Structural basis for the folding of PINK1 by the HSP90–CDC37 chaperone complex

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Abstract

PTEN-induced kinase 1 (PINK1) is a mitochondrial serine/threonine kinase that plays a central role in Parkin-dependent mitophagy. Mutations in PINK1 are associated with autosomal recessive forms of Parkinson's disease. PINK1 is identified as a high-affinity client of the HSP90–CDC37 complex. Stability of PINK1 is regulated by the HSP90–CDC37 chaperone system. However, the molecular mechanism by which HSP90–CDC37 recognizes and facilitate the folding of PINK1 remains unclear. Here, we present a cryogenic electron microscopy (cryo-EM) structure of the human PINK1–HSP90–CDC37 complex. The β5 strand of the PINK1 N-lobe projects into the central channel formed at the interface between two protomers in the HSP90 dimer, which holds the PINK1 kinase domain in a partially unfolded state. The C-lobe and unique C-terminal extension (CTE) of PINK1 is folded. HSP90 covers the CTE of PINK1, which overlaps with interaction sites for TOM5, TOM20, and the intramolecular N-helix. The HPNI motif of CDC37 interacts with the C-lobe of PINK1, mimicking the HPNI motif in the N-lobe of PINK1. A pathogenic mutation L347P is suggested to disrupt the interaction with the HPNI motif of CDC37. Another pathogenic mutation H271Q is located within the HPNI motif of the N-lobe of PINK1. These findings provide structural insights into the folding of PINK1 and its dysfunction in Parkinson's disease.

Introduction

Mutations in the genes encoding PINK1 and Parkin (also known as PARK6 and PARK2, respectively) can cause autosomal recessive Parkinson's disease (1, 2). PINK1 is a mitochondrial serine/threonine kinase that plays a critical role in Parkin-dependent mitophagy to maintain mitochondrial quality (3-5). Under normal conditions, PINK1 is imported into mitochondria in a membrane potential-dependent manner and cleaved by mitochondrial proteases (6-8). During these import and processing steps, the kinase domain of PINK1 remains exposed to the cytosol in an inactive, non-phosphorylated state (9, 10). The cleaved PINK1 is released from mitochondria and subsequently degraded by the ubiquitin–proteasome pathway, resulting in a rapid turnover of PINK1 (3, 4, 11). Proteasome inhibition causes the accumulation of excessive PINK1 in the cytosol and induces the aggregate formation (11, 12). Meanwhile, loss of the membrane potential of damaged mitochondria abrogates the import and processing of PINK1 and facilitates the PINK1 accumulation on the outer mitochondrial membrane to induce Parkin-dependent mitophagy (10). The state of PINK1 is regulated at multiple steps, ensuring precise surveillance of damaged mitochondria.

The accumulated PINK1 on the outer mitochondrial membrane becomes activated through autophosphorylation in trans (10, 13) and phosphorylates both ubiquitin and the ubiquitin-like (UBL) domain of Parkin (14-17). The phosphorylated Parkin interacts with the phosphorylated ubiquitin, which allosterically activates Parkin (16, 18). The activated Parkin promotes polyubiquitination of mitochondrial substrates to initiate the selective clearance of damaged mitochondria (19). A recent cryo-EM study revealed that the PINK1–TOM complex assembles around VDAC2 and promotes activation of PINK1 through trans auto-phosphorylation (20). The C-terminal extension (CTE) of the PINK1 kinase domain, which is unique to PINK1 and characterizes its atypical kinase architecture, interacts with its N-helix as well as with TOM5 and TOM20, which are components of the TOM complex.

A kinome-wide quantitative analysis of the HSP90–CDC37–client interaction identified PINK1 as

a high-affinity substrate of the HSP90–CDC37 complex (21). HSP90 and its co-chaperone CDC37 are a central chaperone complex for protein kinases (22, 23), which is required for the folding and stabilization of a wide range of kinases (21). The stability of PINK1 is influenced by the HSP90–CDC37 chaperone system (24-27). The L347P mutation of PINK1, a Parkinson's disease–associated mutation, disrupts the PINK1–HSP90–CDC37 interaction and leads to rapid degradation of PINK1 (24, 26). Despite the importance of the PINK1 stabilization by the HSP90–CDC37 chaperone complex, its underlying structural mechanism remains to be elucidated.

In this study, we report the cryo-EM structure of the PINK1–HSP90–CDC37 complex at a nominal resolution of 3.08 Å. The kinase domain of PINK1 is held in an immature conformation. The N-lobe of PINK1 remains unfolded, whereas the C-lobe and CTE of PINK1 are folded. HSP90 covers the CTE of PINK1. The C-lobe of PINK1 interacts with the conserved HPNI motif of CDC37, which mimics the HPNI motif present in the folded kinase domain of PINK1. Pathogenic mutations H271Q and L347P of PINK1 affect intramolecular and intermolecular interactions via the HPNI motif, respectively. These findings provide structural insights into the recognition and stabilization of the atypical kinase architecture of PINK1 by the HSP90–CDC37 complex, and highlight structural and mechanistic relationships between chaperone-mediated folding and pathogenic mutations causing Parkinson's disease.

Results

Complex formation between human PINK1 and the HSP90–CDC37 chaperone.

PINK1 consists of an N-terminal mitochondrial targeting sequence (MTS), an outer mitochondrial localization signal (OMS), and a single transmembrane domain (TMD) followed by N-helix and a kinase domain (Fig. 1a). The N-lobe of the kinase domain of PINK1 is composed of five β-strands, the αC-helix and three insertions. The C-lobe and the unique CTE is mainly composed of α-helices. The cleft between the N- and C-lobes forms the ATP-binding pocket. The CTE interacts with the N-helix and forms an interface for binding to components of the TOM complex. HSP90 is a dimeric molecular chaperone. Each protomer contains an ATP-binding N-terminal domain (NTD^{HSP90}), a middle domain (MD^{HSP90}), and a C-terminal dimerization domain (CTD^{HSP90}) (Fig. 1a). The NTD^{HSP90} and MD^{HSP90} are connected by a flexible charged linker (FCL^{HSP90}). Two protomers form a channel through dimerization. This channel adopts a clamp-like structure and accommodates client proteins (Fig. 1b). CDC37 functions as a kinase-specific co-chaperone. The N-terminal domain of CDC37 (NTD^{CDC37}) directly binds to the kinase domain of client proteins (Fig. 1a). The middle domain of CDC37 (MD^{CDC37}) connects to HSP90 through a linker region that extends across the outer surface of the HSP90 dimer, thereby bridging kinases to the chaperone (Fig. 1b).

PINK1 is cleaved by PARL and exposes Phe104 at the N-terminus (7, 11). The phenylalanine residue at the N-terminus is recognized by the E3 ubiquitin ligases (Ubr1, Ubr2, and Ubr4) of the N-end rule pathway as the destabilizing residue and subsequently degraded by the proteasome (11). To express the stabilized fragment of PINK1, we replaced Phe104 with Met, and deleted the N-terminal 103 residues. For cryo-EM analysis, we co-expressed human PINK1 (residues 104–581, F104M), HSP90β, and CDC37 in insect cells and purified the PINK1–HSP90β–CDC37 complex. HSP90 functions as a dimer, in which the CTD^{HSP90} forms a relatively stable dimerization interface (28). The NTD^{HSP90} alternates between the open and closed conformations in response to the ATPase cycle (28). To stabilize the HSP90 dimer during purification,

molybdate was added in the buffer, which is known to trap HSP90 in the closed state that corresponds to its client-bound conformation (29-31). The PINK1–HSP90β–CDC37 complex was first purified by affinity chromatography (Supplementary Fig. 1a), followed by gel filtration chromatography (Supplementary Fig. 1b). The subunit composition of the complex was confirmed by SDS–PAGE (Supplementary Fig. 1b). Fractions corresponding to the peak top of the gel-filtration profile containing PINK1, HSP90β, and CDC37 were collected and rapidly vitrified on EM grids for single-particle cryo-EM analysis.

