Demographic insights for coral restoration

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Abstract

Coral reef decline has prompted a global surge in reef restoration initiatives. The success of initiatives that aim to sustain coral populations or assemblages will depend on demographic principles. Restoration strategies generally follow two demographic pathways: additive approaches, which increase population numbers by introducing recruits, fragments, or adults without altering vital rates; and multiplicative approaches, which enhance vital rates–growth, survival, or reproduction–either extrinsically (e.g., herbivory, habitat protection) or intrinsically (e.g., assisted evolution). We synthesized 28 coral matrix population models spanning morphologically diverse coral species from the Caribbean, Hawaii, and the Great Barrier Reef to quantify additive and multiplicative changes needed to increase population growth. Results highlight two consistent demographic leverage points for which population growth was most sensitive:(1) survival of reproductive adults and (2) successful recruitment. Improving adult survival or recruitment by 20% through multiplicative means produced a 5% increase in population growth rate, depending on the fraction of the population affected. By contrast through additive means, the same population growth increases required ~100 recruit outplants or 5–10 adult outplants per 1,000 individuals in a population annually. For large populations typical of restoration targets (10^5–10^7 individuals), this translates to 10^3–10^4 outplants per year—levels rarely logistically or economically feasible. These findings yield two key implications. First, additive interventions are inefficient for large populations and demand sustained, large-scale effort. Second, strategies enhancing vital rates across broad population segments represent the most effective means of boosting coral abundance, including habitat protection, the alleviation of environmental stressors, and interventions which promote long-term survival and recruitment.

Keywords
Restoration, demography, matrix model, coral, populations

Introduction

Coral reef conservation is transitioning from a focus on species and habitat protection to active restoration (Rinkevich 2019). Many scientists are skeptical that coral restoration can change the course of the coral reef crisis (Hughes et al. 2023), in part because of the vast geographical areas of reefs, numerous coral species, diverse life history strategies, and large coral population sizes (McWilliam et al. 2018, Deitzel et al. 2023). However, modeling efforts suggest that some local-scale actions combined with large-scale action on climate change could result in significantly different reef outcomes (Bay et al. 2017, Committee on Interventions to Increase the Resilience of Coral Reefs 2019, McManus et al. 2021). Indeed, the numbers and scales of coral reef restoration initiatives are growing rapidly (e.g., SECORE, Coral Nurture Program, RRAP, MARS, reviewed by Boström-Einarsson et al. 2020; McLeod at al. 2022; Edwards et al. 2024). Governments and nonprofits are spending millions of dollars on research and logistics supporting restoration (Bayraktarov et al. 2019, Tortolero-Langarica et al. 2020, Knapp et al. 2022, Sam et al. 2021, Dela Cruz & Harrison 2020; reviewed by Watt-Pringle et al. 2024; Segaran et al. 2024; Schmidt-Roach et al. 2025). Assessments of restoration practices are generating mixed reviews among ecologists (Boström-Einarsson et al. 2020, McLeod et al. 2022; Madin et al. 2023, Hughes et al. 2023, Streit et al. 2024; Suggett et al. 2024; Watt-Pringle et al. 2024). Meanwhile, corals continue to fare poorly (Hoegh-Guldberg et al. 2023).

Coral population growth or decline can be projected quantitatively as a matrix of transitions among various life history stages (Hughes 1984), or with higher resolution as an Integral Projection Model (e.g., McWilliam et al. 2023). Similar to most plants and animals, colonial reef building corals can be classified simply into three life history stages (Caswell 1989). The first is the recruitment stage, which represents new individuals entering the population by sexual or asexual reproduction (Hughes and Jackson 1981). Recruits tend to have low growth rates (absolute change in colony area) and exceptionally low survival rates (Carlot et al. 2021). Second is the juvenile stage, which is generally considered the point prior to reproductive maturity at which a colony is large enough to reduce post-settlement mortality and growth becomes steady and predictable (Stearns 1989, Doropoulos et al. 2016). Third is the adult stage where growth slows and colonies become reproductively mature. Reproductive output scales with the number of polyps which, in turn, scales with colony size (Hall and Hughes 1996; Padilla-Gamiño & Gates 2012; Álvarez-Noriega et al 2016; Johnston et al. 2020). A complete population matrix describes transitions between each of these stages (including recruitment, growth, shrinkage, and survival, Fig. 1A), and the dominant eigenvalue of the matrix (known as lambda, λ) reflects the intrinsic growth rate of the population, which can be used as a common currency for comparing population dynamics (Edmunds et al. 2014; Cant et al. 2021). Values above one ($\lambda > 1$) translate to population growth, and those below one ($\lambda < 1$) indicate population decline. Lambda is also a metric of population resilience following disturbances, as it reflects the capacity for a population to grow back to pre-disturbance levels (Madin et al. 2012; McWilliam et al. 2023), and is, therefore, a useful tool for guiding conservation decisions (Crowder et al. 1982; Doak et al. 2021; Edmunds et al. 2014; Edmunds & Riegl 2020).

87
Figure 1. The yearly stage transitions for a population of *Acropora cytherea* sensu Veron 2000 at Lizard
Island on the Great Barrier Reef. (**A**) Most cells show probabilities of moving between stages in one year.
The blue cell represents the number of recruits that return to the population per year from a single adult
for the population growth to be at stasis (λ=1.0). The stable size structure is the expected proportions of
each stage when the population has stabilized. (**B**) The transition cell value required to increase the
intrinsic population growth rate to λ=1.05, a 5% increase. Each cell was calculated independently, while
the others remained constant. (Proportional changes are given in parentheses.) Note that cell values
greater than 1 imply the addition of individuals. On the right are the number of individuals and outplant
cover required for each stage to increase λ=1.05 without changing transition probabilities.
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From a demographic perspective, there are two general approaches for increasing a
population's intrinsic growth rate: (1) the "multiplicative approach," which manipulates matrix
transition probabilities and (2) the "additive approach," which artificially increases population
numbers. Table 1 summarizes some common restoration activities and the demographic
approach used. For example, increasing individual growth, survival, reproductive and
recruitment rates are all multiplicative levers for changing long-term population growth rate, if
those increases are sustained through time. Under traditional coral reef management, such
changes have been conducted by improving the environment (e.g., reducing pollution [Vega
Thurber et al. 2013; Lapointe et al. 2019; Gove et al. 2023], and removing competitors
[Ceccarelli et al. 2018] and predators [Pratchett et al. 2017, Raker et al. 2023]). Nevertheless,

as a response to the increasingly negative effects of climate change, the focus has shifted to
more active interventions such as moving corals or gametes between environments (assisted
migration [Vitt et al. 2009; Hagedorn et al. 2021; Barreto et al. 2023]) and selective breeding
(assisted evolution [van Oppen et al. 2015; Humanes et al. 2021; Drury et al. 2022; Bay et al.
2023]), or by increasing population numbers via outplanting of coral larvae, fragments, or whole
colonies (reviewed by Boström-Einarsson et al. 2020; Edwards et al. 2024). Theoretically,
adding new individuals to the population each year can have the same demographic outcome
as altering survival probabilities, because new individuals contribute to successful growth and
reproduction. However, this can be a very costly strategy (Bayraktarov et al. 2019; Hughes et al.
2023), as those numbers must come from outside the population (e.g., a nursery or corals of
opportunity or previously harvested fragments), and be actively deployed every year. A review
of coral restoration in Indonesia found that 84% of projects reported an additive approach (i.e.,
some form of coral transplantation) and 16% relied on natural larval recruitment upon artificial
reefs (Watt-Pringle et al. 2024). The two demographic approaches are not mutually exclusive;
for example, genetically different or selectively bred corals can be added to potentially increase
survival and reproduction (Drury et al. 2022, Bay et al. 2017). Ultimately, demographic
approaches are needed to assess the relative utility of additive versus multiplicative
modifications to population and their effectiveness for reef conservation.
126
127 **Table 1**. Common approaches for coral restoration and conservation and their demographic approach.

