Sexual dimorphism in pollen foraging and sensory traits in *Heliconius* butterflies

José Borrero[1], David Rivas-Sánchez[2], Daniel Shane Wright[1], Caroline Nicole Bacquet[3], Stephen H. Montgomery[2], Alexander Keller[4] & Richard M. Merrill[1]

Corresponding authors: jose.borrero@lmu.de (J.B.) & merrill@bio.lmu.de (R.M.M)

[1] *Division of Evolutionary Biology, Faculty of Biology, LMU Munich, Germany*

[2] *School of Biological Science, University of Bristol, Bristol, United Kingdom*

[3] *Universidad Regional Amazónica de Ikiam, Tena, Ecuador*

[4] *Cellular and Organismic Networks, Faculty of Biology, LMU Munich, Germany*

Abstract

Sexual dimorphism in foraging behaviour is widespread in insects and may arise from differences in nutritional demands, sensory systems, or cognition. In *Heliconius*, pollen-feeding is an evolutionary innovation among butterflies that supports extended lifespans and sustained reproduction. However, how foraging behaviour varies between the sexes and how it relates to sexually dimorphic traits remains poorly understood. We investigated sex-specific foraging strategies in wild *Heliconius himera*, a highland specialist from southern Ecuador, using field surveys and DNA metabarcoding. Females carried more pollen than males, consistent with higher nutritional demands. Yet both sexes used similar plant richness and composition, indicating that differences reflect foraging effort rather than shifts in plant choice. Gut samples revealed greater pollen diversity and a more consistent community profile than proboscis samples, suggesting they better capture cumulative foraging history. To place these behavioural differences in a sensory context, we quantified sexually dimorphic sensory traits. Males had larger eyes and more ommatidia, whereas females had larger mushroom bodies, brain regions that support associative learning and memory. These patterns are consistent with sexual dimorphism reported across *Heliconius* and may contribute to the observed sex differences in

26   pollen foraging. Our findings highlight how sex-specific foraging differences may arise from

27   effort on shared floral resources and co-occur with divergent sensory and neural investment,

28   offering insights into the ecological basis of intraspecific variation in pollen use.

Introduction

Sexual dimorphism is widespread across animals and often involves divergence in morphology, physiology, and behaviour. Among insects, sex differences in foraging behaviour may arise from distinct dietary needs or sensory biases. Many species exhibit sex-specific specializations in visual and olfactory systems that support different ecological roles, such as mate location in males and resource acquisition in females (Smith et al., 2019). For instance, in the oil bee *Macropis fulvipes*, both sexes forage on the same flowers, but males rely more on visual cues and only collect pollen, whereas while females show a stronger preference for olfactory cues and collect both pollen and floral oils (Dötterl et al., 2011). Similarly, in the fly *Bibio rufiventris*, males and females differ in their responses to floral volatiles over the course of the day, reflecting sex-specific attraction to scent cues (Zhu et al., 2025). Despite such examples, the ecological and sensory drivers of sex-specific foraging behaviour remain poorly understood in most insect taxa.

Pollen-feeding is a widespread and ecologically important behaviour in insects, particularly among bees, flies, and beetles, where it plays a central role in adult nutrition, reproduction, and pollination (Romeis et al., 2005). These lineages have evolved diverse morphological and behavioural adaptations to collect and digest nutrient-rich pollen (Roulston & Cane, 2000; Ruedenauer et al., 2019). Unlike most butterflies that feed only on nectar, *Heliconius* uniquely feed on pollen, a strategy central to their nutrition, reproduction, and mutualism with the plants they pollinate (Gilbert, 1972; Young & Montgomery, 2020). Sex differences in pollen feeding have been documented in some *Heliconius* species, but how these differences align with sexually dimorphic sensory traits, and whether they involve shifts in pollen load versus composition is understudied.

Insects that feed on pollen gain access to amino acids and lipids otherwise scarce in nectar

(Romeis et al., 2005). In *Heliconius*, pollen feeding has been linked to enlarged mushroom

bodies that support sensory integration and learning, possibly facilitating this unique behaviour

(Sivinski, 1989; Montgomery et al., 2016). Both sexes benefit from pollen-derived nutrients that

extend lifespan and sustain reproduction (Dunlap-Pianka et al., 1977; Pinheiro de Castro et al.,

2020). But the direct benefits differ: in females, amino acids and lipids directly support

oogenesis and sustained egg production, and in older females, pollen-feeding helps maintain

adult cyanogenic defences that contribute to Müllerian mimicry (O'Brien et al., 2003, 2004;

Pinheiro de Castro et al., 2025). whereas in males direct benefits appear more limited, where it

mainly supports spermatophore production and mating effort (Boggs et al., 1981; Boggs &

Gilbert, 1979). These differences in direct benefits likely drives a female bias in pollen foraging.

Multiple studies have found evidence for sex differences in pollen feeding in *Heliconius.* Field

observations of *H. charithonia*, a species within the *erato/sara/sapho* clade, have revealed sex-

specific pollen use: females are more often recorded on *Hamelia patens* plants, while males

favor *Lantana camara* (Mendoza-Cuenca & Macías-Ordóñez, 2005). Similarly, broader studies

across *Heliconius* have documented larger pollen loads and longer foraging duration in females

than males (Boggs et al., 1981; Cardoso, 2001; Gilbert, 1991). These patterns mirror broader

trends in other Lepidoptera, where females actively seek out nectar with higher amino acid

content than males (Alm et al., 1990; Erhardt & Rusterholz, 1998; Mevi-Schütz & Erhardt,

2005).

In *Heliconius*, differences in foraging may be influenced by sexually dimorphic visual systems.

Unlike most butterflies, *Heliconius* possess two UV-sensitive opsins, and in the erato/sara/sapho

clade one of these (UVRh1) is linked to the sex chromosome (W) and expressed only in

females, enabling UV wavelength discrimination in females, which may enhance floral detection

capabilities (Chakraborty et al., 2023; Finkbeiner & Briscoe, 2021). Its ecological relevance

remains unclear; for instance, UV sensitivity does not appear to affect oviposition in *H.*

erato and *H. himera* (Borrero et al., 2023). Beyond opsin expression, other visual traits such as

facet number, facet diameter, and corneal area shape acuity and sensitivity and can differ

between sexes (Land, 1997; Wright et al., 2023). As such, integrated studies of foraging

behaviours and sensory traits may help elucidate the basis of sex-specific pollen foraging.

Heliconius himera is a highland specialist inhabiting seasonally dry forests of southern Ecuador

(~1500 m asl). Compared with its lowland relatives, it occupies more open habitats with higher

light intensity, stronger temperature fluctuations, and distinct floral communities, conditions that

likely shape both resource availability and foraging strategies (Descimon & Mast, 1984; Jiggins

et al., 1996; Dell'Aglio, Mena, et al., 2022). Reflecting adaptation to this environment, *H. himera*

differs from lowland *H. erato* in brain morphology and sensory processing, investing more in

antennal lobes and less in visual neuropils (Montgomery & Merrill, 2017). When visual and

olfactory cues conflict, *H. himera* relies more heavily on olfactory input than *H. erato* (Dell'Aglio,

McMillan, et al., 2022), a pattern also seen in other Lepidoptera with altered neural investment

(Stöckl et al., 2016; Borrero et al., 2024). As an incipient species within the *erato/sara/sapho*

clade, it likely shares the sexually dimorphic opsin expression and vision reported in related taxa

(McCulloch et al., 2017; Chakraborty et al., 2023). This combination of distinct sensory

weighting and potential sexually dimorphic vision makes *H. himera* a compelling model for

exploring how environmental adaptation and sex-specific sensory traits shape foraging

behaviour.

Despite its ecological importance, pollen use in *Heliconius* remains incompletely understood.

