The "five divergent amino acids rule" and evolution analysis of

cytochrome *c* oxidase as new methods for *Scolopendra* Linnaeus, 1758

species differentiation (Chilopoda. Scolopendromorpha).

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Abstract

Scolopendra Linnaeus, 1758 is a genus of centipede showing reasonably good morpho-

molecular correlation. This often allows the acceptable species differentiation and

phylogenetic positioning by just using the partial sequence of the cytochrome *c* oxidase

(COI) gene. However, for biological, statistical, or technical reasons, several exceptions

to that fact have been observed, which together with a lack of a solid criterion to

differentiate species at the molecular level, make difficult the interpretation of some

taxonomic and systematic outcomes.

With the scope of providing a standardized system improving the molecular

Scolopendra species delimitation and surpassing some issues related with the mtDNA's

use, a total of 45 representative COI sequences belonging to 22 taxa are tested, but for

the first time, performing distinct amino acid's chains evolution analyses. To illustrate

this, a general evaluation of the genus is firstly provided, while to exemplify some of

the deeper analysis, the cases of *S. paradoxa* Doménech, 2018 and *S. spinosissima*

Kraepelin, 1903 are then here re-explored using some alternative tools.

As a result, the partial COI protein sequences analysis identified, at least, ten amino acid

residues' positions as useful for *Scolopendra* species differentiation, being generally

five divergent residues enough to distinguish taxa ["the five divergent amino acid rule"].

When this premise wasn't fulfilled, the amino acid frequency, exclusivity, or

electrochemical properties and especially the *r* substitution probability test, helped solve

the cases. Among *S. paradoxa* and *S. spinosissima* five diverging residues were found,

with one of them being exclusive to the entire genus for the former taxon. Also, the

substitution probability (*r*) analysis showed strong positive selection for three of these

five divergent amino acids.

As exceptions, *S. dawydoffi* Kronmüller, 2012 has been found indistinguishable from *S.*

multidens Newport, 1844 by this method, suggesting its eventual synonymy, while *S.*

cataracta Siriwut, Edgecombe & Panha, 2016 showed an intraspecific maximum

divergence of 5 amino acids. Finally and at nomenclatural level, the species name *S.*

hainanum Kronmüller, 2012 is emended as *S. hainanensis* while the taxon *S. mojiangica*

Zhang & Chi, is here declared *nomen nudum*, due its original description remains

unavailable.

Introduction

The class Chilopoda is among the oldest groups of terrestrial arthropods (Edgecombe &

Giribet 2007). These soil dweller invertebrates' (Undheim & King 2011; Undheim et al.

2015) are venomous predators widely distributed across the warmer and tropical

ecosystems. They are classified into five distinct orders, which currently include a total

of 3,300 described taxa (Edgecombe & Giribet 2007). Among the 700 valid species of

the order Scolopendromorpha, the most recognizable genus is *Scolopendra* Linnaeus,

1758, which currently comprises around 100 accepted members (Bonato et al. 2016;

Schileyko et al. 2020).

Like many other metazoans and since their first descriptions, species of this

genus have undergone multiple taxonomic and nomenclatural changes (see, for

example, Linnaeus 1758; Kraepelin 1903; Attems 1930; Kronmüller 2012). However,

these classical classification systems, based mainly on morphological and geographical

differences, have been significantly improved by integrating them with more modern

molecular techniques. As a paradigm of this progress, cytochrome *c* oxidase (COI) has

proven to be a reliable bioindicator capable of establishing consistent morpho-molecular

phylogenies across various groups. These have helped resolve some of the difficult

relationships within the genus *Scolopendra* (Edgecombe & Giribet 2008; Vahtera et al.

2012, 2013; Oeyen et al. 2014; Siriwut et al. 2015). Likewise, through the quantitative

analysis of genetic distances in partial COI sequences, it has also been possible to

distinguish species with a reasonable degree of confidence (Siriwut et al. 2016;

Doménech et al. 2018). However, despite generally working well, these methods do not

always reflect an expected correlation between the morphology of all taxa and their

percentage of divergence or phylogenetic position. For example, some studies have

revealed that two congeners, despite being morphologically very different, appear to be

more molecularly similar to each other than to their morphologically closer relatives. In

some cases, even species placed in different genera turned out to be more similar to

each other at the molecular level than to their actual congeners (Vahtera et al. 2012,

2013; Vahtera & Edgecombe 2014; Siriwut et al. 2016; Doménech et al. 2018).

Three main reasons could explain these occasional inconsistencies: first,

technical issues that may affect the preprocessing of samples, PCR, and/or DNA

sequencing, which could ultimately compromise sequence quality. The second reason

relates to the statistical methodology used to carry out the systematic analysis. These

tests may yield significantly different results depending on several factors, such as the

number and length of sequences used, the number and type of species or genera studied,

the alignment method, the statistical approach, or the number of applied replicates and

biases caused by certain bioinformatic phenomena such as long branch attraction (Susko

et al. 2021). However, the third and most important reason may lie in various biological

causes, such as the accumulation of neutral mutations, their saturation, or differing

nucleotide substitution rates (heterotachy) in the COI sequences of each species

(Kimura 1968, 1983; Zuckerkandl & Pauling 1962; Xia 2009).

Furthermore, molecular studies of the genus have generally been conducted

partially; this is by dividing analyses by geographical region (see Oeyen et al. 2014;

Siriwut et al. 2015, 2016; Doménech et al. 2018). This adds to the issue, as there is no

clearly defined threshold to distinguish intra- from interspecific levels for species of the

genus *Scolopendra*. As a result, it is sometimes observed that the same percentage of

molecular divergence between two individuals can be considered either intra- or

interspecific depending on the source taken as reference (compare Oeyen et al. 2014;

Siriwut et al. 2015, 2016; Doménech et al. 2018). Lastly, when comparing different

species, this lack of consensus can also occasionally make it difficult to coherently

reconcile morphological results with various molecular studies.

On the other hand, molecular studies on COI have generally focused on genetic

positioning in trees and complemented by the percentage analysis of genetic

divergences. These studies, without exception, have always been based on nucleotide

sequences. However, phenotypic determination—and consequently, the external

morphology—is given by the proteins translated from these and other genes, with the

peptide chain carrying the "true" evolutionary weight as it determines the adaptation or

elimination of individuals in a specific environment. The peptide chains of the partial

COI sequence have not been studied taxonomically, yet they may offer certain

advantages over nucleotide sequences. Quantitatively, amino acid chains, being two-

thirds shorter than nucleotide chains, could facilitate faster reading and interpretation of

different alignments. Likewise, the analysis of amino acid substitutions, where fewer

changes are expected compared to nucleotide analysis (Kimura 1968, 1983), could be

easier to analyze. This might translate into the use of whole numbers instead of

divergence percentages obtained through more complex statistical methods subject to

other statistical biases. Qualitatively, amino acid chains are more diverse than

nucleotides (20 possible residues vs. only four possible bases), which allows easier

discrimination of changes; making it possible to observe whether these changes are

exclusive to a species or a group of species. These changes, involving electrochemical

and structural modifications in the protein, can be measured, providing new differential

data between sequences. Similarly, amino acid changes carry greater evolutionary and

taxonomic significance than nucleotide changes, as they tend to be more inherently

discriminative (Kimura 1968, 1983). Finally, protein studies would present much less

bias and "noise" than nucleotide-based studies, as they would be minimally influenced

by factors such as neutral mutations or substitution saturation, potentially offering

molecular divergences more consistent with morphology.

Not being the main objective of this study to discern the probable causes

generating the morphomolecular incongruences observed in previous articles based on

mtDNA (Vahtera et al. 2012, 2013; Vahtera & Edgecombe 2014; Siriwut et al. 2016;

Doménech et al. 2018), the present work analyzes for the first time the advantages and

disadvantages of using translated partial COI sequences when applied to the systematics

of the genus *Scolopendra*. To illustrate this, the translated partial COI fragment from a

representative sample set of 21 *Scolopendra* species will be compared under this

alternative molecular perspective. First, general results obtained from various

bioinformatic tests will be presented, along with general conclusions for discriminating

between different species of the genus. Next, as a guide, the methodology for resolving

cases of difficult interpretation from an evolutionary standpoint will be outlined,

exemplified by two species recently redefined by this research group — *S. spinosissima*

Kraepelin, 1903 and *S. paradoxa* Doménech, 2018 — (Doménech et al. 2018;

Doménech 2024). Finally, a simple summary scheme will be provided for the easy

application of the aforementioned methods. The work will conclude with a discussion of

the pros and cons of using these new methods and will present some taxonomic and

nomenclatural observations involving some of the species studied here.

Methods

Pre-processing of materials, nomenclatural considerations, and image treatment.

To elongate the previously trimmed terminal ends and reaffirm their suitability, the

sequences of *Scolopendra spinosissima* Kraepelin, 1903 (KY888682.1), *S. paradoxa*

Doménech, 2018 (MH087425.1, KY888684.1, KY888685.2, KY888686.2), and *S.*

subscrustalis Kronmüller, 2009 (KY888683.2) were re-sequenced following the

protocol outlined in Doménech et al. (2018).