Activity	Demographic Approach	Description	Examples
Allee Effect Mitigation	Multiplicative	Increasing density beyond the minimum required for successful fertilization.	Ricardo et al. 2025
Artificial habitat	Multiplicative	Deployment of artificial structures for attracting coral and other biodiversity with a focus on the recruitment stage.	Knoester et al. 2023; Reichert et al. 2024; Tanaya et al. 2025
Assisted evolution	Multiplicative	A range of interventions that speed up the natural process of coral adaptation that might improve growth and survival.	Van Oppen et al. 2015; Caruso et al. 2021; Drury et al. 2022
Assisted migration	Multiplicative	The delivery of corals with environmental tolerances to another based on anticipated environmental changes.	Van Oppen et al. 2014; Caruso et al. 2021
BioRock Electric Reef	Additive / Multiplicative	Similar to substrate stabilisation, but with the addition of low voltage electricity applied to metal mesh to promote calcification	Goreau & Prong 2017; Goreau 2022
Coral gardening	Additive	The asexual propagation of corals onto a reef typically following a nursery phase.	Page et al. 2018; Tortolero-Langracia et al. 2020; Knapp et al. 2022
Coral repositioning	Additive	Securing dislodged corals back onto the reef. In some cases, corals will be fragmented before being	Garrison & Ward 2008; Nava & Figueroa-

		repositioned. Like coral gardening and transplantation, but without a nursery phase.	Camacho 2017; Sam et al. 2021
Coral seeding	Additive	The settlement of coral larvae or attachment of small fragments onto structures for reef deployment.	Forsman et al. 2015; Lindal 2003; Suzuki et al. 2011
Coral superfoods / probiotics	Additive Multiplicative (if sustained)	Administering nutrients or supplementing coral microbiomes to boost survival and growth. Note that this intervention must be sustained to be multiplicative.	Doering et al. 2021; Peixoto et al. 2017
Coral Transplantation	Additive / Subtractive	Like assisted migration or coral seeding, corals are moved from one place to another, but environmental tolerances are not considered, and adult colonies are generally used. Supplementing existing populations with individuals destined for mortality due to development, without a nursery phase. An umbrella term that can also include coral gardening and repositioning.	Lindal 2003; Kilbane et al. 2008; Rogers et al. 2017; Forrester et al. 2019; Kikuzawa et al. 2021
Early maturity	Multiplicative	Selecting for maturity at smaller sizes.	Randall et al. 2020
Herbivore fishing restrictions	Multiplicative	Decreasing competition for space on the reef and improving recruitment, growth and survival of coral. However, recruitment can be hindered by herbivores in the absence of habitat complexity.	Penin et al. 2011; Bonaldo et al. 2017 Morais & Bellwood 2018; Strain et al. 2018
Larval-based restoration	Additive	Supplying large numbers of coral larvae to damaged reef areas.	Harrison et al. 2021
Macroalgae removal	Multiplicative / Additive	Suppress macroalgae proliferation to facilitate coral recovery. In some cases, paired with urchin releases.	Neilson et al. 2018; Smith et al. 2021
Pollution & sedimentation mitigation	Multiplicative	Generally improving the environment improves all demographic rates.	Donovan et al. 2021; Donovan et al. 2020; Gove et al. 2023
Spawn capture and rearing	Additive	Collecting reproductive gametes during spawning events for culturing.	Dela Cruz & Harrison 2020
Substrate stabilization	Multiplicative	Laying an artificial substrate (e.g., mesh) over areas of unconsolidated reef rubble.	Fox et al. 2019; Chesney & Richards 2022; Ceccarelli et al. 2020
Urchin releases	Multiplicative	Like herbivore fishing restrictions, but potentially more disruptive to coral recruitment.	Neilson et al. 2018; Barrows et al. 2023

Optimal restoration approaches will likely differ among geographic locations and coral taxa with
different sensitivities to demographic change across their life cycle. Coral species exhibit an
array of life history strategies because of energetic trade-offs among growth, survival and
reproductive rates (Edmunds et al. 2014, McWilliam et al. 2023; Stearns 1989). Indeed, this
array of strategies supports coexistence among species (Chesson 2000) and is essential for
reef ecosystem function (Bellwood et al. 2004). For corals, major morphological groups (or
growth forms) tend to reflect life history strategies (Jackson 1982; McWilliam et al. 2023;
Dornelas et al. 2017). For instance, branching and tabular coral growth forms are specialised to
compete for reef space via early and rapid growth (Baird & Hughes 2000), but typically have
lower survival rates across life stages (Madin et al. 2016). Reproductive output varies
dramatically among coral taxa, yet tends to be similar within growth forms, and highest in taxa
which reach large colony sizes (Hall and Hughes 1996; Álvarez-Noriega 2016). Identifying life
stages in which lambda is sensitive to change in specific vital rates is therefore necessary to
design suitable conservation and restoration plans, acknowledging that these may vary across
species.
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In this study, we use demographic modeling to explore how changes to population structures
can shift a population from decline to recovery for the purposes of coral conservation. We
collated existing coral matrix population models capturing a range of geographic locations and
coral morphologies (Table 2) to address a question that all reef resource managers and
restoration practitioners should ask: what changes in transition probabilities or outplant numbers
are needed to increase a population's yearly growth by a given amount? We explicitly compare
two approaches to reef conservation, the additive approach in which colonies are outplanted,
and the multiplicative approach in which vital rates are modified, quantifying the relative effort
needed for each to affect population growth rates. We also examine the different life stages of
coral in which changes in vital rates or abundances have the largest effects on population
growth across different taxa. Our study highlights that demographic approaches are needed to
inform decisions regarding the allocation of effort when attempting to preserve and restore coral
populations.
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159   **Methods**
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We collated 28 projection transition matrices, representing recruit, juvenile, and adult life history
stages, spanning 22 species from six growth forms of reef-building corals (Table 2). Of these
matrices, 11 were already three-by-three stage matrices representing recruits, juveniles and
adults from the Comadre database (Salguero-Gómez et al. 2016). Only complete matrices that
captured unmanipulated population growth were included from Comadre. Another 11 species
matrices were generated from the Trimodal dataset (Lizard Island, Great Barrier Reef; Madin et
al. 2023) by transposing yearly colony growth and survival data into three-by-three matrices
using the stage size classifications in Table 2 (see Supporting Methods, Fig. S1 and Fig. S2).
Six more species matrices were generated from NOAA's Coral Reef Conservation Program-
funded Vital Rate Project across the Hawaiian Archipelago (Rodriguez et al 2022). For those
matrices that began as larger than three-by-three, we selected transition sizes from recruit to

juvenile according to the observed decline in mortality rates (Fig. S1), and from juvenile to adult
according to estimated size of first reproduction (Fig. S2).
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175   **Table 2**. A summary of the 28 matrices by species. Each life history stage shows the range of planar
176   areas for each stage. In parentheses are the areas used to estimate population areas, which are roughly
177   the midpoints of stages. [a]Mercado-Molina et al. (2015); [b]Madin et al. 2023; [c]Hughes & Tanner 2000;
178   [d]Rodriguez et al 2022; [e]Hernández-Pacheco et al. 2011. Growth form was collected from the Coral Trait
179   Database (Madin et al. 2016).

