# Flexible selection of working memory representations to reduce cognitive cost

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# Abstract

Across psychology, economics, and decision science, it is well established that choosing a good internal representation can make complex problems easier to solve[1,2]. This idea, central to theories of bounded rationality[3] and heuristics[4,5], suggests that intelligent behavior depends not only on what is stored in working memory, but on the format in which it is stored. Whether non-human animals flexibly select internal formats, and how such choices are implemented in neural circuits, has remained unclear. Here we show that rats adjust the format of working memory to reduce cognitive load. When a task encourages an action-based code, rats maintain the relevant information as a stable motor plan supported by frontal cortex. When the same task is reformulated to make this code costly, they instead store a sensory trace in auditory cortex. These strategy shifts are rapid and reliable, revealing that rats can switch between distinct neural circuits to store information in the most economical internal format. These results demonstrate that flexible representational selection is not unique to humans[6] but is present in rodents and reflected in circuit-level reallocation of working memory. This establishes a neural basis for classic theories linking problem representation to computational efficiency and provides a path toward a neural circuit-level description of selecting internal formats to reduce cognitive costs.

# Introduction

Across many domains of human reasoning, the difficulty of a problem depends critically on how it is represented. In physics, when modeling a ball spinning around on a string, switching to a rotating frame transforms a complex problem with time-varying forces into a simple one. In arithmetic, reframing $49 + 49$ as $(50 + 50 - 2)$ reduces the computation to a trivial adjustment. In psychology, economics, and decision science, this general principle—*reframing can reduce cognitive load*—lies at the heart of bounded rationality[3], heuristics and biases[4,7], fast-and-frugal decision strategies[5], classical theories of problem representation[1], and modern analyses of strategy and organizational reasoning[2]. These perspectives converge on the idea that cognitive systems simplify problems by selecting an internal representation that makes the required computation tractable. Yet how the brain flexibly selects between representational formats remains poorly understood[8].

Working memory (WM) provides a natural domain in which to study representational selection. WM can store information in distinct internal formats: sensory traces, abstract codes, or prospective motor plans. Depending on the context, these formats have different stability, precision, and computational costs. Resource-rational theories propose that cognitive systems balance these costs against task demands[9–11]. While most theoretical and empirical work has focused on what kind of information to maintain[12] or how deeply to search[13], there is recent evidence that humans select representations that trade off costs and benefits in planning[6]. Whether non-human animals flexibly choose between WM formats to minimize representational burden remains unknown.

To investigate whether working memory formats are flexibly chosen according to representational demands, we designed two parallel auditory delayed-response tasks using an 8-port orienting wall (FIG. 1a,b). The tasks share identical motor requirements but differ in how costly it is to represent the mapping from sensory cue to action. In the *Ego* task, sounds map directly onto egocentric directions, encouraging early construction and maintenance of a motor plan (FIG. 1c:H1, Ego task). In the *Allo* task, sounds map to world-centered target locations, so that generating an egocentric motor vector requires subtracting the start position from the target location, resulting in many possible movement vectors (FIG. 1c:H1, Allo task). Critically, animals could either compute and retain

the motor vector (costly in the Allo task) or instead maintain a sensory WM trace and perform the transformation later (FIG. 1c:H2) or maintain a WM trace about the location of the target port (FIG. 1c:H3). This framework isolates representational-format choice as a key decision variable in reducing WM load (FIG. 1d).

One consequence of this framework is that it encourages us to question one of the most robust findings in neuroscience: reports of persistent, action-predictive activity in mammalian frontal cortex during delayed-response tasks. Such findings, observed in mice[14,15], rats[16,17], monkeys[18–20], and humans[21,22], have supported the view that frontal motor planning is the default mechanism for bridging delays[23,24]. However, the paradigms used in these studies impose strong structural constraints: subjects are head-fixed or held in a fixed posture, and responses are limited to a small set of stereotyped movements. Such designs favor early sensorimotor transformation and maintenance of a motor plan, potentially masking alternative representational formats.

