A morphometric approach to the taxonomic dilemma of *Zonozoe drabowiensis* Barrande, 1872

and *Zonoscutum solum* Chlupáč, 1999 (Upper Ordovician, Czech Republic)

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ABSTRACT

Zonozoe drabowiensis Barrande, 1872 and *Zonoscutum solum* Chlupáč, 1999 are rare and incompletely preserved arthropods from the Upper Ordovician of the Czech Republic. Their classification has been a subject of debate for over a century due to the limited number of specimens, lack of knowledge related to post-cephalic morphology and the absence of clear diagnostic features. They were previously considered as members of Aglaspidida, an extinct group of arthropods from the Cambrian and Ordovician, within Vicissicaudata, a branch of the larger arthropod clade Artiopoda. Herein, we analysed the cephalic outlines of *Zonozoe* and *Zonoscutum* to determine whether their shapes align more closely with vissicaudatans than with other early Palaeozoic arthropods, offering a new morphological perspective on their systematics. We assembled a data set of cephalic outlines each representing one of thirty-three early Palaeozoic species, including *Zonozoe*, *Zonoscutum*, nine euchelicerates, six aglaspidids, three cheloniellids, and a selection of other artiopodans. We quantified their shape using elliptical Fourier analysis. Principal component analysis (PCA) and linear discriminant analysis (LDA) place *Zonozoe* within the vicissicaudatan morphospace, and *Zonoscutum* in their proximity, clearly distinguishing them from euchelicerates. These data add support to the most conservative classification of *Zonozoe* and *Zonoscutum* within Artiopoda, while strengthening the case for a more specific affinity with Vicissicaudata, helping to resolve a 150-year-old taxonomic uncertainty. More broadly, this study demonstrates the value of outline-based morphometrics in aiding systematic hypotheses when discrete characters are unavailable or scarce, offering a reproducible tool for re-evaluating other problematic fossils.

Key words: *Zonozoe drabowiensis*, *Zonoscutum solum,* elliptical Fourier analysis, Vicissicaudata, Artiopoda

INTRODUCTION

Zonozoe drabowiensis Barrande, 1872 and *Zonoscutum solum* are enigmatic arthropods known from the Letná and the Libeň Formations of the upper Ordovician of the Czech Republic (Chlupáč, 1965, 1999b; Rak et al., 2009; Vokáč & Grigar, 2024). Barrande (1872) originally described *Zonozoe* as an arthropod, specifically as a large carapace of an ostracod. Later on, Novák (1887) in his unpublished manuscript considered *Zonozoe* to be a merostomate arthropod close to Synziphosurina (Chelicerata) (Chlupáč, 1963, 1965, 1999b; Rak, 2009). Since then, these fossils have not received much attention until the second half of the twentieth century. Chlupáč (1963) assigned *Zonozoe* to the group Aglaspidida (Walcott, 1911) – a group of extinct marine arthropods, known from the Cambrian and Ordovician strata. However, this assignment remains uncertain (Chlupáč, 1963). It should also be noted that at that time, Aglaspidida was considered as one of the basal-most groups within Xiphosura (Chelicerata). Subsequent studies have revisited the taxon, reiterating its general association with Aglaspidida and/or the Merostomata, but always with a low degree of confidence (Chlupáč, 1965; Hou, 1997; Chlupáč, 1999a; Rak et al., 2009). As for *Zonoscutum*, the only fossil ascribed to this species was described in Chlupáč (1999a), who tentatively assigned it to Aglaspidida.

During the one hundred and fifty years since the discovery of *Zonozoe*, and twenty-five years for *Zonoscutum,* their classification has been complicated by two main factors. Firstly, the material ascribed to both *Zonozoe* and *Zonoscutum* is scarce, incomplete and not well preserved (Ortega Hernández et al., 2013; Van Roy et al., 2022). There are currently only eight known specimens of *Zonozoe* and only one specimen of *Zonoscutum,* all of which are isolated moulds of cephala (Chlupáč, 1999b; Rak et al., 2009; Vokáč & Grigar, 2024), preserved in quartzose sandstones of the Libeň and Letná Formations. In most cases, the specimens preserve only the general vaulted cephalon outline with a raised longitudinal region terminated with a ridge bearing what is a vague indication of the eyes (Rak et al., 2009; Van Roy et al., 2022). The only exception is a single specimen of *Zonozoe* preserving what has been referred to as glabellar furrows (Rak et al.,

2009) and a specimen of *Zonoscutum* preserving similar structures but in a different position

(Chlupáč, 1999a). Secondly, the classification of *Zonozoe* and *Zonoscutum* has been further

complicated by the complex history of the classification of aglaspidid arthropods themselves,

especially during the second half of the twentieth century. Their overall dorsal morphology led

several authors to consider the aglaspidids as an order of the subclass Xiphosura (Størmer, 1955),

grouping Aglaspidida with Xiphosura and Eurypterida, as part of the Merostomata (Raasch, 1939;

Størmer, 1944; Eldredge & Smith, 1974) or to consider Aglaspidida as the sister group to

chelicerates (Weygoldt and Paulus, 1979). This is in contrast to more recent aglaspidid phylogenies,

which emphasize ventral anatomical features as key to understanding their true affinities (Briggs et

al., 1979; Hesselbo, 1992; Ortega□Hernández et al., 2013; Lerosey-Aubril, Paterson, et al., 2017;

Lerosey-Aubril, Zhu, et al., 2017). Currently, they have been placed within the broader clade

Artiopoda, as part of the clade Vicissicaudata (Ortega□Hernández et al., 2013; Lerosey-Aubril,

Paterson, et al., 2017; Lerosey-Aubril, Zhu, et al., 2017). Until very recently, the absence of strong

defining characteristics for the clade has made Aglaspidida prone to being used as a wastebasket

taxon (Plotnick & Wagner, 2006). This could be the case for the poorly preserved *Zonozoe* and

Zonoscutum, which lack strong anatomical evidence supporting placement within aglaspidids. Other

factors that led previous workers to assign *Zonozoe* and *Zonoscutum* to Aglaspidida are likely not

only the general similarity between their cephala, but also the known (or suspected) presence of

aglaspidids in the same paleogeographic region (Chlupáč, 1963, 1965; Van Roy, 2005; Ortega-

Hernandez et al., 2010) and the weak resemblance with other arthropod groups (e.g. position of the

eyes and glabella if compared with trilobites).

