.8; Douglas et al., 2022) of BEAST (v2.7.5; Bouckaert et al., 2019). For the analysis with BEAST, we implemented the substitution models determined to best fit to the 50 partitions by PartitionFinder in the above described analysis. As clock model of the species tree, we used a relaxed clock and estimated its standard deviation and clock rate. As a model of speciation, the calibrated Yule model was used. Estimations of mutation rates in wild *Arabidopsis thaliana* (L.) populations were reported to range between 2 and 5 x10^{-9} site^{-1}year^{-1} (Exposito-Alonso et al. 2018). Even though it is likely that the mutation rates in perennial *Hordeum* species are slightly lower, these estimates are a sufficient proxy for a prior distribution. Therefore, we defined the priors for the clock rates of the species tree and the partitions as a log-normal distribution with a mean (μ) of 0.0035 and a standard deviation of 0.9 (note, these values were transformed to site^{-1} [million years]$^{-1}$). We created three MRCA priors for time calibration. A normal distribution with μ=19.4 and =6 at the split of *P. juncea* and *D. villosum* (Bernhardt et al. 2017), a normal distribution with μ=12 and =3 at the split of *D. villosum* and *Hordeum* (Brassac and Blattner 2015; Bernhardt et al. 2017) and a normal distribution with μ=9.23 and =1.3 at the *Hordeum* root (Bernhardt et al., 2017; Brassac & Blattner, 2015) . Chain lengths were 700 million, storing every 4,000 samples. We used CoupledMCMC (1.2.0; Müller & Bouckaert, 2020) with four chains. All other parameters were left at default. The corresponding xml file can be found in the supplementary material. We ran this analysis three times with different seeds and inspected the log files in Tracer (v1.7.2; Rambaut et al., 2018) to ensure convergence. The three independent runs were combined with Logcombiner (v2.7.6; Rambaut et al., 2018) discarding 10% of the trees of each run as burn-in to create the final collection of tree samples. Treeannotator (v2.7.6; Drummond & Rambaut, 2007) was used to summarize tree samples to a maximum clade credibility tree with 0% burn-in and 0.5 posterior probability limit.

Species' life-history strategies were defined based on (Bothmer et al. 1995) (Tab. S1) after their description was verified by common garden experiments in the Botanical Garden Düsseldorf (Anokye et al. 2025); Longitude: 51.188324, Latitude: 6.800582). We defined annuals as species that senescent within one year after germination while perennial plants survived more than one cycle of seasons and transitioned from vegetative to reproductive growth at least twice. The R package 'ape' was used for this analysis (Paradis and Schliep 2019).

Interspecies hybridization

We extracted all coding sequences from scHOGs in Hordeum which were present in at least two accessions per species or in one accession for the species which were represented by only a single accession. This resulted in a set of 2,909 scHOGs, which in the following will be referred to as the 'two-accession set'. The sequences were aligned and trimmed using the same approach outlined above with the scHOGs present in all accessions.

Phylogenetic network inference was conducted with the maximum pseudo-likelihood approach developed for SNAQ (Solís-Lemus et al. 2017) and implemented in the Julia (v1.10.2; Bezanson et al., 2017) package 'PhyloNetworks' (v0.16.3; Solís-Lemus et al., 2017). Genetrees from the aligned coding sequences of the two-accessions set were estimated with IQtree (v2.2.0; Minh et al., 2020) using the same procedure as outlined above for creating the input of genesortR (Koch 2021). Species quartet concordance factors were calculated from these genetrees. We ran SNAQ 10 independent times with different seeds, allowed reticulations ranging from 0-10 and the species tree as a guide tree. We considered all networks with a pseudo log-likelihood below a certain threshold as candidate networks. This threshold was arbitrary. Yet, in all models, there was a steep increase in pseudo-log-likelihood between networks below and above this threshold, suggesting a clear decrease in statistical support for networks above the threshold (Fig. S5). Reticulations that appeared in at least two networks were considered candidate reticulations (Fig. S6).

For SNP calling, we used the reference genome from the species that was on average the least diverged from all other species. The most suitable reference genome was determined by calculating patristic distances between all accessions from the accessions tree. The species with the lowest average patristic distances to all other species was *H. comosum* (J.Presl), which therefore was used as the common reference genome. Filtered reads from all accessions subjected to RNA sequencing were mapped against the *H. comosum* reference genome (Feng et al. 2025) using STAR (v2.7.10a; Dobin et al., 2013) with default settings. In subsequent filtering steps with SAMtools (v1.3.1; Danecek et al., 2021), only uniquely mapped reads were retained. Variant calling was done with BCFtools (v1.10.2; Danecek et al., 2021) and default parameters. We kept only biallelic SNPs without missing data, QUAL $\geq$ 30, GQ $\geq$ 30, DP $\geq$ 5 and for heterozygous SNP calls DP $\geq$ 30. VCFtools (v0.1.16; Danecek et al., 2011) and BCFtools (v1.10.2; Danecek et al., 2021) were used for variant filtering. The 13 included accessions from Feng et al. (2025) were included by extracting the gene space defined in the gff files of the corresponding species from the respective reference genomes. These sequences were mapped against the *H. comosum* reference genome using Minimap2 with the asm20 option to account for the divergence between species (Li 2018). Only sequences with MAPQ $\geq$ 30 were used to call biallelic, homozygous SNPs with QUAL $\geq$ 30. The two VCF files were merged and the merged file was used for the subsequent procedure.

Linkage disequilibrium (LD) pruning was done with PLINK (v 1.90b7; Purcell et al., 2007). Windows of 50 kb were shifted by five variants, and SNPs with $r^2 \geq 0.5$ within one window were removed. The pruned SNP set was used as input for Dsuite (v0.5r53; Malinsky et al., 2021) to conduct ABBA-BABA tests for candidate reticulations found with SNAQ (Solís-Lemus et al. 2017). In all tests, *Dasypyrum villosum* was the outgroup species. In Dsuite, we used the --ABBAclustering parameter to test whether ABBA patterns in tests with significant D tend to cluster together (Koppetsch et al. 2024).

Biogeography of annual and perennial *Hordeum* species

We downloaded publicly available georeferenced observations of the diploid *Hordeum* species (excluding domesticated barley) included in our phylogeny from GBIF (GBIF.org 2022), the New York Botanical Garden (NANSH; http://sweetgum.nybg.org/science/ih/; accessed 16.08.2022) and the North American Network of Small Herbaria (https://nansh.org/portal/collections/index.php; accessed 16.08.2022). Since public databases of plant occurrences are often contaminated with faulty entries, we restricted data points

of each *Hordeum* species to their geographic distribution as previously described (Bothmer et al. 1995).

Additionally, we selected one random observation from observations that occurred within a 10km radius

to avoid over representation of more frequently monitored areas. Climate variables were downloaded for

all remaining observations from Worldclim 2 (Fick and Hijmans 2017) with a resolution of 2.5 minutes.

Landcover descriptions were retrieved from the European Union's Copernicus Land Monitoring Service

(Land Cover 2019, version 3, 100m raster resolution; Buchhorn et al., 2020). This cleaned data set was

used for the subsequent procedure (Tab. S3). Due to the large variation of data points per species, we cal-

culated the mean of each environmental variable and continued with the species means to avoid biases

due to over- or under-representation of specific species. Because the timing of growth seasons differs

between hemispheres, we classified spring, summer, autumn, and winter according to the conventional

meteorological months for the Northern and Southern Hemispheres, respectively. In the Northern Hemi-

sphere, spring was represented by March-May, summer by June-August, autumn by September-Novem-

ber, and winter by December-February, whereas in the Southern Hemisphere these groupings were shifted

by six months (e.g., spring: September-November). This procedure ensured that seasonal climate

descriptors match major growth cycle patterns across both hemispheres.

