Shared latent representations of speech production for cross-patient speech decoding

Spalding, Z.[1], Duraivel S.[1], Rahimpour S.[2,7], Wang C.[1], Barth K.[1], Schmitz C.[1], Lad S. P.[2], Friedman A. H.[2], Southwell, D. G.[2,3,4], Viventi J.[1,2,3,4,#], & Cogan G. B.[1,2,4,5,6,#]

#Corresponding authors

Author affiliations:

1. Department of Biomedical Engineering, Duke University, Durham, NC
2. Department of Neurosurgery, Duke School of Medicine, Durham, NC
3. Department of Neurobiology, Duke School of Medicine, Durham, NC
4. Duke Comprehensive Epilepsy Center, Duke University, Durham, NC
5. Department of Neurology, Duke School of Medicine, Durham, NC
6. Center for Cognitive Neuroscience, Duke University, Durham, NC
7. Department of Neurosurgery, University of Utah, Salt Lake City, UT

Abstract

Speech brain-computer interfaces (BCIs) can restore communication in individuals with neuromotor disorders who are unable to speak. However, current speech BCIs limit patient usability and successful deployment by requiring large volumes of patient-specific data collected over long periods of time. A promising solution to facilitate usability and accelerate their successful deployment is to combine data from multiple patients. This has proven difficult, however, due to differences in user neuroanatomy, varied placement of electrode arrays, and sparse sampling of targeted anatomy. Here, by aligning patient-specific neural data to a shared latent space, we show that speech BCIs can be trained on data combined across patients. Using canonical correlation analysis and high-density micro-electrocorticography (µECoG), we uncovered shared neural latent dynamics with preserved micro-scale speech information. This approach enabled cross-patient decoding models to achieve improved performance relative to patient-specific models facilitated by the high resolution and broad coverage of µECoG. Our findings support future speech BCIs that are more accurate and rapidly deployable, ultimately improving the quality of life for people with impaired communication from neuromotor disorders.

Abstract: 174/175

36 Correspondence to:
37 Gregory B. Cogan
38 Department of Neurology
39 Duke South, Blue Zone 4519, Durham, NC 27710
40 gregory.cogan@duke.edu
41
42 Jonathan Viventi
43 Biomedical Engineering
44 1149 CIEMAS, 101 Science Dr, Durham, NC 27705
45 j.viventi@duke.edu
46

47 **Running title**: Cross-Patient Speech Decoding

48 **Keywords:** Speech, IEEG, BCI, decoding

Main

Speech is an intricate coordination of muscle movements that shape sounds into words[1].
Paralysis of these muscles by neuromotor disorders such as amyotrophic lateral sclerosis (ALS)
and brainstem stroke can disrupt speech, resulting in debilitating losses to communicative ability
and a reduced quality of life[2–6]. Speech brain-computer interfaces (BCIs) have shown promise in
restoring communication to individuals with neuromotor disorders by using machine learning
models to accurately decode intended speech units directly from brain activity[7–10].

Speech BCIs can be broadly divided into non-invasive approaches, such as
electroencephalography (EEG), and invasive approaches, such as electrocorticography (ECoG).
Non-invasive BCIs offer advantages in safety and ease of deployment but are limited by lower
spatial resolution and susceptibility to noise, which can constrain their ability to resolve fine-
grained articulatory representations required for high-accuracy speech decoding[11,12]. In contrast,
invasive BCIs provide higher spatial and temporal resolution recordings directly from cortical
populations, enabling more accurate decoding of speech-related neural activity[7–10], but introduce
challenges related to surgical implantation, limited and variable electrode coverage, and patient-
specific sampling of neural populations. Invasive recording modalities have become a primary
focus for achieving high-accuracy speech decoding despite these challenges.

A key limitation of invasive speech BCIs is their reliance on patient-specific approaches that
require resource-intensive, individualized data acquisition from each patient – typically spanning
multiple weeks – before their associated machine learning models can be trained to achieve
maximal accuracy[8]. This approach is taken to mitigate the effects of varied sensor placement,
sparse sampling of the targeted anatomy, and neuroanatomical variability across patients[13].
These long training and deployment times make patient use cumbersome and hinder their
adoption as assistive communication devices. Future speech BCIs that successfully combine data
across multiple patients may accelerate training by requiring minimal data collection from new

patients. Such cross-patient speech BCIs could be rapidly deployable and demonstrate greater
robustness across a broader patient population.

For successful cross-patient speech decoding, there must be similarity in the neural
representations of speech production across patient recordings. Invasive speech decoding
studies typically record from the sensorimotor cortex (SMC)[14–16] with intracranial
electroencephalographic devices, such as ECoG[17–20] or intracortical microelectrode arrays[21,22],
from which sampled representations of speech production may differ from patient to patient due
to variability in electrode array placement or individual neuroanatomy. However, recent work has
demonstrated that for motor activity, there is a low-dimensional structure in the time-varying
neural population activity of the motor cortex[23–25], known as latent dynamics, that is shared across
individuals. For example, in non-human primates, the latent dynamics of individual subjects
performing limb movement can be aligned and pooled across animals to train a cross-animal
movement decoding model[26]. We hypothesize that this phenomenon extends to human speech
production. That is, the latent dynamics underlying the motor control of articulator muscles in the
vocal tract to produce speech have a shared representation across separate patients despite
variability in electrode placement and individual neuroanatomy (Fig. 1a). These latent dynamics
can be aligned and pooled across patients to enable training of cross-patient speech BCIs.

We additionally hypothesize that the elucidation of shared latent dynamics requires an accurate
sampling of speech-relevant neural population activity. We have previously shown that accessing
the rich spatiotemporal representations of speech production in the SMC requires precise neural
interfaces with high spatial-specificity and broad coverage. This dense, broad sampling of the
cortex has been achieved using micro-electrocorticographic (µECoG) arrays to record from a
large surface of the brain with a high density of micro-contacts[27], which has allowed resolution of
micro-scale features for both epilepsy[28] and speech production[29]. While surface recording via
µECoG relies on activity in neural local field potentials (LFPs) and cannot access the same multi-
unit activity that previous studies used to investigate these cross-patient motor latent dynamics[26],
previous work has shown strong similarity between latent dynamics extracted from high-frequency
LFP bands and multi-unit sources[30]. Together, these results suggest that µECoG has sufficient
spatial-specificity, coverage, and signal quality to enable successful cross-patient alignment for
speech BCIs.

Here, we investigate cross-patient representations of speech production by performing acute
neural recordings with high-density µECoG arrays over the SMC of eight patients during a speech
repetition task. We use dimensionality reduction techniques to reduce high channel-count µECoG
recordings into low-dimensional latent dynamics, and we subsequently learn linear
transformations that align these latent dynamics across patients. We show that cross-patient
latent dynamics encode speech information relevant to the control of articulator muscles in the
vocal tract. These shared speech encodings enable the training of cross-patient speech decoding
models, which in turn outperform patient-specific decoding models. Lastly, we show that micro-
scale sampling of the SMC with broad coverage is critical for learning the proper alignment
between patients that preserves speech information.

Figure 1. Shared latent dynamics underlying speech production. a. Despite differences in electrode placement and individual neuroanatomy, we propose that the neural population activity that controls articulatory muscles for speech production is shared across separate patients and can be elucidated by aligning their latent dynamics to a common space. **b.** Neuroanatomical location of μECoG arrays for all patients projected onto a common brain (MNI-152, see Fig. S1 for patient-specific array locations). **c.** Speech repetition task. Patients are presented with an auditory non-word stimulus that is composed of three phonemes (e.g. */abe/*) and are instructed to immediately repeat this phoneme sequence. Timing shows statistics of task performance. For information on experimental design see Methods – *Speech task* **d.** Example spectrogram showing high-gamma (HG; 70-150 Hz) power centered around the response onset (t = 0). **e.** Spectrograms across 128-channel (left, S1) and 256-channel (right, S3) μECoG arrays. Black borders indicate channels with significant HG power during patient response relative to a pre-stimulus baseline (FDR-corrected non-parametric permutation tests). The red border indicates the example spectrogram from **d.**

Our results demonstrate improved speech decoding for BCIs facilitated by a shared, cross-patient
representation of speech production encoded in the latent dynamics representing SMC population
activity. These findings enable future speech BCIs with accelerated deployment times and
increased generalizability, resulting in improved quality of life for individuals who have lost the
ability to speak due to neuromotor disorders.

Results

We recorded speech-relevant neural activity from the surface of the SMC using high-density, high
channel-count μECoG arrays in eight patients undergoing awake neurosurgical procedures (four
with 128 channels – 1.33 mm spacing, four with 256 channels – 1.72 mm spacing, see Table S1
for summary of patient information). Fig. 1b shows the electrode array locations of all patients
projected to an average brain (see Fig. S1 for array locations on patient-specific brains).
Recordings were performed intraoperatively while patients were instructed to repeat non-word
sequences of three phonemes, or sound units (e.g. */b/* in bat), immediately after hearing an

auditory presentation of the non-word (Fig. 1c, see Methods and Table S2). Using
spectrotemporal analysis of the recorded neural signals, we observed neural activations around
patient response onset (Fig. 1d). Specifically, we found strong increases in power in the high-
gamma (HG; 70-150 Hz) band, which has been shown to be a robust marker of speech production
in the SMC[8,19,31] that correlates well with multi-unit neural firing[32–34]. Significant HG activations
during patient responses (relative to a pre-stimulus baseline, FDR-corrected non-parametric
permutation test – see Methods) were observed in many electrodes across the entirety of each
µECoG array (significant electrode amounts in Table S3), exhibiting unique spatial activation
patterns (Fig. 1e, all patients in Fig. S3).

Alignment of cross-patient latent dynamics

Speech trials for each patient were grouped into four categories representing types of articulation
corresponding to the phonemes presented in our stimuli (low: /a/, /e/, high: /i/, /u/, labial: /p/, /b/,
/v/, dorsal: /k/, /g/). For simplicity, we classified trials by the articulator type of the first phoneme
in the spoken sequence, resulting in 12 "low" non-words, 14 "high" non-words, 14 "labial" non-
words, and 12 "dorsal" non-words. There were distinct spatiotemporal patterns of HG activity
specific to each type of articulation (Fig. 2a, all patients in Fig. S4). While patterns differ across
articulators, they also differ across patients. This can be seen across all patients, but we illustrate
this difference for two representative patients (S1 and S2 – Fig. 2a). These patient-specific
differences in spatiotemporal speech representations demonstrate the necessity of specialized
alignment techniques in uncovering a shared representation of speech across patients.
148
To extract latent dynamics from these high-dimensional data, we use principal component
analysis (PCA) as a linear dimensionality reduction tool to decompose HG activity (± 500 ms
around speech onset) across the significant channels into a low-dimensional set of principal
components (PCs) while preserving timing information. Articulatory information encoded in the
latent dynamics can be visualized by further projecting the latent dynamics into a two-dimensional
space using t-distributed Stochastic Neighbor Embedding (t-SNE, see Methods), where
separation between clusters for different articulator types was qualitatively apparent (shown here
for S1 – Fig. 2b). Clustering strength of the latent dynamics in this space was quantified using the
silhouette score (for additional clustering strength metrics, see Fig. S5), which showed
significantly better separation of articulatory feature clusters than in a chance distribution where
ground-truth cluster identities were shuffled (Fig. 2c; Mann-Whitney U test, U = 2500.0, $n_1 = n_2$ =
50, p =7.07e-18). This shows that PCA successfully extracts latent dynamics that are informative
to speech.

The latent dynamics showed distinct trajectories through their respective latent spaces, as
visualized in two-dimensional PC space (S1 and S2: Fig. 2d, left). We sought to functionally align
these latent dynamics to a similar space to train speech decoding models that learn a similar
representation of speech production from multiple patient sources. To model this functional
alignment, we used canonical correlation analysis (CCA) to identify linear combinations of patient-

Figure 2. Alignment of latent dynamics across patients. a. Spatiotemporal activation patterns of HG neural activity (z-scored relative to pre-stimulus baseline) across the µECoG arrays for S1 and S2 grouped by different articulatory gestures. Vocal tract diagrams show visual representations of articulatory category (low, high, labial, dorsal). HG traces below the heatmaps show average activity across all channels for each articulator category. The spatiotemporal HG patterns of these articulatory gestures are distinct across patients, motivating a need for alignment for cross-patient grouping. **b.** Latent dynamics are extracted from HG activity using principal component analysis (PCA). Projection of S1 latent dynamics to a 2D space as shown by t-distributed stochastic neighbor embedding (t-SNE) which qualitatively demonstrates the clustering of latent dynamics by articulator type. **c.** Quantification of articulatory clustering strength in **b** by the silhouette score of points in t-SNE space. The silhouette score for latent dynamics is significantly higher than that of a chance distribution generated by shuffling the ground truth articulator labels (***$p < 0.001$, Mann-Whitney U test). **d.** Latent dynamical trajectories for S1 and S2, as visualized in 2D PC space. The trajectories appear dissimilar. Alignment of S2 latent dynamics to S1 is performed with canonical correlation analysis (CCA). Resultant aligned latent dynamics appear more visually similar. **e.** As in **c**, but with data from both S1 and S2. Without alignment, the latent dynamics articulatory clustering strength is above chance ($p < 0.001$, Mann-Whitney U test) but much lower than the patient-specific S1 mean. With alignment, the articulatory clustering strength is significantly higher than both the unaligned case ($p < 0.001$, Mann-Whitney U test) and chance ($p < 0.001$, Mann-Whitney U test), and much closer to the patient-specific S1 mean. **f,** Representational similarity analysis on cross-patient latent dynamics. Across all patients, cross-patient similarity in speech information (Pearson r between representational dissimilarity matrices) represented by latent dynamics are significantly higher when latent dynamics are first aligned with CCA alignment instead of left unaligned (*$p < 0.05$, Wilcoxon signed-rank test).

specific latent dynamics such that they are maximally correlated across patients. This resulted in
patient-aligned latent dynamics that show more similar trajectories through a common latent
space (shown for S1 and S2: Fig. 2d, right). Repeating our previous technique of clustering
strength on latent dynamics, we found weaker across-patient clustering when these latent
dynamics were unaligned relative to patient-specific clustering, indicating differences in articulator
representations across patients (Fig. 2e). With alignment, clustering strength was significantly
higher than when unaligned (Mann-Whitney U test, U = 2500, n_1 = n_2 = 50, FDR-corrected p =
1.06e-17) and much closer to patient-specific values. As t-SNE operates unsupervised, it is
decoupled from any potential leakage of label information that could occur during alignment.
Therefore, silhouette score quantification of clustering in the t-SNE space of aligned latent
dynamics represents a valid quantification of aligned cross-patient clustering, and shows that
CCA alignment can successfully map latent dynamical representations of speech across different
patients.