Later authors commenting on *Zonozoe* and *Zonoscutum* decided to have a more parsimonious

approach. Ortega□Hernández et al. (2013) opted for the exclusion of those taxa from an extensive

phylogenetic analysis of Vicissicaudata due to the lack of compelling taxonomically relevant

features. Van Roy et al. (2022), instead, revised *Zonozoe* and *Zonoscutum* to critically assess their

possible affinity with *Triopus draboviensis* Barrande, 1872. In their re-examination of the material,

the authors did more than simply dismiss the affinity with *Triopus*; they advanced a conservative

and more parsimonious taxonomic classification for both *Zonozoe* and *Zonoscutum*. They suggested

that, at most, the fossils could be broadly classified as Artiopoda, and argued that any attempt to

refine the classification beyond that level would be unwarranted (Van Roy et al., 2022). Known

only from cephalic moulds, both *Zonozoe* and *Zonoscotum* lack the diagnostic features of

Vicissicaudata, all of which are post-cephalic (Lerosey-Aubril, Zhu, et al., 2017; Briggs et al.,

2023). They also lack several defining traits of Aglaspidida, including all post-cephalic diagnostic

characters, most notably the modified posterior appendages, as well as evidence of a biomineralized

cuticle and dorsal ecdysial sutures (Ortega⬜Hernández et al., 2013; Lerosey-Aubril, Zhu, et al.,

2017). As a result, they cannot be confidently classified through standard phylogenetic or

comparative analysis (Ortega⬜Hernández et al., 2013; Van Roy et al., 2022), a conclusion fully

supported by this study. While the enigmatic *Zonozoe* and *Zonoscutum* are unlikely to be key taxa

in contributing to resolving the phylogeny of Aglaspidida, a more accurate assignment of this

species to this group may yield valuable palaeoecological and paleobiogeographical insights.

In order to contribute to resolving the taxonomic dilemma surrounding *Zonozoe* and *Zonoscutum*

more effectively, we adopted an alternative approach to phylogenetic analysis and classical

comparative anatomy, focusing instead on an overlooked well preserved feature in these fossils: the

overall cephalic outline. We compared the outline of the cephalon of *Zonozoe* and *Zonoscutum* with

thirty-three other arthropods. We implemented a principal component analysis (PCA) and linear

discriminant analysis (LDA) on the study of the cephalic outline to extract the maximum

morphological information from the limited material of *Zonozoe* and *Zonoscutum*. This

methodology has been used in paleobiology to compare the outline of different fossils and can

provide information on their taxonomy, ecology, and development (Gevirtz, 1976; Healy-Williams

& Williams, 1981; Burke et al., 1987; Foote, 1989; Pates et al., 2021; Ma et al., 2023; Braig et al.,

2024; Drage & Pates, 2024; Collantes & Pates, in press). A significant advantage of this

methodology over landmark-based approaches is that it does not rely on discrete morphological

characters, facilitating quantification of shape variation when such discrete characters are absent.

Combined with the classical taxonomic work carried out by various authors over the years

(Chlupáč, 1963, 1965, 1999b; Rak, 2009), our approach offers a new line of evidence, contributing

to either confirming or rejecting previous hypotheses about the taxonomic affinities of these fossils.

Both *Zonozoe* and *Zonoscutum* plotted within the Vicissicaudata area of morphospace, and thus the

shape of their cephala concurs with a vicissicaudate affinity.

MATERIALS AND METHODS

Species involved in the study and the criteria of selection

In addition to one *Zonozoe* and one *Zonoscutum* specimen (Figure 1), nine euchelicerates, two

nektaspids, six non-vicissicaudatans artiopodans and fourteen vicissicaudatans arthropods

(including six aglaspidids and three cheloinellids), representing thirty-three species, were included

in our study. Trilobites were excluded for several reasons: 1) While Cambrian and/or Ordovician

nektaspids, euchelicerates, and vicissicaudatans are morphologically rather conservative—at least in

the general shape of their cephala/prosomas (cf. Fig. 1; Lerosey-Aubril et al., 2017; Pérez-Peris et

al., 2021)—this is not the case for trilobites. The cephalic morphology of trilobites is extremely

diverse, particularly during the Ordovician period (e.g., Drage & Pates, 2024; Balseiro et al., 2025),

and such diversity would necessarily introduce substantial noise into our dataset. This study aims to

build on previous taxonomic work and, given the limitations of our methodology, to avoid

introducing additional variability that would likely reduce the predictability of our analyses; (2)

Zonozoe and *Zonoscutum* have never been considered closely related to Trilobita; and (3) the

position of the eyes and glabella in *Zonozoe* and *Zonoscutum* differs markedly from that of any

contemporaneous trilobite.

The most complete and horizontally preserved specimen of *Zonozoe* (Lectotype NM L 23586) was

chosen, while *Zonoscutum* is known from only one specimen (Holotype NM L 33021). The

selection from the literature of other taxa, was made following four main criteria: 1) to include

Early Palaeozoic diversity from various geographic regions, but with particular emphasis on species

coeval and sympatric to *Zonozoe* and *Zonoscutum* (e.g. *Paraeurypterus anatoliensis* Lamsdell et al.

2013); 2) to exclude taxa with aberrant cephalic outlines; 3) to favour those potentially related to

Zonozoe and *Zonoscutum* (e.g. *Eozetetes gemmelli* Edgecombe et al. 2017 and *Chlupacaris dubia*

Van Roy, 2005); and 4) to maximize higher-level taxonomic diversity (e.g. *Carimersa neptuni*

Briggs et al., 2023 and *Lunataspis borealis* Lamsdell et al., 2022). A summary of the species and

specimens included in the analysis, along with inventory numbers, geological age, geographic

provenance, and references to the original sources from which raw images were obtained, is

provided in Table 1, while the specimen sample, one for each high taxonomic group, is shown in

Figure 1.