If working-memory format is flexibly chosen according to representational cost, then relaxing these constraints should reveal strategies beyond frontal motor planning[8]. Indeed, we found that altering representational demands revealed distinct WM strategies. In the Ego task, neurons in the frontal orienting fields (FOF) in secondary motor cortex, exhibited robust prospective coding during the delay and FOF inactivation impaired performance, indicating a motor WM format in FOF. In contrast, in the Allo task, FOF did not exhibit motor-planning nor location-planning signals, and FOF inactivation had no behavioral effect, ruling out FOF based motor or location WM. Instead, auditory cortex exhibited sensory WM signals consistent with a retrospective code. A cost-sensitive dynamical model showed that maintaining a motor vector in the Allo task imposed higher representational load, predicting the observed shift toward a sensory format. When we simplified the Allo task by fixing the start position, animals rapidly reverted to frontal motor planning, and FOF became necessary again.

These results demonstrate that animals choose a WM format that minimizes cognitive load[10,25,26]. This establishes representational format as a core dimension of cognitive strategy selection and reveals a neural mechanism by which distinct cortical circuits are flexibly recruited to implement these formats.

# Results

## Two auditory memory-guided tasks in different reference frames

We separately trained two groups of rats on either the Ego or the Allo task (details of subjects can be found in TABLE S1). The Ego and Allo tasks share the same trial structure. In both tasks, trials started with the illumination of a randomly selected start port. Rats needed to poke into the start port and remain fixated with their nose in start port for 1.5 s. During the fixation period, rats heard one of two 0.75 s sound cues followed by another 0.75 s silent delay period. An auditory 'go' cue signaled the end of fixation, after which rats reported their choice. If they poked into the correct target port (described below) they could collect a reward in the reward port (FIG. 1a).

The correct target port was defined based on task context and sound cue. For the Ego task, sound A indicated that the target was to the left and sound B indicated that the target was to the right of the start port (FIG. 1b-left, FIG. S1a-left). For the Allo task, the bottom left & bottom right port in the wall were the target ports for sound A & B, respectively, regardless of start position (FIG. 1b-right, FIG. S1a-right). Rats performed both tasks significantly above chance (Allo task: 69 ± 1% correct, n=20 rats, 2011 sessions, 412,922 trials. Ego task: 76 ± 2%, n=9 rats, 506 sessions, 85,597 trials).

**Figure 1.** Task design and behavior quantification **a.** Schematic of the task. **b.** Mapping the sounds to the ports in the two tasks. Ego task: sounds map to movement directions. Allo task: sounds map to port locations. **c.** Given the structure of our task and prior literature, we considered three possible working-memory formats associated with representational transformations at different times in the trial (purple color represents for cue, gray for motor, and blue for location formats) H1: A self-centered movement-vector format in the FOF. This is efficient in the Ego task, but costly in the Allo task. H2: A sensory working-memory format in posterior cortex[8]. This format has low WM load in both tasks, but sensory WM is generally less robust than motor WM[27]. H3: A world-centered location-based format in the FOF[28]. This format has high WM load in the Ego task, and low load for Allo. Our task design allowed animals to adopt any of these three formats, enabling us to test whether and how representational demands steer the choice among them. **d.** Schema of working-memory load across tasks for H1 vs. H2. The Ego task (blue dot) requires 1 bit of WM regardless of strategy. The Allo task (orange dot) also requires 1 bit of WM in the sensory frame, but has a higher WM load in the motor frame. The purple and dark green background colors highlighted the preference for using motor or sensory strategies for working memory. The gradient is shifted so that for equal memory load, a motor strategy is favored, reflecting the benefits of movement planning (faster movements) **e.** Histograms of movement times (MT) for the Ego and Allo tasks, for center-start trials. Correct (Top) and error (Bottom) trials. Vertical line: median of MT. **f.** Example key points and vector angles feature for video decoding analysis. **g.** Head direction trajectories from one example session. Thin lines are individual trials. The gray box highlights the fixation period. **h.** Cross-validated (CV) decoding accuracy for choice from posture features across time in a trial. Shade: 95% CI of the mean across subjects. The blue and orange thick bars at the top indicate above-chance accuracy for more than half of the subjects for the Ego and Allo task ($p < .05$, permutation test with FDR correction). The thick grey bar at the top indicates the time when the decoding accuracy was significantly different on the Ego versus Allo task ($p < .05$, two-sample $t$-test with FDR correction).

Chance performance in the task could be considered to be 1/6, since there are 6 ports available on the port wall beside the start port. However, given a task and start port, there were only two possible target ports (associated with the two sounds). An analysis of error trials showed that rats understood the structure of the tasks: most of the errors were made to the port corresponding to the unheard sound cue (Ego task: 94.9%, Allo task: 94.7%, see FIG. S1b). Performance on the Ego task was significantly higher than the Allo task ($t(22) = 4.205$, $p = .0002$, two-sampled $t$-test; FIG. S1g).