Cryo-EM single particle analysis of the human PINK1–HSP90–CDC37 complex in native conditions. Among 1,606,203 particles picked from 4,217 movies without templates with particle diameters ranging from 100 to 200 Å, 1,373,615 particle images were extracted with a box size of 400 pixels (0.724 Å/pixel) and downsampled to a size of 96 pixels (3.02 Å/pixel) by Fourier cropping (Supplementary Fig. 2a). The downsampled images were subjected to reference-free 2D classification, which generated the dimeric projection images. 62 classes with 758,357 particles were used as templates for picking particles with a particle diameter of 200 Å. 1,586,437 particle images were extracted and subjected to the second round of 2D classification, resulting in a set of images similar to that generated in the first round of 2D classification. We selected 38 classes with clear structural features and used 607,867 particles for the *ab initio* 3D reconstruction. Four classes of *ab initio* 3D models were generated and then refined by heterogeneous refinement. The resultant 3D maps from two classes represented the HSP90 dimer, one in a closed state and the other in a widely open state. For the closed state, 199,855 particles were selected from 208,564 particles after excluding junk particles through 2D classification and performed homogeneous refinement and non-uniform refinement. Non-uniform refinement yielded a density map at a nominal resolution of 3.05 Å. For the widely open state, 211,262 particles were selected from 216,240 particles after 2D classification and performed homogeneous refinement and non-uniform refinement. Non-uniform refinement yielded a density map at a nominal

resolution of 3.04 Å.

The cryo-EM analysis revealed two distinct states of the PINK1–HSP90β–CDC37 complex. One state indicates that the HSP90 dimer adopts a closed conformation. In this state, weak density corresponding to the client is visible. The absence of the double-helix of CDC37 suggests that CDC37 has dissociated. In the other state, the HSP90 dimer displays an unusually open conformation at the CTD^{HSP90}. The density corresponding to both PINK1 and CDC37 are not observed. While conformational opening and closing of the NTD^{HSP90} are well established for HSP90 (32), such a pronounced separation of the CTD^{HSP90} has not previously been reported. Considering the possibility that the complex was disassembled during vitrification, we next attempted chemical crosslinking to stabilize the complex.

Cryo-EM single particle analysis of the human PINK1–HSP90–CDC37 complex with chemical crosslinking.

To stabilize PINK1 in complex with HSP90 and CDC37, the eluate from the affinity chromatography was crosslinked with glutaraldehyde at a final concentration of 0.025% (Supplementary Fig. 3a), and further purified by size-exclusion chromatography (Supplementary Fig. 3b). SDS–PAGE analysis confirmed the integrity of the crosslinked complex (Supplementary Fig. 3b), which eluted at a position nearly identical to that of the native complex. Fractions corresponding to the peak were collected and rapidly vitrified on EM grids for cryo-EM analysis.

Among 1,610,071 particles picked from 4,281 movies without templates with particle diameters ranging from 100 to 200 Å, 1,379,120 particle images were extracted with a box size of 400 pixels (0.724 Å/pixel) and downsampled to a size of 96 pixels (3.02 Å/pixel) by Fourier cropping (Supplementary Fig. 4a). The downsampled images were subjected to reference-free 2D classification, which generated the dimeric projection images. 95 classes with 1,325,908 particles were used as templates for picking particles with a particle diameter of 200 Å. 1,605,673 particle images were extracted and subjected to the second round of 2D

classification, resulting in a set of images similar to that generated in the first round of 2D classification. We selected 120 classes with clear structural features and used 1,368,585 particles for the *ab initio* 3D reconstruction. Four classes of *ab initio* 3D models were generated in the first round and refined by heterogeneous refinement. One class comprising 474,125 particles was further subjected to a second round of *ab initio* reconstruction into three classes followed by heterogeneous refinement. After excluding junk particles through 2D classification, 227,582 particles were selected from the 233,619 particles. Homogeneous and non-uniform refinements were performed to generate density maps. To further optimize the map quality, global and local CTF refinements were applied after non-uniform refinement. The subsequent homogeneous refinement, non-uniform refinement, and two rounds of local refinement yielded a 3.20 Å map. Although the second round of local refinement used the map from the first round as a reference to further optimize particle alignment, the resolution showed little improvement. To improve the accuracy of particle identification, we performed particle picking using the neural network–based method Topaz and obtained 1,171,684 particle images. After 2D classification to remove junk particles, 1,163,406 particles were used for *ab initio* 3D reconstruction and heterogeneous refinement. A single class of 454,727 particles was subjected to homogeneous, non-uniform, and local refinements, resulting in a 3.08 Å map. The atomic models of the HSP90–CDC37 complex (PDB 7Z38) and the predicted PINK1 C-lobe (AlphaFold model, AF-Q9BXM7-F1-model_v4) fit well into the density (local resolution range of 2.5 to 8.5 Å) (Supplementary Figs. 4a, b and Table 1).

Overall structure of the human PINK1–HSP90–CDC37 complex.

Cryo-EM density of the closed HSP90 dimer was observed along with protrusions corresponding to the two α-helices of the NTD^{CDC37} (Fig. 1c). The CDC37 linker was traced extending across the outer surface of HSP90, with weak density attributable to the middle domain. Residual density observed near the complex corresponded well to the known position of the C-lobe of a kinase bound to the HSP90–CDC37 complex. The AlphaFold2-

predicted model of PINK1 comprising the C-lobe and CTE (residues 318–581) was fitted into the observed density, although no corresponding density was visible for a part of the activation loop. Additional density was also observed inside the channel formed by the HSP90 dimer, which appeared to be connected to the PINK1 C-lobe.

Interaction between the C-lobe of PINK1 and the NTD^{CDC37} mediated by the HPNI motif.

The density at the interface between the C-lobe of PINK1 and the NTD^{CDC37} is well resolved (Figs. 2a, b). The C-lobe of PINK1 interacts with the NTD^{CDC37} through hydrophobic residues (Figs. 2c-e). The region between His20 and Trp31 of CDC37 are buried within the PINK1 C-lobe (Fig. 2c). Trp31 of CDC37 is positioned in close proximity to Leu353, Asp384 of the DFG motif, and Asp360 of the HRD motif of PINK1 (Fig. 2c). Ile23 of CDC37 forms hydrophobic interactions with Ile382 in β7 and Leu353 in the αE helix of PINK1 (Fig. 2d). His352 in PINK1 forms a π-π stacking interaction with His20 (Fig. 2e).

The loop (residues 20–23) of CDC37 overlaps with the loop (residues 271–274) between insertion2 (ins2) and β4 of the N-lobe of PINK1 in the predicted model (Figs. 3a, b). The primary structures of both loops share the "HPNI" motif (Figs. 3b, c), which is consistent with the established role of this sequence as a critical determinant for the kinase–CDC37 interaction (31, 33). The loop is located on the backside of the ATP-binding pocket and stabilizes the relative positioning of the N- and C-lobes (Fig. 3a). The HPNI motif of PINK1 is evolutionarily conserved (Fig. 3d). The H271Q mutation within this motif in PINK1 has been reported as a Parkinson's disease-associated mutation (Fig. 3b). These observations suggest that the NTD^{CDC37} associates with PINK1 by mimicking the interactions formed with the folded N-lobe.

The β5 strand of the PINK1 N-lobe passing through the HSP90 dimer channel.

The five β strands of the PINK1 N-lobe are a conserved structural element that contributes to the formation of

the ATP-binding pocket, together with the C-lobe. The β5 strand of PINK1 is located immediately upstream of the C-lobe (Fig. 4a). Within the central channel of the HSP90 dimer, we observed residual density, which was assigned to the β5 strand of PINK1 (Figs. 4b-e). The hydrophobic side chains of Phe315, Leu316, and Val317, on β5 of PINK1 are buried in a groove formed by Leu439, Ile517, Tyr520, and Leu611 in one of the two HSP90 protomers ($HSP90^{A}$) (Figs. 4d, e). On the other hand, no density could be clearly assigned to the other four β strands of the PINK1 N-lobe (Figs. 1c, 4c). These findings suggest that the antiparallel β sheet of the PINK1 N-lobe could not assemble in the PINK1–HSP90–CDC37 complex, resulting in unfolding of the N-lobe. The HSP90 dimer captures PINK1 in an intermediate folding state, where the C-lobe and CTE is folded but the N-lobe is unfolded.

