Distinct networks of expressed genes are associated with neophobia in the hippocampus of male

and female Eurasian tree sparrows (*Passer montanus*)

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**Abstract**

Neophobia, avoidance of novel stimuli, is an ecologically and evolutionarily relevant behavioral

trait that varies among individuals and across species. Especially among wild animals, the

neuromolecular mechanisms underlying individual variation in neophobia have not been well

characterized. We examined three neophobic behaviors in captive female and male Eurasian tree

sparrows (*Passer montanus*) from a wild population introduced to the USA in 1870: responses

towards novel objects, novel foods, and repeated presentations of the same initially novel object.

We compared transcriptomic patterns associated with neophobia in three brain regions, the

striatum, dorsal hippocampus, and rostral hippocampus, using differential expression and co-

expression network analyses. We found that the striatum and hippocampus had distinct

transcriptomic profiles, as did the rostral and caudal subregions of the hippocampus, supporting

recent hypotheses that these subregions are functionally specialized. Despite the absence of sex

differences in neophobic behaviors, neophobia-associated gene modules revealed sex-specific

patterns within brain regions. For females, neophobic behaviors more strongly correlated with

gene modules in the caudal hippocampus, a region involved in stress and anxiety, whereas for

males, neophobic behaviors correlated with gene modules in the rostral hippocampus, a region

that may play a larger role in spatial cognition. These modules exhibited significant overlap,

suggesting that neophobic behaviors in both females and males are driven by shared

neurobiological mechanisms, though they exhibit sex-specific patterns of brain region

localization. Further, this work highlights the importance of examining both male and female

animals in neurobiological research.

**Keywords:** neurogenomics, sex difference, transcriptomics, RNAseq, songbird, behavior

**Introduction**

Fearful behavior towards novel stimuli, or neophobia, is found in many animal species,

and can be observed in response to new foods, objects, and environments (Crane & Ferrari,

2017; Kimball & Lattin, 2023b). While an aversive reaction to a dangerous new stimulus can

help ensure an animal's survival, a fearful response to novelty may prevent access to new foods,

nesting sites, or other beneficial resources (Greenberg & Mettke-Hofmann, 2001). Neophobia

has become increasingly relevant for many wild animals as their environments undergo

anthropogenic changes. Research suggests that urban animals are less neophobic than their rural

counterparts (Breck et al., 2019; Huang et al., 2020; Jarjour et al., 2019; Tryjanowski et al.,

2016) and even within urban sites, individuals that experienced more anthropogenic disturbances

were less neophobic than those encountering fewer disruptions (Grunst et al., 2019). Neophobia

has also been tied to invasion success, where reduced neophobia may help invasive species

exploit novel resources (Candler & Bernal, 2015; Cohen et al., 2020; Martin & Fitzgerald, 2005).

However, despite its ecological relevance and the wide variation in neophobia observed across

individuals, populations, and species, the molecular mechanisms underlying this behavior are not

well understood. Therefore, there is a clear need for more studies investigating neurobiological

differences between neophobic and non-neophobic individuals.

Much of what is currently known about the neurobiological basis of neophobia comes

from laboratory rodent models. Although lab-bred rats and mice typically show a preference for

novel over familiar stimuli, they do exhibit neophobia towards some - but not all - novel tastes

and when feeding in a novel environment (hyponeophagia) (Olszewski et al., 2014; Roman et al.,

2009; Shephard et al., 1982; Stehberg et al., 2011). Rodent research has identified several

neurobiological mediators in the glutamate, serotonin, and norepinephrine systems involved in

these types of feeding neophobia (Ramírez-Lugo et al., 2015; Royet et al., 1983; Simpson et al.,

2011; Steketee et al., 1992). However, because these neophobic responses are context- and

stimulus-specific, it is unclear how generalizable these results are to the more widespread

neophobia typically observed in wild animals (Crane & Ferrari, 2017).  Further, because lab

animals tend to be highly inbred, they do not show the wide within-species variation in

neophobia observed in many wild populations. Research in wild-caught or outbred animals is

necessary to reveal the neuroendocrine basis for individual variation in neophobia.

Transcriptomic approaches enable the unbiased identification of gene expression patterns

associated with behavior, extending the scope beyond the handful of neurotransmitters and

hormones typically investigated with more targeted approaches (Bell & Robinson, 2011;

Bengston et al., 2018; Calisi & MacManes, 2015). Transcriptomics has been particularly useful

in revealing what brain regions are involved in behaviors with wide inter-individual variation,

like neophobia. For example, a recent transcriptome study in the invasive house sparrow (*Passer*

*domesticus*) (Lattin et al., 2022) found that neophobic and non-neophobic individuals had

distinct patterns of constitutive differential gene expression in brain regions that included the

hippocampus (a region linked to spatial cognition, novelty responses, and memory) and the

striatum (which is involved in reward and learning) (Güntürkün, 2005; Medina & Reiner, 1995;

Rose & Colombo, 2005; Shinohara & Yasoshima, 2021). Differences between neophobic and

non-neophobic sparrows were especially pronounced in the hippocampus, suggesting this region

may be particularly important in neophobia (Lattin et al., 2022), a conclusion further supported

by a study showing high neuronal activity in the hippocampus after exposure to novel objects

(Kimball et al., 2022). However, this transcriptome work was performed in only one sex, leaving

uncertainty about sex-specific mechanisms.

Although many species - including house sparrows - show no sex differences in

neophobia (Schaffer et al., 2021; Kelly et al., 2020; Krajcir et al., 2024), it is not necessarily the

case that the same neurobiological mechanisms regulate the same behaviors in males and

females. The sexes may converge upon similar behaviors despite divergent neurobiological

processes, due to different life history pressures or endocrine phenotypes (McCarthy et al.,

2012). Neurobiological differences between females and males may emerge in several different

ways. For instance, many sex differences have been found in hippocampal anatomy in mammals

(Brivio et al., 2020; Yagi & Galea, 2019). In laboratory mice, females have greater dendritic

spine density and heightened gene expression responses to receptor knockouts in the

hippocampus than males, likely driven by differences in steroid hormone levels (Oakley et al.,

2024; Shors et al., 2001). In birds, female brown-headed cowbirds (*Molothrus ater*) have a larger

hippocampus and greater neurogenesis than males, potentially associated with females' brood

parasite life history strategy (Guigueno et al., 2016; Sherry et al., 1993). However, food-caching

black-capped chickadees (*Poecile atricapillus*) do not show sex differences in spatial memory or

hippocampal size (Petersen & Sherry, 1996), and recent genome-wide association studies

revealed little effect of sex on the genetic contribution to chickadee spatial cognition (Semenov

et al., 2024). Thus, the inclusion of both female and male subjects in biological research has the

potential to reveal sex-dependent biological processes (Klein et al., 2015; Shansky & Murphy,

2021).

In the present study, we examined potential links between neophobia and constitutive

gene expression in the brain of Eurasian tree sparrows (*Passer montanus*), an invasive songbird

introduced to the USA in 1870 (Widmann, 1889). Birds are among the taxa with the highest

levels of neophobia (Crane & Ferrari, 2017), and like their congener the house sparrow, Eurasian

tree sparrows have wide and repeatable inter-individual variation in neophobia (Krajcir et al.,

2024). Because different types of neophobia tests can sometimes show varying results (Dusang

et al., 2025; Kimball & Lattin, 2023a), neophobia was assessed using three different behavioral

assays: sparrows' responses towards novel objects, novel foods, and repeated exposures of the

same initially novel object. Three weeks after neophobia behavior tests were complete, we

compared gene expression patterns in three different brain regions (striatum, rostral

hippocampus, and caudal hippocampus) in males and females in relation to their behavioral

performance using differential expression and co-expression network analyses. We divided the

hippocampus into subregions because recent evidence suggests that the avian hippocampus

shows specialization along a rostral-caudal axis, similar to that of the dorsal and ventral

hippocampus in rodents (Fanselow & Dong, 2010; Gualtieri et al., 2019; Smulders, 2017). We

predicted that neophobic and non-neophobic individuals would show differences in constitutive

gene expression across the three brain regions, with distinct patterns reflecting their functional

specializations. These patterns could emerge via differences in expression patterns, module

composition or structure, or biological processes, suggesting that there may be multiple

biological routes to the same behavioral outcome. Based on past work showing novel object

exposure increased neuronal activity in the caudal but not rostral portions of the house sparrow

hippocampus (Kimball et al., 2022), we expected the strongest effects of neophobia on

expression in the caudal hippocampus. In the same set of Eurasian tree sparrows, we previously

found no sex differences in responses to any neophobia assays (Krajcir et al., 2024). However,

although neophobia behaviors did not differ by sex, we predicted that the underlying

transcriptomic patterns might show sex specificity.

**Materials and methods**

*Bird capture and behavioral trials*

All animal work was approved by the Louisiana State University Institutional Animal

Care and Use Committee (protocol #10-2021), and Eurasian tree sparrows were collected under

an Illinois State Scientific Collecting Permit. Sparrow capture, transport, housing, and euthanasia

followed bird research guidelines from the Ornithological Council (Fair et al., 2023). Adult

Eurasian tree sparrows (n = 13 males, 11 females) were captured from Tazewell County, Illinois,

United States in March 2023 using mist nets and nest box traps on private landowners' property.