A key feature of our experimental design was that trials started from the center port were identical in the two tasks. For example, in both the Ego and Allo task, when starting from the center port, sound A indicated that the target is down to the left (FIG. 1b, trial type 2). By restricting analyses to trials starting from the center port, 'center-start' trials, all sensory and motor aspects were identical across the two tasks, so any behavioral (or neural) differences must reflect the task context (i.e., differences in strategy that animals are using to solve the two tasks). Here, we restricted the analysis of behavioral data to these center-start trials.

In delayed-response tasks, movement planning is associated with faster movements and small postural adjustments during the delay period[16,29,30]. In line with this, we found that the time from exiting the center port until reaching the chosen port (movement time, MT) was, on average, 41.8 ms shorter on the Ego task trials compared with the Allo task (FIG. 1e top; rank-sum test: U=38.5, $n_{Ego} = 9$, $n_{Allo} = 20$, $p = .016$). From error trials, we observed a similar result: $\Delta MT$ =49.8 ms (FIG. 1e bottom; rank-sum test: U=30, $n_{Ego} = 9$, $n_{Allo} = 20$, $p = .005$) This $\approx$50 ms difference is consistent with the previous reports comparing movements with vs. without planning[15,16,31–33], suggesting that animals in the Allo less prepared for the movement than animals in the Ego task.

Movement planning during the delay period is often associated with postural adjustments[16,29,34]. By estimating posture from video, we observed that rats tended to turn slightly toward their intended target during fixation (FIG. 1g). We trained a logistic classifier to predict the upcoming choice from posture in the two tasks. Choice was more accurately decoded from rats' posture during the delay period and early movement period in the Ego task than in the Allo task (FIG. 1h, left panel). Note that as rats neared the target port we were able to decode their choice perfectly in both tasks, validating our decoding method (FIG. 1h, right panel). These differences in MT and posture were robust across animals (FIG. S1c,d) and were not driven by differences in performance (FIG. S1e). Together, the posture and movement time results were consistent with the hypothesis that rats in the Ego task performed their sensory-motor transformation early, used a prospective motor plan to bridge the delay period (FIG. 1c:H1). In contrast, rats in the Allo task seemed to delay the sensory-motor transformation and used a retrospective memory of the sound (or sound category or location) to bridge the early delay period (FIG. 1c:H2 or H3).

## FOF neurons encode start port in both tasks, but only encode future choice in the Ego task.

In memory-guided 2AFC tasks, FOF activity encodes upcoming choice and causally contributes to task performance[16,17,35–37]. Using the poke wall, we recently found that FOF neurons encode both self-centered movement plans and world-centered target positions[28]. Given the behavioral evidence for different mnemonic strategies between the two tasks, we wanted to investigate how the contribution of the FOF might differ between them. We recorded single-unit activity from the FOF in rats performing both Ego and Allo tasks (Ego task: n = 2887 neurons, 4 rats; Allo task: n = 3325 neurons, 8 rats).

We first explored the encoding of two key task variables in individual neurons on correct

trials: choice and start position. In the Ego task, start position was not essential for solving the task, since the sounds mapped to ports relative to the start position. In the Allo task, rats must combine the start position information and the sound to compute a movement vector to the target. FOF neurons exhibited tuning for task variables to different degrees in the two tasks (Example neurons: FIG. 2a-c,e-g). For each cell, we fit two Poisson generalized linear mixed-effects models (GLMMs) to quantify the choice coding during the delay and start port coding during the whole fixation period (see Methods). We found that a significantly greater proportion of neurons showed selectivity for the upcoming choice in the Ego task compared to the Allo task (Ego task: $864/2589 = 0.33$; Allo task: $374/3186=0.12$; $\chi^2(1, N = 5775) = 396.90$, $p < .001$; FIG. 2d,h). This finding was robust across subjects (FIG. S3). Moreover, 'pure' choice neurons were rare in the Allo task and had relatively weak tuning. The striking difference in choice coding (comparing the spread in the x-axes between FIG. 2d,h) was in contrast to the robust coding for start position in both tasks. The lack of choice coding in the Allo task was surprising given the wide consensus about the role of FOF (and frontal cortex more generally) in delayed-response tasks.