Traditional methods such as field observations and palynological keys have revealed diverse

diets dominated by *Psychotria*, *Psiguria*, and *Lantana* (Boggs et al., 1981; Cardoso, 2001;

Estrada & Jiggins, 2002; Mendoza-Cuenca & Macías-Ordóñez, 2005). However, these methods

are slow, expertise-dependent, and limited in taxonomic resolution. DNA metabarcoding

provides a powerful alternative: targeting regions such as ITS2, enables high-throughput,

species-level identification of mixed pollen samples (Bell et al., 2016; Keller et al., 2015).

Already widely applied in bee pollination studies (Martins et al., 2023; Piko et al., 2021; Wilson

et al., 2021), this method can open new perspectives on *Heliconius* foraging ecology and sex-

specific pollen use.

In this study, we integrate field surveys, DNA metabarcoding, and morphological analyses to

investigate sex differences in foraging behaviour in *H. himera*. Specifically, we tested the

hypothesis that sexes differ in the amount and diversity of pollen collected, reflecting distinct

nutritional demands, foraging strategies, or sensory capabilities. We further quantified eye and

brain morphology to place behavioural differences in a sensory context. This study provides one

of the first molecular characterizations of pollen resource use in *Heliconius* and advances our

understanding of how sexual dimorphism in sensory traits shapes intraspecific variation in

foraging, with implications for pollinator ecology more broadly.

Materials & Methods:

Study sites:

Field surveys were conducted at five sites in Loja Province, Ecuador, around the city of

Vilcabamba (4°15′S, 79°13′W), at elevations of 1500–1700 m (Fig. 1A–B). All sites are

characterized by thorn scrub dry forest dominated by Acacia macracantha and xerophytic

vegetation (Jiggins et al., 1996). Rumi Wilco is a 40-ha private reserve containing ~700 plant

species (Jorgensen & Leon-Yanez, 1999), while Cerro Mandango is a nearby mountain site

dominated by Acacia. The other sites (Quebrada Puliche, Agua de Hierro, and San Pedro) occur

*Figure 1: **Sampling region and pollen foraging behavior in H. himera.** (**A**) Study area in southern Ecuador. (**B**) Overview map showing the location of butterfly survey sites in Loja Province. Points represent individual collection records, with colour indicating sex (green = female, orange = male). (**C**) Satellite map of survey sites, including Rumi Wilco, Cerro Mandango, Quebrada Puliche, and San Pedro. (**D**) Close-up photograph of a H. himera with a visibly pollen-loaded proboscis.*

along small streams with similar vegetation, adjacent to coffee plantations and pastures.

Butterfly sampling:

From April to September 2023, we sampled *H. himera* along trails at all five sites, with most

effort concentrated at Rumi Wilco and Cerro Mandango, which were visited at least five days

per week. Daily sampling took place between 08:00 and 17:00, coinciding with peak activity of

H. himera (Davison et al., 1999), spanning the wet-to-dry season transition.

Each butterfly was captured, marked on the forewing, and released after recording GPS

location, time, sex, body length, pollen load and observer. Pollen load was visually classified as

none, small, or large depending on the proportion of the proboscis covered (Fig. 1D).

Recaptured individuals were noted for movement analyses. In total, 877 butterflies were

recorded (371 females, 506 males).

To test for sex differences in microhabitat, we measured temperature and humidity at capture

sites using HOBO Temp/RH U23-002 data loggers that recorded continuously for at least 24 h

post-capture. Environmental data were obtained for 49 individuals (24 females, 25 males) over

2–3 sampling days per month throughout the study period.

Pollen collection and ITS2 sequencing:

To identify pollen sources, we collected 71 *H. himera* (25 females, 46 males) with hand nets

between July and August 2023. Specimens were placed in entomological envelopes;

proboscises were dissected and stored in PCR tubes, while bodies (containing guts) were

preserved in DMSO/EDTA/NaCl solution. Samples were kept at –15 °C until DNA extraction and

sequencing in February 2024. All proboscises and a subset of guts (13 females, 26 males) were

analysed using ITS2 DNA metabarcoding following the protocol of Sickel *et al.*, (2015), with

positive and negative controls included.

Sequence   reads   were   processed   with   a   custom   bioinformatics   pipeline

(https://github.com/chiras/metabarcoding_pipeline), generating amplicon sequence variants

(ASVs) and assigning taxonomy against curated ITS2 plant databases (Keller et al., 2020;

Quaresma et al., 2024). Remaining unclassified reads were further annotated with the SINTAX

algorithm (Edgar, 2016). Data were analysed in R with the *phyloseq* package (McMurdie &

Holmes, 2012). Analyses were restricted to Streptophyta, and taxa were retained if they reached

≥1% relative read abundance in at least one sample.

Wing reflectance spectrometry and age estimation:

To estimate the relative age of wild individuals, we adapted a spectrometric method developed by (Dell'Aglio et al., 2017), who used wing colour metrics to infer butterfly age. We collected full-spectrum reflectance data (300–750 nm) from red and yellow wing bands of insectary-reared individuals of known age across seven *Heliconius* species (n = 320), including 108 reared *H. himera*. Spectra were measured using an Ocean Optics Flame spectrometer with a PX-2 xenon light source and WS-1 white standard. Reflectance was analysed in R with the pavo package (Maia et al., 2019), and the area under the reflectance curve (AUC) was calculated for each colour patch.

Age was predicted using linear models with log-transformed AUC values as predictors. A combined model including both red and yellow AUCs provided the best fit (adjusted R^2 = 0.13, P < 0.001) and was used to estimate the age of wild *H. himera*. Predicted values were backtransformed to days, and each individual was assigned a relative age score. This continuous relative age metric provided a standardized measure across individuals and was used to test how pollen foraging behaviour varies with age in the wild population.

Eye cuticle morphology:

To test for sexual dimorphism in eye cuticle morphology, we analysed wild-caught and insectary-reared H. himera following established protocols (Wright et al., 2024; Wright & Merrill, 2025b, 2025a). Eyes were dissected, cleaned, mounted on slides, and imaged at 25× magnification. Corneal surface area and facet number were measured using image thresholding and the Analyze Particles function in ImageJ/Fiji (Schindelin et al., 2012). Inter-ocular distance and hind tibia length were also measured to account for body size. Given the strong bilateral symmetry of these traits (Wright et al., 2024), analyses were performed on the left eye unless damaged, in which case the right eye was used.

Brain immunohistochemistry and Imaging:

We examined sexual dimorphism in brain morphology in 43 *H. himera* (21 females, 22 males), combining previously analysed wild individuals from Vilcabamba (Ecuador) and common-garden reared butterflies from Cambridge (UK) (Montgomery & Merrill, 2017) with newly processed specimens reared in Tena (Ecuador; 8 females, 9 males). Brains were labelled using anti-synapsin immunofluorescence (Montgomery et al., 2016; Ott, 2008) imaged with confocal microscopy, and neuropils segmented in Amira. We measured volumes of key optic and central brain regions previously shown to be ecologically and evolutionarily relevant in *Heliconius* (Montgomery et al., 2021; Montgomery & Merrill, 2017), Including the medulla, lamina, lobula, lobula plate, accessory medulla, ventral lobula, antennal lobe, anterior optic tubercle, and mushroom body calyx and lobes plus pedunculus. The rest-of-central-brain (rCB) volume was used as an allometric control. Paired structures were quantified in one hemisphere and doubled, and volumetric data were z-axis corrected using a scaling factor of 1.52 (Montgomery & Ott, 2015) and log10-transformed before analysis.