Material and photography

The studied material of *Zonozoe* and *Zonoscutum* is housed in the palaeontological collections of

the National Museum, Prague (prefix NM L). The photographs of *Zonozoe* were taken using a Sony

Alpha A7R V digital camera equipped with a Canon EF 100 mm f/2.8 Macro lens under normal

lighting conditions, using a stand and a sandbox to hold the sample in place and ensure the

horizontality of both the camera and the specimen. Specimens were coated with ammonium

chloride prior to photography to enhance morphological details.

Geological settings of Zonozoe *and* Zonoscutum

The examined specimens of *Zonozoe* and *Zonoscutum* (Figure 1) come from the Ordovician

deposits of the Prague Basin (Czech Republic, Barrandian area). All *Zonozoe* and *Zonoscutum*

specimens represent internal moulds preserved as imprints in coarse quartzose sandstones, showing

limited anatomical details. The majority of them were collected at the Děd Hill near Beroun (five

specimens of *Zonozoe*; NM L 32986, NM L 23590, NM L 33029, NM L 23586, NM L 32 987) or in

the fields between Trubín and Trubská villages near Králův Dvůr (one specimen of *Zonozoe* NM L

33029; see Drage et al. (2018)for details of the location), in the upper part of the Letná Formation

(Chlupáč, 1965). The only known cephalon of *Zonoscutum* (NM L 33021) also originates from this

latest formation and locality (Chlupáč, 1999a). Additionally, one specimen of *Zonozoe* (NM L

37385) was recovered from the Řevnice Quartzite Member within the Libeň Formation near Kařez

(Rak et al., 2009), and another one has been recently reported from the Řevnice Quartzite Member

near Starý Plzenec (Vokáč & Grigar, 2024).

The Libeň Formation is developed in three different members – the shallow water Řevnice

Quartzite, the sequence of black shales deposited in deeper sea below the wave base in anoxic

conditions, and the pyroclastic material (Havlíček, 1998). The Letná Formation is generally

composed of alternating quartzose sandstones, greywackes, siltstones, and occasional shales, with

sandstones representing a shallow-water environment (Chlupác & Kukal, 1988; Kraft et al., 2023).

Zonozoe and *Zonoscutum* have so far been discovered only in the shallow water facies of Libeň and

Letná Formations. Both Letná and Libeň Formations are of Sandbian age (Upper Ordovician) (Kraft

et al., 2023). From the palaeogeographical perspective, these formations within the Prague Basin

were deposited in high-latitude area near the western margin of Gondwana (Kraft et al., 2023;

Scotese, 2023).

Silhouette drawing, Principal component analysis (PCA), linear discriminant analysis (LDA)

We used photographs of selected specimens to make black silhouettes of carapace outlines on a

white background for selected specimens. Photographs were either taken by the authors in museum

collections or sourced from the literature (see Table 1 and Figure 1). Using Adobe Illustrator 2021,

we manually traced each specimen's outline over the photograph, then exported the silhouette as a

JPG file (Figure 3A). These image files served as input for the Momocs package in R (Bonhomme

et al., 2014; RCoreTeam, 2024), which we used to perform the subsequent outline analyses. JPG

files were imported and converted to outlines, scaled, centered, and sampled at 64-point resolution,

and subjected to elliptical Fourier analysis. The function calibrate_harmonicpower_efourier was

used to determine the number of harmonics required for 99.9% of the harmonic power. Elliptical

Fourier coefficients were visualised using Principal Component Analysis. Linear Discriminant

Analysis was used to model differences between the major groupings in the dataset (Artiopoda

(non-Vicissicaudata), Euchelicerata, Vicissicaudata, *Zonozoe, Zonoscutum*). R code used for the

analysis, and silhouettes in .jpg format, are supplied in Supplementary Information as Additional

file 1 and 2 respectively.

RESULTS

EFA, Principal component analysis (PCA), linear discriminant analysis (LDA)

Fifteen harmonics were retained, providing 99.9% of the harmonic power for the dataset. Principal

component analysis reduced the dimensionality of elliptical Fourier analysis results, with PC1 and

PC2 explaining 87.1% of the variation (67.4% and 19.7% respectively). Higher principal

components (PC3–PC26) explain <5% of the variation each, and <15% overall. All groups overlap

at least to some extent in the PCA space: Euchelicerata, Vicissicaudata, and Non☐Vicissicaudata

artiopodans (Figure 3B). Euchelicerates cluster toward the upper left, while non☐Vicissicaudatan

artiopodans occupy the area close to the origin. Vicissicaudata occupy a broader area in the

morphospace, including most of the area occupied by non-Vicissicaudata, as well as areas more

positive in PC1 and more negative in both PC1 and PC2. Within Vicissicaudata, aglaspidids fall

near the origin of morphospace, except for *Tremaglaspis unite* (Fortey & Rushton, 2003), which

plots with a negative PC1 value (Figures☐3B and C). Cheloniellids form a tight cluster with

negative PC2 values. Both *Zonozoe* and *Zonoscutum* lie within the lower region of the

vicissicaudatan space. *Zonozoe* plots between the cheloniellid *Neostrabops martini* (Caster &

Macke, 1952) and the aglaspidid *Aglaspis barrandei* (Hall, 1862)(Figures☐3B and C), whereas

Zonoscutum lies close to the cheloniellids *Triopus* and *Drabovaspis complexa* (Barrande, 1872) and

the artiopodan *Naraoia* (Figures☐3B and C).