Climate niche spaces were approximated by the hypervolume of the climate variables calculated using

dynamic range boxes with geometric means of individual variables implemented in the R package

'dynRB' (v0.18; Junker et al. 2016). This procedure was done for all accessions across annual species and

for all accessions across perennial species. Additionally, hypervolumes were calculated from accessions

of each species separately, except for *H. erectifolium*, where only a single accession is available. Wil-

coxon rank-sum tests were used to test the significance of each climate niche and variable between annu-

als and perennials.

We modeled monthly temperature and precipitation variation between annual and perennial species using

Bayesian generalized additive mixed models (GAMMs) implemented in the R package 'brms' (Bürkner,

2017). Temperature/precipitation was the response variable, and life history (annual vs. perennial) was in-

cluded as a fixed factor. Smooth functions of month were used to capture cyclic seasonal patterns; i.e.

each month was coded numerically (1-12) to represent the annual cycle, with consecutive numbering

from January (winter) to December, and treated as a cyclic variable. This ensured that December and

January were modeled as adjacent months. Life-history-specific smooths allowed the two groups to differ

in their seasonal temperature profiles. The model included a phylogenetic random effect based on the cor-

relation matrix derived from the species tree to account for non-independence due to shared ancestry.

Model convergence was evaluated using standard diagnostics implemented via the 'brms' package. Pos-

terior predictions were generated for all months and both life histories using species-averaged estimates.

For each month, we calculated the posterior probability that the predicted climate value for annual species

exceeded that for perennials, P(annual > perennial), and interpreted these probabilities as measures of

evidence for systematic differences between life histories. Since no formal thresholds exist, we used the

following guideline: values between 0.50 and 0.75 were considered to indicate no evidence, values from

0.75 to 0.90 moderate evidence, and values above 0.90 strong evidence that annuals exceed perennials.

Because the scale is symmetric, probabilities below 0.50 provide evidence in the opposite direction, with

0.50-0.25 indicating no evidence, 0.25-0.10 moderate evidence, and values below 0.10 strong evidence

that perennials exceed annuals.

To quantify interannual climate variability across the habitats of annual and perennial *Hordeum* species,

we obtained monthly precipitation and minimum and maximum temperature data for the period 1960–

2019 (Fick and Hijmans 2017). For each month, the mean of the minimum and maximum temperatures

was used as a summary measure of monthly temperature. Interannual variability in temperature and pre-

cipitation was estimated as the median absolute deviation across years for each month. Bayesian GAMMs

were then fitted, following the same procedures described above, to test for differences in interannual cli-

matic variability between life-history strategies.

Differences in landcover between annuals and perennials were tested using the non-paramtetric phylogen-
etic ANOVA implemented in the R package 'phytools' (v2.5-2) with the species tree to model ancestral
covariance (Revell, 2024).

The entire procedure was done in R (v4.3.3; R CoreTeam, 2021).

As mentioned above, we focused on iteroparous species that continuously maintain above-ground veget-
ative material and, therefore, excluded the pseudo-annual *H. bulbosum* from this analysis.

Gene copy number variation in the context of life-history strategies

For analyses of gene family sizes, we only used protein sequences from all reference genomes published
by Feng et al. (2025) and added six outgroup species: *Arabidopsis thaliana*, *Oryza sativa*, *Zea mays*,
Sorghum bicolor, *Brachypodium distachyon* and *Dasypyrum villosum* (Tab. S4). The protein sequences of
this collection were used as input for OrthoFinder (v2.5.4; Emms & Kelly, 2019) and processed the same
way as outlined above for the protein sequences from the transcriptome assemblies and the selected
accessions from Feng et al. (2025). The resulting gene families will have the prefix "GENO." in the
following to distinguish them from the first OrthoFinder analysis described above.

Single copy HOGs were extracted from the output and used for the same phylogenetic analyses like
described above for the protein sequences from the transcriptome assemblies and the selected accessions
from Feng et al. (2025) except for the species tree inference, i.e. alignment and trimming → creation of
partition → estimating gene properties for phylogenetic inference → accession based phylogenetic
inference. The resulting accession tree will be called 'genome accession tree' hereafter.

The outgroups and all polyploid specieswere removed from the assignment of proteins to gene family
(N0.tsv) done by OrthoFinder (v2.5.4; Emms & Kelly, 2019). We then counted the number of genes of
the remaining diploid *Hordeum* species and filtered this data set. The data set contained seven annual
Hordeum species, and we wanted to reduce the computational expenses by excluding as many gene
families with large variations of presence/absence within annuals or perennials because these were
unlikely to be functionally linked to differences between life history strategies. We allowed 1-species
error margin and, hence, retained only gene families where at least 6 species had a gene count of ≥ 1.

A gene family was considered a candidate if the gene count of the smallest family in an annual species
exceeded the gene count of the largest family in a perennial species or vice versa. The procedure was
done in R (v4.3.3; R Core Team, 2021) using base functions. Gene features including GO-terms of these
final candidate genes were extracted from the *Hordeum comosum* accession (NGB18417) for consistency
with expression and selection analyses (see below). Gene features and GO-terms were extracted from one
representative gene per family. No GO-term enrichment analysis was conducted for copy number
candidates because only eight candidates had GO-term annotatins.

To be comprehensive, we also conducted the analysis a second time with *H. bulbosum* included. Yet,
these results are not discussed and are only presented in the supplementary files. The same procedures
were followed for the analyses of differential expression and selection explained below.

Differential gene expression in the context of life-history strategies

For the analysis of differential gene expression, only diploid *Hordeum* accessions subjected to RNA
sequencing were used. Read counts were derived from the mapped reads using HTSeq (v2.0; Putri et al.,
2022). Gene counts of accessions were sorted according to assignments to scHOGs done by OrthoFinder
(v2.5.4; Emms & Kelly, 2019) for the analysis of gene copy number variation described above. Genes'
tagwise dispersion and size factors (using "TMM") were estimated with the R package 'edgeR' (v4.2.2;
Chen et al. 2025). Significant differential expression between annual and perennial accessions were tested

by fitting a generalized mixed linear model with negative binomial errors and a log link function
(ngGLMM) which was implemented the R package 'glmmSeq' (v.0.5.5; Lewis et al., 2024). The
phylogenetic relationship among accessions was included as random effect variables to account non-
independence of accessions. The phylogenetic relationship was determined by the first three principal
coordinates of the cophenetic distances of the ML based accession tree from the 87 pruned set of
accessions while excluding the 13 accessions from Feng et al.(2025), because they were not subjected to
RNA sequencing. P-values were corrected for multiple testing with the Benjamini-Hochberg method
(Benjamini and Hochberg 1995). Genes were considered differentially expressed (DE) candidates if their
corrected p-value was ≤ 0.01 and their $|\log_2(\text{fold change})|$ was ≥ 1.

Gene features including GO-terms of these final candidate genes were extracted from the *Hordeum*
comosum accession (NGB18417) because it was the only accession that had a sequence in all scHOGs of
the two-accession set and available gene features from Feng et al. (2025). GO-term enrichment analysis
was done for biological processes using a Fischer's exact test with all genes used for the analysis as
background and candidate genes as   genes of interest. The analysis was done in the R package 'topGO'
(v2.56.0; Rahnenfuhrer & Alexa, 2024). GO-terms were considered significant if the p-values were $\leq$
0.05. The entire procedure was done twice, once with all accessions and once without *H. bulbosum*
accessions.