Statistical analysis:

Pollen presence was modelled with binomial GLMMs including sex and time of day as fixed effects and random intercepts for individual and date. Pollen load categories were analysed with cumulative link models with sex and time of day as fixed effects. Activity patterns were analysed with multinomial regression, modelling sex as a function of hour of day. Body size was tested with linear mixed models including sex as a fixed effect and observer as a random intercept. Movement distances of recaptured individuals were calculated with Vincenty ellipsoid methods and analysed with log-transformed linear models including sex and body length as fixed effects. Microhabitat associations (Temperature (°C) and relative humidity (%)) with sex were examined with binomial GLMMs including temperature and relative humidity as continuous predictors and date as a random intercept.

To test effects of sex, body part (proboscis vs. gut), and predicted age on pollen composition,

we used PERMANOVA on Bray–Curtis dissimilarities. Pollen richness (observed taxa) and

diversity (Shannon index) were modelled with GLMMs, including date as a random effect. Age

was modelled as a continuous predictor; results were compared with likelihood ratio tests and

visualized with estimated marginal means.

Sexual dimorphism in eye and brain morphology was tested using linear models and

standardized major axis (SMA) regression. For eyes, log10-transformed corneal area and facet

number were modelled with sex, $\log_{10}$-transformed hind tibia length, and interocular width as

fixed effects. For neuropils, $\log_{10}$- transformed volumes were modelled with sex, rearing

condition (wild vs. reared, with reared individuals from Tena or Cambridge), and log10 rest-of-

central-brain volume (rCB) as fixed effects. Model residuals were checked for normality, and

fixed effects were evaluated with Type II ANOVA. Likelihood ratio tests assessed the

contribution of predictors, and post hoc contrasts were performed with *emmeans* using

Bonferroni correction. SMA regression (smatr package (Warton et al., 2012)) was used to test

sex-specific differences in slopes, intercepts (elevation shifts), and major axis shifts. Mushroom

body calyx and lobes plus pedunculus were analysed separately, consistent with

previous *Heliconius* studies (Montgomery et al., 2016; Montgomery & Merrill, 2017).

Results:

Sex Differences in Pollen Presence and Load

Our field collections resulted in records from 371 females and 506 males, which we used to test whether the probability of carrying pollen differed between the sexes. Overall, females were more likely to both carry pollen and tend to carry greater quantities compared to males. Specifically, inclusion of sex significantly improved model fit in our binomial GLMM ($\chi^2 = 16.59$, $P < 0.001$). Fitting a cumulative link model analysing pollen load categories, "no pollen", "little pollen" and "lot pollen" (Figure 2A), also revealed that females were significantly more likely to carry both small (estimate = 0.09, z = 4.31, P < 0.001) or large pollen loads (estimate = 0.06, z = 4.00, P = 0.0001) than males.

Figure 2: Pollen loads and temporal activity patterns in H. himera. (A) *Estimated marginal means of predicted probabilities for each pollen load category by sex, derived from a cumulative link mixed model. Letters denote significant pairwise differences between sexes within each category (α = 0.05). (**B**) Proportional distribution of captured males (orange) and females (green) across the day, binned by hour of capture (**C**) H. himera actively foraging on Lantana flowers.*

Pollen richness and diversity differ by body part but not by sex

To investigate the diversity of pollen resources used by *H. himera* and assess potential differences between sexes, sample types, and individual age, we performed DNA metabarcoding on pollen collected from 25 females and 46 males proboscis samples, as well as gut samples from a subset of these individuals (13 females and 26 males). Metabarcoding yielded 1,309,788 high-quality, taxonomically classified reads (mean = 11,907 per sample), detecting 54 families, 122 genera, and 163 species. All samples passed quality filtering and were retained for downstream analysis. The most common taxa included *Lantana camara* (Verbenaceae), *Stellaria media* (Caryophyllaceae), and several Asteraceae species: *Chromolaena odorata*, *Kaunia rufescens*, and *Fleischmannia pycnocephala* (Fig. 3A; Fig S3-S4).

Non-metric multidimensional scaling (NMDS) ordination revealed substantial overlap in pollen composition, with no significant differences between sexes (PERMANOVA: R^2 = 0.0096, F = 1.05, P = 0.378). In contrast, pollen composition differed significantly between body parts (R^2 = 0.0325, F = 3.63, P = 0.001; Fig. 3B), suggesting some differentiation in the pollen sources detected externally versus internally, possibly reflecting variation in sampling windows or digestion. We found no evidence that predicted age explained variation in pollen composition (R^2 = 0.014, F = 1.03, P = 0.415; Fig. S2).

Generalized linear and linear mixed models confirmed that observed plant species richness and Shannon diversity differed significantly by body part, with gut samples containing greater richness (χ^2 = 29.80, df = 1, P < 0.001) and diversity (χ^2 = 15.56, df = 1, P < 0.001) than proboscis samples (Fig. 3C). In contrast, sex had no effect on either metric (richness: χ^2 = 1.00, P = 0.32; Shannon: χ^2 = 0.002, P = 0.97). Predicted age did not influence richness or diversity when modelled as a continuous variable (richness: χ^2 = 0.19, P = 0.66; Shannon: χ^2 = 1.78, P = 0.18).

Figure 3: Diversity and composition of pollen collected from different sexes and tissues. (*A*) *Relative read abundance of the most common plant taxa identified in proboscis and gut samples. Bars represent the mean relative abundance across individuals, with colors indicating plant families. (**B**) NDMS plot across samples, showing clustering by body part (proboscis = purple, gut = green) and sex (female = circles, male = triangles). 95% confidence ellipses are shown for each group. (**C**) Observed richness and Shannon diversity of pollen samples collected from proboscis and gut tissues of H. himera, separated by sex.*

261 **Daily activity, movement, habitat use and age**

262 Given the sex differences in pollen presence and load size, we next tested for variation in body

263 size, daily activity, movement, and microhabitat use (Fig. S1). Females averaged 24.4 mm in

264 length (SD = 1.66, IQR = 23–26), while males averaged 25.3 mm (SD = 1.55, IQR = 24–26).

265 Linear models confirmed males were larger (χ^2 = 129.73, P < 0.001; Fig. S1)

266 Daily activity also differed (LR = 19.11, P < 0.001), with females captured earlier in the day (Fig.

267 2B). To test whether time of day independently influenced pollen presence, we included capture

268 time as a fixed effect in the binomial GLMM. After accounting for sex (χ^2 = 16.59, P < 0.001),

269 capture time did not improve the model (χ^2 = 0.46, P = 0.50).

270 Movement was analysed from 109 recaptured females and 162 males. Recaptured distances

271 varied widely but were comparable: females moved a mean of 87.2 m (median = 24.8, SD =

272 176.0) and males 66.9 m (median = 36.2, SD = 95.1). A linear model of log-transformed

273 distances showed no effect of sex on movement distance (χ^2 = 0.63, P = 0.61; Fig. S1).

274 Both sexes were recaptured under similar microclimatic conditions. Females were recaptured at

275 a mean temperature of 25.8 °C (median = 25.9, SD = 3.5) and 50.9% relative humidity (median

276 = 48.0, SD = 16.8), while males were recaptured at 26.4 °C (median = 26.2, SD = 3.4) and

277 49.5% relative humidity (median = 48.3, SD = 15.0). These distributions overlapped strongly,

278 with interquartile ranges of 23.3–27.9 °C for females and 24.1–29.3 °C for males. Binomial

279 GLMMs confirmed that neither temperature (χ^2 = 1.05, df = 1, P = 0.31) nor humidity (χ^2 = 0.11,

280 P = 0.75) significantly affected the probability of capturing either sex (Fig. S1).

281 **Sexual dimorphism in eye and brain morphology**

282 We tested for sexual dimorphism in eye and brain morphology (Fig. 4). Males had larger eyes

283 than females ($F_{1,39}$ = 26.25, P < 0.001). SMATR analysis showed no sex difference in slope

284 between corneal area and tibia length but a significant elevation shift (Wald χ^2 = 68.82, P <

0.001), with males consistently having larger eyes for a given tibia length (Fig. 4A). Facet

number was also higher in males ($F_{1,40}$ = 6.07, P = 0.018). Using corneal area as the allometric

control, SMATR again detected no sex difference in slope or elevation but a significant major

axis shift (Wald χ^2 = 21.23, P < 0.001), showing that males had larger eyes and thus more

facets (Fig. 4D).