Linear discriminant analysis is dominated by LD1 (74.6% of the trace), with LD2 (13.9%), with

remaining LDs providing only a relatively small amount of information. Our LDA (Figure 3D)

separates Euchelicerata from all other taxa but cannot separate vicissicaudates from non-

vicissicaudate artiopodans (Table 2). Leave-one-out cross-validation was 48.4% (15/31) for the

whole dataset, but 77.8% (7/9) for Euchelicerata. Non Vicissicaudata artiopodans fall into the

vicissicaudatans space, with leave-one-out cross-validation poor for both these groups, with most

incorrect assignments placing vicissicaudatans in the non-vicissicaudatan space and vice versa

(Table 2). Both *Zonozoe* and *Zonoscutum* plot at the lower right part of the LDA space, outside of

all other groups but close to the Vicissicaudata/non-Vicissicaudata areas, far from the Euchelicerata

area (Figure 3D).

DISCUSSION

Our outline analyses provide some morphological support for an artiopodan affinity for both

Zonozoe and Zonoscutum. This was the recently lowest taxonomic level assigned to this species

through the weak morphological traits with taxonomic relevance available. The PCA does not

explicitly support an affinity with Aglaspidida, particularly in the case of *Zonoscutum.* Instead, both

taxa are placed closer to cheloniellids than to other groups, in an intermediated position towards

aglaspidids and some non-vicissicaudatan artiopodans (Figures 3B and C).

Although our analysis places *Zonozoe and Zonoscutum* near the cheloniellid cluster, we interpret

these results cautiously. Key differences in cephalic morphology, which are most apparent in three-

dimensional views, are not fully captured by our two-dimensional dataset, and both *Zonoscutum*

and cheloniellids follow in this case. They show, respectively, a highly vaulted and extremely

flattened cephalon, which may have been more affected by this methodological constraint,

respectively to what was retrieved for *Zonozoe* and the other involved taxa. We believe this is the

explanation behind the results of our PCA, concerning *Zonoscutum* clustering closer to the well-

defined cheloniellids *Triopus* and *Drabovaspis* than to any other taxa. Based on this, we consider

our method less effective for taxa with extremely vaulted or flattened cephala, such as cheloniellids

and *Zonoscutum*. This limitation might have been addressed by using 3D volume data, but such data

are difficult to obtain for fossils held in collections across multiple continents, and will also be

impacted by flattening during or after fossilisation. Thus, we do not consider the results of our

analyses strong enough to question (Van Roy et al., 2022), who rejected a close relationship

between *Zonozoe*, *Zonoscutum*, and cheloniellids, especially with the coeval and sympatric *Triopus*.

On the other hand, our analyses show low support for the alternative hypothesis of euchelicerate

affinities, as euchelicerates are separated in the PC space from the other groups, with *Zonozoe* and

Zonoscutum plotting far from euchelicerates and within the Vicissicaudata area.

The closest non-cheloniellid arthropod to *Zonoscutum,* is *Naroia*, followed by the euchelicerate

Brachyopterus and the aglaspidid *A. barrandei*. Those results do not allow us to state further

interpretations for *Zonoscutum*. The results for *Zonozoe,* instead, present some clearer insight. The

closest taxa in our PCA to *Zonozoe* are the cheloniellid *Neostrabops* and the aglaspidid *A.*

barrandei. Ortega-Hernández et al. (2013) assigned *Neostrabops* as the less derived Cheloniellida,

on the basis of a shared "presence of rounded genal angles, the anterior boundaries of the trunk

reflexed anterolaterally and a reduced head shield". However, the absence of a well-developed

thoracic axis, already led Van Roy (2006) to suggest it may represent the less derived form of the

group. Other studies (e.g., Legg et al., 2013; Lerosey-Aubril, Zhu, et al., 2017) also placed

Neostrabops as a basal cheloniellid, but again relied on limited synapomorphies. The lack of

preservation of key traits, such as furcal rami or the morphology of terminal tergites, hinders

reliable comparison with better-known forms like *Triopus* or *Cheloniellon calmani* Broili, 1932.

The proximity of *Zonozoe* and *Neostrabops* in our PCA may therefore reflect more on the uncertain

taxonomy of *Neostrabops* than on *Zonozoe* itself. Both fall within the vicissicaudatan morphospace,

closer to aglaspidids than to other cheloniellids. Based on our results, we suggest that the

taxonomical assignment of *Neostrabops* could be reconsidered towards Aglaspidida after new

investigations or, in a more parsimonious way, towards Xenopoda.

On the other hand, the close PCA placement of *Zonozoe* (and *Neostrabops*) with *A. barrandei*

supports a possible affinity with aglaspidids. While we do not draw a definitive conclusion, this

result is consistent with previous observations, such as the position of the eyes and glabellar

furrows behind them, which, while not strongly diagnostic (Van Roy et al., 2022), align with our

morphometric findings and the exclusion of a possible eucheledicerate affinity retrieved in our

analyses (Figure 3).

Our LDA results, though more influenced by predefined categories, provide additional insight.

Euchelicerates are distinguished from Artiopoda, however vicissicaudatans and non-

vicissicaudatans cannot be clearly separated (Figure 3D; Table 2). This was an unexpected finding

for our outline data, given the superficial similarity in dorsal morphology between some early

euchelicerates and vicissicaudatans like aglaspidids (e.g. *P. anatoliensis* and *T. unite*). Previous

authors have even suggested the affinities of euchelicerates with aglaspidids based on such

similarities (Størmer L. 1955), but our LDA analyses were able to discriminate the two groups.

Both taxa of interest, *Zonoscutum* and *Zonozoe,* are positioned closer to non-vicissicaudatan

artiopods than to vicissicaudatans, in contrast with our PCA results, however, they are clearly

separated from Euchelicerata. These results reinforce the more recent interpretation of aglaspidids,

and by extension *Zonozoe* and *Zonoscutum*, as artiopodans, most likely vicissicaudatans, by clearly

distinguishing them from euchelicerates.

