Title: History of divergence and gene flow shaping geographic variation in Andean warblers (*Myioborus*)

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Abstract

Studying how genetic variation is structured across space, and how it relates to divergence in

phenotypic traits relevant to reproductive isolation, is important for our overall understanding of

the speciation process. We used reduced-representation genomic data (ddRAD-seq) to examine

patterns of genetic variation across the full distribution of an Andean warbler species complex

(*Myioborus ornatus–melanocephalus)*, which includes a known hybrid zone between two

strikingly different plumage forms. Genetic structure largely reflects geographic variation in

head plumage, some of which corresponds to major topographic barriers in the Andes. We also

found evidence of isolation by distance shaping genetic patterns across the group's broad

latitudinal range. We found that *chrysops* and *bairdi*, two taxa with marked plumage differences

that have a known hybrid zone, were characterized by low overall genetic divergence. Based on

our cline analyses of both plumage and genomic hybrid indices, this hybrid zone extends for

approximately 250 km, where advanced generation hybrids are likely most common. We also

identified a slight difference in the centers of the plumage and genomic clines, potentially

suggesting the asymmetric introgression of *chrysops*-like plumage traits. By studying genetic

variation in a phenotypically complex group distributed across a topographically complex area,

which includes a hybrid zone, we were able to show how both geographic features and

potentially sexually selected plumage traits may play a role in species formation in tropical

mountains

Keywords: *phylogeography, hybrid zone, reduced representation genomic data, geographic*

variation, Parulidae.

Introduction

Geographic variation in traits relevant to reproductive isolation is often viewed as a snapshot of

the early stages of population divergence (Mayr, 1942; Price, 2008), making it a focus for

understanding the process of speciation. To study this process, it is useful to evaluate how

phenotypic variation in such traits is related to the underlying population genetic structure

(Edwards et al., 2016; Zamudio et al., 2016) A common expectation is that phenotypic and

genetic divergence are positively correlated: in the absence of strong divergent selection,

extended periods of isolation are typically required for phenotypic differentiation to evolve

(Nosil, 2008; Price, 2008; Winger & Bates, 2015). It follows that range disjunctions associated

with topographic and ecological barriers likely to have restricted historical gene flow are

expected to be accompanied by genetic and phenotypic discontinuities (Edwards et al., 2016).

However, geographic variation in phenotypic traits is not always concordant with genetic

structure. This discordance could be driven, for example, by strong selection on some traits

across environmental gradients over short time scales (Walsh et al., 2017), or complex histories

of selection and gene flow leading to phenotypic convergence (Márquez et al., 2020). A better

understanding of how species diversify across space requires studying such histories of isolation,

and how these relate to divergence in traits, particularly those that might be important for mate

choice and, consequently, reproductive isolation.

Patterns of plumage differentiation—thought to be important for reproductive isolation in

birds (Price, 2008; Uy et al., 2018)— among allopatric populations within species are pervasive

among Andean birds (Graves, 1988; Remsen, 1984). Although geographic variation in plumage

and genetic structure in Andean birds is often associated with current topographic and ecological

barriers (Gutiérrez-Pinto et al., 2012; Prieto-Torres et al., 2018), many of these barriers tend to

be permeable and unstable. Dispersal over such barriers could potentially lead to introgressive

hybridization that connecting otherwise diverging populations (Winger, 2017). Additionally,

differentiated populations can come into renewed contact when the barriers themselves change

over time. In the particular case of the Andes, climatic fluctuations (Ramírez-Barahona &

Eguiarte, 2013) and, potentially, volcanic activity (Sanín et al., 2024) have promoted cycles of

fragmentation and reconnection of elevational vegetation belts, creating opportunities for contact

among periodically isolated populations (Vuilleumier, 1969). Examining these complex histories

of divergence and gene flow in geographically variable and widely distributed Andean taxa

contributes to a more comprehensive understanding of how bird species are formed and

maintained in this biodiversity hotspot (Rahbek et al., 2019; Sonne et al., 2022).

Here, we used a reduced representation genomic data set from an Andean bird species

complex of *Myioborus* warblers (Parulidae) characterized by striking geographic variation in

head color patterns to gain insights into differentiation in geographically isolated populations and

the processes that occur after differentiated populations come into secondary contact. This young

(<1 my; Cuervo, 2013) species complex inhabits the high Andes from Bolivia to Venezuela, and

is characterized by differences in head color patterns despite shallow genetic differentiation in

mitochondrial DNA (Céspedes-Arias et al., 2021; Pérez-Emán, 2005). Seven plumage groups

have been described as subspecies belonging to two species, *Myioborus ornatus* and *M.*

melanocephalus, distributed mostly allopatrically along different sections of the Andes (Cuervo

& Céspedes Arias, 2023; Zimmer, 1949). According to this classification scheme, *M. ornatus*

includes subspecies *ornatus* and *chrysops*, and *M. melanocephalus* includes subspecies *bairdi,*

griseonuchus, malaris, melanocephalus and *bolivianus*. Recent taxonomic revisions, however,

suggest that a four-species classification scheme might better capture what is known about

genetic and plumage variation in this complex (Cuervo & Céspedes Arias, 2023). Regardless of

the species delimitation scheme, analyses based on the ND2 mitochondrial gene have revealed

extensive haplotype sharing among groups with conspicuous plumage differences, potentially

suggesting a history of rapid plumage evolution or repeated mitochondrial introgression

(Céspedes-Arias et al., 2021).

Our previous work in regions where the ranges of *chrysops* and *bairdi* abut, on the border

between Colombia and Ecuador, revealed that these two taxa hybridize widely across more than

200 km where individuals with a variety of intermediate head color patterns occur (Céspedes-

Arias et al., 2021). Although hybridization there has been extensive, the existence of a sharp

plumage cline, limited to a relatively narrow zone within these taxa's broader ranges (20–30%),

suggests the action of some form of selection maintaining their differentiation (Barton & Hewitt,

1985; Brelsford & Irwin, 2009; Harrison, 1986). Here, we leveraged this previous systematic

collection and detailed plumage characterization of the *chrysops-bairdi* hybrid zone (Céspedes-

Arias et al., 2021) to gain a deeper understanding of hybridization dynamics using genomic data.

We examined patterns of genomic variation within this warbler species complex at two

different spatial scales: the entire distribution of the species complex, which corresponds to a

broad latitudinal band in the Andes (approximately 3800 km in geographic distance), and the

hybrid zone existing between two markedly different plumage groups as outlined above. By

focusing on these two different scales, we can address questions related to multiple processes

relevant to the formation and maintenance of species. First, leveraging a broad geographic scale

sampling of individuals representing all plumage forms in the complex, we asked: (1) How is

genetic structure related to geographic distance, topography, and plumage variation along the

Andes? At a finer spatial scale, in the hybrid zone between two plumage forms, we asked: (2.1)

Does the geographic pattern of introgression mirror plumage variation across the hybrid zone?

And (2.2) is there evidence for non-neutral processes such as selection against hybrids based on

patterns of genomic variation? Our genome-wide data coupled with dense geographic sampling

substantially improves our ability to reconstruct and understand the diversification history of

these colorful montane warblers, which was previously limited to studies using only a single

mitochondrial gene.

Methods

Study species and sampling

We used tissue samples (*n*=227, Figure 1) collected across the distribution of the *M. ornatus-melanocephalus* complex, by us (Céspedes-Arias et al., 2021), or previously deposited in museum collections (Supplementary Table 1). Samples encompassed all (seven) currently taxonomically recognized plumage forms and a denser sampling in the *bairdi-chrysops* hybrid zone area, along the border between Colombia and Ecuador (Figure 1A). We also obtained data for eight individuals as outgroups corresponding to three other species in *Myioborus*: *M. pictus* (n = 3), *M. miniatus* (n = 3) and *M. brunniceps* (n = 2) which vary in their phylogenetic and geographic proximity to the *M. ornatus-melanocephalus* complex (Lovette et al., 2010; Pérez-Emán, 2005).