To further examine the ability of CCA to align representations of speech across all patients, we
used representational similarity analysis (RSA) to quantify cross-patient similarity in latent
dynamical representations of articulatory information. Specifically, we measured the Pearson
correlation between representational dissimilarity matrices (RDMs) constructed from cross-
patient latent dynamics with and without alignment (Fig. S6; see Methods). To better capture
speech information and task structure in these RDMs, we grouped trials by their full, three-length
articulator sequence (e.g. "labial-low-dorsal") instead of simply by the first articulator in the spoken
sequence. Across all patients, we found that correlation between RDMs significantly improved
when latent dynamics were aligned to a common patient space prior to RDM construction instead
of unaligned in separate patient spaces (Fig. 2f; Wilcoxon signed-rank test, W = 0, n = 8, p = 0.01).
Additionally, we found that alignment does not trivially result in an increased correlation between
articulatory RDMs, as RDMs constructed from aligned latent dynamics with permuted articulatory
labels (and thus no relevant organization of articulatory information) showed significantly lower
correlations than those constructed with ground truth articulatory labels (Fig. S7; Wilcoxon signed-
rank test, W = 0, n = 8, FDR-corrected p = 0.01). To further show that speech-relevant neural
activity is important to cross-patient alignment, we also quantified articulatory representational
similarity in a control condition using surrogate data. Specifically, we replaced the true neural data
with surrogate data generated by the Tensor Maximum Entropy (TME) method, which is randomly
generated with the constraint of preserving the first and second order statistical structure of our
neural data (equal mean and covariance across trials, channels, and time; see Methods). As such,
this surrogate data is random with respect to the speech sequences spoken during trials and
should not encode any relevant articulatory information. Cross-patient representational similarity
from this surrogate data control was significantly lower than that of our original aligned latent
dynamics (Fig. S7; Wilcoxon signed-rank test, W = 0, n = 8, FDR-corrected p = 0.01), indicating
that speech information encoded in the true neural activity is important to successful cross-patient
alignment. These results demonstrate the success of CCA in aligning latent dynamics across
patients to elucidate a shared representation of speech production.

Figure 3. **Cross-patient alignment preserves spatial representations of articulatory features. a.** Spatial map of electrode tuning to different articulator categories for S1 (128 channels), as quantified by the ROC-AUC score from univariate electrode decoding. Different articulators have distinct spatial representations over the array. **b.** Same as in **a**, but for patient S3 (256 channels). Cross-patient projection enabled reconstruction of a source patient's (S3) articulator tuning in a target patient's (S1) space. **c.** (Left) Quantification of Pearson correlation between reconstructed articulator maps from source patient S3 and ground-truth articulator maps for target patient S1. Reconstructed maps are more similar when using CCA alignment than without alignment across all articulator types. Horizontal dashed lines indicate an interval of trivial correlation values, represented by 95% confidence intervals on correlation values from a null distribution of reconstructions performed with permuted CCA weights ($n = 100$). Correlation values above this interval suggest non-trivial alignment to the target articulator map, while correlation values below this interval suggest misalignment, or non-trivial anti-correlation. (Right) Similarity in articulator map reconstructions for all target patients ($n = 8$, each dot is a target patient). For each target patient, reconstructed maps were averaged across all source patients within articulator type to produce a resultant map to compare against the ground-truth articulator map. Reported similarity values for each target patient are the average similarity across all four articulator types. Similarity in reconstructed maps from aligned latent dynamics are significantly higher than those without alignment (***$p < 0.001$, Wilcoxon signed-rank test). **d.** Articulator decoding from cross-patient data. Support vector machine (SVM) decoding models are trained to classify articulator type from latent dynamics. There is no significant difference in accuracy between patient-specific SVMs and SVMs trained on S1's data then tested on data from other patients that have been aligned and reconstructed to S1's space ($p = 0.79$, exact two-sided paired permutation test). Without alignment, cross-patient decoding accuracy is significantly lower (**$p < 0.01$, exact two-sided paired permutation test).

Cross-patient alignment preserves spatial articulator maps

Previous work leveraging the high spatial resolution of μECoG showed micro-scale patterns of spatial tuning to different articulator types[29]. As the alignment of latent dynamics showed that articulator representations could be co-aligned temporally across patients, we hypothesized that spatial tuning to articulatory features would also be preserved following alignment. To investigate this, we first calculated single-electrode tuning to the four articulator categories. For each electrode, we trained a linear discriminant analysis decoding model to classify articulator type. We quantified the tuning of an electrode to an articulator by its receiver-operator-characteristic area-under-the-curve (ROC-AUC) value. Fig. 3a shows the spatial arrangement of electrode-articulator tunings for S1, where distinct spatial maps for each articulator type are visible.

To investigate the cross-patient preservation of these maps, we used CCA alignment to project
spatial articulator information from one patient's space to another. We first used PCA to extract
latent dynamics from two patients and learned a CCA transformation to align them. The
transformations learned in this process enable the projection of data from a source patient's
"electrode-space" to a target patient's electrode-space, which we call cross-patient projection.
Specifically, electrode-level data can be projected to the source patient's latent space with the
source patient's PCA transform, (optionally) aligned to the target patient's latent space, and then
projected to the target patient's electrode-space by applying the inverse of target patient's PCA
transform. We illustrate this analysis with S3 as the source patient and S1 as the target patient
(Fig. 3b). After learning the PCA and CCA transformations, the electrode-articulator tunings for
the source patient (Fig. 3b, left) were projected to the target patient's space. We denote these
projected spatial maps as reconstructed articulator maps (Fig. 3b, right). The Pearson correlation
between ground-truth articulator maps and reconstructed articulator maps was calculated to
quantify similarity in the reconstructed spatial tunings. We found that the similarity between
reconstructed and target articulator maps was higher when cross-patient projection was
performed with alignment than without (Fig. 3c, left). Additionally, we found that the target patient's
articulator maps could be reconstructed with high similarity for certain articulator categories (low,
labial, dorsal) by averaging reconstructed maps across all source patients (Fig. S8 – illustrated
for target patient S1), showing that articulator information in one patient is well-represented across
multiple patient sources. Across all target patients, we found significantly higher similarity between
reconstructed and ground-truth maps with alignment (Fig. 3c, right) than without (Wilcoxon
signed-rank test, $W = 3$, $n = 8$, $p = 0.04$), indicating that cross-patient alignment results in improved
preservation of spatial articulator tuning. To confirm this similarity between ground-truth and
reconstructions of articulator maps with alignment was indeed driven by shared speech
information encoded in the recorded neural activity, we performed a control analysis again using
TME-generated surrogate data in place of our original neural data (Fig. S9). When decomposition
and alignment transformations were learned using this surrogate data that is random with respect
to speech information, we found reduced mean correlations between ground truth and
reconstructed articulator maps with alignment across all patients and articulators (Fig. S9a; mean:
surrogate aligned = -0.09, aligned = 0.16), though with no significant effect (FDR-corrected $p = $
0.07, exact two-sided paired permutation test), which may be due to our limited number of
patients.

We validated the preservation of articulatory information across patients by training support vector
machine (SVM) decoding models to classify articulatory features from patients that had not been
used for training. Specifically, we trained a singular SVM on HG signals (± 500 ms around speech
onset) from one patient and evaluated its accuracy when tested on all other patients projected to
the single patient's space, both with alignment and without (Fig. 3d – illustrated for S1). As a
baseline, we compared cross-patient accuracies to the accuracy of patient-specific SVMs for each
patient. We found both patient-specific and cross-patient decoding on aligned reconstructed data
yielded above chance articulator decoding accuracies (chance = 0.25; mean: patient-specific =
0.39, Wilcoxon signed-rank test, $W = 1$, $n = 7$, $p = 0.047$; mean: aligned reconstruction = 0.43,
Wilcoxon signed-rank test, $W = 0$, $n = 7$, FDR-corrected $p = 0.047$). Additionally, we found no
significant difference between patient-specific and aligned cross-patient decoding accuracies (n

= 7, FDR-corrected *p* = 0.79, exact two-sided paired permutation test), indicating that the aligned
cross-patient SVM performs just as well as the patient-specific SVM despite the separate patient
sources. Without alignment, there was a significant decrease in cross-patient decoding accuracy
relative to the aligned context (mean unaligned reconstruction = 0.23; *n* = 7, FDR-corrected *p* =
0.002, exact two-sided paired permutation test), which was expected due to the dissimilarity found
between reconstructed articulator maps without alignment and ground-truth articulator maps.
There was no significant difference between unaligned cross-patient decoding accuracies and
chance accuracy (chance = 0.25; Wilcoxon signed-rank test, W = 4, *n* = 7, FDR-corrected *p* =
0.11), demonstrating the necessity of alignment in elucidating shared neural representations of
speech. We repeated this aligned decoding analysis with our TME-generated surrogate data to
again confirm that success of the aligned reconstructions was driven by speech information
encoded in the original neural data (Fig. S9b). We observed a significant decrease in cross-patient
decoding accuracies on aligned reconstructed data in the surrogate control relative to that of our
original data (mean: surrogate aligned reconstruction = 0.27, aligned reconstruction = 0.43; *n* =
7, FDR-corrected *p* = 0.01, exact two-sided paired permutation test). Overall, we have
demonstrated that cross-patient alignment preserves important spatial characteristics of
articulatory features in the SMC, enabling successful cross-patient speech decoding.

Expanded cross-patient datasets improve speech decoding accuracy

Given the preservation of speech information in aligned, cross-patient latent dynamics, we
investigated if cross-patient datasets could be used to address data limitations in speech BCI
training and improve their performance. With a selected target patient, additional data from other
patients can be aligned to this target patient and appended to the target datasets, enabling vastly
expanded training datasets relative to patient-specific contexts. To test this, we trained SVM
models on aligned cross-patient latent dynamics during speech (total combined dataset size
across all patients: ~8.2 minutes utterance duration, ~68.5 minutes experiment duration, see
Table S3 for amount of speech data per patient) and we evaluated their accuracy on a held-out
set from the target patient using 20-fold cross-validation. We then compared these results to
patient-specific latent dynamics and unaligned cross-patient latent dynamics. While thus far, our
analyses focused on decoding articulatory features, here we evaluated the accuracy of classifying
distinct phoneme units present within these articulator categories (e.g. /k/, /a/, /b/) and in all three
position of the spoken phoneme sequence to mimic translational BCI use cases where phonemes
are rapidly decoded and strung together into words and sentences.

We found significant differences in decoding accuracy across patient-specific, unaligned cross-
patient, and aligned cross-patient decoding contexts (Fig. 4a; 9-way phoneme decoder, chance
= 0.11; repeated measures ANOVA, $F_{2,14}$ = 12.59, *p* = 0.007). In decoding models trained on
unaligned cross-patient latent dynamics, individual phoneme decoding accuracy was significantly
lower relative to patient-specific decoding (unaligned cross-patient: mean = 0.19, 95% CI = [0.15,
0.24]; patient-specific: mean = 0.24, 95% CI = [0.17, 0.31]; follow-up FDR-corrected paired t-test,
corrected *p* = 0.03; effect size *d* = 0.57). Models trained on aligned cross-patient latent dynamics,

Figure 4. Expanded cross-patient datasets improve speech decoding accuracy. a. Cross-patient phoneme decoding across all eight patients. Each line represents cross-patient decoding aligned to a specific target patient's space. In blue, SVM models are trained and tested on patient-specific data. In red, SVM models are trained on latent dynamics from all eight patients without alignment and tested on held-out patient-specific data from the target patient, resulting in a significant decrease in decoding accuracy (*$p < 0.05$, repeated measures ANOVA and follow-up FDR-corrected paired t-tests). In purple, SVM models are trained on latent dynamics from all patients with CCA alignment and tested on held-out patient specific data from the target patient, improving phoneme decoding accuracy relative to unaligned and patient-specific cases ($p < 0.05$, repeated measures ANOVA and follow-up FDR-corrected paired t-tests). The gray dashed line indicates chance levels of decoding. **b.** Cross-patient decoding for target patients S1, S2, and S3. Aligned cross-patient accuracies were significantly higher than both unaligned and patient-specific decoding contexts in each target patient (***$p < 0.001$, post-hoc Tukey HSD test following one-way ANOVA). **c.** Patient-specific and aligned cross-patient decoding repeated as the percentage of data (trials) in the target patient dataset is subsampled. Accuracies shown here are aggregated across all patients (Fig. S16 for all patients separately). Even with limited target patient data to learn latent dynamics alignment, the cross-patient decoding outperformed patient-specific decoding (**$p < 0.01$ at all subsampling ratios, exact two-sided permutation test). **d.** Aligned cross-patient decoding as the amount of additional cross-patient data (trials) is varied for patient S5. Little additional data was needed to increase the phoneme decoding accuracy beyond patient-specific levels. Diminishing gains in decoding accuracy are observed as additional cross-patient data is added. **e.** Relationship between cross-patient improvement (percent change between aligned cross-patient and patient-specific decoding) and target patient-decoding accuracy. Using a linear fit we found a significant negative relationship ($p < 0.05$, $r = -0.77$), showing cross-patient gain gets smaller as target patient-specific accuracy increases.

however, exhibited strong phoneme decoding accuracy – even outperforming patient-specific
models (aligned cross-patient: mean = 0.31, 95% CI = [0.22, 0.40]; follow-up FDR-corrected
paired t-test, corrected p = 0.01; effect size d = 0.55). Fig. 4b shows cross-patient decoding
accuracies separated by target patient for a few sample patients (results for all target patients in
Fig. S10). One patient (S3) in particular received an immense benefit from this additional cross-
patient data (mean: aligned cross-patient = 0.53, patient-specific = 0.29), which we attribute to

the initial lack of patient-specific data available (S3 had approximately one-third the amount of
patient-specific data as the other patients), so the additional data provided critical repetitions to
enable sufficient training. To validate that these increased cross-patient decoding accuracies
were not due to the alignment procedure artificially inducing structure in cross-patient data that
was not originally present, we generated surrogate data that preserved the first and second order
statistical structure of our neural data (equal mean and covariance across trials, channels, and
time) but was otherwise random (see Methods) and aligned this data to the target patient instead.
We hypothesized that this surrogate neural data contained similar statistical structure but lacked
speech-relevant latent dynamics and thus would fail to decode speech information from the target
patient following alignment to the target patient's space. Models trained on only this aligned
surrogate data yielded accuracies with no significant difference from chance across all target
patients (Fig. S11; Wilcoxon signed-rank test, $W = 8$, $n = 8$, $p = 0.20$), indicating alignment itself
does not artificially yield improved phoneme decoding accuracy. These results show that speech
decoding models can learn a shared representation of phoneme information from separate patient
sources, substantiating the success of cross-patient speech BCIs.

