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Designing Convergent Overlapping Genes with Transformer Encoder Models and

Lightweight Structural Proxies

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Abstract

Overlapping genes allow multiple proteins to be encoded from a single DNA sequence, including convergent (antisense; tail-to-tail) orientations across three reading frames (phases 0, 1, and 2), with phase 1 most frequently observed in nature. Designing such overlaps is challenging due to codon degeneracy, phase-specific biases, and the need to preserve structural integrity for both proteins. Here, a purpose-built transformer encoder is introduced, trained on a balanced synthetic dataset of convergent overlaps spanning diverse prokaryotic genomes and GC contents. Controlled amino acid substitutions were incorporated during training to enhance model generalization, particularly for phase 1 overlaps. At inference, Monte Carlo dropout enabled uncertainty-aware sampling of synonymous codon solutions, which were iteratively refined using a windowed, multi-objective optimization framework. Candidate overlaps were scored using composite weighting across secondary structure preservation, substitution similarity, alignment identity, and ESM-2 contact map similarity, with SSIM applied as a rapid proxy for structural fidelity. This approach generated convergent overlaps across all phases, with phase 1 showing the highest success rates. Optimization trajectories revealed distinct dynamics, with

secondary structure preservation steadily increasing despite its lower weight. External validation

using SwissProt proteins stratified by AlphaFold2 pLDDT confidence supported generalization

to proteins with differing rigidity, yielding high secondary structure preservation in silico. These

results demonstrate that transformer models trained directly at the nucleotide level, when

coupled with uncertainty-aware inference and lightweight structural proxies, can support the

computational design of synthetic overlapping genes without requiring full structural prediction.

This framework offers a scalable path for phase-specific, codon-aware overlap design under

realistic constraints.

Introduction

The study of overlapping genes is an important area of interest in prokaryotic genetics, attributed

to their ability to encode multiple functional products from a single DNA sequence (1–4). For

example, a single gene sequence might code for two distinct proteins, each playing a different

role in cellular function. Advancements in identifying and designing these overlapping sequences

promise the creation of genetic constructs that are both more efficient and compact, potentially

boosting the functionality and stability of engineered organisms (5,6). This phenomenon not only

poses challenges but also opens new avenues in genetic engineering and synthetic biology (3).

Overlapping genes have been described using different approaches, but can be broadly grouped

into unidirectional (sense; encoded on the same strand, $\rightarrow \rightarrow$) or convergent (antisense; encoded

on the opposite strand) orientations (**Fig 1A**); convergent overlaps can be further refined into

"head-to-head" ($\leftarrow \rightarrow$) or "tail-to-tail" ($\rightarrow \leftarrow$) orientations (7). Convergent overlapping genes

occur in the same reading frame (phase 0), or frameshifted by one (phase 1) or two (phase 2)

nucleotides. For example, in phase 2, convergent overlaps share their third (degenerate) codon

positions. Research into convergent (tail-to-tail; antisense) overlapping gene sequences in

prokaryotes has included analyses on the length distribution among overlapping and near-

overlapping genes (8), investigations into their prevalence (and misannotations) within

prokaryotic genomes (9,10), as well as the characterization of convergent gene pairs (11–18).

Notably, both convergent and unidirectional overlaps exhibit a non-uniform distribution with a

significant phase bias, predominantly favoring phase 1 overlaps (8,19,20). While the majority of

convergent overlaps fall into phase 2, when excluding 4 nucleotide overlaps, nearly half are

instead found in phase 1 (8). These studies collectively highlight the phase preference and

distribution patterns of naturally occurring gene overlaps.

The successful prediction of overlapping nucleotide sequences encoding two amino acid

sequences poses unique computational challenges, and has been previously explored (6,21–25);

the landscape of available methods has been summarized (22) with additional information

pertaining to more recently released programs provided in **Supplementary Table 1**. An early

solution for the design of unidirectional overlapping protein sequences included design of an

algorithm to identify the shortest DNA sequence which could encode two amino acid sequences

(26). More recently, examples included a dynamic programming algorithm (6) and a subsequent

method that combines dynamic programming with a hidden Markov model (HMM) (5,27). The

strategy was further extended to arbitrary pairs of natural protein domains (28). These

approaches have collectively shown that it is possible to design fully embedded overlapping

genes that encode functional proteins with high homology to starting sequences, underscoring

the potential for engineered overlapping genes in prokaryotic genetics. Another approach, termed

overlapping, alternate-frame insertion (OAFI) demonstrated the ability to design unidirectional

overlaps through insertion of one gene into an alternate reading frame of another by targeting

sites that may tolerate insertions (29). Recently, Byeon et al. (2025) demonstrated that deep

generative models can be used to design synthetic overlapping genes spanning distinct protein

families, achieving high in silico and experimental success rates even under the constraints of the

standard genetic code (30). Their results indicate that sequence space may contain many viable

overlapping solutions, suggesting broader applicability of computational approaches to both

unidirectional and convergent overlaps.

Transformer models have demonstrated superior performance in various sequence modeling

tasks due to their ability to capture long-range dependencies and contextual information (31–33).

This study leverages a novel application of the transformer encoder-based model architecture to

predict synthetic convergent overlapping genes in prokaryotes. Unlike dynamic programming

approaches, transformers utilize self-attention mechanisms to process the entire input sequence

simultaneously (32), enabling the model to capture patterns and dependencies that span across

long sequences. In this study, Monte Carlo (MC) dropout is applied during inference to enhance

prediction performance. Dropout, more generally, is a method of reducing overfitting in deep

neural networks during training (34). MC dropout retains dropout activity during inference,

effectively sampling from a distribution of possible model predictions (34–36). This provides a

practical Bayesian approximation of model uncertainty (37), which is especially advantageous

for predicting overlapping genes where the degeneracy of the genetic code permits many distinct

but functionally viable nucleotide encodings. By sampling multiple plausible solutions, MC

dropout enables exploration of the many synonymous coding possibilities inherent to the genetic

code, increasing the likelihood of identifying overlaps that preserve the structural and functional

integrity of both proteins.

The objectives of this study are to (i) evaluate the ability of transformer encoder models to

recover convergent overlaps across all three phases, (ii) quantify the impact of overlap length

and sequence variability on prediction success, and (iii) assess multi-objective optimization as an

alternative to full structural prediction in improving overlap fidelity.

Methods

Synthetic Overlapping Gene Dataset Construction for Model Training

While naturally occurring convergent overlaps have been systematically assessed and

documented (9), there is no readily available database of convergent overlapping genes with

equally distributed overlap lengths. Given this limitation, a database of synthetic convergent

overlaps was generated using a diverse set of open reading frames (ORFs) from prokaryotic

genomes. Synthetic convergent overlapping genes were created using ORF sequences from 24

different genomes from prokaryotes (**Supplementary Table 2**), using the R (version 4.3.1) and

Python programming languages (version 3.8.8). Genomes were selected to have a wide range of

genomic GC content (ranging from 23.3% to 74.9%, with a mean of 52.41%), with the aim of

controlling for potential amino acid compositional bias driven by GC content (38). The between-

strain coding GC content (calculated using the set of ORF sequences from each strain) ranged

from 20.7% to 72.7%, with a mean of 50.5%.

Briefly, for each strain, sequences were downloaded from the NCBI database

(https://www.ncbi.nlm.nih.gov/genome/) and each FASTA file was processed to extract ORFs,

which were stored in separate list for each strain (**Fig 1B**). ORF sequences were filtered to

remove sequences shorter than 450 nucleotides, to exclude sequences with insufficient

opportunity for varied random sampling during overlap construction. A function was defined to

randomly select in-frame segments from the ORF sequence lists, to include a primary and

secondary sequence either with or without a convergent overlapping region. First, a contiguous

in-frame 312-nucleotide length section from each ORF sequence was extracted and used as the

basis for the primary sequence. The primary sequence segments were modified to ensure they

contained start and randomly selected stop codons, as these were removed during initial

sequence processing.

The reverse complement of this synthetic gene was selected to generate the secondary sequence

with a defined convergent overlap length. A uniform set of overlap lengths was generated for

each length by extracting the reverse complement, adding a stop codon, and removing any in-

frame stop codons with a random codon. Both the added stop codon and in-frame replacements

were selected such that stop codons in the forward direction were not introduced. However,

owing to the methodology employed, a portion of generated secondary sequences with

overlapping sequences contained more than one stop codon and were subsequently filtered from

the dataset to ensure all sequences contained only one stop codon and encoded a single amino

acid sequence.