The specific epithet of the taxon *Scolopendra hainanum* Kronmüller, 2012 ***nom.***

imperfect. is considered an original incorrect spelling (Art. 32.5 ICZN) due to

grammatical gender disagreement (Art. 11.9.1.3, 31.1.2, 31.2, 32.2, and 32.5 ICZN).

Therefore, the name is justifiably emended here (Art. 19.2 ICZN) and is consistently

presented throughout the text as *S. hainanensis* ***nom. correct***.—in agreement with the

species' author, C. Kronmüller; May/June 2020, pers. comm. Likewise, the *taxon S.*

mojiangica attributed to Zhang & Chi is here considered a ***nomen nudum*** (Art. 12 and

14, ICZN) as its original description is unavailable (NCBI: txid2023220). Since

morphologically this species is not clearly distinguishable from other Asian congeners,

this taxonomic unit is provisionally kept separate from the rest to avoid attribution bias.

The creation and editing of images (contrast and brightness adjustment, notes

and references in illustrations) were carried out using Adobe Photoshop CS6.

Bioinformatic analysis of partial COI sequences.

A total of 383 COI sequences assigned to the genus *Scolopendra* were retrieved from

the BOLD V4 database (Ratnasingham & Hebert 2007). The partial COI sequences

were batch-translated into proteins using the getorf program from the EMBOSS tools

(Rice et al. 2007), applying the invertebrate mitochondrial genetic code and discarding

ORFs with fewer than 200 residues. To align the selected partial COI sequences

translated into proteins, the ClustalW program implemented in BioEdit (Hall 2011) and

MEGA X was used (Figure 1A, Supplementary files 1–3). The protein alignments were

manually checked, discarding sequences with gaps, ambiguous residues, or those that

did not align properly. Subsequently, some representative sequences from each species

were selected proportionally to the number of available sequences and with the greatest

possible geographical/geological distance between them. The taxonomic identity of the

selected sequences was verified or modified according to current nomenclature. The

divergence table of the selected protein products was generated using MEGA X

(Supplementary file 4).

The amino acid substitution probability (r) (Table 1) was calculated using a

trimmed alignment of 41 partial COI protein sequences and a substitution matrix

estimated and implemented in MEGA X (Kumar et al. 2018). The sequence logos

(Figure 1B) were generated using the WebLogo online tool

(https://weblogo.berkeley.edu/logo.cgi). Amino acid frequencies were calculated using

MEGA X, and the heat map of amino acid frequencies along with hierarchical clustering

(HCL) was generated using MeV software (Multiple Experiment Viewer,

www.mev.tm4.org) (Figure 1C). Amino acid sequence changes were evaluated with

reference to the haplotype of the translated partial sequence belonging to the holotype of

S. paradoxa (S. paradoxa.CEUA017-MR0000).

194 The statistical selection of the models that best fit the amino acid substitution

195 study was performed using the Find Best Substitution Model tool implemented in

196 MEGA X, assuming the lowest AICc scores (corrected Akaike Information Criterion).

197 Evolutionary analysis was carried out in MEGA X. The maximum likelihood (ML)

198 phylogenetic tree (Guindon et al. 2010) was constructed using the mitochondrial general

199 reversible model (mtREV) (Le et al. 2017) with 1000 bootstrap replicates (Figure 2). A

200 consensus tree was subsequently obtained and rooted with selected outgroups. In the

201 final stage, node support was evaluated, and genetic affinities were discussed in relation

202 to current nomenclature and taxonomy.

203 The summary diagram outlining the intra- and interspecific boundaries of COI

204 partial sequences translated into proteins for the genus *Scolopendra* was created using

205 PowerPoint (Microsoft Office 365) (Figure 3).

206
207 **Abbreviations**
208
209 **Institutions:**
210 **BMB-DENR** - Biodiversity Management Bureau - Department of Environment and
211 Natural Resources, Philippines.
212 **CEUA**- Colección Entomológica de la Universidad de Alicante San Vicent del Raspeig,
213 Alacant, Spain.
214 **PAE** - Philippines Association of Entomologists, Inc., Philippines.
215 **PNM** - Philippine National Museum, Philippines.
216 **PNU** - Philippine Normal University, Philippines.
217 **WRD-DENR** - Wildlife Resources Division - Department of Environment and Natural
218 Resources, Philippines.
219
220 **Molecular Section:**
221 **A**, alanine; **COI**, cytochrome *c* oxidase; **E**, glutamic acid; **L**, leucine; **M,** methionine; **N**,
222 asparagine; **S**, serine; **T**, threonine.
223

Others: ICZN; International Code of Zoological Nomenclature.

Results

Sequence Filtering

The first step in the bioinformatic analysis was to approach *bona fide* sequences and

eliminate those with the evident errors. To do so, strict quality control criteria were

applied to the sequences extracted from the BOLD database (see Materials section).

Briefly, of the 383 sequences initially obtained, 22 sequences were discarded as they did

not actually belong to the genus *Scolopendra*. Of the "29" *Scolopendra* species initially

available, 14 sequences were removed for having fewer than 200 residues, including

two representative species: *S. cf. amazonica* Bücherl, 1946 and *S. leki* (Waldock and

Edgecombe, 2012). Additionally, another 17 sequences were eliminated for being non-

translatable into a coding protein. A third filter removed 61 sequences and five more

species (*S. lufengia* Kang, Liu, Zeng, Deng, Luo, Chen & Chen, 2017; *S. nigrocapitis*

Zhang & Wang, 1999; *S. mojiangica*; *S. valida* Lucas, 1840; and the remaining

sequences of *S. cretica* Lucas, 1853), as they yielded defective protein alignments.

Finally, from the extracted dataset, only a total of 269 sequences were deemed

suitable, from which 45 sequences belonging to 21 different species were selected

(Supplementary File 1).

Partial COI protein sequences variation in the genus *Scolopendra*.

Although we have found a high amino acid identity regions in our *Scolopendra* partial

protein COI genes' alignment, we have identified ten residues' sites that present a great

variability in the relative position number: 21, 24, 52, 104, 105, 112, 115, 129, 131 and

176; being also the relatives positions 249, 255, 256 and 265 other probable amino acid

variable hot spots (Fig. 1B, Supplementary files 2 and 3).

250 Constantly, determinate amino acid combinations in the proteins' variable

251 fraction have being observed at intraspecific and interspecific levels. These let to

252 establish differences between species with a quite elevate degree of confidence As a

253 general rule, when comparing two sequences, five or more amino acid residue changes

254 in the studied COI fragment (equivalent to 2.2% or more) indicate an interspecific range

255 between the two amino acid chains. However, as an exception to this clear

256 intra/interspecific division — equivalent to a mitochondrial DNA divergence gap —

257 only 14 exceptions were observed among the 820 alignments analyzed. Therefore, this

258 simple method of counting divergent amino acids is capable of distinguishing different

259 species of the genus *Scolopendra* with an approximate accuracy of 98.29%

260 (Supplementary file 4)

261 At the intraspecific level, all alignments uniformly showed between zero and

262 four amino acid changes. The only exception was found in the alignment between the

263 holotype of *S. cataracta* from Laos and a conspecific individual from China, which

264 showed five residue substitutions. Additionally, although not counted here to avoid

265 potential attribution/sequencing biases, two singular cases were detected far beyond

266 these general observations: 1) *S. subspinipes* Leach, 1816, despite aligning correctly,

267 showed either only two or nine changes respectively, depending on whether the unique

268 CUMZ sequence was used or not; and 2) *S. pinguis* Pocock, 1891, a species under

269 suspected cryptic speciation (Siriwut et al. 2015, 2016), which exhibited an extremely

270 unusual range of between two and fourteen amino acid variations in the studied COI

271 fragment (Supplementary file 4).

At the interspecific level, all alignments consistently showed five or more

substitutions (hereafter referred to as the "5 divergent amino acid rule") (Supplementary

file 4). However, two different types of exceptions were detected: 1) Presumed distinct,

but morphologically very similar species, with all their alignments falling within the

intraspecific range (four or fewer divergent residues), such as *S. multidens* Newport,

1844 / *S. dawydoffi* Kronmüller, 2012, which consistently aligned with zero to three

amino acid substitutions; and 2) Clearly distinct species whose respective alignments

fell within both the intra- and interspecific range (all with between four and five

substitutions): *S. cataracta* / *S. hainanensis*, *S. lufengia* / *S. dawydoffi* and *S. cingulata*

when aligned with *S. lufengia* and *S. cataracta*. In the interspecific range, no

divergences involving zero to three residue substitutions were observed between

morphologically distinct species, leading to the conclusion that this number of changes

occurs exclusively within the intraspecific range. Moreover, without exception, all

morphologically distinct species with overlapping substitutions in the intra/inter range

consistently had at least one alignment with four or five, or more divergent residues,

depending on the sequences analyzed. This suggested that when this rare ranges overlap

occurs, the analyzed taxa consistently support the presence of two distinct species

(Supplementary file 4).