When aligning the latent dynamics of one patient to those of a new target patient, it is unclear
how much new data from the target patient must be collected to enable sufficient alignment to the
target patient's latent space. We expect that this alignment can be learned with minimal amounts
of data – much less than needed to sufficiently train a complex patient-specific decoding model –
as CCA is a linear alignment method. To test this, we repeated our cross-patient decoding
analysis while subsampling the number of trials from the target patient at intervals of 5%
(excluding S3 due to a previously mentioned lack of trials) used for alignment and decoder training
(Fig. 4c, Fig. S16 for all patients separately). Across all patients, we found that CCA yielded
successful alignment that enabled cross-patient speech decoding with significantly higher
accuracies than patient-specific contexts as the subsampling ratio decreased ($n = 8$, $p = 0.008$ at
all subsampling ratios with FDR correction, exact two-sided permutation tests), even for target
patient datasets with only 5% of their original trials (~7 trials/0.5 min.). This demonstrates the
feasibility of cross-patient alignment with minimal data collection from new patients.

To quantify the relationship between the amount of additional cross-patient data and decoding
performance, we repeatedly trained cross-patient SVMs while varying the amount of additional
cross-patient data trials available. For each patient, we randomly subsampled a fixed number of
trials equally from each other patient. We then aligned the subsampled trials to the target patient,
trained a cross-patient SVM model, and evaluated its performance on a held-out test set from the
target patient with 20-fold cross-validation. The results from this analysis for an illustrative patient
(S5) are shown in Fig. 4d (all patients in Fig. S17). We chose to use S5 as the representative
patient here, instead of S1 as we have done previously, as S5 better portrayed the increases in
aligned decoding accuracy as additional cross-patient data was added that we saw across nearly
every other patient. We found patient-specific performance (mean accuracy = 0.17) was
exceeded after only 4.4 minutes of cross-patient data, with cross-patient decoding accuracy
increasing to a mean of ~0.22 (chance = 0.11) with all cross-patient data used. We observed
diminishing gains in decoding accuracy as further cross-patient data was added for S5, as shown
by a linear fit between the log-transformed amount of cross-patient data and aligned cross-patient
decoding accuracy ($r^2 = 0.94$; $p = 3.0e\text{-}11$; Fig. S18). However, this may be an effect of the

simplicity of the SVM models used (relative to state-of-the-art speech decoding models). We
expect that more complex models would be able to better use this additional cross-patient data
to further improve decoding accuracy.

We found varied improvements in aligned cross-patient decoding accuracy by patient. We
hypothesized that this inter-patient variability may be related to the patient-specific decoding
accuracy of the target patient, as patient-specific decoding models in patients with low accuracies
may be insufficiently trained and stand to benefit more from additional cross-patient training
samples. To investigate this, we plotted the relationship between each target patient's patient-
specific decoding accuracy and the percent change in decoding accuracy when trained with
aligned cross-patient data relative to patient-specific data, which we denote the cross-patient
improvement (Fig. 4e). For this analysis, we chose to exclude S3 due to the confound of dataset
size, as it has one-third of the data of the other patients. Using a linear fit, we found a significant
negative relationship between cross-patient improvement and patient-specific decoding accuracy
of the target patient (r = -0.77, p = 0.04), indicating patients with lower patient-specific decoding
accuracy saw a larger benefit to decoding accuracy with cross-patient data. We further
investigated pairwise effects on inter-subject variability using a leave-one-out cross-patient
training strategy (Fig. S19a; again, excluding S3). Specifically, for each target patient, we added
all available cross-patient data to the target patient's training set except for a single held-out
patient and reported the percent decrease in cross-patient decoding accuracy relative to the
accuracy with the full cross-patient training set. This allowed us to investigate changes in cross-
patient accuracy on the level of a single source patient to look for additional drivers of inter-subject
variability in cross-patient accuracy. Using linear regression, we found a significant relationship
between the patient-specific decoding accuracy of the target patient and the left-out change in
cross-patient accuracy (Fig. S19b; r^2 = 0.51, FDR-corrected p = 3.89e-7), indicating that target
patients with higher patient-specific accuracy are more robust to left-out data than target patients
with lower patient-specific accuracy. Surprisingly, we found no significant effect of left-out patient-
specific accuracy (Fig. S19c; r^2 = 0.05, FDR-corrected p = 0.24) or distance between electrode
arrays in neuroanatomical space (Fig. S19d; r^2 = 0.01, FDR-corrected p = 0.64; S4 and S7
additionally excluded to constrain analysis to left hemisphere). These results align with Fig. 4e
and the view that insufficient training of these lower accuracy models results in more benefit from
(or more reliance on) cross-patient data.

Cross-patient alignment improves speech decoding in a simulated real-time environment

Thus far, the presented alignment and decoding analyses were performed in offline decoding
contexts that use non-causal windows of neural activity centered around patient responses.
However, speech BCI systems must operate causally and without access to patient response
timing. As such, the translatability of our cross-patient alignment methodology to more relevant
real-time speech decoding contexts remains unclear.

To investigate the effects of cross-patient alignment on real-time speech decoding performance,
we applied an adapted alignment and decoding strategy on simulated real-time neural recordings
from our intraoperative dataset. We first extracted HG power within short windows of the neural
recording (20 ms, as in previous real-time speech BCI work[7]) for each trial. HG data from a

Figure 5. Cross-patient alignment improves real-time speech decoding performance a. Schematic of simulated real-time speech decoding pipeline. (Left) High-gamma (HG) signals are extracted in 20 ms windows to simulate streaming of real-time features. We use a sliding, causal decoding window (280 ms, 80 ms stride) to predict a timeseries of phonemes from 0.5 to 3.5 seconds following stimulus onset. A decoding model (recurrent neural network) is trained with a combination of patient-specific and aligned cross-patient data. We use connectionist temporal classification (CTC) loss to train the model to predict a sequence of phonemes from the output timesteps. (Right) Phoneme predictions from a sample trial. The top plot shows the timeseries of phoneme output probabilities. The trial starts with blank predictions during silence, then predicts a sequence of three phonemes (/g/, /a/, /b/) after. The bottom diagram shows the predicted sequence (/gab/) compared to the true sequence (/kab/). The mistakenly predicted first phoneme results in a phoneme error rate (PER) of 33%. Despite this mistake, the predicted probability of the /k/ phoneme is nearly that of the /g/ phoneme, indicating confusion between these similar phonemes. **b.** CTC-RNN decoding performance, measuring phoneme error rate between predicted and true sequences. There is no significant difference between chance and patient-specific PERs ($p = 0.81$, FDR-corrected Wilcoxon signed-rank test). However, unaligned cross-patient PERs are significantly lower than patient-specific (*$p < 0.05$, FDR-corrected Wilcoxon signed-rank test) and aligned cross-patient PERs are significantly lower than both patient-specific and unaligned PERs ($p < 0.05$, FDR-corrected Wilcoxon signed-rank test. **c.** PER in patient-specific and aligned cross-patient contexts as the percentage of target patient data (trials) is subsampled, averaged across all patients. Cross-patient PERs are significantly lower than patient-specific PERs for all subsampling ratios ($p < 0.05$, FDR-corrected Wilcoxon signed-rank test). Cross-patient gains remain even with as little as 5% of the original patient-specific dataset. **d.** PER in aligned cross-patient decoding as the amount of additional cross-patient data is varied, averaged across all patients. PER starts at patient-specific values with little cross-patient data, then steadily decreases as more cross-patient data is added. Aligned cross-patient PERs are significantly lower than patient-specific PERs after ~38.8 minutes of cross-patient data are added ($p < 0.05$, FDR-corrected Wilcoxon signed-rank test).

designated training period was used to learn decomposition and alignment matrices between
source and target patients with our PCA-CCA alignment method, which we used to increase the
size of the target patient's training dataset with aligned data from source patients as in Fig. 4. We
then trained a recurrent neural network (RNN) model with connectionist temporal classification[35]
(CTC) loss to output a timeseries of phoneme predictions from causal, sliding windows of HG
power without knowledge of patient speech onset timing (Fig. 5a). To evaluate performance of
the CTC-RNN model with and without cross-patient alignment, we analyzed the phoneme error

rate (PER) between predicted and true phoneme sequences on a held-out test set from the target
patient when the model was trained on patient-specific, unaligned cross-patient, and aligned
cross-patient data.

We again found that decoding models trained on aligned cross-patient data outperformed both
those trained on patient-specific and unaligned cross-patient data (Fig. 5b, all target patients
separately in Fig. S20). Across all patients (excluding S3 due to a limited data amount – see
Methods), we found no significant difference between PER in a chance distribution, obtained from
shuffling label identity across trials, and patient-specific training (mean: chance = 88.4%, patient-
specific = 87.1%; Wilcoxon signed-rank test, $W = 12$, $n = 7$, FDR-corrected $p = 0.81$), likely due
to our limited amount of data paired with the complex CTC-RNN decoding model resulting in
insufficient training. Surprisingly, we found that CTC-RNNs trained on unaligned cross-patient
data achieved significantly lower PERs than in the patient-specific context (mean unaligned cross-
patient = 82.5%; Wilcoxon signed-rank test, $W = 0$, $n = 7$, FDR-corrected $p = 0.02$). This was the
opposite of the effect that we observed in our offline decoding analysis using SVM models (Fig.
4a), suggesting feature transformations within the RNN model may better elucidate shared
patterns across data from unaligned patient sources than simpler SVM models. With models
trained on aligned cross-patient data, we saw a further decrease in PER relative to the unaligned
context across all patients (mean aligned cross-patient = 79.4%; Wilcoxon signed-rank test, $W =$
0, $n = 7$, FDR-corrected $p = 0.02$). We saw particularly strong improvements when using aligned
cross-patient data in S2, with PER decreasing from 84.2% in the patient-specific context to 67.9%
in the aligned cross-patient context. These results confirm that our observed increases in speech
decoding performance following cross-patient alignment translate to real-time decoding contexts.

To quantify changes in CTC-RNN performance with variable amounts of data, we subsampled
target and cross-patient data while repeating our simulated real-time alignment and decoding
method. First, we subsampled the amount of target patient data available to CTC-RNN models
trained on either patient-specific or aligned cross-patient data at intervals of 5% (Fig. 5c, all
patients separately in Fig. S21). We found that aligned cross-patient decoding significantly
outperformed patient-specific decoding for all target data subsampling ratios (Wilcoxon signed-
rank test, $n = 7$, $W = 1$, FDR-corrected $p = 0.03$ for a subsampling ratio of 60%, $W = 0$, FDR-
corrected $p = 0.02$ for all other subsampling ratios). Importantly, even with only 5% of the original
patient-specific data (~5 trials/0.3 min.), aligned cross-patient decoding outperformed patient-
specific decoding. As such, there is little "barrier to entry" required in the data collection before
cross-patient gains are observed. Next, given a full patient-specific dataset (~6.3 min. experiment
duration – see Methods), we evaluated decoding performance as we subsampled the amount of
cross-patient data added to test the necessary amount of cross-patient data needed to outperform
patient-specific decoding (Fig. 5d, all patients separately in Fig. S22). We found that aligned
cross-patient decoding achieved significantly lower PERs than patient-specific decoding after
~38.8 minutes of cross-patient data were added (Wilcoxon signed-rank test, $n = 7$, $W = 0$, FDR-
corrected $p = 0.04$ for cross-patient data > 38.8 min., $p > 0.05$ for lesser amounts of cross-patient
data). To estimate performance as additional cross-patient data is added, we performed a linear
fit between the amount of added cross-patient data and log-transformed PER ($r^2 = 0.98$, $p = 6.6e$-
12, Fig. S23). Using this fit, we would achieve a 25% average PER with 14.6 hours of cross-
patient data and a 10% average PER with 25.3 hours of cross-patient data. These results align

well with learning curves composed entirely of patient-specific data shown in an ECoG speech
BCI in previous literature by Metzger and colleagues[8], suggesting that cross-patient data can be
a valid substitute for long-term collection of patient-specific data.

To ensure the timing required to learn cross-patient alignment transformations is feasible in real-
time contexts, we quantified the time to learn CCA transformation matrices for each target patient.
We found an average time of 5.8 seconds, with up to ~10 seconds maximum, to calculate
alignment between a target patient with dataset size of ~6.3 min. and ~60 min. of cross-patient
data (Fig. S24a). This alignment only needs to be learned once following the collection of training
data for the target patient. As such, this time should be framed in the context of the overall
experiment duration, in which a one-time delay of 10 seconds is negligible compared to the 6.3
min. of prior data collection, meaning that learning CCA alignment can easily be integrated into
data collection paradigms. Further, we have taken the approach of aligning cross-patient data to
the space of the target patient, resulting in no change to real-time prediction latency, as there is
no need to transform the real-time-acquired data from the target patient during evaluation periods
prior to input to the decoding model. However, to account for cases in which the user may wish
to align to a space other than the target patient, thus necessitating a linear transformation by a
learned alignment matrix, we quantified the additional latency required for a linear transformation
of the input data prior to input to the CTC-RNN (Fig. S24b). We found an average timing of 0.04
ms to apply a linear transformation to the acquired target patient data. In comparison to the CTC-
RNN prediction latencies ranging from 1 to 7 ms this was a negligible increase to prediction
latency, especially given the 80 ms latency budget defined by the stride of our decoding window.
Overall, timing requirements of cross-patient alignment via CCA are compatible with requirements
of real-time data collection and latency budgets.

High-resolution and broad coverage are critical to alignment

To fully assess the benefits of high spatial sampling and coverage for cross-patient decoding, we
quantified the spatial sampling of the SMC required to sufficiently elucidate speech-relevant latent
dynamics that can be aligned across patients. We subsampled various spatial characteristics of
our µECoG arrays and calculated both the cross-patient decoding accuracy of SVM models
trained on aligned latent dynamics and the cross-patient representational similarity (via RSA) in
the extracted latent dynamics at each subsampled point. We separately investigated the influence
of subsampling electrode density, array coverage, and electrode contact size.

To investigate the effects of electrode density, as represented by the distance between electrodes
(pitch), we used Poisson disk sampling to randomly sample individual electrodes across the array
while constraining the minimum distance between sampled electrodes to approximate the cortical
sampling of lower-density ECoG arrays. Simulated pitch ranged from 10 mm, the pitch of standard
clinical ECoG devices, to 1.5 mm. To quantify the speech information encoded in the pitch-
subsampled data, we repeated our cross-patient phoneme decoding analysis from Fig. 4 at each
effective pitch value. We observed increases in phoneme decoding accuracy as pitch decreased
(an increase in spatial sampling) in both patient-specific and aligned cross-patient contexts (Fig.