Sparrows were transported to Louisiana State University overnight (to minimize stress) in a

modified pet carrier with multiple food and water dishes and perches. Once birds arrived at

Louisiana State University, they were placed in long-term housing cages in a vivarium facility on

campus. The cages were supplied with multiple perches of differing materials, a dust bathing tray

filled with sand, a plastic pine branch to use as a hide, water, and food (a mix of seeds, vitamin-

supplemented food pellets, and grit). Birds were acclimated to captivity for three weeks before

beginning behavior trials. Neophobia behavior trials are further detailed in Krajcir et al. (2024).

Birds were doubly housed until three days before behavioral trials. Because Eurasian tree

sparrows are not sexually dimorphic, we did not know sex at the time of paired housing. We

paired individuals randomly, so some were in same-sex and some were in mixed-sex pairs. None

of the pairs showed aggression during shared housing. During experimental trials, birds were

singly housed to avoid any effects of social interactions on neophobia (Kelly et al., 2020).

Sparrows were cared for daily by the research team and animal husbandry staff, and weighed

every two weeks to ensure body mass was maintained.

Neophobia in captive Eurasian tree sparrows was quantified using the same methods

previously used in house sparrows (Kimball et al., 2022). Briefly, food was removed 30 min

before lights out in the room, and trials conducted the following morning 30 min after lights on.

Neophobia was quantified as latency to first approach and feed from a food dish presented either

with a novel object (a purple plastic egg, white partial cover, red-painted dish, yellow pipe

cleaners, or blinking light) or a novel food item (chopped kiwi, shredded cheese, peanut butter,

or fruit-flavored cereal) during 1-h trials. Multiple types of novel objects and foods were used to

ensure we measured individuals' general response to novelty, rather than specific responses to

one particular stimulus, which may be a type of pseudoreplication (Kimball and Lattin 2023b).

Control approach and feeding times were determined by presenting birds with their normal food

dish filled with familiar foods. Control trials were used to ensure that behavioral responses were

specific to the novel stimulus, rather than a generalized aversion to the testing procedure; indeed,

all sparrows ate relatively quickly when only familiar foods were present during control trials (on

average $214 \pm 224$ s to approach and $254 \pm 232$ s to feed).

Food neophobia, object neophobia, and habituation to repeated exposures of an initially

novel object were tested. Object neophobia was tested during week one (n = 3 novel object trials

n = 2 control trials per bird, presented in random order), food neophobia was tested during week

two (n = 4 novel food trials and n = 2 control trials per bird, presented in random order), and

object habituation was tested during week three (n = 4 trials using the same initially novel object

and n = 1 control trial per bird, with the control trial first). There is some debate in the literature

about what measure best reflects an animal's neophobia: its responses to novel stimuli alone or

the difference in its responses to novel compared to control stimuli (e.g., Many Birds Project

2025). Because we found that average responses to novel objects and foods and the difference

from average control responses were correlated ($R^2 > 0.71$, p < 0.0001 for all measures), we used

sparrows' average responses to novel stimuli for ease of interpretation. All six behavioral

measures of neophobia (average latency to approach the food dish when novel objects were

present, average latency to feed when novel objects were present, average latency to approach

the food dish when it contained novel food, average latency to eat novel food, average latency to

approach the food dish during novel object habituation trials, and average latency to feed during

novel object habituation trials) were highly and significantly correlated among individuals when

measured as a group, as well as when males and females were examined separately, except for

novel food approach and novel food feeding in females (Table S1). To correlate different

neophobia-related behaviors with gene expression (see WGCNA below), we averaged among

trials for novel objects, food, and object habituation for each individual.

*RNAseq tissue preparation and sample collection*

Three weeks after neophobia trials ended, sparrows were euthanized using an overdose of

isoflurane anesthesia followed by rapid decapitation. Our goal was to assess differences in

constitutive gene expression in the brains of neophobic and non-neophobic birds, rather than

differences due to behavioral trials. This three week period was chosen because this is the typical

length of time we use to habituate wild animals to new conditions such as lab housing, and

because there is good evidence that this length of time is sufficient from some physiological

changes to adjust (Dickens and Romero 2009). After decapitation, brains were removed, flash-

frozen on powdered dry ice, and stored at -80°C until sectioned coronally on a cryostat (Cryostar

NX50, Thermo Fisher). Tissue was embedded in Optimal Cutting Temperature compound

(VWR 95057-838) in block molds and placed in the cryostat to be sliced at -22°C. Slices were

mounted directly onto slides (Superfrost Plus, VWR 48311-703) in two alternating series of 50

μm and 200 μm slices (Lattin et al., 2022). The 50 μm slides were dried overnight at 4°C, stained

with thionin, which stains Nissl substance and DNA, and coverslipped. The 200 μm slides were

chilled on dry ice and stored at -80°C until tissue punching.

Brain tissue punches were taken from the three target regions: striatum, caudal

hippocampus, and rostral hippocampus (Figure 1A). Target brain slices for punching in the 200

μm slides were determined using landmarks in the adjacent 50 μm thionin-stained slides, using

published songbird brain atlases as guides (Nixdorf-Bergweiler & Bischof, 2007; Stokes et al.,

1974). Striatum punches came from slices that also contained the quinto-frontal tract, which

corresponds to plates 3 and 4 from the Stokes canary atlas (1974). Two 1 mm diameter punches

(Fine Science Tools 18035–01, 15 G) were taken from each hemisphere of the striatum on one

target slice (i.e., four total punches), ~3 mm from the rostral tip of the brain. Rostral

hippocampus was punched on slices where the septomesencephalic tract (TrSM) was evident, ~5

mm into the brain, which corresponds to plate 9 from the Stokes canary atlas (1974). Ten 0.5 mm

diameter punches (Fine Science Tools 18035-50, 19G) were collected from both hemispheres of

the rostral hippocampus across three consecutive brain slices. Caudal hippocampus was punched

from slices where the cerebellum first came in, ~8 mm into the brain, which corresponds to

plates 20 and 21 from the Stokes canary atlas (1974). Tissue was collected from both

hemispheres of four consecutive slices using the 0.5 mm diameter tissue punch (10-16 punches

total). All punches from the same target brain region and subject were combined in sterile

D/RNAse Free 1.6 mL centrifuge tubes and stored at -80°C until RNA extraction. Tissue

punching tools were sterilized between subjects and brain regions using D/RNAse Free solution

(Cole-Parmer□EW-78901-43) followed by submersion in 95% molecular-grade ethanol, DEPC-

treated water, and 95% molecular-grade ethanol.

**Figure 1.** (A) Anatomical landmarks in hand-drawn coronal sections of Eurasian tree sparrow

brains depicting the three target regions (shown in blue) as well as the relative size and location

of the punches (dashed circles). The top image (1) is the most anterior slice and depicts the

striatum (StM) and its landmark, the quinto-frontal tract (QF). The middle image (2) depicts the

rostral hippocampus (rHP) along with its target landmark, the septomesencephalic tract (TrSM),

as well as a rough depiction of the optic tectum (TeO). The bottom image (3) is the most

posterior of the drawings, and depicts the caudal hippocampus (cHP), along with its target

landmark, the cerebellum (Cb) at its first emergence. (B) Principal component analysis of gene

expression patterns, showing the first two principal components (PC), which explain 72% of the

variance. Points represent male (triangles) and female (circles) sparrows across three brain

regions: striatum (red), caudal hippocampus (yellow), and rostral hippocampus (blue). Sample

size = 13 males and 11 females per region.

*RNA extraction and library preparation*

RNeasy Lipid Tissue Mini Kits (Qiagen; 1023539) were used to extract RNA from brain

tissue and RNA concentrations and quality assessed using a Bioanalyzer system (Agilent 5400).

Some samples initially resulted in RNA concentrations too low for library preparation.

Therefore, 13 samples were evaporated down in a sterile incubator until target concentrations for

library preparation were achieved. The incubator was sterilized with D/RNase free and 100%

molecular-grade ethanol before being brought to 45°C. Samples were evaporated for no more

than 3 h. Final sample quality was assessed using the Bioanalyzer, and average total RIN score

for all samples was 8.6 (range: 6.7-9.7).

*RNA sequencing, alignment, and mapping*

We submitted total RNA to Novogene for sequencing using 150 bp paired-end

sequencing (NovaSeq X Plus system) to generate an average of ~52.4 million reads per sample

(Table S2). After filtering to remove adapters and low-quality nucleotides, average reads per

sample was ~49.7 million. The error rate of sequencing for all samples was 0.01 and the G-C

percentage was 48%.We used the zebra finch genome (GCA_003957565.4) as a reference (Rhie

et al., 2021), as it is the most closely related species with a well-annotated genome (Hauber,

Louder, & Griffith, 2021; Kumar et al., 2022). We aligned reads using HISAT2 (Mortazavi et al.,

2008), for a 28.6% (range: 25.7-30.7%) mapping rate, with roughly ~14 million reads per

sample. We used StringTie to assemble transcripts (Pertea et al., 2015). We ran a principal

component analysis (PCA) to visualize transcriptomic patterns across brain regions and sexes

(Figure 1B).

