MycoMobilome: A community-focused non-

redundant database of transposable element

consensus sequences for the fungal kingdom

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Abstract

Transposable elements (TEs) are found in nearly all eukaryotic genomes. Despite

significant advances in the sequencing of genomes, TE resources remain sparse,

leading to a lack of traceability, reproducibility, and duplication of effort when

annotating TEs. Here, we focus on the fungal kingdom and present MycoMobilome, a

non-redundant database of TE consensus sequences systematically curated using a

set of 4,309 genomes covering all major clades. The initial database contains 276,641

consensus sequences after filtering to remove putative host genes and low-quality

consensus sequences. We provide a consistent naming convention to surface

information on the confidence in the classification including potential conflicting ORF

functions, along with metadata to enable evaluation of TEs of interest and to determine

whether further curation work is required on a case-by-case basis. Finally, we provide

guidelines for community contributions, and encourage researchers to deposit new or

curated sequences, which will be incorporated into future MycoMobilome releases.

Introduction

Annotation of transposable elements (TEs) is essential for the analysis of genomes even

if the focus is primarily on coding sequences. TE annotation relies on the initial

discovery of TE families, termed *de novo* curation, followed by homology-based

annotation using tools to recognise all members of each TE family. Various approaches

exist for *de novo* curation, with the most common including RepeatModeler2 (1) and

TEdenovo of REPET (2), whilst there are also databases providing previously curated TE

families that can be used to annotate genomes, including Dfam (3, 4), TREP (5), and

RepBase (6).

The fungal kingdom spans almost a billion years of evolution, exhibits

remarkable phenotypic and genomic diversity, and contains species found in nearly all

ecological niches. The availability of genome resources for species spanning the

kingdom is accelerating, partly due to efforts including the 1000 fungal genomes

initiative (https://1000.fungalgenomes.org) and broader efforts of the Darwin Tree of

Life and Earth Biogenome projects (7, 8). Despite these growing efforts, curated and

multi-species TE resources for the fungal kingdom are almost non-existent. For

example, in Dfam version 3.9, the only curated TEs are found for *Septoria linicola* and

Zymoseptoria tritici (https://dfam.org/). Dedicated efforts to curate TE libraries for

individual species include the plant pathogens *Magnaporthe oryzae* (9) and *Z. tritici*

(10). However, cross-referencing TE annotations across species has not been

attempted, leading to reproducibility issues and duplication of effort as researchers

perform annotations largely on a per-species basis.

Here, we present MycoMobilome

(https://github.com/TobyBaril/MycoMobilome), a non-redundant database of TE

consensus sequences covering species across the fungal kingdom. We created

MycoMobilome using all publicly available genome resources for fungi and provide

researchers with a systematically generated TE consensus library with a persistent

naming scheme to improve consistency and reproducibility. Consensus sequences are

provided with labels to show whether their classification is supported by open reading

frames (ORFs), and whether these consistently match the provided classification.

Applying MycoMobilome classification to primary fungal models, we find that the

genome fraction annotated as TEs increases by between +7.20% in *M. oryzae* to

+13.41% in *Cryptococcus neoformans* compared to existing community TE

annotations.

MycoMobilome is a key resource to facilitate investigations into TEs across the

fungal kingdom and provides easy-to-implement methods for researchers to contribute

new or updated sequences with attribution. Consistent application of MycoMobilome

will improve consistency in TE naming conventions, whilst reducing the duplication of

effort often accompanying repeat annotation in new genome assemblies.

84   Materials and Methods

85   *Initial Curation for MycoMobilome*

All publicly available genomes and associated proteomes for the fungal kingdom were

sourced using Mycotools (version 0.32.3)(11). Genomes were manually filtered to

remove those under embargo, resulting in a final set of 4,309 genome assemblies.

Genome assembly metadata is provided in the MycoMobilome database

(https://doi.org/10.5281/zenodo.17037469).