Figure 6. **High-resolution sampling with broad coverage is required for successful cross-patient alignment. a.** Effect of electrode density. (Top) Cross-patient phoneme decoding with varied pitch (distance between electrodes). Accuracy increases across patient-specific and aligned cross-patient decoding contexts as pitch decreases. Aligned cross-patient accuracies were only significantly higher than patient-specific accuracies for effective pitch less than 3 mm ($p < 0.05$ for pitch ϵ [2, 1.5] mm, exact two-sided paired permutation test, *p<0.05). (Bottom) Cross-patient representational similarity, as measured by the Pearson correlation between representational dissimilarity matrices, increases as the pitch decreases. Each line represents a single target patient, where correlations are averaged across all non-target patients aligned to the target patient. **b.** Effect of array coverage. As in **a,** but for subsampled grids of varying size. (Top) Phoneme decoding accuracy similarly increases as grid size increases. Aligned cross-patient accuracies were only significantly higher than patient-specific accuracies for grid sizes greater than 6x12 (~8 mm x 17 mm*; p < 0.05* for grid sizes ϵ [6x12, 7x14, 8x16], exact two-sided paired permutation test). (Bottom) Cross-patient representational similarity increases as grid size increases. **c.** Effect of contact size. As in **a,** but with varying contact sizes obtained by spatially-averaging different amount of neighboring micro-contacts. (Top) Phoneme decoding accuracy increases as contact size decreases. The mean phoneme decoding accuracy of aligned cross-patient models is higher than patient-specific models for contact sizes under 3x3 (~3 mm diameter) however there is no significant difference after correction for multiple comparisons ($p > 0.05$ for all contact sizes, exact two-sided paired permutation test). (Bottom) Cross-patient representational similarity generally increases as contact size decreases.

6a, top; Fig. S25 for all patients separately). Crucially, we found that aligned cross-patient models
significantly outperformed patient-specific models for only pitch values under 3 mm ($n = 8$, FDR-
corrected $p = [0.04, 0.04]$ for pitch ϵ [1.5, 2] mm, respectively, $p > 0.05$ for all other pitch values,
exact two-sided paired permutation test). We additionally found that the representational similarity
between aligned latent dynamics of separate patients increased as the effective pitch decreased
across all patients, with more rapid increases for smaller pitch values (Fig. 6a, bottom). These
results indicate that sufficient sampling of cross-patient latent dynamics necessitates high-density
neural interfaces.

To evaluate the necessity of neural interfaces with broad coverage of the SMC, we selected sub-
grids of electrodes from our full electrode arrays and repeated the analyses in Fig. 5a. Grid sizes
ranged from 2x4 (~2 mm x 5 mm) to 8x16 electrodes (~11 mm x 23 mm). Grid-subsampled
phoneme decoding gradually increased as coverage increased for both decoding contexts (Fig.
6b, top; Fig. S26 for all patients separately). Aligned cross-patient models had significantly higher
accuracies than patient-specific contexts for only grid sizes 6x12 and larger ($n = 8$, FDR-corrected

p = [0.04, 0.04, 0.04] for grid sizes ϵ [6x12, 7x14, 8x16], respectively, *p > 0.05* for all other grid
sizes, exact two-sided paired permutation test), which corresponds to coverages greater than ~8
mm x 17 mm, showing that broad coverage of the SMC is critical for successful sampling of cross-
patient latent dynamics. Similarly, we found that the representational similarity between aligned
cross-patient latent dynamics increased as the size of the subsampled grids increased across all
patients, showing better similarity with broader coverage (Fig. 6b, bottom).

Finally, we quantified the effects of contact size by extracting HG information from recorded
signals averaged together across neighboring electrode contacts to approximate larger contacts.
Contact sizes ranged from 8x8 micro-contacts (~11 mm diameter) to a single micro-contact (200
µm). We found that decoding accuracy increases as contact size decreases for both decoding
contexts (Fig. 6c, top; Fig. S27 for all patients separately). While the decoding accuracy of cross-
patient SVMs was higher than the patient-specific decoding accuracy for contact sizes under 3x3
(~3 mm), there was no significant difference after correction for false discovery rate (*n* = 8*, p >*
0.05 for all contact sizes, exact two-sided paired permutation test). Representational similarity in
aligned latent dynamics generally increased as contact size decreased (Fig. 6c, bottom). These
results suggest that electrode density and array coverage play a more important role in properly
sampling SMC cross-patient latent dynamics during speech.

Discussion

Speech BCIs have recently demonstrated impressive performance in rapidly and accurately
translating neural signals to words in patients with an inability to speak due to neuromotor
disorders. However, these BCIs typically require long periods of patient-specific data collection
that limit their deployability and effectiveness. In this work, we investigated shared, cross-patient
representations of speech production in the SMC to reduce the deployment time of speech BCIs
by enabling cross-patient decoding models. While cross-patient speech decoding has been
previously investigated using transfer learning[14], as of this writing, this is the first study, to our
knowledge, to use neural activity aligned across human patients for speech decoding. Our results
support the idea that cross-patient models can reach sufficient data requirements for advanced
decoding methodologies with minimal data collection from any individual patient and can be easily
transferred across patients, resulting in robust and rapidly deployable speech BCIs that are more
accessible to a broader range of patients.

A novel aspect of this study is the investigation of shared latent dynamics in human speech.
Previously, the alignment of latent dynamics across individuals has been constrained to non-
human primates and rodents performing limb movement tasks[26]. Here, we show this phenomenon
of shared neural activity across individuals extends to the human motor speech system, where
the implications for translational BCI contexts stand to provide substantial benefits for training
machine learning models and accelerating deployment following device implantation. Despite
differences between limb movement and speech production, the presence of shared speech
latent dynamics is a logical extension as both movement contexts pertain to the alignment of low-
level neural motor representations underlying highly-coordinated muscle movements.

We hypothesized that neural population activity underlying speech articulator movement is well-
represented in patients' latent dynamics and shared across patients. This is an important
motivation for the use of functional alignment in our study, as the transformations learned through
our alignment procedure do not simply reflect efforts of domain adaptation between a target and
source patient. Instead, this method aims to exploit the existence of a shared neural manifold of
speech motor control to account for differences in neural sampling across patients that arises
from variability in neuroanatomy and electrode placement. This distinction is critical, as it allows
for principled pooling of neural data across patients while maintaining a consistent decoding
model, and provides a complementary alternative to existing transfer learning strategies in neural
decoding.

To extract latent dynamics from HG activity across many channels, we used PCA as a
dimensionality reduction method to approximate latent dynamics. Further, to perform alignment
between latent dynamics from different patients, we used CCA to learn linear transformations
between latent dynamics. We chose these methods for their simple and interpretable
transformations, where aligned latent dynamics are the result of spatial filters applied to the
original electrode space. However, PCA and CCA are limited because they 1) fail to capture
relevant nonlinear latent representations of cortical neural activity during motor control[24,39] 2) treat
data from different time points as independent observations, which discards potential temporal
information that could be useful in learning a better alignment across patients. Recent dynamical
systems frameworks have been developed that use deep representation learning to elucidate
nonlinear latent dynamics while explicitly modeling temporal information, such as Latent Factor
Analysis for Dynamical Systems (LFADS)[39] and MAnifold Representation Basis LEarning
(MARBLE)[40]. Future studies should investigate nonlinear cross-patient representations of speech
production extracted from these advanced dynamical systems models to determine if these
changes improve alignment quality between patients. However, study designs must be wary of
limited interpretability, relative to linear decomposition methods, with deep learning models.
Additionally, the amount of data per patient required to successfully learn latent dynamics should
be considered in these deep models, as a steep data requirement to learn and align latent
dynamics is antithetical to the desired rapid deployability for translational BCI contexts.

We found that alignment of latent dynamics by CCA preserved spatial properties of speech
information. In many cases, we found strong correlations between ground-truth articulator maps
and articulator maps projected to that patient's space from another source patient's space.
However, we found some cases of negative correlation, and thus misalignment, when
reconstructing articulator maps with weak representations, suggesting this alignment may fail to
properly capture weakly represented speech information. Statistically, CCA alignment is designed
to maximize correlation between cross-patient latent dynamics but does not enforce a one-to-one
spatial correspondence. As a result, reconstruction into an individual patient's electrode space
can introduce distortions, particularly when the shared latent space captures only a subset of the
variance present in the target patient. As described earlier, this may be a failure of the linear latent
dynamics being unable to approximate necessary nonlinear activity to accurately preserve
observed spatial activations[24]. From a neurophysiological perspective, these discrepancies may
reflect true, minor inter-patient variability in speech motor representations. While similar cortical
regions are engaged across individuals, the combined differences in anatomy, electrode

coverage, and functional organization may not be fully explained by a shared motor control
manifold. As such, these observations suggest that while a shared latent structure supports cross-
patient generalization, it may not fully capture all patient-specific features of neural organization.

Regardless, we found that this alignment enabled successful cross-patient decoding, with models
trained on additional cross-patient data consistently outperforming those trained on patient-
specific data alone. In some patients, these improvements were substantial. These analyses,
however, were conducted on a relatively small dataset, combining recordings from eight patients
with limited data per individual. We expect that scaling to larger and more diverse patient cohorts
will further enhance decoding performance, particularly when paired with more advanced models
such as recurrent neural networks (RNNs) and other deep learning approaches.

By systematically varying the composition of the cross-patient training set, we observed significant
inter-patient variability in decoding gains. Notably, the strongest predictor of improvement was the
baseline decoding accuracy of the target patient, whereas neither the identity of source patients
nor the neuroanatomical distance between electrode arrays had a significant effect. However, this
neuroanatomical distance analysis did not consider any effect of cortical regions-of-interest
(ROIs), which we suspect play an important role in finding shared or distinct latent dynamical
representations of speech. It would be difficult to implement such an analysis with this dataset
due to the homogeneity of motor area coverage with our patients' arrays. However, we believe
future work should consider a comprehensive ROI-based analysis with stereo-
electroencephalography (SEEG), as the distributed coverage of SEEG recordings would be well
suited to answer this question, assuming electrode density and sampling within ROIs are sufficient
to extract relevant speech-related latent dynamics. Overall, these findings suggest that cross-
patient alignment provides the greatest benefit when patient-specific models are limited by
insufficient data or lower signal quality, effectively acting as a form of data augmentation that
stabilizes decoding in these settings.

Our CTC-RNN results demonstrate that cross-patient alignment can substantially improve real-
time speech decoding performance in contexts that more closely reflect operational BCI systems.
Using simulated real-time µECoG recordings and causal sliding windows of HG activity, we found
that recurrent neural network models with CTC loss trained on aligned cross-patient data
consistently outperformed both patient-specific and unaligned cross-patient models. Importantly,
these benefits were observed even with minimal amounts of target patient data, indicating that
cross-patient alignment can reduce the data collection burden required to achieve accurate
decoding. Performance improvements scaled predictably with the amount of aligned cross-patient
data, and log-extrapolation to long-term dataset sizes suggested that cross-patient data could
serve as a viable substitute for extensive patient-specific training.

Critically, the latency required to compute alignment transformations and apply them to incoming
neural data is minimal relative to the overall experimental timescale and decoding window,
demonstrating that this approach is compatible with real-time BCI constraints. These findings
suggest that cross-patient alignment provides a practical and effective means to leverage shared
neural structure across individuals, stabilizing decoding in low-data regimes while maintaining
real-time feasibility. Together, these results underscore the potential of cross-patient approaches

to accelerate BCI deployment, reduce patient-specific training requirements, and support robust
speech decoding in clinically relevant contexts.

By subsampling various spatial characteristics of our µECoG arrays, we found that neural
interfaces with both high resolution and broad coverage of the SMC are best suited to align latent
dynamics and train cross-patient speech decoding models. Neural interfaces in current speech
BCIs[7,8] typically prioritize only one of these characteristics, as intracortical microelectrode arrays
offer high electrode density but limited coverage and ECoG grids offer wider coverage but with
limited spatial sampling. While our results suggested potentially limited generalization to non-
µECoG neural interfaces because of degraded sampling of the neural signal, previous work by
Safaie and colleagues has shown successful alignment of motor latent dynamics across multiple
individuals from neural recordings with small intracortical microelectrode arrays[26], showing
feasibility with reduced coverage. Additionally, future studies should consider alignment across
neural interface types. As cross-patient alignment is only calculated on dimensionality-reduced
latent dynamics, alignment could still be successful with a heterogeneity of neural interfaces
across patients, which would increase data availability for cross-patient alignment. Nonetheless,
our current findings motivate the use of µECoG for cross-patient speech decoding as it enables
both dense and broad sampling.

It is important to note that the current study is limited to the analysis of cross-patient latent
dynamics during overt speech in able-bodied speakers. These patients differ functionally from
individuals with impaired speech function due to neuromotor disorders like ALS or brainstem
stroke, where neural signal related to speech may be weaker, due to potential degradation of
cortical structures necessary for speech production and monitoring[41,42], or organized differently
than those with intact speech as a whole due to compensatory reorganization of neural motor
pathways[43,44]. As such, it remains unknown if the preservation of speech latent neural dynamics
that we observed here (in individuals without speech dysfunction) will be present across
individuals with varying levels of motor system impairments. However, recent speech BCI studies
have found that articulatory representations of speech may be well preserved in patients with
impaired speech many years after diagnosis or injury with ALS[7] or brainstem stroke[8], respectively,
suggesting that the shared articulatory latent dynamics we observed here may indeed be relevant
to the translational development of speech BCIs.

Together, our results demonstrate that speech-relevant neural population dynamics are shared
across human patients, providing a foundation for more efficient and scalable speech BCI
systems. With cross-patient alignment based on latent dynamics, future speech BCIs may
achieve faster adaptation with minimal patient-specific data, resulting in improved patient usability
and performance. Most importantly, cross-patient speech BCIs will improve communicative ability
and quality of life for individuals with paralysis from neuromotor disorders.

Methods

All our research studies were approved by the Institutional Review Board of the Duke University
Health System under the following protocol ID:

Pro00072892: Studying human cognition and neurological disorders using μECoG electrodes.

Participants

This study included neural data collected intraoperatively from eight awake patients (mean age =
57.4, one female) with fully intact speech at the Duke University Medical Center (DUMC).
Intraoperative recordings were performed while patients S1, S2, S4, and S5 underwent surgical
interventions for movement disorders by deep brain stimulator (DBS) implantation and patients
S3, S6, S7, and S8 were undergoing brain tumor resection. All subjects were native English
speakers. Written informed consent was obtained prior to surgery in accordance with the
Institutional Review Board of the Duke University Health System.