Species	Region	Matrices	Recruit (cm^2)	Juvenile (cm^2)	Adult (cm^2)	Growth form
Acropora cervicornis [a]	Carribean	2	0-100 (50)	100-1000 (550)	>1000 (1500)	Arborescent
Acropora cf. digitifera [b]	GBR	1	0-150 (75)	150-300 (225)	>300 (375)	Digitate
Acropora cytherea [b]	GBR	1	0-500 (250)	500-2000 (1250)	>2000 (3000)	Tabular
Acropora humilis [b]	GBR	1	0-200 (100)	200-300 (250)	>300 (350)	Digitate
Acropora hyacinthus [b]	GBR	1	0-200 (100)	200-1000 (600)	>1000 (1400)	Tabular
Acropora intermedia [b]	GBR	1	0-150 (75)	150-500 (325)	>500 (675)	Arborescent
Acropora millepora [b]	GBR	1	0-150 (75)	150-300 (225)	>300 (375)	Corymbose
Acropora nasuta [b]	GBR	1	0-150 (75)	150-300 (225)	>300 (375)	Corymbose
Acropora robusta [b]	GBR	1	0-200 (100)	200-500 (350)	>500 (650)	Arborescent
Acropora spathulata [b]	GBR	1	0-150 (75)	150-700 (425)	>700 (1050)	Corymbose
Agaricia agaricites [c]	Carribean	2	0-20 (10)	20-100 (60)	>100 (150)	Laminar
Goniastrea pectinata [b]	GBR	1	0-20 (10)	20-200 (110)	>200 (300)	Massive
Goniastrea retiformis [b]	GBR	1	0-20 (10)	20-200 (110)	>200 (300)	Massive
Helioseris cucullata [c]	Carribean	2	0-20 (10)	20-100 (60)	>100 (150)	Laminar
Montipora capitata [d]	*Hawaii*	1	0-20 (10)	20-100 (60)	>100 (150)	Laminar

Species	Region	Matrices	Recruit (cm^2)	Juvenile (cm^2)	Adult (cm^2)	Growth form
Montipora patula [d]	*Hawaii*	1	0-20 (10)	20-100 (60)	>100 (150)	Laminar
Orbicella annularis [b,e]	Carribean	4	0-50 (25)	50-150 (100)	>150 (225)	Massive
Pocillopora meandrina [d]	*Hawaii*	1	0-150 (75)	150-700 (425)	>700 (1050)	Corymbose
Porites astreoides [a]	Carribean	1	0-50 (25)	50-150 (100)	>150 (225)	Massive
Porites lichen [d]	*Hawaii*	1	0-50 (25)	50-150 (100)	>150 (225)	Massive
Porites lobata [d]	*Hawaii*	1	0-50 (25)	50-150 (100)	>150 (225)	Massive
Porites lutea [d]	*Hawaii*	1	0-50 (25)	50-150 (100)	>150 (225)	Massive

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To compare the demographic effects of restoration across coral taxa, we assumed that all
populations were at stasis when matrix data were collected—neither growing nor declining. To
do this, we modified the recruitment rates of taxa (which were not recorded in field studies) and
used a least squares function and the *optimize* function in R to estimate the amount of
recruitment generated by adults that would result in a population with λ=1 (R Core Team 2025;
Fig. S3A). Estimated levels of recruitment per adult colony required to make λ=1 were
consistent with other levels reported in the literature (Madin et al. 2012; McWilliam et al. 2023).
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We used a least squares function to estimate transition cell changes (multiplicative) and
outplant numbers (additive) required to achieve population growth increases of 1, 2, 5 and 10%.
For the multiplicative approach, we increased a given transition cell value, while holding all other
cells constant, until attaining levels of λ=1.01, 1.02, 1.05 or 1.1 (e.g., Fig. S3A). Cell values were
allowed to increase above 1, which is theoretically the maximum transition probability for all
cells that do not reproduce. However, we only report changes for the species for which cell
values reflecting transition probabilities were one or less. We did not include the calculated
changes for "shrinkage" cells, as the perturbation resulted in impossibly large values. That is,
increasing population growth rate by elevating transitions from a large stage to a smaller stage
can only occur when total survival probability in the resulting stage is above one, which can only
be accomplished by adding individuals to the population, such as by reproduction.
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For the additive approach, we increased each stable size structure (SS) value, while holding all
others constant, until the summed matrix multiplication of the baseline transition matrix and the
SS reached 1.01, 1.02, 1.05 or 1.1 (e.g., Fig. S3B). The amount of SS increase for a stage was
multiplied by 1000 to express the number of outplants required per 1,000 individuals in the
population. We also present these numbers in terms of the outplanted coral area per year (i.e.,
the planar area of coral required from an outside source), which was done by multiplying the

number of individuals in each stage by the midpoint stage sizes in Table 2. We multiplied these
values by 10,000 to express as outplant area per hectare (10,000 m^2) of population cover. Note
that additive metrics only reflect the levels required in the first year; that is, proportionally more
additions are required as the population size increases.
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We estimated typical coral population sizes and population domain areas (i.e., the total area of
living coral in a population) using NOAA's National Coral Reef Monitoring Program (NCRMP)
data on coral colony density across the US-affiliated central Pacific. The NCRMP effort includes
a stratified random survey of reef sites across 37 islands which enumerates coral colonies,
returning colony-level taxonomic identification, colony density, and survey sector reef area
(Towle et al. 2022). From this data set, we highlight 80 taxa that represent the most-robust
species-level identifications present in the dataset and then relied on the survey design
estimates of sector-level colony number; that is, the estimated census population size at the
sector scale (Smith et al 2011; Winston et al 2019). The NCRMP survey design assesses adult
colony density in three depth-dependent strata per sector (0-6 m, 6-18 m, 18-30 m), so sector-
level abundance (population size) for each representative species was calculated as the product
of strata-level mean colony density and stratum area, summed across the three strata in each
sector (Smith et al 2011; Winston et al 2019). Here, the sector represents roughly 10s of km of
coastline (i.e., island scale for small islands [e.g., Jarvis, Pagan]) or side-of-island scale for
larger islands (e.g., Oʻahu, Guam). Previous work on the modeled distribution of juvenile colony
density, show an apparent stock-recruitment at sector scale, which suggests this scale is a
reasonable proxy for a demographically coupled population (Couch et al 2023). While we are
presenting projection matrices from three regions (Caribbean, Australia, Central US-Pacific), we
only present the estimates of population sizes from the US Pacific.
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232 **Results**
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Across the 34 population matrices, yearly survival probabilities tended to increase as colonies
got larger (Fig. 2). Growth probabilities declined slightly with increasing size, and growth from
recruit to adult (skipping the juvenile phase) had the lowest probability, and it was only observed
in some massive and laminar species populations. Shrinkage (partial mortality or fission) to
smaller stages tended to occur more for the adult stage. By way of example, Fig. 1A shows the
matrix for *Acropora cytherea* and the blue transition cell represents the estimated number of
successful recruits generated by an average adult colony per year; in this case, for the
population to remain in stasis, only 8-9 of the hundreds of thousands of larvae generated by an
adult colony must recruit back into the population. The green cells show probabilities of
surviving and growing to larger stages; yellow cells show probabilities of surviving and
remaining at the same stage; and red cells show surviving and shrinking via partial mortality or
fission to smaller stages. The vector to the right of the transition matrix in Fig. 1A is the
population's stable size structure (i.e., the transition matrix's eigenvector): the proportion of
colonies at each stage once the population has stabilized (i.e., 76% of the colonies would be
recruits, 17% juveniles and 7% adults; Caswell 1981).
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Figure 2. Boxplots for transition cells values for the 34 species matrices. The color scheme reflects that of
Fig. 1 with survival probabilities in yellow, growth and survival probabilities in green, shrinkage in red, and
contribution of adults to recruitment in blue.

Demographic models captured the changes in colony numbers and transition values needed to
shift populations from stasis to growth. Using *Acropora cytherea* as an example, we find that
increasing population size by 5% (equivalent to increasing λ to 1.05) would require outplanting
of either 140 recruits, 64 juvenile or 5 adult colonies per 1000 colonies in a population per year
(Fig. 1B). Similarly, for a summed planar area of 100 m², we find that 3% of this area (3 m²)
would need to be outplanted as new adult corals each year to increase population growth rate
by 5%. Taking a multiplicative approach, a 5% increase in λ can be achieved by increasing
recruit survivorship from 0.25 to 0.6 (assuming all other transition probabilities remain the same;
Fig. 1B), equivalent to a 2.4-fold increase in recruit survival rates (parentheses, Figure 1B). On
the other end of the life-stage spectrum, increasing survival of adult colonies from 57% to 78%
achieves a 5% increase in population growth, equivalent to a 1.36-fold increase in adult survival.
Finally, 5% increases in population growth of *A. cytherea* can be achieved by increasing the
survival of juvenile colonies that grow into adults and the probability of recruitment; each of
these requiring an approximately 1.7-fold change.