We also found sex differences in brain morphology, particularly in higher-order integrative

regions (Fig. 4B–F). Linear models revealed that females had significantly larger mushroom

body calyces (MBCA; χ^2_1 = 36.0, P < 0.001), after accounting for overall brain size and rearing

condition (χ^2_2 = 2.57, P =0.28). SMATR analysis confirmed a significant elevation shift in the

scaling relationship between MBCA volume and overall brain size (Wald χ^2_1 = 34.25, P < 0.001),

indicating that females consistently had larger MBCA for a given brain size (Fig. 4C). A similar

pattern, with larger female investment. was observed for the mushroom body lobes and

pedunculus (Fig. 4F). Linear models showed significant effects of sex (χ^2_1 = 29.5, P < 0.001),

after accounting for overall brain size and rearing condition (χ^2_2 = 49.8, P < 0.0001). SMATR

analysis detected a significant elevation shift in the scaling relationship (Wald χ^2_1 = 8.17, P

=0.01).

By contrast, optic lobes showed no sex differences (χ^2_1 = 0.91, P = 0.34; Wald χ^2_1 = 1.15, P =

0.28; Fig. 4B), nor did analyses of individual visual neuropils (Supplementary Table 1). Antennal

lobes also showed no significant differences (χ^2_1 = 3.6, P = 0.06; Wald χ^2_1 = 0.23, P = 0.63; Fig.

4E). Full model results are provided in Supplementary Table 1.

*Figure 4: **Sexual dimorphism in eye and brain morphology.** Males = orange; females = green. All traits are log$_{10}$-transformed. (**A**) Corneal area shows a significant elevation shift, with males having larger eyes. (**B, E**) Optic lobe (OL) and antennal lobe (AL) volumes scale with rest-of-central-brain volume (rCB) but show no sex differences. (**C, F**) Mushroom body calyx (MBCA) and mushroom body lobes and pedunculus (MBLOBES+PED) exhibit a significant elevation shift, with females having larger volumes than males. (**D**) Facet count shows a significant major axis shift, with males having more facets. Lines indicate sex-specific SMATR regressions (solid) or common slopes (dashed). Insets show the corresponding anatomical structures: eye (gray), OL (yellow/orange), MB (red), AL (blue).*

Discussion

Systematic field sampling combined with DNA metabarcoding revealed high floral diversity and

clear sex differences in pollen foraging in wild *H. himera*. Females carried larger pollen loads

than males, consistent with sex-specific nutritional demands; yet both sexes used similar plant

richness and composition, indicating shared floral resources and differences in effort rather than

choice. Notably, gut samples contained higher pollen diversity and more consistent community

profiles than proboscis samples, suggesting they better capture cumulative foraging history. We

further confirmed sexual dimorphism in eye morphology and mushroom body volume,

consistent with patterns reported across Heliconius (Couto et al., 2023; Wright et al., 2024).

While we did not directly test functional links, these sensory traits may contribute to sex

differences in foraging effort. Overall, our study provides one of the first molecular

characterizations of pollen resource use in Heliconius, showing that sex-specific foraging

reflects effort on shared floral resources and may be shaped by divergent sensory investment.

The floral resources used by *H. himera* differed markedly from those of lowland *Heliconius*.

Metabarcoding revealed a diverse diet of 163 plant species spanning, genera and 54 families.

While *Lantana camara* (*Verbenaceae*) was the most frequent resource, *Asteraceae* were also

well represented, particularly *Chromolaena* and *Kaunia*, which produce amino acid–rich nectar

and are widely visited by butterflies and other pollinators (Layek et al., 2022; Rathnayake &

Wijetunga, 2016; Wist & Davis, 2008). By contrast, *Gurania* and *Psiguria* (*Cucurbitaceae*),

central to the foraging ecology of lowland *Heliconius (Estrada & Jiggins, 2002; Jiggins, 2017)*,

are entirely absent in the range and diet of *H. himera*. Instead, *H. himera* relied heavily on

Lantana, highlighting its role as a core floral resource in montane dry forests. Larval resource

use also reflects this environment: *H. himera* oviposits on *Passiflora* species of the *Decaloba*

subgenus, which tolerate dry conditions and occur in open habitats (Ulmer & MacDougal, 2004).

Thus, both larval and adult foraging are shaped by the montane dry-forest environment: larvae

exploit drought-tolerant host plants, while adults rely on a broad array of floral resources.

Together, these patterns suggest that *erato* clade species, including *H. himera,* have retained a

generalist strategy across life stages, facilitating colonisation of high-altitude habitats.

In our study, both sexes of *H. himera* experienced similar environmental conditions and

movement patterns, indicating shared microhabitat use and strong spatial overlap in access to

floral resources. In patchy, resource-limited montane habitats, such overlap may favour a

generalist foraging strategy over sex-specific specialization. Accordingly, sex-specific nutritional

demands appear to be met through differences in foraging effort rather than consumption of

distinct floral species. In other *Heliconius*, microhabitat differences often underlie floral

preferences rather than intrinsic species identity (Cardoso, 2001; Estrada & Jiggins, 2002). For

example, *H. melpomene* and *H. cydno* rely on *Lantana* and *Psychotria*, respectively, due to

habitat use rather than innate preference (Jiggins, 2017). Likewise, long-term studies

of *Argynnis idalia* show that sex differences in floral visitation diminish when males and females

forage during overlapping phenological windows (Chmielewski et al., 2023). By contrast, *H.*

charithonia exhibits clear sex-specific floral preferences (Mendoza-Cuenca & Macías-Ordóñez,

2005). The absence of strong floral partitioning in *H. himera* therefore suggests that sex-specific

strategies are expressed mainly through pollen collection effort, as an adaptive response to

ecological constraints of montane environments. While *H. himera* belongs to the

erato/sara/sapho clade, where sexual dimorphism in UV opsin expression is common, its opsin

profile remains unknown (Chakraborty et al., 2023; McCulloch et al., 2016, 2017). A reduction or

absence of such dimorphism could further contribute to convergence in floral use between

sexes.

Our comparison of gut and proboscis samples confirmed that tissue type strongly affects pollen

detection and must be considered in dietary inference. Gut samples showed higher richness

and diversity, likely reflecting a longer integration window and cumulative foraging history or

more passive uptake from nectar. Proboscis samples were often dominated by *Lantana*,

suggesting its importance as a core pollen source actively processed through prolonged

proboscis manipulation and digestion (Eberhard et al., 2007; Gilbert, 1972; Harpel et al., 2015).

Meanwhile, floral taxa visited primarily for nectar may not persist externally, indicating that gut

samples capture longer-term foraging, whereas proboscis samples reflect more recent plant

interactions.

Beyond these tissue-specific insights, our results highlight the power of DNA metabarcoding for

ecological research in butterflies. This approach enabled species-level identification of complex

pollen mixtures from small tissue samples, detecting a broad taxonomic spectrum not easily

resolved by classical palynology (Bell et al., 2016). The successful use of DMSO-preserved

samples further demonstrates its potential for archived collections, opening opportunities to

compare historical and spatial variation in pollen use, shifts in foraging behaviour, and

interspecific differences across the *Heliconius* clade. For example, seasonal or local changes in

pollen availability may influence investment in cyanogenic glycosides or egg-laying effort

(Jiggins, 2017; Young & Montgomery, 2020). Leveraging DNA metabarcoding across temporal

and environmental gradients can thus deepen our understanding of the ecological and

evolutionary dynamics structuring pollinator–plant interactions in tropical ecosystems.