Interaction of the PINK1 CTE with HSP90

The CTE of PINK1 is composed of a short helix (αJ) followed by three α-helices (αK–αM) connected by the loops, which is characteristic of a unique structural element not found in conventional kinases. In the present complex structure, the CTE of PINK1 is folded and positioned back onto the kinase domain (Figs. 1c, 5a). The C-lobe and CTE of PINK1 are accommodated in the cleft formed by the MD^{HSP90} and FCL^{HSP90} (Figs. 1c, 5a). The position of the αK helix in the AlphaFold2 model of PINK1 would clash with HSP90 (Fig. 5b). The αK helix is suggested to rotate to avoid the clash with HSP90. Leu539 on the αK helix of PINK1 hydrophobically interacts with Trp312 of HSP90 (Fig. 5c). The αJ–αK linker and αM helix of PINK1 are positioned close to the FCL^{HSP90} (Fig. 5d). Due to discontinuous density, the atomic model of the FCL^{HSP90} could not be built (Fig. 5d). The interaction between PINK1 and the FCL^{HSP90} appears to be relatively weak. To explore the conformational heterogeneity of the FCL^{HSP90}, we performed 3D Variability Analysis (3D VA) in CryoSPARC. This analysis visualized relative motions of the FCL^{HSP90} to the CTE of PINK1 in accordance with lower local resolution in the periphery of the complex (Supplementary Fig. 5). These structural features indicate that the

FCL^{HSP90} is loosely bound to the CTE of PINK1.

Competitive binding of the PINK1 CTE to HSP90 and the TOM complex.

PINK1 forms a complex with the translocase of the outer membrane (TOM) and VDAC on the outer membrane of depolarized mitochondria (20). In the PINK1–TOM–VDAC complex, the CTE of PINK1 interacts with the N-helix of PINK1 and the TOM20 and TOM5 components of the TOM complex (Figs. 6a, b). The CTE of PINK1 faces the MD^{HSP90} and FCL^{HSP90} in the present PINK1–chaperone complex structure. To clarify the difference in the interaction of the PINK1 CTE with HSP90 and the TOM complex, we aligned and superposed the structures of the HSP90–CDC37 and TOM–VDAC complexes, using the PINK1 CTE as the reference (Figs. 6c, d). In both complex structures, the N-helix of PINK1 makes contact with the αK and αL helices. In the TOM–VDAC complex, TOM20 interacts with the N-helix and αK helix of PINK1, whereas TOM5 interacts with the αM helix of PINK1. The positions of the N-helix of PINK1 and TOM20 partially overlap with that of the MD^{HSP90} (Fig. 6c). The position of TOM5 overlaps with that of the FCL^{HSP90} (Fig. 6d). These findings suggest that the interactions of PINK1 with the HSP90–CDC37 and TOM complexes are competitive and mutually exclusive.

Discussion

PINK1 has been identified as a high-affinity client of the HSP90–CDC37 chaperone complex by a kinome-wide quantitative screen and mass spectrometry (11, 21, 25, 26). HSP90 inhibitors such as 17-AAG and geldanamycin decrease the half-life and protein levels of PINK1 under normal conditions (26, 27). On the other hand, when mitochondrial depolarization and HSP90 inhibition occur simultaneously, different cellular dynamics of PINK1 have been observed: the amount of PINK1 protein and ubiquitin phosphorylation are decreased in some studies (34, 35), whereas the half-life of PINK1 is prolonged and its ability to recruit Parkin

remains, regardless of HSP90 inhibition, in other studies (27, 36). Thus, the role of HSP90 in regulating PINK1 under mitochondrial depolarization conditions remains controversial. Nonetheless, there is a consensus that PINK1 stability is strongly influenced by the HSP90 chaperone system under normal conditions. In this study, we determined the cryo-EM structure of the human PINK1–HSP90–CDC37 complex. In the present structure, the C-lobe and the CTE of PINK1 are already folded, whereas the N-lobe of PINK1 likely remains unfolded. This structure shows that the HSP90–CDC37 complex holds a folding intermediate of PINK1, which stabilizes key structural elements prior to full kinase domain maturation. The intermediate state suggests a stepwise folding mechanism of the kinase domain of PINK1.

CDC37 functions as a kinase-specific co-chaperone of HSP90, which associates with immature kinases and facilitates their delivery to HSP90 (23). The HPNI amino acids sequence of CDC37 is the critical conserved motif for kinase recognition (33). Disruption of this motif by mutation reduces binding affinity for client kinases, thereby impairing their stability and maturation in the cell (31, 37). The present cryo-EM structure of the PINK1–HSP90–CDC37 complex shows that the HPNI motif of CDC37 also interacts with the C-lobe of PINK1 in a manner similar to the HPNI motif of PINK1 in the AlphaFold2-predicted PINK1 structure.

The L347P and H271Q mutations of PINK1 have been identified in patients with Parkinson's disease (38, 39). The L347P mutation decreases the stability of PINK1 due to impaired interaction with the HSP90–CDC37 complex (24, 26). In the present structure, Leu347 of PINK1 is located on the αE helix of the C-lobe, which interacts with the HPNI motif of CDC37. The L347P mutation may distort αE and disrupt its interaction with CDC37. On the other hand, the H271Q mutation is located within the HPNI motif of the N-lobe of PINK1. The H271Q mutation potentially interferes with the π–π stacking between His271 and His352. We previously reported that the H271Q mutant of PINK1 is defective in complex formation and autophosphorylation upon mitochondrial depolarization, thereby impairing the recruitment and activation of

Parkin on damaged mitochondria (4, 10, 13). The H271Q mutation is suggested to affect the integrity of the kinase domain through destabilization of the interaction between the N-lobe and C-lobe, which likely hinders proper folding. Our findings provide the molecular basis for familial Parkinson's disease linked to PINK1 mutations.

A recent cryo-EM study has revealed the structure of a PINK1–TOM–VDAC array that promotes PINK1 stagnation to mitochondria and activation through trans-autophosphorylation (20). The CTE of PINK1 interacts with the N-helix and with TOM5 and TOM20, components of the TOM complex. The present PINK1–HSP90–CDC37 structure shows that the HSP90–CDC37 complex shields the CTE of PINK1. As the position of the HSP90–CDC37 complex relative to the CTE of PINK1 overlaps with those of TOM5 and TOM20 in the PINK1–TOM–VDAC array, the HSP90–CDC37 complex is likely to dissociates from PINK1 when PINK1 is recruited to damaged mitochondria. Release from the HSP90–CDC37 complex would enable folding of the N-lobe of PINK1 and intramolecular interactions between the N-helix and CTE of PINK1, thereby facilitating maturation of the kinase domain. In addition, the CTE of PINK1 can interact with components of the TOM complex, which in turn permits the activation of PINK1 through trans-autophosphorylation. Since the HSP90–CDC37 complex does not directly promote the activation of PINK1 upon mitochondrial depolarization, the effect of the HSP90 inhibition on PINK1 is still unclear. However, these findings suggest that the HSP90–CDC37 complex maintains PINK1 in the intermediate state under normal conditions, which might prime PINK1 for the activation upon mitochondrial depolarization.

Here, we propose a model in which PINK1 is held in an intermediate conformation by the HSP90–CDC37 complex and that the kinase domain of PINK1 completes folding through interactions with the TOM complex and dissociation from the HSP90–CDC37 complex on the outer membrane of damaged mitochondria (Fig. 7). The HSP90–CDC37 system involves other co-chaperones and post-translational modifications. The phosphorylation of CDC37 at Ser13 by casein kinase 2 (CK2) has been shown to be a prerequisite for the

interaction between CDC37 and client kinases (40, 41). In contrast, protein phosphatase 5 (PP5) promotes dissociation of client kinases from the HSP90–CDC37 complex by dephosphorylating CDC37 at Ser13 (41, 42). In addition, activator of HSP90 ATPase activity 1 (Aha1) is a co-chaperone known to stimulate the ATP hydrolysis of HSP90 (43, 44). Further studies are required to understand that co-chaperones and post-translational modifications of the HSP90–CDC37 complex fine-tune the PINK1 stability and activity. Our results provide a structural basis for PINK1 stabilization and reveal a mechanism by which the HSP90–CDC37 complex facilitates PINK1 folding and protects its unengaged interface.