The more conspicous differences among plumage groups in the *ornatus-melanocephalus* complex correspond to the presence/absence and extent of colored plumage patches in the head, as illustrated in Figure 1B. We note that there are different degrees of plumage differentiation and diagnosability based on our own assessments (Cuervo & Céspedes Arias, 2023). For example, the subspecies *malaris, bolivianus* and *melanocephalus* are very similar in plumage, and some of the described diagnostic traits do not seem to vary in a way that is discrete and corresponds with these groups (Cuervo & Céspedes Arias, 2023).

We also note that the geographic limits of some groups are also not clearly defined, specifically of the *bairdi* and *griseonuchus* groups which have only subtle differences in in the amount of yellow vs black on the sides of the face, as well as the presence and extent of a black nuchal band behind the rufous crown (Chapman, 1927; Cuervo & Céspedes Arias, 2023; Zimmer, 1949). Although birds from northern Peru (Dept. of Cajamarca) and central Ecuador are clearly distinct (Supplementary Figure 1), it is unclear whether a phenotypic transition is discrete

or clinal across intervening southern Ecuador and there is a lack of sampling across this area to

evaluate these possibilities. However, to inform our discussion on patterns of genetic variation,

we inspected most of the available specimens collected in this area, either directly or through

photographs, to describe variation in these two plumage traits (Supplementary Figure 1).

Laboratory procedures

DNA extraction and library preparation

We extracted genomic DNA from pectoral muscle (n=232) and blood samples (n=3) employing

a modified phenol-chlorophorm protocol (Sambrook, 1987), using phase-lock tubes. Each DNA

extraction was subsequently purified using homemade Sera-Mag Magnetic Speed-beads for

magnetic nucleic acid extraction (MagNA). To obtain reduced representation genomic data, we

used double-digest restriction site-associated DNA sequencing (ddRAD) following Peterson et

al. (2012), and using modifications from Thrasher et al. (2018). The genomic libraries were then

sequenced on one HiSeq2500 Illumina lane (100 bp SE) by the Genomics Core Facility of

Cornell University.

Bioinformatics pipeline

Quality assessment, filtering and demultiplexing

We assessed sequence quality using FastQC (Andrews, 2010). We used the *process_radtags*

function implemented in STACKS 2.66 (Rochette et al., 2019) to demultiplex the reads and

apply quality filters. Specifically, we filtered out all reads with uncalled bases (--*clean*), those

with low phred quality scores *(--quality*, *--score-limit* 20) and those that did not pass the Illumina

chastity/purity filter (--*filter-illumina*). After demultiplexing, we checked the number of reads

available per individual, and only kept those samples with 200,000 or more reads

(Supplementary Figure 2). Using this threshold, we discarded 11 individuals, mostly

corresponding to the *ornatus* group and tissues collected around ten years prior to this study

(range of number of reads for these individuals = 960–150,152). Despite discarding these

samples, 23 high-quality samples for the *ornatus* group remained in downstream analyses. After

excluding these individuals from downstream analyses, we kept a total of 242,204,894 reads

corresponding to 224 individuals (three represented twice in the library).

Integrated assembly and SNP calling

After demultiplexing, we assembled the RAD reads *de-novo* using functions implemented in

STACKS 2.66 (Rochette et al., 2019), and later integrated position information for the loci in the

catalog from a reference genome (Paris et al., 2017; Rivera-Colón & Catchen, 2022). This

integrated approach performs better than directly mapping reads onto a reference genome in

some scenarios, for example, if the reference genome is relatively distant to the study species as

is our case (Paris et al., 2017). We used the chromosome-level assembly of the *Setophaga*

coronata genome (Baiz et al., 2021) as reference, the highest quality available genome within

Parulidae. The time to the most recent common ancestor between *Setophaga* and *Myioborus* is

estimated between 5–7 mya (Barker et al., 2015).

Optimization of de novo assembly parameters

Before assembling the reads, we conducted an optimization of the parameters *M* and *n* used by

denovo_map.pl (Rochette et al., 2019)*,* for three different assemblies: one including

representatives from the outgroups, one that only included individuals within the *ornatus-*

melanocephalus complex, and one including only samples relevant for the hybrid zone analyses

(*chrysops*, individuals in the hybrid zone, *bairdi,* and *griseonuchus*, Figure 1) (see

Supplementary Material and Supplementary Table 2 for details).

Full de novo assembly run

We then ran *denovo_map.pl* for the entire dataset with the chosen parameter values, using again

the three population maps (n = 224 full assembly, n = 216 ingroup only assembly, and n = 123

for the hybrid zone assembly). For the population maps corresponding to the three different

assemblies, we classified individuals into one of the following groups: *ornatus*, *chrysops*,

"hybrid zone", *bairdi, griseonuchus, malaris, melanocephalus*, and *bolivianus* (populations_2

column in Supplementary Table 1), plus *pictus, miniatus*, and *brunniceps* for the assembly

including the outgroups. For the population map purposes the hybrid zone group corresponds to

a geographic group encompassing populations in and near the hybrid zone, irrespective of the

plumage phenotype of the specimens (see the map in Figure 1).

183   After the assemblies were completed, we assessed the per sample coverage and

percentage of reads incorporated (Rivera-Colón & Catchen, 2022). The assembly corresponding

to only individuals within the *melanocephalus-ornatus* complex resulted in a catalog of 91,650

loci, with a mean effective coverage per sample of 59.1x (SD = 13.2x, min = 18.3x, max = 97.4).

The "hybrid zone" assembly resulted in a catalog of 70,281 loci, with a mean effective coverage

per sample of 61.6x (SD =12.1x, min = 22.5x, max = 99.4x). The assembly including outgroups

resulted in a catalog of 97,533 loci, with a mean effective coverage per sample of 59.3x (SD =

13.2x, min = 18.1x, max = 98.5x). Four individuals with a mean effective coverage lower than

20x in assemblies in which they were included initially were excluded from downstream

analyses (see Supplementary Table 1; Supplementary Figure 3).

Mapping to the reference genome

After obtaining the *de novo* catalog, we mapped loci to the reference genome by first indexing

the *S. coronata* genome using *bwa index* (Li & Durbin, 2009). Then we used *bwa mem* to align

the *de novo* catalog to the reference genome. Finally, we used the *stacks-integrate-alignments*

script to generate a catalog with position information (Rivera-Colón & Catchen, 2022; Rochette

et al., 2019). After filtering based on mapping quality and alignment coverage (using default

values), 60,523 loci (66% of the total number of loci in *de novo* catalog) were retained for the

ingroup assembly, 46,036 (66% of the total number of loci in *de novo* catalog) were retained for

the "hybrid zone" assembly, and 65,537 (67% of the total number of loci in *de novo* catalog) for

the full assembly.