**Figure 2.** Single neuron quantification for choice and start port tuning **a-c** Example neurons from the Ego task. The rasters and PSTHs were aligned to the sound cue onset. The rasters were sorted by different start ports (indicated by green-orange colors) and upcoming choices (red solid line for right choices, blue dashed line for left choices). The top PSTH was grouped by start port, the lower by upcoming choice. The bottom row (firing rate versus start port position) summarizes the relationship among the neural activity, start port position and choice. The lines are fits of a linear model (`Hz ~ start_port*choice`) to the data (dots with error bars represent mean $\pm$ SEM). **d** Distribution of coefficients of start-port (y) vs. choice (x). Black circled data show where the three example neurons shown in panel a-c are located in the scatter plot. Colors of the points indicate the significance of tuning (see legend). **e-g.** same as panel a-c, but from the Allo task **h.** same as panel d, but from the Allo task.

## Choice coding in the FOF is persistent and stable in the Ego but not the Allo task

The single-cell selectivity analysis provided a useful first indication that the FOF plays distinct roles across the two tasks. To ensure that sensory and motor variables were fully matched, we focused subsequent analyses on center-start trials to examine the dynamics of choice and start coding. At the single neuron level, from the end of the cue period (0-0.75 s) until after the go signal, there was a significantly larger fraction of neurons with choice coding in Ego than in Allo animals (Fig. 3a).

Visualizing the encoding of task variables as *proportion of selective neurons* can obscure a representation that is weak in each cell but robust in the population[38]. We employed pseudopopulation decoding to examine choice coding in population space. The decoding accuracy of the Ego task ramped up quickly over the trial, with a similar time-course as the rise in individually significant cells, and remained higher than the Allo task throughout the trial (Fig. 3b, $p < .01$, permutation test). Moreover, choice coding in the Allo task was significantly above chance only slightly before the go cue. The single-cell and pseudopopulation results strongly suggest that the Allo task is not supported by the FOF.

To better understand the stability of choice coding, we trained decoders at one time in the trial and then tested them at other times (Fig. 3d). We found that decoding for choice in the Ego task was stable across the entire delay period (Fig. 3d, left panel, within red square), but absent in the Allo task (Fig. 3d, right panel, within red square). The strong and stable choice coding in the Ego task is consistent with previous findings in delayed-response tasks in M2[28,39]. However, the absence of choice coding in the Allo task was surprising, especially given that the analyses were restricted to center-start trials. This provided further evidence that animals in the Allo task were delaying the sensory-motor transformation until the end of the delay period. To reiterate, the lack of choice coding in the FOF in the Allo task on center-start trial implies that the FOF activity, *in any reference frame*, does not encode upcoming choice, ruling out H1 (FOF motor WM) and H3 (FOF location WM) for the Allo task

We also examined the dynamics of the start position coding in FOF at both the single cell and population level. At the single-cell level, throughout the trial, we observed more cells significantly encoding the start position in the Allo task than in the Ego task (Fig. 3e). Decoding performance for start position from the pseudopopulation in the Allo task was strong and stable for the entire fixation period. Decoding for the Ego task was also strong. However, it was significantly worse than the Allo task during most of the early cue and delay period (Fig. 3f, $p < .01$, permutation test). To better understand the dynamics of the start position coding in FOF, we performed cross-time decoding for pseudopopulations of Ego and Allo task (Fig. 3g). We found that, in both Ego and Allo tasks, start position at any time during fixation could be decoded from any other time during fixation, indicating a stable start-position code. However, consistent with the single unit findings, start position was decoded better in the Allo task (Fig. 3g).

One limitation of pseudopopulation analysis is that it may miss true encoding that depends on correlations between neurons, masked by large trial-by-trial fluctuations. To address this, we performed moment-by-moment leave-one-trial-out decoding of start position and choice at the single-session level for both Ego and Allo. The results were consistent with both single-cell and pseudopopulation analyses: start position decoding was strong in both tasks but choice decoding was significantly stronger and earlier in the Ego task (Fig. 3c, Fig. S4). Overall, the earliest time with above-chance single-trial-decoding in the Ego task was before the go cue (From cue onset: $1.10 \pm 0.08$ s, n=51 sessions), which was much earlier than the Allo task, which on average emerged after the go cue ($1.67 \pm$

0.04 s, n=44 sessions, comparison: $p < .001$, permutation test, FIG. 3c). Taken together, the analysis of FOF activity on center-start trials showed clear differences between the two tasks. In the Ego task, FOF activity predicted upcoming choice, while in the Allo task there is a noticeable absence of choice activity until just before the go cue. Together with our demonstration of strong start-position coding in the Allo task (and the lack of correlation between start and choice coding in the Allo task or between choice coding and behavior performance, FIG. S4F-H), our results suggest an FOF independent strategy for solving the Allo task.