Moreover, upon examining the amino acid sequence alignments (Supplementary

files 2 and 3), it was also found that 12 out of the 21 *Scolopendra* species had one or

more unique residues in their sequence. These exclusive variations were observed

consistently in species represented by multiple sequences, suggesting that the presence

of such unique amino acids is, by itself, diagnostic of the species possessing them.

Other amino acids were shared only by specific groups of morphologically related

species, such as *S. subcrustalis* Kronmüller, 2009 and *S. laeta* Haase, 1887, or by

morphologically and geographically distant species such as *S. mutilans* L. Koch, 1878,

S. polymorpha Wood, 1861, and *S. viridis* Say, 1821, indicating a character shared by a

group of species. When these group-characteristic but not species-exclusive amino acids

were found, they were also observed to be useful for distinguishing species—especially

when the taxa were not morphologically and/nor geographically related.

On the other hand, in terms of amino acid composition, the different translated

partial COI protein sequences also showed significant similarity among *Scolopendra*

species (Figure 1C). However, slight variations in composition were observed, which

made it possible to establish hierarchical groupings and, in turn, distinguish species by

positioning them in a way that differs from the phylogenies (Figure 2). By this method,

only some species showed non-monophyletic groupings.

At the phylogenetic level, the tree inferred based on protein chains (Figure 2)

showed similar results in species placement when compared to mtDNA trees (check

Siriwut et al. 2015, 2016; Oeyen et al. 2014; Doménech et al. 2018). However, branch

positioning was more sensitive to amino acid variations, while node supports were

correspondingly weaker in protein-based phylogenetic reconstructions. Ultimately, it

was found that trees based on COI protein-coding sequences were somewhat less useful

for establishing phylogenies than those based on mtDNA (see Siriwut et al. 2015, 2016;

Oeyen et al. 2014; Doménech et al. 2018).

At the taxonomic level, the molecular results showed a significant number of

amino acid substitutions (>4) between *S. mutilans* and its closest relative, *S.*

subspinipes, reaffirming that these entities are not conspecific taxa, contrary to what has

been suggested in recent literature (Vahtera et al. 2013; Kang et al. 2017; Han et al.

2018) (Supplementary File 4). Conversely, the findings by Kang et al. (2017) which

reported no molecular differences between *S. subspinipes* and *S. hainanensis*, are not

supported here (residue differentiation >4; Supplementary File 4), as our results were

consistent with them being distinct species, aligning with their clear morphological

differences (Kronmüller 2012; Siriwut et al. 2016).

In parallel, experimental alignments (unpublished), which include relatively

short but partially well-conserved sequences, revealed that the morphological similarity

between *S. morsitans* Linnaeus, 1758 and *S. cf. amazonica* is strongly supported at the

molecular level, suggesting a likely synonymy (residue substitution = 1). Meanwhile,

species that appear morphologically similar, such as *S. japonica* and *S. negrocapitis*

Zhang & Wang, 1999, as well as *S. mojiangica*, produced inconclusive results (three to

five amino acid substitutions), showing the unsolved taxonomical and nomenclature

situation for these cases.

COI protein sequence variations between *S. paradoxa* and *S. spinosissima*, and

final considerations

The analysis of COI protein sequence variations was conducted using the DNA

sequences of *S. paradoxa* and *S. spinosissima* obtained in Doménech et al. (2018)

(Supplementary File 1). We found that the protein sequences derived from the holotype

and paratypes of *S. paradoxa* encoded exactly the same protein sequence

(Supplementary Files 2 and 3). Therefore, only one consensus sequence of *S. paradoxa*

was used for the following analyses. Figure 1A shows the pairwise alignment of the

partial COI protein sequences of *S. paradoxa* and *S. spinosissima*. As shown in the

alignment, the partial COI proteins displayed a high degree of identity (97.8% of

aligned amino acids in Supplementary Files 2 and 3). However, we identified five

amino acid residue differences between the two species: 19T>A, 105T>S, 108L>M,

115N>S, and 169E>S (Figures 1A–B, Table 1). Of these five substitutions, two—

105T>S and 115N>S—were located in regions of high variability observed across the

sequences (Supplementary File 2). The variations 19T>A, 108L>M, and 169E>S

involved relevant changes in the physicochemical properties of the amino acids (Table

1), which could suggest changes in the structure, function, and adaptation of the

resulting COI protein.

To ensure that the evolutionary process between these two species—of which

few sequences are available—is not random or an exception to the "five amino acid

rule," we calculated the substitution probability of these five residues using a total of 41

partial COI protein sequences of *Scolopendra* (Table 1). We found that substitutions

19T>A, 105T>S, and 115N>S had a substitution probability greater than one,

suggesting strong positive selection (adaptive or diversifying) at these three amino acid

positions (Table 1). In contrast, the substitutions at residues 108L>M and 169E>S

showed substitution probabilities below one, suggesting moderate and strong negative

(purifying) selection, respectively (Table 1). These data support the conclusion that the

observed amino acid divergence cannot be explained by genetic drift alone and

highlight the selective pressures involved in the diversification of these two species.

Among all the *Scolopendra* species represented in this study, *S. paradoxa*

consistently and exclusively exhibited a glutamic acid (E) at relative position 169,

making this amino acid a standalone diagnostic character for the species

(Supplementary File 3). As a group, *S. paradoxa-S. spinosissima* did not exhibit unique

group-specific amino acids that differentiated them from the rest of the species.

However, *S. spinosissima* shared characteristic amino acids with *S. calcarata* Porat,

1876, while *S. paradoxa* shared group-specific amino acids with *S. laeta*—none of

which were found together in any other species.

In the hierarchical classification of amino acid frequencies (Figure 1C), *S.*

paradoxa and *S. spinosissima* were grouped into non-sister sections, suggesting the

separation of these species by the use of this method. Similarly to this, the inferred

protein phylogenetic reconstruction showed *S. paradoxa* and *S. spinosissima* as close

but not strictly sister species. *Scolopendra paradoxa* displayed strong monophyletic

node support including its four sequences, whereas the position of *S. spinosissima*

remained weakly supported—precisely as reported in previous literature (Doménech et

al. 2018).

In line with earlier findings (Doménech et al. 2012), all these showed molecular

analyses concerning *S. spinosissima* and *S. paradoxa* once again conclude that these

taxonomic entities are objectively and definitively two distinct species, separated by

evolutionary selective pressures rather than by chance.

Finally, to facilitate species discrimination within *Scolopendra* using this alternative

molecular method based on partial proteins translated from COI fragments, we provide a

summary diagram of the results obtained, including noted exceptions and recommendations for

particular situations (Figure 3).

Discussion

In this study, the translated partial COI gene has been confirmed as a valuable

biomarker for the identification and differentiation of species within the genus

Scolopendra. The analysis of amino acids in the partial COI protein fragment showed

that sequences belonging to different species differ by five or more residues. However,

some exceptions to this "rule" were detected. To clarify most of these ambiguous

inter/intraspecific cases, additional analysis of exclusive and characteristic divergent

amino acids—their frequency, electrochemical properties, and especially the *r*

substitution probability test—proved to be useful tools for resolving these exceptional

scenarios.

Compared to traditional divergence studies based on mtDNA percentage

differences, the intra/interspecific discrimination of amino acid substitutions expressed

as natural numbers proved to be more easily interpretable, computationally less

demanding, and less affected by methodological and biological variation. Likewise, the

r substitution probability test showed a high degree of confidence in resolving complex

cases of taxon differentiation, providing measurable insights into the evolutionary

history of these species. On the other hand, the phylogenetic tree inferred from amino

acid chains, while yielding good results on its own, also proved to be no more accurate

than phylogenetic reconstructions based on mtDNA (Figure 2). In this case, the shorter

and more similar sequences, along with the statistical weight of any amino acid

substitution, made this type of analysis—though robust—more prone to fluctuations

than the more "tolerant" trees built from mtDNA data. Meanwhile, heat map and

hierarchical clustering, although useful in distinguishing species by amino acid

composition in most cases, were not as precise as divergence studies, but still served as

valuable supporting tools for the overall analysis. In contrast, the study of exclusive or

group-characteristic amino acids proved to be significant for the separation and

discrimination of *Scolopendra* species. However, this latter method should be

interpreted with caution in species with a low number of sequences analyzed and/or a

restricted geographic distribution.

Proportionally, it has been shown that interspecific mtDNA divergence

percentages do not correspond to those of amino acids (i.e., compare the divergences of

S. spinosissima and *S. paradoxa*, with respective differences of 19% for mtDNA and

2.2% for the coding protein; see Supplementary File 4 and Doménech et al. 2018). This

result aligns with Kimura's neutral theory (1983), which states that most DNA

mutations do not affect codon translation. Therefore, compared to most nucleotide

changes, an amino acid substitution is much more influential on the structure and function of the resulting protein, and consequently, on species decline or differentiation. Thus, due to its inherently lower susceptibility to the random effects of evolution, a species differentiation method based on proteins, such as the one proposed here, could represent an alternative to mtDNA-based methods, offering certain interpretative advantages and a high level of reliability.