SNP datasets

We obtained different SNP datasets to be used in downstream analyses using the *populations*

program from STACKS 2.66 (Rochette et al., 2019). For some analyses, as specified below, we

did further filtering of SNPs using *vcftools* (Danecek et al., 2011). For the ingroup assembly, we

obtained two different SNP datasets: one including one SNP per locus, and one that included all

variant and non-variant sites per locus (i.e., full haplotypes only for fineRADStructure, see

below). In both cases we applied the following filters: *--min-samples-overall* 0.8 (locus needs to

be present in 80% of all individuals to be retained), *--min-mac* 5 (minimum allele count of 5),

and *--max-obs-het* 0.7 (maximum observed heterozygosity of 0.7). For the full haplotype dataset,

we used the same parameters and included the *--filter-haplotype-wise* flag. We decided to apply

a minimum allele count (*--min-mac* 5) filter to exclude SNPs that may be artifacts of genotyping

errors, but kept a relatively low value to avoid excluding all low frequency SNPs which also

contain relevant information for population genetic analyses (Linck & Battey, 2019). Details on

how the SNPs datasets from the three different assemblies were obtained are in the

Supplementary information. In short, we used the same filtering parameters for the hybrid zone

assembly and for the assembly including outgroups we used *--min-mac 2* and *--min-samples-per-*

pop 0.5

Based on the assembly for the ingroup only, and after filtering, we obtained 5,054 loci,

composed of 73,786 sites, including 22,132 variant sites for the full haplotype dataset. For the

ingroup (one SNP per locus) dataset, we obtained the same number of loci and kept 4,800 variant

sites.

Analyses of diversification within the *ornatus-melanocephalus* complex

We described and visualized genetic structure in order to test its relationship with plumage and

geography. To evaluate how genetic structure related to current taxonomic patterns of variation

in head color patterns (see Figure 1), we also inferred subspecies-level phylogenies and

admixture graphs. Additionally, we estimated the level of differentiation between plumage

groups in terms of F_{ST}. Lastly, we used a spatially explicit model to evaluate how genetic

variation within the *ornatus-melanocephalus* complex is structured across their range.

Phylogenetic and admixture graph inference

With the goal of inferring the overall relationships among plumage groups, we employed two

methods to build phylogenies and infer admixture graphs. For this set of analyses, we excluded

all individuals from or adjacent to the hybrid zone (in white, Figure 1) and the *griseonuchus-*

bairdi transition area (arrows, Figure 1), and used the assembly that includes outgroups (*M.*

pictus, M. miniatus, M. brunniceps). For phylogenetic approaches, we included all outgroup

individuals (total n=110), while for the admixture graph inference, we only included one

outgroup (total n=105).

raxml-ng: We used RAxML-NG (Kozlov et al., 2019) to infer maximum-likelihood trees from

concatenated ddRAD loci, including both variant and invariant sites. Because individual

fragments are short (~150 bp), we concatenated all loci into a supermatrix (Leaché & Oaks,

2017). A VCF including invariant sites was generated with *populations* using the parameters --

vcf-all --min-mac 2 *--min-populations* 10 *--min-samples-per-pop* 0.5, then converted to Nexus

with *vcf2phylip*. (https://github.com/edgardomortiz/vcf2phylip). Then, we ran *raxml-ng* using

this nexus alignment and the GTR + G model of nucleotide substitution with no partitions. To

search for the tree with the highest likelihood, we ran 20 independent runs, 10 of them starting

from the maximum parsimony tree and 10 from random trees. From these searches, the tree with

the highest likelihood was selected among the 20 slightly different trees. We calculated branch

support with 100 bootstrap replicates.

snapper: To infer evolutionary relationships among plumage groups under a phylogenetic

framework, we used *snapper* (Stoltz et al., 2021). *Snapper* infers a "species" tree using diffusion

models and takes as input biallelic SNPs that are assumed to be unlinked, bypassing the necessity

of building individual gene trees, similar to site-based coalescent methods (Bryant et al., 2012).

We classified individuals into groups corresponding to the seven subspecies described in Figure

1. For this analysis, we generated a SNP dataset from the assembly including outgroups (*--min-*

mac 2, *--max-obs-het* 0.7, --min-populations 10, *--min-samples-per-pop* 0.5) filtered to only keep

biallelic SNPs using *vcftools* and then converted the resulting *vcf* files to nexus using the

vcf2phylip script. Using the nexus alignments as input, we prepared the *.xml* files in Beauti 2.7.6

(Bouckaert et al., 2019). We ran four independent chains with a length of 15,000,000 steps, and a

thinning interval of 1,500. We set a gamma prior for the birth rate (λ), and left default values for

the remaining priors. We increased the weights of two operators (gamma mover and rate mixer)

to 10. Stationarity was assessed using Tracer 1.7.1 (Rambaut et al., 2014), based on a visual

exploration of trace plots and ESS values (ESS > 500 was obtained for all parameters.) After

exploring trace plots, we set the burn-in at 10%.

TreeMix (OrientAGraph): As an alternative way to describe the historic relationships among

plumage groups, we used *OrientAGraph* (Molloy et al., 2021), an approach based on the

TreeMix algorithm, which implements a graph-based model that uses genome-wide allele

frequencies to infer population splits and admixture events (Pickrell & Pritchard, 2012). This

method detects evidence of historical introgression between plumage groups and can better

describe the history of populations when not fully bifurcating. For input, we applied the same

SNP filtering as in the *snapper* analysis, adjusting only *--min-populations* to 8 and excluding

SNPs with >30% missing data using *--max-missing* 0.7 in VCFtools. The filtered VCF was

converted to TreeMix format with *vcf2treemix.py*.

(https://github.com/CoBiG2/RAD_Tools/blob/master/vcf2treemix.py).

We ran *OrientAGraph* testing for a range of migration edges (*m* parameter), running 5

iterations per *m*, with *m* values ranging from 1 to 6. We used *M. miniatus* (n = 3) as outgroup to

root the tree and incorporated a *bairdi-chrysops* known migration event using the *-cor_mig* and -

climb flags, and a 0.5 migration weight. This was done to account for the high likelihood that

there are signs of introgression even away from the hybrid zone. After running all iterations for

this range of m values, we used *OptM* (Fitak, 2021) to evaluate the optimal value of *m* in each

case, using an approximation based on linear regressions between *m* values and likelihood. We

made a small modification to the *optm* function to be able to account for the known migration

event. Based on this linear regression criteria, we chose the optimal *m* values and included those

results here.

Genetic structure within the ornatus-melanocephalus complex

PCA: To summarize genetic variation in the *ornatus-melanocephalus* complex, we conducted a

genomic PCA using functions implemented in *gdsmt* and *SNPrelate* (Zheng, 2013) for R. To

conduct this PCA, we used only one SNP per locus, including only biallelic SNPs. To account

for the effect of missing data on the PCA (Yi & Latch, 2022), we (1) visualized the amount of

missing data in our PC plots, to evaluate the possibility that high missing data samples are

clustered together and/or have values biased towards the origin, (2) ran an additional PCA only

including SNPs with a missing rate < 0.1, and (3) ran an additional PCA using a different *vcf* as

input with more strict filtering for missing data when running populations (*--min-samples-overall*

0.9). We also conducted a PCA for a subset of the individuals to describe in more detail genetic

variation in the hybrid zone and adjacent areas (*chrysops*, hybrid zone individuals, *bairdi*, and

griseonuchus).

fineRADStructure: As a complementary approach to visualize genetic structure within the

ornatus-melanocephalus complex, we used *fineRADStructure*, which estimates a coancestry

matrix and an associated distance tree based on full haplotype information from RADseq data

(Malinsky et al., 2018). This method has particularly high resolution in inference of recent

shared ancestry, and allows one to describe nested patterns of genetic structure (Malinsky et al.,

2018). To run *finestructure* (part of the *fineRADStructure* pipeline), we used the output from the

populations program containing both variant and invariant sites (i.e., full haplotypes). No

population prior was specified for the run, which was set for 200,000 iterations with a burn-in of

100,000 (-x 100,000, -y 100,000), and a thinning interval of 1,000 (-z 1000).