*Identification of differentially expressed genes*

263            We first identified sex- and region-specific genes that were differentially expressed

between individuals showing the most consistent behavioral traits. We categorized individuals as

"consistently neophobic" or "consistently non-neophobic" if their latency to approach or feed

was above (more neophobic) or below (less neophobic) the median for novel object, novel object

habituation, and novel food trials. Across all 6 behavioral trials, there were 7 individuals (n = 4

males; n = 3 females) that were consistently neophobic and 7 individuals (n = 4 males; n = 3

females) that were consistently non-neophobic (Figure S4). We acknowledge this binary

approach reduced sample size, but it allowed us to focus on the most pronounced differences in

gene expression associated with neophobia. Although latencies across trials were highly

correlated, behavioral responses may have varied in non-binary ways; therefore, we also used

gene co-expression networks to analyze the full spectrum of individual variation in each trial (see

below).

275            Differentially expressed genes (DEGs) and normalized values were determined using

DESeq2 v.1.42.0 in R (Love et al., 2014). Transcripts with < 10 counts in the smallest group

sample size (n = 3 for females and n = 4 for males) were filtered out (n = 8,888 transcripts

retained for female caudal hippocampus; n = 8,976 male caudal hippocampus; n = 8,957 female

rostral hippocampus; n = 9,180 male rostral hippocampus; n = 8,869 female striatum; n = 8,772

male striatum). We performed Wald tests for each gene comparing non-neophobic vs neophobic

(reference) individuals by sex within each brain region. Log2 fold change estimates were shrunk

using the *apeglm* function in DESeq2. P-values were corrected using Benjamini-Hochberg

corrections and FDR ≤ 0.05 were considered differentially expressed.

284            DEGs were assessed for enrichment of biological process Gene Ontology (GO) terms

using PANTHER (Mi et al. Thomas, 2019) with a Fisher's Exact test and a cut-off of FDR ≤

0.05. GO terms were further summarized with REVIGO (http://revigo.irb.hr/), which clusters

GO terms based on semantic similarity (similarity threshold = 0.7).

*Construction of weighted gene co-expression networks*

Next, we performed a weighted gene co-expression network analysis (WGCNA) in each

brain region (n = 24 individuals; 11 females and 13 males) to determine how whole networks of

putatively co-regulated genes, rather than individual genes, relate to the full range of variation in

latency to approach and feed during trials (Langfelder & Horvath, 2008). We created networks

within brain regions because gene expression patterns were more region- than sex-specific

(Figure 1B). Using the normalized counts from DESeq2, we filtered out genes with low counts

(< 10 norm counts in the smallest group size; n = 11 for females) and low variability (25% least

variable transcripts) within the hippocampus and striatum (n = 6,640 transcripts retained in cHp

and rHp; n = 6,528 in striatum). Sample clustering was performed to detect sample-level outliers.

We then generated a signed hybrid network by selecting a soft threshold power ($\beta$) = 3 for the

caudal hippocampus, $\beta$ = 9 for the rostral hippocampus, and $\beta$ = 3 for the striatum in accordance

with scale-free topology (signed $R^2 > 0.80$; Figure S1). We used a biweight mid-correlation

(bicor) function and modules were calculated using a minimum module size of 30. A threshold

of 0.25 (merging modules with a correlation $\geq 0.75$) was used to merge modules in Dynamic

Tree Cut (Figure S2). Genes not assigned to a module were classified to the color gray. All other

gene modules were assigned a random color.

*Identifying gene networks of interest*

Expression levels for each module were summarized by the first principal component

(module eigengene), which we used to test for an association between modules and the average

time to approach or feed in each behavioral assay using the bicor function. We examined these

relationships across and within sexes. We additionally included sex and body size (wing chord,

tarsus length, and post-captivity mass) to account for these potentially confounding variables.

We assessed each trait-associated module (defined as $p < 0.05$ and bicor $> |0.60|$) for enrichment

of biological process GO terms using PANTHER (Mi et al., 2019). We only assessed genes with

a significant module membership (MM; the correlation between the gene expression profile and

module eigengene), defined as $p < 0.05$ and MM $> 0.60$.

We identified intramodular hub genes in trait-associated modules, as these genes are

potential drivers of the patterns highlighted in these modules. We defined intramodular hub

genes as those with a high trait-based gene significance (the correlation between a gene's

expression profile and a trait; GS > |0.60|) for at least one module-related trait of interest and

high module membership (MM > 0.80), with a threshold of $p < 0.05$ for both (Horvath & Dong,

2008). We additionally looked for overlap between intramodular hub genes and DEGs as these

genes have strong trait-related expression, making them core candidate hub genes. We identified

whether these genes were found in the most enriched GO terms and visualized networks using

Cytoscape (Shannon et al., 2003). Networks were filtered to only show edges supported by

protein-protein interaction (PPI) data from the Search Tool for the Retrieval of Interacting

Genes/Proteins (STRING) online database (https://string-db.org/) (Szklarczyk et al., 2019), with

a combined score threshold of 0.4. We calculated the degree score for each gene, and those in the

top 10% were classified as PPI hub genes.

To assess functional similarity across the two hippocampal subregions, we looked for

consensus between the caudal and rostral hippocampus network modules by assessing similarity

in module structure and gene membership. Modules were considered overlapping if they shared a

significant number of genes, as determined by Fisher's exact test. This approach allowed us to

identify modules representing similar transcriptional profiles across brain regions. We

additionally examined whether neophobia-related expression patterns emerged across brain

regions by performing a permutational multivariate analysis of variance (PERMANOVA). This

multivariate approach assesses group differences in the combined eigengene profile instead of

testing each module separately. We compared sample-level module eigengenes from all

behavioral trait-related modules across sex, neophobia phenotype, and their interaction in

consistently neophobic and non-neophobic sparrows (n = 3 neophobic and 4 non-neophobic

males; n = 3 neophobic and 3 non-neophobic females) (Figure S4). Module eigengenes

summarize the expression pattern of a module and reflect how strongly a sample aligns with the

overall module profile, allowing comparisons of co-expression patterns across networks within

samples. We used the "adonis2" function in the R package *vegan* (Oksanen et al., 2019), with

Euclidean distance, and visualized sample differences using PCA.

**Results**

*Distinct transcriptomic profiles by brain region*

One of our goals was to examine whether Eurasian tree sparrows showed distinct patterns

of constitutive gene expression in the striatum as well as in different parts of the hippocampus.

Out of 8,935 genes, the majority (89.1%) were expressed in all three brain regions, though each

region had ~100-200 uniquely expressed genes (Figure S3). A PCA revealed that the three brain

regions had distinct transcriptomic profiles (Figure 1B). PC1 explained 63% of the variance and

partitioned the striatum from both hippocampal regions, while PC2 explained 9% of the variance

and partitioned the hippocampal regions. Overall, these data suggest that the caudal and rostral

hippocampus had distinct transcriptomic profiles, but were more similar to each other than to the

striatum.

*Differentially expressed genes in neophobic and non-neophobic individuals by region and sex*

Another goal was to compare sex- and region-specific differences in constitutive gene

expression in consistently neophobic and non-neophobic individuals. As published previously

(Krajcir et al., 2024), there were no sex differences in neophobia behaviors (p > 0.45) (Figure

S4). Neither sex nor neophobia phenotype affected sparrows' captive body mass (p > 0.07 at all

time points). The greatest differences in constitutive gene expression were observed in the caudal

hippocampus of females, followed by the rostral hippocampus of males (Figure S5). In females,

there were n = 257 DEGs between non-neophobic and neophobic individuals in the caudal

hippocampus (Figure S6A), n = 2 DEGs in the rostral hippocampus, and n = 4 DEGs in the

striatum. In males, there were n = 2 DEGs between non-neophobic and neophobic individuals in

the caudal hippocampus, n = 64 DEGs in the rostral hippocampus (Figure S6B), and n = 6 DEGs

in the striatum. Only four DEGs were present in multiple contrasts, including neurotensin (NTS)

in the male striatum and female caudal hippocampus, LOC100225836 in female striatum and

caudal hippocampus, NBEAL2 in female striatum and caudal hippocampus, and NAPG in male

rostral hippocampus and female caudal hippocampus (for all DEGs see Table S3).

Significantly enriched biological processes were only identified for DEGs in the female

caudal hippocampus (Table S5). DEGs upregulated in neophobic females relative to non-

neophobic females were most significantly enriched for "cytoplasmic translation" and "ribosome

biogenesis", while DEGs upregulated in non-neophobic females were most enriched for "neuron

cellular homeostasis" and "learning or memory", as well as other processes like "cellular

responses to BDNF" and "Gamma-aminobutyric acid (GABA) metabolic processes". While no

GO biological processes were found, several of the DEGs in other regions are involved in

neuroendocrine processes (e.g., SCG2 and NTS in the male striatum and RIT2 in the male caudal

hippocampus), neuron signaling (e.g., GRM2 in the female striatum and GRIK2 in male rostral

hippocampus), and stress responsiveness (e.g., OPRK1 in female rostral hippocampus).