Changes in transition values needed to achieve 5% increase in population growth varied across
the 34 matrices in the analysis, though some consistencies emerged across different life
histories and biogeographic regions (Fig. 3). Species for which required transition values
became greater than one (Fig. 3A, red points) indicated that the multiplicative approach was not
a variable option. For instance, improving adult survivorship in order to increase population
growth rate by 5% was only viable for 79% of our populations. Considering only the population
for which a strategy is possible, two transition cells—adult survival and recruitment—required
the least change in value to achieve a 5% increase in population growth (Fig. 3B), which were
also reflected in matrix elasticities for adult survival and recruitment that quantify the relative
sensitivity of population growth to the various transitions (Fig. S4; Caswell 1989). Increases in
successful recruitment or adult survival present the most consistent, demographically efficient
means to increase population growth, with the caveat that the adult survival approach is only
viable for 27 of the 34 populations. The small changes in recruitment and adult survival needed

to boost population growth were largely consistent across coral growth forms (Fig. 3C).
Changes required for other transition cells were higher and more variable, reflecting the
sensitivity of growth forms to different changes in demographic rates. For example, the low
growth in massive taxa and the low juvenile survival in laminar taxa required higher necessary
changes in transition probability to boost population growth (Fig 3c; Fig S5). Meanwhile, higher
recruit survival in massive taxa meant that smaller shifts in recruit survival were needed relative
to other growth forms (Fig 3; Fig S5). The small changes in recruitment and adult survival
needed to boost population growth were also consistent across biogeographic regions (Figure
3D). With the exception that the Caribbean showed greater efficacy (lower transition multiplier)
of elevated recruitment survival for some species, but not all as reflected by high levels of
variation.

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Figure 3. The multiplicative approach. (**A**) The cell values required to increase population growth rate by
5% (λ=1.05) summarized for all species. Transition values cannot exceed one for cells that do not
generate individuals (red points) unless individuals are artificially added. The percent of species for which
changing cell values is possible are given. (**B**) The proportional change in cell values required to increase
population growth rate by 5%, only for species for which the approach is possible. Proportional change is
also shown for (**C**) species growth form and (**D**) biogeographic region. The color scheme reflects that of
Fig. 1 with survival probabilities in yellow, growth and survival probabilities in green, and contribution of
adults to recruitment in blue.
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Increasing population growth by adding numbers required approximately 100 recruits or
juveniles per 1000 colonies in the population in the first year (Fig. 4A, left), or 5–10 adults; and
this number would increase proportionally as the population size increased. These numerical
additions translate into similar amounts of live coral area for recruits and adults of approximately
2–5 m^2 yearly for every 100 m^2 of adult coral area in a population (Fig. 4A, right). In contrast,

juveniles require approximately an order of magnitude more coral material since they require
large numbers of corals (comparable with recruits) but with substantially increased areas of 20–
500 cm^2 (Table 2). These results were consistent among growth forms, with notable deviations
for massive and laminar growth forms. For example, significantly more recruits or juveniles
would be required relative to adults for the laminar populations due to low early survival rates,
suggesting that a strategic focus on adults may more efficiently elevate population growth.
Conversely, fewer recruits would be required to elevate population growth for massive species
relative to other growth forms, likely because of higher rates of recruit survival (Fig. 2). Desired
levels of population growth rate scale approximately linearly with the amount of coral material
added to the population each year (Fig. 4B). For instance, for a population size of 1000
individuals, adding one adult per year is estimated to increase population growth by 1% and
adding 10 adults would increase it by 10%.
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Figure 4. The numbers approach. (**A**) Boxplots of the number of outplants (left) and outplant area (right) for each life history stage required to increase yearly population growth rate by 5% (λ=1.05), summarised by growth form. (**B**) Boxplots of the number of outplants per 1000 individuals (left) and outplant area per 100 m² of population area (right) for each life history stage required to increase yearly population growth rate by 1, 2, 5, and 10% for all species.

Both additive and multiplicative restoration approaches depend on the scale of implementation
(Fig. 5). The number of yearly adults that need to be outplanted to achieve a 5% increase in
population growth differs by two orders of magnitude across species (Fig. 5B). Assuming that
sector-scale population sizes seen across the US-affiliated Pacific are reasonable estimates for
other populations, and based on mean estimates of census population size (~625,000
individuals, Fig. 5A), a minimum of 26 (*Acroopora millepora*) and a maximum of 28,000
(*Montastrea annularis*) adult colony outplants per year are necessary to alter the dynamics of a
population. Orders of magnitude greater numbers of additional recruits are necessary to alter
population growth by 5%, with a minimum of 33,000 and a maximum of 630,000 recruits added
to the natural rates every year, based on the average population size in the dataset (Fig. 5C).
Such numbers highlight the difficulty of altering population growth at large (regional) scales
using the additive approach.

341
Figure 5. (**A**) Population abundance for 80 species estimated from NOAA NCRMP Pacific benthic survey
data. Comparison of approaches relative to species population sizes. (**B**) The number of yearly adult

outplants required to increase population growth rate by 5%, with each colored line representing one of
the 34 examined transition matrices. The species at either end of the range are labeled. (**C**) The number
of yearly recruit-sized outplants needed to increase population growth by 5% for each of the 34 matrices.
(**D**) Population-scale reef area estimates from NOAA Pacific benthic survey data. (**E**) The change in adult
survival probability needed to increase population growth rate by 5% when considering the proportion of
the population affected by a conservation measure for each of the 34 matrices. Points indicate the
theoretical limit; that is, where adult survival probability is 100%. (**F**) The change in recruitment probability
needed to increase population growth rate by 5% for each of the 34 matrices.
352
Elevating adult survival probability is also likely to be most effective if the benefit can be
implemented across the entire population (Fig. 5D). However, when we model a realistic
intervention that does not benefit the entire coral population, we can show that elevated vital
rates required to meet a 5% shift in population growth rate (lambda) are relatively stable when
only subsets of the population are affected (Fig. 5D-E). Indeed, for many taxa the necessary
improvement to survival remains stable (< 1.5-fold increase) as long as > 50% of the population
is affected (Fig. 5E). Similarly, a five-fold increase in recruitment was sufficient to shift lambda to
1.05 for all taxa as long as > 25% of the population was affected by the changes (Fig. 5F).
Nevertheless, the number of additional recruits required becomes exceedingly high if small
portions of the population are impacted (Fig 5F). Moreover, we identify a critical threshold in
survival beyond which a 5% change in lambda is theoretically impossible because survival has
already reached 100% (points in Fig. 5E). For many taxa, this threshold occurs between 25-
50% of the population affected, demonstrating the importance of capturing large proportions of
the population to achieve meaningful shifts in lambda, a feat that requires conservation action to
be implemented across large areas (Fig 5D).
368
369   **Discussion**
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Our analysis suggests that modifying vital rates at critical stages of the life cycle is the most
effective way to grow coral populations. This "multiplicative" approach focuses on modifying
factors that influence survival, growth and recruitment, and shows considerable promise if it can
be applied over large portions of the effective population (Fig. 5D-F). A wide range of
conservation measures can be employed to modify vital rates (Table 1), yet demographic
models suggest that those which affect adult survival and recruitment rates are likely to be the
most successful because they require the least change. If applied to the whole population, a
1.2-fold increase in survival (20% increase) or a 1.5-fold increase in recruitment (50%, on
average) was sufficient to boost population growth by 5%. In contrast, relying solely on
outplanting additional colonies to modify coral population growth rate—the additive approach—
is likely to be inefficient for most species, particularly those with high abundances (Fig. 5A-C).
While it was possible to boost population growth by adding colonies, we find that yearly
additions of 5-10% of the existing population size in recruit form, or 0.5-1% of the existing
population size in adult form, were necessary to alter population trends (Fig. 4). Genetic models
suggest that a similar level of adult transplantation (2-5% of the population per year) could
prevent extinction, but not population decline, under moderate emissions scenarios (formerly
RCP6.0) only if transplanted corals were pre-adapted to warming conditions (Bay et al. 2017).