### In the Ego task, planning is in a self-centered frame that generalizes across start positions

Having shown that, for center-start trials in the Ego task, FOF persistently codes for choice during delay period, we wanted to better understand the reference frame of this code: specifically, is there a common left/right planning subspace for all start positions? Alternatively, each start position could have distinct planning activity for left and right choices. To visualize choice planning encoding at the population level, we analyzed FOF activity in an activity space where each dimension corresponds to activity of one simultaneously recorded neuron. We estimated a 'coding direction'[14,40]), $CD$, from the delay period of center-start trials along which activity maximally discriminated future choice (FIG. 3h, Methods). When left-out trials were projected to the coding direction, FOF population activity in the Ego task strongly separated by choice (FIG. 3i). However, in the Allo task, the same approach failed to show clear discrimination between choices (FIG. 3j). This recapitulates our earlier finding (FIG. 3b), but at the single-trial level, rather than in the pseudopopulation.

We then asked how the coding direction defined from center-start trials generalized to other start positions. In the Ego task, the coding direction generalized across start positions (FIG. 3k; FIG. S5a). This result showed that the FOF activity during the delay period represents an egocentric plan that is robust to variation in start position. As expected, in the Allo task, using the uninformative coding direction derived from center-start trials resulted in consistently poor decoding for other start positions (FIG. 3l; FIG. S5b). For the Ego task, the decoding accuracy during the delay period for 10-folds center-start trials was $.816 \pm .124$ SD Using the center-start decoding other start positions, the accuracy was .774 $\pm$ .138 SD (FIG. 3m, n=60 sessions, total least square (TLS) regression slope=1.20, 95%CI [0.78, 1.72], $p = .008$ from bootstrap for slope$> 0$; $R^2 = 0.6341$). This result confirmed that in the Ego task, the activity pattern in FOF was the same for planning leftward/rightward movement across different start positions, i.e. planning in an egocentric reference frame. In the Allo task, decoding during the delay period was low on center-start trials ($.59 \pm$ .12 SD), and even lower when generalizing to other start-positions ($.52 \pm .04$ SD; FIG. 3n, n=70 sessions, TLS regression slope=0.07 [-0.03,0.17] 95%CI; $p = 0.081$ from bootstrap; $R^2 = 0.03$).

### Silencing FOF impairs performance in the Ego task, but not the Allo task

The electrophysiological results that FOF supports a movement planning strategy for the Ego but not for the Allo task. To causally test this we expressed stGTACR2[41] in pyramidal neurons bilaterally in FOF and implanted fibers for delivery of light laser (FIG. S10d,e). A sham fiber was also implanted at the posterior part of the skull. We used this sham fiber to habituate animals to tethering and laser delivery before any FOF silencing sessions. Even after habituation, we interleaved FOF sessions with control sessions where we connected to

**Figure 3.** FOF population encoding of choice and start port information **a.** Development of choice-dependent activity over the time course of the trial. Lines indicate % of cells at each time with significant left/right choice selectivity on center-start trials. Grey bar indicates when the % selective cells was significantly different between the Ego vs. Allo task ($p < .01$, Chi-Sq test, FDR Correction) **b.** Cross-validated decoding accuracy for choice, across the trial, averaged over 100 pseudo populations. Shade is 99% CI The blue and orange thick bars at the top indicate above-chance accuracy for the Ego and Allo task. Grey bar indicates significant difference between Ego vs. Allo tasks ($p < .05$, permutation test with Bonferroni correction for number of time points). **c.** Distribution of choice decoding latency of each session. Dots are sessions. Error bars are 95% CI. **d.** Across-time cross-validated choice decoding accuracy averaged across 100 pseudopopulations. The decoders were trained at one time window (x-axis) and tested at another (y-axis). Contours, $p < .01$ (extreme pixel-based test). The white and red box highlight the fixation and delay period, respectively. **e.** As in (a) but for start-port selectivity over the trial. **f.** Cross-validated decoding $R^2$ for start position, across the trial, averaged over 100 pseudo populations. Shade is 99% CI. Grey bar shows significant difference between Ego and Allo tasks ($p < .05$, permutation test with Bonferroni correction). **g.** Similar to **d** but for start port decoding. **h.** Schematic of single-trial choice coding and generalization. The coding direction (CD) for estimated from center-start choices and then used to predict trials started from other ports. **i.** 95% CI of FOF neural activity projected to the CD (n=60 sessions) for left-out center-start trials in Ego. Blue/Red are left/right trials. **j.** same as **i** for the Allo task ((n=70 sessions). **k.** like **i**, but for trials other than center-start trials projected onto the center-start CD. **l.** as in **k**) but for the Allo task. **m.** Comparing CV-decoding accuracy of center-started trials against out-of-sample accuracy of trials started from other start ports for each session. Cross indicates mean and SD of decoding accuracy of each axis. **n.** same as **m** for the Allo task.