Calculation of F_{ST}: To estimate overall genetic differentiation among plumage groups, we

calculated pairwise F_{ST} between subspecies (i.e., plumage and geographic groups). We excluded

all individuals from the hybrid zone from this analysis. Given the uncertainty in the *bairdi-*

griseonuchus limits (see Results), we also excluded individuals from the southern Ecuadorian

provinces (as in the *snapper* analyses). We calculated the Weir and Cockerham estimator (Weir

& Cockerham, 1984) using *vcftools* (Danecek et al., 2011) in 25,000 bp windows. To test for

potential discordance between nuclear and mitochondrial divergence, we evaluated whether

SNPs-based pairwise F_{ST} values were correlated with those estimated from the ND2 gene

(Céspedes et al. 2021). To assess the correlation between these two matrices, we implemented a

Mantel Test based on a Pearson's correlation using the function *mantel* implemented in the R

package *vegan* (Oksanen et al., 2007)

To test for isolation-by-distance (IBD), we also calculated pairwise windowed F_{ST}

between localities as defined in Supplementary Table 1 and evaluated if these are correlated with

the pairwise geographic distances. Geographic distances between localities were calculated using

the function *rdist.earth* implemented in the R package *fields* (Nychka et al., 2025). We also

evaluated the correlation between these matrices using a Mantel test based on a Pearson's

correlation as implemented in *vegan*.

Admixture: We employed admixture (Alexander et al., 2009) to estimate individual genetic

ancestries using a maximum-likelihood approach at three different geographic scales: whole

complex, hybrid zone and adjacent populations (*bairdi, chrysops*, hybrid zone, *griseonuchus*),

and hybrid zone only (*bairdi*, *chrysops*, hybrid zone).

Prior to running admixture, we calculated relatedness between all individuals and then

excluded individuals that appeared closely related. Specifically, we used the *--relatedness2* flag

in *vcftools* to export a table of pairwise relatedness values using the KING kinship estimator

(Manichaikul et al., 2010). We then identified pairs of individuals with a relatedness higher than

0.177 (i.e. likely first-degree relatives) and excluded one of them from admixture analyses. We

then transformed the VCF files to *.bed* format by using plink2 (Chang et al., 2015). Additionally,

we performed a SNP window linkage-disequilibrium pruning using plink2 *(--indep-pairwise*,

followed by *--extract*), on windows of 50 SNPs and using a 0.3 threshold. This was done for both

scales of analyses, and then admixture was run on the thinned datasets.

For all admixture runs, we included the *–cv* flag, to perform the cross-validation

procedure to choose the optimal k (i.e. number of source populations) value. For the whole

complex analyses, we ran admixture for values of $k\{1…10\}$, and for the other two scales, we

used values of $k\{1…6\}$. For each value of k, we ran 10 iterations starting with a randomly

generated seed. We analyzed variation across runs using *pong* (Behr et al., 2016) and generated

figures using the *mapmixture* (Jenkins, 2024) package for R.

FEEMS: To more directly evaluate how genetic variation is shaped across the Andes, we used a

spatially explicit model based approach: *FEEMS* (Marcus et al., 2021)

(https://github.com/VivaswatS/feems). This approach is based on a model of IBD and builds

continuous migration surfaces based on a geographic grid, visualizing areas of inferred low and

high gene flow (Marcus et al., 2021). To run FEEMS, we prepared four input files: (1) *plink* file

with genotype data for all individuals in the *ornatus-melanocephalus* complex (same set as for

whole-complex admixture analyses), (2) text file with geographic coordinates for all specimens

with genotype data (Supplementary Table 1), (3) a global grid with triangular cells of

approximately 7,774 km^2, obtained using the package *dggridR* for R (res=8; Barnes et al. 2024),

and (4) outer extent boundary for the grid, which was generated as a polygon that roughly

delineates the montane areas occupied by *Myioborus* and encompasses all sampling sites. The

chosen resolution of the grid allowed for most individuals in the same sampling site to be

assigned to a unique deme, based on preliminary explorations. We used the cross-validation

approach (Marcus et al., 2021) to choose the penalty parameters to be used in the optimization

procedure (λ and λ_q; https://github.com/VivaswatS/feems/blob/main/docsrc/notebooks/cross-

validation.ipynb). After preliminary explorations, the migration surface presented here was

generated using $\lambda = 0.17$ and $\lambda_q = 100$, which minimized the cross-validation error (although

lower values appeared roughly as adequate, Figure 6).

Analysis of genetic variation across the *bairdi* x *chrysops* hybrid zone

Calculation of a genomic hybrid index and genetic composition of the hybrid zone

We estimated a genomic hybrid index and interclass heterozygosity using functions implemented in *triangulaR* (Wiens & Colella, 2025). As noted above, the southern parental population was defined based on our own results and includes individuals from central to south Ecuador that do not cluster genetically with *griseonuchus* (see Results). In the SNP dataset for this analysis, we included all SNPs per locus.

To identify ancestry-informative SNPs, we chose an allele frequency difference of 0.5 between parental populations. Higher thresholds resulted in very few (i.e., less than 100) or no SNPs being retained, and there were no alleles fixed in each parental population. Then, to describe the genetic composition of the hybrid zone, we used the values of genome-wide hybrid indices and observed interclass heterozygosity for all individuals, and plotted these in a triangle plot (Cronemberger et al., 2020; Liu et al., 2022; Lopez et al., 2021).

Geographic cline analyses

To describe the geographic extent of the hybrid zone and evaluate whether genetic and plumage variation were concordant, we fitted geographic clines using *HZAR* (Derryberry et al., 2014). We fitted and compared clines for genomic hybrid indices—corresponding to admixture proportions and estimated using *gghybrid*—and plumage hybrid indices estimated previously for these individuals (Céspedes-Arias et al., 2021). For each of these "traits", we fit a model that estimates cline width (w) and center (c) but has fixed values for mean and variance of the trait values at the cline extremes ($\mu L, \mu R, varL, varR$). Both models were run with a tune value of 1.5, and for each of these we ran 3 chains of 2 million iterations discarding the first 10% as burn-in. All runs were performed randomizing starting parameters. To evaluate whether plumage and genomic clines

are coincident and concordant, we compared the 95% confidence interval of the center and width

estimates, which were calculated aggregating the three independent chains for each model.

Results

Evolutionary relationships among plumage groups as inferred from phylogenies and

admixture graphs

We employed two different phylogenetic approaches to describe evolutionary relationships

among the seven plumage groups that define the study taxa. Using a maximum-likelihood

approach with all loci concatenated (21,304 variant sites), we found that most individuals of the

same subspecies clustered together (Figure 1, panel C), yet we did not observe complete

reciprocal monophyly in most groups. The *chrysops* group was the exception*,* as it appeared as a

clade nested within the *ornatus* population. Notably, however, support values were generally

low, implying that not many sites are phylogenetically informative as expected for a scenario of

recent divergence and/or ongoing gene flow. Generally, we observed that individuals from

geographically adjacent subspecies are intermixed in the tree.

The *snapper* subspecies-level tree (Figure 2A), inferred using 3,105 biallelic SNPs,

showed strong support for a monophyletic "southern" group, encompassing three *M.*

melanocephalus subspecies (*malaris, melanocephalus, bolivianus*) characterized by having

spectacles and a black crown. The taxon *griseonuchus*, from north of the Marañón River Valley,

characterized by having a reddish crown that is phenotypically very similar to *bairdi*, was

recovered as sister to a strongly supported clade including the more northerly groups from

Colombia and Ecuador. Within this expanded "northern" group, *chrysops* was recovered as sister

to *ornatus*, and these, in turn, are sister to *bairdi* (Figure 2A). Note that, unlike the maximum-

likelihood tree, we *a priori* assigned individuals to groups for these analyses, which can bias the

phylogenetic estimation given that the limits among these populations are not fully defined.