For each genome, a library of putative TE consensus sequences was generated

using `earlGreyLibconstruct` in Earl Grey (v4.4.0)(12), configured with Dfam curated

elements only (v3.7)(3, 4). All 4,309 *de novo* consensus libraries were subsequently

combined into a single FASTA file containing 773,843 sequences. These sequences

were clustered using the `easy-cluster` scalable cascaded clustering approach in

MMseqs2 (13) with `--min-seq-id 0.8 -c 0.8 --cov-mode 1 --cluster-reassign` to cluster

sequences to the 80-80-80 TE family rule (14), resulting in 354,315 non-redundant

consensus sequences. The representative sequence for each cluster was extracted

and labelled with the source genome (*i.e.* the genome from which the consensus

sequence originated).

Autonomous TEs encode domains for their selfish activity, and the identity of

these open reading frames (ORFs) can be used to classify TEs (14–16). Conversely,

multicopy host proteins can erroneously be curated as repetitive sequences by

automated TE curation methods, given their repetitive occurrence in host genomes. We

translated all six frames of each TE consensus sequence using `transeq -clean -frame

6` in EMBOSS (v.6.6.0)(17). We identified matches to known host proteins present in

the Fungi partition of RefSeq release 228 (18) using Diamond BLASTp (v 2.1.11) (19) with

`--sensitive --matrix BLOSUM62 --evalue 1e-3`. Following this, we detected similarity

to characterised TE proteins using two complementary approaches. First, we used

HMMscan in HMMER (v3.4)(20) with `-E 10 --noali` to detect homology to known TE

protein domain hmm models supplied in ProfilesBankForREPET_Pfam35.0_GypsyDB as

part of the REPET software suite (2, 21, 22). Hits were filtered to retain those with

fseq_evalue $\leq$ 0.001 and fseq_bitscore $\geq$ 50. Next, we used BLASTp (v2.14.1+)(23) to

detect homology to known TE protein domains found in RepeatPeps.lib, which is part of

RepeatMasker (v4.1.9)(24) and is used to classify TEs using the RepeatClassifier

module in the initial *de novo* curation step in Earl Grey. A minimum e-value of 1e-3 was

defined, and nested hits were removed to retain the highest quality protein hit for each

query, followed by combining of adjacent and overlapping hits. Potential host gene hits

were identified using the approach developed by (25). A TE consensus sequence was

designated as a putative host gene if either: (i) there were hits to RefSeq queries and no

hits to known TE queries; (ii) there were hits to both RefSeq queries and known TE

queries, but at least 90 residues aligned to a RefSeq query did not overlap with

alignments to known TE queries. Consensus sequences with no hits were retained in

the TE library as putative non-autonomous TEs. In total, 24,571 consensus sequences

were identified as potential host genes and removed.

To further refine the database, poor-quality TE consensus sequences were

filtered. We define a poor-quality consensus sequence as <120bp in length, as these

are likely to be incomplete and poor quality. Further, majority-rule consensus

sequences can sometimes contain unknown nucleotides, indicated with N in the

sequence. We removed all sequences with $\geq$5% N using `seqtk comp`

(https://github.com/lh3/seqtk). In total, 53,103 consensus sequences were removed

for not meeting the quality thresholds, resulting in a final database size of 276,642

putative TE consensus sequences.

The majority of TE consensus sequences in publicly available databases remain

uncurated. Despite this, these sequences remain a widely used resource. To enable the

community to evaluate TE annotation quality of newly analyzed genomes, we introduce

an automated classification of each TE consensus against protein hits to known TEs

(Tables are provided in the MycoMobilome database). Each TE consensus was labelled

with a two-letter code providing the level of evidence supporting the TE classification:

`_PE` for protein evidence that matches the given classification, `_DA` for protein

evidence that contradicts the given classification, and `_NE` for no protein evidence,

which includes non-autonomous TEs.

Finally, each TE consensus in MycoMobilome was given a unique name with the

following format: `MycMob1.0_family-[unique_family_number]-

[six_letter_species_code]_[protein

evidence]#[high_level_TE_classification]/[sub_level_TE_classification] @[genus

species]`.