**Figure 2**. (A) Module-trait relationships in the Eurasian tree sparrow caudal hippocampus were

identified using bicor tests in WGCNA. The colors of each gene module were assigned

randomly. Associations are shown for sex (females as reference) and latency to approach in

behavioral trials across and within sexes. "Object Approach" refers to the latency to approach a

food dish with a novel object; "Food Approach" a food dish containing a novel food; and

"Habituation Approach" a food dish with the same (initially novel) object over four days. For

each individual, we averaged across multiple trials for each novel object, novel food, and

habituation approach time measure. Correlation coefficients are shown with p-values in

parentheses. The colors on the heatmap are determined by the magnitude and direction of the

correlation coefficient, with darker shades of red indicating a stronger positive correlation and

darker shades of blue indicating a stronger negative correlation. (B) Expression of genes in the

blue (top) and turquoise (bottom) modules were significantly associated with neophobic

behaviors in females but not males. Relative expression of genes with MM > 0.8 is shown across

food approach times (in seconds). Gray lines connect genes across samples. Z-scores reflect

standardized gene expression: $Z = (x - \mu)/\sigma$, where $\mu$ and $\sigma$ are the mean and standard

deviation across all samples and x is the expression of the gene in a specific sample.

*Trait-associated modules in the caudal hippocampus*

We also used co-expression network analyses to examine which genes in the

hippocampus and striatum were most associated with the full range of neophobic or non-

neophobic behaviors. Within the caudal hippocampus, WGCNA constructed 9 modules (Table

S5), several of which demonstrated trait-related patterns in females but not males (Figure 2A).

The blue module was positively associated with novel food and novel object habituation

approach times in females, while the turquoise module was negatively associated with these

traits (Figures 2A, 2B). Genes in the blue module were most enriched for terms linked to

translation (e.g., "cytoplasmic translation" and "ribosome assembly") and energy production

(e.g., "aerobic respiration" and "proton motive force-driven mitochondrial ATP synthesis")

(Figure 3A; Table S6), suggesting co-regulated genes involved in these processes are linked to

slower approach times to novel food items and slower habituation to novel objects in females.

Several of the intramodular hub genes in the blue module were differentially expressed between

consistently neophobic and non-neophobic females (n = 38 genes), of which 12 genes were also

PPI hub genes (Figure S7A), and the overlap between these intramodular hub genes and

differentially expressed genes was significant (Fisher's exact test, $p < 0.01$). These core

candidate hub genes in the blue module were linked to some of the most enriched pathways

(Figure 3A), suggesting they could be drivers of these processes.

Genes in the turquoise module were enriched for terms like "learning or memory",

"chemical synaptic transmission", and "insulin receptor signaling pathway" (Figure 3B; Table

S6), suggesting co-regulated genes involved in these processes are linked to faster approach

times to novel food items and repeated presentations of the same initially novel object in

females. Several of the intramodular hub genes in the turquoise module were also differentially

expressed between consistently neophobic and non-neophobic females (n = 78 genes), of which

14 genes were also PPI hub genes (Figure S7B), and the overlap between these intramodular hub

genes and differentially expressed genes was significant (Fisher's exact test, $p < 0.0001$). These

strong candidate hub genes were again linked to some of the most enriched biological processes

(Figure 3B). Additional trait-related modules included the green and yellow modules that had

genes expressed higher in males compared to females, and the magenta module that had genes

with elevated expression in females relative to males (Figure S8). The magenta module was also

negatively correlated with novel food feeding time (Figure S8). Functional enrichment results for

all trait-associated modules can be found in Table S6.

**Figure 3**. Barplot of the 10 most significant (i.e., lowest FDR) Gene Ontology (GO) biological

processes and GOChord plot of associated genes in the (A) blue and (B) turquoise modules of

the caudal hippocampus of Eurasian tree sparrows. Genes are linked to their assigned GO

biological processes via colored ribbons and ordered according to their module membership. All

genes shown are intramodular hubs and were differentially expressed between consistently

neophobic and non-neophobic individuals.

*Trait-associated modules in the rostral hippocampus*

In the rostral hippocampus, WGCNA constructed 8 modules (Table S7), several of which

demonstrated trait-related patterns in males but not females (Figures 4A, S9). In males, the

greenyellow module was positively associated with novel food approach time, while the black

module was negatively associated with this behavior (Figure 4B). The purple module was

negatively associated with novel object approach time and time to feed in the presence of a novel

object in males (Figures 4A, S9). Genes in the greenyellow module were most enriched for terms

related to cholesterol biosynthesis, steroid metabolism, and neurotransmitters ("GABA metabolic

process" and "glutamate secretion") (Figure 5A; Table S6), suggesting co-regulated genes

involved in these processes are associated with slower approach times to novel food items in

males. One intramodular hub gene in the greenyellow module was differentially expressed

between consistently neophobic and non-neophobic males (SLC1A3), and it was also a PPI hub

gene (Figure S10A). SLC1A3 is linked to the following enriched GO terms: "transepithelial

transport", "GABA metabolic process", and "behavior". Genes in the black module were

enriched for terms related to transcription and post-transcriptional modifications, specifically

mRNA splicing (Figure 5B; Table S6), suggesting co-regulated genes involved in these

processes are linked to faster approach times to novel food items in males. Four intramodular

hub genes in the black module were differentially expressed between consistently neophobic and

non-neophobic males (MED13, SRSF7, RBBP6, and LOC100229022), of which SRSF7 was

also a PPI hub gene (Figure S10B). SRSF7 is linked to the following enriched GO terms:

"regulation of mRNA splicing, via spliceosome" and "mRNA cis splicing, via spliceosome".

Genes in the purple module were most enriched for a post-translational modification (i.e.,

"peptidyl-serine phosphorylation"; Table S6), suggesting this process could be linked to faster

novel object approach and feeding times in males. One intramodular hub gene in the purple

module was also differentially expressed between consistently neophobic and non-neophobic

males (MACF1). Additional trait-related modules included the green and brown modules that

had genes expressed higher in males and females, respectively (Figure S9). Of note, several

critical stress-related genes, including CRH, NR3C1, and CRHR2 were assigned to the brown

module. Functional enrichment results for all trait-associated modules can be found in Table S6.

**Figure 4.** (A) Module-trait relationships in the sparrow rostral hippocampus were identified

using bicor tests in WGCNA. The colors of each gene module were assigned randomly.

Associations are shown for sex (females as reference) and latency to approach in behavioral

trials across and within sexes. "Object Approach" refers to the latency to approach a food dish

with a novel object; "Food Approach" a food dish containing a novel food; and "Habituation

Approach" a food dish with the same (initially novel) object over four days. For each individual,

we averaged across multiple trials for each novel object, novel food, and habituation approach

time measure. Correlation coefficients are shown with p-values in parentheses. The colors on the

heatmap are determined by the magnitude and direction of the correlation coefficient, with

darker red shades indicating a stronger positive correlation and darker blue shades indicating a

stronger negative correlation. (B) Expression of genes in the black (top) and greenyellow

(bottom) modules were significantly associated with neophobic behaviors in males but not

females. Relative expression of genes with MM > 0.8 is shown across food approach times (in

seconds). Gray lines connect genes across samples. Z-scores reflect standardized gene

expression: $Z = (x − μ)/σ$, where μ and σ are the mean and standard deviation across all

samples and x is the expression of the gene in a specific sample.

**Figure 5.** Barplot of the most significant (lowest FDR) Gene Ontology biological processes in
the (A) greenyellow and (B) black modules of the rostral hippocampus of Eurasian tree
sparrows.

*Trait-associated modules in the striatum*

One outlier sample was removed from the striatum dataset (Figure S11). WGCNA
constructed 12 modules in the striatum (Table S8), several of which demonstrated trait-related
patterns in both males and females (Figures S12A, S13). The lightcyan module was negatively
associated with multiple behavioral metrics in males, while the midnight blue, purple, and black
modules were positively associated with neophobia metrics in males. The brown and red
modules were positively associated with neophobia behavioral metrics in females. The most
significant relationships were between the brown module and female novel object habituation
approach time and the lightcyan module and male novel food approach time (Figure S12B).