The amino acid sequence pairs with known overlaps were modified to incorporate up to 4 of

BLOSUM62 weighted amino acid changes. The changes included altering amino acid sequences

such that 1) the internal reverse stop codon was removed from both amino acid sequences

(primary and secondary), while keeping the known overlap for the original amino acid

sequences, or 2) the internal reverse stop codon was removed from both amino acid sequences

(primary and secondary), while also introducing additional amino acid modifications per

sequence. This yielded modified primary and secondary amino acid sequences, with an

unmodified known convergent overlap nucleotide sequence capable of encoding amino acid

sequences with high amino acid sequence identity to the primary and secondary amino acid

sequences.

The length of the input amino acid sequences was limited to 104 amino acids to manage

computational complexity during training and inference phases. Each amino acid was separated

by a space and the overall included a terminal asterisk (*) and the primary and secondary

sequences were concatenated to generate a single input sequence for training.

To balance the representation of different overlap lengths, data were stratified based on the

length of the overlap. The overlapping gene training data were balanced, ensuring equal

representation for each strain and each length of nucleotide overlap (specifically, convergent

overlaps of length 199 to 312 nucleotides). The final dataset was composed of synthetic gene

overlaps, each represented by a pair of modified amino acids sequences with a known target

convergent overlap sequence. This dataset served as the training set for the transformer encoder-

based models.

Initial Model Description and Training

The synthetic overlapping gene dataset was used to train transformer-encoder based models with

the aim of generating convergent (tail-to-tail) overlaps for any two specified amino acid

sequences. A total of 8,892,000 combined pairs, corresponding to 78,000 pairs per convergent

overlap length.

Transformers have demonstrated superior performance in various sequence modeling tasks,

particularly where understanding long-range dependencies is important (31–33). As such, this

study utilizes a PyTorch based transformer encoder model for overlap prediction (39).

Tokenization and text vectorization were performed using the TensorFlow Keras API (40), and

data were split into training and validation sets. During training, the models each utilized 156

embedding dimensions, 13 attention heads, 192 feed-forward network dimensions, 5 blocks, a

dropout rate of 0.1, and a total of 802,983 trainable parameters (**Fig. 2D**). Models were compiled

using the Adam optimizer with cross-entropy as the loss function.

Each model was trained for 3 epochs with a batch size of 32 on a single NVIDIA GeForce GTX

5070 Ti GPU with 16GB of memory, and reached a validation accuracy of ≥90% by the third

and final epoch.

Test Dataset Generation and Model Inference

Amino acid pairs were generated as described above for preparation of the training dataset, but

without the final modification step; however, a different set of strains (n = 12) were selected than

those used for training, with a wide range of genomic GC content (ranging from 23.6% to 74.1%,

with a mean of 50.2%) (**Supplementary Table 3**). Given that inference was performed using

only coding sequences, the coding sequence GC content was calculated for each strain, and the

corresponding mean GC value was used for subsequent analyses (varying from 24.2% to 72.1%,

and a mean of 50.0%).

It is noted that inference was performed using the trained models along with the same tokenizer

to vectorize the input sequence. Using a "greedy decoder" (41), the token with the highest

probability at each step of the sequence generation process was selected.

MC dropout was implemented during inference to allow exploration of the uncertainty in model

predictions (eg, variation in predicted overlap sequences), leveraging the benefits of Bayesian

inference, as previously described (34,36,42). During inference, depending on the objective,

feedforward dropout was set to 0.275 for initial predictions.

An algorithm was developed to generate overlapping gene sequences from input amino acid

sequences using the trained models. The algorithm first back-translated the input amino acid

sequences into a nucleotide sequence, leveraging *E. coli* codon usage frequencies to reflect the

probabilistic distribution of codons for each amino acid (43). After back-translation, the models

were used to predict potential overlaps within these nucleotide sequences. Predicted overlaps

were then integrated into the original nucleotide sequences, resulting in a collection of modified

sequences that incorporated the predicted overlapping regions downstream of the remaining

back-translated nucleotide sequence. These modified sequences (ie, the back-translated and

predicted regions) were then translated into amino acid sequences to assess the accuracy of the

predictions. In instances where the correct amino acid sequence was not identified, additional

rounds of prediction and back-translation were performed. This iterative process continued until

either an overlap yielding amino acid sequences meeting the pre-specified criteria was identified,

or the specified number of inference rounds were completed without a predicted overlap (defined

as an unsuccessful prediction).

An alignment score was calculated for each sequence prediction. The predicted sequence was

aligned against the known overlap using the Needleman-Wunsch algorithm (implemented via

pairwise2 from Biopython (44)). For sequences with a known overlap and a predicted overlap

sequence, the calculated alignment score was based on the length of the overlap, normalizing

scores to a scale of 0 to 1 to facilitate comparison across predictions.

Secondary Structure Prediction, Substitution Score, and Structure Prediction

The prediction of secondary structures was performed using S4PRED, which enabled predictions

based on an input amino acid sequence alone without the need for multiple sequence alignments

or known homologous sequences (45). S4PRED outputs were formatted as "ss2" files, from

which the predicted secondary structure sequence was extracted for downstream analyses. The

model employs a simplified three-state classification—α-helix (H), β-sheet (E), and coil/loop

(C), which corresponds to a coarse-grained version of the DSSP (Dictionary of Secondary

Structure of Proteins) annotations (46). Minor code modifications were implemented to enable

efficient batch processing, substantially reducing runtime. These predicted structures were used

to evaluate the structural compatibility of convergent overlapping gene candidates. Substitution

matrices utilized were BLOSUM62 (47,48) and ProtSub (49). Protein structures were predicted

either with AlphaFold3 or ColabFold (50–52).

Optimization of predicted sequences

An iterative, windowed optimization procedure was used to convert paired amino-acid inputs

into convergent (tail-to-tail) overlapping nucleotide sequences while preserving fold-relevant

features of both proteins and maintaining sequence similarity. Input amino-acid sequences were

first back-translated to an initial nucleotide scaffold using *E. coli* codon usage frequencies. The

region targeted for overlap design was tiled into partially overlapping amino-acid windows; in

the experiments reported here windows were 10 amino acids with an 8-amino-acid stride (10 AA

window, 8 AA stride; model receptive field used = 105 AA). During optimization, only

nucleotides mapping to the current window were permitted to change; flanking nucleotides were

held fixed except at user-designated residues enforced by token-level biases (for example to

preserve active-site residues or motifs). Bracketed constraints on the forward strand were

mirrored on the reverse-complement strand so that enforced bases remained consistent with the

convergent orientation.

Within each window a transformer-encoder was executed in an uncertainty-aware mode: Monte-

Carlo (MC) dropout was retained at inference to generate diverse synonymous codon proposals.

Token-level fixed-logit (base) biases were applied at specified nucleotide positions to enforce

required bases during sampling; combined forward/reverse fixed-logit maps ensured constraints

were respected on both strands. Each sampled nucleotide pair was translated to amino-acid

sequences and duplicate amino-acid solutions were removed prior to scoring.

Structural compatibility was assessed using single-sequence secondary structure prediction

(S4PRED; three-state H/E/C). Secondary structure preservation was reported as the percentage

of residues for which a candidate's predicted state matched the original sequence prediction.

Substitution similarity was computed using BLOSUM62 and normalized to the original

sequence's self-score to yield a percent similarity; global alignment identity was also measured.

ESM-2 (53) (model: esm2_t12_35M_UR50D) was used to generate residue-residue contact

probability matrices, having demonstrated utility as a fast and effective single-sequence

prediction feature (54). Contact map similarity was assessed using the structural similarity index

(SSIM) (55), computed between ESM-2-predicted contact maps of candidate overlap sequences

and their respective originals. SSIM quantifies local and global structural pattern agreement

(range 0-1, with 1 indicating identical maps). Raw values were linearly scaled to a percentage (0-

100) to yield the ESM-2 contact map score. This metric served as a rapid, computationally

efficient proxy for structural preservation, enabling large-scale overlap design iterations while

retaining sensitivity to perturbations in contact map topology.