Selection signatures in the context of life-history strategies

Coding sequences from the two-accession set were subjected to codon-aligned sequence alignment using
MACSE (v2.07; Ranwez et al., 2018) with parameter -local_realign_init 1 and -local_realign_dec 1. Each
of these sequences were used as input to CODEML implemented in PAML (v4.10.7; Yang, 2007) to
model D_n/D_s across the accession tree, including only accessions from *Hordeum* species, which allowed
us to control for phylogenetic non-independence. We ran the null model (M_0) and the branch model (M_{alt}),
where accessions from annual species build the foreground branches.

The two-accession set was used to create gene trees with the 'estimatePhangornTreeAll' function with
GTR as model of nucleotide substitution implemented in the R package 'RERconverge' (v0.3.0;
Kowalczyk et al., 2019). These gene trees were then used to test for convergent rate shifts associated with
annual and perennial life histories and annuals as foreground clades, which can indicate repeated
selection. The procedure was done with 'RERconverge'.

The resulting p-values of both methods were corrected for multiple testing with the Benjamini-Hochberg
method (Benjamini and Hochberg 1995). If a gene showed corrected p-values ≤ 0.05 in both methods it
was considered a candidate gene putatively under divergent selection in annual and perennial *Hordeum*
species. Extraction of gene features and GO-term enrichment analysis were done the same way as
described for DE analysis above. The entire procedure was done twice, once with all accessions and once
without *H. bulbosum* accessions.

Data and resource availability

900   Raw data, a selection of intermediate results, supplements and scripts were stored in the DataPlant Hub
901   https://git.nfdi4plants.org/timo.hellwig/evolution_and_divergent_genetic-
902   mechanisms_of_annual_and_perennial_hordeum_species.

Acknowledgements

We gratefully acknowledge computational support from the Zentrum für Informations- und Medientechnologie (ZIM), in particular the HPC team operating the HILBERT high-performance computing cluster at Heinrich-Heine-Universität Düsseldorf.

This work was funded by the European Research Council (ERC) under the European Union's Horizon Europe research and innovation programme (PERLIFE, No. 101002085) and the Deutsche Forschungsgemeinschaft (DFG) under (1) Germany's Excellence Strategy (EXC-2048/1, Project ID: 390686111) and (2) the Collaborative Research Centre/Transregio (TRR 341, Project ID: 456082119).

Authors' Contributions

TH: Conceptualization, Investigation, Methodology, Software, Formal analysis, Visualization, Writing - original draft, Data curation, Project administration. ND: Investigation. EBH: Methodology. MvK: Conceptualization, Funding acquisition, Resources, Writing - review & editing, Project administration, Supervision.

Supplementary Figures

Figure S1: Summary statistics of the read filtering process. Each dot represents one accession.

Figure S2: Schematic representation of the data creation workflow.

Figure S3: Phylogenetic tree of the unpruned set of accessions created by fasttree during the first OrthoFinder analysis. Accessions in red were removed due to their possible mislabeling.

Figure S4: Value of the six gene-partition properties against their sorted position. Order was determined according to their phylogenetic usefulness estimated by genesortR.

Figure S5: Pseudo-log-likelihood of phylogenetic network analysis using SNAQ averaged over repetitions (A) and per per number of reticulations (B-G) with networks below the threshold (green dotted line). Selected candidate networks are depicted in Fig. S6.

Figure S6: Candidate phylogenetic networks (A-H). Coloured lines indicate major (dark blue; $\gamma > 0.5$) and minor parent edges (light blue; $\gamma < 0.5$) of putative hybridization events. Their corresponding numbers represent the inheritance proportion (γ).

Figure S7: Maximum likelihood phylogenetic tree based on previously published genomes (designated as "genome accession tree" in the manuscript). To emphasize the branching order despite highly variable branch-lengths, the tree is rendered as a cladogram.

Figure S8: Species tree with mean (A) and 95% HPD interval (B) of divergence times as node labels.

Figure S9: Climatic niche spaces of individual Hordeum species separated by life-cycle strategy and estimated as n-dimensional hypervolumes.

Figure S10: Results from differential expression analysis of annual vs perennial accessions. Volcano plots (top row), heatmap of candidate genes (middle row; colours correspond to log 10(fold change) and graph of GO-term enrichment analysis (bottom row). Left column with and right column without *H. bulbosum* accessions. GO-terms are coloured according to p-values from yellow (high) to red (low).

Figure S11: Graph of GO-term enrichment analysis with (left column) and without (right column) *H. bulbosum* accessions. GO-terms are coloured according to p-values from yellow (high) to red (low).

Supplementary Tables

Table S1: List of plant material used for transcriptome assemblies.

Table S2: Pan-Hordeum consortion accessions used to complement the detection of orthologs

Table S3: Environmental variables of publicly available Hordeum observations after cleaning.

Table S4: Data sources of reference genomes used for creating the maximum likelihood phylogenetic tree based on previously published genomes(designated as "genome accession tree" in the manuscript; Fig. S7)

Table S5: Summary statistics of scHOG partitions inferred by genesortR.

Table S6: ABBA-BABA test results of selected candidate reticulations

Table S7: List of candidate genes from analyses of divergent genetic mechanisms and their assignment to eight major functional categories.

Table S8: List of orthogroups with output statistics of selection analysis without H. bulbosum accessions

Table S9: Gene copy number of each species and description of one representative H. comosum gene of each gene family that showed copy number variation between annual (red) and perennial (blue) Hordeum species.

Table S10: Members of gene families with divergent gene copy numbers between annual and perennial Hordeum species

Table S11: List of genes included in differential expression analysis without H. bulbosum accessions

Table S12: List of significant GO-terms of genes with significant signatures of selection with and without H. bulbosum accessions

Table S13: List of significant GO-terms of genes with significant signatures of selection with and without H. bulbosum accessions

Table S14: List of orthogroups with output statistics of selection analysis with H. bulbosum accessions

Table S15: List of genes included in differential expression analysis with H. bulbosum accessions

References

Amasino R. 2010. Seasonal and developmental timing of flowering. *The Plant Journal* [Internet] 61:1001–1013. Available from: https://doi.org/10.1111/j.1365-313X.2010.04148.x

Andrews S. 2010. A Quality Control Tool for High Throughput Sequence Data [Online]. *[last accessed 07.08.2025]* [Internet]. Available from: https://www.bioinformatics.babraham.ac.uk/projects/fastqc/

Anokye M, Hellwig T, Haraldsson EB, Schüller R, Döring N, von Korff M. 2025. Trait variations in annual and perennial Hordeum are structured by phylogeny and shaped by life history. *in preparation.*

Arif M, Mahmood N, Nawaz AF, El-Sheikh MA, Alansi S, Ahmad P. 2025. Comprehensive genome-wide characterization of NAC transcription factors in Barley influence insights into stress tolerance and evolutionary dynamics. *Sci Rep* [Internet] 15:24257. Available from: https://doi.org/10.1038/s41598-025-89045-6

Aslam M, Fakher B, Jakada BH, Cao S, Qin Y. 2019. SWR1 Chromatin Remodeling Complex: A Key Transcriptional Regulator in Plants. *Cells* [Internet] 8:1621. Available from: https://doi.org/10.3390/cells8121621

Bahrami F, Arzani A, Rahimmalek M. 2021. Tolerance to high temperature at reproductive stage: Trade-offs between phenology, grain yield and yield-related traits in wild and cultivated barleys. *Plant Breeding* [Internet] 140:812–826. Available from: https://doi.org/10.1111/pbr.12953

Bapteste E, van Iersel L, Janke A, Kelchner S, Kelk S, McInerney JO, Morrison DA, Nakhleh L, Steel M, Stougie L, et al. 2013. Networks: expanding evolutionary thinking. *Trends in Genetics* [Internet] 29:439–441. Available from: https://doi.org/10.1016/j.tig.2013.05.007