Higher pollen loads in females likely reflect reproductive and physiological demands. This

pattern aligns with earlier observations in other *Heliconius* species, where females forage more

actively and carry larger pollen loads than males (Boggs et al., 1981; Cardoso, 2001; Mendoza-

Cuenca & Macías-Ordóñez, 2005). Female *Heliconius* require a steady intake of amino acids

and lipids for oogenesis and to maintain cyanogenic glycosides that serve as chemical defenses

for adults and offspring (Dunlap-Pianka et al., 1977; Gilbert, 1972; O'Brien et al., 2003; Pinheiro

de Castro et al., 2025). These demands cannot be met through nectar alone, making pollen

essential for replenishing protein reserves. Males also benefit from amino acids for

spermatophore production and maintenance (Boggs et al., 1981; Boggs & Gilbert, 1979), but

carried lower pollen loads, possibly reflecting weaker nutritional demands, greater time

investment in mate search, or greater efficiency in resource extraction. Female-biased foraging

extends beyond *Heliconius*: across Lepidoptera, females preferentially use nutrient-rich

resources supplying amino acids, lipids, and sterols required for egg production (Alm et al.,

1990; Chmielewski et al., 2023; Erhardt & Rusterholz, 1998; O'Brien et al., 2004; Mevi-Schütz &

Erhardt, 2005). This sex-specific investment pattern is consistent with optimal foraging theory,

where allocation tracks net fitness benefits (Houston, 1995). More broadly, sex-based

differences in resource use are an important source of intraspecific variation that may influence

pollination networks and community dynamics (Kishi & Kakutani, 2020; Smith et al., 2019,

2021).

We used full-spectrum wing reflectance to estimate the relative age of wild butterflies, adapting

a model trained on known-age individuals. Although wing wear and colouration are widely used

as age proxies in Lepidoptera (Dell'Aglio et al., 2017; Kemp, 2006), they are influenced by

environmental exposure and behaviour, so reflectance-based estimates should be interpreted

cautiously. In our dataset, predicted age did not explain variation in pollen richness or diversity.

This contrasts with earlier studies reporting age-related peaks in pollen load (Boggs et al., 1981)

and suggests that age effects in *Heliconius* may be weak or context dependent.

Our analyses revealed pronounced sexual dimorphism in the visual system, with males

possessing significantly larger eyes and more facets than females. This mirrors patterns across

insects, where males often exhibit enlarged eyes or acute zones that improve spatial resolution

and sensitivity to moving stimuli, aiding in mate detection and tracking (Somanathan et al.,

2017; Wright et al., 2024; Ziemba & Rutowski, 2000). In butterflies, such traits may help males

navigate visually complex habitats, track mates more efficiently, or evade predators while

exposed during mate search. By contrast, female *H. himera* may rely more on chromatic cues

during foraging, especially in species where dimorphic UV opsin expression supports female-

specific flower detection (Chakraborty et al., 2023; Finkbeiner & Briscoe, 2021). Alternatively,

females may accumulate larger pollen loads through greater persistence at flowers; recent

behavioural work shows that *Heliconius* females often revisit flowers repeatedly, leading to

increased pollen accumulation (Dell'Aglio et al., 2024). Both mechanisms are plausible, and

direct behavioural data from *H. himera* will be needed to determine their relative importance.

Among insects, neural investment reflects ecological demands such as foraging, navigation,

and reproduction (Smid & Vet, 2016). In *Heliconius*, mushroom bodies, centres for sensory

integration, learning, and memory, are markedly enlarged compared to other Lepidoptera, a

pattern linked to the evolution of pollen feeding and long-term spatial memory (Sivinski, 1989;

Montgomery et al., 2016; Young & Montgomery, 2020; Couto et al., 2023). In our study, *H.*

himera females had significantly larger mushroom body volumes than males, consistent with the

Heliconius-wide pattern identified by Couto et al. (2023) but absent in outgroup *Heliconiini*. One

possibility is that female-biased mushroom bodies reflect greater foraging demands. However,

their functional basis remains uncertain: laboratory assays of learning and memory

in *Heliconius* have not detected sex effects (Hodge et al., 2025; Toure et al., 2020; Young et al.,

2024). Alternatively, larger mushroom bodies may arise as a by-product of higher reproductive

investment. Females typically carry larger pollen loads and engage in prolonged or repeated

flower visits (Dell'Aglio et al., 2024), and the nutrients obtained may enable greater allocation to

neural tissue. At present, these explanations remain speculative, and there is no direct evidence

that larger mushroom bodies enhance cognitive or foraging performance in females.

Together, our findings show that female *H. himera* carry larger pollen loads than males, yet both

sexes exploit a similar diversity and composition of floral resources, indicating that sex-specific

differences arise from foraging effort rather than plant choice. In patchy montane environments,

such a generalist strategy may facilitate colonisation of high-altitude habitats. Gut samples

proved particularly informative, capturing a broader and more integrated record of pollen use

than proboscis samples and underscoring the importance of tissue choice in dietary inference.

We also documented sexual dimorphism in eye and brain morphology, consistent with patterns

across *Heliconius*, although the functional significance of these traits remains unresolved.

Finally, our use of DNA metabarcoding highlights the value of molecular tools for reconstructing

butterfly diets and linking sex-specific foraging to broader questions of intraspecific variation and pollination ecology.

Acknowledgements

We are grateful to the Ministerio del Ambiente, Agua y Transición Ecológica for permission to collect butterflies in Ecuador (MAATE-DBI-CM-2021-0176; MAATE-DNB-CM-2021-0176). We thank Alicia Cerchiai for allowing us access to Bosque y Vegetación Protectores Rumi Wilco. Alicia Cerchiai, Alejandro Mancino, Verónica Romero, and Veronika Schottenheim contributed to plant and butterfly stock maintenance. Yi Peng Toh assisted with brain imaging.

Portions of the text were revised with assistance from OpenAI's ChatGPT (GPT-4, accessed July 2025) to improve clarity and style. All content was critically reviewed and finalized by the authors.

Funding information

Research Funded by ERC Starter Grant 851040 to R.M.M. and a scholarship to D.F.R.S. from the Ministerio de Ciencia, Tecnología e Innovación (Convocatoria No. 860).

Conflict of interest

The authors have no conflict of interest to declare.

Author Contributions

J.B.: conceptualization, data curation, formal analysis, investigation, visualization, writing—original draft, writing—review and editing; D.F.R.S.: conceptualization, investigation, writing—review and editing; D.S.W.: methodology, writing—review and editing; S.H.M.: conceptualization, supervision, writing—review and editing; A. K.: resources, formal analysis, writing—review and editing.; R.M.M.: conceptualization, resources, supervision, writing—original

459   draft, writing—review and editing.