Candidates were ranked by a fixed weighted composite score across all experiments: 0.15

(secondary structure preservation), 0.15 (substitution similarity), 0.1 (alignment identity), and 0.6

(ESM2 contact map similarity). Although this weighting scheme was not exhaustively

optimized, empirical testing with randomly selected SwissProt proteins indicated that it yielded

stable rankings across experiments and supported overlap generation while balancing structural

fidelity with sequence similarity (**Figs. S3, S4, and S5**). The top-ranked candidate for the

window was integrated, and optimization proceeded to the next window. Multiple full passes

across windows were permitted for refinement. After the first structurally successful candidate,

global feedforward and attention dropout levels were reduced to bias sampling from exploratory

search toward fine-tuning, though the weighting scheme itself remained unchanged.

Efficiency measures implemented for reproducibility and speed included: (i) caching of

secondary structure predictions so identical amino-acid sequences were scored only once, and

(ii) caching of ESM-2 embeddings and contact maps. All candidate metadata (attempt, window,

per-candidate metrics, and secondary structure predictions) were retained in memory and

exported at the end of each input row to permit analysis.

Mutual information, PCA, and robustness testing

Terminal regions (104 amino acids) from 250 randomly selected SwissProt-derived proteins

were used as seeds to generate ensembles of mutated sequences. For each seed, trajectories were

created by stepwise amino acid substitutions sampled from the 20-letter alphabet, introducing

one mutation per step for up to 75 iterations. This yielded a progressive set of variant sequences

per seed protein. Each sequence in the ensemble was then scored for global alignment identity,

ESM-2 contact map similarity, and secondary structure match relative to the reference. These

metric vectors formed the input for subsequent mutual information (MI) and principal

component analyses.

MI was estimated after quantile discretization of continuous scores into 20 bins to evaluate

pairwise dependence among objectives. To reduce sensitivity to binning, MI was also estimated

using a k-nearest-neighbor approach. Statistical significance of discrete MI values was assessed

using permutation testing with 1,000 random shuffles. Principal component analysis (PCA) was

applied to z-scored metric vectors (alignment, ESM-2 contact map similarity, secondary

structure match) to examine the dimensional structure of variation. Robustness checks included

varying the number of bins (5-40) for MI and bootstrap resampling of PCA explained variance.

Statistical Analyses and Figure Preparation

The R programming language (version 4.3.1) and the ggplot2 package (56,57) were used to

perform statistical analyses and generate figures. Prediction success rates are reported as

percentages with exact 95% confidence intervals calculated using the Clopper–Pearson method.

Correlations between similarity metrics were assessed using Pearson's correlation coefficient

with associated p-values. Relationships between secondary structure composition (coil, helix,

sheet) and prediction performance were examined using LOESS smoothing with 95–99%

confidence bands. For external validation experiments, performance metrics were stratified by

AlphaFold2 pLDDT confidence brackets and overlap phase. Group differences were evaluated

using nonparametric pairwise contrasts with adjusted p-values (Bonferroni correction). Effect

sizes are reported as absolute percentage-point differences between groups, alongside confidence

intervals. Data are presented as mean values with ranges or confidence intervals where

appropriate. Figures were generated with ggplot2, with additional overlays and schematics

prepared in Inkscape (version 1.3.2).

Declaration of Generative AI and AI-assisted technologies

During the preparation of this work, ChatGPT (5.0; OpenAI) was used to assess readability and

language, and as a tool to generate code snippets and assist in debugging. After using this

tool/service, the author reviewed and edited the content as needed and takes full responsibility

for the content of the publication.

Code and data availability

The code used for synthetic overlap dataset construction, model training, and inference are

available at: https://github.com/protosome/convergent_overlaps_aa_change

Results

Prediction of convergent overlapping genes from amino acid sequence pairs from a

dedicated transformer-encoder model set

While prior work has established that it is technically feasible to generate overlapping genes

from paired amino acid sequences, the performance of dedicated transformer-encoder models for

this task has not been systematically evaluated. In this study, transformer-based models were

trained to generate convergent overlapping nucleotide sequences from amino acid pairs, and their

ability to recover overlaps across all three phases was assessed. These models were designed to

identify convergent overlaps capable of encoding both input proteins, using recent advances in

transformer-based sequence modeling to address the large solution space introduced by the

degeneracy of the genetic code (31,58). Prediction quality was evaluated using alignment

identity scores and secondary structure preservation metrics, allowing comparison across overlap

lengths from 199 to 312 nucleotides.

Initial filtering was performed using a conservative alignment score cutoff of 34% (28,59).

Without Monte Carlo dropout (ie, dropout enabled during inference), successful predictions were

substantially reduced, and repeated inference runs converged to identical codon solutions. This

indicated that deterministic decoding restricted the search space to narrow, low-diversity

outcomes. By contrast, applying Monte Carlo dropout during inference expanded sequence

diversity, producing multiple distinct codon assignments that encoded the same amino acid

sequences. This stochastic sampling increased the likelihood of recovering overlaps that satisfied

both similarity and structural thresholds, underscoring the importance of uncertainty-aware

inference in navigating the combinatorial complexity of convergent overlap prediction.

To further assess training requirements, the effect of imperfect overlaps and amino acid

substitutions in the training data was examined. Models trained only on unaltered overlaps

exhibited markedly reduced success rates, particularly for phase 1 (**Figs. 3D, 3E**). In a shared test

set of 5,000 amino acid sequence pairs without a known convergent overlap, training with

modified sequences (stop codon removed plus 1–4 substitutions per sequence) substantially

improved prediction rates: phase 0, 95.3% (94.7-95.9%); phase 2, 86.7% (85.7-87.6%); and

phase 1, 81.6% (80.5-82.6%). In contrast, models trained on unaltered overlaps performed

significantly worse: phase 0, 73.7% (72.4-74.9%); phase 2, 49.1% (47.7-50.5%); and phase 1,

9.7% (8.9-10.6%). These findings are consistent with the hypothesis that incorporating

controlled sequence variation during training increased model flexibility and suggest improved

generalization, particularly when coupled with Monte Carlo dropout during inference. On this

basis, models trained with amino acid substitutions were selected for all subsequent analyses.

The outputs of these models were further evaluated using BLOSUM62 and ProtSub substitution

matrices. For overlap lengths of 310-312 nucleotides, similarity scores were strongly correlated

with amino acid identity ($r = 0.88$; $p < 0.0001$), and the matrices themselves were highly

correlated with each other ($r = 0.98$; $p < 0.0001$).

In addition to substitution metrics, alignment scores and secondary structure scores were

compared across phases. Phase 1 overlaps exhibited the highest average alignment and

secondary structure scores, phase 0 displayed intermediate values, and phase 2 was generally the

lowest. However, score distributions overlapped considerably, suggesting that phase-dependent

differences were not absolute. The near-identical secondary structure score distributions for

phase 0 and phase 2 further indicated that structural outcomes may be more sensitive to

sequence-specific context than to phase alone.

To broaden the analysis, overlap lengths from 199 to 312 nucleotides were examined across a

larger set of amino acid pairs. Phase-dependent patterns persisted: phase 1 overlaps were

increasingly recoverable at shorter lengths, and the relative rank order of secondary structure and

alignment scores across phases remained consistent. These findings suggest that phase 1 overlaps

may be predicted to be structurally and codon-compatible favorable, and that performance of the

transformer encoder model set is influenced by overlap length and the diversity of sequences

available for training.

High Secondary Structure Scores Are Linked to Coil-Dominated Precursor Sequences

To assess whether the secondary structure composition of precursor amino acid sequences

influences the predicted quality of convergent overlaps, the proportion of coil (C), β-sheet (E),

and α-helix (H) structures in each input was compared against the model's predicted secondary

structure score. For each amino acid pair, structural proportions were calculated separately and

then averaged across both sequences. This approach reflects the biological premise that overlap

feasibility arises not from a single sequence in isolation, but from the combined structural

constraints and flexibilities of the two proteins that must be co-encoded (60,61).

Across all three overlap phases, sequences with higher coil content exhibited elevated secondary

structure scores (**Fig. 3C**). This trend was most apparent in phase 0 and phase 1, where LOESS-

smoothed curves showed a modest increase in secondary structure score with increasing coil

content, followed by a sharper rise beyond approximately 70% coil. In phase 2, a similar increase

was observed at high coil levels, but the trend was less consistent, with greater variability at

intermediate values. By contrast, α-helix content was inversely associated with secondary

structure score. The most pronounced effect occurred in phase 0, where increasing helix content

corresponded to a marked decline in secondary structure scores. A similar, though less steep,

trend was observed in the other phases. For β-sheet content, no consistent relationship with

secondary structure score was observed; across all three phases, smoothed curves were largely

flat, indicating that sheet content neither strongly promoted nor hindered predicted structural

quality.