Bastow R, Mylne JS, Lister C, Lippman Z, Martienssen RA, Dean C. 2004. Vernalization requires epigenetic silencing of FLC by histone methylation. *Nature* [Internet] 427:164–167. Available from: https://doi.org/10.1038/nature02269

Benjamini Y, Hochberg Y. 1995. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. *Journal of the Royal Statistical Society: Series B (Methodological)* [Internet] 57:289–300. Available from: https://doi.org/10.1111/j.2517-6161.1995.tb02031.x

Bernhardt N, Brassac J, Kilian B, Blattner FR. 2017. Dated tribe-wide whole chloroplast genome phylogeny indicates recurrent hybridizations within Triticeae. *BMC Evolutionary Biology* [Internet] 17:141. Available from: https://doi.org/10.1186/s12862-017-0989-9

Bezanson J, Edelman A, Karpinski S, Shah VB. 2017. Julia: A Fresh Approach to Numerical Computing. *SIAM Rev.* [Internet] 59:65–98. Available from: https://doi.org/10.1137/141000671

Blattner FR. 2004. Phylogenetic analysis of Hordeum (Poaceae) as inferred by nuclear rDNA ITS sequences. *Molecular Phylogenetics and Evolution* [Internet] 33:289–299. Available from: https://doi.org/10.1016/j.ympev.2004.05.012

Blattner FR. 2006. Multiple intercontinental dispersals shaped the distribution area of Hordeum (Poaceae). *New Phytologist* [Internet] 169:603–614. Available from: https://doi.org/10.1111/j.1469-8137.2005.01610.x

Blattner FR. 2009. Progress in phylogenetic analysis and a new infrageneric classification of the barley genus Hordeum (Poaceae: Triticeae). *Breeding Science* [Internet] 59:471–480. Available from: https://doi.org/10.1270/jsbbs.59.471

Boege K, Marquis RJ. 2005. Facing herbivory as you grow up: the ontogeny of resistance in plants. *Trends in Ecology & Evolution* [Internet] 20:441–448. Available from: https://doi.org/10.1016/j.tree.2005.05.001

Böhlenius H, Huang T, Charbonnel-Campaa L, Brunner AM, Jansson S, Strauss SH, Nilsson O. 2006. CO/FT regulatory module controls timing of flowering and seasonal growth cessation in trees. *Science* [Internet] 312:1040–1043. Available from: https://doi.org/10.1126/science.1126038

Bolle C. 2004. The role of GRAS proteins in plant signal transduction and development. *Planta* [Internet] 218:683–692. Available from: https://doi.org/10.1007/s00425-004-1203-z

Bothmer R von, Jacobsen N, Baden C, Jorgensen RB, Linde-Laursen I. 1995. An ecogeographical study of the genus Hordeum (2nd edition). Available from: https://hdl.handle.net/10568/104311

Bothmer RV, Flink J, Jacobsen N, Kotimäki M, Landström TA. 1983. Interspecific hybridization with cultivated barley (Hordeum vulgare L.). *Hereditas* [Internet] 99:219–244. Available from: https://doi.org/10.1111/j.1601-5223.1983.tb00895.x

Bouckaert R, Vaughan TG, Barido-Sottani J, Duchêne S, Fourment M, Gavryushkina A, Heled J, Jones G, Kühnert D, Maio ND, et al. 2019. BEAST 2.5: An advanced software platform for Bayesian evolutionary analysis. *PLOS Computational Biology* [Internet] 15:e1006650. Available from: https://doi.org/10.1371/journal.pcbi.1006650

Boyko JD, Hagen ER, Beaulieu JM, Vasconcelos T. 2023. The evolutionary responses of life-history strategies to climatic variability in flowering plants. *New Phytologist* [Internet] 240:1587–1600. Available from: https://doi.org/10.1111/nph.18971

Brassac J, Blattner FR. 2015. Species-Level Phylogeny and Polyploid Relationships in Hordeum (Poaceae) Inferred by Next-Generation Sequencing and In Silico Cloning of Multiple Nuclear Loci. *Syst Biol* [Internet] 64:792–808. Available from: https://doi.org/10.1093/sysbio/syv035

Buchhorn M, Smets B, Bertels L, De Roo B, Lesiv M, Nandin-Erdene Tsendbazar, Martin Herold, Steffen Fritz. 2020. Copernicus Global Land Service: Land Cover 100m: collection 3: epoch 2019: Globe. Available from: https://zenodo.org/record/3939050

Bürkner P.-C. (2017). brms: An R Package for Bayesian Multilevel Models Using Stan. *Journal of Statistical Software [Internet] 80*(1), 1-28. https://doi.org/10.18637/jss.v080.i01

Bushmanova E, Antipov D, Lapidus A, Prjibelski AD. 2019. rnaSPAdes: a de novo transcriptome assembler and its application to RNA-Seq data. *GigaScience* [Internet] 8:giz100. Available from: https://doi.org/10.1093/gigascience/giz100

Calone R, Mircea D-M, González-Orenga S, Boscaiu M, Lambertini C, Barbanti L, Vicente O. 2022. Recovery from Salinity and Drought Stress in the Perennial Sarcocornia fruticosa vs. the Annual Salicornia europaea and S. veneta. *Plants* [Internet] 11:1058. Available from: https://doi.org/10.3390/plants11081058

Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* [Internet] 10:421. Available from: https://doi.org/10.1186/1471-2105-10-421

Carter B, Bishop B, Ho KK, Huang R, Jia W, Zhang H, Pascuzzi PE, Deal RB, Ogas J. 2018. The Chromatin Remodelers PKL and PIE1 Act in an Epigenetic Pathway That Determines H3K27me3 Homeostasis in Arabidopsis. *The Plant Cell* [Internet] 30:1337–1352. Available from: https://doi.org/10.1105/tpc.17.00867

Chapin FS, Schulze E-D, Mooney HA. 1990. The Ecology and Economics of Storage in Plants. *Annual Review of Ecology and Systematics* [Internet] 21:423–447. Available from: https://www.jstor.org/stable/2097032

Chapman EA, Thomsen HC, Tulloch S, Correia PMP, Luo G, Najafi J, DeHaan LR, Crews TE, Olsson L, Lundquist P-O, et al. 2022. Perennials as Future Grain Crops: Opportunities and Challenges. *Front Plant Sci* [Internet] 13:898769. Available from: https://doi.org/10.3389/fpls.2022.898769

Charnov EL, Schaffer WM. 1973. Life-History Consequences of Natural Selection: Cole's Result Revisited. *The American Naturalist* [Internet] 107:791–793. Available from: https://doi.org/10.1086/282877

Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics* [Internet] 34:i884–i890. Available from: https://doi.org/10.1093/bioinformatics/bty560

Chen Y, Chen L, Lun ATL, Baldoni PL, Smyth GK. 2025. edgeR v4: powerful differential analysis of sequencing data with expanded functionality and improved support for small counts and larger datasets. *Nucleic Acids Res* [Internet] 53:gkaf018. Available from: https://doi.org/10.1093/nar/gkaf018

Choi JY, Platts AE, Fuller DQ, Hsing (邢禹依) Y-I, Wing RA, Purugganan MD. 2017. The Rice Paradox: Multiple Origins but Single Domestication in Asian Rice. *Molecular Biology and Evolution* [Internet] 34:969–979. Available from: https://doi.org/10.1093/molbev/msx049

Christiansen MW, Gregersen PL. 2014. Members of the barley NAC transcription factor gene family show differential co-regulation with senescence-associated genes during senescence of flag leaves. *Journal of Experimental Botany* [Internet] 65:4009–4022. Available from: https://doi.org/10.1093/jxb/eru046