Overall, we found stronger signals of admixture events between the northern relative to

the southern group (Figure 2B). Depending on the *m* value, we recovered slightly different

topologies, though we note that these incongruencies were restricted to the relationships between

the northern group taxa, namely *griseonuchus, bairdi*, *chrysops*, and *ornatus* taxa (Figure 2B).

Here we present the inferences for m = 1 and m = 3 as these were the optimal values based on

the linear regressions fitting the likelihood vs *m* curves (Supplementary Figure 1). Both graphs

consistently recovered a monophyletic "southern" group (*malaris*, *melanocephalus* and

bolivianus), which was consistent with the topology of the *snapper* tree. In the m = 1 graph

(Figure 2B), *griseonuchus* and *bairdi* appeared as sister, and this clade was, in turn, sister to the

southern group. In contrast, in the m=3 graph, *bairdi* was sister to *chrysops*, and thus a migration

edge between these taxa was not estimated despite the known migration event parameter. Two

edges with high migration weight were inferred, between *griseonuchus* and *bairdi,* and

griseonuchus and the *bairdi-chrysops* node. An additional edge, with low migration weight, was

inferred between the outgroup and the *bolivianus-melanocephalus* group (Figure 2). We note,

however, that the outgroup has a very low sample size (n = 3) and thus do not find this inference

to be very reliable.

Genetic structure within the *ornatus-melanocephalus* complex

The PCA, based on 4,800 biallelic SNPs, showed that individuals generally cluster in relation to

plumage groups and geographic origin (Figure 3), but some phenotypically distinct groups were

very close in the PC1-2 space (particularly *chrysops, ornatus* and *bairdi*). As in the phylogenetic

analyses, one can distinguish clearly the "southern" group, including all individuals from

populations south of the Marañón River Valley (*bolivianus, melanocephalus, malaris,* Figure 3),

which occupied a wide range of PC1 values (Figure 3). There was also an identifiable northern

cluster (*chrysops, bairdi*, and hybrid zone individuals) that was, however, not fully distinct from

a third cluster comprised of *griseonuchus* individuals. Within this northern cluster, the plumage

groups *ornatus, chrysops, bairdi,* and individuals from the hybrid zone ("northern", hereafter)

appeared very close in PC1-2 space. As expected, individuals from the hybrid zone (in gray,

Figure 3) fell intermediate between their parental populations. The *ornatus* group appeared as a

more defined cluster when considering PC3 (Supplementary Figure 5). Notably, some

individuals putatively classified as *bairdi*, collected in the Ecuadorian provinces of Loja and

outlining ridges in the Amazon, appeared to cluster with *griseonuchus* (Figure 2). We observed a

similar pattern in the hybrid zone plus *griseonuchus* PCA (Supplementary Figure 6). We

obtained similar results when using a SNP set of the whole complex obtained with a stricter

filtering for the PCA (Supplementary Figure 7).

456   Two main clusters are supported in the coancestry matrix resulting from

fineRADstructure: populations occurring from south Ecuador to Bolivia (*griseonuchus*

plus "southern" group), and those occurring from Ecuador to Colombia ("northern" group)

(Figure 3C), which contrasted with the topology inferred by *snapper*. Within the first group,

griseonuchus and the southern group appear as differentiated clusters. The few exceptions to this

pattern are a few putatively *bairdi* individuals that clustered with *griseonuchus*, but were

collected in the southern edge of the distribution of this plumage group (where it is replaced by

griseonuchus, Figures 1 and 3) in the Cordillera del Cutucú and Loja Province of Ecuador. We

detected two main groups within the "northern" group (Ecuador and Colombia): one

corresponded to *ornatus* (occurring in the Eastern Andes of Colombia) and the other to the

hybridizing taxa in the complex and individuals from the hybrid zone.

Admixture analyses including all individuals in the *ornatus-melanocephalus* complex

revealed genetic breaks along the distribution of these warblers that were overall consistent with

fineRADStructure (Figure 4, A). For example, when assuming K = 2 and K = 3 the "southern"

group appeared as an identifiable cluster. For K = 2, which is the optimal K value based on

cross-validation error values (Supplementary Figure 8, top panel), the group *griseonuchus* also

clustered with the southern group (Figure 4, A1). For K = 3, a "southern" plus *griseonuchus*

group can be also identified, and there was an additional genetic break roughly located in the

described *bairdi-chrysops* hybrid zone in southern Colombia (Figure 4, B1).

The results of an admixture analyses focused on the hybrid zone and adjacent

populations, when assuming K = 2, showed a genetic break that clearly coincides with the

chrysops-bairdi hybrid zone (Figure 4, B and C), as previously identified based on plumage

variation (see map in Figure 1). However, when also considering *griseonuchus*, the transition

between these genetic groups is less sharp and displaced to the south (Figure 4, panel B). In both

cases there was some east-west differentiation, with localities to the west of the Ecuadorian

Andes showing a higher mean admixture proportion corresponding to the southern cluster

(Figure 4, B1 and B3). When using K = 3 for the analyses including *griseonuchus* (a *K* that

corresponds to the number of identifiable plumage groups), two genetic clusters were identified

in Ecuador, and one in Colombia, and one of these genetic breaks did again roughly correspond

to the *bairdi-chrysops* hybrid zone (Figure 4, B2). In both cases the *K* value with the lowest

cross-validation is *K* = 1 (Supplementary Figure 7).

Values of pairwise F_{ST} between populations showed an overall correspondence with

geographic distance: subspecies that are far apart in space show higher genetic differentiation

(e.g., *ornatus* vs *bolivianus,* Figure 5). We observed higher pairwise differentiation between taxa

belonging to the "southern" versus "northern" groups, which is consistent with all other analyses

describing genetic structure. In contrast, there was very low differentiation among the three

subspecies occurring south of the Marañón River Valley ("southern" group), which all have

similar head coloration. However, there was also relatively low levels of differentiation within

the "northern" plus *griseonuchus* group, which encompasses striking plumage diversity,

particularly between the two hybridizing taxa, *bairdi* and *chrysops, and bairdi* and *griseonuchus*

(Figure 5). There was a positive correlation (Figure 5, Mantel statistic r = 0.75, p-value = 0.001)

between pairwise F_{ST} calculated based on our ddRAD SNPs and the ND2 mitochondrial gene

(Céspedes-Arias et al., 2021). Lastly, as expected if patterns of genetic variation reflect IBD, we

found a positive correlation between pairwise F_{ST} between localities and geographic distance

(Figure 5, Mantel statistic r = 0.65, p-value = 0.001). Using the same classification scheme for

individuals as that to calculate F_{ST}, we estimated genetic diversity statistics. We found that

nucleotide diversity is very similar among groups, ranging from 0.076 to 0.091 (Supplementary

Table 3).

The migration surfaces inferred by FEEMS suggest that there are areas within the

distribution of these warblers in which genetic differentiation cannot be fully explained by IBD

(i.e., areas of inferred low gene flow; Figure 6). These areas of low migration coincided mostly

with two geographic breaks that dissect the distribution of the complex (Figure 1, Figure 6),

specifically the Magdalena River Valley and the Marañón River Valley, and with the hybrid

zone. In contrast, most of the range south of the North Peruvian low showed mid to high values

of inferred effective migration, which can indicate that genetic structure along this area is well

explained by IBD alone. We acknowledge that low sampling density in certain regions,

particularly in central and southern Peru, may have influenced the inferred migration surface

(Figure 6). However, cross-validation results indicate that the selected regularization parameter

provides a good fit while accounting for unobserved nodes (Supplementary Figure 9). With that

caveat in mind, the FEEMS results are broadly consistent with the patterns of genetic structure

revealed by our other analyses.