To evaluate whether these relationships extended to sequence-level similarity, mean alignment

scores were also plotted against precursor structural compositions. No meaningful trends were

observed for any secondary structure class (**Fig. S1**). Alignment scores remained relatively

constant regardless of coil, β-sheet, or α-helix content, suggesting that precursor structure did not

systematically influence the model's ability to recover the reference overlap sequence.

Multi-objective optimization and computational design of EGFP/AmpR convergent

overlaps

Although the transformer-encoder models were capable of generating convergent overlaps for

paired amino acid sequences, the secondary structure preservation achieved in initial predictions

was variable and generally lower than values reported in prior work (60). To address this

limitation, an iterative sequence optimization algorithm was implemented to refine model

outputs using a multi-objective framework that did not rely on MSA availability (**Fig. 4A**).

The optimization procedure employed a windowed feedforward Monte Carlo (MC) dropout

strategy to generate multiple candidate solutions for each sequence window (**Fig. 4B**). Candidate

overlaps were then evaluated against two complementary structural criteria: 1) predicted

secondary structure states derived from S4PRED (62), providing local fold-relevant constraints,

and 2) ESM-2 contact map similarity (63), which encodes sequence-level context that may

reflect long-range residue dependencies. A key feature of the framework was the ability to

enforce selective preservation of amino acid residues when required. This was achieved by

applying a feedforward mask to specific codon positions, constraining the model to maintain

defined residues (for example, active-site motifs) while permitting synonymous variation

elsewhere. In practice, this allowed critical sequence features to be retained while enabling

exploration of codon-level degeneracy to optimize overlap formation.

Relationships among the chosen objectives were next quantified using mutual information (MI)

and principal component analysis (PCA). Both approaches indicated that alignment identity,

ESM-2 contact map similarity, and predicted secondary structure capture overlapping but distinct

constraints. Discrete MI (20 quantile bins) showed that alignment carried moderate information

about both ESM-2 contact map similarity (MI = 0.393) and secondary structure match (MI =

0.381), whereas the MI between ESM-2 contact map similarity and secondary structure

similarity was lower (MI = 0.240), consistent with partial independence. Continuous MI

estimates (kNN regression) produced similar results (0.509, 0.508, and 0.340, respectively). All

associations were highly significant in permutation tests (p = 0.001). PCA of z-scored metrics

supported this interpretation (**Fig. S2**). A single axis (PC1) explained 72.4% of variance (95%

CI: 71.8-73.0) and loaded positively on all three objectives, reflecting a general similarity

dimension. PC2 (18.4%; 95% CI: 18.0-18.9) contrasted ESM-2 contact map similarity with

secondary structure similarity, while PC3 (9.2%) primarily captured sequence-only variance.

Thus, although alignment dominates the shared signal, secondary structure preservation and

ESM-2 contact map-based similarity provide orthogonal information that cannot be reduced to

alignment alone. This justifies their inclusion as separate objectives during overlap optimization.

Metric behavior under progressive sequence divergence was also assessed by subjecting 250

randomly sampled SwissProt proteins to iterative single-residue substitution trajectories (75 steps

per sequence). Across trajectories, all four similarity measures declined with increasing

mutational distance, but the rate and pattern of decay differed by metric (**Fig. S3**). Alignment

identity and BLOSUM62 similarity decreased rapidly and near-linearly, consistent with their

direct dependence on residue-level identity. By contrast, ESM-2 contact map similarity and

secondary structure similarity showed more gradual declines. These distinct decay profiles

support the interpretation that the objectives capture complementary aspects of overlap fidelity.

Weighting experiments further clarified the contributions of each objective. When the relative

weights assigned to secondary structure similarity, ESM-2 contact map similarity, alignment

identity, and BLOSUM62 substitution similarity were varied across randomly selected SwissProt

pairs with very high AlphaFold2 pLDDT values ($\geq$90), distinct trade-off dynamics were

observed (**Fig. S4**). Increased weighting on secondary structure similarity consistently improved

secondary structure preservation but reduced alignment and substitution similarity, whereas

embedding-focused weightings enhanced ESM-2 contact map similarity at the expense of

secondary structure similarity. Intermediate combinations (eg, 0.15-0.4 secondary structure with

0.6 ESM-2 contact map similarity) yielded more balanced outcomes across the four metrics.

Variance patterns indicated that secondary structure weighting stabilized optimization

trajectories, while ESM-2 contact map similarity remained comparatively variable across

candidate sequences.

A convergent overlap between enhanced green fluorescence protein (EGFP) and a TEM-1 β-

lactamase (ampicillin resistance marker from pCVD004; hereafter AmpR) was designed and

refined under the same objectives as a biologically relevant test case (64,65). The combination of

MC dropout sampling, secondary structure prediction, and embedding-based scoring consistently

improved fold preservation of computationally designed EGFP/AmpR 311 nucleotide (phase 1)

overlaps (**Fig. 4C; Fig. 5**). Compared to unoptimized outputs (ie, sequences generated from the

first window), optimized sequences displayed higher secondary structure similarity rates and

increased ESM-2 contact map similarity, demonstrating that overlap designs can be improved

using lightweight, MSA-independent objectives. During windowed optimization, alignment and

BLOSUM62 metrics remained largely constant or decreased across windows, reflecting their

role in constraining candidate sequences to a biologically plausible subset of sequence space. In

contrast, secondary structure similarity generally increased throughout optimization, despite

being weighted at only 0.15 in the composite score. ESM-2 contact map similarity increased

modestly in the early windows before plateauing. These findings suggest that fold-relevant

secondary structure metrics may act as the principal discriminative driver of optimization

progress, with alignment- and embedding-based metrics providing a stabilizing baseline.

Following optimization, a pair of amino acids with the highest combined score was selected for

structural comparison using predictions from AlphaFold3 (50), while selecting amino acid

sequences in both pairs for preservation based on predicted importance (eg, active site). The

AlphaFold3-predicted models of designed overlaps were highly similar to reference predictions

(**Fig. 4C**), with TM-scores of 0.98 (EGFP) and 0.90 (AmpR), the latter with deviations generally

localized to a disordered coil not located in the overlapping terminal region (66).

Performance of this algorithm in predicting overlapping genes using pLDDT as an

orthogonal proxy for intrinsic structure rigidity

Analyses on prokaryote-derived amino acid sequence pairs indicated that secondary structure

preservation increased with coil content and decreased with helix content, with sheet content

showing no consistent effect. To determine whether this pattern reflected a generalizable

property of overlap design for convergent overlap derived from terminal amino acid sequences,

an external validation set was assembled from SwissProt proteins stratified by AlphaFold2 (AF2)

per-residue confidence (pLDDT), an orthogonal proxy for intrinsic structural rigidity (52,67–69).

SwissProt sequences were filtered to 105–200 amino acids to align with the 105-AA receptive

field and to bound computational cost, then binned into Low-intermediate (50-69), Confident

(70-89), and Very high (90-100) pLDDT groups. For each bracket, 100 non-redundant amino-

acid pairs were formed, and convergent overlaps were designed across phases 0, 1, and 2 at 310,

311, and 312 nt using the same transformer-encoder, windowed Monte-Carlo (MC) dropout

inference, objective weights, and constraint handling described previously.

Quantitative analysis of secondary structure composition (determined using S4PRED) across

pLDDT confidence bins revealed clear trends. Looking specifically at the terminal AA regions,

as pLDDT confidence increased from the Low-intermediate to the Very high bins, coil content

declined substantially (from ~64% to ~41%), while both helix and sheet content rose (helix from

~29% to ~39%; sheet from ~6% to ~20%) (**Figs. 6A, 6B**). These shifts suggest that AlphaFold's

higher-confidence regions correspond generally to more ordered structural states, and reinforce

the interpretation of pLDDT as an approximate proxy for structural order and rigidity (50,70,71).

These AF2-stratified findings were consistent with the earlier composition-based analysis using

prokaryotic ORFs in this study. Sequence pairs with lower pLDDT (coil-enriched) were

associated with slightly higher secondary structure preservation, whereas higher-confidence

inputs tended to show reduced variation (**Fig. 6C**). Nevertheless, the overall high secondary

structure scores across all three brackets emphasize that the algorithm was able to recover

structurally faithful overlaps even for proteins predicted by AlphaFold2 to adopt stable, well-defined folds. Alignment identity and substitution similarity remained generally stable across brackets, indicating that secondary structure preservation was the primary dimension along which flexibility exerted its effect.