Christiansen MW, Matthewman C, Podzimska-Sroka D, O'Shea C, Lindemose S, Møllegaard NE, Holme IB, Hebelstrup K, Skriver K, Gregersen PL. 2016. Barley plants over-expressing the NAC transcription factor gene HvNAC005 show stunting and delay in development combined with early senescence. *Journal of Experimental Botany* [Internet] 67:5259–5273. Available from: https://doi.org/10.1093/jxb/erw286

Chun I, Kim HJ, Hong S, Kim Y-G, Kim M-S. 2023. Structural basis of DNA binding by the NAC transcription factor ORE1, a master regulator of plant senescence. *Plant Comm* [Internet] 4. Available from: https://doi.org/10.1016/j.xplc.2022.100510

Cole LC. 1954. The Population Consequences of Life History Phenomena. *The Quarterly Review of Biology* [Internet] 29:103–137. Available from: https://doi.org/10.1086/400074

Cornwallis CK, Griffin AS. 2024. A Guided Tour of Phylogenetic Comparative Methods for Studying Trait Evolution. *Annual Review of Ecology, Evolution, and Systematics* [Internet] 55:181–204. Available from: https://www.annualreviews.org/content/journals/10.1146/annurev-ecolsys-102221-050754

Crevillén P, Dean C. 2011. Regulation of the floral repressor gene FLC: the complexity of transcription in a chromatin context. *Current Opinion in Plant Biology* [Internet] 14:38–44. Available from: https://www.sciencedirect.com/science/article/pii/S1369526610001172

Crews TE, Carton W, Olsson L. 2018. Is the future of agriculture perennial? Imperatives and opportunities to reinvent agriculture by shifting from annual monocultures to perennial polycultures. *Global Sustainability* [Internet] 1. Available from: https://doi.org/10.1017/sus.2018.11

Crews TE, Rumsey BE. 2017. What Agriculture Can Learn from Native Ecosystems in Building Soil Organic Matter: A Review. *Sustainability* [Internet] 9:578. Available from: https://doi.org/10.3390/su9040578

Curaba J, Talbot M, Li Z, Helliwell C. 2013. Over-expression of microRNA171 affects phase transitions and floral meristem determinancy in barley. *BMC Plant Biology* [Internet] 13:6. Available from: https://doi.org/10.1186/1471-2229-13-6

Dainat J. 2020. AGAT: Another Gff Analysis Toolkit to handle annotations in any GTF/GFF format. *(Version v0.8.0). Zenodo https://www.doi.org/10.5281/zenodo.3552717.*

Danecek P, Auton A, Abecasis G, Albers CA, Banks E, DePristo MA, Handsaker RE, Lunter G, Marth GT, Sherry ST, et al. 2011. The variant call format and VCFtools. *Bioinformatics* [Internet] 27:2156–2158. Available from: https://doi.org/10.1093%2Fbioinformatics%2Fbtr330

Danecek P, Bonfield JK, Liddle J, Marshall J, Ohan V, Pollard MO, Whitwham A, Keane T, McCarthy SA, Davies RM, et al. 2021. Twelve years of SAMtools and BCFtools. *GigaScience* [Internet] 10:giab008. Available from: https://doi.org/10.1093/gigascience/giab008

De Kort H, Prunier JG, Ducatez S, Honnay O, Baguette M, Stevens VM, Blanchet S. 2021. Life history, climate and biogeography interactively affect worldwide genetic diversity of plant and animal populations. *Nat Commun* [Internet] 12:516. Available from: https://doi.org/10.1038/s41467-021-20958-2

De Souza JG, Da Silva JV. 1987. Partitioning of Carbohydrates in Annual and Perennial Cotton (Gossypium hirsutum L.). *Journal of Experimental Botany* [Internet] 38:1211–1218. Available from: https://www.jstor.org/stable/23691801

DeSalle R, Riley M. 2020. Should Networks Supplant Tree Building? *Microorganisms* [Internet] 8:1179. Available from: https://doi.org/10.3390/microorganisms8081179

Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S, Batut P, Chaisson M, Gingeras TR. 2013. STAR: ultrafast universal RNA-seq aligner. *Bioinformatics* [Internet] 29:15–21. Available from: https://doi.org/10.1093/bioinformatics/bts635

D'Odorico P, Carr JA, Laio F, Ridolfi L, Vandoni S. 2014. Feeding humanity through global food trade. *Earth's Future* [Internet] 2:458–469. Available from: https://doi.org/10.1002/2014EF000250

Doebley J, von Bothmer R, Larson S. 1992. Chloroplast Dna Variation and the Phylogeny of Hordeum (poaceae). *American Journal of Botany* [Internet] 79:576–584. Available from: https://doi.org/10.1002/j.1537-2197.1992.tb14595.x

Dong Z, Xiao Y, Govindarajulu R, Feil R, Siddoway ML, Nielsen T, Lunn JE, Hawkins J, Whipple C, Chuck G. 2019. The regulatory landscape of a core maize domestication module controlling bud dormancy and growth repression. *Nat Commun* [Internet] 10:3810. Available from: https://doi.org/10.1038/s41467-019-11774-w

Douglas J, Jiménez-Silva CL, Bouckaert R. 2022. StarBeast3: Adaptive Parallelized Bayesian Inference under the Multispecies Coalescent. *Systematic Biology* [Internet] 71:901–916. Available from: https://doi.org/10.1093/sysbio/syac010

Drummond AJ, Rambaut A. 2007. BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evolutionary Biology* [Internet] 7:214. Available from: https://doi.org/10.1186/1471-2148-7-214

Duan E, Wang Y, Liu L, Zhu J, Zhong M, Zhang H, Li S, Ding B, Zhang X, Guo X, et al. 2016. Pyrophosphate: fructose-6-phosphate 1-phosphotransferase (PFP) regulates carbon metabolism during grain filling in rice. *Plant Cell Rep* [Internet] 35:1321–1331. Available from: https://doi.org/10.1007/s00299-016-1964-4

Dubos C, Stracke R, Grotewold E, Weisshaar B, Martin C, Lepiniec L. 2010. MYB transcription factors in Arabidopsis. *Trends in Plant Science* [Internet] 15:573–581. Available from: https://doi.org/10.1016/j.tplants.2010.06.005

El-Rabey HA, Badr A, Schäfer-Pregl R, Martin W, Salamini F. 2002. Speciation and Species Separation in Hordeum L. (Poaceae) Resolved by Discontinuous Molecular Markers. *Plant Biol (Stuttg)* [Internet] 4:567–575. Available from: https://doi.org/10.1055/s-2002-35435

Emms DM, Kelly S. 2019. OrthoFinder: phylogenetic orthology inference for comparative genomics. *Genome Biology* [Internet] 20:238. Available from: https://doi.org/10.1186/s13059-019-1832-y

Ewels P, Magnusson M, Lundin S, Käller M. 2016. MultiQC: summarize analysis results for multiple tools and samples in a single report. *Bioinformatics* [Internet] 32:3047–3048. Available from: https://doi.org/10.1093/bioinformatics/btw354

Exposito-Alonso M, Becker C, Schuenemann VJ, Reiter E, Setzer C, Slovak R, Brachi B, Hagmann J, Grimm DG, Chen J, et al. 2018. The rate and potential relevance of new mutations in a colonizing plant lineage. *PLOS Genetics* [Internet] 14:e1007155. Available from: https://journals.plos.org/plosgenetics/article?id=10.1371/journal.pgen.1007155