Hybrid zone

To describe the geographic extent of hybridization and the genetic composition of the *bairdi* x

chrysops hybrid zone, we identified ancestry-informative markers to estimate genomic hybrid

indexes and interclass heterozygosity for individuals across the hybrid zone and parental *bairdi*

and *chrysops* populations. Using a relatively low threshold for allele frequency difference (0.5),

we identified only 29 sites that were ancestry informative from the total of 14,301, and none of

these were fixed between the parental populations. This result complicates the interpretation of

the triangle plots (see below) and highlights the low level of genetic differentiation between

these two hybridizing taxa. Because of the scarcity of ancestry informative sites in our ddRAD

data set, which can affect the accuracy of genomic hybrid index estimations, we opted to conduct

cline analyses with admixture proportions (with K=2) instead.

We found that the head coloration and genomic hybrid index, defined as the admixture

proportions in Figure 4 (panel C3) had different centers but similar widths (Figure 7, panel A).

The admixture proportion cline had a width of 281.6 km (95% confidence interval [CI]: 245.0–

319.8km), compared to 240.6 km for head coloration (95% confidence interval [CI]: 277.5–

311.1 km). Given the substantial overlap in confidence intervals, we do not interpret this 41 km

difference in width as significant. However, we found a 37 km difference in the estimates of the

center between the plumage and admixture proportion clines, for which the 95% confidence

intervals did not overlap. For admixture proportions the cline was centered around 348.8 km

(95% confidence interval [CI]: 335.4–362.7 km) in our sampling transect, while the plumage

cline was centered around 311.1 km (95% confidence interval [CI]: 300.7–322.6 km). Therefore,

based on our *HZAR* analyses, the plumage cline appeared to be displaced towards the south (i.e.,

towards Tungurahua, Ecuador, the origin of the sampling transect), implying that the *chrysops*-

like plumage traits may be introgressing into *bairdi*. However, there was an overall strong

correlation between the plumage hybrid indices and admixture proportion across the hybrid zone

and allopatric populations (Figure 7, panel B; linear model; *p-value* < 0.001, *m* = 0.9, *R-squared*

= 0.8). We identified the outliers of this linear regression as those individuals with residuals >2

SD units from the mean, finding that most were below the regression line (Figure 7).

Specifically, we identified six individuals that had a more *chrysops*-like plumage from the hybrid

zone area (i.e., lower) relative to what the model predicts based on the admixture proportion,

which was consistent with the cline analysis results that shows that *chrysops*-like plumage is

introgressing into *bairdi*.

To describe the genetic composition of the *bairdi-chrysops* hybrid zone, we built triangle

plots (interclass heterozygosity vs hybrid index) based on the 29 ancestry-informative markers

that we identified (Figure 7, panel C). The individuals from the hybrid zone are distributed

towards the center of the triangle, many below the line that defines what value combinations are

possible under Hardy-Weinberg Equilibrium which is likely a product of the low allele

frequency difference threshold. We note that individuals from populations away from the hybrid

zone (Figure 7, in green and orange) are not located in the bottom corners of the triangle plot,

which is expected given that the allele frequency difference threshold is lower than one. If

interpreted considering the caveats described above, the distribution of individuals in the triangle

is suggestive of a hybrid zone mostly composed of advanced generation hybrids, without many

F1s.

Discussion

We used reduced-representation genomic data to study genetic variation at two different spatial

scales: the full distribution of a geographically variable warbler complex (*M. ornatus-*

melanocephalus) and a known hybrid zone between two of these plumage forms. At both scales,

our set of thousands of SNPs allowed us to explore genetic structure with a resolution that

mtDNA loci could not provide in previous studies (Céspedes-Arias et al., 2021; Pérez-Emán,

2005). We found evidence of isolation-by-distance driving genetic structure across the

latitudinally extensive range of these warblers, and genetic divergence increased where ranges

are separated by known geographic barriers. These clear genetic breaks coincided with observed

differences in head plumage coloration among populations. Notably, one transition between

groups includes a geographically restricted hybrid zone between *bairdi* and *chrysops*, likely

comprised primarily of advanced generation hybrids, where differences in genetic ancestry and

head coloration are correlated.

Below we discuss how our findings relate to the history of divergence and gene flow

among these populations, with a focus on how genetic variation imperfectly covaries with

plumage traits, and how it is structured across geography. We then discuss how genetic variation

is structured across the hybrid zone, how it relates to patterns of variation in head coloration in

this area, and whether there is evidence of selection against hybrids maintaining this seemingly

extensive hybrid zone.

Geographic variation in plumage color and genetic structure

Our qualitative assessments based on historical plumage descriptions and examinations of study

skins suggest that for these *Myioborus*, patterns of geographically clustered phenotypic variation

coincide with phylogeographic structure uncovered in the present study. First, we found that

individuals belonging to the same subspecies usually form genetic clusters, and that all major

genetic breaks within the species complex coincide with a transition between clearly

differentiated plumage groups, although we note that we lack data quantifying the degree of

plumage differentiation between subspecies (e.g. Winger and Bates 2015). In addition,

subspecies with subtle plumage differentiation (Cuervo and Céspedes Arias 2023), specifically

those occurring south of the Marañón River Valley (Figure 1), all characterized by black crown

and spectacles, show little genetic differentiation from one another.

592   Previous examinations of genetic variation in mitochondrial loci (Céspedes-Arias et al.,

2021; Pérez-Emán, 2005) found extensive haplotype sharing and little genetic differentiation

among the subspecies *ornatus* and the hybridizing *bairdi* and *chrysops*, which are all

characterized by striking plumage differences (see Figure 1). In contrast, in our analyses of

ddRAD data, which encompass thousands of SNPs across the genome, these subspecies are each

identifiable genetic clusters, albeit with shallow genetic differentiation. Taken together, we

interpret these results as evidence of rapid and recent diversification of head plumage patches in

the *ornatus-melanocephalus* complex, as seen in other Neotropical taxa (Campagna et al., 2012,

2017). Our results also highlight the usefulness of reduced representation genomic data to study

the evolutionary history of young radiations (Aguillon et al., 2018; Toews, Campagna, et al.,

2016), in which few loci might be insufficient to detect genetic differentiation and describe

evolutionary relationships among phenotypically distinct populations (Friis et al., 2016;

Stryjewski & Sorenson, 2017). We note that lack of differentiation in mtDNA between

phenotypically distinct groups could also be the result of mitochondrial introgression (Irwin et

al., 2009; Toews & Brelsford, 2012), although the strong positive correlation of F_{ST} for ND2 and

our ddRAD loci does not support a scenario of extensive mitochondrial introgression in the

group.

Genetic structure in the region of the Andes spanning from central Ecuador to northern

Peru (north of the Marañón River Valley) is complex and not clearly associated with a single

topographic feature. This area also broadly corresponds to the transition between *griseonuchus*

and *bairdi* plumage (see Figure 1, Supplementary Figure 1)*,* two subspecies characterized by a

rufous crown and spectacles. Despite their overall similarity, these two groups show

differentiation in at least two head plumage traits: the presence of a black band behind the rufous

crown and the extent of black in the cheeks and lores (Zimmer 1949; Curson et al. 1994; World

and International 2019; Cuervo and Céspedes Arias 2023). Our qualitative assessments of

plumage variation (Supplementary Figure 1), however, suggest that the variation in these traits

might not be completely discrete. Moreover, we found that individuals from Loja and the

Cordillera del Cutucú (Ecuador), show substantial variation in both putatively diagnostic

plumage traits. Therefore, these individuals, which are genetically clustered with *griseonuchus,*

do not uniformly resemble the individuals from Cajamarca, Peru (Supplementary Figure 1).