Genes in the brown module were most enriched for terms linked to cellular transport and
organization (i.e., "regulation of vesicle-mediated transport", "positive regulation of organelle
organization", and "regulation of catabolic processes"; Table S6), suggesting co-regulated genes
involved in these processes are linked to greater neophobia in females. There was no overlap
between DEGs and intramodular hub genes in the brown module. Genes in the lightcyan module
had no enriched terms, but one intramodular hub gene (HTT) was also differentially expressed
between consistently neophobic and non-neophobic males, and it was also a PPI hub gene
(Figure S14). Genes in the midnight blue module were enriched for terms related to sensory
development (i.e., "visual system development" and "inner ear auditory receptor cell
differentiation") and signaling (i.e., notch and BMP signaling pathway terms), suggesting genes
linked to these processes are elevated in more neophobic males, but there was no overlap

between DEGs and intramodular hub genes. The black module was also most enriched for

sensory system terms (i.e., "trigeminal nerve structural organization", "branchiomotor neuron

axon guidance", "G protein-coupled opioid receptor signaling pathway", "sensory perception of

pain", "adult behavior", and "sensory system development") and signaling processes (i.e.,

"neuropeptide signaling pathway" and "regulation of neuronal synaptic plasticity"), but there

was no overlap between DEGs and intramodular hub genes. Genes in the purple module had no

enriched terms, but one intramodular hub gene was also differentially expressed between

consistently neophobic and non-neophobic males (SCG2). The red module was most enriched

for mRNA splicing, suggesting genes linked to this process were elevated in more neophobic

females, but there was no overlap between DEGs and intramodular hub genes. Additional trait-

related modules included the yellow module that had genes expressed higher in males than

females. Functional enrichment results for all trait-associated modules can be found in Table S6.

*Patterns across brain regions*

Motivated by initial observations of transcriptional similarity, and to better understand

shared patterns across the two subregions of the hippocampus and their sex-specific associations,

we examined consensus between rostral and caudal hippocampal modules. We found significant

gene overlap between neophobia-associated modules in the female caudal hippocampus and

male rostral hippocampus, but relationships between neophobia and module gene expression

were often reversed in the two sexes (Figure S14). Significant overlap in trait-associated modules

included the blue module in the caudal hippocampus and the black module in the rostral

hippocampus, the turquoise module in the caudal hippocampus and the greenyellow module in

the rostral hippocampus, and the yellow module in the caudal hippocampus and green module in

the rostral hippocampus (Figure 6). The blue-black module overlap involved 278 genes

(constituting 77.9% of the black module genes) that were positively correlated with novel food

approach times in the female caudal hippocampus (Figure 2B), but negatively correlated with

this trait in the male rostral hippocampus (Figure 4B; Figure S15A). Genes found in both

modules that had a MM $> 0.80$ and were significantly associated with behavioral traits were

linked to transcription and post-transcriptional (e.g., mRNA splicing) processes (Figure S15

A), suggesting that these processes may be elevated in the rostral hippocampus of less neophobic

males and in the caudal hippocampus of more neophobic females. The turquoise-greenyellow

overlap involved 212 genes (constituting 88.3% of the greenyellow module genes) that were

negatively correlated with novel food approach times in the female caudal hippocampus (Figure

2B), but positively correlated with this trait in the male rostral hippocampus (Figure 4B; Figure

S15A). Trait-related genes with high MM found in both modules were linked to chemical

synaptic transmission and processes related to GABA and dopamine (Figure S16B), suggesting

these processes may be elevated in the caudal hippocampus of less neophobic females and in the

rostral hippocampus of more neophobic males. The yellow-green overlap involved 184 genes

that were elevated in males in both brain regions.

We also examined whether neophobia-related expression patterns emerged across brain

regions using the most consistently neophobic and non-neophobic individuals, as previously

categorized for the differential expression analysis. We found that module eigengene profiles

across brain regions significantly differed by behavioral phenotype. PERMANOVA results

showed a significant main effect of neophobia ($F_{1,9} = 3.44$, p = 0.006), but not sex ($F_{1,9} = 0.98$, p

= 0.45). There was a trend for the neophobia-by-sex interaction to be significant (neophobia by

sex: $F_{1,9} = 1.96$, p = 0.08), thus we analyzed the principal component scores within each sex

separately. Among males, more neophobic sparrows had significantly lower PC1 scores ($F_{1,5} =$

13.04, p = 0.015), but no difference in PC2 ($F_{1,5} = 1.87$, p = 0.23). Among females, more

neophobic sparrows had significantly lower PC2 scores ($F_{1,4} = 10.71$, p = 0.03), but not PC1 ($F_{1,4}$

= 0.003, p = 0.96). These differences were evident in the PCA, where the first two PCs explained

61.1% of the total variance in trait-related module eigengenes (Figure S17A). Lower PC1 scores

in neophobic males reflected relatively lower expression of genes in the lightcyan module

(striatum) and black module (rostral hippocampus), and higher expression in the greenyellow

module (rostral hippocampus) (Figure S17B). In contrast, lower PC2 scores in neophobic

females reflected higher expression in the brown module (striatum) and blue module (caudal

hippocampus), and lower expression in the turquoise module (caudal hippocampus) (Figure S17

B). These results suggest that region-specific co-expression patterns are not isolated but instead

co-occur within individuals in a behavior- and sex-dependent manner, likely reflecting

coordinated transcriptional patterns across multiple brain regions.

**Figure 6.** Correspondence between gene co-expression modules in the caudal and rostral hippocampus of Eurasian tree sparrows. Each row represents a module from the caudal hippocampus, and each column represents a module from the rostral hippocampus. Cell values indicate the number of transcripts shared between modules. Color intensity reflects the significance of module overlap; darker red color indicates a more significant overlap as calculated using Fisher's exact test and shown as -log10(p-value).

**Discussion**

We identified region-specific, constitutive transcriptomic patterns associated with neophobia in female and male Eurasian tree sparrows (*Passer montanus*). The striatum and hippocampus had distinct transcriptomic profiles, as did the rostral and caudal subregions of the hippocampus. The number of differentially expressed genes between neophobic and non-neophobic individuals varied across brain regions and sexes. However, the full complexity of the molecular mechanisms underlying neophobic behaviors was better captured using WGCNA, which revealed that neophobia was associated with similar gene modules between sexes, but their expression was localized to different hippocampal subregions in males and females. In females, neophobic behaviors were associated with gene modules in the caudal hippocampus that were enriched for synaptic transmission, steroid metabolism, and post-transcriptional processes,

whereas in males, neophobic behaviors were associated with similar gene modules in the rostral

hippocampus. Altogether, these findings suggest that although neophobic behaviors are similar

between sexes, they may be driven by region-specific neurobiological pathways. Below, we

discuss these results and their implications for our understanding of the neurotranscriptomic

mechanisms related to neophobia.

*Brain regions had distinct transcriptomic profiles, differentiating the caudal and rostral*

*hippocampus*

Brain region was a major driver of variation in gene expression. Though most genes were

expressed in the striatum, caudal hippocampus, and rostral hippocampus, each region had ~100-

200 uniquely expressed genes. We found that the caudal and rostral hippocampus had distinct

transcriptomic profiles, supporting the hypothesis that the avian hippocampus may show

subregional specialization along a rostral-caudal axis, similar to the dorsal-ventral axis of the

rodent hippocampus (Fanselow & Dong, 2010; Gualtieri et al., 2019; Smulders, 2017).

Specifically, the rostral hippocampus may play a larger role in spatial cognition, while the caudal

hippocampus may be more involved in anxiety, stress, and avoidance responses (Madison et al.,

2023). A recent study in black-capped chickadees also supports the functional specialization of

these regions in songbirds, as the caudal hippocampus received more inputs from amygdalar

regions involved in emotional responses, whereas the rostral hippocampus received more inputs

from the thalamus, which plays a key role in relaying sensory and motor information (Applegate

et al., 2023). To our knowledge, our study is the first to confirm that the rostral and caudal

hippocampus express different transcriptomic profiles in birds. Future transcriptomic

comparisons between the rostral and caudal subregions can further clarify whether different parts

of the hippocampus are specialized for different functions in birds as well as mammals.

The striatum was most differentiated from the caudal and rostral hippocampus, as would

be predicted based on its distinct structure and functions. Though subregional specializations of

the striatum have not been well-explored in birds (Mezey & Csillag, 2002), the striatum of

mammals is typically divided into the dorsal striatum, encompassing the caudate and putamen,

and the ventral striatum, which includes the nucleus accumbens and the olfactory tubercle

(Voorn et al., 2004). Future work could explore the transcriptomic profiles of striatal structures

in birds as a means to better characterize potential striatal subregions.

*Neophobia-associated gene modules differed by brain region and sex*

While prior work with this same group of Eurasian tree sparrows (as well as studies of

the congeneric house sparrow) found no sex difference in neophobic behaviors (Ensminger et al.,

2012; Kelly et al., 2020; Krajcir et al., 2024), we identified sex-specific patterns of neophobia-

associated gene expression. Specifically, neophobic behaviors correlated with gene expression in

the caudal hippocampus of females and the rostral hippocampus of males. Both categorical and

continuous comparisons of gene expression related to neophobia revealed this sex-by-region

interaction. Though more work is needed to understand the drivers of sex-specific gene

expression in Eurasian tree sparrows, one hypothesis is that not only does the hippocampus show

subregional specialization in birds, but that neural processes in these subregions vary by sex.