Depending on the population size, the number of new colonies needed can be exceedingly
large (Fig. 5A-C).
390
The population size affected by a conservation initiative strongly influences both the
multiplicative and additive approaches, which ultimately depend on the spatial scale of
conservation. At the likely scale of demographically connected populations (i.e. stock-
recruitment scale) we estimate the census population sizes of most coral species to be in the
range of 10^5 to 10^7 individuals (Fig. 5A). These island-scale values are lower than many Pacific
coral population estimates, as estimates that span full species ranges suggest that roughly two-
thirds of species have population sizes exceeding 100 million colonies and one-fifth exceeding a
billion colonies (Dietzel et al. 2021). Quantifying population growth at this more local scale is
challenging, and more precise estimates would require information on dispersal dynamics and
spatial differences in vital rates (Hunter & Caswell et al. 2005). Nevertheless, our simplified
extrapolation of population growth suggests that outplanting as few as 500 adult colonies per
year for rare species, or as many as 50,000 for common species, was necessary for a 5%
increase in population growth at the scale of the NCRMP sectors (Fig. 5B-C). This result
demonstrates the challenges of this strategy for typical coral population sizes unless a
substantial amount of coral material is available, or the population is small or limited to an area
with high levels of self-recruitment, like an embayment. In contrast, multiplicative reef
conservation measures, which focus on enhancing vital rates, can often effectively capture large
components of a coral population. For example, networks of Marine Protected Areas can cover
a large geographic space (> 1000 hectares; Selig & Bruno 2010; Smith et al. 2025) and thus
boost coral vital rates (e.g., improved herbivory and coral recruitment) across a large portion of
a coral population. Similarly, reducing greenhouse gas emissions is likely to benefit most, if not
all, of a coral population by reducing mortality from climate stressors, such as recurrent
bleaching (Hughes et al. 2023; Jury et al. 2024).
414
For local interventions, multiplicative approaches also offer the greatest chance of boosting
coral population growth at the site or reef scale, especially if they limit the need for costly yearly
intervention. For example, artificial structures can boost local recruitment by providing substrate,
modifying water flow, and enhancing settlement probabilities (Nguyen et al. 2022; Higgins et al.
2022; Fig. 6). Our calculations suggest that if a small fraction of the incoming recruits
experience a 10-fold or more increase in settlement success (as has been shown possible, Fig.
6), this is more than sufficient to boost local population growth by 5%. Coral populations on
reefs can still be vast (e.g., a 1-hectare reef with one coral per m^2 has a population of 10,000),
thus requiring hundreds of new recruits and multiple adult colonies to be outplanted every year
to modify population growth, even at a small scale. Boosting local population growth is
fundamentally dependent on recruitment, which is highly variable over time and space owing to
the open nature of coral populations. Understanding this variability and source-sink
metapopulation dynamics will ultimately be crucial for a demographically informed approach to
coral conservation (Crowder et al. 2000; Hansen 2011).
429
We find two critical demographic transitions whereby small changes had strong effects on
population growth. First, adult survival universally required the least change in probability to

increase population growth. This result was expected, given that corals produce many offspring
that mostly perish (Type III survivorship; Crowder et al. 1982; Tsounis & Edmunds 2016). For
example, a 20% increase in average adult survival can enhance population growth by 5% (Fig.
2B), which is remarkably consistent across growth forms (Fig. 2C). Past studies suggest that
focusing on adult survivorship provides the most efficient use of resources, delivering significant
population benefits for smaller interventions. For instance, protecting mature female turtles from
trawling led to population increases in the endangered loggerhead turtle (Crouse et al. 1987).
However, our analysis suggests that this approach is not viable for all species populations, with
21% of the populations requiring increases in survival surpassing 100% (Fig. 3A;
Supplementary Table 1). Furthermore, increasing adult survivorship across large areas for
sessile corals is complex, and would typically involve addressing environmental stressors like
thermal anomalies or mitigating local stressors (Ateweberhan et al. 2013; Good & Bahr 2021;
Donovan et al. 2021; Gove et al. 2023). Increasing adult survivorship is also an important focus
of many restoration strategies (Table 2), including substrate stabilization (Fox et al. 2019;
Chesney & Richards 2022; Ceccarelli et al. 2020), macroalgal removal (Neilson et al. 2018;
Smith et al. 2021), artificial evolution (Van Oppen et al. 2015; Caruso et al. 2021; Drury et al.
2022), and artificial migration (Van Oppen et al. 2014; Caruso et al. 2021).
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Increasing recruitment success was the second leverage point for enhancing population
growth(Fig. 3B). Despite the impressive fecundity of corals, recruitment and post-recruitment
processes represent possibly the largest bottleneck for coral population growth (Arnold et al.
2010; Ritson-Williams et al. 2009; Tsounis & Edmunds 2016; Doropoulos et al. 2021).We find
that population growth can be consistently increased by 5% with a less-than 2-fold improvement
in recruitment. Recruitment operates differently to other life stage transitions, because it is a
multiplier of population numbers, rather than a probability of transition for a single individual, and
so multiplicative changes are not bounded by one and are potentially easier to manipulate via
management actions (Randall et al. 2020, Harrison et al. 2021). Given the large potential for
larvae supply, capturing just a small fraction of wasted larvae has extraordinary potential for
scalable restoration. Spawn capture and nursery-rearing of recruits is promising (Dela Cruz &
Harrison 2020), but still faces the same limitations as coral gardening, relying on consistent
outplanting to maintain its efficacy (Table 1). However, improving the passive capture of even a
small fraction of wasted larvae has extraordinary potential for restoration. For instance, by
conservatively improving settlement by just 2-fold, population growth could be increased by
10%. Reef managers could achieve such improvements by enhancing environmental
conditions, such as reducing sedimentation or contaminants (Nalley et al. 2021), promoting
herbivorous fishes and urchins, restoring suitable habitats, stabilizing unconsolidated hard
substrate, or deploying settlement substrates for coral larvae (Table 1). For instance, data from
settlement structures with cryptic recesses routinely demonstrate improvements of 50-200-fold
over natural reefs (Doropoulos et al 2016, Reichert et al.2025; Fig. 6). While density dependent
processes (i.e., crowding and competition) act to thin these early life gains, the chances that
increased numbers of juveniles make it through the early life bottleneck increases, sometimes
dramatically (e.g., 20-50-fold after one year; Reichert et al 2025).

Figure 6. The density of coral recruits on natural reefs (Reichert et al. 2025), traditional settlement tiles
(Bramanti and Edmunds 2016; Edmunds 2022; Brown 2004; Friedlander and Brown 2006), and
structures designed with cryptic recesses (Doropoulos et al 2016; Reichert et al. 2025). See supplemental
methods for details.
480
This paper considered various approaches to coral restoration from demographic first principles.
The collation of simple population matrix models, and a straightforward analysis looking at the
influence of multiplicative and additive changes for coral populations, demonstrates the salient
features of current restoration approaches and their likely success. There are contexts in which
"additive" restoration at achievable scales can improve population outcomes, especially in small,
restricted populations or those experiencing Allee effects. However, in many coral populations,
raising population growth rates above replacement by simply outplanting more colonies into
declining populations will require efforts at scales unlikely to be cost-effective. On the other
hand, our demographic analysis suggests that "multiplicative" efforts to the rates of survivorship
and/or recruitment through combinations of traditional management and process-focused active
interventions are likely the more effective strategy for responding to declining coral
populations.Every coral conservation effort is unique and context-dependent, but we hope our
demographic framework will provide resource managers and restoration practitioners a
framework to understand and predict how restoration efforts will translate into meaningful
outcomes.
496
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498
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501
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503
504 Data and code are available at https://github.com/jmadinlab/demographic-insights
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References

Álvarez-Noriega M, Baird AH, Dornelas M, Madin JS, Cumbo VR, Connolly SR (2016) Fecundity and the demographic strategies of coral morphologies. Ecology 97:3485–3493

Amir C (2023) Measuring Coral Vital Rates using TagLab Semi-automatic Coral Annotation and Temporal Linking across Fixed Sites: Standard Operating Procedures and Time Savings Estimate.