On the other hand, from a general perspective, it is noteworthy that approximately one-third of the available COI sequences of Scolopendra exhibited various issues that hindered proper alignment. The intrinsic difficulty of COI sequencing—which includes artifacts such as nuclear mitochondrial DNA segments (NUMTs) (Rogers & Griffiths-Jones 2012)—combined with human-related factors like species misidentification, statistical errors, and biological elements such as the accumulation of neutral mutations, excessive substitution saturation, or the phenomenon of heterotachy (Kimura 1968, 1983; Zuckerkandl & Pauling 1962; Xia 2009), may ultimately disrupt molecular analysis. All of these elements should be evaluated to improve the quality of future analyses aimed at species differentiation. Furthermore, aside from the sequences reviewed directly in this study, the quality of the remaining protein sequences filtered using different methods can only be approximated here, and cannot be guaranteed with certainty. For this reason, some of the "irregular" cases tentatively treated as exceptions in this study (see Results section) may, in fact, be explained by the aforementioned errors. A higher standard of rigor and standardized filtering applied prior to sequence publication could be key to improving the overall quality of COI databases.

In the species-level analysis, our study found that some sequences of *S. cataracta* exhibited the highest molecular variability within the genus, consistent with

the findings of Siriwut et al. (2015), making it the only intraspecific exception to the

"five amino acid rule." However, despite passing the selection filters, this exception

could potentially be linked to an error, given the geographic-molecular discrepancies

observed among the available sequences of this species, thus requiring further study. On

the other hand, the morphologically very similar S. multidens and S. dawydoffi (Siriwut

et al. 2016) also could not be separated using this method, which might suggest that the

minor morphological differences actually represent intrinsic variability within a single

taxon. The possible synonymy of these two species should be confirmed through a study

of the type specimens. Likewise, the sequences of *S. subspinipes* and *S. pinguis* should

be re-examined due to the excessively high intraspecific distances observed in this

study. Additionally, the validity of the few sequences available for *S. cingulata*, *S.*

cataracta, and *S. lufengia* should also be assessed, as their placement falls ambiguously

within either the intra- or interspecific range. If no errors are confirmed in these few

cases, such deviations should be interpreted solely as a reflection of the progressive

evolutionary effect in certain specimens—representing a transitional and certainly

exceptional stage relative to the divergence ranges clearly defined by the five amino

acid rule.

Regarding *S. spinosissima* and *S. paradoxa*, the comparison of the translated

COI ortholog gene revealed five clear and consistent differences in their amino acid

composition and in certain electrochemical properties (Figure 1A and B, Table 1,

Supplementary Files 2 and 3). Moreover, the *r* substitution probability analysis rejected

the null hypothesis (genetic drift) for all amino acid substitutions and confirmed that

none of them could be explained by any random process. All these genetic results

provide evidence of a speciation process driven by selection between *S. spinosissima*

and *S. paradoxa*, findings that further support the morphological differences and
taxonomic separation of both species (Doménech et al. 2018, Doménech 2024).

The genetic results presented here represent a step forward in the distinction of
Scolopendra species. Although consistent, the conclusions of this analysis were based
on representative sequences from approximately 20–25% of the described species
within the genus, and are therefore subject to further refinement. Revisiting the
questionable and excluded sequences, along with incorporating more sequences from
both these and other unexamined taxa, could enhance the accuracy of this new and
simple methodology aimed at improving the molecular differentiation of *Scolopendra*
species.

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In accordance with the Philippine authorities, the undersigned authors of this article
wish to explicitly emphasize that access to any type of biological resources in this
nation always requires appropriate research permits from the DENR.

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Tables

Table 1. Position and chemical proprieties of divergent amino acid residues between *S. paradoxa* and *S. spinosissima* COI protein sequence. Colum *r* is probability of substitution. The numbering of the amino acids refers to the positions indicated in the supplementary file 1. *=essential amino acids.

Residue position	Location on highly variable regions	S. paradoxa		S. spinosissima		Probability of substitution (*r* value)
		Amino acids	Electrochemical Characteristics	Amino acids	Electrochemical Characteristics	
19	No	T*	Polar, non-charged	A	Non-polar	1.83
105	Si	T*	Polar, non-charged	S	Polar, non-charged	1.69
108	No	L*	Non-polar	M*	Polar, non-charged	0.47
115	Si	N	Polar, non-charged	S	Polar, non-charged	1.79
169	No	E	Negatively charged; Polar	S	Polar, non-charged	0.11

Figures

Figure 1. Molecular analysis of the partial COI protein sequence. **A** Pairwise alignment of the partial COI protein sequence of *S. paradoxa* / *S. spinosissima*. Identical amino acid residues are shaded in black, and residues with more than 70% similarity with the rest of the studied sequences are shaded in grey. Asterisks indicate amino acid substitutions between these two species. Blue lines mark sites where residues show high variability within the genus *Scolopendra*. **B** Logos representing 11 polymorphic residues found in the 41 partial COI protein sequences (blue asterisk). Polymorphic residues from the partial COI protein sequences of *S. paradoxa* / *S. spinosissima* are included (red asterisk). Red asterisks underlined in blue indicate polymorphic residues shared between the 22 *Scolopendra* spp. and the *S. paradoxa* / *S. spinosissima* COI protein sequences. **C** Heatmap and hierarchical clustering of amino acid frequency composition from the 45 partial COI protein sequences of *Scolopendra*. Hierarchical clustering was calculated using Pearson's average correlation of amino acid frequency composition for each species. Colored branches highlight the close amino acid frequency relationships of *S. paradoxa* (orange) and *S. spinosissima* (green).

Figure 2. Maximum Likelihood phylogenetic reconstruction based on the protein-coding region of the partial COI gene translated into proteins, including 45 selected *Scolopendra* taxa along with six additional members of the tribe Scolopendrini and two species from the subfamily Otostigminae. Bootstrap values are shown for each node. Colored branches in the tree represent the genetic affinities of *S. paradoxa* (blue) and *S. spinosissima* (red) in relation to their congeners (thin black branches). Bold branches and thick black branches indicate the positions of other members of the *Scolopendrini* tribe and the Otostigminae species (outgroups), respectively.

694   **Figura 3.** Summary scheme of the results, exceptions, and recommendations for
695   determining intra- or interspecific differences between two or more translated protein
696   sequences from the partial COI gene in the genus *Scolopendra*.

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Supplementary files

Supplementary file 1. Summary of the 45 COI sequences from *Scolopendra* species, along with eight from other Scolopendromorpha, obtained from the Barcode of Life Data System (BOLD) and used in the molecular analyses. Each sample includes the institutional abbreviation of the voucher, type status, voucher code, locality, and GenBank accession number for the selected partial COI gene sequence.

Sample name (species and specimens' voucher)	Locality	COI GenBank accession
Scolopendra spinosissima **Kraepelin, 1903**		
S.spinosissima.CEUA016.Mr0009	Philippines	KY888682.1
Scolopendra paradoxa **Doménech, 2018**		
S.paradoxa.CEUA017.Mr0000 (Holotype)	Philippines	MH087425.1
S.paradoxa.CEUA016.Mr0006 (Paratype 9)	Philippines	KY888684.1
S.paradoxa.CEUA016.Mr0007 (Paratype 10)	Philippines	KY888685.2
S.paradoxa.CEUA016.Mr0008 (Paratype 11)	Philippines	KY888686.2
Scolopendra afer **(Meinert,1886)**		
S.afer.IZ130688	Tanzania	KF676520.1
Scolopendra alternans **Leach, 1816**		
S.alternans.IZ130689	United States of America	KF676521.1
Scolopendra calcarata **Porat, 1876**		
S.calcarata.SYSU:Chilo112	Vietnam	AB614399.1
S.calcarata.CUMZ00418	Thailand	KU512630.1
Scolopendra cataracta **Siriwut, Edgecombe & Panha , 2016**		
S.cataracta.CUMZ00316 (Holotype)	Laos	KR705672.1
S.cataracta.CUMZ00317 (Paratype)	Laos	KR705633.1
Scolopendra cingulata **Latreille, 1829**		
S.cingulata.ZFMKMYR01593	France	KJ812087.1