CONCLUSION

Our outline⬚based morphometric analyses provide quantitative support for placing *Zonozoe* and

Zonoscutum within artiopodans. Although they plot closer to cheloniellids in the morphospace of

two-dimensional carapace outlines, this could reflect methodological limits due to

three-dimensional cephalic shape being compressed. Both PCA and LDA consistently separate

Vicissicaudata, Aglaspidida, *Zonozoe* and *Zonoscutum* from euchelicerates. Within PCA space, they

plot closest to vicissicaudata artiopods. Taken together, these results support a vicissicaudatan

affinity for these taxa and reject an euchelicerates affinity. Overall, our method has proven to be a

useful alternative for classifying arthropod fossils when discrete characters required for

phylogenetic methods are absent. This is often the case with moulds, which rarely preserve fine

anatomical details. While our approach is less precise than formal phylogenetic analysis, it allows

for the inclusion of specimens that would otherwise be excluded from broader evolutionary or

paleoecological studies. In our case, it provides evidence for the presence of non-cheloniellid

vicissicaudatans in the Letná Formation, based on poorly preserved specimens that would likely be

overlooked using only traditional qualitative methods.

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AUTHOR CONTRIBUTIONS

L.Lu. conceptualized the original idea and wrote the first draft of the manuscript. L.Lu., L.La. and

L.C. collected the data. S.P. analysed the data. L.La. and L.C., and S.P. prepared the figures and

wrote sections of the material and methods, results, and discussion. Y.L., J.B. and M.N. were

involved in the discussion and final editing of the manuscript. All authors read and approved the

final version of the manuscript.

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Figure 1. Representative cephala of *Zonozoe drabowiensis*, *Zonoscutum solum*, euchelicerate

arthropods, vicissicaudatan cheloniellids, vicissicaudatan aglaspidids, and non-vicissicaudatan

artiopodans featured in this study. A1, *Zonozoe drabowiensis*, dorsal view; A2, *Zonozoe*

drabowiensis, frontal view; B, *Zonoscutum solum*, dorsal view, modified from Van Roy et al.

(2022); C, Unnamed Xiphosura from the Fezouata biota, modified from (Lamsdell et al., 2013). D,

Eozetetes gemmelli, modified from (Edgecombe et al., 2017); E, *Neostrabops martini*, modified

from (Caster & Macke, 1952); F, *Aglaspis barrandei*, modified from (Hesselbo, 1992). Scale bar

equals 5 mm.

Figure 2. Geographical and stratigraphical position of the localities where *Zonozoe*

***drabowiensis* and *Zonoscutum solum* were collected.** A, Map of the Czech republic; B,

Stratigraphical column of the upper Ordovician (modify after (Van Roy et al., 2022)).

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Figure 3. Data and Results of Principal Component Analysis and Linear Discriminant

Analysis of elliptical Fourier coefficients. A, Cephalic outline of *Zonozoe drabowiensis*,

Zonoscutum solum and the other specimens involved in the study; colors of silhouettes match colors

for points in panel C. B, Position of *Zonoscutum* and *Zonozoe* in relation to major taxonomic

groupings: Euchelicerata, Artiopoda (not including Vicissicaudata), and Vicissicaudata. Grey

convex hulls show subgroups within Vicissicaudata, specifically Aglaspidida and Cheloniellida.

Coloured marks along x and y axis indicate PC1 and PC2 values of individual specimens plotted in

the space, with positions of specimens shown by filled circles. Grey outlines show variation in

cephalic outline across PC1 and PC2. Histogram in bottom right shows amount of variation

explained by PC1, PC2, and PC3-5; C, results by individual outline of PCA analyses. Abbreviation

'unnamedfez' refers to an unnamed Xiphosura from the Fezouata biota (Van Roy et al., 2010;

Lamsdell & Ocon, 2025); D, Position of *Zonoscutum* and *Zonozoe* in relation to major taxonomic

groupings: Euchelicerata, Artiopoda (not including Vicissicaudata), and Vicissicaudata in Linear

Discriminant space. Coloured circles show positions of individual specimens. Histograms in B and

D the shows the proportion of the trace for each principal component (B, C) or linear discriminant

(D).

Table 1. List of specimens used for the outline analyses. Figures refer to figure number in

Reference column from which outline was taken.