Bracket- and phase-specific effects were evident across all three metrics, consistent with the secondary structure composition of the pLDDT bins. Across overlap phases, sequences in the Low-intermediate (50-69) bracket generally outperformed those in the Confident (70-89) and Very high (90-100) brackets, with the gap being most pronounced in phase 2 (**Fig. 6D**). In phase 1, the bracket effect was clear for secondary structure preservation and for the combined score relative to Very high (90-100), whereas pairwise differences were not evident for ESM-2 contact map similarity; Low-intermediate (50-69) also did not differ from Confident (70-89) for the combined score. Overall, phase 1 maintained the highest preservation, while phase 2 exhibited the largest bracket separations. Although group differences were statistically significant, effect sizes varied by metric: within a phase, combined and ESM-2 contact map similarity differences were typically ~1-4 percentage points, whereas secondary structure differences reached ~5-8 points in some phase 2 contrasts. All groups maintained mean secondary structure preservation above 85%. Full pairwise estimates with confidence intervals and adjusted p-values are provided in **Supplementary Tables 4 and 5**.

While statistically significant differences were observed between pLDDT brackets, these differences were relatively small, and overall secondary structure preservation remained consistently high across all conditions (**Fig. 6E**). These results reinforce the generalizability of the earlier coil-associated trend, but also highlight the robustness of the multi-objective,

uncertainty-aware optimization framework in achieving high predicted structural preservation

regardless of precursor rigidity.

Discussion

Overlapping gene architectures have long been recognized as both a source of evolutionary

constraint and an avenue for novel functionality (72–80). In addition to their evolutionary

importance, overlapping structures can be purposefully engineered under realistic codon and

structural constraints. The present study shows that phase-specific convergent overlaps can be

computationally generated and optimized using a transformer encoder-based multi-objective

framework without requiring full 3D prediction at the optimization stage; targeted 3D checks

were applied post hoc. This framework offers a complementary tool that could expand the range

of strategies available for synthetic overlap design, particularly where phase, length, and

nucleotide-level control are critical.

Recent work has demonstrated that deep generative protein models can discover viable

overlapping solutions under genetic code constraints. Byeon et al. (2025) showed that pretrained

amino acid-space generative models can design synthetic overlapping genes across multiple

reading frames, validating expression experimentally using Gibbs sampling with codon-

compatibility constraints (30). Xu et al. (2025) further demonstrated that pretrained generative

protein language models (ESM-3) can yield viable entangled protein pairs through structure-

conditioned inverse folding and CAMEOS-based entanglements, filtered by cross-entropy and

Potts energy scores (81). Importantly, Xu et al. established that pretrained generative models are

capable of identifying functional overlap-compatible solutions, albeit in a limited experimental

context (the InfA/AroB system) and without explicit codon-level optimization.

The framework presented here builds on this momentum but takes a different route. Rather than

relying on pretrained amino acid-space sampling, it employs a purpose-built, phase-generalizable

transformer encoder model set trained on a balanced synthetic dataset of convergent overlaps

spanning diverse GC contexts and overlap lengths. Multi-objective optimization is integrated

directly into the inference loop, with candidate overlaps iteratively refined through a windowed,

uncertainty-aware (MC dropout) search. Evaluation criteria span predicted secondary structure

preservation (S4PRED), amino acid substitution similarity (BLOSUM62), pairwise alignment

identity, and ESM-2 contact map similarity. This approach emphasizes codon-level control while

maintaining structural fidelity, enabling overlap recovery without requiring full 3D structural

prediction at each iteration.

MC dropout was retained during inference to enable stochastic sampling of synonymous codon

solutions, thereby broadening the effective search space (35). Candidate overlaps were ranked

using a fixed composite score that balanced structural preservation, substitution similarity,

alignment identity, and embedding-based metrics. This weighting scheme served as a practical

approach for navigating competing design constraints without requiring full structural prediction

at each iteration, and was most closely aligned with prior work arguing that synthetic overlap

design requires explicit balancing of competing biological constraints and objectives (5,60).

While other multi-objective optimization strategies could be considered in future work (such as

ε-constraint methods (82,83), adaptive scalarization (84,85), or evolutionary multi-objective

optimizers (86)), the fixed weighted framework proved sufficient to achieve high overlap fidelity

under the conditions evaluated.

The synthetic convergent overlapping gene dataset construction method developed as part of this

study provides a structured framework for exploring overlapping gene design in silico, as it

permits generation of diverse and uniformly distributed overlaps. However, one potential

limitation of this approach is the reliance on synthetic, rather than natural, overlapping genes for

model training, which were generated using ORFs from a diverse set of prokaryotic strains. This

may introduce a bias affecting the predictive ability and variability in sequence output. Despite

this concern, the method employed to generate synthetic overlapping genes is in principle

analogous to the formation of new overlaps through the process of gene extension following the

loss of a stop codon (8). This results in synthetic overlapping genes where the second (extended)

strand's encoded amino acid composition is derived from the reverse complement of the first

strand, without undergoing further selection. As recently reported, there is extensive evidence

suggesting functionality in prokaryotic convergent (antisense) proteins, indicating that non-

coding RNA regions could also encode functional proteins (87). For same-strand overlaps, a

significant difference in overall composition compared to non-overlapping genes has been

previously observed (88), along with a bias towards disorder-promoting amino acids (89). The

extent to which this affects convergent overlapping genes remains unclear. Future work will

therefore aim to (i) characterize compositional differences between natural and synthetic

overlaps, (ii) extend design to longer proteins beyond the 199-312 nucleotide range considered

here, and (iii) experimentally validate whether in silico-designed overlaps from this framework

maintain expression, folding, and function in vivo.

In summary, this study establishes that phase-aware transformer models trained directly at the

nucleotide level, when coupled with uncertainty-aware inference and multi-objective

optimization, can recover computationally designed convergent overlapping genes under realistic

genomic constraints. By integrating codon-level sampling with structural and substitution-based

objectives, this framework complements recent amino acid-space generative approaches while

offering length- and phase-specific control. These findings provide a computational foundation

for experimentally testable designs and point toward a scalable strategy for compact, synthetic

gene architectures.

Fig 1. **Depiction of convergent overlapping genes in three reading frames and the process**
developed to computationally design synthetic convergent overlaps from coding sequences.

(A) Sequences are represented for convergent (tail-to-tail) and unidirectional (same-strand)
overlapping genes in each of the three reading frames (phase 0, phase 1, and phase 2). Note,
phase 0 unidirectional overlaps are not represented as these are considered as an alternative start
site for the same gene (90). In each phase, the reading frame was shifted by one nucleotide. Start
and stop codons are indicated with green and red shading, respectively. (B) The general process
for creating synthetic convergent overlapping genes is presented, starting with a pool of ORF
sequences filtered by length (≥450 base pair [bp]). The primary sequence and in-frame 105-bp
fragment were randomly selected. A randomly selected stop codon from the reverse complement
(rc), and the fragment reverse complement was extended to that stop codon to serve as the
downstream sequence of the secondary sequence. Given the downstream secondary sequence
length, another sequence was randomly selected from the same gene list. A size appropriate
fragment was selected from that gene with length equal to the difference between the desired
gene length and the upstream overlap fragment (ie, the reverse complement of the primary
sequence to the selected stop codon). The upstream fragment was then combined with the
downstream fragment to generate the complete secondary sequence. The resultant primary and
secondary nucleotide sequences, sharing a known convergent overlapping region, were then
translated to amino acid (aa) sequences (either modified or unmodified) for subsequent use in
model training or inference.

Fig 3. Training data evaluation of secondary structure preservation and convergent
overlap-phase performance.

(A, D) Results for unmodified amino acid sequences. (A) Scatter density plot of secondary
structure (SS) score versus alignment score, with contours and marginal density plots showing
the distribution by overlap phase (0 = red, 1 = green, 2 = blue). (D) Fraction of successful
predictions across overlap lengths (310, 311, and 312 nucleotides), stratified by phase. Success
rates are reported with exact 95% Clopper–Pearson confidence intervals. (B, E) Results for
modified amino acid sequences. (B) Scatter density plot of SS score versus alignment score with
the same phase-coloring scheme. (E) Fraction of successful predictions across overlap lengths
for modified sequences, stratified by phase. Success rates are reported with exact 95% Clopper–

Pearson confidence intervals. (C) Relationship between sequence-level SS composition (coil [C], sheet [E], helix [H]) and mean SS score across phases 0, 1, and 2. Dashed lines indicate LOESS regression fits with 99% confidence bands.