Barreto MM, Schmidt-Roach S, Zhong H, Aranda M (2023) Assessing the feasibility of assisted migration of corals in the Red Sea. Front Mar Sci 10:1181456

Baums IB, Baker AC, Davies SW, Grottoli AG, Kenkel CD, Kitchen SA, Kuffner IB, LaJeunesse TC, Matz MV, Miller MW, Parkinson JE, Shantz AA (2019) Considerations for maximizing the adaptive potential of restored coral populations in the western Atlantic. Ecological Applications 29:e01978

Bay RA, Rose NH, Logan CA, Palumbi SR (2017) Genomic models predict successful coral adaptation if future ocean warming rates are reduced. Sci Adv 3:e1701413

Bay et al. (2023). CORDAP R&D Technology Roadmap for Understanding the Natural Adaptation and Assisted Evolution of Corals to Climate Change. CORDAP. cordap.org

Bayraktarov E, Stewart-Sinclair PJ, Brisbane S, Boström-Einarsson L, Saunders MI, Lovelock CE, Possingham HP, Mumby PJ, Wilson KA (2019a) Motivations, success, and cost of coral reef restoration. Restor Ecol 27:981–991

Bayraktarov E, Stewart-Sinclair PJ, Brisbane S, Boström-Einarsson L, Saunders MI, Lovelock CE, Possingham HP, Mumby PJ, Wilson KA (2019b) Motivations, success, and cost of coral reef restoration. Restoration Ecology 27:981–991

Boström-Einarsson L, Babcock RC, Bayraktarov E, Ceccarelli D, Cook N, Ferse SCA, Hancock B, Harrison P, Hein M, Shaver E, Smith A, Suggett D, Stewart-Sinclair PJ, Vardi T, McLeod IM (2020) Coral restoration – A systematic review of current methods, successes, failures and future directions. PLoS ONE 15:e0226631

Bramanti L, Edmunds PJ (2016) Density-associated recruitment mediates coral population dynamics on a coral reef. Coral Reefs 35:543–553

Brown EK (2004) Reef coral populations: Spatial and temporal differences observed at. six reefs off West Maui. Ph.D. Thesis, University of Hawai'i, 277 pp.

Caruso C, De Souza MR, Kahkejian V, Davidson JM, Matsuda S, Madin JS, Drury C (2025) Short-term stress testing predicts subsequent natural bleaching variation. Coral Reefs

Caruso C, Rocha De Souza M, Ruiz-Jones L, Conetta D, Hancock J, Hobbs C, Hobbs C, Kahkejian V, Kitchen R, Marin C, Monismith S, Madin J, Gates R, Drury C (2022) Genetic patterns in Montipora capitata across an environmental mosaic in Kāne'ohe Bay, O'ahu, Hawai'i. Molecular Ecology 31:5201–5213

Ceccarelli DM, McLeod IM, Boström-Einarsson L, Bryan SE, Chartrand KM, Emslie MJ, Gibbs MT, Gonzalez Rivero M, Hein MY, Heyward A, Kenyon TM, Lewis BM, Mattocks N, Newlands M, Schläppy M-L, Suggett DJ, Bay LK (2020) Substrate stabilisation and small structures in coral restoration: State of knowledge, and considerations for management and implementation. PLoS ONE 15:e0240846

Chesson P (2000) Mechanisms of Maintenance of Species Diversity. Annual Review of Ecology and Systematics 31:343–366

Clark S, Edwards AJ Coral transplantation as an aid to reef rehabilitation: evaluation of a case
study in the Maldive Islands. 13
Clark S, Edwards AJ Use of Artificial Reef Structures to Rehabilitate Reef Flats Degraded by
Coral Mining in the Maldives. 21
Committee on Interventions to Increase the Resilience of Coral Reefs, Ocean Studies Board,
Board on Life Sciences, Division on Earth and Life Studies, National Academies of
Sciences, Engineering, and Medicine (2019) A Decision Framework for Interventions to
Increase the Persistence and Resilience of Coral Reefs. National Academies Press,
Washington, D.C.
Couch CS, Oliver TA, Dettloff K, Huntington B, Tanaka KR, Vargas-Ángel B (2023) Ecological
and environmental predictors of juvenile coral density across the central and western
Pacific. Front Mar Sci 10:1192102
Crouse DT, Crowder LB, Caswell H (1987) A Stage-Based Population Model for Loggerhead
Sea Turtles and Implications for Conservation. Ecology 68:1412–1423
Crowder LB, Lyman SJ, Figueira WF, Priddy J (2000) Source-sink population dynamics and the
problem of siting marine reserves. BULLETIN OF MARINE SCIENCE 66:
De Roos AM, Boukal DS, Persson L (2006) Evolutionary regime shifts in age and size at
maturation of exploited fish stocks. Proc R Soc B 273:1873–1880
Doak DF, Waddle E, Langendorf RE, Louthan AM, Isabelle Chardon N, Dibner RR, Keinath DA,
Lombardi E, Steenbock C, Shriver RK, Linares C, Begoña Garcia M, Funk WC,
Fitzpatrick SW, Morris WF, DeMarche ML (2021) A critical comparison of integral
projection and matrix projection models for demographic analysis. Ecological
Monographs 91:e01447
Donovan MK, Adam TC, Shantz AA, Speare KE, Munsterman KS, Rice MM, Schmitt RJ,
Holbrook SJ, Burkepile DE (2020) Nitrogen pollution interacts with heat stress to
increase coral bleaching across the seascape. Proc Natl Acad Sci USA 117:5351–5357
Donovan MK, Burkepile DE, Kratochwill C, Shlesinger T, Sully S, Oliver TA, Hodgson G,
Freiwald J, Van Woesik R (2021) Local conditions magnify coral loss after marine
heatwaves. Science 372:977–980
Dornelas M, Madin JS, Baird AH, Connolly SR (2017) Allometric growth in reef-building corals.
Proceedings of the Royal Society B: Biological Sciences 284:20170053
Doropoulos C, Evensen NR, Gómez-Lemos LA, Babcock RC (2017) Density-dependent coral
recruitment displays divergent responses during distinct early life-history stages. R Soc
open sci 4:170082
Doropoulos C, Roff G, Bozec Y, Zupan M, Werminghausen J, Mumby PJ (2016) Characterizing
the ecological trade-offs throughout the early ontogeny of coral recruitment. Ecological
Monographs 86:20–44
Drury C, Caruso C, Quigley K (2022) Selective Breeding to Enhance the Adaptive Potential of
Corals. In: Van Oppen M.J.H., Aranda Lastra M. (eds) Coral Reef Conservation and
Restoration in the Omics Age. Springer International Publishing, Cham, pp 71–84
Edmunds P, Riegl B (2020) Urgent need for coral demography in a world where corals are
disappearing. Mar Ecol Prog Ser 635:233–242
Edmunds PJ (2022) Finding signals in the noise of coral recruitment. Coral Reefs 41:81–93