Intraoperative neural recording

Intraoperative neural recordings were performed using custom-designed micro-
electrocorticographic (μECoG) arrays that were fabricated by Dyconex Micro Systems
Technology (Basserdorf, Switzerland). Recordings were performed during awake neurosurgical
procedures for movement disorders (deep brain stimulator implantation – DBS) and tumor
resection. Two different μECoG designs were used. First, for DBS implantation surgeries, a
narrow (11 mm x 21 mm) μECoG array with 128 micro-contacts (200 μm contact diameter, 1.33
mm center-to-center spacing between contacts) was inserted through a burr hole and placed in
subdural space over the target sensorimotor cortex (SMC). For tumor resection surgeries, larger
(38 mm x 21 mm) μECoG arrays with 256 micro-contacts (200 μm contact diameter, 1.72 mm
center-to-center spacing between contacts) were placed subdurally, directly over the SMC
through a craniotomy window. Both array types used gold electrodes, with a platinum-iridium
coating (in all patients except S1), embedded in a liquid crystal polymer thin-film (LCP-TF) substrate
(Impedance information for all arrays in Fig. S2). Arrays were sterilized with ethylene oxide gas
prior to surgical implantation. μECoG arrays were connected to custom headstages using Intan
Technologies RHD chips to amplify and digitize recorded neural signals. These headstages were
then connected to an Intan Technologies RHD recording controller via micro high-definition
multimedia interface (μHDMI) cables. Recorded neural data were processed by the Intan RHX
v3.0 data acquisition software (Intan Technologies, Inc.). Recordings were analog filtered
between either 0.1 Hz and 7,500 Hz or 0.1 Hz and 10,000 Hz. Recordings were digitized at 20
ksps.

To visualize μECoG array location for each patient, electrode locations were co-registered with
patient cortical reconstructions that were generated from preoperative MRI images using
Freesurfer's *recon-all-clinical* function[45]. For DBS implantation surgeries, array location was
confirmed with an intraoperative CT scan following recording that was aligned to the reconstructed
preoperative MRI volume. Electrodes were localized from the CT scan using BioImage Suite[46].
For tumor resection surgeries, array location was registered to the preoperative MRI with Brainlab
Neuronavigation (Munich, Germany) coordinates marked at array corners. Brainlab coordinates
were imported into BioImage Suite, where intermediary electrode locations were interpolated to
match the known electrode spacing of the electrode array (due to missing Brainlab coordinates
for subject S8, electrode location was manually determined based on intraoperative images pre-

and post-array placement). In both DBS and tumor resection cases, electrode locations were
subsequently projected to the leptomeningeal surface of the brain. Electrode arrays of all patients
were visualized together by projection to a common brain (MNI-152, Fig. 1) and separately on
patient-specific brains (Fig. S1).

Speech task

Patients performed a speech repetition task in the acute intraoperative neural recording setting.
In a single trial, patients are presented with an auditory stimulus representing one of 52 unique
non-words via a computer speaker and asked to verbally repeat the sounds they hear. Non-words
were sequences of three phonemes designed to sample the phonotactics of American English.
Sequences were composed of three of nine possible phonemes (four vowels: /a/, /e/, /i/, /u/ and
five consonants: /b/, /p/, /v/, /g/, /k/) arranged in consonant-vowel-consonant (e.g. /k/, /a/, /b/) or
vowel-consonant-vowel (e.g. /a/, /b/, /e/) format (See Table S2 for all stimuli). Trials were
composed of stimulus presentation (stimulus duration: ~0.5 s) followed by a three second
response window with 250 ms jitter in between trials. Patients responded ~1.1 s after the
beginning of the stimulus presentation, with their response lasting ~0.45 s. Trials were presented
across three consecutive blocks (52 trials per block for each unique non-word token). Each block
corresponded to ~0.5 min. of utterance duration. Stimuli order was randomly shuffled within
blocks. Trial start times were marked using flashing lights recorded by a photodiode on the task
presentation laptop (Thorlabs, Inc.). Patient responses were recorded with a lavalier microphone
(Movophoto, Inc.) connected to a pre-amplifier (Behringer). Both photodiode and microphone data
were sent via a BNC cable/connector to auxiliary channels on the Intan RHD recording controller
for synchronization with recorded neural data at 20 ksps. The speech task was designed using
Psychtoolbox in MATLAB R2014a. Phoneme labels (identity, start time, stop time) for both
auditory stimuli and patients responses were first automatically generated using the Montreal
Forced Aligner[47] and then manually corrected using the Audacity audio-editing software[48].

Neural data preprocessing

723 Recorded neural data was decimated to 2 ksps using a finite impulse response anti-aliasing filter
724 (Kaiser window, MATLAB's *resample* function) and common-averaged-referenced (CAR) to
725 remove common noise across channels. Electrodes with large impedance values (>1 Mohm)
726 were excluded from CAR. After CAR, trials with large transient activity (>50 µV between
727 timepoints) were discarded. All preprocessing was done in MATLAB R2021a.

Spectrogram visualization

Spectrograms were calculated for each electrode to visualize spectrotemporal neural activity
during patient responses (Fig. 1d-e). Spectral information was extracted using 500 ms windows
with a 50 ms step-size and 10 Hz frequency smoothing using a multi-taper time-frequency
analysis method[49]. Spectrograms during patient responses were normalized by a baseline period
500 ms prior to auditory stimulus presentation by dividing response spectrograms by the average
baseline power-spectral density to remove *1/f* broadband activity. Normalized response

spectrograms were averaged across trials for each electrode and the log-power was calculated
around patient response onset (-1 to 2 s) to yield neural activity in units of decibels.

High-gamma extraction

The high-gamma band (HG: 70-150 Hz) of the neural signal was extracted to estimate neural
population dynamics in the SMC. HG signals have previously been shown to encode rich speech
production information[8,19,31] and strong correlations with neuron spiking activity[32–34], suggesting a
valid estimation of speech-relevant SMC population dynamics. Following preprocessing, neural
signals were bandpass filtered using frequency-domain Gaussian filters centered at eight
logarithmically spaced center frequencies (with logarithmically scaled bandwidths around each
center frequency) between 70 and 150 Hz[50]. Specifically, the neural timeseries were decomposed
into their corresponding frequency representations with the fast Fourier transform (FFT) and
multiplied in frequency space with a Gaussian kernel centered around each center frequency to
select and smooth information from the frequencies in the desired band. The inverse FFT was
then performed to convert bandpass filtered signals at each band back to the time domain.
Envelopes of signals in each band were calculated as the absolute value of the Hilbert transform
and subsequently averaged across the eight bands. The resultant timeseries was then
downsampled to 200 Hz with a finite impulse response anti-aliasing filter (Kaiser window,
MATLAB's *resample* function). For visualization of HG activity across arrays (Fig. 2a), HG activity
was z-score normalized to the baseline period (500 ms prior to auditory stimulus presentation) for
each trial. For all other analyses (latent dynamics extraction, decoding), HG activity was
normalized by subtracting the average HG envelope of the baseline period. Analyses in this work
used HG activity in a one second window (-500 ms to 500 ms) around patient response onset.
Channels with significant speech HG activity were determined by statistical comparison between
response and baseline periods (see *Statistical Analysis*). HG extraction was performed in
MATLAB R2021a.

Generation of surrogate data

As a control condition for many analyses, we repeated these analyses using data generated from
the Tensor Maximum Entropy (TME) procedure[51] to for each patient instead of their true data. In
the following text, we briefly adapt the description of the method as presented by Elsayed and
Cunningham. TME-generated surrogate data preserves the second-order statistics (mean and
covariance) of the data with respect to each dimension of the data (trials, time, and channels),
while otherwise being maximally random. To perform this, the method generates a probability
distribution *p* that has maximum entropy *H* under the constraints of equal mean and variance to
the original data for each dimension. That is, for surrogate data D_{surr} composed of *s* trials, *t*
timepoints, and *n* channels (shape = (*s, t, n*)), we have:

$D_{surr} \sim p, D_{surr} \in \mathbb{R}^{s,t,n}$ (sampling of surrogate data)
$E_p[D_{surr}] = M = [\mu_s, \mu_t, \mu_n]$ (preservation of original data means)
$\bar{D}_{surr} = D_{surr} - M$ (mean subtraction for covariance equations below)
$E_p[\sum_j^t \sum_k^n \bar{D}_{surr}(:,j,k) \ \bar{D}_{surr}(:,j,k)^T] = \Sigma_s$ (preservation of original data covariance in trials)
$E_p[\sum_i^s \sum_k^n \bar{D}_{surr}(i,:,k) \bar{D}_{surr}(i,:,k)^T] = \Sigma_t$ (preservation of original data covariance in time)

$E_p\left[\sum_i^s \sum_j^t \overline{D}_{surr}(i,j,:)\overline{D}_{surr}(i,j,:)^T\right] = \Sigma_n$ (preservation of original data covariance in channels)

$\hat{p} = \text{argmax}_p H(p) = \text{argmax}_p - \int p(S)\log(p(S))dS, \int p(S)dS = 1$ (maximum entropy constraint)

Where μ_x and Σ_x are the mean and covariance for dimension *x* of the data (trials, time, channels),
respectively, $E_p[.]$ is the expectation over probability distribution *p,* and $\hat{p}$ is the optimal probability
distribution to sample surrogate data, which is essentially a 3D Gaussian distribution with
standard deviation controlled by the covariance preservation and maximum entropy constraints.

As these constraints only impose structure with respect to covariance in each dimension, and
randomness elsewhere, latent dynamics extracted from TME data should not encode speech-
relevant information. As such, our control analyses allow us to evaluate alignment without the
effect of speech information while still controlling for important aspects of the neural signal such
as autocorrelation effects and correlation between channels. Surrogate data generation was
performed with code from https://github.com/gamaleldin/TME in MATLAB R2021a.

Latent dynamics extraction

We extracted latent dynamical activity from estimated SMC population activity as measured in
HG activity around response onset for each electrode and trial (data array of the dimensions – *s,*
t, n), where *s* is the number of trials, *t* is the number of timepoints, and *n* is the number of significant
electrodes/channels). We used principal component analysis (PCA, scikit-learn[52]) to reduce the
high-dimensional channel space into a low-dimensional space composed of *k* principal
components (PCs), where each PC represented a linear combination of activity across the *n*
channels. *k* was determined as the number of components explaining 90% of the variance in the
original data. This yielded patient-specific latent dynamics (data array of the dimensions – *s, t, k*)
as the projection of the original HG activity on the *k*-dimensional latent space. Extraction of latent
dynamics was performed in Python 3.10.

Clustering of articulatory features in the latent space was visualized by further projecting latent
dynamics into a two-dimensional subspace (Fig. 2b) using t-distributed Stochastic Neighbor
Embedding (t-SNE, scikit-learn, perplexity = 30), a widely used technique for visualization of high-
dimensional data[53]. t-SNE decomposition of latent dynamics yielded a data array of dimensions
(*s*, 2). Visualization was performed by plotting each trial as a point in 2D t-SNE space and coloring
each trial point by its articulatory category. Quantification of clustering strength was performed by
calculating the Silhouette Coefficient (scikit-learn, distance = Euclidean) for each trial in t-SNE
space, which measures the difference between inter- and intra-cluster distances[54]. The score is
more positive when the inter-cluster distance is greater than intra-cluster distance. The reported
silhouette score was calculated as the average of all positive-valued Silhouette Coefficient trials[29].
A distribution was generated by calculating silhouette scores across many iterations (*n* = 50) of
projection to 2D t-SNE space. A chance distribution was generated by repeating the silhouette
score analysis for latent dynamics with randomly permuted articulator labels. Clustering with
additional metrics is shown in Fig. S4.

Alignment of latent dynamics

Single unaligned latent dynamic trajectories were visualized for S1 and S2 by plotting the first two
PC values at each timepoint during the speech utterance window, averaged across all trials (Fig.
2c). We refer to these patient-specific latent dynamics as "unaligned", as there has been no
explicit functional alignment of these potentially different patient latent spaces to improve similarity
between them. Alignment of these latent dynamics was performed using canonical correlation
analysis (CCA) to find linear combinations of the latent dynamics PCs that resulted in maximal
correlation between the latent dynamics of the two patients. CCA alignment is performed in a
similar fashion as in[25], which is described below and generally follows MATLAB's *canoncorr*
function. Alignment of latent dynamics was performed in Python 3.10 with numpy.

Starting with the latent dynamics of patients S1 (s_1, t, k_1) and S2 (s_2, t, k_2), we require
correspondence between trials s_1 and s_2 across these patients. As patients may be missing
different trials due to experimental noise or outlier removal, we enforced uniformity in trials by
condition averaging the trials into categories representing the unique phoneme sequences (s_{ca},
at most 52 unique sequences) presented during the task and sub-selecting only the phoneme
sequences present in both patients (i.e. if S1 has no trials for the sequences */abe/*, trials for */abe/*
in S2 will not be considered for alignment). Condition-averaged trials were folded together with
the time dimension to create latent dynamics matrices L_1, L_2 of dimensions (s_{ca}*t, k_1) and (s_{ca}*t,
k_2), respectively. We chose to condition average by phoneme sequence to enforce that the same
phoneme sequences should have similar latent dynamics trajectories. We enforce this constraint
by organizing L_1 and L_2 such that the same row/sample in each matrix corresponds to the same
condition and timepoint. That is, we perform a single CCA fit between each pair of patients, where
our condition constraint is enforced by matrix construction of L_1 and L_2. Note that while label
information is used to condition-averaged trials to create correspondence across patients, label
information is not used in the following learning of CCA transformations to map between patient
latent spaces.