Because of limited sampling in the area, we are unable to draw stronger conclusions, but our data

suggest that there is some discordance between plumage and genetic variation, and evidence of

introgression based on admixture graphs. Despite some differentiation in at least these two

plumage traits, our genetic results are concordant with substantial historical or contemporary

gene flow among populations in this area. This suggests that the geographic range of *bairdi* is

potentially outlined by two hybrid zones, one to the north with *chrysops* (Céspedes-Arias et al.,

2021), and to the south with *griseonuchus.* Evaluating this hypothesis of hybrid zones at the

range edges, as observed in barn swallows (Scordato et al., 2017)*,* will require a systematic

collection of specimens across the *bairdi-griseonuchus* boundary.

Genetic structure across space: the influence of geographic distance and topographic

features along the Andes

Several of our analyses support a strong correlation between genetic variation and geographic

location of samples and support an important role of isolation-by-distance driving genetic

structure in *Myioborus* along the Andes. Patterns of IBD are close to ubiquitous in nature

(Aguillon et al., 2017; Seeholzer & Brumfield, 2018; Vos & Velicer, 2008), but see Meirmans,

2012), and are often taken as the null model for genetic differentiation if organisms have limited

dispersal given the scale of their range, as is the case for these small warblers with a

latitudinally-expansive distribution range. However, IBD does not account fully for the observed

patterns of genetic variation. Specifically, we found evidence of discrete genetic—and

phenotypic—clusters, and of reduced migration (relative to IBD) across some areas along the

Andes, which in turn correspond to lowland valleys such as the Marañón and the Magdalena

River Valleys. These deep, lowland gaps interrupt the continuity of humid high-elevation forest

and scrub (Graham et al., 2010) that these *Myioborus* inhabit in the present, and also coincide

with genetic and phenotypic breaks within other taxa that inhabit similar elevation bands

(Gutiérrez-Pinto et al., 2017; Martínez-Gómez et al., 2023; Winger & Bates, 2015). These two

topographic features were identified in a comparative study as being associated with allopatric

events across multiple Andean bird genera (Hazzi et al., 2018). These genetic breaks, clearly tied

to topographic features, also correspond with striking discontinuities in head coloration traits

(see Figure 1).

Other areas associated with clear genetic breaks observed in the inferred migration

surface were not clearly associated with topographic features that interrupt the present-day

continuity of humid, high-elevation forests. Specifically, low migration is inferred in a large area

ranging from the border between Ecuador and Colombia down to southern Ecuador. We suggest

that this pattern emerges results partially because this broad area includes the *chrysops-bairdi*

hybrid zone, in which genetic differentiation relative to distance is expected to be higher than

what is expected under pure IBD, as two somewhat differentiated subspecies come into contact.

The extension of this area of low inferred migration to south Ecuador is consistent with our

fineRADStructure, PCA and admixture results that suggest that there is substantial genetic

differentiation between individuals in close proximity in this area. Specifically, we found that

birds from isolated mountain spurs such as the Cordillera del Cutucú (southeastern Ecuador) and

Cordillera del Condor (Peru-Ecuador border) clustered genetically with birds from the main

Andes in Cajamarca (northwestern Peru), and Loja (southern Ecuador), instead of the more

geographically proximate individuals from the eastern Ecuadorian Andes in Morona-Santiago

(see Figure 2). The Cordillera del Cutucú and Cordillera del Cóndor are outlying mountain

ranges in the Amazonian foothills, separated from the main Andes by the valleys of the rivers

Upano and Zamora, respectively (Pozo-Zamora et al., 2022; Robbins et al., 1987). Our results

suggest that these valleys, and surrounding lowland areas, have likely acted as dispersal barriers

for these higher-elevation warblers. On the other hand, the patterns of genetic variation described

here indicate that there has likely been substantial gene flow connecting the populations of

warblers inhabiting isolated peaks in these mountain ranges (separated by the Santiago River

Valley; Robbins et al. 1987; Pozo-Zamora et al. 2022), and the eastern slope of the Andes in

northern Peru.

Hybrid zone dynamics

Based on our reduced-representation genomic data set, the hybridizing *chrysops* and *bairdi* are

characterized by very shallow genetic divergence, which contrasts with their striking plumage

differentiation. Moreover, across thousands of SNP sites, we only identified 29 ancestry

informative sites with substantial allele frequency differences between parental populations. This

scarcity of ancestry informative markers limits our ability to draw definitive conclusions about

the genetic composition of this hybrid zone but highlights the low level of genetic differentiation

between *chrysops* and *bairdi*.

This hybrid zone likely formed by secondary contact of two formerly isolated plumage

forms. The disconnection and subsequent reconnection of habitat that could have shaped this

process may have followed elevational shifts in the cloud forest belt during the Pleistocene

(Flantua et al., 2014; Ramírez-Barahona & Eguiarte, 2013), or potentially been driven by

volcanic activity that modified topographic connectivity (Sanín et al., 2024). An investigation of

the evolutionary origins of this hybrid zones will require both a better understanding of the

patterns of connectivity of cloud forest in the area where the *chrysops-bairdi* hybrid zone is

centered, as well as an estimation of the time since secondary contact, which could be obtained

based on inferences of ancestry tract length in whole genome data (Pool & Nielsen, 2009).

Our geographic cline analyses using plumage and genomic data suggest that the hybrid

zone extends for over 200 km, similar to the findings from its original description based on face

color traits (Céspedes-Arias et al., 2021). We do not find statistically significant differences in

the width of the admixture proportion (i.e., genomic) and plumage geographic clines. Taken

together, plumage and genomic patterns of variation thus suggest that hybridization between

chrysops and *bairdi* has been extensive, albeit still restricted to a fraction of the entire

distribution of these taxa (approximately 30% of our sampling transect) which is continuous at

least from central Ecuador to the north of the Central Andes of Colombia. Therefore, the

geographic extent of hybridization is unlikely to be constrained by physical or ecological barriers

that limit dispersal beyond the hybrid zone (e.g., DeRaad et al., 2023), raising the question of

what factors are restricting the hybrid zone to this area.

The lack of information on natal dispersal distances as well as a the wide range of

plausible times since secondary contact constrains our ability to generate a direct comparison of

the observed cline widths with that expected under neutral diffusion (Barton and Gale 1993).

With the available data we are thereby unable to evaluate whether the *chrysops* x *bairdi* hybrid

zone corresponds to a stable tension zone, in which the width of clines is constrained by

selection against hybrids. A second possibility is that the observed sharp genomic and plumage

clines are not maintained by selection, but instead that this is a young and potentially expanding

hybrid zone, and that the differentiation between *chrysops* and *bairdi* might become eroded over

time (Kearns et al., 2018).

Using a range of plausible dispersal distances inferred from another *Myioborus* species

(Céspedes-Arias et al., 2021; Mumme, 2015), we can approximate expectations under neutrality

(Barton & Gale, 1993). For a cline width of 281 km, the neutral diffusion model predicts that

~14,337 generations would be required assuming a dispersal distance of 935 m, or ~53,278

generations with a dispersal distance of 485 m. In principle, building upon our systematic

collection of specimens across the *chrysops* x *bairdi* hybrid zone, the alternative of an expanding

hybrid zone could be revisited by comparing plumage and genomic clines based on specimens

collected across different time points. However, given that the expansion of this hybrid zone

under neutral diffusion with plausible dispersal distances is expected to be slow (i.e. 0.5 km over

50 generations), time between sampling periods would need to be very long to detect changes.