S.cingulata.ZFMKMYR01561	Hungary	KJ812079.1
S.cingulata.ZFMKSco01	Austria	KJ812067.1
Scolopendra dawydoffi **Kronmüller, 2012**		
S.dawydoffi.CUMZ00294.1	Thailand	KR705635.1
S.dawydoffi.CUMZ00294.2	Thailand	KR705634.1
Scolopendra dehaani **Brandt, 1840**		
S.dehaani.CUMZ00333	Laos	KR705673.1
S.dehaani.CUMZ00325	Thailand	KR705656.1
S.dehaani.CUMZ00331	Cambodia	KR705686.1
S.dehaani.CUMZ00252	Thailand	KR705683.1
Scolopendra hardwickei **Newport, 1844**		
S.hardwickei.FIMN-Shardwickei	N/a (¿India?)	KF676523.1
Scolopendra japonica **L. Koch, 1878**		
S.japonica.CUMZ00319	Japan	KR705679.1
S.japonica.SYSU:Chilo206	Japan	AB672741.1
Scolopendra laeta **Haase, 1887**		
S.laeta.MCZ.AM.IZ130696	Australia	KF676541.1
Scolopendra morsitans **Linnaeus, 1758**		
S.morsitans.CUMZ00344	Thailand	KR705664.1
S.morsitans.CUMZ00342	Thailand	KR705666.1
S.morsitans.SYSU:Chilo208	Taiwan	AB672739.1
S.morsitans.SYSU:Chilo005	Taiwan	AB614397.1
Scolopendra multidens **Newport, 1844**		
S.multidens.SYSU:Chilo118	Vietnam	AB672738.1

Scolopendra mutilans **L. Koch, 1878**		
S.mutilans.NIAS18224	Korea	MG991739.1
S.mutilans.NIAS8227	Korea	MG991742.1
Scolopendra oraniensis **Lucas, 1846**		
S.oraniensis.ZFMKMYR00568	Morocco	KJ812088.1
Scolopendra pinguis **Pocock, 1891**		
S.pinguis.CUMZ00304	Laos	KR705642.1
S.pinguis.CUMZ00309	Laos	KR705646.1
S.pinguis.CUMZ00314	Thailand	KR705648.1
Scolopendra polymorpha **Wood, 1861**		
S.polymorpha.IZ130561	United States of America	KF676525.1
Scolopendra subcrustalis **Kronmüller, 2009**		
S.subcrustalis.CEUA016.MR0010	Philippines	KY888683.2
Scolopendra subspinipes **Leach, 1816**		
S.subspinipes.IZ131452	Martinique	HQ402554.1
S.subspinipes.CUMZ00315	Singapur	KR705636.1
Scolopendra viridis **Say, 1821**		
S.viridis.IZ130727	United States of America	DQ201431.1
Scolopendromorpha sequences used for the inferred phylogenetic tree		
Asanada brevicornis **Meinert, 1886**		
Asanada.brevicornis.CES-08951	India	JN004014.1
Akymnopellis chilensis **(Gervais, 1847)**		
Akymnopellis.chilensis.IZ-130824	Chile	KF676538.1
Cormocephalus (_Cormocephalus_) _westwoodi_ **(Newport, 1844)**		

Cormocephalus.westwoodi.CES-07258	Australia	JN004004.1
Cormocephalus (*Campylostigmus*) *orientalis* **Ribaut, 1923**		
Campylostigmus.orientalis.MCZ.IZ-130623	New Caledonia	HQ402542.1
Arthrorhabdus formosus **Pocock, 1891**		
Arthrorhabdus.formosus.MCZ.IZ-130617	South Africa	HQ402539.1
Hemiscolopendra marginata **(Say, 1821)**		
Hemiscolopendra.marginata.MCZ.IZ-130659	United States of America	HQ402548.1
Ethmostigmus rubripes **Brandt, 1840**		
Ethmostigmus.rubripes.MCZ.IZ-130653	Australia	KF676542.1
Rhysida immarginata **Porat, 1876**		
Rhysida.immarginata.SYSU:Chilo-050	Taiwan: Kinmen, Lieyu	AB614648.1

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                                           70        80        90        100       110       120
                                    ....|....|....|....|....|....|....|....|..**..**|.*..*....|
S.paradoxa.CEUA017.MR0000           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSALVENGAGTG
S.paradoxa.CEUA016.MR0006           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSALVENGAGTG
S.paradoxa.CEUA016.MR0007           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSALVENGAGTG
S.paradoxa.CEUA016.MR0008           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSALVENGAGTG
S.spinosissima.CEUA016.MR0009       IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLSLLMSSALVENGAGTG
S.cataracta.CUMZ00316               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLTSALVESGAGTG
S.cataracta.CUMZ00317               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLSSALVESGAGTG
S.dehaani.CUMZ00333                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLASALVENGAGTG
S.dehaani.CUMZ00325                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLASALVENGAGTG
S.dehaani.CUMZ00331                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLASALVENGAGTG
S.dehaani.CUMZ00252                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLASALVENGAGTG
S.subspinipes.CUMZ00315             IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSITLLLASTLMENGAGTG
S.subspinipes.IZ131452              IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLTSALVENGAGTG
S.mutilans.NIAS18224                IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLTSALIENGAGTG
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S.japonica.CUMZ00319                IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLLSSAMVENGAGTG
S.japonica.SYSU:Chilo206            IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLSSAMVENGAGTG
S.subcrustalis.CEUA016.MR0010       IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLASALVENGAGTG
S.laeta.MCZ.AM.IZ130696             IFFMVMPIMIGGFGNWLIPLMLGAPDMAFPRMNNLSFWLLPPSLALLLASAMVESGAGTG
S.morsitans.CUMZ00344               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSAMVENGAGTG
S.morsitans.CUMZ00342               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSAMVENGAGTG
S.morsitans.SYSU:Chilo208           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSAMVENGAGTG
S.morsitans.SYSU:Chilo005           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSAMVENGAGTG
S.pinguis.CUMZ00304                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLASAMIESGAGTG
S.pinguis.CUMZ00309                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLMTSAMMENGAGTG
S.pinguis.CUMZ00314                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLASAVIESGAGTG
S.calcarata.SYSU:Chilo112           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLFPPSLMLLLSSAMDESGASTG
S.calcarata.CUMZ00418               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLSLLLASAALESGAGTG
S.calcarata.NHMUK010305528          IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLSLLLASAALESGAGTG
S.multidens.SYSU:Chilo118           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLLSSALVENGAGTG
S.dawydoffi.CUMZ00294.1             IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLLSSALVENGAGTG
S.dawydoffi.CUMZ00294.2             IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLLSSALVENGAGTG
S.cingulata.ZFMKMYR01593            IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLSSALVENGAGTG
S.cingulata.ZFMKMYR01561            IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLSSALVENGAGTG
S.cingulata.ZFMKSco01               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLSSALVENGAGTG
S.oraniensis.ZFMKMYR00568           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLMSSALVESGAGTG
S.alternans.IZ130689                IFFMVMPIMVGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLLSSGLVESGAGTG
S.polymorpha.IZ130561               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLLSSALVESGAGTG
S.viridis.IZ130727                  IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLILLLSSAMVENGAGTG
S.hardwickei.FIMNShardwickei        IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLMASAMVESGAGTG
S.afer.IZ130688                     IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMMLMGSSIVESGAGTG
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S.paradoxa.CEUA017.MR0000           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSALVENGAGTG
S.paradoxa.CEUA016.MR0006           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSALVENGAGTG
S.paradoxa.CEUA016.MR0007           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSALVENGAGTG
S.paradoxa.CEUA016.MR0008           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSALVENGAGTG
S.spinosissima.CEUA016.MR0009       IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLSLLMSSALVESGAGTG
S.cataracta.CUMZ00316               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLTSALVESGAGTG
S.cataracta.CUMZ00317               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLSSALVESGAGTG
S.dehaani.CUMZ00333                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLIASALVENGAGTG
S.dehaani.CUMZ00325                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLIASALVENGAGTG
S.dehaani.CUMZ00331                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLIASALVENGAGTG
S.dehaani.CUMZ00252                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLIASALVENGAGTG
S.subspinipes.CUMZ00315             IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSITLLIASTLMENGAGTG
S.subspinipes.IZ131452              IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLITSALVENGAGTG
S.mutilans.NIAS18224                IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLTSALIENGAGTG
S.mutilans.NIAS8227                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLTSALIENGAGTG
S.japonica.CUMZ00319                IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLLSSAMVENGAGTG
S.japonica.SYSU:Chilo206            IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLSSAMVENGAGTG
S.subcrustalis.CEUA016.MR0010       IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLIASALVENGAGTG
S.laeta.MCZ.AM.IZ130696             IFFMVMPIMIGGFGNWLIPLMLGAPDMAFPRMNNLSFWLLPPSLALLIASAMVESGAGTG
S.morsitans.CUMZ00344               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSAMVENGAGTG
S.morsitans.CUMZ00342               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSAMVENGAGTG
S.morsitans.SYSU:Chilo208           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLLSSAMVENGAGTG
S.morsitans.SYSU:Chilo005           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLIASAMVENGAGTG
S.pinguis.CUMZ00304                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLIASAMVENGAGTG
S.pinguis.CUMZ00309                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLMTSAMMENGAGTG
S.pinguis.CUMZ00314                 IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLIASAVLESGAGTG
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S.calcarata.CUMZ00418               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLSLLIASAALESGAGTG
S.calcarata.NHMUK010305528          IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLSLLIASAALESGAGTG
S.multidens.SYSU:Chilo118           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLISSALVENGAGTG
S.dawydoffi.CUMZ00294.1             IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLSSALVENGAGTG
S.dawydoffi.CUMZ00294.2             IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLSSALVENGAGTG
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S.cingulata.ZFMKSco01               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLLSSALVENGAGTG
S.oraniensis.ZFMKMYR00568           IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLMSSALVESGAGTG
S.alternans.IZ130689                IFFMVMPIMVGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLISSGLVESGAGTG
S.polymorpha.IZ130561               IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLMLLISSALVESGAGTG
S.viridis.IZ130727                  IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLILLLSSAMVESGAGTG
S.hardwickei.FIMNShardwickei        IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSLTLLMASAMVESGAGTG
S.afer.IZ130688                     IFFMVMPIMIGGFGNWLVPLMLGAPDMAFPRMNNLSFWLLPPSIMLLMGSSIVESGAGTG
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S.paradoxa.CEUA016.MR0006	WAVKITAILLLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGH-----
S.paradoxa.CEUA016.MR0007	WAVKITAILLLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFG------
S.paradoxa.CEUA016.MR0008	WAVKITAILLLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFG------
S.spinosissima.CEUA016.MR0009	WAVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPE---
S.cataracta.CUMZ00316	WAVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.cataracta.CUMZ00317	WAVKITAILLLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.dehaani.CUMZ00333	WAVMITAILLLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.dehaani.CUMZ00325	WAVMITAILLLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.dehaani.CUMZ00331	WAVMITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.dehaani.CUMZ00252	WAVMITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.subspinipes.CUMZ00315	WAVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.subspinipes.IZ131452	WAVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.mutilans.NIAS18224	WSVLITTILLLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLF----------
S.mutilans.NIAS8227	WSVKITTILLLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLF----------
S.japonica.CUMZ00319	WSVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.japonica.SYSU:Chilo206	WSVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLF----------
S.subcrustalis.CEUA016.MR0010	WAVGITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPTGGGDPILYQHLFWFFGHPE---
S.laeta.MCZ.AM.IZ130696	WAVGITAILLLFSLPVLAGAITMLLTDRNFNTSFFDPTGGGDPILYQHLFWFFGHPEVYI
S.morsitans.CUMZ00344	WAVGITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.morsitans.CUMZ00342	WAVGITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.morsitans.SYSU:Chilo208	WAVGITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLF----------
S.morsitans.SYSU:Chilo005	WAVGITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLF----------
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S.pinguis.CUMZ00309	WAVLITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.pinguis.CUMZ00314	WAVLITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
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S.calcarata.CUMZ00418	WAVLITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.calcarata.NHMUK010305528	WAVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.multidens.SYSU:Chilo118	WAVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLF----------
S.dawydoffi.CUMZ00294.1	WAVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.dawydoffi.CUMZ00294.2	WAVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.cingulata.ZFMKMYR01593	GSVKITAILLLLLSLPVLAGXITMLLKDRNFNTSFFDPAGGGDPILYBHLYWX-------
S.cingulata.ZFMKMYR01561	WSVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGD-----------------
S.cingulata.ZFMKSco01	WSVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGD-----------------
S.oraniensis.ZFMKMYR00568	WAVKITTILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWF-------
S.alternans.IZ130689	WAVKITTILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.polymorpha.IZ130561	WAVKITTILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.viridis.IZ130727	WAVKITTILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYQHLFWFFGHPEVYI
S.hardwickei.FIMNShardwickei	WAVKITAILLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDP-----------------
S.afer.IZ130688	WAVGITTVLLLLSLPVLAGAITMLLTDRNFNTSFFDPAGGGDPILYXHLFWFFGHPEVYI