Edmunds PJ, Burgess SC, Putnam HM, Baskett ML, Bramanti L, Fabina NS, Han X, Lesser
MP, Madin JS, Wall CB, Yost DM, Gates RD (2014) Evaluating the causal basis of
ecological success within the scleractinia: an integral projection model approach. Marine
Biology 161:2719–2734
Edwards A, Guest J, Humanes A (2024) Rehabilitating coral reefs in the Anthropocene. Current
Biology 34:R399–R406
Edwards AJ, Coral Reef Targeted Research & Capacity Building for Management Program
(2010) Reef rehabilitation manual. Coral Reef Targeted Research & Capacity Building
for Management Program, St. Lucia, Qld.
Figueiredo J, Baird AH, Harii S, Connolly SR (2014) Increased local retention of reef coral
larvae as a result of ocean warming. Nature Clim Change 4:498–502
Forsman ZH, Page CA, Toonen RJ, Vaughan D (2015) Growing coral larger and faster: micro-
colony-fusion as a strategy for accelerating coral cover. PeerJ 3:e1313
Friedlander A, Brown EK (2006) Hanalei Bay, Kaua'i marine benthic communities since 1992:
spatial and temporal trends in a dynamic Hawaiian coral reef ecosystem. Technical
Report HCSU-003, Pacific Aquaculture and Coastal Resources Center.
Gascoigne J, Lipcius R (2004) Allee effects in marine systems. Mar Ecol Prog Ser 269:49–59
Gove JM, Williams GJ, Lecky J, Brown E, Conklin E, Counsell C, Davis G, Donovan MK,
Falinski K, Kramer L, Kozar K, Li N, Maynard JA, McCutcheon A, McKenna SA, Neilson
BJ, Safaie A, Teague C, Whittier R, Asner GP (2023) Coral reefs benefit from reduced
land–sea impacts under ocean warming. Nature 621:536–542
Hagedorn M, Page CA, O'Neil KL, Flores DM, Tichy L, Conn T, Chamberland VF, Lager C,
Zuchowicz N, Lohr K, Blackburn H, Vardi T, Moore J, Moore T, Baums IB, Vermeij MJA,
Marhaver KL (2021) Assisted gene flow using cryopreserved sperm in critically
endangered coral. Proc Natl Acad Sci USA 118:e2110559118
Hall VR, Hughes TP (1996) Reproductive Strategies of Modular Organisms: Comparative
Studies of Reef- Building Corals. Ecology 77:950–963
Hansen A (2011) Contribution of source–sink theory to protected area science. In: Liu J., Hull
V., Morzillo A.T., Wiens J.A. (eds) Sources, Sinks and Sustainability. Cambridge
University Press, pp 339–360
Harrison PL, Dela Cruz DW, Cameron KA, Cabaitan PC (2021) Increased Coral Larval Supply
Enhances Recruitment for Coral and Fish Habitat Restoration. Front Mar Sci 8:750210
Higgins E, Metaxas A, Scheibling RE (2022) A systematic review of artificial reefs as platforms
for coral reef research and conservation. PLoS ONE 17:e0261964
Hughes TP, Baird AH, Dinsdale EA, Moltschaniwskyj NA, Pratchett MS, Tanner JE, Willis BL
(2000) Supply-side ecology works both ways: the link between benthic adults, fecundity,
and larval recruits. Ecology 81:2241–2249
Hughes TP, Baird AH, Morrison TH, Torda G (2023) Principles for coral reef restoration in the
anthropocene. One Earth 6:656–665
Hughes TP, Jackson JBC (1980) Do Corals Lie About Their Age? Some Demographic
Consequences of Partial Mortality, Fission, and Fusion. Science 209:713–715
Humanes A, Beauchamp EA, Bythell JC, Carl MK, Craggs JR, Edwards AJ, Golbuu Y, Lachs L,
Martinez HM, Palmowski P, Paysinger F, Randle JL, Van Der Steeg E, Sweet M,

Treumann A, Guest JR (2021) An Experimental Framework for Selectively Breeding
Corals for Assisted Evolution. Front Mar Sci 8:669995
Jury CP, Bahr KD, Cros A, Dobson KL, Freel EB, Graham AT, McLachlan RH, Nelson CE, Price
JT, Rocha De Souza M, Shizuru L, Smith CM, Sparagon WJ, Squair CA, Timmers MA,
Vicente J, Webb MK, Yamase NH, Grottoli AG, Toonen RJ (2024) Experimental coral
reef communities transform yet persist under mitigated future ocean warming and
acidification. Proc Natl Acad Sci USA 121:e2407112121
Kenkel CD, Traylor MR, Wiedenmann J, Salih A, Matz MV (2011) Fluorescence of coral larvae
predicts their settlement response to crustose coralline algae and reflects stress. Proc R
Soc B 278:2691–2697
Knapp ISS, Forsman ZH, Greene A, Johnston EC, Bardin CE, Chan N, Wolke C, Gulko D,
Toonen RJ (2022) Coral micro-fragmentation assays for optimizing active reef
restoration efforts. PeerJ 10:e13653
Lapointe BE, Brewton RA, Herren LW, Porter JW, Hu C (2019) Nitrogen enrichment, altered
stoichiometry, and coral reef decline at Looe Key, Florida Keys, USA: a 3-decade study.
Mar Biol 166:108
Levitan DR, Fukami H, Jara J, Kline D, McGovern TM, McGhee KE, Swanson CA, Knowlton N
(2004) MECHANISMS OF REPRODUCTIVE ISOLATION AMONG SYMPATRIC
BROADCAST-SPAWNING CORALS OF THE MONTASTRAEA ANNULARIS SPECIES
COMPLEX. Evolution 58:308–323
Madin JS, Anderson KD, Andreasen MH, Bridge TCL, Cairns SD, Connolly SR, Darling ES,
Diaz M, Falster DS, Franklin EC, Gates RD, Hoogenboom MO, Huang D, Keith SA,
Kosnik MA, Kuo C-Y, Lough JM, Lovelock CE, Luiz O, Martinelli J, Mizerek T, Pandolfi
JM, Pochon X, Pratchett MS, Putnam HM, Roberts TE, Stat M, Wallace CC, Widman E,
Baird AH (2016) The Coral Trait Database, a curated database of trait information for
coral species from the global oceans. Scientific Data 3:160017
Madin JS, Baird AH, Baskett ML, Connolly SR, Dornelas MA (2020) Partitioning colony size
variation into growth and partial mortality. Biology Letters 6:5
Madin JS, Baird AH, Connolly SR, Dornelas MA, Álvarez-Noriega M, McWilliam MJ, Barbosa M,
Blowes SA, Cetina-Heredia P, Christie AP, Cumbo VR, Diaz M, Emms MA, Graham E,
Hansen D, Hisano M, Howells E, Kuo C, Palmer C, Hong JTC, Zhi En Teo T, Woods RM
(2023a) Six years of demography data for 11 reef coral species. Ecology 104:e4017
Madin JS, Baird AH, Dornelas M, Connolly SR (2014) Mechanical vulnerability explains size-
dependent mortality of reef corals. Ecology Letters 17:1008–1015
Madin JS, Hughes TP, Connolly SR (2012) Calcification, Storm Damage and Population
Resilience of Tabular Corals under Climate Change. PLoS ONE 7:e46637
Madin JS, McWilliam M, Quigley K, Bay LK, Bellwood D, Doropoulos C, Fernandes L, Harrison
P, Hoey AS, Mumby PJ, Ortiz JC, Richards ZT, Riginos C, Schiettekatte NMD, Suggett
DJ, Van Oppen MJH (2023b) Selecting coral species for reef restoration. Journal of
Applied Ecology 60:1537–1544
McLeod IM, Hein MY, Babcock R, Bay L, Bourne DG, Cook N, Doropoulos C, Gibbs M,
Harrison P, Lockie S, Van Oppen MJH, Mattocks N, Page CA, Randall CJ, Smith A,
Smith HA, Suggett DJ, Taylor B, Vella KJ, Wachenfeld D, Boström-Einarsson L (2022)

Coral restoration and adaptation in Australia: The first five years. PLoS ONE
17:e0273325

McManus LC, Forrest DL, Tekwa EW, Schindler DE, Colton MA, Webster MM, Essington TE,
Palumbi SR, Mumby PJ, Pinsky ML (2021) Evolution and connectivity influence the
persistence and recovery of coral reefs under climate change in the Caribbean,
Southwest Pacific, and Coral Triangle. Global Change Biology 27:4307–4321

McWilliam M, Dornelas M, Álvarez-Noriega M, Baird AH, Connolly SR, Madin JS (2023) Net
effects of life-history traits explain persistent differences in abundance among similar
species. Ecology 104:e3863

Mumby PJ, Sartori G, Buccheri E, Alessi C, Allan H, Doropoulos C, Rengiil G, Ricardo G (2024)
Allee effects limit coral fertilization success. Proc Natl Acad Sci USA 121:e2418314121

Nguyen LT, Tran PD, Nguyen KQ (2022) An effectiveness of artificial coral reefs in the
restoration of marine living resources. Regional Studies in Marine Science 49:102143

Peixoto RS, Voolstra CR, Baums IB, Camp EF, Guest J, Harrison PL, Montoya-Maya PH,
Pollock FJ, Smith DJ, Wangpraseurt D, Banaszak AT, Chui APY, Shah N, Moore T,
Fabricius KE, Vardi T, Suggett DJ (2024) The critical role of coral reef restoration in a
changing world. Nat Clim Chang 14:1219–1222