We found that cline centers are not fully coincident, as that of the plumage cline is

displaced to the south of the genomic cline center, implying that the *chrysops*-like plumage is

introgressing into *bairdi.* Assuming that most of our ddRAD loci can be considered neutral

markers, our results indicate that head coloration might be under selection across this hybrid

zone (Gay et al., 2008). Specifically, the observed pattern would be consistent with a scenario in

which the *chrysops*-like plumage is favored across the hybrid zone, which would lead to the

asymmetric introgression of this phenotype towards the south. Asymmetric introgression of

plumage traits has been observed in other avian hybrid zones (Baldassarre et al., 2014; Lim et al.,

2024; Semenov et al., 2021), and in two of these cases, behavioral evidence supported the idea

that these traits provide fitness advantages in the context of mate choice (Baldassarre & Webster,

2013; Long et al., 2024; Stein & Uy, 2006).

Even though the difference in plumage and genomic cline center is statistically

significant, we found it to be relatively small (38 km; approximately 4% of the entire sampling

transect). This contrasts with much more marked differences in center estimations found in other

hybridizing birds like *Motacilla* wagtails (Semenov et al., 2021) and *Malurus* fairy-wrens

(Baldassarre et al., 2014). We acknowledge that this minor difference could be an artifact of the

way in which both of our hybrid indexes were calculated (e.g. with the scoring system giving

equal weight to pre-defined patches; Céspedes-Arias et al., 2021), and potentially low sampling

sizes in some of the localities. However, this slight difference could be the result of selection on

the head plumage patches. Even though the role of these color patches has not been studied in

these warblers yet, it is plausible that they may be relevant for intraspecific communication and

mate choice as seen in head plumage patterns in many other bird species (e.g., Marchetti 1992;

Andersson et al. 1998; Scott and Deag 1998; Rémy et al. 2010). Future behavioral studies could

evaluate these hypotheses of selection favoring *chrysops*-like head color patterns across this

hybrid zone.

Besides describing the geographic extent of hybridization, we used our reduced-

representation genomic data set to gain insights into the genetic composition of the *chyrsops x*

bairdi hybrid zone. However, as noted above, the scarcity of ancestry informative sites in our

dataset complicates the interpretation of these results, as it compromises the accuracy of genomic

hybrid indexes and interclass heterozygosity estimations (Gompert et al., 2024; Wiens & Colella,

2025). Setting a low allele frequency threshold, which was necessary in our case, can also result

in a tendency of individuals to cluster near the center of triangle plots (Wiens & Colella, 2025).

Bearing these caveats in mind, we interpret our triangle plot results as suggestive of a hybrid

zone composed mostly of advanced generation hybrids, with no F1s sampled, given the lack of

individuals with intermediate hybrid indexes and very high interclass heterozygosity (Capblancq

et al., 2020; DeRaad et al., 2023; Natola et al., 2021). This is consistent with the existence of a

high diversity of plumage phenotypes in the hybrid zone, as well as the lack of contemporary

geographic contact between parental *chrysops* and *bairdi* populations (i.e., we do not observe

"pure" individuals of both species in the same sites) (Céspedes-Arias et al., 2021). Given the

shallow genetic divergence between *chrysops* and *bairdi*, a more detailed examination of the

genetic composition of this hybrid zone will necessitate whole-genome sequencing, capturing

more ancestry informative sites. Specifically, as in other examples of hybridizing taxa with very

little genomic divergence, whole-genome sequencing may be needed to identify these regions

and sites that are highly differentiated and study hybridization dynamics in detail (Aguillon et al.,

2021; Poelstra et al., 2014; Toews, Taylor, et al., 2016).

 Figures

Figure 1: Sampling localities of specimens included in the study **(A)**, which encompass all major plumage groups **(B)**. The color of the dots corresponds to study taxa. The white dots correspond to areas around the *bairdi-chrysops* hybrid zone as identified based on plumage patterns (Céspedes-Arias et al., 2021). The subspecies *malaris, melanocephalus*, and *bolivianus* are here represented by one illustration. In the map, elevation is denoted by the gray scale in the background, with the elevation in which these *Myioborus* occur shown in an intermediate shade of gray. The localities marked with an asterisk correspond to those in Loja and outlining ridges in Morona-Santiago that are discussed in the main text, and numbers mark important topographic features. **(C)** Maximum-likelihood phylogeny estimated with our dataset of concatenated ddRAD loci, showing how overall relationships somewhat reflect geography (outgroups are cropped out of the figure here). The branches are colored by subspecies as in the map, with only bootstrap values over 70 shown (hybrid zone samples not included in the analysis). Illustrations: Andrés Montes Rojas.

Figure 2. Evolutionary relationships within groups in the *Myioborus ornatus-melanocephalus* complex as inferred by snapper and *OrientAGraph*. **(A)** The main figure shows the phylogenetic relationships between taxa, with the blue topology corresponding to the most frequent in the posterior distribution, followed by red. **(B)** Admixture graphs from *OrientAGraph* with 1(top) and 3 (bottom) migration edges (*m*). The migration edges are shown as colored arrows, and the hue denotes the migration weight.

Figure 3. Genetic structure within the *ornatus-melanocephalus* complex. (**A**) The distribution of individuals in the PC1/PC2 space showing clustering based on plumage groups, which are denoted by the color of the dots. (**B**) Strong correlation between PC1 and latitude. In both cases (A and B) the size of the dot is inversely proportional to the proportion of missing data. (**C**) fineRADStructure coancestry matrix, and associated distance tree (above) showing clusters of individuals that overall correspond to plumage groups, which are denoted by the colored dots that correspond to groups as in Figure 1. The asterisk highlights a small number of putatively *bairdi* individuals, all from the south end of Ecuador, that genetically cluster with *griseonuchus* (from northern Peru). The color scale denotes values of coancestry, with blue and black tones corresponding to very high levels of coancestry.

Figure 4. Genetic clusters (**A**) across the entire distribution of the *ornatus-melanocephalus* complex and (**B**) the hybrid zone and adjacent populations as shown by the mean admixture proportions per locality. The dataset and *K* value for each plot is specified at the top of each map. **(C) Admixture prop**barplots showing the admixture proportions per individual, which are organized into localities from south to north (localities are divided by white dotted lines).

Figure 5. Genetic differentiation among Andean *Myioborus* taxa. **(A)** Mean pairwise F_{ST} values between subspecies (i.e., plumage and geographic groups). **(B)** Correlation between pairwise F_{ST} calculated with ddRAD SNPs (this study) and the ND2 mitochondrial gene from the same set of samples (Cespedes-Arias *et al* 2021). **(C)** Positive correlation between geographic distance and F_{ST} between localities supports a role of isolation-by-distance shaping genetic variation in the *ornatus-melanocephalus* complex.

Figure 6. Estimated effective migration graph across the entire distribution of the *Myioborus ornatus-melanocephalus* complex, highlighting areas of restricted gene flow. The edges of the grid are colored by estimated effective migration, from burnt orange (low migration) to blue (high migration). In the grid, the black dots correspond to the sampled nodes and the size of the dot correspond to the sample size (*N*). We highlight three areas of low migration, two geographic barriers and the hybrid zone where genetically distinct taxa have come into contact.

Figure 7. Genetic and plumage variation across the *bairdi x chrysops* hybrid zone. **(A)** The plumage and genomic clines have different estimated centers. **(B)** Photographs of specimens collected for which we calculated a plumage hybrid index, showing a sample of the phenotypic diversity across the hybrid zone, and "parental" phenotypes at both extremes. Photos: Andrés Cuervo, Paulo Pulgarín, David Ocampo, and Laura Céspedes. **(C)** We observed an overall a strong correlation between the genomic hybrid index (*bairdi* admixture proportion) and the plumage hybrid index. Most individuals that deviate from this relationship (outliers marked in color) are individuals with a more chrysops-like plumage than expected given the admixture proportion. **(F)** Triangle plot showing the genetic composition of the hybrid zone. The x axis corresponds to a genomic hybrid index as calculated using 29 ancestry informative markers. Parentals are shown in lime green and orange.

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