Let d be the minimum rank of L_1 and L_2. CCA alignment first centers L_1 and L_2 by mean
subtraction. Following, QR decompositions are calculated for both L_1 and L_2 as

$$L_1 = Q_1 R_1,$$
$$L_2 = Q_2 R_2 ,$$

where Q is an orthonormal basis for the corresponding latent dynamics. Correlations between the
orthonormal bases are represented by the inner product matrix $Q_1{}^T Q_2$. Singular value
decomposition on this matrix yields

$$Q_1^T Q_2 = USV^T,$$

where U and V are the left and right singular vectors, respectively, which define the
transformations of Q_1 and Q_2 to a shared, cross-patient space with maximal pairwise correlation.
The U and V matrices (truncated to sizes (k_1, d) and (k_2, d), respectively) can be better understood
in the context

$$M_1 = R_1^{-1} U,$$

$$853 \qquad\qquad\qquad\qquad M_2 = R_2^{-1}V,$$

854   where M_1 and M_2 are projection matrices mapping L_1 and L_2 to the shared, cross-patient space,
855   respectively. Transformation is performed via the matrix multiplication

$$857 \qquad\qquad\qquad\qquad L_1^a = L_1 M_1,$$
$$858 \qquad\qquad\qquad\qquad L_2^a = L_2 M_2,$$
$$856 \qquad\qquad\qquad\qquad L_{2\to1}^a = L_2 M_2 M_1^{-1},$$

where L_1^a *and* L_2^a are the aligned latent dynamics of S1 and S2, respectively, where the canonical
correlations between these are the diagonal elements of S above. L_1^a *and* L_2^a are aligned in a
cross-patient space that requires transformations of both input patients. In this work, we instead
choose to align patients to the space of a desired target patient to enable alignment from any
number of target patients to a single common space. The alignment of S2's latent dynamics to
S1's space, denoted $L_{2\to1}^a$, is calculated by first aligning S2 to the common cross-patient space
and applying the inverse of S1's projection matrix (M_1) to map to S1's space. As such, multiple
patients can be aligned to S1's space by first calculating pairwise CCA alignment and projecting
all patients to S1's target space.

The clustering analysis with silhouette score quantification mentioned above was repeated on a
dataset composed of both S1's and S2's data (Fig. 2d). Without alignment, latent dynamics were
separately extracted from S1 and S2, concatenated (truncating the number of PCs in the patient
with higher-dimensional latent dynamics to ensure consistency in dimensions for concatenation),
and reduced to a 2D space with t-SNE. While we refer to these latent dynamics as "unaligned",
referencing a lack of explicit functional alignment between patients, the latent dynamics are
effectively "variance-aligned", in that there is assumed correspondence between PCs that explain
the same relative levels of variance with each patient-specific latent space (e.g. PC 1 in S1 is
assumed to correspond with PC 1 in S2). With alignment, S2 latent dynamics were aligned to
S1's space, aligned latent dynamics were concatenated, and t-SNE decomposition to a 2D space
was applied. In both cases, silhouette score quantification was identical to the patient-specific
case.

Representational similarity analysis (RSA)[55] between unaligned and aligned patients was
performed to quantify shared representations of speech production in aligned latent dynamics
across all patients. For a given patient, we first construct a representational dissimilarity matrix
(RDM). The latent dynamics from that patient (shape=(s, t, k)) are grouped by condition
(articulator sequences in this analysis, e.g. "labial-low-dorsal" representing a sequence of three
articulators) and averaged within these conditions to create a 2D data matrix of shape (s_{ca}, t^*k),
where latent components and time have been folded together to a single dimension representing
latent dynamics. For each pair of conditions, we calculate the Pearson correlation (r) between
these flattened representative latent dynamics and represent their dissimilarity as $1 - r$. The RDM
is then constructed from these pairwise dissimilarities between different conditions within a patient
(Fig. S6). Between a given source and target patient, each with RDMs constructed from latent
dynamics, we calculate the representational similarity between the latent dynamics of these two
patients as the correlation between their RDMs, indicating similarities in how articulatory speech

information is represented in their respective latent dynamics. For each target patient, we quantify
the average representational similarity between all source patients when RDM construction is
performed using unaligned, patient-specific latent dynamics and latent dynamics aligned from the
source patient to the target patient (Fig. 2f). We additionally calculated representational similarity
when aligning surrogate data that is random with respect to speech (see *Generation of surrogate*
data) and when labels were randomly permuted before RDM calculation (Fig. S7). This RSA
analysis   was   performed   in   Python   3.10   using   the   *rsatoolbox*   package
(https://github.com/rsagroup/rsatoolbox).

Single-electrode articulator tuning

Univariate tuning of electrodes to different articulator types was performed similar to Duraivel et
al. (2023)[29] (Fig. 3a). Briefly, for each patient and each electrode in that patient's µECoG array,
we trained a PCA-LDA (linear discriminant analysis) decoding model to predict articulator type
from the timeseries HG data for the current electrode. This model first projected input data to a
low-dimensional subspace explaining some percentage of the variance with PCA, then learned
to classify articulator type from low-dimensional features. Evaluation and training were performed
with 5-fold nested cross-validation. The outer folds, splitting training and testing data, were used
to measure performance as the receiver-operator-characteristic-area-under-the-curve (ROC-
AUC) for the binary classification of each articulator type. ROC-AUC scores were average across
all 5-test folds within articulator type. The inner folds, splitting training and validation data, were
used to optimize the PCA variance percentage hyperparameter. Grid search was performed on a
one-dimensional vector of PCA variance percentages (10% to 100%, 5% step size) and the best
performing percentage across all validation folds was used in the current test set fold. Maps for
each articulator type (trial counts: low = 12, high=14, labial=14, dorsal=12).   and patient were
generated by repeating this process for all individual electrodes. Single-electrode articulator
tuning was performed in Python 3.10 with the scikit-learn library.

Cross-patient projection

To project single-electrode articulator maps from a source patient's space to a target patient's
space (Fig. 3b), we first learned decompositions from HG signals to latent dynamics with PCA
and pairwise alignment with CCA for each patient as described above (*Alignment of latent*
dynamics), but using HG signals from all channels instead of only significant channels. We then
applied these transformations to spatially decompose, optionally align, and spatially reconstruct
articulator maps across patients. We note that alignment is learned from patient HG signals, not
ROC-AUC scores, presenting no conflict of data leakage when projecting cross-patient articulator
maps composed of these ROC-AUC values. The following equation summarizes this cross-
patient projection process,

928
$$R^{S \to T} = (R^S - \bar{R}^S)M_{PCA}^S(M_{CCA}^{S \to T})M_{PCA}^{T(-1)} + \bar{R}^S$$

where $R^{S \to T}$ is the source patient's articulator map projected to the target patient's space, R^S is
the source patient's articulator map, $\bar{R}^S$ is the mean value of the source patient's articulator map,
M_{PCA}^X is the PCA decomposition for patient X (source or target), $M_{CCA}^{S \to T}$ is the source-to-target CCA

alignment matrix that can optionally be applied to perform cross-patient projection with alignment.
When performing unaligned articulator map reconstructions, set $M_{CCA}^{S \rightarrow T} = I$. We also provide the
following text description of this process for additional context. Decomposition was performed by
passing flattened articulator maps through the source patient's PCA. For compatibility between
this PCA, which expects data centered around zero, and ROC-AUC values in the range [0.5, 1],
we subtracted the mean ROC-AUC value across all channels to de-bias the ROC-AUC values.
This enabled articulator maps to be scaled to proper feature ranges for transformation by the
previously learned PCA. When aligned reconstructions were performed, alignment transformation
matrices were applied to align the latent source patient articulator maps to the target patient's
latent space. With alignment, transformation to a different sized latent space is handled
automatically. Without alignment, we enforced that each PCA used the same number of
components (n = 30) instead of the number of components explaining 90% of the original variance
for dimensionality consistency. Finally, the PCA transform from the target patient was inversely
applied to map (optionally aligned) latent articulator maps up to the target patient's electrode
space. The mean ROC-AUC across all source patient channels (subtracted prior to transformation
via source patient PCA) was then added back in to return ROC-AUC values to the proper domain.
Fig. 3b (right) visualizes the reconstructions of S1 articulator maps by S3. Fig. S6 shows the
reconstructed S1 articulator maps averaged across all source patients. We note that this
reconstruction does not consider any neuroanatomical information in the projection. It is purely a
reconstruction in electrode-level space, where we are interested in similarities between the
reconstructed and ground-truth maps on an electrode-to-electrode level.

To compute the similarity between ground-truth and reconstructed articulator maps (Fig. 3c), the
Pearson correlation was computed between ground-truth and reconstructed articulator maps.
This is analogous to computing the cosine similarity between mean-subtracted ground truth and
reconstructed articulator maps. Fig. 3c (left) quantifies the similarity for reconstructions of S1
articulator maps by only S3. In Fig. 3c (right), for each target patient, articulator maps from all
source patients were considered by separately reconstructing these maps and averaging the
maps across source patients as a single reconstructed map per articulator type prior to calculating
similarity. We additionally calculated the correlation between reconstructed and ground-truth
articulator maps when projecting cross-patient surrogate data (see *Generation of surrogate data*)
that is random with respect to articulatory information (Fig. S9a).

We quantified the articulator decoding performance of models tested on aligned and unaligned
reconstructed data (Fig. 3d). All decoding models used were PCA-SVM (support vector machine)
models. This combined a PCA layer for feature size reduction (as in PCA-LDA models described
above) with a bagged SVM classifier (scikit-learn, C=1, gamma=(n_features*feature_variance)$^{-1}$,
balanced class weight) using a radial basis function kernel. No additional feature scaling beyond
PCA reduction was performed prior to SVM input. SVM PCA explained variance was set at a fixed
80% value. Articulator decoding performance (4 classes: low, high, labial, dorsal; chance = 0.25)
was quantified as the balanced accuracy across predictions from all test folds with 20-fold cross-
validation. The articulator identity for the first phoneme of the repeated three-phoneme sequence
was predicted by the models. First, we trained baseline patient-specific decoding models for each
patient, where both training and testing data were composed of data from a single patient. We

then projected HG data from each source patient's (S2-S8) to S1's electrode space by inversely
applying S1's PCA transformation to the source patient's latent dynamics, either unaligned or
aligned to S1's space. Cross-patient decoding was performed by applying the decoding model
trained on S1's data to each source patient's reconstructed HG data as test data. We additionally
performed cross-patient articulator decoding when projecting cross-patient surrogate data (see
Generation of surrogate data) that is random with respect to articulatory information (Fig. S9b).
Similar to above, cross-patient decoding was performed by applying the decoding model trained
on S1's real data, to each source patient's surrogate data reconstructed in S1's space as test
data.

Cross-patient phoneme decoding

We trained PCA-SVM decoding models (as described above in *Cross-patient projection*) to
predict phoneme (9 classes: */a/, /e/, /i/, /u/, /b/, /p/, /v/, /g/, /k/*; chance = 0.11) identity in a target
patient from combined cross-patient data across all patients in our dataset (Fig. 4a). Parameters
and evaluation methods are identical to those defined above, but with phonemes as the labels
instead of articulators. Phonemes were predicted at all three positions of the phoneme sequence.
Phoneme decoding accuracy was reported as the balanced accuracy across predictions from all
20 test folds across 50 iterations to generate a distribution across different training and testing
splits. Cross-validation folds were generated across the trial dimension of our data. For results
aggregated across all patients (Fig. 4b), the average balanced accuracy across all 50 iterations
was reported as a single value per patient. For all decoding contexts (patient-specific, unaligned
cross-patient, aligned cross-patient), training and test splits were only performed on the target
patient. That is, in cross-patient contexts, cross-patient data was only aligned to the target
patient's training data (when alignment was performed) and added to the training set, such that
model accuracy was quantified on held-out data from a single target patient, as would be the
clinical use-case for cross-patient speech BCIs. This alignment to only the training set of the target
patient additionally prevented data leakage between the training and testing data. Only HG data
from significant channels were used for cross-patient phoneme decoding. Cross-patient phoneme
decoding was performed in Python 3.10, primarily with the scikit-learn library.

We started by calculating the phoneme decoding accuracies of patient-specific models for each
target patient as a baseline. Models were trained on unaligned cross-patient data by first
separately decomposing all patient HG data to latent dynamics explaining 90% of the variance of
the input HG data with PCA. For the target patient, PCA decomposition was learned on the
training set and applied to the test set to avoid information leakage. Unaligned latent dynamics
across all patients were concatenated as additional observations/trials into a single dataset
(truncated to the minimum number of PCs across all patients for consistency of latent
dimensionality). A PCA-SVM decoding model was then trained to predict phoneme identity from
the expanded cross-patient training dataset. As mentioned above, phoneme decoding
performance of the PCA-SVM was quantified on the test set containing only data from the target
patient. Models were trained on aligned cross-patient data by repeating the same process for
unaligned cross-patient data and applying CCA alignment after decomposing all patient data to
latent dynamics. CCA alignment was calculated between each source patient and the target
patient to align all source patient data to the same target patient space (this was additionally

beneficial as no additional transformation of the test data in the target patient's space was
required). As with PCA, CCA alignment was only learned between the training set of the target
patient and cross-patient sources to prevent information leakage between training and testing
sets. Fig. 4b shows cross-patient phoneme decoding results S1, S2, S3. The results across all
patients are shown in Fig. S7.

As a control condition for cross-patient decoding, we aligned surrogate data that is random with
respect to phoneme information (see *Generation of surrogate data*) to each target patient instead
of true data from other patients (Fig. S11). Surrogate data were generated for each of the eight
patients, where seven surrogate patients were aligned to true target patient data for each target
patient. Alignment and subsequent decoding were done the same as with all true data.

For subsampling of target patient data (Fig. 4c, Fig. S9 for all patients separately), patient-specific
and aligned cross-patient phoneme decoding were performed as described above while a
percentage of the target patient's training trials were subsampled prior to alignment (for cross-
patient models) and decoding. Subsampling was performed by randomly and uniformly sampling
trials from the target patient's training dataset until the desired number of trials were obtained.
Reported phoneme decoding accuracy is the average balanced decoding accuracy for each of
the eight target patients.

For subsampling of cross-patient data (Fig. 4d, Fig. S10 for all patients separately), aligned cross-
patient models were trained as described above while varying the amount of cross-patient data
aligned to the target patient. We equally subsampled n trials from each patient, where n ranged
from 5 trials to the maximum number of trials from the current patient with a step size of 25 (i.e.
5, 30, 55, …). Cross-patient trials were sampled equally from each patient to avoid effects of
patient identity during sampling. Each point reports the phoneme decoding accuracy across 50
iterations of sampling. Fig. 4d visualizes these results for target patient S5. The average patient-
specific decoding accuracy for S5 is overlaid for comparison. To show diminishing gains in
decoding accuracy as the amount of cross-patient data increased, we performed a linear fit to
S5's log-transformed decoding accuracy vs. cross-patient trials curve using scipy (Fig. S11).