A.

EGFP Sequence (terminal sequences bolded):
MVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYGKLTLKFICTTGKLPVPWPTLVTTLTYGVQCFSRYPDHMKQ
HDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTLVNRIELKGIDFKEDG**NILGHKLEYN[Y]NS[H]NVYIMADKQ
KNGIKVNFKIRHNIEDGSVQLADHYQQNTPIGDGPVLLPDNHYLS[T]Q[S]ALSKDPNEKRDHMVLL[E]FVTAAG
ITLGMDELYK**

AmpR Sequence (terminal sequences bolded):
MSIQHFRVALIPFFAAFCLPVFAHPETLVKVKDAEDQLGARVGYIELDLNSGKILESFRPEERFPMMSTFKVLLCGAVLSRIDA
GQEQLGRRIHYSQNDLVEYSPVTEKHLTDGMTVRELCSAAITMSDNTAANLLLTTIGGPKELTAFLHNMGDHVTRLDRWEP
**ELNEAIPNDERDTTMPAAMATTLRKLLTGELLTLASRQQLIDWMEADKVAGPLLRSALPAGWFIA[DKSGAG]ERGSR
GIIAALGPDGKPSRIVVIYTTGSQATMDERNRQIAEIGASLIKHW**

> **Input sequences**
> The terminal 104 amino acids from both are extracted and concatenated to define the input sequences. Brackets define positions to retain.

> **Transformer encoder model inference, sequence evaluation, and optimization**
>
> > **Pass 1 (initial window):**
> > Generate a diverse initial set of overlaps, subject to MC dropout across entire sequence
> >
> > **Subsequent passes/windows:**
> > Generate overlaps subject to localized MC dropout, using pair with highest combined score from that window as the next starting sequence pair
>
> > **Evaluation of AA sequences:**
> > At each stage, candidate overlaps are scored against their native sequence using a composite metric that integrates:
> > - **Secondary structure** (S4PRED)
> > - **Long-range interactions** (ESM-2 contact map)
> > - **Pairwise alignment**
> > - **Substitution scoring** (BLOSUM62)
> > This combined score serves as the objective function guiding both filtering and selection

> **Select AA sequence pair with highest average combined score**

B.

EGFP AA Seq, with convergent overlap

AmpR AA Seq, with convergent overlap

C.

TM-Score: 0.98   TM-Score: 0.90

Fig 4. In silico design of convergent overlap with EGFP and AmpR

Overview of the multi-objective optimization process for EGFP and AmpR proteins using the transformer encoder framework. (A) Schematic of the transformer encoder–based workflow. The terminal 104 amino acids of EGFP and AmpR (across all tested overlap lengths) are concatenated and passed through the model. Outputs are scored by a composite metric integrating alignment, substitution, secondary structure preservation, and long-range interactions. Bracketed amino acids from the input are preserved to enforce positional constraints. The final sequence pair is selected from the output table by the highest average combined score. (B) Multiple-sequence alignment of all output pairs (n=3,624) for EGFP and AmpR across both optimization passes and windows (top to bottom). Diagonal dropout patterns correspond to localized MC dropout applied to the overlap region. Variability reflects bracket-constrained codon degeneracy introduced on the non-fixed partner sequence. (C) Predicted tertiary structures generated with AlphaFold3 for EGFP (left) and AmpR (right). Native and overlapping terminal regions relevant to the 311-nt overlap are highlighted (native in red, overlapped in blue); non-overlapping regions are shown in gray.

Fig 5. Trajectories of composite and component metrics during computational optimization of EGFP/AmpR convergent overlaps

Optimization performance for EGFP and AmpR across two passes, each consisting of 13 windows. Metrics shown include the combined score (top left), which integrates secondary structure (SS), ESM-2 contact map similarity, and alignment components, as well as the individual SS (top right), ESM-2 (bottom left), and amino acid alignment (bottom right) scores. Solid red (EGFP) and blue (AmpR) lines denote mean values, and shaded regions represent the minimum-maximum range across sequences within each window.

Fig 6. Comparison of computational overlap predictions across pLDDT brackets, with structural composition and optimization outcomes

Analyses are based on random sampling of SwissProt protein pairs (n=100 for each bracket) from the AlphaFold2 Protein Structure Database (AF2). (A) Distribution of average pLDDT values for the terminal 104 residues across sampled sequences, with red dashed lines marking the confidence thresholds. (B) Counts of sequences assigned to pLDDT confidence brackets, shown for both total and terminal regions (internal white bars denote terminal regions). (C) Ternary plots of amino acid composition (helix [H], sheet [E], and coil [C]) for the primary and secondary sequence sets, colored by pLDDT bracket (low-intermediate, confident, and very high). Circles with black centers denote the centroid (mean position of all sequences within a

given pLDDT bracket) in ternary space. (D) Optimization performance across convergent overlap phases (0, 1, 2) for Pass 1 (P1) and Pass 2 (P2). Plots show average secondary structure (SS Avg) and ESM-2 contact map similarity (ESM Avg) scores by optimization window, with shaded regions representing the range of values stratified by pLDDT bracket. (E) Density distributions of individual metric scores (SS, ESM, and Alignment) for the highest combined score sequences (primary and secondary) across phases 0, 1, and 2.

References

1. Barrell BG, Air GM, Hutchison CA. Overlapping genes in bacteriophage φX174. Nature. 1976 Nov;264(5581):34–41.

2. Keese PK, Gibbs A. Origins of genes: "big bang" or continuous creation? Proc Natl Acad Sci USA. 1992 Oct 15;89(20):9489–93.

3. Wright BW, Molloy MP, Jaschke PR. Overlapping genes in natural and engineered genomes. Nat Rev Genet. 2022 Mar;23(3):154–68.

4. Grainger DC. The unexpected complexity of bacterial genomes. Microbiology. 2016 July 1;162(7):1167–72.

5. Blazejewski T, Ho HI, Wang HH. Synthetic sequence entanglement augments stability and containment of genetic information in cells. Science. 2019 Aug 9;365(6453):595–8.

6. Opuu V, Silvert M, Simonson T. Computational design of fully overlapping coding schemes for protein pairs and triplets. Sci Rep. 2017 Nov 20;7(1):15873.

7. Sabath N, Graur D, Landan G. Same-strand overlapping genes in bacteria: compositional determinants of phase bias. Biology Direct. 2008;3(1):36.

8. Kingsford C, Delcher AL, Salzberg SL. A Unified Model Explaining the Offsets of Overlapping and Near-Overlapping Prokaryotic Genes. Molecular Biology and Evolution. 2007 Sept;24(9):2091–8.

9. Pallejà A, Harrington ED, Bork P. Large gene overlaps in prokaryotic genomes: result of functional constraints or mispredictions? BMC Genomics. 2008;9(1):335.

10. Zehentner B, Ardern Z, Kreitmeier M, Scherer S, Neuhaus K. Evidence for Numerous Embedded Antisense Overlapping Genes in Diverse *E. coli* Strains [Internet]. 2020 [cited 2024 July 12]. Available from: http://biorxiv.org/lookup/doi/10.1101/2020.11.18.388249

11. Hücker SM, Vanderhaeghen S, Abellan-Schneyder I, Scherer S, Neuhaus K. The Novel Anaerobiosis-Responsive Overlapping Gene ano Is Overlapping Antisense to the Annotated Gene ECs2385 of Escherichia coli O157:H7 Sakai. Front Microbiol. 2018 May 14;9:931.

12. Vanderhaeghen S, Zehentner B, Scherer S, Neuhaus K, Ardern Z. The novel EHEC gene asa overlaps the TEGT transporter gene in antisense and is regulated by NaCl and growth phase. Sci Rep. 2018 Dec 14;8(1):17875.

13. Zehentner B, Ardern Z, Kreitmeier M, Scherer S, Neuhaus K. A Novel pH-Regulated, Unusual 603 bp Overlapping Protein Coding Gene pop Is Encoded Antisense to ompA in Escherichia coli O157:H7 (EHEC). Front Microbiol. 2020 Mar 20;11:377.

14. Behrens M, Sheikh J, Nataro JP. Regulation of the Overlapping *pic/set* Locus in *Shigella flexneri* and Enteroaggregative *Escherichia coli*. Infect Immun. 2002 June;70(6):2915–25.