**Figure 4.** Effects of optogenetic silencing the FOF. **a.** Illustration for virus injection and fiber implantation location at bilateral FOF and a sham fiber implantation above the skull (Left). Coronal sections of a rat brain expressing stGtACR2-FusionRed (Right). **b.** Timeline of bilateral silencing of FOF during two different trial epochs (cue and delay). **c.** Performance modulation for cue period laser effect. Each FOF session is one point. **d.** Same as **c** but for the delay period. **e.** Comparison of cue period silencing between Sham and FOF sessions. Y axis: fraction of correct choices for laser-on-trials - laser-off-trials. Dot and error bar reflect mean and 95% bootstrapped CI of the mean across sessions. **f.** Same as **e** but for the delay period.

the sham fiber (FIG. 4a left). Optogenetically, we silenced bilateral FOF on 25% of trials by delivering 473nm blue laser (FIG. 4a right) during either the cue period or delay period (FIG. 4b).

For each session, we compared the performance, $p$(correct), between laser-off and laser-on trials during the cue and delay epochs. During the cue period, there was a small decrease in performance in both tasks (FIG. 4c). In contrast, during the delay period, nearly all Ego-task sessions exhibited a marked reduction in performance on laser-on trials relative to laser-off trials, whereas in the Allo there was no apparent difference between performance on laser-on vs. laser-off trials (FIG. 4d).

To more rigorously quantify the effect of silencing the FOF, we fit logistic generalized linear mixed-effects models (GLMMs) to quantify the impact of optogenetic inactivation of FOF on performance compared to sham sessions, during the cue and delay periods of the Ego or Allo task, separately (cue/delay × Ego/Allo = 4 models). For succinctness, $p$ values of likelihood ratio tests are reported in the corresponding figures. All details of the GLMMs are reported in methods and statistical appendix.

Silencing FOF in the cue period caused small but significant impairments in both tasks (FIG. 4e). However, delay period FOF silencing produced a larger performance impairment in the Ego task (FIG. 4f left). Consistent with weak choice coding for the Allo task in FOF, we didn't observe significant performance impairment for delay period FOF silencing for the Allo task (FIG. 4f right). Thus, silencing FOF during the delay period significantly impaired the Ego task but not the Allo task performance. Moreover, the FOF silencing effect was also observed at the individual subject level (FIG. S6c,d; TABLE S2).

However, comparing two groups to baseline is not the same as directly comparing them to each other[42]. To directly compare the FOF silencing effect between the two tasks, we designed a mixed-effect logistic model that considered performance fluctuation, task baseline difference, visual effect of the laser, and most importantly, the interaction between the two tasks and FOF silencing. By dropping the interaction term and comparing the reduced model, we rigorously quantified the difference in the effect of silencing FOF between the two tasks. We considered the cue period and the delay period silencing separately. For the cue period, we found no difference between the full and reduced model, indicating that the effect of silencing was not significantly different between the two tasks (p=.569, LRT). For the delay period, there was a significant difference (p=.005, LRT). Thus, during the short-term memory period (i.e. delay period), the silencing FOF generated a significantly greater impairment in performance in the Ego versus the Allo task. Detailed statistical results can be found in TABLE S3 and the statistic appendix.

One potential confound is that the overall performance on the Allo task is worse than the Ego task, which could make it hard to detect a significant effect of silencing. To address that concern, we re-ran our analysis using a subset of sessions from each task within the same range of performance (60 to 67 % correct). Even in this subset of the data, silencing FOF impaired Ego performance significantly more than Allo performance (p=.043, FIG. S6f). Note that this is very conservative, as typically more difficult tasks are more sensitive to perturbations[43].