Materials and methods

Plasmids

Human PINK1 (residues 104–581, accession No. NM_032409.3), in which Phe104 was replaced by Met, was cloned with a C-terminal His_8 tag into the BamHI and EcoRI sites downstream of the PH promoter in the pFastBacDual vector. Human HSP90β (accession No. NM_001271969.2) and human CDC37 (accession No. NM_007065.4) were cloned into the BamHI and EcoRI sites under the PH promoter and into the KpnI and XhoI sites under the P10 promoter, respectively.

Sample preparation for cryo-EM

PINK1(F104M)-His_8, HSP90β, and CDC37 were co-expressed in Sf9 cells using the Bac-to-Bac system (Invitrogen) with X-tremeGENE HP (Roche) as the transfection reagent for P1 virus generation. The P2 virus was amplified to produce 20 mL of P3 virus, which was subsequently used to infect 200 mL of Sf9 cells. The infected cells were incubated at 27 °C for 3 days. The infected Sf9 cells were harvested by centrifugation at 2,000 × g for 5 min at 4 °C and washed twice with cold phosphate buffered saline (PBS). The cell pellets were resuspended in a lysis buffer (50 mM Tris-HCl, pH 8.0, 150 mM NaCl, 10 mM $MgCl_2$, 10 mM KCl, 20 mM Na_2MoO_4, 20 mM imidazole, and 0.5% Triton X-100) and lysed by sonication on ice for 5 min. The lysate was clarified by centrifugation at 2,000 × g for 10 min at 4 °C. The cleared lysate was loaded onto a Ni-NTA agarose (Qiagen) column pre-equilibrated with the lysis buffer. The column was washed with the lysis buffer (first wash), without Triton X-100 (second wash). The bound protein was eluted with 50 mM Tris-HCl buffer (pH 8.0) containing 150 mM NaCl and 10 mM $MgCl_2$, 10 mM KCl, 20 mM Na_2MoO_4, and 300 mM imidazole. For purification of the native complex, the elution sample was further purified by a size-exclusion chromatography on a ENrich™ SEC 650 column (10 × 300 mm; Bio-Rad) equilibrated with 50 mM Tris-HCl buffer (pH 8.0) containing 150 mM NaCl and 10 mM $MgCl_2$, 10 mM KCl, and 20 mM Na_2MoO_4. A fraction

enriched in PINK1 (F104M)-His_8, HSP90β, and CDC37 was concentrated to 0.67 g/L using an Amicon Ultra-4 centrifugal filter unit (100 kDa MWCO) for cryo-EM analysis. For preparation of the crosslinked complex, the elution sample was concentrated tenfold and diluted three times using an Amicon Ultra-4 centrifugal filter unit (100 kDa MWCO), thereby exchanging the buffer to the crosslinking buffer (50 mM HEPES-Na, pH 8.0, 150 mM NaCl, 10 mM $MgCl_2$, 10 mM KCl, and 20 mM Na_2MoO_4). Crosslinking was performed by adding glutaraldehyde at a final concentration of 0.025% and incubating at room temperature for 30 min. The reaction was quenched with Tris-HCl buffer (pH 8.0) at a final concentration of 0.1 M. The crosslinked sample was further purified by size-exclusion chromatography on an ENrich™ SEC 650 column (10 × 300 mm) equilibrated with 50 mM Tris-HCl (pH 8.0), 150 mM NaCl, 10 mM $MgCl_2$, 10 mM KCl, and 20 mM Na_2MoO_4. The fraction enriched in the crosslinked complex was concentrated to 0.30 g/L using an Amicon Ultra-0.5 centrifugal filter unit (3 kDa MWCO).

Cryo-EM grid preparation and data collection

Cu grids (R1.2/1.3, 300 mesh, Quantifoil) with holey carbon films were hydrophilized using a JEC-3000FC Auto Fine Coater (JEOL) at 7 Pa, 10 mA, and 10 sec. A 3 μL aliquot of 0.67 g/L intact protein solution or 0.30 g/L crosslinked protein solution was added to the grids using a Vitrobot Mark IV (Thermo Fisher Scientific) at a temperature of 8 °C and 100% humidity. The grids were then immersed in liquid ethane and rapidly frozen under the following conditions: waiting time of 0 sec, blotting time of 5 sec, and blotting force of 15 for the intact sample, and waiting time of 0 sec, blotting time of 3 sec, and blotting force of 15 for the crosslinked sample. Data collection was carried out on a Glacios cryo-transmission electron microscope (Thermo Fisher Scientific) operating at 200 kV, equipped with a Falcon 4 electron detector at Kyoto University. A total of 4,217 or 4,281 movies were automatically collected using EPU software. Movies were collected at a target defocus range of −0.8 to −1.6 μm and a nominal magnification of ×190,000, corresponding to a calibrated

pixel size of 0.724 Å/pixel. Each movie was recorded with an exposure time of both 4.31 s, subdivided into both 66 frames with a total electron dose of 50 e^{-1} $Å^{-2}$. All processing was performed in CryoSPARC v.4.3.1, v4.4.1, and v.4.7.0 (45).

Cryo-EM data processing for the native PINK1–HSP90–CDC37 complex

The collected micrographs were processed using patch motion correction and patch CTF estimation. 1,606,203 particles were initially picked using blob picker (minimum particle diameter: 100 Å, maximum particle diameter: 200 Å). After 2D classification, 758,357 particles were selected based on the average 2D images of the protein and used as templates for template picker (particle diameter: 200 Å). 1,816,520 particles were picked. 607,867 particles selected from the average 2D images of the protein and subjected to *ab initio* reconstruction. Heterogeneous refinement was performed using four 3D references, generated two class with a well-defined structure. 199,855 or 211,262 particles were selected after 2D classification further refined by homogeneous and non-uniform refinement. Local refinement following non-uniform refinement resulted in a 3.05 Å or 3.04 Å resolution 3D map of the PINK1–HSP90–CDC37 complex. Overall resolution estimates correspond to a Fourier shell correlation of 0.143 using an optimized mask that is automatically determined after refinement.

Cryo-EM data processing for the crosslinked PINK1–HSP90–CDC37 complex

The collected micrographs were processed using patch motion correction and patch CTF estimation. 1,610,071 particles initially were picked using blob picker (minimum particle diameter: 100 Å, maximum particle diameter: 200 Å). 1,325,908 particles were selected from the average 2D images of the protein and used as templates for template picker (particle diameter: 200 Å), resulting in the picked of 1,857,354 particles. 1,368,585 particles were selected from the average 2D images of the protein and subjected to an *ab initio*

reconstruction and heterogeneous refinement into four 3D classes. One of the resulting classes was further refined through a second round of *ab initio* reconstruction and heterogeneous refinement into three 3D classes. The selected particles were further cleaned by 2D classification, resulting in 227,582 particles. These particles were processed through homogeneous refinement, non-uniform refinement, global and local CTF refinement, homogeneous refinement, non-uniform refinement, and two rounds of local refinement, yielding a final map at 3.20 Å resolution. Additionally, particle picking was performed using Topaz (46), resulting in 1,171,684 particles. 1,163,406 particles were selected after 2D classification and subjected to *ab initio* reconstruction into four classes, followed by heterogeneous refinement. 454,727 particles were selected and refined by homogeneous and non-uniform refinement. Local refinement following non-uniform refinement resulted in a 3.08 Å resolution 3D map of the PINK1–HSP90–CDC37 complex. Overall resolution estimates correspond to a Fourier shell correlation of 0.143 using an optimized mask that is automatically determined after refinement. Local resolution maps were obtained using local resolution estimation. 3D variability analysis (47) was performed on the final 454,727 particle set, using a mask that encompassed the PINK1, solving for 3 orthogonal principal modes over 20 iterations with a 8 Å resolution filter. The resulting variability was visualized using 3D variability display in cluster mode, with the number of frames/clusters set to six.

Model building

Model building was performed using Coot (48) and UCSF ChimeraX (49). The initial model of the HSP90–CDC37 complex used a part of the cryo-EM structure of the HSP90–CDC37 complex (PDB 7Z38) (31). The initial model of the C-lobe and CTE of PINK1 used AlphaFold model (AF-Q9BXM7-F1-model_v4). These models fitted to the cryo-EM map using UCSF ChimeraX. The final structure was refined using Phenix (50). All figures were created using UCSF ChimeraX.