Notably, our finding in birds aligns with numerous studies from rodents showing that cognitive

performance-related behaviors are associated with neurogenesis in the ventral hippocampus of

females, and in the dorsal hippocampus of males (Dalla et al. 2009, Yagi et al. 2016, Yagi &

Galea 2019). In rats, there are numerous sex differences in hippocampal anatomy that include

dendritic spine density in the CA1 subregion (Shors et al., 2001; Woolley et al., 1990),

morphology of CA3 pyramidal neurons (Juraska et al., 1989), and dendritic branching in the

dentate gyrus (Juraska et al., 1985). Parallel differences may exist in the hippocampus of male

and female birds, but this remains to be determined. This rodent work, together with the present

study, suggest that sex differences in hippocampal processes may be conserved across

vertebrates.

Our findings highlight that neophobia is regulated by sex-specific regionalization of gene

expression, and suggest putative candidates for genes and processes that may play a conserved

role in neophobia across species. In the caudal hippocampus, the most neophobic females had

higher expression of genes involved in energy production and transcription/mRNA

splicing/translation (blue module). The latter processes could increase transcriptomic diversity

and functional complexity (Mauger & Scheiffele, 2017), yielding more diverse molecular routes

to respond to novel stimuli. In contrast, non-neophobic females had higher expression of genes

involved in various forms of signaling activity (turquoise module). This included GO terms

related to learning or memory, synaptic transmission, and GABA biosynthesis. GABA is a key inhibitory neurotransmitter, and its synthesis depends on glutamic acid decarboxylases (GAD1 and GAD2), both of which were differentially expressed hub genes. Expression of these enzymes is highly correlated with GABA levels and inhibitory synaptic neurotransmission (Dicken et al., 2015; Lee et al., 2019), which shape neuronal activity and contribute to contextual learning in the hippocampus (Lovett-Barron et al., 2014). Additional differentially expressed hub genes included NTSR1 and CNR1, which are implicated in hippocampal learning and memory (Li et al., 2011; Ruiz-Contreras et al., 2013), as well as in modulating stress responses and anxiety-like behaviors (Kyriatzis et al., 2024; Martin et al., 2002). Thus, differences in the constitutive expression of these genes may influence how individuals respond to novel and potentially threatening stimuli. In the rostral hippocampus, the most neophobic males showed higher expression of genes involved in signaling activity, including GABA and steroids (greenyellow module), but lower expression of genes linked to transcription/mRNA splicing (black module). Note that although we measured gene expression three weeks after behavioral trials ended, it is possible that exposure to novel stimuli initiated lasting effects on neuroplasticity that were still ongoing at the time of tissue collection. Alternatively, these genes may have differed in expression prior to the behavioral trials, and may explain some of the variation in neophobic responses. Regardless, these patterns suggest that neophobic males rely on specific signaling pathways rather than broad transcriptional flexibility in a hippocampal subregion involved in spatial processing, whereas non-neophobic females demonstrate a similar strategy but in a subregion associated with stress and anxiety.

Overall, our results suggest that basic neurobiological processes vary among individuals with different behavioral phenotypes, and that some parallel mechanisms may regulate neophobia in both sexes but in different brain regions and in opposite directions. This supports so-called 'Type III' sex differences (sensu (McCarthy et al., 2012)), where convergent behaviors are underlain by divergent neurobiological processes in females and males. What remains to be determined is how these individual and sex-specific differences in gene expression arise; is variation driven by innate neurobiological processes, or is it shaped by flexibly interacting with the environment? Activational and/or organizational effects of sex steroids could underlie these sex differences (Adkins-Regan, 2005). From a life-history standpoint, female songbirds are often more sensitive to stress-induced reproductive suppression, which could reflect sex-specific

differences in brain regions involved in responding to potential stressors, like the hippocampus
(Calisi et al., 2018; Hirschenhauser et al., 2003; Ouyang et al., 2012; Schmidt et al., 2014). As
more studies (finally!) include both males and females, we encourage sex contextualist
approaches (Pape et al., 2024; Richardson, 2022; Smiley et al., 2024) that assess the possibility
of sex-related variation in neurobiology even in the absence of behavior differences, as we would
not have discovered these sex-specific associations between neophobia and neural gene
expression otherwise.

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<div align="center">**References**</div>

Adkins-Regan, E. (2005). *Hormones and Animal Social Behavior*. Princeton, NJ: Princeton
    University Press.

Applegate, M. C., Gutnichenko, K. S., & Aronov, D. (2023). Topography of inputs into the
    hippocampal formation of a food-caching bird. *Journal of Comparative Neurology,
    531*(16), 1669-1688. doi:https://doi.org/10.1002/cne.25533

Bell, A. M., & Robinson, G. E. (2011). Behavior and the dynamic genome. *Science, 332*(6034),
    1161. doi:10.1126/science.1203295

Bengston, S. E., Dahan, R. A., Donaldson, Z., Phelps, S. M., van Oers, K., Sih, A., & Bell, A. M.
    (2018). Genomic tools for behavioural ecologists to understand repeatable individual
    differences in behaviour. *Nat Ecol Evol, 2*(6), 944-955. doi:10.1038/s41559-017-0411-4

Breck, S. W., Poessel, S. A., Mahoney, P., & Young, J. K. (2019). The intrepid urban coyote: a
comparison of bold and exploratory behavior in coyotes from urban and rural
environments. *Sci Rep, 9*(1), 2104. doi:10.1038/s41598-019-38543-5

Brivio, E., Lopez, J. P., & Chen, A. (2020). Sex differences: Transcriptional signatures of stress
exposure in male and female brains. *Genes, brain and behavior, 19*(3), e12643.

Calisi, R. M., Austin, S. H., Lang, A. S., & MacManes, M. D. (2018). Sex-biased transcriptomic
response of the reproductive axis to stress. *Horm Behav, 100*, 56-68.
doi:10.1016/j.yhbeh.2017.11.011

Calisi, R. M., & MacManes, M. D. (2015). RNAseq-ing a more integrative understanding of
animal behavior. *Current Opinion in Behavioral Sciences, 6*, 65-68.
doi:https://doi.org/10.1016/j.cobeha.2015.09.007

Candler, S., & Bernal, X. E. (2015). Differences in neophobia between cane toads from
introduced and native populations. *Behavioral Ecology, 26*(1), 97-104.
doi:10.1093/beheco/aru162

Cohen, T. M., Kumar, R. S., Nair, M., Hauber, M. E., & Dor, R. (2020). Innovation and
decreased neophobia drive invasion success in a widespread avian invader. *Animal
Behaviour, 163*, 61-72. doi:10.1016/j.anbehav.2020.02.012

Crane, A. L., & Ferrari, M. C. O. (2017). Patterns of predator neophobia: a meta-analytic review.
*Proc Biol Sci, 284*(1861). doi:10.1098/rspb.2017.0583

Dicken, M. S., Hughes, A. R., & Hentges, S. T. (2015). Gad1 mRNA as a reliable indicator of
altered GABA release from orexigenic neurons in the hypothalamus. *Eur J Neurosci,
42*(9), 2644-2653. doi:10.1111/ejn.13076

Dickens, M.J., Romero, L.M. (2009). Wild European Starlings (*Sturnus vulgaris*) adjust to
captivity with sustained sympathetic nervous system drive and a reduced fight-or-flight
response. *Physiological and Biochemical Zoology,* 82(5),603–10.

Dusang, B. A., Henry, M. S., Kimball, M. G., Cochran, E. B., Wilson, M. B., & Lattin, C. R.
(2025). Spatial neophobia is still not correlated with object neophobia in wild-caught
house sparrows (Passer domesticus). *Royal Society Open Science, 12*(5), 250220.

Ensminger, A. L., Westneat, D. F. (2012). Individual and sex differences in habituation and
neophobia in house sparrows (Passer domesticus). *Ethology, 118*(11), 1085-1095.
doi:10.1111/eth.12009

Fair, J. E., Paul, E., Jones, J., & Bies, L. (2023). *Guidelines to the Use of Wild Birds in Research*.
Washington D.C.: Ornithological Council.

Fanselow, M. S., & Dong, H. W. (2010). Are the dorsal and ventral hippocampus functionally
distinct structures? *Neuron, 65*(1), 7-19. doi:10.1016/j.neuron.2009.11.031

Greenberg, R., & Mettke-Hofmann, C. (2001). Ecological aspects of neophobia and neophilia in
birds. In V. Nolan & C. F. Thompson (Eds.), *Current Ornithology* (Vol. 16). Boston,
MA: Springer.