Pratchett M, Caballes C, Wilmes J, Matthews S, Mellin C, Sweatman H, Nadler L, Brodie J,
Thompson C, Hoey J, Bos A, Byrne M, Messmer V, Fortunato S, Chen C, Buck A,
Babcock R, Uthicke S (2017) Thirty Years of Research on Crown-of-Thorns Starfish
(1986–2016): Scientific Advances and Emerging Opportunities. Diversity 9:41

Raker C, Olmeda-Saldaña M, Williams SM, Weil E, Prada C (2023) Use of predator exclusion
cages to enhance Orbicella faveolata micro-fragment survivorship and growth during
restoration. Front Mar Sci 10:1122369

Randall C, Negri A, Quigley K, Foster T, Ricardo G, Webster N, Bay L, Harrison P, Babcock R,
Heyward A (2020) Sexual production of corals for reef restoration in the Anthropocene.
Mar Ecol Prog Ser 635:203–232

Rapuano H, Shlesinger T, Roth L, Bronstein O, Loya Y (2023) Coming of age: Annual onset of
coral reproduction is determined by age rather than size. iScience 26:106533

Reichert J, Jorissen H, Drury C, Hancock JR, Haynes C, Nims AD, Rova LH, Schiettekatte
NMD, Madin JS (2025) Helix recesses boost coral larvae settlement and survival.
Biological Conservation 311:111407

Ricardo G, Doropoulos C, Babcock RC, Buccheri E, Khalil A, Mumby PJ (2025) Critical
thresholds of adult patch density and spacing during coral fertilization. Nat Ecol Evol

Rodriguez C (2021) Extracting Coral Vital Rate Estimates at Fixed Sites Using Structure-from-
Motion Standard Operating Procedures.

Rodriguez C "Measuring the Impact of Thermal Stress on Coral Resilience in Hawai'i Using
Large-Area Imagery." (2022). CSUMB Masters Thesis

Schopmeyer SA, Lirman D, Bartels E, Gilliam DS, Goergen EA, Griffin SP, Johnson ME, Lustic
C, Maxwell K, Walter CS (2017) Regional restoration benchmarks for Acropora
cervicornis. Coral Reefs 36:1047–1057

Smith SG, Swanson DW, Chiappone M, Miller SL, Ault JS (2011a) Probability sampling of stony
coral populations in the Florida Keys. Environ Monit Assess 183:121–138

Smith SG, Swanson DW, Chiappone M, Miller SL, Ault JS (2011b) Probability sampling of stony
coral populations in the Florida Keys. Environ Monit Assess 183:121–138
Speare KE, Enright LN, Aplin A, Adam TC, Edmunds PJ, Burkepile DE (2025) Early life stage
bottleneck determines rates of coral recovery following severe disturbance. Ecology
106:e4510
Streit RP, Morrison TH, Bellwood DR (2024) Coral reefs deserve evidence-based management
not heroic interference. Nat Clim Chang 14:773–775
Terence P. Hughes (1984) Population Dynamics Based on Individual Size Rather than Age: A
General Model with a Reef Coral Example. The American Naturalist 123:778–795
Torres WI, Holstein DM, Putnam HM, Edmunds PJ, Puritz JB, Toonen RJ, Hench JL (2025)
Post-disturbance recovery dynamics of connected coral subpopulations. Theor Ecol 18:6
Tortolero-Langarica JJA, Rodríguez-Troncoso AP, Cupul-Magaña AL, Rinkevich B (2020)
Micro-Fragmentation as an Effective and Applied Tool to Restore Remote Reefs in the
Eastern Tropical Pacific. IJERPH 17:6574
Towle EK, Donovan EC, Kelsey H, Allen ME, Barkley H, Blondeau J, Brainard RE, Carew A,
Couch CS, Dillard MK, Eakin CM, Edwards K, Edwards PET, Enochs IC, Fleming CS,
Fries AS, Geiger EF, Grove LJ, Groves SH, Gorstein M, Heenan A, Johnson MW,
Kimball J, Koss JL, Kindinger T, Levine A, Manzello DP, Miller N, Oliver T, Samson JC,
Swanson D, Vargas-Ángel B, Viehman TS, Williams ID (2022) A National Status Report
on United States Coral Reefs Based on 2012–2018 Data From National Oceanic and
Atmospheric Administration's National Coral Reef Monitoring Program. Front Mar Sci
8:812216
Tsounis G, Edmunds PJ (2016) The potential for self-seeding by the coral Pocillopora spp. in
Moorea, French Polynesia. PeerJ 4:e2544
Van Oppen MJH, Oliver JK, Putnam HM, Gates RD (2015) Building coral reef resilience through
assisted evolution. Proc Natl Acad Sci USA 112:2307–2313
Vega Thurber RL, Burkepile DE, Fuchs C, Shantz AA, McMinds R, Zaneveld JR (2014) Chronic
nutrient enrichment increases prevalence and severity of coral disease and bleaching.
Global Change Biology 20:544–554
Veron JEN (2000) Corals of the world.<NZ> Vol. 1: XII, 463 S. zahlr. Ill., Kt.<NZ> Vol. 2 VIII,
429 S. zahlr. Ill., Kt.<NZ> Vol. 3 VIII, 490 S. zahlr. Ill., Kt. Australian Institute of Marine
Science, Townsville
Vitt P, Havens K, Hoegh-Guldberg O (2009) Assisted migration: part of an integrated
conservation strategy. Trends in Ecology & Evolution 24:473–474
Watt-Pringle R, Razak TB, Jompa J, Ambo-Rappe R, Kostaman AN, Smith DJ (2024) Coral reef
restoration in Indonesia: lessons learnt from the world's largest coral restoration nation.
Biodivers Conserv 33:2675–2707
Winston M, Couch C, Ferguson M, Huntington B, Swanson DW, Vargas-Ángel B (2019)
Ecosystem Sciences Division Standard Operating Procedures: Data Collection for Rapid
Ecological Assessment Benthic Surveys, 2018 Update. 6.50 MB
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763 **Supporting Materials**
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Supporting Methods–A key uncertainty with the matrices used in this study were the transition
boundaries between recruits, juveniles and adults. Boundaries for the Comadre matrices were
set during the design of the original studies and these cannot be changed. Survival rates in the
recruit stage were consistently lower than the juvenile stage (Fig. 2), and the boundary between
juvenile and adult was in the vicinity of reported mature colony sizes in the literature (Table 1).
For the Trimodal species we are able to fine-tune the boundaries based on fecundity and
survival data. For instance, we defined the recruit-juvenile boundary as the colony size where
survival rate stabilized (Fig. S1) and the juvenile-adult boundary as the colony size where
approximately 75% of colonies were reproductive (Fig. S3).

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775 **Figure S1**. Yearly survival probability for Trimodal (Madin et al. 2023) and NOAA (Rodriguez et
776 al 2021) species as a function of colony size showing transition boundaries used for recruit-
777 juvenile (solid vertical lines) and juvenile adult (dashed lines corresponding with Fig. S2). Blue
778 lines are loess smoothers to illustrate underlying size-survival patterns.

779
780 **Figure S2**. Size at reproduction for the 11 trimodal species (Madin et al. 2023) with logistic
781 model fits (blue lines). We used the approximate size at which 75% of colonies are reproductive
782 as the juvenile-adult boundary for transition matrices.

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A. Multiplicative

B. Additive

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786 **Figure S3**. A demonstration of the matrix operations for (**A**) multiplicative and (**B**) additive
787 approaches to increasing year on year population growth by 5% for *Acropora cytherea*. $\odot$
788 represents the Hadamard product and • represent the matrix product. Σ represents the sum.
789 This example represents best estimates of recruit survival change in A and yearly recruit
790 outplanting in B. The values circled in yellow were found using least square optimization in R.

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794 **Figure S4**. Elasticities.
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799   **Figure S5**. Boxplots for transition cells for the 34 species matrices summarized by growth form.
800   Color coding matches Fig. 1.
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