461   **Data Availability Statement**

462   All data and analysis are available from the Dryad Digital Repository.

References

Alm, J., Ohnmeiss, T. E., Lanza, J., & Vriesenga, L. (1990). Preference of cabbage white butterflies and honey bees for nectar that contains amino acids. *Oecologia*, *84*(1), 53–57. https://doi.org/10.1007/BF00665594

Bell, K. L., de Vere, N., Keller, A., Richardson, R. T., Gous, A., Burgess, K. S., & Brosi, B. J. (2016). Pollen DNA barcoding: Current applications and future prospects. *Genome*, *59*(9), 629–640. https://doi.org/10.1139/gen-2015-0200

Boggs, C., & Gilbert, L. E. (1979). Male Contribution to Egg Production in Butterflies: Evidence for Transfer of Nutrients at Mating. *Science*, *206*(4414), 83–84. https://doi.org/10.1126/science.206.4414.83

Boggs, C., Smiley, J., & Gilbert, L. (1981). Patterns of pollen exploitation by Heliconius butterflies. *Oecologia*, *48*, 284–289. https://doi.org/10.1007/BF00347978

Borrero, J., Mogollon Perez, E., Wright, D. S., Lozano-Urrego, D., Rueda-Muñoz, G., Pardo-Diaz, C., Salazar, C., Montgomery, S. H., & Merrill, R. M. (2024). Weighting of sensory cues reflect changing patterns of visual investment during ecological divergence in Heliconius butterflies. *Biology Letters*, *20*(10), 20240377. https://doi.org/10.1098/rsbl.2024.0377

Borrero, J., Wright, D. S., Bacquet, C. N., & Merrill, R. M. (2023). Oviposition behavior is not affected by ultraviolet light in a butterfly with sexually-dimorphic expression of a UV-sensitive opsin. *Ecology and Evolution*, *13*(7), e10243. https://doi.org/10.1002/ece3.10243

Cardoso, M. (2001). Patterns of Pollen Collection and Flower Visitation by Heliconius Butterflies in Southeastern Mexico. *Journal of Tropical Ecology*, *17*(5), 763–768. https://doi.org/10.1017/S0266467401001572

Chakraborty, M., Lara, A. G., Dang, A., McCulloch, K. J., Rainbow, D., Carter, D., Ngo, L. T.,

Solares, E., Said, I., Corbett-Detig, R. B., Gilbert, L. E., Emerson, J. J., & Briscoe, A. D.

(2023). Sex-linked gene traffic underlies the acquisition of sexually dimorphic UV color

vision in Heliconius butterflies. *Proceedings of the National Academy of Sciences*,

120(33), e2301411120. https://doi.org/10.1073/pnas.2301411120

Chmielewski, M. W., Naya, S., Borghi, M., Cortese, J., Fernie, A. R., Swartz, M. T., Zografou, K.,

Sewall, B. J., & Spigler, R. B. (2023). Phenology and foraging bias contribute to sex-

specific foraging patterns in the rare declining butterfly Argynnis idalia idalia. *Ecology*

and Evolution, *13*(7), e10287. https://doi.org/10.1002/ece3.10287

Couto, A., Young, F. J., Atzeni, D., Marty, S., Melo-Flórez, L., Hebberecht, L., Monllor, M., Neal,

C., Cicconardi, F., McMillan, W. O., & Montgomery, S. H. (2023). Rapid expansion and

visual specialisation of learning and memory centres in the brains of Heliconiini

butterflies. *Nature Communications*, *14*(1), Article 1. https://doi.org/10.1038/s41467-023-

39618-8

Davison, A., McMillan, W. O., Griffin, A. S., Jiggins, C. D., & Mallet, J. L. B. (1999). Behavioral

and physiological differences between two parapatric Heliconius species. *Biotropica*,

31(4), 661–668. https://doi.org/10.1111/j.1744-7429.1999.tb00415.x

Dell'Aglio, D., Akkaynak, D., McMillan, W. O., & Jiggins, C. D. (2017). Estimating the age of

Heliconius butterflies from calibrated photographs. *PeerJ*, *5*, e3821.

https://doi.org/10.7717/peerj.3821

Dell'Aglio, D. D., Mena, S., Mauxion, R., McMillan, W. O., & Montgomery, S. H. (2022).

Divergence in *Heliconius* flight behaviour is associated with local adaptation to different

forest structures. *Journal of Animal Ecology*, *91*(4), 727–737.

https://doi.org/10.1111/1365-2656.13675

Dell'Aglio, D., Denise, McMillan, W. O., & Montgomery, S. H. (2022). Shifting balances in the

weighting of sensory modalities are predicted by divergence in brain morphology in

incipient species of Heliconius butterflies. *Animal Behaviour*, *185*, 83–90. https://doi.org/10.1016/j.anbehav.2022.01.003

Dell'Aglio, D., McMillan, O. W., & Montgomery, S. (2024). Using motion-detection cameras to monitor foraging behaviour of individual butterflies. *Ecology and Evolution*, *14*(7), e70032. https://doi.org/10.1002/ece3.70032

Descimon, H., & Mast, D. M. (1984). Semispecies relationships between Heliconius erato cyrbia Godt. And H. himera Hew. In southwestern Ecuador. *The Journal of Research on the Lepidoptera*, *22*(4), 229–237. https://doi.org/10.5962/p.266821

Dötterl, S., Milchreit, K., & Schäffler, I. (2011). Behavioural plasticity and sex differences in host finding of a specialized bee species. *Journal of Comparative Physiology A*, *197*(12), 1119–1126. https://doi.org/10.1007/s00359-011-0673-2

Dunlap-Pianka, H., Boggs, C. L., & Gilbert, L. E. (1977). Ovarian Dynamics in Heliconiine Butterflies: Programmed Senescence versus Eternal Youth. *Science*, *197*(4302), 487–490. https://doi.org/10.1126/science.197.4302.487

Eberhard, S. H., Hrassnigg, N., Crailsheim, K., & Krenn, H. W. (2007). Evidence of protease in the saliva of the butterfly *Heliconius melpomene* (L.) (Nymphalidae, Lepidoptera). *Journal of Insect Physiology*, *53*(2), 126–131. https://doi.org/10.1016/j.jinsphys.2006.11.001

Edgar, R. C. (2016). *SINTAX: A simple non-Bayesian taxonomy classifier for 16S and ITS sequences* (p. 074161). bioRxiv. https://doi.org/10.1101/074161

Erhardt, A., & Rusterholz, H.-P. (1998). Do peacock butterflies (Inachis io L.) detect and prefer nectar amino acids and other nitrogenous compounds? *Oecologia*, *117*, 536–542.

Estrada, C., & Jiggins, C. D. (2002). Patterns of pollen feeding and habitat preference among Heliconius species. *Ecological Entomology*, *27*(4), 448–456. https://doi.org/10.1046/j.1365-2311.2002.00434.x

Finkbeiner, S. D., & Briscoe, A. D. (2021). True UV color vision in a female butterfly with two UV

opsins. *Journal of Experimental Biology*, *224*(18), jeb242802.

https://doi.org/10.1242/jeb.242802

Gilbert, L. E. (1972). Pollen Feeding and Reproductive Biology of *Heliconius* Butterflies.

Proceedings of the National Academy of Sciences, *69*(6), 1403–1407.

https://doi.org/10.1073/pnas.69.6.1403

Gilbert, L. E. (1991). Biodiversity of a Central American Heliconius community: Pattern, process,

and problems. *Plant-Animal Interactions: Evolutionary Ecology in Tropical and*

Temperate Regions, 403–427.

Harpel, D., Cullen, D. A., Ott, S. R., Jiggins, C. D., & Walters, J. R. (2015). Pollen feeding

proteomics: Salivary proteins of the passion flower butterfly, *Heliconius melpomene*.

Insect Biochemistry and Molecular Biology, *63*, 7–13.

https://doi.org/10.1016/j.ibmb.2015.04.004

Hodge, E. A., Alcalde Anton, A., Bestea, L., Hernández, G., Margareth Aguilar, J., Farnworth, M.