Data availability

The coordinates and maps of the PINK1–HSP90–CDC37 complex have been deposited in the Protein Data Bank/Electron Microscopy Data Bank under the accession codes of 9X4X/EMD-66567. Other data are available from the corresponding authors upon reasonable request.

Acknowledgements

Cryo-EM data collection was performed using a Glacios cryo-TEM with the support of the Cryo-EM Facility, Institute for Life and Medical Sciences, Kyoto University. Joint Usage/Research Center Program of the Institute for Life and Medical Sciences at Kyoto University (to T.N.). This work was supported by JSPS/MEXT KAKENHI (21K15084, 24H01894 to K.O. and 18H05501 to S.F.)

Author contributions

K.O.-Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Software, Visualization, Writing – original draft.

H.Y.-Investigation, Writing – review and editing.

A.O.-Investigation, Writing – review and editing.

S.G.-Investigation, Resources

Y.N.-Investigation, Resources.

Y.S.-Resources, Supervision

T.N.-Resources, Supervision

S.F.-Supervision, Funding acquisition, Software, Project administration, Writing – review and editing.

Additional information

We declare no competing financial interests.

Figure legends

Figure 1. Cryo-EM structure of the PINK1–HSP90–CDC37 complex.

(a) Domain organizations of human PINK1, CDC37, and HSP90β. MTS, N-terminal mitochondrial targeting sequence; OMS, outer mitochondrial localization signal; TMD, transmembrane domain; CTE, C-terminal extension; NTD, N-terminal domain; MD, middle domain; CTD, C-terminal domain. **(b)** Schematic diagram of the PINK1–HSP90–CDC37 complex. The coloring scheme is the same as in (a). **(c)** Cryo-EM density map and structure of the PINK1–HSP90–CDC37 complex at 3.08 Å resolution. The coloring scheme is the same as in (a).

Figure 2. Interaction of the PINK1 C-lobe with CDC37 mediated by its loop of the N-terminal domain loop.

(a) Overall view of the cryo-EM structure of the PINK1–HSP90–CDC37 complex. The box indicates an area including the locations of the views shown in (b-e) **(b)** Close-up view of the density map at the interface between PINK1 (pink) and CDC37 (green). **(c)** Close-up view of the PINK1–CDC37 interaction around Trp31 of CDC37. The structure (top) and corresponding map (bottom) are shown. **(d)** Close-up view of the PINK1–CDC37 interaction around Ile23 of CDC37. The structure (top) and corresponding map (bottom) are shown. **(e)** Close-up view of the PINK1–CDC37 interaction around His31 of CDC37. The structure (top) and corresponding map (bottom) are shown.

Figure 3. Comparison of the HPNI motif between CDC37 and PINK1.

(a) Comparison between the NTD^{CDC37}–$C\text{-}lobe^{PINK1}$ interaction (cryo-EM) and $N\text{-}lobe^{PINK1}$–$C\text{-}lobe^{PINK1}$ interaction (AlphaFold2). The boxes indicate the loop containing the HPNI motif. **(b)** Close-up view of the

HPNI motifs of CDC37 (green; cryo-EM) and PINK1 (red; AlphaFold2). **(c)** Amino-acid sequence alignment of the HPNI motif-containing loop of human PINK1 and CDC37. *, identical residues; :, residues with strongly similar properties; ., residues with weakly similar properties. **(d)** Amino-acid sequence alignment of the HPNI motif-containing loop of PINK1 among representative vertebrates. *, identical residues; :, residues with strongly similar properties; ., residues with weakly similar properties.

Figure 4. Insertion of the β5 strand of the PINK1 N-lobe into the interfacial channel of the HSP90 dimer.

(a) The AlphaFold2-predicted model (AF-Q9BXM7-F1-model_v4) of human PINK1. The N-lobe contains the five-stranded β sheet (β1–β5). (**b)** Overall view of the cryo-EM structure of the PINK1–HSP90–CDC37 complex. **(c)** Close-up view of the unfolded segment of the PINK1 N-lobe, which is inserted into the channel at the interface between two protomers in the HSP90 dimer. **(d, e)** Close-up view of the density map and structures around the unfolded segment of the PINK1 N-lobe.

Figure 5. Interface between PINK1 and HSP90.

(a) The structure and density map of the interface between PINK1 and HSP90. The two boxes indicate the locations of the views shown in (c) and (d) **(b)** Superposition of the cryo-EM and AlphaFold2-predicted PINK1 structures. **(c)** Close-up view of the structure and density map around Trp312 of HSP90 and Leu539 of PINK1. **(d)** Close-up view of the interaction between the PINK1 CTE and HSP90 FCL.

Figure 6. Comparison of the spatial arrangement of HSP90–CDC37 and TOM–VDAC relative to PINK1.

(a) Cryo-EM structure of the PINK1–TOM–VDAC complex (PDB 9EIH). **(b)** Close-up views of PINK1–TOM5–TOM20 in the PINK1–TOM–VDAC complex. The other components of the complex are omitted for clarity. **(c)** Comparison between the PINK1–TOM20 and PINK1–HSP90 interactions. In the PINK1–TOM20

interface (left panel), TOM20 interacts with both the PINK1 CTE and N-helix, which also interact with each other. In the PINK1–HSP90 interface (middle panel), the FCL^{HSP90} covers the PINK1 CTE. The superposition (right panel) shows that the PINK1 N-helix and TOM20 overlap with the NTD^{HSP90} and MD^{HSP90}. **(d)** Comparison between the PINK1–TOM5 and PINK1–HSP90 interactions. In the PINK1–TOM5 interface (left panel), the TOM5 interacts with the PINK1 CTE. The superposition (right panel) shows that TOM5 overlaps with the FCL^{HSP90}.

Figure 7. Model for PINK1 stabilization by the HSP90–CDC37 complex and transition to its active state on depolarized mitochondria.

(a) In the cytosol, the HSP90–CDC37 complex captures the folding intermediate of PINK1, where the N-lobe remains unfolded due to binding of the isolated β5 strand to HSP90, whereas the C-lobe and CTE is folded. The CTE is covered with FCL^{HSP90}. The HPNI motif of CDC37 interacts with the C-lobe of PINK1, mimicking the HPNI motif of the N-lobe. **(b)** TOM20 and TOM5 compete with HSP90–CDC37 for binding to the PINK1 CTE, displacing the chaperone complex from PINK1. **(c)** Upon mitochondrial depolarization, the dissociation of the HSP90–CDC37 complex allows the PINK1 CTE to interact with the N-helix and further with TOM20 and TOM5, along with the N-lobe folding. Finally, PINK1 dimerizes and becomes activated in a trans-autophosphorylation fashion.