15. Balabanov VP, Kotova VYu, Kholodii GY, Mindlin SZ, Zavilgelsky GB. A novel gene, *ardD* , determines antirestriction activity of the non-conjugative transposon Tn *5053* and is located antisense within the *tniA* gene. FEMS Microbiol Lett. 2012 Dec;337(1):55–60.

16. Delaye L, DeLuna A, Lazcano A, Becerra A. The origin of a novel gene through overprinting in Escherichia coli. BMC Evol Biol. 2008;8(1):31.

17. Fellner L, Bechtel N, Witting MA, Simon S, Schmitt-Kopplin P, Keim D, et al. Phenotype of *htgA* (*mbiA*), a recently evolved orphan gene of *Escherichia coli* and *Shigella* , completely overlapping in antisense to *yaaW*. FEMS Microbiol Lett. 2014 Jan;350(1):57–64.

18. Graf F, Zehentner B, Fellner L, Scherer S, Neuhaus K. Three Novel Antisense Overlapping Genes in E. coli O157:H7 EDL933. Gilk SD, editor. Microbiol Spectr. 2023 Feb 14;11(1):e02351-22.

19. Cock PJA, Whitworth DE. Evolution of Gene Overlaps: Relative Reading Frame Bias in Prokaryotic Two-Component System Genes. J Mol Evol. 2007 Apr;64(4):457–62.

20. Cock PJA, Whitworth DE. Evolution of Relative Reading Frame Bias in Unidirectional Prokaryotic Gene Overlaps. Molecular Biology and Evolution. 2010 Apr 1;27(4):753–6.

21. Pavesi A. Prediction of two novel overlapping ORFs in the genome of SARS-CoV-2. Virology. 2021 Oct;562:149–57.

22. Nelson CW, Ardern Z, Wei X. OLGenie: Estimating Natural Selection to Predict Functional Overlapping Genes. Tamura K, editor. Molecular Biology and Evolution. 2020 Apr 3;msaa087.

23. Schlub TE, Buchmann JP, Holmes EC. A Simple Method to Detect Candidate Overlapping Genes in Viruses Using Single Genome Sequences. Malik H, editor. Molecular Biology and Evolution. 2018 Oct 1;35(10):2572–81.

24. Chlebek JL, Leonard SP, Kang-Yun C, Yung MC, Ricci DP, Jiao Y, et al. Prolonging genetic circuit stability through adaptive evolution of overlapping genes. Nucleic Acids Research. 2023 July 21;51(13):7094–108.

25. Decrulle AL, Frénoy A, Meiller-Legrand TA, Bernheim A, Lotton C, Gutierrez A, et al. Engineering gene overlaps to sustain genetic constructs in vivo. Braun EL, editor. PLoS Comput Biol. 2021 Oct 8;17(10):e1009475.

26. Wang B, Papamichail D, Mueller S, Skiena S. Two Proteins for the Price of One: The Design of Maximally Compressed Coding Sequences. In: Carbone A, Pierce NA, editors. DNA Computing [Internet]. Berlin, Heidelberg: Springer Berlin Heidelberg; 2006 [cited 2024 July 10]. p. 387–98. (Lecture Notes in Computer Science; vol. 3892). Available from: http://link.springer.com/10.1007/11753681_31

27. Manuel Martí J, Hsu C, Rochereau C, Xu C, Blazejewski T, Nisonoff H, et al. GENTANGLE: integrated computational design of gene entanglements. Elofsson A, editor. Bioinformatics. 2024 June 21;btae380.

28. Wichmann S, Scherer S, Ardern Z. Biological factors in the synthetic construction of overlapping genes. BMC Genomics. 2021 Dec;22(1):888.

29. Leonard SP, Halvorsen T, Lim B, Park DM, Jiao Y, Yung M, et al. Creating overlapping genes by alternate-frame insertion [Internet]. 2024 [cited 2024 Nov 29]. Available from: http://biorxiv.org/lookup/doi/10.1101/2024.11.07.622342

30. Byeon GW, Expòsit M, Baker D, Seelig G. Design of overlapping genes using deep generative models of protein sequences [Internet]. Synthetic Biology; 2025 [cited 2025 Aug 8]. Available from: http://biorxiv.org/lookup/doi/10.1101/2025.05.06.652464

31. Choi SR, Lee M. Transformer Architecture and Attention Mechanisms in Genome Data Analysis: A Comprehensive Review. Biology. 2023 July 22;12(7):1033.

32. Vaswani A, Shazeer N, Parmar N, Uszkoreit J, Jones L, Gomez AN, et al. Attention Is All You Need [Internet]. arXiv; 2023 [cited 2023 Oct 18]. Available from: http://arxiv.org/abs/1706.03762

33. Lin T, Wang Y, Liu X, Qiu X. A survey of transformers. AI Open. 2022;3:111–32.

34. Srivastava, N, Hinton, G, Krizhevsky, A, Sutskever, I, Salakhutdinov, R. Dropout: A Simple Way to Prevent Neural Networks from Overfitting. JMLR. 2014;15(56):1929−1958.

35. Gal Y, Ghahramani Z. Dropout as a Bayesian Approximation: Representing Model Uncertainty in Deep Learning. Proceedings of The 33rd International Conference on Machine Learning. 2016;(48):pp 1050-1059.

36. Lemay A, Hoebel K, Bridge CP, Befano B, De Sanjosé S, Egemen D, et al. Improving the repeatability of deep learning models with Monte Carlo dropout. npj Digit Med. 2022 Nov 18;5(1):174.

37. Seoh R. Qualitative Analysis of Monte Carlo Dropout [Internet]. arXiv; 2020 [cited 2024 July 13]. Available from: https://arxiv.org/abs/2007.01720

759   38. Singer GAC, Hickey DA. Nucleotide Bias Causes a Genomewide Bias in the Amino Acid
760   Composition of Proteins. Molecular Biology and Evolution. 2000 Nov 1;17(11):1581–8.

761   39. Paszke A, Gross S, Massa F, Lerer A, Bradbury J, Chanan G, et al. PyTorch: An Imperative
762   Style, High-Performance Deep Learning Library [Internet]. arXiv; 2019 [cited 2024 July 13].
763   Available from: https://arxiv.org/abs/1912.01703

764   40. TensorFlow Developers. TensorFlow [Internet]. Zenodo; 2023 [cited 2024 Feb 24]. Available
765   from: https://zenodo.org/doi/10.5281/zenodo.10126399

766   41. Zarrieß S, Voigt H, Schüz S. Decoding Methods in Neural Language Generation: A Survey.
767   Information. 2021 Aug 30;12(9):355.

768   42. Graczyk KM, Pawłowski J, Majchrowska S, Golan T. Self-normalized density map (SNDM)
769   for counting microbiological objects. Sci Rep. 2022 June 22;12(1):10583.

770   43. Subramanian K, Payne B, Feyertag F, Alvarez-Ponce D. The Codon Statistics Database: A
771   Database of Codon Usage Bias. Saitou N, editor. Molecular Biology and Evolution. 2022
772   Aug 3;39(8):msac157.

773   44. Cock PJA, Antao T, Chang JT, Chapman BA, Cox CJ, Dalke A, et al. Biopython: freely
774   available Python tools for computational molecular biology and bioinformatics.
775   Bioinformatics. 2009 June 1;25(11):1422–3.

776   45. Moffat L, Jones DT. Increasing the accuracy of single sequence prediction methods using a
777   deep semi-supervised learning framework. Xu J, editor. Bioinformatics. 2021 Nov
778   5;37(21):3744–51.

779   46. Kabsch W, Sander C. Dictionary of protein secondary structure: Pattern recognition of
780   hydrogen-bonded and geometrical features. Biopolymers. 1983 Dec;22(12):2577–637.

781   47. Henikoff S, Henikoff JG. Amino acid substitution matrices from protein blocks. Proc Natl
782   Acad Sci USA. 1992 Nov 15;89(22):10915–9.

783   48. Eddy SR. Where did the BLOSUM62 alignment score matrix come from? Nat Biotechnol.
784   2004 Aug;22(8):1035–6.

785   49. Jia K, Jernigan RL. New amino acid substitution matrix brings sequence alignments into
786   agreement with structure matches. Proteins. 2021 June;89(6):671–82.

787   50. Abramson J, Adler J, Dunger J, Evans R, Green T, Pritzel A, et al. Accurate structure
788   prediction of biomolecular interactions with AlphaFold 3. Nature. 2024 June
789   13;630(8016):493–500.

790   51. Mirdita M, Schütze K, Moriwaki Y, Heo L, Ovchinnikov S, Steinegger M. ColabFold:
791   making protein folding accessible to all. Nat Methods. 2022 June;19(6):679–82.