Fedak G. 1985. Wide Crosses in *Hordeum*. In: Rasmusson DC, editor. Agronomy Monographs. Madison, WI, USA: American Society of Agronomy, Crop Science Society of America, Soil Science Society of America. p. 155–186. Available from: https://doi.org/10.2134/agronmonogr26.c7

Feng J-W, Jiang C, Strejčková B, Szecówka M, Haraldsson E, Lux T, An C, Bayer M, Bertrand YJK, Chudecká M, Corrales SV, Čegan R, Fiebig A, Gao G, Haghi R, Himmelbach A, Houben A, Jin Y, Krak K, Komatsuda T, Kuo Y-T, Mahelka V, Mayer KFX, Melonek J, Pecinka A, Sakuma S, Šimková H, Small I, Spannagl M, Stein N, Toegelová H, Waugh R, Wicker T, Zhang H, Zhu Z, Šafář J, Kopecký D, Blattner FR, Yang P, von Korff M, Mascher M. 2025. A genus-wide pangenome of Hordeum. 28 October 2025. PREPRINT (Version 1). Research Square. Available from: https://doi.org/10.21203/rs.3.rs-7766969/v1

Fick SE, Hijmans RJ. 2017. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology* [Internet] 37:4302–4315. Available from: https://doi.org/10.1002%2Fjoc.5086

Friedman J. 2020. The Evolution of Annual and Perennial Plant Life Histories: Ecological Correlates and Genetic Mechanisms. *Annu. Rev. Ecol. Evol. Syst.* [Internet] 51:461–481. Available from: https://doi.org/10.1146/annurev-ecolsys-110218-024638

Friedman J, Willis JH. 2013. Major QTLs for critical photoperiod and vernalization underlie extensive variation in flowering in the Mimulus guttatus species complex. *New Phytologist* [Internet] 199:571–583. Available from: https://doi.org/10.1111/nph.12260

Fuerst D, Shermeister B, Mandel T, Hübner S. 2023. Evolutionary Conservation and Transcriptome Analyses Attribute Perenniality and Flowering to Day-Length Responsive Genes in Bulbous Barley (Hordeum bulbosum). *Genome Biology and Evolution* [Internet] 15:evac168. Available from: https://doi.org/10.1093/gbe/evac168

Garnier E. 1992. Growth Analysis of Congeneric Annual and Perennial Grass Species. *Journal of Ecology* [Internet] 80:665–675. Available from: https://www.jstor.org/stable/2260858

GBIF.org. 2022. GBIF.org (August , 16) GBF Occurence Download. Available from: https://doi.org/10.15468/dl.9dyt27

Gonzalez-Paleo L, Parmigiani C, Pastor-Pastor A, Vilela AE. 2024. From acquisitive to conservative: The ontogenetic trajectory of root syndrome of annual and perennial co-generic species. *Acta Oecologica* [Internet] 123:103996. Available from: https://doi.org/10.1016/j.actao.2024.103996

Grabherr MG, Haas BJ, Yassour M, Levin JZ, Thompson DA, Amit I, Adiconis X, Fan L, Raychowdhury R, Zeng Q, et al. 2011. Full-length transcriptome assembly from RNA-Seq data without a reference genome. *Nat Biotechnol* [Internet] 29:644–652. Available from: https://doi.org/10.1038/nbt.1883

Grime JP. 1977. Evidence for the Existence of Three Primary Strategies in Plants and Its Relevance to Ecological and Evolutionary Theory. *The American Naturalist* [Internet] 111:1169–1194. Available from: https://doi.org/10.1086/283244

Gruner P, Miedaner T. 2021. Perennial Rye: Genetics of Perenniality and Limited Fertility. *Plants* [Internet] 10:1210. Available from: https://doi.org/10.3390/plants10061210

Guindon S, Dufayard J-F, Lefort V, Anisimova M, Hordijk W, Gascuel O. 2010. New Algorithms and Methods to Estimate Maximum-Likelihood Phylogenies: Assessing the Performance of PhyML 3.0. *Systematic Biology* [Internet] 59:307–321. Available from: https://doi.org/10.1093/sysbio/syq010

Haas B. 2024. Transdecoder. *Available online at: https://github.com/TransDecoder/TransDecoder [last accessed: 07.08.2025].*

Heidel AJ, Kiefer C, Coupland G, Rose LE. 2016. Pinpointing genes underlying annual/perennial transitions with comparative genomics. *BMC Genomics* [Internet] 17:921. Available from: https://doi.org/10.1186/s12864-016-3274-1

Hepworth J, Dean C. 2015. Flowering Locus C's Lessons: Conserved Chromatin Switches Underpinning Developmental Timing and Adaptation1. *Plant Physiol* [Internet] 168:1237–1245. Available from: https://doi.org/10.1104/pp.15.00496

Herron SA, Rubin MJ, Ciotir C, Crews TE, Van Tassel DL, Miller AJ. 2020. Comparative Analysis of Early Life Stage Traits in Annual and Perennial Phaseolus Crops and Their Wild Relatives. *Front. Plant Sci.* [Internet] 11. Available from: https://doi.org/10.3389/fpls.2020.00034

Hibbins MS, Hahn MW. 2022. Phylogenomic approaches to detecting and characterizing introgression. *Genetics* [Internet] 220:iyab173. Available from: https://doi.org/10.1093/genetics/iyab173

Hjertaas AC, Preston JC, Kainulainen K, Humphreys AM, Fjellheim S. 2023. Convergent evolution of the annual life history syndrome from perennial ancestors. *Front. Plant Sci.* [Internet] 13. Available from: https://doi.org/10.3389/fpls.2022.1048656

Hu F, Wang D, Zhao X, Zhang T, Sun H, Zhu L, Zhang F, Li L, Li Q, Tao D, et al. 2011. Identification of rhizome-specific genes by genome-wide differential expression Analysis in Oryza longistaminata. *BMC Plant Biology* [Internet] 11:18. Available from: https://doi.org/10.1186/1471-2229-11-18

Hu FY, Tao DY, Sacks E, Fu BY, Xu P, Li J, Yang Y, McNally K, Khush GS, Paterson AH, et al. 2003. Convergent evolution of perenniality in rice and sorghum. *Proceedings of the National Academy of Sciences* [Internet] 100:4050–4054. Available from: https://doi.org/10.1073/pnas.0630531100

Jagadish SVK. 2020. Heat stress during flowering in cereals – effects and adaptation strategies. *New Phytologist* [Internet] 226:1567–1572. Available from: https://doi.org/10.1111/nph.16429

Jakob SS, Blattner FR. 2006. A Chloroplast Genealogy of Hordeum (Poaceae): Long-Term Persisting Haplotypes, Incomplete Lineage Sorting, Regional Extinction, and the Consequences for Phylogenetic Inference. *Molecular Biology and Evolution* [Internet] 23:1602–1612. Available from: https://doi.org/10.1093/molbev/msl018

Jin Y, Du X, Jiang C, Ji W, Yang P. 2024. Disentangling sources of gene tree discordance for Hordeum species via target-enriched sequencing assays. *Molecular Phylogenetics and Evolution* [Internet] 199:108160. Available from: https://doi.org/10.1016/j.ympev.2024.108160

Jörgensen RB. 1986. Relationships in the barley genus (Hordeum): An electrophoretic examination of proteins. *Hereditas* [Internet] 104:273–291. Available from: https://doi.org/10.1111/j.1601-5223.1986.tb00541.x

Junker RR, Kuppler J, Bathke AC, Schreyer ML, Trutschnig W. 2016. Dynamic range boxes – a robust nonparametric approach to quantify size and overlap of n-dimensional hypervolumes. *Methods in Ecology and Evolution* [Internet] 7:1503–1513. Available from: https://doi.org/10.1111/2041-210X.12611

Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermiin LS. 2017. ModelFinder: fast model selection for accurate phylogenetic estimates. *Nat Methods* [Internet] 14:587–589. Available from: https://doi.org/10.1038/nmeth.4285

Katoh K, Misawa K, Kuma K, Miyata T. 2002. MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Research* [Internet] 30:3059–3066. Available from: https://doi.org/10.1093/nar/gkf436

Kebrom TH, Brutnell TP, Hays DB, Finlayson SA. 2010. Vegetative axillary bud dormancy induced by shade and defoliation signals in the grasses. *Plant Signal Behav* [Internet] 5:317–319. Available from: https://doi.org/10.4161/psb.5.3.11186

Kiefer C, Severing E, Karl R, Bergonzi S, Koch M, Tresch A, Coupland G. 2017. Divergence of annual and perennial species in the Brassicaceae and the contribution of cis-acting variation at FLC orthologues. *Molecular Ecology* [Internet] 26:3437–3457. Available from: https://doi.org/10.1111/mec.14084

Koch NM. 2021. Phylogenomic Subsampling and the Search for Phylogenetically Reliable Loci. *Molecular Biology and Evolution* [Internet] 38:4025–4038. Available from: https://doi.org/10.1093/molbev/msab151

Koller D, Highkin HR. 1960. Environmental Control of Reproductive Development in Hordeum Bulbosum, a Perennial Pasture Grass. *American Journal of Botany* [Internet] 47:843–847. Available from: https://doi.org/10.1002/j.1537-2197.1960.tb07174.x

Koppetsch T, Malinsky M, Matschiner M. 2024. Towards Reliable Detection of Introgression in the Presence of Among-Species Rate Variation. *Systematic Biology* [Internet] 73:769–788. Available from: https://doi.org/10.1093/sysbio/syae028

Kowalczyk A, Meyer WK, Partha R, Mao W, Clark NL, Chikina M. 2019. RERconverge: an R package for associating evolutionary rates with convergent traits. *Bioinformatics* [Internet] 35:4815–4817. Available from: https://doi.org/10.1093/bioinformatics/btz468

Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B. 2017. PartitionFinder 2: New Methods for Selecting Partitioned Models of Evolution for Molecular and Morphological Phylogenetic Analyses. *Molecular Biology and Evolution* [Internet] 34:772–773. Available from: https://doi.org/10.1093/molbev/msw260

Lewis M, Goldmann K, Sciacca E. 2024. glmmSeq: General Linear Mixed Models for Gene-Level Differential Expression. Available from: https://myles-lewis.github.io/glmmSeq/

Li H. 2018. Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics* [Internet] 34:3094–3100. Available from: https://doi.org/10.1093/bioinformatics/bty191

Li Z, Lathe RS, Li J, He H, Bhalerao RP. 2022. Towards understanding the biological foundations of perenniality. *Trends in Plant Science* [Internet] 27:56–68. Available from: https://doi.org/10.1016/j.tplants.2021.08.007

Li Z, Ma R, Bhalerao RP. 2024. Perenniality: the tale of three MADS-box genes. *Cell Res* [Internet] 34:753–754. Available from: https://doi.org/10.1038/s41422-024-01001-7

Loka D, Harper J, Humphreys M, Gasior D, Wootton-Beard P, Gwynn-Jones D, Scullion J, Doonan J, Kingston-Smith A, Dodd R, et al. 2019. Impacts of abiotic stresses on the physiology and metabolism of cool-season grasses: A review. *Food and Energy Security* [Internet] 8:e00152. Available from: https://doi.org/10.1002/fes3.152

Lundgren MR, Des Marais DL. 2020. Life History Variation as a Model for Understanding Trade-Offs in Plant–Environment Interactions. *Current Biology* [Internet] 30:R180–R189. Available from: https://doi.org/10.1016/j.cub.2020.01.003

Ma Z, Hu L, Jiang W. 2024. Understanding AP2/ERF Transcription Factor Responses and Tolerance to Various Abiotic Stresses in Plants: A Comprehensive Review. *International Journal of Molecular Sciences* [Internet] 25:893. Available from: https://doi.org/10.3390/ijms25020893

Malinsky M, Matschiner M, Svardal H. 2021. Dsuite - Fast D-statistics and related admixture evidence from VCF files. *Molecular Ecology Resources* [Internet] 21:584–595. Available from: https://doi.org/10.1111/1755-0998.13265

Marillia EF, Scoles GJ. 1996. The use of RAPD markers in Hordeum phylogeny. *Genome* [Internet] 39:646–654. Available from: https://doi.org/10.1139/g96-082

Mascher M, Wicker T, Jenkins J, Plott C, Lux T, Koh CS, Ens J, Gundlach H, Boston LB, Tulpová Z, et al. 2021. Long-read sequence assembly: a technical evaluation in barley. *The Plant Cell* [Internet] 33:1888–1906. Available from: https://doi.org/10.1093/plcell/koab077

Mason-Gamer RJ, White DM. 2024. The phylogeny of the Triticeae: Resolution and phylogenetic conflict based on genomewide nuclear loci. *American Journal of Botany* [Internet] 111:e16404. Available from: https://doi.org/10.1002/ajb2.16404

Melzer S, Lens F, Gennen J, Vanneste S, Rohde A, Beeckman T. 2008. Flowering-time genes modulate meristem determinacy and growth form in Arabidopsis thaliana. *Nat Genet* [Internet] 40:1489–1492. Available from: https://doi.org/10.1038/ng.253

van der Merwe MJ, Groenewald J-H, Stitt M, Kossmann J, Botha FC. 2010. Downregulation of pyrophosphate: d-fructose-6-phosphate 1-phosphotransferase activity in sugarcane culms enhances sucrose accumulation due to elevated hexose-phosphate levels. *Planta* [Internet] 231:595–608. Available from: https://doi.org/10.1007/s00425-009-1069-1

Minh BQ, Schmidt HA, Chernomor O, Schrempf D, Woodhams MD, von Haeseler A, Lanfear R. 2020. IQ-TREE 2: New Models and Efficient Methods for Phylogenetic Inference in the Genomic Era. *Molecular Biology and Evolution* [Internet] 37:1530–1534. Available from: https://doi.org/10.1093/molbev/msaa015

Mishra RC, Grover A. 2016. ClpB/Hsp100 proteins and heat stress tolerance in plants. *Critical Reviews in Biotechnology* [Internet] 36:862–874. Available from: https://doi.org/10.3109/07388551.2015.1051942

Morrison DA. 2005. Networks in phylogenetic analysis: new tools for population biology. *International Journal for Parasitology* [Internet] 35:567–582. Available from: https://doi.org/10.1016/j.ijpara.2005.02.007

Müller NF, Bouckaert RR. 2020. Adaptive Metropolis-coupled MCMC for BEAST 2. *PeerJ* [Internet] 8:e9473. Available from: https://doi.org/10.7717/peerj.9473

Paradis E, Schliep K. 2019. ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics* [Internet] 35:526–528. Available from: https://doi.org/10.1093/bioinformatics/bty633

Petersen G, Seberg O. 1998. Molecular Studies on the Phylogeny of the Genus Barley (Hordeum; Poaceae). In: Karp A, Isaac PG, Ingram DS, editors. Molecular Tools for Screening Biodiversity: Plants and Animals. Dordrecht: Springer Netherlands. p. 437–440. Available from: https://doi.org/10.1007/978-94-009-0019-6_78

Petersen G, Seberg O. 2003. Phylogenetic Analyses of the Diploid Species of Hordeum (Poaceae) and a Revised Classification of the Genus. *sbot* [Internet] 28:293–306. Available from: https://doi.org/10.1043/0363-6445-28.2.293