Species	Number	Age and origin	Clade	Reference	Figures
Zonoscutum solum	NM L 33021	Upper Ordovician, Czech Republic	Suspected Aglaspidida	(Van Roy et al., 2022)	Fig. 8C
Zonozoe drabowiensis	NM L 23586	Upper Ordovician, Czech Republic	Suspected Aglaspidida	Present work	Fig. 1E
Beckwithia typa	BPM 1060	Late Cambrian, Utah	Aglaspidida	(Lerosey-Aubril, Paterson, et al., 2017)	Fig. 9F
Chlupacaris dubia	NMSG.2005.103.1	Upper Ordovician, Morocco	Aglaspidida	(Van Roy, 2005)	Fig. 3A
Tremaglaspis unite	NHMIA56	Upper Ordovician, Wales	Aglaspidida	(Fortey & Rushton, 2003)	Fig. 1B
Glypharthrus trispinicaudatus	NIGPAS 165044	Late Cambrian, South China	Aglaspidida	(Lerosey-Aubril, Paterson, et al., 2017)	Fig. 3 G
Chraspedops modesta	MPM 11938	Late Cambrian, US	Aglaspidida	(Hesselbo, 1992)	Fig. 18, 4
Aglaspis barrandei	AMNH 39193	Late Cambrian, Wisconsin	Aglaspidida	(Hesselbo, 1992)	Fig. 2, 5
Xiphosura from Fezouata	YPM IP 532152	Lower Ordovician, Morocco	Euchelicerata	(Lamsdell & Ocon, 2025)	Fig. 2, 7
Paraeurypterus anatoliensis	MTANHMSETR 10-İZ-01-1	Upper Ordovician, Turkey	Euchelicerata	(Lamsdell et al., 2013)	Fig. 3A
Lunataspis borealis	MM I-4583	Upper Ordovician, Canada	Euchelicerata	(Lamsdell et al., 2022)	Fig. 2, 1
Pseudoniscus falcatus	UK. NHMUK PI	Early Silurian, Scottland	Euchelicerata	(Bicknell & Pates, 2020)	Fig. 9A
Kasibelinurus amicorum	AM F68969	Devonian, Australia	Euchelicerata	(Bicknell & Pates, 2020)	Fig. 11A
Kwanyinaspis maotianshanensis	ELI-12004001	Early Cambrian, South China	Artiopoda	(Zhang & Shu, 2005)	Fig. 2A-D
Pygmaclypeatus daziensis	YKLP 11427	Early Cambrian, South China	Artiopoda	(Schmidt et al., 2025)	Fig. 1A
Thulaspis tholops	MGUH 34172a	Early Cambrian, North Greenland	Artiopoda	(Berks et al., 2023)	Fig. 1A
Retifacies abnormalis	JS-840	Early Cambrian, South China	Artiopoda, Trilobitomorpha	(Lin et al., 2024)	Fig. 1A
Kuamaia lata	YKLP 17296	Early Cambrian, South China	Artiopoda, Helmetiidae	(O'Flynn et al., 2024)	Fig. 2A
Emeraldella brocki	USNM 136641	Middle Cambrian, Canada	Artiopoda, Vicissicaudata	(Stein & Selden, 2012)	Fig. 3B
Tardisia broedeae	FMNH PE 88856	Late Carboniferous, Illinois	Artiopoda, Vicissicaudata	(McCoy et al., 2025)	Fig. 1D
Eozetetes gemmelli	SAM P48369a	Early Cambrian, Australia	Artiopoda, Vicissicaudata	(Edgecombe et al., 2017)	Fig. 2A
Carimersa neptuni	OUMNH PAL-C.376503	Early Silurian, England	Artiopoda, Vicissicaudata	(Briggs et al., 2023)	Fig. 1A
Sidneyia minor	YKLP 12435	Early Cambrian, South China	Artiopoda, Vicissicaudata	(Du et al., 2023)	Fig. 2A
Triopus draboviensis	NM L 16736	Upper Ordovician, Czech Republic	Cheloniellida	(Van Roy et al., 2022)	Fig. 2B
Drabovaspis complexa	NM L 23577	Upper Ordovician, Czech Republic	Cheloniellida	(Van Roy et al., 2022)	Fig. 9A
Neostrabops martini	25560	Late Cambrian, Missuri	Cheloniellida	(Caster & Macke, 1952)	Fig. 2

Lunataspis borealis	ROM IP 64616	Upper Ordovician, Canada	Euchelicerata	(Lamsdell et al., 2022)	Fig. 1A
Brachyopterus stubblefieldi	D. 3124	Middle Ordovician, Wales	Euchelicerata	(Størmer, 1951)	Fig. 1
Orcanopterus manitoulinensis	ROM 56450	Upper Ordovician, Canada	Euchelicerata	(Stott et al., 2005)	Fig. 3.1
Chasmataspis laurencii	USNM 125099	Middle Ordovician, US	Euchelicerata	(Dunlop et al., 2003)	Fig. 1B
Arthroaspis bergstroemi	MGUH 30388	Early Cambrian, Greenland	Artiopoda, Conciliterga	(Stein et al., 2013)	Fig. 2E
Naraoia magna	ROMIP 64557	Middle Cambrian, Canada	Artiopoda, Nektaspida	(Mayers et al., 2019)	Fig. 15B
Naraoia arcana	ROMIP 64515	Middle Cambrian, Canada	Artiopoda, Nektaspida	(Mayers et al., 2019)	Fig. 16C

367　**Table 2.** Cross validation table following Linear Discriminant Analysis, showing the classification

368　of groups and the class accuracy. Class accuracy and classification not displayed for *Zonoscutum*

369　and *Zonozoe* as they are represented only by a single specimen.

	Euchelicerata	Non-Vicissicadata	Vicissicaudata	*Zonoscutum*	*Zonozoe*	Class accuracy
Euchelicerata	7	0	1	0	1	77.8%
Non-Vicissicadata	2	1	5	0	0	12.5%
Vicissicaudata	2	5	7	0	0	50%

References

Barrande J. (1872). *Systême silurien du centre de la Bohême: trilobites, crustacés divers et poissons: supplément* (Vol. 1). chez l'auteur et l'éditeur.

Berks H. O., Lunde Nielsen M., Flannery-Sutherland J., Thorshøj Nielsen A., Park T. Y. S., & Vinther J. (2023). A possibly deep branching artiopodan arthropod from the lower Cambrian Sirius Passet Lagerstätte (North Greenland). *Papers in Palaeontology*, *9*(3), e1495.

Bicknell R. D., & Pates S. (2020). Pictorial atlas of fossil and extant horseshoe crabs, with focus on Xiphosurida. *Frontiers in Earth Science*, *8*, 98.

Bonhomme V., Picq S., Gaucherel C., & Claude J. (2014). Momocs: outline analysis using R. *Journal of Statistical Software*, *56*, 1-24.

Braig F., Torres G., Giménez L., & Haug J. T. (2024). Outline analysis as a new method for investigating development in fossil crabs. *Palaeontologia Electronica*, *27*(1), 1-13.

Briggs D., Bruton D., & Whittington H. (1979). Appendages of the arthropod. *Aglaspis spinifer*.

Briggs D. E., Siveter D. J., Siveter D. J., Sutton M. D., Legg D., & Lamsdell J. C. (2023). A vicissicaudatan arthropod from the Silurian Herefordshire Lagerstätte, UK. *Royal Society Open Science*, *10*(8), 230661.

Broili F. (1932). Eine neue Crustacee aus dem rheinischen Unterdevon. *Sitzungsberichte der Mathematisch-Naturwissenschaftlichen (Abteilung) Klasse der Bayerischen Akademie der Wissenschaften zu Munchen*, 27–38.

Burke C. D., Full W. E., & Gernant R. E. (1987). Recognition of fossil fresh water ostracodes: Fourier shape analysis. *Lethaia*, *20*(4), 307-314.

Caster K. E., & Macke W. B. (1952). An aglaspid merostome from the Upper Ordovician of Ohio. *Journal of Paleontology*, *26*(5), 753-757.