Table 1 Data collection and refinement statistics

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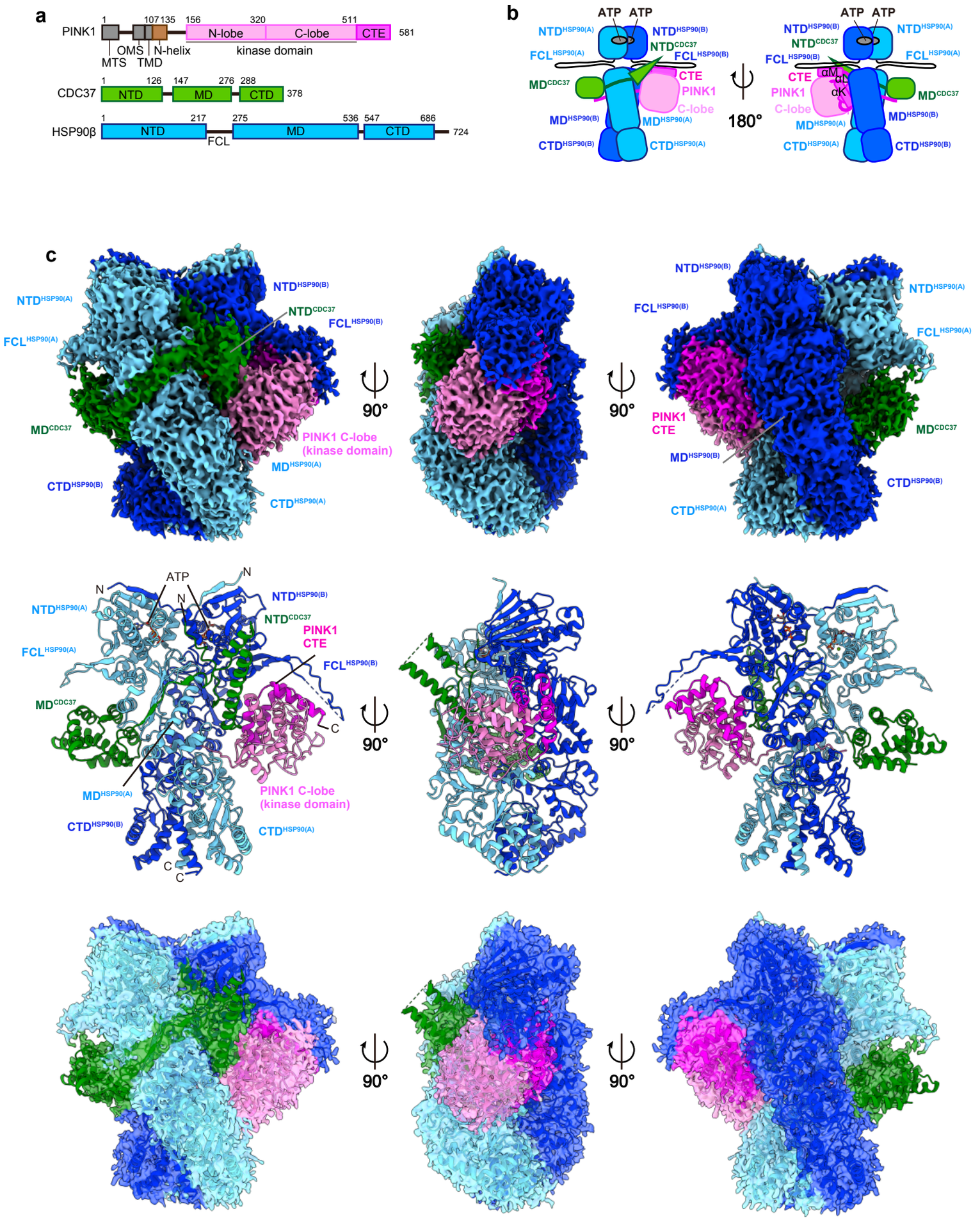

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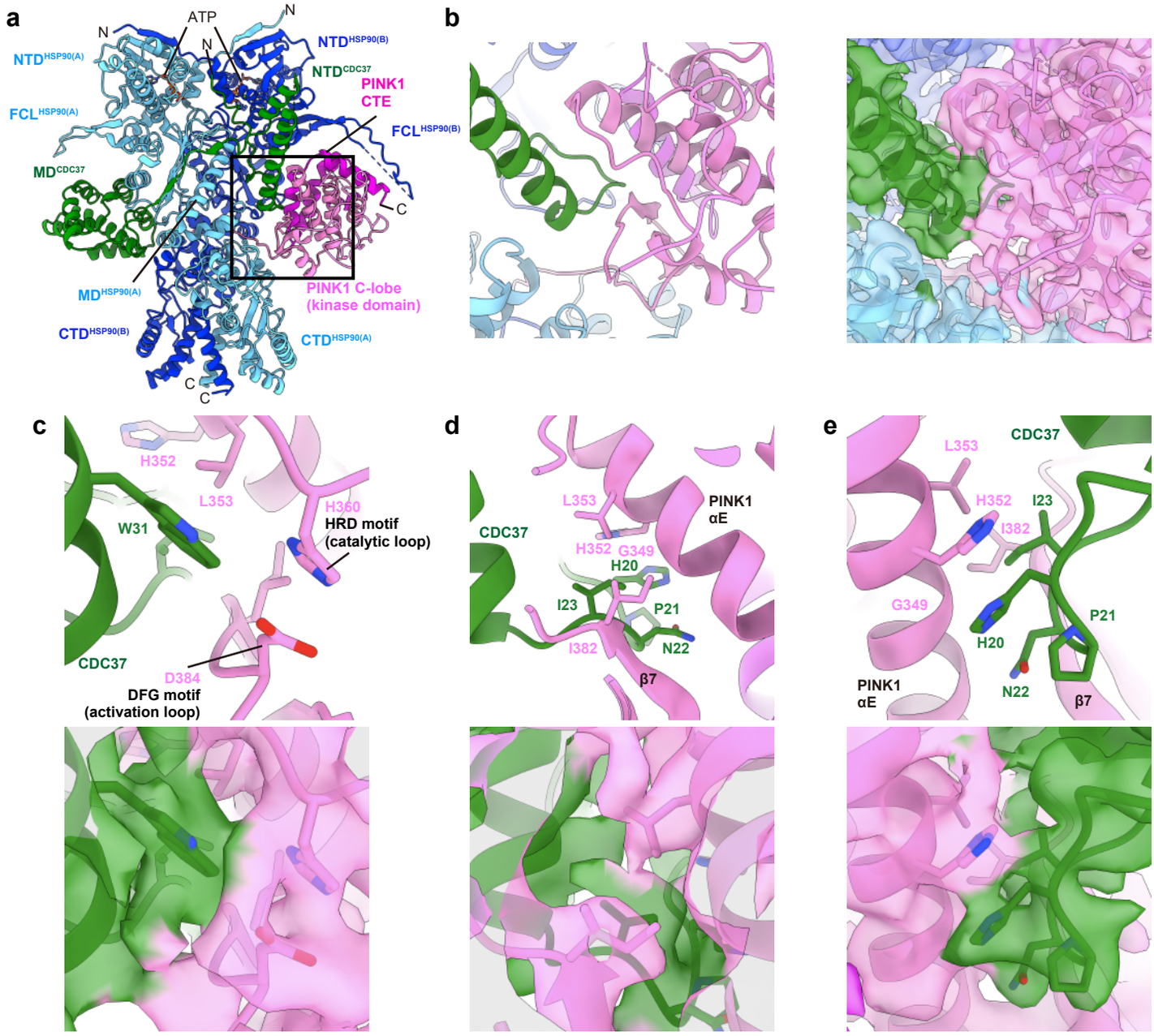

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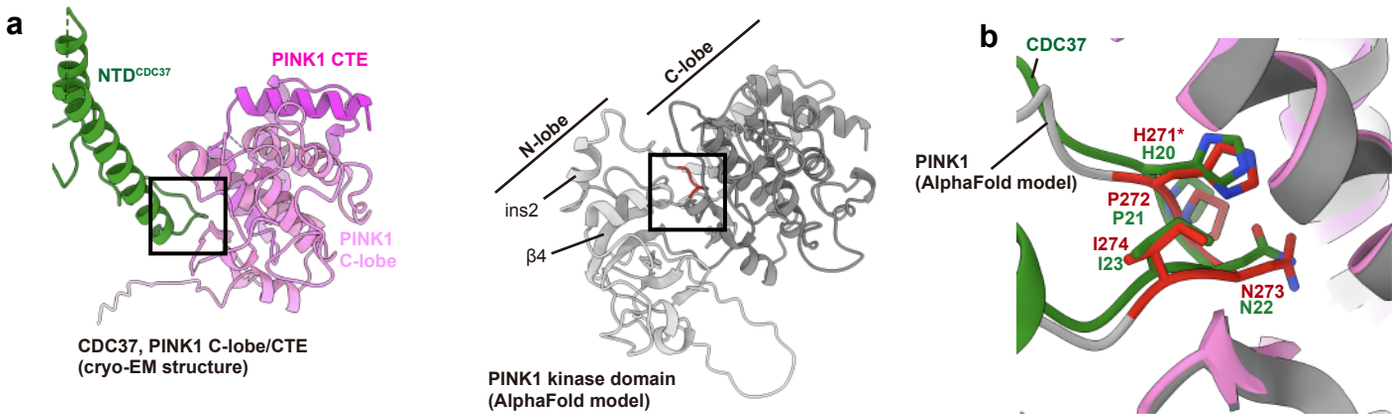

c

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PINK1 261- SKRGPKQLAPHPNIIR--VLRAFTSS -284
CDC37  10- IEVSDDEDETHPNIDTASLFRW-RH- -33
            : . .:   ****    ::*
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d