S., Dalbosco Dell'Aglio, D., McMillan, W. O., & Montgomery, S. H. (2025). Modality-

specific long-term memory enhancement in *Heliconius* butterflies. *Philosophical*

Transactions of the Royal Society B: Biological Sciences, *380*(1929), 20240119.

https://doi.org/10.1098/rstb.2024.0119

Houston, A. I. (1995). Energetic constraints and foraging efficiency. *Behavioral Ecology*, *6*(4),

393–396. https://doi.org/10.1093/beheco/6.4.393

Jiggins, C. D. (2017). *The Ecology and Evolution of Heliconius Butterflies*. Oxford University

Press. https://doi.org/10.1093/acprof:oso/9780199566570.001.0001

Jiggins, C. D., McMillan, W. O., Neukirchen, W., & Mallet, J. (1996). What can hybrid zones tell

us about speciation? The case of Heliconius erato and H. himera (Lepidoptera:

Nymphalidae). *Biological Journal of the Linnean Society*, *59*(3), 221–242.

https://doi.org/10.1111/j.1095-8312.1996.tb01464.x

Keller, A., Danner, N., Grimmer, G., Ankenbrand, M., Von Der Ohe, K., Von Der Ohe, W., Rost, S., Härtel, S., & Steffan-Dewenter, I. (2015). Evaluating multiplexed next-generation sequencing as a method in palynology for mixed pollen samples. *Plant Biology*, *17*(2), 558–566. https://doi.org/10.1111/plb.12251

Keller, A., Hohlfeld, S., Kolter, A., Schultz, J., Gemeinholzer, B., & Ankenbrand, M. J. (2020). BCdatabaser: On-the-fly reference database creation for (meta-)barcoding. *Bioinformatics*, *36*(8), 2630–2631. https://doi.org/10.1093/bioinformatics/btz960

Kemp, D. J. (2006). Heightened phenotypic variation and age-based fading of ultraviolet butterfly wing coloration. *Evolutionary Ecology Research*, *8*(3), 515–527.

Kishi, S., & Kakutani, T. (2020). Male Visitors May Decrease Modularity in Flower–Visitor Networks. *Frontiers in Ecology and Evolution*, *8*. https://doi.org/10.3389/fevo.2020.00124

Land, M. (1997). Visual Acuity in Insects. *Annual Review of Entomology*, *42*, 147–177. https://doi.org/10.1146/annurev.ento.42.1.147

Layek, U., Das, A., & Das, U. (2022). Floral biology, floral volatile organic compounds and floral visitors of Chromolaena odorata, an invasive alien species in West Bengal, India. *Biodiversitas Journal of Biological Diversity*, *23*(4). https://doi.org/10.13057/biodiv/d230447

Maia, R., Gruson, H., Endler, J. A., & White, T. E. (2019). pavo 2: New tools for the spectral and spatial analysis of colour in r. *Methods in Ecology and Evolution*, *10*(7), 1097–1107. https://doi.org/10.1111/2041-210X.13174

Martins, A. C., Proença, C. E. B., Vasconcelos, T. N. C., Aguiar, A. J. C., Farinasso, H. C., de Lima, A. T. F., Faria, J. E. Q., Norrana, K., Costa, M. B. R., Carvalho, M. M., Dias, R. L., Bustamante, M. M. C., Carvalho, F. A., & Keller, A. (2023). Contrasting patterns of foraging behavior in neotropical stingless bees using pollen and honey metabarcoding. *Scientific Reports*, *13*(1), Article 1. https://doi.org/10.1038/s41598-023-41304-0

McCulloch, K. J., Osorio, D., & Briscoe, A. D. (2016). Sexual dimorphism in the compound eye of Heliconius erato: A nymphalid butterfly with at least five spectral classes of photoreceptor. *Journal of Experimental Biology*, *219*(15), 2377–2387. https://doi.org/10.1242/jeb.136523

McCulloch, K. J., Yuan, F., Zhen, Y., Aardema, M. L., Smith, G., Llorente-Bousquets, J., Andolfatto, P., & Briscoe, A. D. (2017). Sexual Dimorphism and Retinal Mosaic Diversification following the Evolution of a Violet Receptor in Butterflies. *Molecular Biology and Evolution*, *34*(9), 2271–2284. https://doi.org/10.1093/molbev/msx163

McMurdie, P. J., & Holmes, S. (2012). Phyloseq: A bioconductor package for handling and analysis of high-throughput phylogenetic sequence data. In *Biocomputing 2012* (pp. 235–246). World Scientific.

Mendoza-Cuenca, L., & Macías-Ordóñez, R. (2005). Foraging polymorphism in *Heliconius charitonia* (Lepidoptera: Nymphalidae): morphological constraints and behavioural compensation. *Journal of Tropical Ecology*, *21*(4), 407–415. https://doi.org/10.1017/S0266467405002385

Mevi-Schütz, J., & Erhardt, A. (2005). Amino Acids in Nectar Enhance Butterfly Fecundity: A Long-Awaited Link. *The American Naturalist*, *165*(4), 411–419. https://doi.org/10.1086/429150

Montgomery, S. H., & Merrill, R. M. (2017). Divergence in brain composition during the early stages of ecological specialization in Heliconius butterflies. *Journal of Evolutionary Biology*, *30*(3), 571–582. https://doi.org/10.1111/jeb.13027

Montgomery, S. H., Merrill, R. M., & Ott, S. R. (2016). Brain composition in Heliconius butterflies, posteclosion growth and experience-dependent neuropil plasticity. *Journal of Comparative Neurology*, *524*(9), 1747–1769. https://doi.org/10.1002/cne.23993

Montgomery, S. H., & Ott, S. R. (2015). Brain composition in Godyris zavaleta, a diurnal

butterfly, Reflects an increased reliance on olfactory information. *Journal of Comparative*

Neurology, *523*(6), 869–891. https://doi.org/10.1002/cne.23711

Montgomery, S. H., Rossi, M., McMillan, W. O., & Merrill, R. M. (2021). Neural divergence and

hybrid disruption between ecologically isolated *Heliconius* butterflies. *Proceedings of the*

National Academy of Sciences, *118*(6), e2015102118.

https://doi.org/10.1073/pnas.2015102118

O'Brien, D. M., Boggs, C. L., & Fogel, M. L. (2003). Pollen feeding in the butterfly Heliconius

charitonia: Isotopic evidence for essential amino acid transfer from pollen to eggs.

Proceedings of the Royal Society of London. Series B: Biological Sciences, *270*(1533),

2631–2636. https://doi.org/10.1098/rspb.2003.2552

O'Brien, D. M., Boggs, C. L., & Fogel, M. L. (2004). Making eggs from nectar: The role of life

history and dietary carbon turnover in butterfly reproductive resource allocation. *Oikos*,

105(2), 279–291. https://doi.org/10.1111/j.0030-1299.2004.13012.x

Ott, S. R. (2008). Confocal microscopy in large insect brains: Zinc–formaldehyde fixation

improves synapsin immunostaining and preservation of morphology in whole-mounts.

Journal of Neuroscience Methods, *172*(2), 220–230.

https://doi.org/10.1016/j.jneumeth.2008.04.031

Piko, J., Keller, A., Geppert, C., Batáry, P., Tscharntke, T., Westphal, C., & Hass, A. L. (2021).