Chlupáč I. (1963). Report on the merostomes from the Ordovician of Central Bohemia. *Věstník Ústředního ústavu geologického*, *38*(6), 399-403.

Chlupáč I. (1965). Xiphosuran merostomes from the Bohemian Ordovician. *Sborník geologických věd, Paleontologie*, *5*, 7-38.

Chlupáč I. (1999a). Some problematical arthropods from the Upper Ordovician Letná formation of Bohemia. *Journal of Geosciences*, *44*(1-2), 79-92.

Chlupáč I. (1999b). Unusual arthropods from the Bohemian Ordovician-a review. *ACTA-UNIVERSITATIS CAROLINAE GEOLOGICA*(1/2), 393-396.

Chlupác I., & Kukal Z. (1988). Possible global events and the stratigraphy of the Palaeozoic of the Barrandian (Cambrian-Middle Devonian, Czechoslovakia). *Sborník geologických věd. Geologie*(43), 83-146.

Collantes L., & Pates S. (in press). Isoxys carbonelli and the palaeoenvironmental disparity of Isoxys during Cambrian Stage 3. *Historical Biology*.

Drage H. B., Laibl L., & Budil P. (2018). Postembryonic development of Dalmanitina, and the evolution of facial suture fusion in Phacopina. *Paleobiology*, *44*(4), 638-659.

Drage H. B., & Pates S. (2024). Distinct causes underlie double-peaked trilobite morphological disparity in cephalic shape. *Communications Biology*, *7*(1), 1490.

Du K., Bruton D. L., Yang J., & Zhang X. (2023). An early Cambrian Sidneyia (Arthropoda) resolves the century-long debate of its head organization. *Science China Earth Sciences*, *66*(3), 521-527.

Dunlop J. A., Anderson L. I., & Braddy S. J. (2003). A redescription of Chasmataspis laurencii Caster & Brooks, 1956 (Chelicerata: Chasmataspidida) from the Middle Ordovician of Tennessee, USA, with remarks on chasmataspid phylogeny. *Earth and Environmental Science Transactions of the Royal Society of Edinburgh*, *94*(3), 207-225.

Edgecombe G. D., Paterson J. R., & Garcia-Bellido D. C. (2017). A new aglaspidid-like euarthropod from the lower Cambrian Emu Bay Shale of South Australia. *Geological Magazine*, *154*(1), 87-95.

Eldredge N., & Smith L. (1974). Revision of the suborder Synziphosurina (Chelicerata, Merostomata): with remarks on merostome phylogeny. *American Museum novitates, no. 2543*, 1– 41.

Foote M. (1989). Perimeter-based Fourier analysis: a new morphometric method applied to the
trilobite cranidium. *Journal of Paleontology*, *63*(6), 880-885.
Fortey R. A., & Rushton A. W. (2003). A new aglaspidid arthropod from the Lower Ordovician of
Wales. *Palaeontology*, *46*(5), 1031-1038.
Gevirtz J. L. (1976). Fourier analysis of bivalve outlines: implications on evolution and autecology.
Journal of the International Association for Mathematical Geology, *8*, 151-163.
Hall J. (1862). *On a new crustacean from the Potsdam Sandstone*.
Havlíček V. (1998). Paleozoic of the Barrandian (Cambrian to Devonian). 41–79.
Healy-Williams N., & Williams D. F. (1981). Fourier analysis of test shape of planktonic
foraminifera. *Nature*, *289*(5797), 485-487.
Hesselbo S. P. (1992). Aglaspidida (Arthropoda) from the upper Cambrian of Wisconsin. *Journal of
Paleontology*, 885-923.
Hou X.-G. (1997). Arthropods of the lower Cambrian Chengjiang fauna, southwest China. *Fossils
and Strata*, *45*, 1-116.
Kraft P., Linnemann U., Mergl M., Bruthansová J., Laibl L., & Geyer G. (2023). Ordovician of the
Bohemian Massif.
Lamsdell J. C., Hoşgör İ., & Selden P. A. (2013). A new Ordovician eurypterid (Arthropoda:
Chelicerata) from southeast Turkey: evidence for a cryptic Ordovician record of Eurypterida.
Gondwana Research, *23*(1), 354-366.
Lamsdell J. C., Isotalo P. A., Rudkin D. M., & Martin M. J. (2022). A new species of the Ordovician
horseshoe crab Lunataspis. *Geological Magazine*, 1-5.
Lamsdell J. C., & Ocon S. B. (2025). Segmentation in early Xiphosura and the evolution of the
thoracetron. *Journal of Paleontology*, 1-20.
Legg D. A., Sutton M. D., & Edgecombe G. D. (2013). Arthropod fossil data increase congruence of
morphological and molecular phylogenies. *Nature communications*, *4*(1), 1-7.
Lerosey-Aubril R., Paterson J. R., Gibb S., & Chatterton B. D. (2017). Exceptionally-preserved late
Cambrian fossils from the McKay Group (British Columbia, Canada) and the evolution of
tagmosis in aglaspidid arthropods. *Gondwana Research*, *42*, 264-279.
Lerosey-Aubril R., Zhu X., & Ortega-Hernández J. (2017). The Vicissicaudata revisited–insights
from a new aglaspidid arthropod with caudal appendages from the Furongian of China.
Scientific Reports, *7*(1), 1-18.
Lin W., Pates S., Losso S. R., & Fu D. (2024). Intraspecific variation of early Cambrian (stage 3)
arthropod Retifacies abnormalis revealed by morphometric analyses. *Frontiers in Ecology
and Evolution*, *12*, 1336365.
Ma J., Pates S., Wu Y., Lin W., Liu C., Wu Y., Zhang M., & Fu D. (2023). Ontogeny and brooding
strategy of the early Cambrian arthropod Isoxys minor from the Qingjiang biota. *Frontiers in
Ecology and Evolution*, *11*, 1174564.
Mayers B., Aria C., & Caron J. B. (2019). Three new naraoiid species from the Burgess Shale, with
a morphometric and phylogenetic reinvestigation of Naraoiidae. *Palaeontology*, *62*(1), 19-
50.
McCoy V. E., Herrera F., Wittry J., Mayer P., & Lamsdell J. C. (2025). A possible vicissicaudatan
arthropod from the Late Carboniferous Mazon Creek Lagerstätte. *Geological Magazine*,
162, e3.
Novák O. (1887). Merostomata. *Archive of the National Museum, Unpublished manuscript, Prague*.
O'Flynn R. J., Williams M., Liu Y., Hou X., Guo J., & Edgecombe G. D. (2024). The early Cambrian
Kuamaia lata, an artiopodan euarthropod with a raptorial frontal appendage. *Journal of
Paleontology*, *98*(5), 808-820.
Ortega-Hernandez J., Braddy S. J., & Rak Š. (2010). Trilobite and xiphosuran affinities for putative
aglaspidid arthropods Caryon and Drabovaspis, Upper Ordovician, Czech Republic.
Lethaia, *43*(3), 427-431.
Ortega-Hernández J., Legg D. A., & Braddy S. J. (2013). The phylogeny of aglaspidid arthropods
and the internal relationships within Artiopoda. *Cladistics*, *29*(1), 15-45.
Pates S., Daley A. C., Legg D. A., & Rahman I. A. (2021). Vertically migrating Isoxys and the early
Cambrian biological pump. *Proceedings of the Royal Society B*, *288*(1953), 20210464.