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Species (PINK1)
Homo_sapiens      SKRGPKQLAPHPNIIRVLRAFTSS
Mus_musculus      SRDGPKQLAPHPNIIRVFRAFTSS
Gallus_gallus     HVLGRKRLRPHPNIIQVIRAFTSS
Danio_rerio       FGAVPKRLSAHPNVITVYRAFTAE
Bos_taurus        PKGGPKQLAPHPNIIRVLRAFTSS
Macaca_mulatta    SKRGPKQLAPHPNIIRVLRAFTSS
Pan_troglodytes   SKRGPKQLAPHPNIIRVLRAFTSS
                       *:*  ***:* * ****:.
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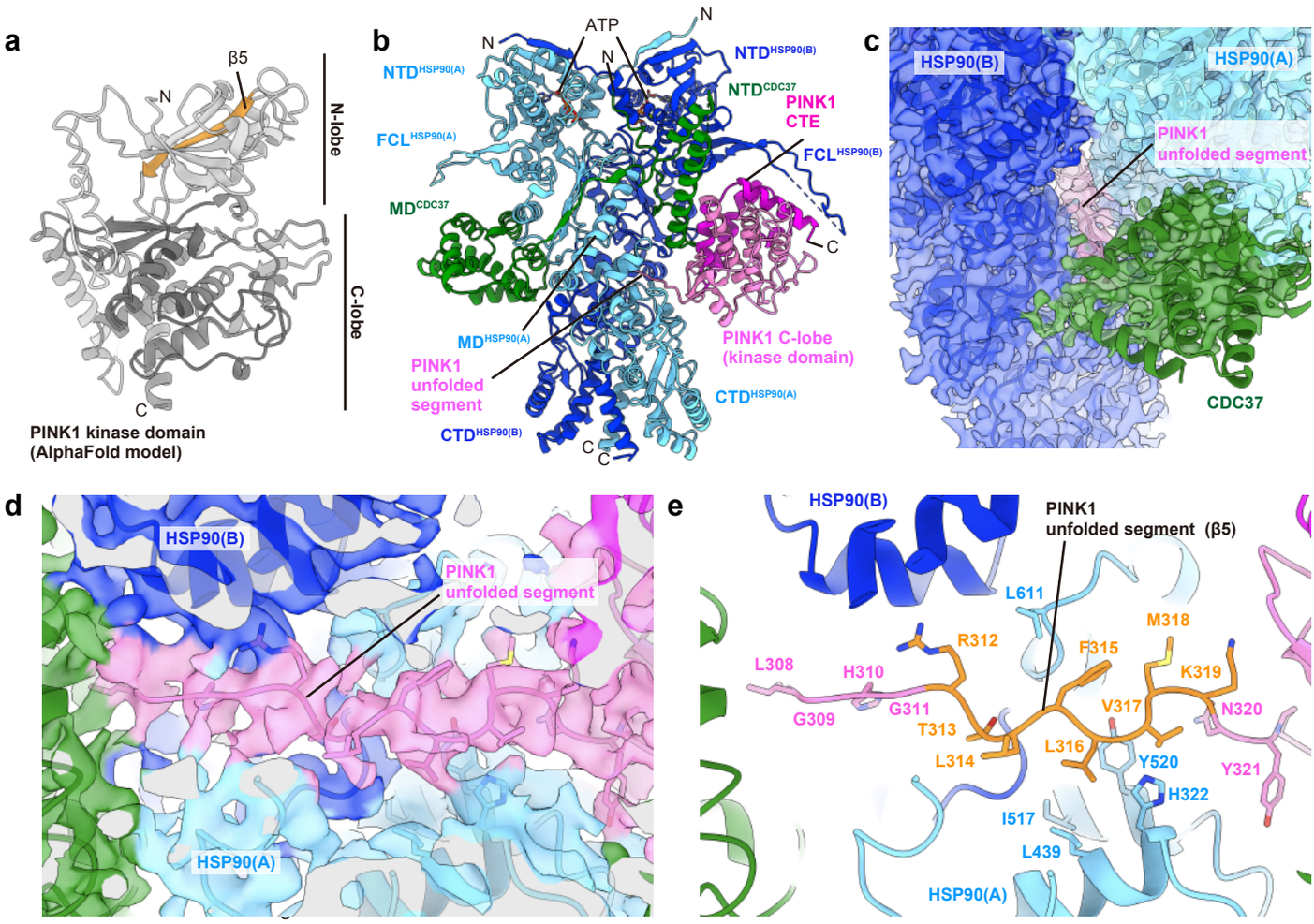
Okatsu et al. Figure 3



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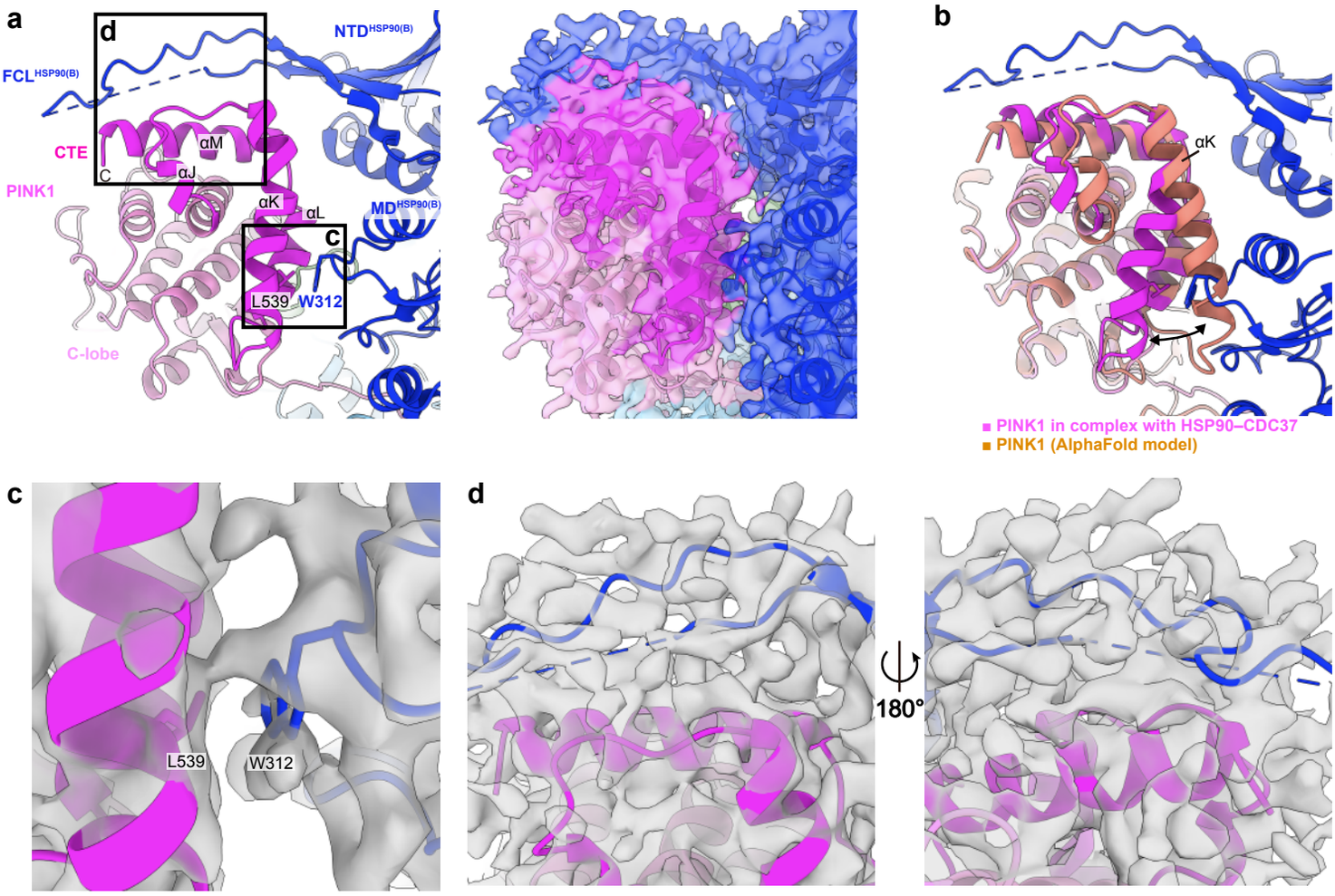