Interestingly, despite the differences of FOF silencing on choice, in both tasks, silencing FOF during the delay reliably slowed reaction times and reduced fixation violations (FIG. S7b,f), confirming that the manipulation was behaviorally effective in both contexts. This suggests that while the contribution of FOF to upcoming choice is distinct between the two tasks, FOF may be contributing to general movement timing and impulsivity[44] in both tasks.

## Auditory cortex retrospective activity supports the Allo task

In the Ego task, we showed that FOF maintains short-term memory in the delay period. The mechanism supporting short-term memory in the Allo task, however, remains unclear. To identify which brain regions carry short-term memory signals in this context, we recorded from several regions that have been associated with spatial and non-spatial forms of short-term memory: medial prefrontal cortex (mPFC)[45–47], the superior colliculus (SC)[48], and the auditory cortex (AC)[49].

As expected, we found strong cue-related responses in AC, with some neurons clearly responding to the individual clicks of the stimuli (FIG. 5b). We also observed neurons that were selective for the sound cue during the silent delay period (FIG. 5a). To quantify cue coding across the population, we fit a Poisson GLMM for each neuron (`Firing rate ~ cue + (1|start port)`; see Methods), using the regression coefficient for cue as an index of selectivity. This analysis revealed that 51% of AC neurons were significantly selective during the cue period, while 18% were significantly selective during the delay period (FIG. 5b green and blue, 18% of 1866 neurons, binomial test against 0.05, $p < .001$). Notably, some delay-selective neurons showed no cue-period response (FIG. 5a, left), suggesting that delay activity is not simply a persistence of cue responses, but could be a distinct representation.

Error trials provided an opportunity to address two key questions: first, whether AC activity represents the retrospective cue or the prospective choice (i.e., whether the sign of coding was preserved or inverted), and second, whether a weaker coding strength during the delay could explain behavioral errors. During the cue period, although AC neurons maintained strong selectivity in error trials and also preserved the same direction, the selectivity was significantly worse on errors compared to correct trials (TLS regression slope = 0.61, 99% CI [0.58, 0.64] from bootstrapped over cells, $R^2 = 0.86$; FIG. 5c, left). This suggests that while AC is clearly encoding the sensory cue, since the slope is positive, levels of engagement or arousal may influence sensory encoding and behavior, since the slope is significantly less than 1. During the delay, neurons continued to show cue-related signals in error trials (positive slope), but with substantially weaker strength compared with correct trials (TLS regression slope = 0.28, 99% CI [0.19, 0.36], 99% CI, $R^2 = 0.23$; FIG. 5c, right). At the population level, single-trial decoding accuracy from simultaneously recorded neurons confirmed this retrospective coding and revealed the link to behavior: cue-period decoding was equally strong in correct and error trials (correct: 0.96, 95% CI [0.94, 0.97]; error: 0.943 [0.89, 0.96], bootstrapped over sessions; Wilcoxon signed-rank test, $p = 0.091$; FIG. 5d-Left), but during the delay decoding was significantly weaker in error trials (correct: 0.64, 95% CI [0.60, 0.68]; error: 0.58 [0.55, 0.62], 95% CI; Wilcoxon signed-rank test, $p = 0.0034$; FIG. 5d right). These results suggest that AC encodes the auditory cue both during the cue and the delay, and that these representations contribute to task performance.

We compared the time course of cue decoding accuracy between AC and FOF on correct trials only, where the cue and choice have a one-to-one mapping. Decoding in AC rose immediately after cue onset, reached near-perfect accuracy throughout the cue epoch, and remained above chance well into the delay. In contrast, decoding in FOF emerged only very late in the delay (FIG. 5f). We then quantified decoding strength across all four regions at two representative time-points during the delay. The decoding from AC recordings was significantly higher than FOF, SC, and mPFC (t = 0.95 s: $p < .01$ for all comparisons; t = 1.05 s: $p < .05$ for all comparisons, ranksum test, see statistical appendix; FIG. 5g). Together, these results indicate that AC provides the strongest and earliest population-level representation of the cue in the Allo task during the delay period.