Grunst, A. S., Grunst, M. L., Pinxten, R., & Eens, M. (2019). Personality and plasticity in
neophobia levels vary with anthropogenic disturbance but not toxic metal exposure in
urban great tits: Urban disturbance, metal pollution and neophobia. *Sci Total Environ,*
*656*, 997-1009. doi:10.1016/j.scitotenv.2018.11.383

Gualtieri, F., Armstrong, E. A., Longmoor, G. K., D'Eath, R. B., Sandilands, V., Boswell, T., &
Smulders, T. V. (2019). Unpredictable chronic mild stress suppresses the incorporation of
new neurons at the caudal pole of the chicken hippocampal formation. *Sci Rep, 9*(1),
7129. doi:10.1038/s41598-019-43584-x

Guigueno, M. F., MacDougall-Shackleton, S. A., & Sherry, D. F. (2016). Sex and seasonal
differences in hippocampal volume and neurogenesis in brood-parasitic brown-headed
cowbirds (Molothrus ater). *Dev Neurobiol, 76*(11), 1275-1290. doi:10.1002/dneu.22421

Güntürkün, O. (2005). The avian 'prefrontal cortex' and cognition. *Current Opinion in*
*Neurobiology, 15*(6), 686-693. doi:http://dx.doi.org/10.1016/j.conb.2005.10.003

Hauber, M. E., Louder, M. I., & Griffith, S. C. (2021). Neurogenomic insights into the
behavioral and vocal development of the zebra finch. *Elife, 10*. doi:10.7554/eLife.61849

Hirschenhauser, K., Winkler, H., & Oliveira, R. F. (2003). Comparative analysis of male
androgen responsiveness to social environment in birds: the effects of mating system and
paternal incubation. *Horm Behav, 43*(4), 508-519. doi:10.1016/s0018-506x(03)00027-8

Horvath, S., & Dong, J. (2008). Geometric interpretation of gene coexpression network analysis.
*PLoS Comput Biol, 4*(8), e1000117. doi:10.1371/journal.pcbi.1000117

Huang, P., St.Mary, C. M., & Kimball, R. T. (2020). Habitat urbanization and stress response are
primary predictors of personality variation in northern cardinals (Cardinalis cardinalis).
*Journal of Urban Ecology, 6*(1), juaa015. doi:10.1093/jue/juaa015

Jarjour, C., Evans, J. C., Routh, M. l., & Morand-Ferron, J. (2019). Does city life reduce
neophobia? A study on wild black-capped chickadees. *Behavioral Ecology, 31*(1), 123-
131. doi:10.1093/beheco/arz167

Juraska, J. M., Fitch, J. M., Henderson, C., & Rivers, N. (1985). Sex differences in the dendritic
branching of dentate granule cells following differential experience. *Brain research,*
*333*(1), 73-80.

Juraska, J. M., Fitch, J. M., & Washburne, D. L. (1989). The dendritic morphology of pyramidal
neurons in the rat hippocampal CA3 area. II. Effects of gender and the environment.
*Brain research, 479*(1), 115-119.

Kelly, T. R., Kimball, M. G., Stansberry, K. R., & Lattin, C. R. (2020). No, you go first:
phenotype and social context affect house sparrow neophobia. *Biology Letters, 16*,
20200286.

Kimball, M. G., Gautreaux, E. B., Couvillion, K. E., Kelly, T. R., Stansberry, K. R., & Lattin, C.
R. (2022). Novel objects alter immediate early gene expression globally for ZENK and
regionally for c-Fos in neophobic and non-neophobic house sparrows. *Behav Brain Res,*
*428*, 113863. doi:10.1016/j.bbr.2022.113863

Kimball, M. G., & Lattin, C. R. (2023a). Exploration of A novel environment is not correlated
with object neophobia in wild-caught house sparrows (Passer domesticus). *Behav*
*Processes, 210*, 104913. doi:10.1016/j.beproc.2023.104913

Kimball, M. G., & Lattin, C. R. (2023b). The "Seven Deadly Sins" of Neophobia Experimental
Design. *Integr Comp Biol*. doi:10.1093/icb/icad127

Klein, S. L., Schiebinger, L., Stefanick, M. L., Cahill, L., Danska, J., De Vries, G. J., . . .
Woodruff, T. K. (2015). Sex inclusion in basic research drives discovery. *Proceedings of*
*the National Academy of Sciences, 112*(17), 5257-5258.

Krajcir, K. J., Kelly, T. R., Kimball, M. G., Cochran, E. B., Stansberry, K. R., Dusang, B. A., . . .
Lattin, C. R. (2024). Eurasian tree sparrows are more food neophobic and habituate to
novel objects more slowly than house sparrows. *Biological Invasions*.
doi:10.1007/s10530-024-03403-5

Kumar, S., Suleski, M., Craig, J. M., Kasprowicz, A. E., Sanderford, M., Li, M., . . . Hedges, S.
B. (2022). TimeTree 5: An Expanded Resource for Species Divergence Times. *Mol Biol*
*Evol, 39*(8). doi:10.1093/molbev/msac174

Kyriatzis, G., Khrestchatisky, M., Ferhat, L., & Chatzaki, E. A. (2024). Neurotensin and
neurotensin receptors in stress-related disorders: Pathophysiology & novel drug targets.
*Current neuropharmacology, 22*(5), 916-934.

Langfelder, P., & Horvath, S. (2008). WGCNA: an R package for weighted correlation network
analysis. *BMC Bioinformatics, 9*, 559. doi:10.1186/1471-2105-9-559

Lattin, C. R., Kelly, T. R., Kelly, M. W., & Johnson, K. M. (2022). Constitutive gene expression
differs in three brain regions important for cognition in neophobic and non-neophobic
house sparrows (Passer domesticus). *PLoS One, 17*(5), e0267180.
doi:10.1371/journal.pone.0267180

Lee, S. E., Lee, Y., & Lee, G. H. (2019). The regulation of glutamic acid decarboxylases in
GABA neurotransmission in the brain. *Arch Pharm Res, 42*(12), 1031-1039.
doi:10.1007/s12272-019-01196-z

Li, J., Chen, C., Chen, C., He, Q., Li, H., Li, J., . . . Dong, Q. (2011). Neurotensin receptor 1
gene (NTSR1) polymorphism is associated with working memory. *PLoS One, 6*(3),
e17365.

Love, M. I., Huber, W., & Anders, S. (2014). Moderated estimation of fold change and
dispersion for RNA-seq data with DESeq2. *Genome Biol, 15*(12), 550.
doi:10.1186/s13059-014-0550-8

Lovett-Barron, M., Kaifosh, P., Kheirbek, M. A., Danielson, N., Zaremba, J. D., Reardon, T. R.,
. . . Losonczy, A. (2014). Dendritic inhibition in the hippocampus supports fear learning.
*Science, 343*(6173), 857. doi:10.1126/science.1247485

Madison, F. N., Bingman, V. P., Smulders, T. V., & Lattin, C. R. (2023). A bird's eye view of
the hippocampus beyond space: Behavioral, neuroanatomical, and neuroendocrine
perspectives. *Horm Behav, 157*, 105451. doi:10.1016/j.yhbeh.2023.105451

ManyBirds Project, Miller, R., Šlipogor, V., Caspar, K.R., Lois-Milevicich, J., Soulsbury, C., et
al. (2025). A large-scale study across the avian clade identifies ecological drivers of
neophobia. *PLoS Biol,* 23(10): e3003394. https://doi.org/10.1371/journal.pbio.3003394

Martin, L. B., & Fitzgerald, L. (2005). A taste for novelty in invading house sparrows, Passer
domesticus. *Behavioral Ecology, 16*(4), 702-707. doi:10.1093/beheco/ari044

Martin, M., Ledent, C., Parmentier, M., Maldonado, R., & Valverde, O. (2002). Involvement of
CB1 cannabinoid receptors in emotional behaviour. *Psychopharmacology, 159*(4), 379-
387.

Mauger, O., & Scheiffele, P. (2017). Beyond proteome diversity: alternative splicing as a
regulator of neuronal transcript dynamics. *Curr Opin Neurobiol, 45*, 162-168.
doi:10.1016/j.conb.2017.05.012

McCarthy, M. M., Arnold, A. P., Ball, G. F., Blaustein, J. D., & De Vries, G. J. (2012). Sex
differences in the brain: the not so inconvenient truth. *J Neurosci, 32*(7), 2241-2247.
doi:10.1523/JNEUROSCI.5372-11.2012

Medina, L., & Reiner, A. (1995). Neurotransmitter organization and connectivity of the basal
ganglia in vertebrates: implications for the evolution of basal ganglia. *Brain Behav Evol,*
*46*(4-5), 235-258. doi:10.1159/000113277

Mezey, S., & Csillag, A. (2002). Selective striatal connections of midbrain dopaminergic nuclei
in the chick (Gallus domesticus). *Cell and tissue research, 308*(1), 35-46.

Mi, H., Muruganujan, A., Ebert, D., Huang, X., & Thomas, P. D. (2019). PANTHER version 14:
more genomes, a new PANTHER GO-slim and improvements in enrichment analysis
tools. *Nucleic Acids Res, 47*(D1), D419-D426. doi:10.1093/nar/gky1038

Mortazavi, A., Williams, B. A., McCue, K., Schaeffer, L., & Wold, B. (2008). Mapping and
quantifying mammalian transcriptomes by RNA-Seq. *Nature Methods, 5*(7), 621-628.
doi:10.1038/nmeth.1226

Nixdorf-Bergweiler, B. E., & Bischof, H.-J. (2007). *A stereotaxic atlas of the brain of the zebra*
*finch, Taeniopygia guttata, with special emphasis on telencephalic visual and song*
*system nuclei in transverse and sagittal sections. Bethesda (MD).*

Oakley, R. H., Riddick, N. V., Moy, S. S., & Cidlowski, J. A. (2024). Imbalanced glucocorticoid
and mineralocorticoid stress hormone receptor function has sex-dependent and
independent regulatory effects in the mouse hippocampus. *Neurobiology of stress, 28*,
100589.