Comparison of existing TE annotation approaches with

MycoMobilome

We wanted to assess the extent to which sampling TEs across fungal diversity enables

improved detection of TE-derived genome content. To do this, we selected three well-

studied organisms and compared TE annotation using MycoMobilome against the

traditional approaches employed by the research communities working with each of

these organisms. We compared our standardised approach with MycoMobilome

against (i) annotation with known fungal TEs from RepBase and Dfam, as used in a

recent study on *C. neoformans* (26); (ii) a community annotation resource for *Candida*

albicans

(http://www.candidagenome.org/download/gff/C_albicans_SC5314/Assembly22/); (iii)

annotation using a curated TE library with RepeatMasker for *M. oryzae* (9).

For *C. neoformans*, we adopted the same approach used in the original study

and annotated the genome assembly for isolate CNA3 H99 (GCF_000149245.1) using

RepeatMasker (v4.1.9) configured with Dfam (v3.9) and RepBase RepeatMasker Edition

(release 20181026) and the options `-species fungi -norna -no_is`. For *M. oryzae*, we

sourced the reference genome assembly MG8 (GCF_000002495.2) and annotated this

with RepeatMasker using the TE consensus library provided with the publication (9). For

C. albicans, we obtained the genome assembly in FASTA format and the community

feature annotation in GFF format for genome assembly SC5314 Assembly 22 from the

Candida genome portal

(http://www.candidagenome.org/download/gff/C_albicans_SC5314/Assembly22/). To

obtain a GFF of repeat regions, we filtered the feature GFF for features named

long_terminal_repeat, repeat_region, and retrotransposon.

For comparison, we adopted our standardised approach as recommended with

MycoMobilome. We annotated all three genome assemblies with

`earlGreyAnnotationOnly` using Earl Grey (v6.3.2), providing MycoMobilome as the

input library, with all other settings left as default.

Hits that are shared and unique to each methodological approach were

identified using BEDTools intersect (v2.31.1)(27), and resultant feature coverage was

calculated in R (v4.4.3)(28) with tidyverse (29). For shared annotations, the width of the

annotation in each case was calculated, rather than using a single length of annotation

across compared methodologies, to account for variation in annotation size between

methods. Venn diagrams were generated using the ggVennDiagram package (30).

Results & Discussion

MycoMobilome assesses TE evidence across the fungal kingdom

The MycoMobilome release contains 276,642 consensus sequences, of which 39,265 have classifications supported by protein evidence. This provides a curated, non-redundant resource for the diversity of TEs across the fungal tree of life. In particular for clades without previous TE annotation efforts, this represents a very substantial expansion of genomic datasets. We performed filtering steps to stringently remove any consensus sequences with the potential to be host genes, as well as poor-quality consensus sequences. Following these efforts, we provide three versions of the database: a full database, a subset containing only TE consensus sequences with protein evidence, and a subset containing only TE consensus sequences lacking protein evidence. The compressed database files are provided via Zenodo (https://doi.org/10.5281/zenodo.17037469) in FASTA format, ensuring compatibility with a broad range of bioinformatics approaches. We provide guidance to non-specialists on how to best implement the MycoMobilome dataset into TE annotation pipelines (https://github.com/TobyBaril/MycoMobilome). We also provide a record of the genomes used to curate TEs across fungal diversity, and tables of hits to known TE proteins from multiple sources to cross-reference evidence supporting TE classifications for sequences of interest.

A community-based curation effort through continuous submissions

We also provide a MycoMobilome community (https://zenodo.org/communities/mycomobilome), hosted on Zenodo, with the aim of encouraging users to contribute new or improved (*i.e.* manually curated) TE consensus sequences to future releases. TE curation takes considerable work and expertise, which researchers should be recognised for. By adopting the Zenodo community approach, user contributions are assigned persistent DOIs, enabling contributors to be cited and recognised for their contributions to the wider genomics community. We provide guidance on how to contribute to MycoMobilome on the GitHub page, maximising ease-of-use. Further, hosting such a resource on Zenodo reduces risks associated with longevity of the database. The nature of MycoMobilome as a database in FASTA format limits file sizes and enables efficient storage and reuse without extensive infrastructure or resource requirements.