To investigate inter-patient variability in aligned cross-patient decoding, we quantified each target
patient's percent change in decoding accuracy from patient-specific performance to aligned cross-
patient decoding with all source patients added, which denote as cross-patient improvement. We
plotted this cross-patient improvement against the decoding accuracy of the target patient and
found a significant relationship using a linear fit between the two variables (Fig. 4e). As mentioned
in the main text, we chose to exclude S3 from this analysis and following inter-patient variability
due to its small dataset size, as it would be unclear cross-patient data would disproportionately
increase accuracy in this dataset with its limited number of trials. We additionally investigated
inter-patient variability using a leave-one-out cross-patient analysis. Here, for a given target
patient, we trained an aligned cross-patient model with all available source patients except one,
which was parametrically varied as the left-out patient. Then we quantified the benefit of that left-
out patient's data to the target patient's data as the percent change in accuracy between aligned
cross-patient training with all source patients and aligned cross-patient training without the left-
out patient, which we call the left-out accuracy change (Fig. S19a). Similar to Figure 4e, we

regressed the left-out accuracy change against effects of 1) target patient-specific accuracy (Fig.
S19b)., 2) left-out patient-specific accuracy (Fig. S19c)., and 3) distance between electrode arrays
in neuroanatomical space (Fig. S19d). All relationships against left-out accuracy change were
assessed with linear fits between the left-out accuracy change and the desired variable. P-values
of each linear fit were FDR corrected over all fits performed in the leave-one-out analysis,

Simulated real-time data generation

We simulated the real-time collection of neural features from our offline speech data to evaluate
the effectiveness of our proposed alignment method in contexts more applicable to speech BCI
deployment. To accomplish this, we adapted the methods of Willett and colleagues[7] to our high-
gamma processing and phoneme decoding pipeline. We began with 2ksps neural data split into
trials by alignment to the presentation of auditory stimuli during the speech task (as real-time
methods would not have knowledge of patient response times to use response-aligned data). We
split each trial into 20 ms bins (40 timepoints per bin at 2 ksps), applied a common average
reference within bins, and performed a modified HG filtering method within these short time bins.
HG filtering methods for offline, trial-based data typically employ a Hilbert transform to extract the
envelope of the logarithmically-spaced and sized frequency bands in the HG range (see *High-*
gamma extraction), which can be difficult to accurately implement in low-latency contexts with
short processing bins. Because of this, we chose to calculate the root-mean-square (RMS) HG
power in each of these 20 ms bins instead. We again bandpass the signal into 8 logarithmically-
spaced and sized frequency bands as with our offline data, then calculate RMS power in each
band as simply the square root of the mean of the squared signal. Bandpass filtering was
performed causally using a 4th order Butterworth filter designed to match frequency band
characteristics of the offline filtering frequency bands[50]. We finally average across these bands to
calculate a single HG power value for each channel. This is repeated for every 20 ms bin within
each trial, effectively downsampling the output HG power to a sampling rate of 50 sps. Here, trials
were initially defined 0 s to 3.5 seconds relative to stimulus onset. However, we trimmed the initial
500 ms to remove filter artifacts, resulting in trial definition as 0.5 s to 3.5 s relative to stimulus
onset.

To mimic clinical speech BCI deployment contexts with designated training and evaluation
periods, we split our three-block speech task into training blocks (first two blocks) and a testing
block (last block). Due to the single block of data collection for patient S3, we chose to exclude
them as a target patient in our simulated real-time decoding analyses (though we still used their
data to increase training set size in other target patients). Following simulated real-time processes
of the data from the training period, we performed channel and trial outlier removal identically to
our offline methods (see *Neural data preprocessing*). Identified bad channels from the training
period were saved and excluded from the data in the test period by imputing their values with the
mean across all good channels. Real-time processed HG data aligned to the stimulus were
normalized (mean-subtraction) by a pre-stimulus baseline (-500 ms to 0 ms relative to stimulus
onset as with offline data). Due to the short time period of experimental data collection, we chose
to save this baseline HG data from the training period of normalization of the test data as well
instead of using rolling normalization methods present in studies with longer collection and
evaluation periods.

Connectionist temporal classification recurrent neural network (CTC-RNN) decoding

Following simulated real-time processing of our training period and testing period data, we trained
stacked recurrent neural network (RNN) models with connectionist temporal classification loss
(CTC) to continuously predict phoneme identity throughout the trials. We used CTC loss and
decoded stimulus-presented phonemes (instead of patient response phonemes), as in previous
speech BCI work[7,8], to avoid use of information on patient-response timing which would not be
available in real-time speech BCI settings. Specifically, we set the label for each speech trial as
the sequence of phonemes presented in the auditory stimulus. Then from 0.5 s to 3.5 s post-
stimulus onset, we decoded phoneme identity using a causal sliding window of HG power features
(280 ms window size, 80 ms stride, i.e. 14 sample window size, 4 sample stride) as performed by
Willett and colleagues[7]. Training with CTC loss enables the comparison of different length
sequences in "decoding window" space (predicted phoneme every 80 ms) to "stimulus sequence"
space (three-length sequence of phonemes) by collapsing repeatedly predicted phonemes and
ignoring periods of "blank" predictions with no associated phoneme. In patient-specific decoding
cases, we trained models to directly translate real-time processed HG data across all channels to
phoneme sequences via our stacked RNN model with CTC loss. In unaligned cross-patient
decoding cases, high-channel count data was first reduced to patient-specific latent spaces via
PCA, pooled together into a single training set across patients, the trained to predict phoneme
sequences with the stacked RNN model. To enable data pooling across different latent sizes,
latent spaces were truncated to the minimum latent size across all patients. Latent size was
determined by the number of components explaining 90% of the variance in the input data.
However, for patient S5, noise in a single channel that was not removed by our initial channel
removal resulted in a single component explaining greater than 90% of the data's variance and
poor subsequent alignment. To remedy this, we fixed component selection to 30 components to
extract a broader range of components that would be better suited for subsequent alignment in this
patient. As latent space reduction was performed on data from the training period, this manual
removal is still valid in the context of clinical speech BCI deployment, as a similar analysis and
correction of data could be performed between training and testing periods. In aligned cross-
patient decoding cases, decoding performed as in the unaligned case, but with alignment of all
source patient latent dynamics to the target patient space prior to model training. In both unaligned
and aligned cases, learned dimensionality reduction and alignment linear transformations were
saved for application to validation or test set data.

CTC-RNN models were trained to optimize phoneme error rate (PER) between true phoneme
sequences and predicted phoneme sequences. PER was defined as the number of insertions,
substitutions, or deletions necessary to convert the predicted sequence to the true sequence,
divided by the true sequence length and converted to a percentage. In all decoding contexts
(patient-specific, unaligned, aligned), we trained the model over 300 epochs. We applied the
following data augmentations to our training data:

- **Temporal warping**: Each trial was temporally warped by randomly stretching or
compressing the time axis using linear interpolation. The signal was first resampled to a
randomly scaled duration (factor range: 0.8–1.2) and then resampled back to the original
length, introducing local temporal distortions while preserving overall sequence duration.

-    **Temporal masking**: A contiguous segment of each trial (10% of the total duration) was
randomly selected and set to zero, simulating transient signal dropout and encouraging
robustness to missing temporal information.
-    **Time shifting**: Each trial was circularly shifted along the time axis by a random offset (±20
samples), preserving signal structure while introducing temporal misalignment to improve
invariance to onset timing.
-    **Additive noise**: Additive Gaussian noise ($\sigma = 0.01$) was applied to the signal to simulate
measurement noise and improve robustness to low-amplitude perturbations.
-    **Amplitude scaling**: Each trial was multiplied by a random scalar factor (range: 0.9–1.1),
introducing variability in overall signal amplitude to promote invariance to differences in
amplitude.

We split a validation set from our training data to monitor PER during training and loaded the
model weights from the model with lower PER during training for use in evaluation on the test set
(Fig. 6b). We performed hyperparameter optimization for each patient and decoding context
separately to optimize CTC-RNN performance. Hyperparameter optimization was implemented
with random search and 10-fold cross-validation over training trials (see Tables S6-8 for patient
hyperparameters in patient-specific, unaligned, and aligned cases, respectively). As the
sequence length of predicted phoneme sequence is variable, a theoretical chance value of PER
based on phoneme composition is less defined for this analysis. As such, we calculated an
empirical chance distribution for each patient by performing patient-specific CTC-RNN training
with permuted labels and evaluation on unmodified test data. As with our offline decoding
analysis, we additionally performed CTC-RNN decoding evaluations while subsampling the
amount of target patient data (Fig. 5c) and cross-patient data (Fig. 5d). We projected PER with
larger cross-patient data amounts by performing a linear fit between aligned decoding PER and
log-transformed amounts of cross-patient data using scipy's *linregress* function. All CTC-decoding
analyses were performed in Python 3.10 via Pytorch-lightning v2.3.3 and Pytorch v2.4.0.

Pitch subsampling

Lower electrode densities were simulated by using Poisson disk sampling[56] to sub-select
electrodes from the μECoG array constrained by a minimum distance between electrodes. Given
a desired pitch as this constraining distance, we calculated the number of electrodes that needed
to be sampled to achieve this pitch as:

$$n = XY/p^2,$$

where *n* is the number of electrodes, *X* and *Y* are the length and width of the μECoG array, and
p is the desired pitch. We rounded *n* to the nearest integer in implementation. We only
considered subsampled electrodes with significant HG activity during the patient response for
subsequent cross-patient analyses. In these cross-patient analyses, all patients were
subsampled to the desired pitch, not just the target patient.

Cross-patient decoding on pitch-subsampled data (Fig. 5a, top; Fig. S13 for all patients
separately) was performed as described previously (*Cross-patient phoneme decoding*) with

PCA-SVM models, but with pitch subsampling of the HG data prior to latent dynamics
extraction, alignment, and decoding. At each of the 50 training iterations, a new subsampling of
electrodes at the current pitch was selected to mitigate effects of electrode identity.
Representational similarity between cross-patient latent dynamics from subsampled arrays (Fig.
5a, bottom) was calculated similarly to previous analyses (*Alignment of latent dynamics*) except
at each pitch, the representational similarity was averaged across 50 iterations to mitigate
effects of electrode identity during sampling.

Grid subsampling

Smaller electrode array coverages were simulated by sampling rectangular sub-grids of size *n* x
2n (*n ϵ [2,8]*) from the total electrode array. While maximum electrode array sizes exceeded 8x16
for patients with arrays of 256 electrodes, we only considered sub-grids up to 8x16 for consistency
across all patients. For a given sub-grid size, we consider all possible starting locations that fit
within the total electrode array. That is, for an 8x16 electrode array with a sub-grid size of 6x12,
there would be a total of 15 possible sub-grids to select from. Only electrodes with significant HG
activity during the patient response were selected from each sub-grid.

Cross-patient decoding from sub-grids (Fig. 5b, top; Fig. S14 for all patients separately) was again
performed as previously described with PCA-SVM models and followed a similar approach to the
cross-patient correlation analysis for sub-grid selection. For a given sub-grid size, training was
repeated for all possible target patient sub-grids and a random sub-grid from each source patient
was selected at each iteration. At each sub-grid size, phoneme decoding accuracy was averaged
across all target patient sub-grids. Representational similarity between cross-patient latent
dynamics from sub-grids (Fig. 5b, bottom) were extracted similarly to previous analyses. For each
target patient and sub-grid size, we extracted latent dynamics from HG in the sub-selected
electrodes and aligned them across patients. We iterated over all possible sub-grids of the given
size for the target patient. For each possible sub-grid, we selected a random sub-grid (uniformly)
from the possibilities for each source patient. Following, representational similarity across aligned
latent dynamics was calculated as previously described. For each sub-grid size, the reported
correlation was averaged across all target patient sub-grids and across all source patients,
yielding a single averaged representational similarity value for each target patient.

Spatial-averaging

Larger contact sizes were simulated by averaging data together from neighboring contacts with
varying sizes. Contact sizes ranged from 1x1 (unaveraged micro-contact used in previous
analyses in this work) to 8x8 squares. To simulate neural recording with these larger contacts,
raw signals were averaged together on these contacts and HG was extracted from the resultant
spatially-averaged neural signal. Only spatially-averaged contacts with significant HG activity
during the patient response (relative to a baseline period 500 ms prior to stimulus presentation)
were considered for subsequent analyses (see *Statistical Analysis*). Note that this spatial-
averaging method does not account for changes in electrode impedance for different sized
contacts. For contact sizes that did not span the total electrode array (e.g. Two 8x8 contacts span

an 8x16 electrode array, but 7x7 contacts do not), electrodes were centered within the electrode
array.

Cross-patient decoding on spatially-averaged data (Fig. 5c, top; Fig. S15 for all patients
separately) was performed as described previously (*Cross-patient phoneme decoding*) with PCA-
SVM models, but with the spatially-averaged data used for latent dynamics extraction, alignment,
and decoding. Cross-patient representational similarity in latent dynamics for spatially averaged
data (Fig. 5c, bottom) was again calculated similar to previous analyses, but with spatial-
averaging of data prior to latent dynamics extraction and alignment. As the spatial-averaging
results in a singular, deterministic sub-selection (unlike pitch and grid subsampling), the
representational similarity in latent dynamics was not repeated over multiple iterations. Instead, a
single representational similarity value (Pearson *r* between RDMs) was calculated of each target
patient and source patient and averaged across all source patients for each target patient (as in
Alignment of latent dynamics). For each contact size, representational similarity was reported
across a distribution of all target patients.

Statistical analysis

All statistical analyses were performed in MATLAB R2021a and Python 3.10 (scipy and
statsmodels). All permutation tests mentioned below use the difference in means as the test
statistic. All false discovery rate (FDR) control mentioned below implements the Benjamini-
Hochberg method[57].

Fig. 1: To identify electrodes with significant HG activation during patient responses, we
compared the average (over time) HG power around the patient response onset (-250 ms to 250
ms) to a pre-stimulus baseline period (-500 ms to 0 ms relative to stimulus presentation) for each
electrode using a FDR-corrected 10,000-iteration one-sided permutation test.

Fig. 2: For latent dynamics clustering strength (quantified by the silhouette score), comparisons
between distributions (articulatory features vs chance) were performed with Mann-Whitney U
(MWU) test. MWU tests were also used for cluster strength comparisons when combining data
from both S1 and S2, where FDR correction was applied. Differences in paired distributions for
cross-patient representational similarity of latent dynamics were compared with a Wilcoxon
signed-rank test.

Fig. 3: Reconstructed articulator map similarity was compared between aligned and unaligned
contexts with a Wilcoxon signed-rank test. Cross-patient articulator decoding accuracy
distributions were compared with exact two-sided, paired permutation tests and FDR correction.
Comparison of cross-patient articulator decoding accuracies to chance accuracy was performed
using Wilcoxon signed-rank tests corrected with FDR correction.

Fig. 4: Differences in aggregated results for all target patients were tested with a repeated
measures ANOVA and follow-up paired t-tests with false discovery control. Within target patients,
differences in distributions of phoneme decoding accuracy were compared across decoding
contexts (patient-specific, unaligned cross-patient, aligned cross-patient) using a one-way

analysis of variance (ANOVA) and post-hoc Tukey tests. Effect sizes were reported as Cohen's
d. For target patient data subsampling, an exact two-sided permutation test was performed at
each target data subsampling ratio. FDR correction was applied across all subsampling ratio
tests. A linear fit was performed between cross-patient gain and target patient-decoding accuracy
to determine a significant relationship between the two variables.