Oksanen, J., Blanchet, F. G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., . . . Wagner, H.
(2019). vegan: Community Ecology Package. https://CRAN.R-
project.org/package=vegan.

Olszewski, P. K., Ulrich, C., Ling, N., Allen, K., & Levine, A. S. (2014). A non-peptide oxytocin
868            receptor agonist, WAY-267,464, alleviates novelty-induced hypophagia in mice: insights
869            into changes in c-Fos immunoreactivity. *Pharmacology Biochemistry and Behavior, 124*,
870            367-372.

871    Ouyang, J. Q., Quetting, M., & Hau, M. (2012). Corticosterone and brood abandonment in a
872            passerine bird. *Animal Behaviour, 84*(1), 261-268. doi:10.1016/j.anbehav.2012.05.006

873    Pape, M., Miyagi, M., Ritz, S. A., Boulicault, M., Richardson, S. S., & Maney, D. L. (2024). Sex
874            contextualism in laboratory research: Enhancing rigor and precision in the study of sex-
875            related variables. *Cell, 187*(6), 1316-1326. doi:https://doi.org/10.1016/j.cell.2024.02.008

876    Pertea, M., Pertea, G. M., Antonescu, C. M., Chang, T. C., Mendell, J. T., & Salzberg, S. L.
877            (2015). StringTie enables improved reconstruction of a transcriptome from RNA-seq
878            reads. *Nat Biotechnol, 33*(3), 290-295. doi:10.1038/nbt.3122

879    Petersen, K., & Sherry, D. F. (1996). No sex difference occurs in hippocampus, food-storing, or
880            memory for food caches in black-capped chickadees. *Behavioural brain research, 79*(1-
881            2), 15-22.

882    Ramírez-Lugo, L., Zavala-Vega, S., Pedroza-Llinas, R., Núñez-Jaramillo, L., & Bermúdez-
883            Rattoni, F. (2015). Effects of glutamate and its metabotropic receptors class 1 antagonist
884            in appetitive taste memory formation. *Behavioural Brain Research, 284*, 213-217.

885    Rhie, A., McCarthy, S. A., Fedrigo, O., Damas, J., Formenti, G., Koren, S., . . . Jarvis, E. D.
886            (2021). Towards complete and error-free genome assemblies of all vertebrate species.
887            *Nature, 592*(7856), 737-746. doi:10.1038/s41586-021-03451-0

888    Richardson, S. S. (2022). Sex contextualism. *Philosophy, Theory, and Practice in Biology, 14*, 2.
889            doi:https://doi.org/10.3998/ptpbio.2096

890    Roman, C., Lin, J.-Y., & Reilly, S. (2009). Conditioned taste aversion and latent inhibition
891            following extensive taste preexposure in rats with insular cortex lesions. *Brain research,*
892            *1259*, 68-73.

893    Rose, J., & Colombo, M. (2005). Neural correlates of executive control in the avian brain. *PLoS*
894            *Biol, 3*(6), e190. doi:10.1371/journal.pbio.0030190

895    Royet, J.-P., Gervais, R., & Araneda, S. (1983). Effect of local 6-OHDA and 5, 6-DHT injections
896            into the rat olfactory bulb on neophobia and learned aversion to a novel food.
897            *Behavioural brain research, 10*(2-3), 297-309.

Ruiz-Contreras, A. E., Carrillo-Sánchez, K., Gómez-López, N., Vadillo-Ortega, F., Hernández-
Morales, S., Carnevale-Cantoni, A., . . . Prospéro-García, O. (2013). Working memory
performance in young adults is associated to the AATn polymorphism of the CNR1 gene.
*Behavioural brain research, 236*, 62-66.

Schaffer, A., Caicoya, A.L., Colell, M., Holland, R., von Fersen, L., Widdig, A., Amici, F.
(2021). Neophobia in 10 ungulate species-a comparative approach. *Behav Ecol Sociobiol.*
75(7):102. doi: 10.1007/s00265-021-03041-0.

Schmidt, K. L., MacDougall-Shackleton, E. A., Soma, K. K., & MacDougall-Shackleton, S. A.
(2014). Developmental programming of the HPA and HPG axes by early-life stress in
male and female song sparrows. *General and Comparative Endocrinology, 196*, 72-80.
doi:https://doi.org/10.1016/j.ygcen.2013.11.014

Semenov, G. A., Sonnenberg, B. R., Branch, C. L., Heinen, V. K., Welklin, J. F., Padula, S. R., .
. . Taylor, S. A. (2024). Genes and gene networks underlying spatial cognition in food-
caching chickadees. *Current Biology, 34*(9), 1930-1939. e1934.

Shansky, R. M., & Murphy, A. Z. (2021). Considering sex as a biological variable will require a
global shift in science culture. *Nature neuroscience, 24*(4), 457-464.

Shephard, R., Buxton, D., & Broadhurst, P. (1982). β-adrenoceptor antagonists may attenuate
hyponeophagia in the rat through a serotonergic mechanism. *Pharmacology Biochemistry*
*and Behavior, 16*(5), 741-744.

Sherry, D. F., Forbes, M., Khurgel, M., & Ivy, G. O. (1993). Females have a larger hippocampus
than males in the brood-parasitic brown-headed cowbird. *Proceedings of the National*
*Academy of Sciences, 90*(16), 7839-7843.

Shinohara, K., & Yasoshima, Y. (2021). Inactivation of the ventral hippocampus facilitates the
attenuation of odor neophobia in rats. *Behavioural Brain Reseach, 401*, 113077.
doi:10.1016/j.bbr.2020.113077

Shors, T. J., Chua, C., & Falduto, J. (2001). Sex differences and opposite effects of stress on
dendritic spine density in the male versus female hippocampus. *Journal of Neuroscience,*
*21*(16), 6292-6297.

Simpson, K. L., Weaver, K. J., de Villers-Sidani, E., Lu, J. Y.-F., Cai, Z., Pang, Y., . . . Lin, R.
C. (2011). Perinatal antidepressant exposure alters cortical network function in rodents.
*Proceedings of the National Academy of Sciences, 108*(45), 18465-18470.

Smiley, K. O., Munley, K. M., Aghi, K., Lipshutz, S. E., Patton, T. M., Pradhan, D. S., . . . Sun,

S. D. (2024). Sex diversity in the 21st century: Concepts, frameworks, and approaches for

the future of neuroendocrinology. *Hormones and Behavior, 157*, 105445.

doi:https://doi.org/10.1016/j.yhbeh.2023.105445

Smulders, T. V. (2017). The avian hippocampal formation and the stress response. *Brain Behav*

*Evol, 90*(1), 81-91. doi:10.1159/000477654

Stehberg, J., Moraga-Amaro, R., & Simon, F. (2011). The role of the insular cortex in taste

function. *Neurobiology of learning and memory, 96*(2), 130-135.

Steketee, J. D., Silverman, P. B., & Swann, A. C. (1992). Noradrenergic receptor mechanisms in

neophobia. *Psychopharmacology, 106*(1), 136-142.

Stokes, T. M., Leonard, C. M., & Nottebohm, F. (1974). The telencephalon, diencephalon, and

mesencephalon of the canary, Serinus canaria, in stereotaxic coordinates. *Journal of*

*Comparative Neurology, 156*, 337-374.

Szklarczyk, D., Gable, A. L., Lyon, D., Junge, A., Wyder, S., Huerta-Cepas, J., . . . Mering, C.

V. (2019). STRING v11: protein-protein association networks with increased coverage,

supporting functional discovery in genome-wide experimental datasets. *Nucleic Acids*

*Res, 47*(D1), D607-D613. doi:10.1093/nar/gky1131

Tryjanowski, P., Moller, A. P., Morelli, F., Biadun, W., Brauze, T., Ciach, M., . . . Zduniak, P.

(2016). Urbanization affects neophilia and risk-taking at bird-feeders. *Sci Rep, 6*, 28575.

doi:10.1038/srep28575

Voorn, P., Vanderschuren, L. J., Groenewegen, H. J., Robbins, T. W., & Pennartz, C. M. (2004).

Putting a spin on the dorsal–ventral divide of the striatum. *Trends in neurosciences,*

*27*(8), 468-474.

Widmann, O. (1889). History of the House Sparrow Passer domesticus and the European Tree

SparrowPasser montanus at Saint Louis, Mo. *Washington DC: US Department of*

*Agriculture, Division of Economic Ornithology and Mammology*.

Wolak, M.E., Fairbairn, D.J., Paulsen, Y.R. (2012). Guidelines for estimating repeatability.

*Methods Ecol Evol* 3:129–137.

Woolley, C. S., Gould, E., Frankfurt, M., & McEwen, B. S. (1990). Naturally occurring

fluctuation in dendritic spine density on adult hippocampal pyramidal neurons. *Journal of*

*neuroscience, 10*(12), 4035-4039.

Yagi, S., & Galea, L. A. (2019). Sex differences in hippocampal cognition and neurogenesis.

961             *Neuropsychopharmacology, 44*(1), 200-213.