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                                          250        260        270        280
                                 ....|....|....|....|....|....|....|....|....|..
S.paradoxa.CEUA017.MR0000        ------------------------------------------------
S.paradoxa.CEUA016.MR0006        ------------------------------------------------
S.paradoxa.CEUA016.MR0007        ------------------------------------------------
S.paradoxa.CEUA016.MR0008        ------------------------------------------------
S.spinosissima.CEUA016.MR0009    ------------------------------------------------
S.cataracta.CUMZ00316            LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGNLGFVV
S.cataracta.CUMZ00317            LILPGFGMISHIISQQSGKKESFGTLGMMYAMGAIGILGFVV
S.dehaani.CUMZ00333              LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.dehaani.CUMZ00325              LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.dehaani.CUMZ00331              LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.dehaani.CUMZ00252              LILPGFGMISHIISQQSGKKETFGTLGMIYAMAAIAILGFVV
S.subspinipes.CUMZ00315          LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.subspinipes.IZ131452           LILPGFGMISHIISQQSGKKEPF------------------
S.mutilans.NIAS18224             ------------------------------------------------
S.mutilans.NIAS8227              ------------------------------------------------
S.japonica.CUMZ00319             LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.japonica.SYSU:Chilo206         ------------------------------------------------
S.subcrustalis.CEUA016.MR0010    ------------------------------------------------
S.laeta.MCZ.AM.IZ130696          LILPGFGMISHIISHYSGKKEPFGTLGMIYAMAAIGILGFVV
S.morsitans.CUMZ00344            LILPGFGMISHIISHYSGKKEPFGTLGMIYAMAAIGILGFVV
S.morsitans.CUMZ00342            LILPGFGMISHIISHYSGKKEPFGTLGMIYAMAAIGILGFVV
S.morsitans.SYSU:Chilo208        ------------------------------------------------
S.morsitans.SYSU:Chilo005        ------------------------------------------------
S.pinguis.CUMZ00304              LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.pinguis.CUMZ00309              LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.pinguis.CUMZ00314              LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.calcarata.SYSU:Chilo112        ------------------------------------------------
S.calcarata.CUMZ00418            LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.calcarata.NHMUK010305528       LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.multidens.SYSU:Chilo118        ------------------------------------------------
S.dawydoffi.CUMZ00294.1          LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.dawydoffi.CUMZ00294.2          LILPGFGMISHIISQQSGKKEPFGTLGMIYAMAAIGILGFVV
S.cingulata.ZFMKMYR01593         ------------------------------------------------
S.cingulata.ZFMKMYR01561         ------------------------------------------------
S.cingulata.ZFMKSco01            ------------------------------------------------
S.oraniensis.ZFMKMYR00568        ------------------------------------------------
S.alternans.IZ130689             LILPGFGMVSHIISQQSGKKEPFGALGMIYAMAAIGILGFVV
S.polymorpha.IZ130561            LILPGFGMVSHIISQQSGKKEPFGSLGMIYAMAAIGILGFVV
S.viridis.IZ130727               LILPGFGMVSHIISHQCGKKEPFGALGMIYAMASIGILGFVV
S.hardwickei.FIMNShardwickei     ------------------------------------------------
S.afer.IZ130688                  LILPGFGMVSHIISQESGSSEP------------------
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719
720

721   **Supplementary file 3.** Alignment of 45 amino acid sequences from the partial COI
722   gene belonging to 22 different species of the genus *Scolopendra*. Asterisks indicate
723   residues common to all studied species. The color coding helps identify amino acids that
724   are   unique   to   a   species   and   characteristic   of   a   group   of   species.

#	Species/Abbrv
1.	S.paradoxa.CEUA017.MR0000
2.	S.paradoxa.CEUA016-Mr0006
3.	S.paradoxa.CEUA016-Mr0007
4.	S.paradoxa.CEUA016-Mr0008
5.	S.spinosissima.CEUA016.MR0009
6.	S.cataracta.CUMZ00316
7.	S.cataracta.CUMZ00317
8.	S.dehaani.CUMZ00333
9.	S.dehaani.CUMZ00325
10.	S.dehaani.CUMZ00331
11.	S.dehaani.CUMZ00252
12.	S.subspinipes.CUMZ00315
13.	S.subspinipes.IZ131452
14.	S.mutilans.NIAS18224
15.	S.mutilans.NIAS8227
16.	S.japonica.CUMZ00319
17.	S.japonica.SYSU:Chilo206
18.	S.subcrustalis.CEUA016MR0010
19.	S.laeta.MCZ.AM.IZ130696
20.	S.morsitans.CUMZ00344
21.	S.morsitans.CUMZ00342
22.	S.morsitans.SYSU:Chilo208
23.	S.morsitans.SYSU:Chilo005
24.	S.pinguis.CUMZ00304
25.	S.pinguis.CUMZ00309
26.	S.pinguis.CUMZ00314
27.	S.calcarata.SYSU:Chilo112
28.	S.calcarata.CUMZ00418
29.	S.calcarata.NHMUK010305528
30.	S.multidens.SYSU:Chilo118
31.	S.dawydoffi.CUMZ00294.1
32.	S.dawydoffi.CUMZ00294.2
33.	S.cingulata.ZFMKMYR01593
34.	S.cingulata.ZFMKMYR01561
35.	S.cingulata.ZFMKSco01
36.	S.oraniensis.ZFMKMYR00568
37.	S.alternans.IZ130689
38.	S.polymorpha.IZ130561
39.	S.viridis.IZ130727
40.	S.hardwickei.FMNShardwickei
41.	S.afer.IZ130688