52. Jumper J, Evans R, Pritzel A, Green T, Figurnov M, Ronneberger O, et al. Highly accurate protein structure prediction with AlphaFold. Nature. 2021 Aug 26;596(7873):583–9.

53. Lin Z, Akin H, Rao R, Hie B, Zhu Z, Lu W, et al. Evolutionary-scale prediction of atomic-level protein structure with a language model. Science. 2023 Mar 17;379(6637):1123–30.

54. Singh J, Litfin T, Singh J, Paliwal K, Zhou Y. SPOT-Contact-LM: improving single-sequence-based prediction of protein contact map using a transformer language model. Martelli PL, editor. Bioinformatics. 2022 Mar 28;38(7):1888–94.

55. Zhou Wang, Bovik AC, Sheikh HR, Simoncelli EP. Image quality assessment: from error visibility to structural similarity. IEEE Trans on Image Process. 2004 Apr;13(4):600–12.

56. Wickham H. ggplot2: Elegant Graphics for Data Analysis [Internet]. New York, NY: Springer New York; 2009 [cited 2024 Mar 22]. Available from: https://link.springer.com/10.1007/978-0-387-98141-3

57. Wickham H, Averick M, Bryan J, Chang W, McGowan L, François R, et al. Welcome to the Tidyverse. JOSS. 2019 Nov 21;4(43):1686.

58. Zhang S, Fan R, Liu Y, Chen S, Liu Q, Zeng W. Applications of transformer-based language models in bioinformatics: a survey. Bateman A, editor. Bioinformatics Advances. 2023 Jan 5;3(1):vbad001.

59. Rost B. Twilight zone of protein sequence alignments. Protein Engineering, Design and Selection. 1999 Feb;12(2):85–94.

60. Wichmann S, Scherer S, Ardern Z. Biological factors in the synthetic construction of overlapping genes. BMC Genomics. 2021 Dec;22(1):888.

61. Lebre S, Gascuel O. The combinatorics of overlapping genes. 2016 [cited 2023 Dec 29]; Available from: https://arxiv.org/abs/1602.04971

62. Moffat L, Jones DT. Increasing the accuracy of single sequence prediction methods using a deep semi-supervised learning framework. Xu J, editor. Bioinformatics. 2021 Nov 5;37(21):3744–51.

63. Lin Z, Akin H, Rao R, Hie B, Zhu Z, Lu W, et al. Evolutionary-scale prediction of atomic-level protein structure with a language model. Science. 2023 Mar 17;379(6637):1123–30.

64. Zhang G, Gurtu V, Kain SR. An Enhanced Green Fluorescent Protein Allows Sensitive Detection of Gene Transfer in Mammalian Cells. Biochemical and Biophysical Research Communications. 1996 Oct;227(3):707–11.

65. Taton A, Unglaub F, Wright NE, Zeng WY, Paz-Yepes J, Brahamsha B, et al. Broad-host-range vector system for synthetic biology and biotechnology in cyanobacteria. Nucleic Acids Research. 2014 Sept 29;42(17):e136–e136.

66. Zhang Y, Skolnick J. Scoring function for automated assessment of protein structure template quality. Proteins. 2004 Dec;57(4):702–10.

67. Alderson TR, Pritišanac I, Kolarić Đ, Moses AM, Forman-Kay JD. Systematic identification of conditionally folded intrinsically disordered regions by AlphaFold2. Proc Natl Acad Sci USA. 2023 Oct 31;120(44):e2304302120.

68. Tunyasuvunakool K, Adler J, Wu Z, Green T, Zielinski M, Žídek A, et al. Highly accurate protein structure prediction for the human proteome. Nature. 2021 Aug 26;596(7873):590–6.

69. Binder JL, Berendzen J, Stevens AO, He Y, Wang J, Dokholyan NV, et al. AlphaFold illuminates half of the dark human proteins. Current Opinion in Structural Biology. 2022 June;74:102372.

70. Abbas U, Chen J, Shao Q. Assessing Fairness of AlphaFold2 Prediction of Protein 3D Structures [Internet]. Bioinformatics; 2023 [cited 2025 Sept 14]. Available from: http://biorxiv.org/lookup/doi/10.1101/2023.05.23.542006

71. Guo HB, Perminov A, Bekele S, Kedziora G, Farajollahi S, Varaljay V, et al. AlphaFold2 models indicate that protein sequence determines both structure and dynamics. Sci Rep. 2022 June 23;12(1):10696.

72. Ardern Z. Alternative Reading Frames are an Underappreciated Source of Protein Sequence Novelty. J Mol Evol. 2023 Oct;91(5):570–80.

73. Cassan E, Arigon-Chifolleau AM, Mesnard JM, Gross A, Gascuel O. Concomitant emergence of the antisense protein gene of HIV-1 and of the pandemic. Proc Natl Acad Sci USA. 2016 Oct 11;113(41):11537–42.

74. Chirico N, Vianelli A, Belshaw R. Why genes overlap in viruses. Proc R Soc B. 2010 Dec 22;277(1701):3809–17.

75. Pavesi A. Origin, Evolution and Stability of Overlapping Genes in Viruses: A Systematic Review. Genes. 2021 May 26;12(6):809.

76. Pavesi A. Asymmetric evolution in viral overlapping genes is a source of selective protein adaptation. Virology. 2019 June;532:39–47.

77. Pavesi A, Romerio F. Extending the Coding Potential of Viral Genomes with Overlapping Antisense ORFs: A Case for the De Novo Creation of the Gene Encoding the Antisense Protein ASP of HIV-1. Viruses. 2022 Jan 14;14(1):146.

78. Safari M, Jayaraman B, Yang S, Smith C, Fernandes JD, Frankel AD. Functional and structural segregation of overlapping helices in HIV-1. eLife. 2022 May 5;11:e72482.

79. Fernandes JD, Faust TB, Strauli NB, Smith C, Crosby DC, Nakamura RL, et al. Functional Segregation of Overlapping Genes in HIV. Cell. 2016 Dec;167(7):1762-1773.e12.

80. Krakauer DC. Stability and Evolution of Overlapping Genes. Evolution. 2000 June;54(3):731–9.

81. Xu C, Chlebek JL, Allen JE, Nisonoff H, Park DM. Generative Protein Design for Overlapping Genes. In OpenReview; 2025. Available from: https://openreview.net/forum?id=35

82. On a Bicriterion Formulation of the Problems of Integrated System Identification and System Optimization. IEEE Trans Syst, Man, Cybern. 1971 July;SMC-1(3):296–7.

83. Miettinen K. Nonlinear Multiobjective Optimization [Internet]. Boston, MA: Springer US; 1998 [cited 2025 Sept 9]. (Hillier FS, editor. International Series in Operations Research & Management Science; vol. 12). Available from: http://link.springer.com/10.1007/978-1-4615-5563-6

84. Marler RT, Arora JS. Survey of multi-objective optimization methods for engineering. Structural and Multidisciplinary Optimization. 2004 Apr 1;26(6):369–95.

85. Das I, Dennis JE. Normal-Boundary Intersection: A New Method for Generating the Pareto Surface in Nonlinear Multicriteria Optimization Problems. SIAM J Optim. 1998 Aug;8(3):631–57.

86. Deb K, Pratap A, Agarwal S, Meyarivan T. A fast and elitist multiobjective genetic algorithm: NSGA-II. IEEE Trans Evol Computat. 2002 Apr;6(2):182–97.

87. Ardern Z, Neuhaus K, Scherer S. Are Antisense Proteins in Prokaryotes Functional? Front Mol Biosci. 2020 Aug 14;7:187.

88. Pavesi A, Vianelli A, Chirico N, Bao Y, Blinkova O, Belshaw R, et al. Overlapping genes and the proteins they encode differ significantly in their sequence composition from non-overlapping genes. Jan E, editor. PLoS ONE. 2018 Oct 19;13(10):e0202513.

89. Rancurel C, Khosravi M, Dunker AK, Romero PR, Karlin D. Overlapping Genes Produce Proteins with Unusual Sequence Properties and Offer Insight into De Novo Protein Creation. J Virol. 2009 Oct 15;83(20):10719–36.

90. Fonseca MM, Harris DJ, Posada D. Origin and Length Distribution of Unidirectional Prokaryotic Overlapping Genes. G3 Genes|Genomes|Genetics. 2014 Jan 1;4(1):19–27.