Fig. 5: Differences between chance, patient-specific, unaligned, and aligned CTC-RNN decoding
was performed with multiple FDR-corrected Wilcoxon signed-rank tests. At each 5% interval in
the target patient subsampling analysis, a Wilcoxon signed-rank test was performed to assess
differences in error rates between aligned and patient-specific results across all patients. FDR-
correction was performed across all subsampling points. Similarly, for the cross-patient
subsampling analysis, Wilcoxon-signed rank tests were performed at each added cross-patient
data value to compare aligned cross-patient performance to the single distribution of patient-
specific decoding performance. FDR-correction was again performed across all subsampling
points.

Fig. 6: Statistical testing was performed to compare differences between aligned cross-patient
and patient-specific decoding accuracies for different subsampling methods. For each
subsampling method (pitch, grid, and spatial-averaging) we performed an exact two-sided, paired
permutation test at each subsampled point with FDR correction across all points.

Data Availability

The data from µECoG recordings that support the decoding analysis are available via the Data
Archive for The Brain Initiative (DABI; accession code: <To be updated after acceptance>) under
restricted access (restrictions will be exempted after appropriate modifications of the IRB protocol:
Pro0072892). The access can be obtained upon written requests from the corresponding authors
(gregory.cogan@duke.edu, j.viventi@duke.edu). Source data are provided with this paper.

Code Availability

Code for cross-patient alignment analyses and notebooks recreating the figures in this work can
be found at https://github.com/coganlab/cross_patient_speech_decoding.

1284

1285

1286

1287

1288

1289

References

1. Levelt, W. J. M. *Speaking: From Intention to Articulation*. (The MIT Press, 1989). doi:10.7551/mitpress/6393.001.0001.

2. Mitchell, J. & Borasio, G. Amyotrophic lateral sclerosis. *The Lancet* **369**, 2031–2041 (2007).

3. Gowda, S. N., Munakomi, S. & De Jesus, O. Brainstem Stroke. in *StatPearls* (StatPearls Publishing, Treasure Island (FL), 2024).

4. Lulé, D. *et al.* Life can be worth living in locked-in syndrome. in *Progress in Brain Research* vol. 177 339–351 (Elsevier, 2009).

5. Haig, A. J., Katz, R. T. & Sahgal, V. Mortality and complications of the locked-in syndrome. *Arch. Phys. Med. Rehabil.* **68**, 24–27 (1987).

6. Felgoise, S. H., Zaccheo, V., Duff, J. & Simmons, Z. Verbal communication impacts quality of life in patients with amyotrophic lateral sclerosis. *Amyotroph. Lateral Scler. Front. Degener.* **17**, 179–183 (2016).

7. Willett, F. R. *et al.* A high-performance speech neuroprosthesis. *Nature* **620**, 1031–1036 (2023).

8. Metzger, S. L. *et al.* A high-performance neuroprosthesis for speech decoding and avatar control. *Nature* **620**, 1037–1046 (2023).

9. Moses, D. A. *et al.* Neuroprosthesis for Decoding Speech in a Paralyzed Person with Anarthria. *N. Engl. J. Med.* **385**, 217–227 (2021).

10. Card, N. S. *et al.* An Accurate and Rapidly Calibrating Speech Neuroprosthesis. *N. Engl. J. Med.* **391**, 609–618 (2024).

11. Panachakel, J. T. & Ramakrishnan, A. G. Decoding Covert Speech From EEG-A Comprehensive Review. *Front. Neurosci.* **15**, 642251 (2021).

12. Lopez-Bernal, D., Balderas, D., Ponce, P. & Molina, A. A State-of-the-Art Review of EEG-Based Imagined Speech Decoding. *Front. Hum. Neurosci.* **16**, 867281 (2022).

13. Dabagia, M., Kording, K. P. & Dyer, E. L. Aligning latent representations of neural activity.

Nat. Biomed. Eng. **7**, 337–343 (2022).

14. Makin, J. G., Moses, D. A. & Chang, E. F. Machine translation of cortical activity to text with

an encoder–decoder framework. *Nat. Neurosci.* **23**, 575–582 (2020).

15. Chartier, J., Anumanchipalli, G. K., Johnson, K. & Chang, E. F. Encoding of Articulatory

Kinematic Trajectories in Human Speech Sensorimotor Cortex. *Neuron* **98**, 1042-1054.e4

(2018).

16. Metzger, S. L. *et al.* Generalizable spelling using a speech neuroprosthesis in an individual

with severe limb and vocal paralysis. *Nat. Commun.* **13**, 6510 (2022).

17. Kellis, S. *et al.* Decoding spoken words using local field potentials recorded from the cortical

surface. *J. Neural Eng.* **7**, 056007 (2010).

18. Herff, C. *et al.* Brain-to-text: decoding spoken phrases from phone representations in the

brain. *Front. Neurosci.* **9**, (2015).

19. Bouchard, K. E., Mesgarani, N., Johnson, K. & Chang, E. F. Functional organization of

human sensorimotor cortex for speech articulation. *Nature* **495**, 327–332 (2013).

20. Pei, X. *et al.* Spatiotemporal dynamics of electrocorticographic high gamma activity during

overt and covert word repetition. *NeuroImage* **54**, 2960–2972 (2011).

21. Stavisky, S. D. *et al.* Neural ensemble dynamics in dorsal motor cortex during speech in

people with paralysis. *eLife* **8**, e46015 (2019).

22. Wilson, G. H. *et al.* Decoding spoken English from intracortical electrode arrays in dorsal

precentral gyrus. *J. Neural Eng.* **17**, 066007 (2020).

23. Churchland, M. M. *et al.* Neural population dynamics during reaching. *Nature* **487**, 51–56

(2012).

24. Gallego, J. A., Perich, M. G., Miller, L. E. & Solla, S. A. Neural Manifolds for the Control of

Movement. *Neuron* **94**, 978–984 (2017).

25. Gallego, J. A., Perich, M. G., Chowdhury, R. H., Solla, S. A. & Miller, L. E. Long-term

stability of cortical population dynamics underlying consistent behavior. *Nat. Neurosci.* **23**,

260–270 (2020).

26. Safaie, M. *et al.* Preserved neural dynamics across animals performing similar behaviour.

Nature **623**, 765–771 (2023).

27. Chiang, C.-H. *et al.* Flexible, high-resolution thin-film electrodes for human and animal

neural research. *J. Neural Eng.* **18**, 045009 (2021).

28. Barth, K. J. *et al.* Flexible, high-resolution cortical arrays with large coverage capture

microscale high-frequency oscillations in patients with epilepsy. *Epilepsia* **64**, 1910–1924

(2023).

29. Duraivel, S. *et al.* High-resolution neural recordings improve the accuracy of speech

decoding. *Nat. Commun.* **14**, 6938 (2023).

30. Gallego-Carracedo, C., Perich, M. G., Chowdhury, R. H., Miller, L. E. & Gallego, J. Á. Local

field potentials reflect cortical population dynamics in a region-specific and frequency-

dependent manner. *eLife* **11**, e73155 (2022).

31. Cheung, C., Hamilton, L. S., Johnson, K. & Chang, E. F. The auditory representation of

speech sounds in human motor cortex. *eLife* **5**, e12577 (2016).

32. Leonard, M. K. *et al.* Large-scale single-neuron speech sound encoding across the depth of

human cortex. *Nature* **626**, 593–602 (2024).

33. Ray, S. & Maunsell, J. H. R. Different origins of gamma rhythm and high-gamma activity in

macaque visual cortex. *PLoS Biol.* **9**, (2011).

34. Ray, S., Crone, N. E., Niebur, E., Franaszczuk, P. J. & Hsiao, S. S. Neural Correlates of

High-Gamma Oscillations (60–200 Hz) in Macaque Local Field Potentials and Their

Potential Implications in Electrocorticography. *J. Neurosci.* **28**, 11526–11536 (2008).

35. Graves, A., Fernández, S., Gomez, F. & Schmidhuber, J. Connectionist temporal

classification: labelling unsegmented sequence data with recurrent neural networks. in

Proceedings of the 23rd international conference on Machine learning - ICML '06 369–376

(ACM Press, Pittsburgh, Pennsylvania, 2006). doi:10.1145/1143844.1143891.

36. Sussillo, D., Churchland, M. M., Kaufman, M. T. & Shenoy, K. V. A neural network that finds

a naturalistic solution for the production of muscle activity. *Nat. Neurosci.* **18**, 1025–1033

(2015).

37. Buzsáki, G., Anastassiou, C. A. & Koch, C. The origin of extracellular fields and currents —

EEG, ECoG, LFP and spikes. *Nat. Rev. Neurosci.* **13**, 407–420 (2012).

38. Dubey, A. & Ray, S. Cortical Electrocorticogram (ECoG) Is a Local Signal. *J. Neurosci.* **39**,

4299–4311 (2019).

39. Pandarinath, C. *et al.* Inferring single-trial neural population dynamics using sequential auto-

encoders. *Nat. Methods* **15**, 805–815 (2018).

40. Gosztolai, A., Peach, R. L., Arnaudon, A., Barahona, M. & Vandergheynst, P. MARBLE:

interpretable representations of neural population dynamics using geometric deep learning.

Nat. Methods **22**, 612–620 (2025).

41. Shellikeri, S., Keith, J., Black, S. E., Zinman, L. & Yunusova, Y. Neuropathology of Speech

Network Distinguishes Bulbar From Nonbulbar Amyotrophic Lateral Sclerosis. *J.*

Neuropathol. Exp. Neurol. **79**, 284–295 (2020).

42. Zaninotto, A. L. *et al.* Speech motor impairment in ALS is associated with multiregional

cortical thinning beyond primary motor cortex. *Front. Neurol.* **15**, 1451177 (2024).

43. Pang, X. *et al.* Reorganization of Dynamic Network in Stroke Patients and Its Potential for

Predicting Motor Recovery. *Neural Plast.* **2024**, 9932927 (2024).

44. Tilton-Bolowsky, V., Stockbridge, M. D. & Hillis, A. E. Remapping and Reconnecting the

Language Network after Stroke. *Brain Sci.* **14**, 419 (2024).

45. Dale, A. M., Fischl, B. & Sereno, M. I. Cortical Surface-Based Analysis. *NeuroImage* **9**, 179–

194 (1999).

46. Papademetris, X. *et al.* BioImage Suite: An integrated medical image analysis suite: An

update. *Insight J.* https://doi.org/10.54294/2g80r4 (2006) doi:10.54294/2g80r4.

47. McAuliffe, M., Socolof, M., Mihuc, S., Wagner, M. & Sonderegger, M. Montreal Forced

Aligner: Trainable Text-Speech Alignment Using Kaldi. in *Interspeech 2017* 498–502 (ISCA,

2017). doi:10.21437/Interspeech.2017-1386.

48. Audacity Team. Audacity. Muse Group (2024).

49. Cogan, G. B. *et al.* Manipulating stored phonological input during verbal working memory.

Nat. Neurosci. **20**, 279–286 (2017).

50. Mesgarani, N., Cheung, C., Johnson, K. & Chang, E. F. Phonetic Feature Encoding in

Human Superior Temporal Gyrus. *Science* **343**, 1006–1010 (2014).

51. Elsayed, G. F. & Cunningham, J. P. Structure in neural population recordings: an expected

byproduct of simpler phenomena? *Nat. Neurosci.* **20**, 1310–1318 (2017).

52. Fabian Pedregosa *et al.* Scikit-learn: Machine Learning in Python. *J. Mach. Learn. Res.* **12**,

2825–2830 (2011).

53. Maaten, L. van der & Hinton, G. Visualizing Data using t-SNE. *J. Mach. Learn. Res.* **9**,

2579–2605 (2008).

54. Rousseeuw, P. J. Silhouettes: A graphical aid to the interpretation and validation of cluster

analysis. *J. Comput. Appl. Math.* **20**, 53–65 (1987).

55. Kriegeskorte, N. Representational similarity analysis – connecting the branches of systems

neuroscience. *Front. Syst. Neurosci.* https://doi.org/10.3389/neuro.06.004.2008 (2008)

doi:10.3389/neuro.06.004.2008.

56. Bridson, R. Fast Poisson disk sampling in arbitrary dimensions. in *ACM SIGGRAPH 2007*

sketches 22 (ACM, San Diego California, 2007). doi:10.1145/1278780.1278807.

57. Benjamini, Y. & Hochberg, Y. Controlling the False Discovery Rate: A Practical and

Powerful Approach to Multiple Testing. *J. R. Stat. Soc. Ser. B Stat. Methodol.* **57**, 289–300

(1995).

Acknowledgements

J.V., G.B.C., S.P.L., A.H.F., D.G.S, S.D., Z.S. were supported by NIH R01DC019498 and DoD W81XWH-21-0538. S.R. was supported by an NRCDP NIH K12 award. J.V., G.B.C., C.W., S.D. were supported by NIH CTSA UL1TR002553. K.B. was supported by the National Science Foundation (Graduate Research Fellowship Program DGE-1644868). C.S. was supported by the National Science Foundation (Graduate Research Fellowship Program DGE-2139754) J.V. and G.B.C. were supported by NIH UG3NS120172, G.B.C, J.V., S.P.L, A.H.F, D.G.S. were supported by NIH R01NS129703, and G.B.C., J.V., S.P.L., S.D. were supported by an internal Duke Institute for Brain Sciences Incubator Award. We thank Nicole Liddle, Palee Abeyta, and Anna Thirakul for help with consenting participants, Seth Foster for help with the experimental design, Steve Harward for help with surgical preparation, and Daniel Sexton for help with surgical preparation and neuroanatomical localizations of electrode arrays. With tremendous gratitude, we thank the patients who volunteered their time for our study.

Author Contributions

Z.S. performed all analyses, performed statistical testing, and generated figures. S.D. performed significant preprocessing of the neural data and assisted in analysis design. G.B.C. and S.R. designed the experimental task. S.R., S.P.L., A.H.F, D.G.S performed intraoperative electrode implantation and neural data acquisition. C.W., K.B., C.S., and J.V. were involved in µECoG electrode design and setting up the intraoperative data acquisition process. Z.S., S.D, K.B., C.S., and J.V., and G.B.C performed intraoperative neural data collection. Z.S., J.V., and G.B.C wrote the manuscript with feedback from other co-authors. G.B.C. was involved in clinical supervision. J.V., and G.B.C. designed and supervised the human intraoperative study.

Competing Interests

The authors declare no competing interests.