Pérez-Peris, F., Laibl, L., Lustri, L., Gueriau, P., Antcliffe, J. B., Enright, O. G. B., & Daley, A. C.
(2021). A new nektaspid euarthropod from the Lower Ordovician strata of Morocco.
Geological Magazine, 158(3), 509-517.
Plotnick R. E., & Wagner P. J. (2006). Round up the usual suspects: common genera in the fossil
record and the nature of wastebasket taxa. *Paleobiology*, *32*(1), 126-146.
Raasch G. O. (1939). Cambrian merostomata. *Special Papers. Geological Society of America*, *19*,
1-146.
Rak S. (2009). The occurrence of non– trilobite arthropods from the quartzites of Letná Formation
from the vicinity of Beroun. *Geologie, paleonto– logie, speleologie–Ceský kras*, *35*, 14-25.
Rak Š., Bergström J., Fatka O., & Budil P. (2009). The Upper Ordovician arthropod Zonozoe
drabowiensis Barrande (Libeň and Letná formations, Sandbian, Barrandian area, Czech
Republic). *Bulletin of Geosciences*, *84*, 185-188.
RCoreTeam. (2024). R: A Language and Environment for Statistical Computing. R Foundation for
Statistical Computing. In. Vienna, Austria.
Schmidt M., Schoenemann B., Hou X., Melzer R. R., & Liu Y. (2025). Pygmaclypeatus daziensis, a
unique lower Cambrian arthropod with two different compound eye systems.
Communications Biology, *8*(1), 317.
Scotese C. R. (2023). Ordovician plate tectonic and palaeogeographical maps.
Stein M., Budd G. E., Peel J. S., & Harper D. A. (2013). Arthroaspis n. gen., a common element of
the Sirius Passet Lagerstätte (Cambrian, North Greenland), sheds light on trilobite ancestry.
BMC evolutionary Biology, *13*(1), 99.
Stein M., & Selden P. A. (2012). A restudy of the Burgess Shale (Cambrian) arthropod *Emeraldella*
brocki and reassessment of its affinities. *Journal of Systematic Palaeontology*, *10*(2), 361-
383.
Størmer L. (1944). On the relationships and phylogeny of fossil and recent Arachnomorpha: a
comparative study on Arachnida, Xiphosura, Eurypterida, Tribolita, and other fossil
Arthropoda. *Skrifter Utgitt av Det Norske Videnskaps-Academi I Oslo. IMatematisk-*
Naturvidenskapelig Klasse, *5*, 1–158.
Størmer L. (1951). A new eurypterid from the Ordovician of Montgomeryshire, Wales. *Geological*
Magazine, *88*(6), 409-422.
Størmer L. (1955). Merostomata in RC Moore, ed., Treatise on invertebrate paleontology, Pt. P,
Arthropoda 2: Geol. Soc. America and Univ. In: Kansas Press.
Stott C. A., Tetlie O. E., Braddy S. J., Nowlan G. S., Glasser P. M., & Devereux M. G. (2005). A new
eurypterid (chelicerata) from the Upper Ordovician of Manitoulin Island, Ontario, Canada.
Journal of Paleontology, *79*(6), 1166-1174.
Van Roy P. (2005). An aglaspidid arthropod from the Upper Ordovician of Morocco with remarks on
the affinities and limitations of Aglaspidida. *Earth and Environmental Science Transactions*
of the Royal Society of Edinburgh, *96*(4), 327-350.
Van Roy P., Orr P. J., Botting J. P., Muir L. A., Vinther J., Lefebvre B., El Hariri K., & Briggs D. E.
(2010). Ordovician faunas of Burgess Shale type. *Nature*, *465*(7295), 215-218.
Van Roy P., Rak Š., Budil P., & Fatka O. (2022). Redescription of the cheloniellid euarthropod
Triopus drabowiensis from the Upper Ordovician of Bohemia, with comments on the
affinities of Parioscorpio venator. *Geological Magazine*, *159*(9), 1471-1489.
Vokáč V., & Grigar L. (2024). New finding of the fossil arthropod Zonozoe drabowiensis Barrande,
1872 in the Řevnice Quartzite (Libeň Formation, Upper Ordovician, Sandbian) bear Starý
Plzenec (Prague Basin, Czech Republic). *Erica*, *31*, 83-87.
Walcott C. D. (1911). Cambrian geology and paleontology II. Middle Cambrian Merostomata.
Smithsonian Miscellaneous Collections, *57*(2), 17–40.
Zhang X., & Shu D. (2005). A new arthropod from the Chengjiang Lagerstätte, Early Cambrian,
southern China. *Alcheringa*, *29*(2), 185-194.

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