Poppenwimer T, Mayrose I, DeMalach N. 2023. Revising the global biogeography of annual and perennial plants. *Nature* [Internet] 624:109–114. Available from: https://doi.org/10.1038/

Prjibelski A, Antipov D, Meleshko D, Lapidus A, Korobeynikov A. 2020. Using SPAdes De Novo Assembler. *Current Protocols in Bioinformatics* [Internet] 70:e102. Available from: https://doi.org/10.1002/cpbi.102

Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MAR, Bender D, Maller J, Sklar P, Bakker PIW de, Daly MJ, et al. 2007. PLINK: A Tool Set for Whole-Genome Association and Population-Based Linkage Analyses. *The American Journal of Human Genetics* [Internet] 81:559–575. Available from: https://doi.org/10.1086%2F519795

Putri GH, Anders S, Pyl PT, Pimanda JE, Zanini F. 2022. Analysing high-throughput sequencing data in Python with HTSeq 2.0. *Bioinformatics* [Internet] 38:2943–2945. Available from: https://doi.org/10.1093/bioinformatics/btac166

R. Core T. 2021. R: A Language and Environment for Statistical Computing. *R Foundation for Statistical Computing, Vienna, Austria* [Internet]. Available from: https://www.R-project.org/

Rahnenfuhrer J, Alexa A. 2024. topGO: Enrichment Analysis for Gene Ontology. Available from: https://doi.org/10.18129/B9.bioc.topGO

Rajhathy T, Morrison JW. 1962. Cytogenetic studies in the genus hordeum vi. the murinum-complex. *Can. J. Genet. Cytol.* [Internet] 4:240–247. Available from: https://doi.org/10.1139/g62-029

Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA. 2018. Posterior Summarization in Bayesian Phylogenetics Using Tracer 1.7. *Systematic Biology* [Internet] 67:901–904. Available from: https://doi.org/10.1093/sysbio/syy032

Ramsay NA, Glover BJ. 2005. MYB–bHLH–WD40 protein complex and the evolution of cellular diversity. *Trends in Plant Science* [Internet] 10:63–70. Available from: https://doi.org/10.1016/j.tplants.2004.12.011

Ranwez V, Douzery EJP, Cambon C, Chantret N, Delsuc F. 2018. MACSE v2: Toolkit for the Alignment of Coding Sequences Accounting for Frameshifts and Stop Codons. *Molecular Biology and Evolution* [Internet] 35:2582–2584. Available from: https://doi.org/10.1093/molbev/msy159

Revell LJ. 2024. phytools 2.0: an updated R ecosystem for phylogenetic comparative methods (and other things). *PeerJ [Internet]* 12:e16505 https://doi.org/10.7717/peerj.16505

Ross J, Li Y, Lim E-K, Bowles DJ. 2001. Higher plant glycosyltransferases. *Genome Biology* [Internet] 2:reviews3004.1. Available from: https://doi.org/10.1186/gb-2001-2-2-reviews3004

Shang Y, Yuan L, Di Z, Jia Y, Zhang Z, Li S, Xing L, Qi Z, Wang X, Zhu J, et al. 2020. A CYC/TB1-type TCP transcription factor controls spikelet meristem identity in barley. *Journal of Experimental Botany* [Internet] 71:7118–7131. Available from: https://doi.org/10.1093/jxb/eraa416

Solís-Lemus C, Bastide P, Ané C. 2017. PhyloNetworks: A Package for Phylogenetic Networks. *Molecular Biology and Evolution* [Internet] 34:3292–3298. Available from: https://doi.org/10.1093/molbev/msx235

Soltis DE, Soltis PS. 2003. The Role of Phylogenetics in Comparative Genetics. *Plant Physiol* [Internet] 132:1790–1800. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC526274/

Stearns SC. 1992. The Evolution of Life Histories. Oxford, New York: Oxford University Press

Steenwyk JL, Iii TJB, Li Y, Shen X-X, Rokas A. 2020. ClipKIT: A multiple sequence alignment trimming software for accurate phylogenomic inference. *PLOS Biology* [Internet] 18:e3001007. Available from: https://doi.org/10.1371/journal.pbio.3001007

Stein O, Granot D. 2019. An Overview of Sucrose Synthases in Plants. *Front. Plant Sci.* [Internet] 10. Available from: https://doi.org/10.3389/fpls.2019.00095

Sung S, Amasino RM. 2004. Vernalization in Arabidopsis thaliana is mediated by the PHD finger protein VIN3. *Nature* [Internet] 427:159–164. Available from: https://doi.org/10.1038/nature02195

Svitashev S, Bryngelsson T, Vershinin A, Pedersen C, Säll T, von Bothmer R. 1994. Phylogenetic analysis of the genus Hordeum using repetitive DNA sequences. *Theoret. Appl. Genetics* [Internet] 89:801–810. Available from: https://doi.org/10.1007/BF00224500

Vallebona C, Mantino A, Bonari E. 2016. Exploring the potential of perennial crops in reducing soil erosion: A GIS-based scenario analysis in southern Tuscany, Italy. *Applied Geography* [Internet] 66:119–131. Available from: https://doi.org/10.1016/j.apgeog.2015.11.015

Wang H, Seiler C, Sreenivasulu N, von Wirén N, Kuhlmann M. 2022. INTERMEDIUM-C mediates the shade-induced bud growth arrest in barley. *Journal of Experimental Botany* [Internet] 73:1963–1977. Available from: https://doi.org/10.1093/jxb/erab542

Wang R, Farrona S, Vincent C, Joecker A, Schoof H, Turck F, Alonso-Blanco C, Coupland G, Albani MC. 2009. PEP1 regulates perennial flowering in Arabis alpina. *Nature* [Internet] 459:423–427. Available from: https://doi.org/10.1038/nature07988

Xing J-X, Zang Q-L, Ye Z-L, Qi L-W, Yang L, Li W-F. 2024. Overexpression of Larch SCL6 Inhibits Transitions from Vegetative Meristem to Inflorescence and Flower Meristem in Arabidopsis thaliana (L.) Heynh. *Plants* [Internet] 13:1232. Available from: https://doi.org/10.3390/plants13091232

Yang Z. 2007. PAML 4: Phylogenetic Analysis by Maximum Likelihood. *Molecular Biology and Evolution* [Internet] 24:1586–1591. Available from: https://doi.org/10.1093/molbev/msm088

Zhai D, Zhang L-Y, Li L-Z, Xu Z-G, Liu X-L, Shang G-D, Zhao B, Gao J, Wang F-X, Wang J-W. 2024. Reciprocal conversion between annual and polycarpic perennial flowering behavior in the Brassicaceae. *Cell* [Internet] 187:3319-3337.e18. Available from: https://doi.org/10.1016/j.cell.2024.04.047

Zhang S, Huang G, Zhang Y, Lv X, Wan K, Liang J, Feng Y, Dao J, Wu S, Zhang L, et al. 2023. Sustained productivity and agronomic potential of perennial rice. *Nat Sustain* [Internet] 6:28–38. Available from: https://doi.org/10.1038/s41893-022-00997-3

Zhang X, Wang H, Sun H, Li Y, Feng Y, Jiao C, Li M, Song X, Wang T, Wang Z, et al. 2023. A chromosome-scale genome assembly of Dasypyrum villosum provides insights into its application as a broad-spectrum disease resistance resource for wheat improvement. *Molecular Plant* [Internet] 16:432–451. Available from: https://doi.org/10.1016/j.molp.2022.12.021

Zhao B, Wang J-W. 2024. Perenniality: From model plants to applications in agriculture. *Molecular Plant* [Internet] 17:141–157. Available from: https://doi.org/10.1016/j.molp.2023.12.011