MycoMobilome improves TE detection in important fungal models

By sampling TEs across fungal diversity, we increase the proportion of well-studied host genomes that is attributed to TEs (Figure 1, Supplementary Table 1). For the major rice and wheat pathogen, *M. oryzae*, which has a well-curated TE library, and much attention focused on TE dynamics, MycoMobilome increases the proportion of the host genome annotated as TEs by 7.20% (2.95Mb). The use of Earl Grey to annotate TEs also increases the proportion of the genome annotated as TEs for loci detected both in previous annotations and MycoMobilome by 10.57% (*i.e.* "shared"; 801.1kb). The improved annotation is likely due to the incorporation of defragmentation and overlap

resolution steps, consistent with previous observations (12). When annotating the

genome of *C. neoformans* with MycoMobilome, we find an increase of host genome

proportion annotated as TEs of 13.41% (2.54Mb) compared to the previous annotation,

which relied on fungal TEs hosted in Dfam and RepBase (Figure 1). In addition, a similar

pattern is observed in *C. albicans*, where annotation using MycoMobilome increases

the proportion of the genome recognised as TE by 9.82% (2.81Mb) compared to the

current community resource based on within-species TE characterisation. Hence,

MycoMobilome provides a substantial increase in well-characterized TEs even in

comparatively well-studied fungal models.

TEs can persist in host genomes over long evolutionary time, over which they will

experience mutation and degradation leading to the accumulation of genomic fossils,

challenging our ability to detect these sequences as TE-derived. However, in related

lineages, these TE families may persist and be retained with higher levels of identity and

at higher copy numbers, enabling their detection. By sampling TEs across deep

evolutionary time and using this information to generate a non-redundant TE library, we

were able to detect homology to TEs that may evade detection using single-species *de*

novo approaches, which require high TE copy numbers to generate an initial consensus

sequence. We show that the use of MycoMobilome increases the proportion of the host

genome annotated as TEs, demonstrating the power of sampling TEs across the fungal

kingdom, and over deep evolutionary time, to be able to detect TE-derived sequences

across fungal diversity.

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Figure 1. The use of MycoMobilome increases the fraction of genomes annotated as
*transposable elements in three key fungal pathogen models. **A.** Venn diagrams*
illustrating the number of base pairs for each genome assembly that were annotated as
TEs using either classical approaches or MycoMobilome (with Earl Grey). Depth of
colour and plot area is proportional to the percentage of total base pairs annotated as
TE across all categories. Numbers in brackets show the percentage of all annotated
base pairs that are found in each category, as labelled. Area of Venn diagrams is
*proportional to total genome size. **B.** TE annotations split by approaches with which*
they are identified, and the classification of each annotated TE, as indicated in the key.
Discrepancies in TE coverage at shared loci arise due to the defragmentation and
overlap resolution steps automatically performed with Earl Grey following TE annotation
with MycoMobilome.

262 Conclusions

Here, we introduce MycoMobilome, a non-redundant database of TE consensus

sequences for the fungal kingdom. We aim to provide this database as a key starting

point to facilitate genomic investigations in lineages spanning the fungal kingdom. Key

features include consistent naming conventions, metadata on the quality of TE

classifications and an easy-to-follow tutorial for the community to process additional

genomes. We welcome contributions to the MycoMobiliome database and provide

citable recognition for submitted TE annotations. We encourage the fungal research

community to engage and expand the database scope to the benefit of all.

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Data Availability

355 The MycoMobilome database has been deposited in the Zenodo database under DOI

356 10.5281/zenodo.17037468 (https://doi.org/10.5281/zenodo.17037468). User guidance

357 and documentation is hosted on GitHub (https://github.com/TobyBaril/MycoMobilome)

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366 T.B. and D.C. conceived and coordinated the study. T.B. performed analyses. D.C.

367 provided funding and supervised the work. T.B and D.C. wrote the manuscript.

369 Conflict of Interest Disclosure

370 The authors declare no conflicts of interest.