727
728

Species/Abbrv

1. S.paradoxa.CEUA017.MR0000
2. S.paradoxa.CEUA016-Mr0006
3. S.paradoxa.CEUA016-Mr0007
4. S.paradoxa.CEUA016-Mr0008
5. S.spinosissima.CEUA016.MR0009
6. S.cataracta.CUMZ00316
7. S.cataracta.CUMZ00317
8. S.dehaani.CUMZ00333
9. S.dehaani.CUMZ00325
10. S.dehaani.CUMZ00331
11. S.dehaani.CUMZ00252
12. S.subspinipes.CUMZ00315
13. S.subspinipes.IZ131452
14. S.mutilans.NIAS18224
15. S.mutilans.NIAS8227
16. S.japonica.CUMZ00319
17. S.japonica.SYSU:Chilo206
18. S.subcrustalis.CEUA016MR0010
19. S.laeta.MCZ.AM.IZ130696
20. S.morsitans.CUMZ00344
21. S.morsitans.CUMZ00342
22. S.morsitans.SYSU:Chilo208
23. S.morsitans.SYSU:Chilo005
24. S.pinguis.CUMZ00304
25. S.pinguis.CUMZ00309
26. S.pinguis.CUMZ00314
27. S.calcarata.SYSU:Chilo112
28. S.calcarata.CUMZ00418
29. S.calcarata.NHMUK010305528
30. S.multidens.SYSU:Chilo118
31. S.dawydoffi.CUMZ00294.1
32. S.dawydoffi.CUMZ00294.2
33. S.cingulata.ZFMKMYR01593
34. S.cingulata.ZFMKMYR01561
35. S.cingulata.ZFMKSco01
36. S.oraniensis.ZFMKMYR00568
37. S.alternans.IZ130689
38. S.polymorpha.IZ130561
39. S.viridis.IZ130727
40. S.hardwickei.FIMNShardwickei
41. S.afer.IZ130688

729   **Supplementary file 4.** Divergence table obtained from 45 amino acid sequences of the

730   partial COI gene belonging to 22 different species of the genus *Scolopendra*. Note that

731   out of 820 alignments, only 14 exceptions to the five amino acids rule were found, eight

732   of which correspond to sequences from the same species.

	S.paradoxa.CEUA017.MR0000	S.paradoxa.CEUA016-Mr0006	S.paradoxa.CEUA016-Mr0007	S.paradoxa.CEUA016-Mr0008	S.spinosissima.CEUA016.MR0009	S.hainanensis.CMMI20190702020	S.hainanensis.CMMI20180922009	Scolopendra.sp.china.CMMI20190702007	Scolopendra.sp.china.CMMI20190418008
S.paradoxa.CEUA017.MR0000									
S.paradoxa.CEUA016-Mr0006	0,000								
S.paradoxa.CEUA016-Mr0007	0,000	0,000							
S.paradoxa.CEUA016-Mr0008	0,000	0,000	0,000						
S.spinosissima.CEUA016.MR0009	0,023	0,023	0,023	0,023					
S.hainanensis.CMMI20190702020	0,062	0,062	0,062	0,062	0,052				
S.hainanensis.CMMI20180922009	0,062	0,062	0,062	0,062	0,052	0,000			
Scolopendra.sp.china.CMMI20190702007	0,067	0,067	0,067	0,067	0,062	0,014	0,014		
Scolopendra.sp.china.CMMI20190418008	0,067	0,067	0,067	0,067	0,062	0,014	0,014	0,000	
S.cataracta.CUMZ00316	0,057	0,057	0,057	0,057	0,048	0,024	0,024	0,038	0,038
S.cataracta.CUMZ00317	0,048	0,048	0,048	0,048	0,038	0,019	0,019	0,028	0,028
S.dehaani.CUMZ00333	0,053	0,053	0,053	0,053	0,053	0,028	0,028	0,043	0,043
S.dehaani.CUMZ00325	0,053	0,053	0,053	0,053	0,053	0,028	0,028	0,043	0,043
S.dehaani.CUMZ00331	0,053	0,053	0,053	0,053	0,053	0,028	0,028	0,043	0,043
S.dehaani.CMMI20180922002	0,052	0,052	0,052	0,052	0,052	0,033	0,033	0,047	0,047
S.subspinipes.CUMZ00315	0,067	0,067	0,067	0,067	0,072	0,053	0,053	0,057	0,057
S.subspinipes.IZ131452	0,049	0,049	0,049	0,049	0,049	0,029	0,029	0,044	0,044
S.mutilans.NIAS18224	0,093	0,093	0,093	0,093	0,093	0,087	0,087	0,093	0,093
S.mutilans.NIAS8227	0,082	0,082	0,082	0,082	0,082	0,077	0,077	0,082	0,082
S.mutilans.CMMI20190331001	0,082	0,082	0,082	0,082	0,082	0,082	0,082	0,087	0,087
S.alcyona.MST02	0,048	0,048	0,048	0,048	0,048	0,053	0,053	0,062	0,062
S.alcyona.MST03	0,057	0,057	0,057	0,057	0,057	0,048	0,048	0,057	0,057
S.japonica.CUMZ00319	0,057	0,057	0,057	0,057	0,057	0,043	0,043	0,053	0,053
S.japonica.SYSU:Chilo206	0,058	0,058	0,058	0,058	0,058	0,043	0,043	0,053	0,053
S.lufengia.CMMI20181030005	0,033	0,033	0,033	0,033	0,033	0,047	0,047	0,062	0,062
S.lufengia.CMMI20181228010	0,047	0,047	0,047	0,047	0,047	0,057	0,057	0,072	0,072
S.subcrustalis.CEUA016MR0010	0,056	0,056	0,056	0,056	0,066	0,067	0,067	0,072	0,072
S.laeta.MCZ.AM.IZ130696	0,082	0,082	0,082	0,082	0,082	0,077	0,077	0,087	0,087
S.morsitans.CUMZ00344	0,053	0,053	0,053	0,053	0,062	0,072	0,072	0,077	0,077
S.morsitans.CUMZ00342	0,053	0,053	0,053	0,053	0,062	0,072	0,072	0,077	0,077
S.morsitans.SYSU:Chilo208	0,058	0,058	0,058	0,058	0,068	0,078	0,078	0,083	0,083
S.morsitans.SYSU:Chilo005	0,048	0,048	0,048	0,048	0,058	0,068	0,068	0,073	0,073
S.pinguis.CUMZ00304	0,072	0,072	0,072	0,072	0,067	0,077	0,077	0,077	0,077
S.pinguis.CUMZ00309	0,067	0,067	0,067	0,067	0,062	0,077	0,077	0,077	0,077
S.pinguis.CUMZ00314	0,068	0,068	0,068	0,068	0,063	0,073	0,073	0,078	0,078
S.calcarata.SYSU:Chilo112	0,119	0,119	0,119	0,119	0,108	0,124	0,124	0,135	0,135
S.calcarata.CUMZ00418	0,091	0,091	0,091	0,091	0,076	0,082	0,082	0,087	0,087
S.calcarata.NHMUK010305528	0,091	0,091	0,091	0,091	0,076	0,082	0,082	0,087	0,087
S.multidens.SYSU:Chilo118	0,053	0,053	0,053	0,053	0,053	0,053	0,053	0,068	0,068
S.dawydoffi.CUMZ00294.1	0,043	0,043	0,043	0,043	0,043	0,043	0,043	0,057	0,057
S.dawydoffi.CUMZ00294.2	0,043	0,043	0,043	0,043	0,043	0,043	0,043	0,057	0,057
S.cingulata.ZFMKMYR01593	0,071	0,071	0,071	0,071	0,071	0,066	0,066	0,081	0,081
S.cingulata.ZFMKMYR01561	0,045	0,045	0,045	0,045	0,045	0,040	0,040	0,055	0,055
S.cingulata.ZFMKSco01	0,044	0,044	0,044	0,044	0,044	0,029	0,029	0,044	0,044
S.oraniensis.ZFMKMYR00568	0,059	0,059	0,059	0,059	0,039	0,059	0,059	0,074	0,074
S.alternans.IZ130689	0,057	0,057	0,057	0,057	0,048	0,072	0,072	0,087	0,087
S.polymorpha.IZ130561	0,053	0,053	0,053	0,053	0,043	0,053	0,053	0,067	0,067
S.viridis.IZ130727	0,088	0,088	0,088	0,088	0,088	0,083	0,083	0,088	0,088
S.hardwickei.FIMNShardwickei	0,050	0,050	0,050	0,050	0,035	0,040	0,040	0,045	0,045
S.afer.IZ130688	0,118	0,118	0,118	0,118	0,103	0,113	0,113	0,129	0,129