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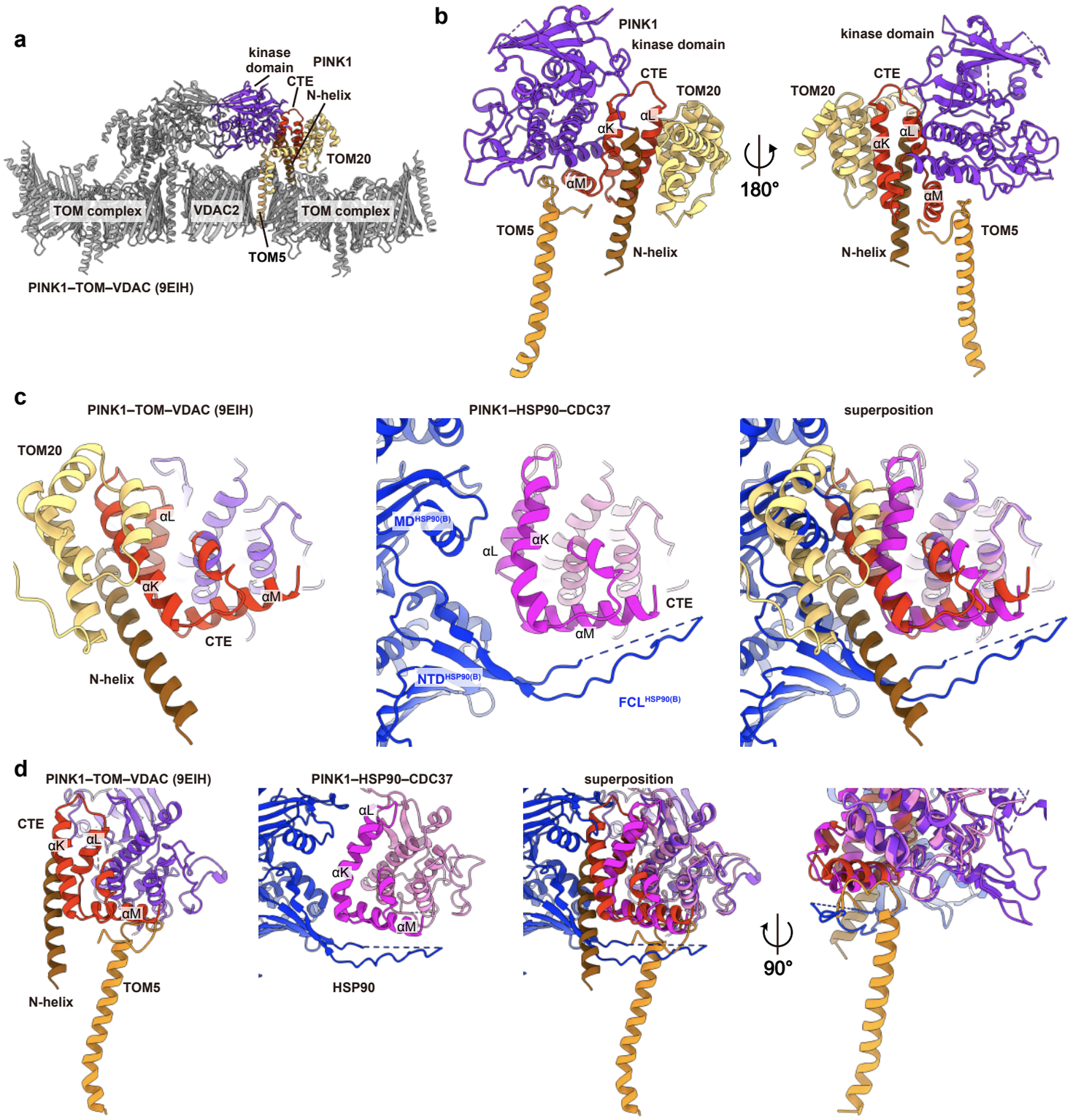

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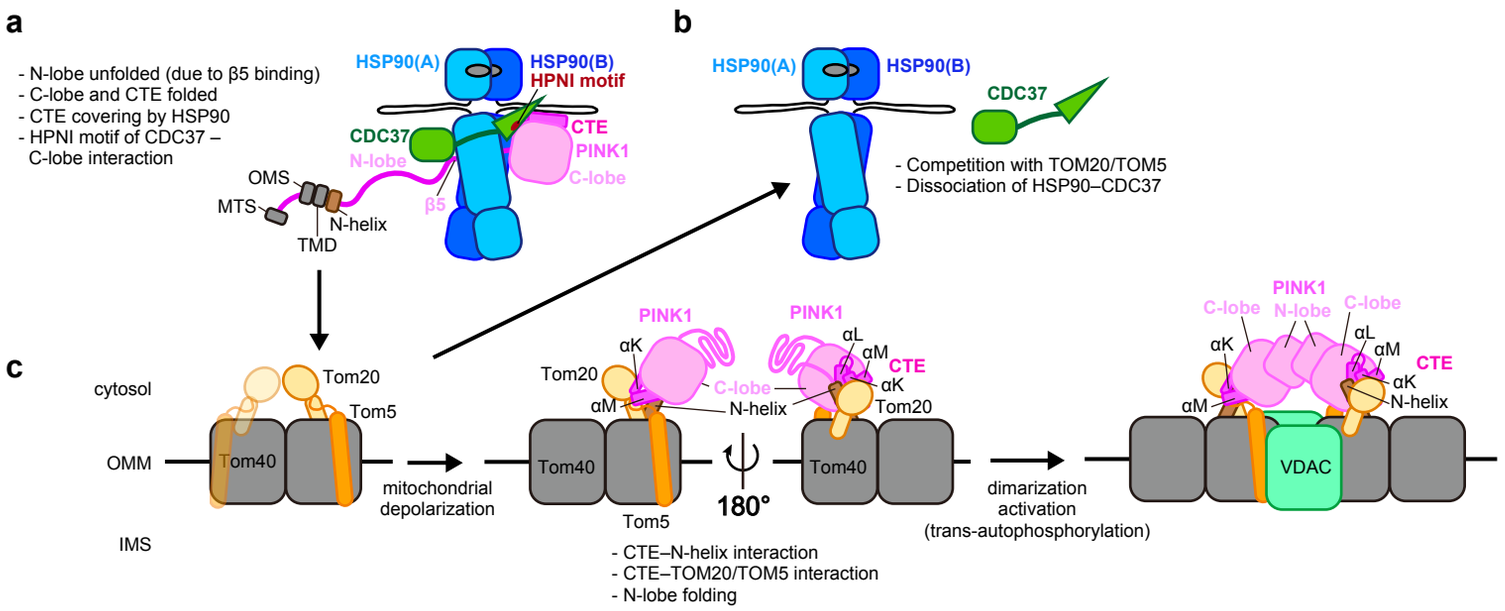

okatsu et al. Figure 7

Table 1

	PINK1–HSP90–CDC37 complex
Data collection and processing	
Magnification	190,000
Voltage (kV)	200
Electron exposure (e^-/Å^2)	50.0
Defocus range (μm)	-0.8 – -1.6
Pixel size (Å)	0.724
Symmetry imposed	C1
Initial particle images (no.)	1,610,071
Final particle images (no.)	454,727
Map resolution (Å)	3.08
FSC threshold	0.143
Refinement	
Initial model used	7Z38, AF-Q9BXM7-F1-model_v4
Model resolution (Å)	
FSC 0.143, unmasked/masked	3.06/3.19
Model composition	
Non-hydrogen atoms	14,094
Protein residues	1,732
Ligand	4
B factors (Å^2), min/max/mean	
Protein	80.30/370.77/177.18
Ligand	88.67/97.64/90.87
RMS deviation	
Bond length (Å)	0.006
Bond angle (°)	0.826
Molprobity score	1.97
Clash score	16.40
Rotamer outliers (%)	0.00
Ramachandran plot	
Favored (%)	96.20
Allowed (%)	3.80
Disallowed (%)	0.00