Effects of three flower field types on bumblebees and their pollen diets. *Basic and*

Applied Ecology, *52*, 95–108. https://doi.org/10.1016/j.baae.2021.02.005

Pinheiro de Castro, E. C., Demirtas, R., Orteu, A., Olsen, C. E., Motawie, M. S., Zikan Cardoso,

M., Zagrobelny, M., & Bak, S. (2020). The dynamics of cyanide defences in the life cycle

of an aposematic butterfly*:* Biosynthesis versus sequestration. *Insect Biochemistry and*

Molecular Biology, *116*, 103259. https://doi.org/10.1016/j.ibmb.2019.103259

Pinheiro de Castro, E. C., McPherson, J., Jullian, G., Mattila, A., Bak, S., Montgomery, S., &

Jiggins, C. (2025). Pollen-feeding delays reproductive senescence and maintains toxicity

of Heliconius erato. *Peer Community Journal*, *5*. https://doi.org/10.24072/pcjournal.546

Quaresma, A., Ankenbrand, M. J., Garcia, C. A. Y., Rufino, J., Honrado, M., Amaral, J.,

Brodschneider, R., Brusbardis, V., Gratzer, K., Hatjina, F., Kilpinen, O., Pietropaoli, M.,

Roessink, I., Van Der Steen, J., Vejsnæs, F., Pinto, M. A., & Keller, A. (2024). Semi-

automated sequence curation for reliable reference datasets in ITS2 vascular plant DNA

(meta-)barcoding. *Scientific Data*, *11*(1), 129. https://doi.org/10.1038/s41597-024-02962-

5

Rathnayake, D. G. R. M. M. K., & Wijetunga, W. M. G. A. S. T. B. (2016). Species Composition

and Visiting Frequencies of Flower Visitors of Chromolaena odorata in a Dry Zone

Forest Patch of Sri Lanka. *Psyche: A Journal of Entomology*, *2016*(1), 8746251.

https://doi.org/10.1155/2016/8746251

Romeis, J., Städler, E., & Wäckers, F. (2005). Nectar- and pollen-feeding by adult herbivorous

insects. In *Environmental Microbiology—ENVIRON MICROBIOL* (pp. 178–219).

https://doi.org/10.1017/CBO9780511542220.008

Roulston, T. H., & Cane, J. H. (2000). Pollen nutritional content and digestibility for animals.

Plant Systematics and Evolution, *222*(1), 187–209. https://doi.org/10.1007/BF00984102

Ruedenauer, F. A., Spaethe, J., van der Kooi, C. J., & Leonhardt, S. D. (2019). Pollinator or

pedigree: Which factors determine the evolution of pollen nutrients? *Oecologia*, *191*(2),

349–358. https://doi.org/10.1007/s00442-019-04494-x

Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., Preibisch, S.,

Rueden, C., Saalfeld, S., Schmid, B., Tinevez, J.-Y., White, D. J., Hartenstein, V., Eliceiri,

K., Tomancak, P., & Cardona, A. (2012). Fiji: An open-source platform for biological-

image analysis. *Nature Methods*, *9*(7), 676–682. https://doi.org/10.1038/nmeth.2019

Sickel, W., Ankenbrand, M. J., Grimmer, G., Holzschuh, A., Härtel, S., Lanzen, J., Steffan-Dewenter, I., & Keller, A. (2015). Increased efficiency in identifying mixed pollen samples by meta-barcoding with a dual-indexing approach. *BMC Ecology*, *15*(1), 20. https://doi.org/10.1186/s12898-015-0051-y

Sivinski, J. (1989). Mushroom body development in nymphalid butterflies: A correlate of learning? *Journal of Insect Behavior*, *2*(2), 277–283. https://doi.org/10.1007/BF01053299

Smid, H. M., & Vet, L. E. (2016). The complexity of learning, memory and neural processes in an evolutionary ecological context. *Current Opinion in Insect Science*, *15*, 61–69. https://doi.org/10.1016/j.cois.2016.03.008

Smith, G. P., Bronstein, J. L., & Papaj, D. R. (2019). Sex differences in pollinator behavior: Patterns across species and consequences for the mutualism. *Journal of Animal Ecology*, *88*(7), 971–985. https://doi.org/10.1111/1365-2656.12988

Smith, G. P., Gardner, J., Gibbs, J., Griswold, T., Hauser, M., Yanega, D., & Ponisio, L. C. (2021). Sex-associated differences in the network roles of pollinators. *Ecosphere*, *12*(12), e03863. https://doi.org/10.1002/ecs2.3863

Somanathan, H., Borges, R. M., Warrant, E. J., & Kelber, A. (2017). Visual Adaptations for Mate Detection in the Male Carpenter Bee Xylocopa tenuiscapa. *PLOS ONE*, *12*(1), e0168452. https://doi.org/10.1371/journal.pone.0168452

Stöckl, A., Heinze, S., Charalabidis, A., El Jundi, B., Warrant, E., & Kelber, A. (2016). Differential investment in visual and olfactory brain areas reflects behavioural choices in hawk moths. *Scientific Reports*, *6*(April), 1–10. https://doi.org/10.1038/srep26041

Toure, M. W., Young, F. J., McMillan, W. O., & Montgomery, S. H. (2020). Heliconiini butterflies can learn time-dependent reward associations. *Biology Letters*, *16*(9), 20200424. https://doi.org/10.1098/rsbl.2020.0424

Ulmer, T., & MacDougal, J. M. (2004). *Passiflora: Passionflowers of the world*. Timber Press (OR).

Warton, D. I., Duursma, R. A., Falster, D. S., & Taskinen, S. (2012). Smatr 3– an R package for estimation and inference about allometric lines. *Methods in Ecology and Evolution*, *3*(2), 257–259. https://doi.org/10.1111/j.2041-210X.2011.00153.x

Wilson, R. S., Keller, A., Shapcott, A., Leonhardt, S. D., Sickel, W., Hardwick, J. L., Heard, T. A., Kaluza, B. F., & Wallace, H. M. (2021). Many small rather than few large sources identified in long-term bee pollen diets in agroecosystems. *Agriculture, Ecosystems & Environment*, *310*, 107296. https://doi.org/10.1016/j.agee.2020.107296

Wist, T. J., & Davis, A. R. (2008). Floral Structure and Dynamics of Nectar Production in Echinacea pallida var. Angustifolia (Asteraceae). *International Journal of Plant Sciences*, *169*(6), 708–722. https://doi.org/10.1086/533602

Wright, D. S., Manel, A. N., Guachamin-Rosero, M., Chamba-Vaca, P., Bacquet, C. N., & Merrill, R. M. (2023). Quantifying visual acuity in Heliconius butterflies. *Biology Letters*, *19*(12), 20230476. https://doi.org/10.1098/rsbl.2023.0476

Wright, D. S., & Merrill, R. M. (2025a, July). *Protocol for ImageJ/FIJI analysis of eye, tibia and abdomen measurements of Lepidoptera*. OSF. https://doi.org/10.17605/OSF.IO/FM8VP

Wright, D. S., & Merrill, R. M. (2025b, July). *Protocol: Lepidoptera Eye Dissection & Mounting*. OSF. https://doi.org/10.17605/OSF.IO/EWBS9

Wright, D. S., Rodriguez-Fuentes, J., Ammer, L., Darragh, K., Kuo, C.-Y., McMillan, W. O., Jiggins, C. D., Montgomery, S. H., & Merrill, R. M. (2024). Selection drives divergence of eye morphology in sympatric Heliconius butterflies. *Evolution*, qpae073. https://doi.org/10.1093/evolut/qpae073

Young, F. J., Alcalde Anton, A., Melo-Flórez, L., Couto, A., Foley, J., Monllor, M., McMillan, W. O., & Montgomery, S. H. (2024). Enhanced long-term memory and increased mushroom body plasticity in Heliconius butterflies. *iScience*, *27*(2), 108949. https://doi.org/10.1016/j.isci.2024.108949

Young, F. J., & Montgomery, S. H. (2020). Pollen feeding in *Heliconius* butterflies: The singular

evolution of an adaptive suite. *Proceedings of the Royal Society B: Biological Sciences*,

287(1938), 20201304. https://doi.org/10.1098/rspb.2020.1304

Zhu, H., Qu, Y., Yuan, K., You, Y., Ren, B., Wester, P., & Rusman, Q. (2025). Temporal variation

in floral scent emission of a woody plant and flower visiting behaviour of male and

female flies. *Oikos, 2025*(5), e10727. https://doi.org/10.1111/oik.10727

Ziemba, K., & Rutowski, R. (2000). Sexual Dimorphism in Eye Morphology in a Butterfly

(Asterocampa leilia; Lepidoptera, Nymphalidae). *Psyche: A Journal of Entomology*, *103.*

https://doi.org/10.1155/2000/54503

728