S.cataracta.CUMZ00316
S.cataracta.CUMZ00317
S.dehaani.CUMZ00333
S.dehaani.CUMZ00325
S.dehaani.CUMZ00331
S.dehaani.CMMI20180922002
S.subspinipes.CUMZ00315
S.subspinipes.IZ131452
S.mutilans.NIAS18224
S.mutilans.NIAS8227
S.mutilans.CMMI20190331001
S.alcyona.MST02
S.alcyona.MST03
S.japonica.CUMZ00319
S.japonica.SYSU:Chilo206

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0,033 0,028 0,000 0,000
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0,024 0,024 0,029 0,029 0,029 0,029 0,044
0,082 0,077 0,077 0,077 0,077 0,077 0,098 0,065
0,072 0,067 0,067 0,067 0,067 0,067 0,087 0,054 0,009
0,072 0,067 0,067 0,067 0,067 0,067 0,087 0,054 0,009 0,000
0,043 0,043 0,053 0,053 0,053 0,053 0,072 0,044 0,087 0,077 0,077
0,043 0,043 0,053 0,053 0,053 0,053 0,072 0,044 0,087 0,077 0,077 0,009
0,048 0,028 0,048 0,048 0,048 0,048 0,072 0,034 0,087 0,077 0,077 0,048 0,038
0,048 0,028 0,048 0,048 0,048 0,048 0,073 0,034 0,088 0,078 0,078 0,048 0,038 0,000
0,033 0,033 0,038 0,038 0,038 0,038 0,062 0,034 0,082 0,072 0,072 0,028 0,028 0,038 0,038
0,043 0,043 0,043 0,043 0,043 0,043 0,067 0,044 0,082 0,082 0,082 0,038 0,038 0,048 0,048
0,057 0,062 0,057 0,057 0,057 0,057 0,057 0,049 0,103 0,093 0,092 0,062 0,062 0,072 0,073
0,077 0,082 0,082 0,082 0,082 0,082 0,082 0,080 0,124 0,113 0,113 0,093 0,082 0,093 0,093
0,067 0,067 0,072 0,072 0,072 0,072 0,082 0,070 0,118 0,108 0,108 0,053 0,053 0,062 0,063
0,067 0,067 0,072 0,072 0,072 0,072 0,082 0,070 0,118 0,108 0,108 0,053 0,053 0,062 0,063
0,073 0,073 0,078 0,078 0,078 0,078 0,088 0,075 0,124 0,114 0,114 0,058 0,058 0,068 0,068
0,063 0,063 0,068 0,068 0,068 0,068 0,078 0,065 0,114 0,103 0,103 0,048 0,048 0,058 0,058
0,087 0,077 0,082 0,082 0,082 0,082 0,087 0,080 0,108 0,098 0,098 0,103 0,093 0,087 0,088
0,077 0,072 0,072 0,072 0,072 0,072 0,072 0,065 0,098 0,087 0,087 0,098 0,087 0,072 0,073
0,083 0,078 0,078 0,078 0,078 0,078 0,083 0,065 0,103 0,093 0,093 0,098 0,093 0,093 0,088
0,130 0,114 0,130 0,130 0,130 0,130 0,156 0,134 0,146 0,135 0,135 0,140 0,130 0,119 0,119
0,082 0,082 0,093 0,093 0,093 0,092 0,103 0,085 0,118 0,108 0,107 0,093 0,087 0,093 0,093
0,082 0,082 0,093 0,093 0,093 0,092 0,103 0,085 0,118 0,108 0,108 0,107 0,093 0,087 0,093 0,093
0,043 0,043 0,048 0,048 0,048 0,048 0,078 0,044 0,098 0,088 0,088 0,043 0,043 0,043 0,043
0,033 0,033 0,033 0,033 0,033 0,033 0,062 0,039 0,093 0,082 0,082 0,043 0,043 0,033 0,033
0,033 0,033 0,033 0,033 0,033 0,033 0,062 0,039 0,093 0,082 0,082 0,043 0,043 0,033 0,033
0,060 0,060 0,071 0,071 0,071 0,071 0,097 0,066 0,097 0,092 0,092 0,050 0,060 0,071 0,071
0,030 0,030 0,040 0,040 0,040 0,040 0,066 0,035 0,071 0,061 0,061 0,030 0,030 0,040 0,040
0,019 0,019 0,029 0,029 0,029 0,029 0,054 0,024 0,064 0,054 0,054 0,019 0,019 0,029 0,029
0,059 0,049 0,064 0,064 0,064 0,064 0,069 0,054 0,090 0,080 0,080 0,080 0,080 0,069 0,069
0,057 0,062 0,077 0,077 0,077 0,077 0,098 0,075 0,113 0,103 0,103 0,067 0,077 0,082 0,083
0,053 0,043 0,062 0,062 0,062 0,062 0,087 0,054 0,087 0,077 0,077 0,057 0,067 0,062 0,063
0,078 0,068 0,088 0,088 0,088 0,088 0,088 0,070 0,108 0,098 0,098 0,073 0,063 0,063 0,063
0,050 0,045 0,050 0,050 0,050 0,050 0,066 0,055 0,103 0,092 0,092 0,071 0,061 0,055 0,055
0,118 0,113 0,124 0,124 0,124 0,124 0,145 0,123 0,150 0,140 0,140 0,124 0,129 0,129 0,130
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S.lufengia.CMMI2018I030005
S.lufengia.CMMI2018I228010
S.subcrustalis.CEUA016MR0010
S.laeta.MCZ.AM.IZ130696
S.morsitans.CUMZ00344
S.morsitans.CUMZ00342
S.morsitans.SYSU:Chilo208
S.morsitans.SYSU:Chilo005
S.pinguis.CUMZ00304
S.pinguis.CUMZ00309
S.pinguis.CUMZ00314
S.calcarata.SYSU:Chilo112
S.calcarata.CUMZ00418
S.calcarata.NHMUK010305528
S.multidens.SYSU:Chilo118

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0,014
0,047  0,057
0,072  0,082  0,048
0,048  0,057  0,053  0,077
0,048  0,057  0,053  0,077  0,000
0,053  0,063  0,058  0,078  0,014  0,014
0,043  0,053  0,048  0,073  0,005  0,005  0,009
0,077  0,093  0,082  0,093  0,098  0,098  0,103  0,093
0,072  0,087  0,082  0,103  0,093  0,093  0,098  0,088  0,033
0,073  0,088  0,078  0,093  0,098  0,098  0,099  0,088  0,028  0,043
0,108  0,124  0,151  0,156  0,135  0,135  0,140  0,130  0,093  0,103  0,099
0,077  0,087  0,081  0,098  0,093  0,093  0,098  0,088  0,043  0,057  0,048  0,103
0,077  0,087  0,081  0,098  0,093  0,093  0,098  0,088  0,043  0,057  0,048  0,103  0,000
0,024  0,033  0,058  0,088  0,058  0,058  0,063  0,053  0,088  0,083  0,078  0,119  0,073  0,073
0,014  0,028  0,057  0,082  0,057  0,057  0,063  0,053  0,087  0,072  0,083  0,119  0,087  0,087  0,019
0,014  0,028  0,057  0,082  0,057  0,057  0,063  0,053  0,087  0,072  0,083  0,119  0,087  0,087  0,019
0,060  0,071  0,092  0,125  0,087  0,087  0,087  0,081  0,130  0,119  0,114  0,158  0,119  0,119  0,071
0,030  0,040  0,061  0,092  0,066  0,066  0,071  0,061  0,087  0,082  0,076  0,131  0,087  0,087  0,040
0,019  0,029  0,049  0,080  0,054  0,054  0,059  0,049  0,090  0,080  0,074  0,127  0,085  0,085  0,029
0,049  0,064  0,069  0,085  0,085  0,085  0,090  0,080  0,064  0,054  0,049  0,106  0,080  0,080  0,059
0,053  0,062  0,087  0,098  0,082  0,082  0,088  0,078  0,072  0,087  0,078  0,130  0,082  0,082  0,068
0,048  0,057  0,077  0,098  0,072  0,072  0,078  0,068  0,067  0,082  0,073  0,108  0,082  0,082  0,048
0,058  0,068  0,073  0,093  0,073  0,073  0,078  0,068  0,078  0,078  0,088  0,130  0,083  0,083  0,063
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0,124  0,134  0,140  0,145  0,140  0,140  0,146  0,135  0,124  0,124  0,130  0,173  0,129  0,129  0,119
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0,000										
0,066	0,066									
0,035	0,035	0,046								
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0,054	0,054	0,097	0,066	0,059						
0,062	0,062	0,087	0,061	0,059	0,069					
0,057	0,057	0,081	0,061	0,049	0,049	0,048				
0,068	0,068	0,108	0,076	0,064	0,064	0,088	0,048			
0,050	0,050	0,094	0,067	0,055	0,055	0,082	0,066	0,087		
0,129	0,129	0,153	0,131	0,122	0,101	0,113	0,072	0,114	0,120	