We next tested whether cue coding during the delay occupied the same or a distinct

**Figure 5.** Auditory cortex (AC) activity in the Allo task. **a.** Example neurons from the AC recorded in the Allo task. Spike raster plots and PSTHs of three representative neurons show different dynamics for cue coding in the delay period (Left), cue period (Middle), or both (Right). Notation is the same as FIG. 2e-g. **b.** Distribution of coefficients of single AC neuron cue coding for cue period and delay period spike count. Example neurons from (a) are circled. **c.** Comparing cue selectivity on correct vs. error trials during the cue (left-purple) and delay (right-cyan). Each point is a single neuron in AC. **d.** Comparing cue decoding on correct and error trials using 10-fold cross-validation during the cue (left-purple) and delay (right-cyan). Points and error bars indicate mean and 95% CI. **e.** Schematic of recorded regions: frontal orienting field (FOF), medial prefrontal cortex (mPFC), auditory cortex (AC), and superior colliculus (SC) **f.** Averaged cue (or choice) decoding accuracy for correct trials across sessions in FOF and AC. The purple and red thick bar at the top indicate when the accuracy was significantly above chance for each region ($p < .01$, permutation test). Grey bar indicates when AC accuracy was significantly higher than FOF ($p < .05$ with FDR test, permutation test). **g.** Cumulative distribution of cue/choice decoding accuracy across sessions (correct trials) in each region, shown for two representative times after cue onset (Left: 0.95 s and right: 1.05 s). **h.** Cross-time decoding of cue (or choice) on correct trials. Similar to FIG. 3d. The white and red boxes highlighted the cue and delay period. Contours, $p < .01$ (extreme pixel-based test)

**Figure 6.** Dynamical model of cost-sensitive WM format selection. **a.** Schematic of the model. The selective excitation dynamically shifts between FOF and AC depending on the cost of the strategy. **b.** Cost landscape. The blue dot represents performing the Ego task with an FOF-dependent self-centered strategy. The orange square represents performing the Allo task with an FOF-dependent self-centered strategy. The orange dot represents performing the Allo task with an AC-dependent sound strategy. The red dotted line highlighted the contour for cost=3 bits. **c.** Distribution of $w_{FOF}$ (weight of using FOF network) in 'expert' models from 1000 simulations. Error bars represent 95% of distribution. **d.** Example activity from the FOF node in the Ego task (left) and the Allo task (right). Solid line represent the mean activity for each choice. Light traces represent activity from selected example trials. **e.** Time-course of decoding accuracy in the FOF network for left/right choice, colored differently for Allo and Ego tasks. Lines represent the mean, shaded areas are 95% CI. **f.** Example FOF node activity during silencing in the Ego task during the delay period. Similar to d, solid traces show the mean activity for future left/right choices, while dashed lines represent the mean activity without silencing. Shaded areas indicate the silencing period (0.75 to 1.5s). **g.** Effect of simulated optogenetic silencing the FOF node for both Ego and Allo tasks. Circles and error bars represent the mean and 95% CI, respectively. **h.** Examples of decoding results for time during the trial from Ego and Allo simulations. **i.** Time decoding mean $R^2$ for the Allo and Ego tasks from FOF model activity. Error bar represent SEM. **j.** Time decoding $R^2$ for the Allo and Ego tasks from FOF recording data. Circles for individual recording sessions. Violins show kernel density estimation for $R^2$ across all sessions. Error bars: SEM

subspace compared with the cue period. Cross-temporal decoding, restricted to correct trials to avoid confounds between cue and choice, showed that AC representations were highly stable within the cue epoch, but after cue offset transitioned into a distinct subspace that remained stable throughout the delay (Fig. 5h). This demonstrates that delay-period memory was not simply a persistence of cue responses, but was recoded into a separate representational space, consistent with rotational dynamics previously described in auditory cortex[50]. Such a cue-to-delay subspace shift was unique to AC: in FOF, SC, and mPFC, decoding either emerged only late in the delay or remained weak. At the population level, session-wise decoding confirmed that AC provided stronger memory-related signals compared to FOF, mPFC and SC, especially during early delay period.

## A dynamical model of cost-sensitive working memory

Given the vast literature that implicates frontal motor cortex (including FOF) in similar memory-guided tasks[24,51,52], we were surprised to discover that performance in the Allo task was independent of FOF. However, it is well-known that frontal cortex dependent premotor planning is not the only form of working-memory[8]. For example, in delayed-comparison tasks, sensory features during the delay can be observed in sensory and prefrontal cortices[49,53,54]. Our recordings from AC suggest that animals in the Allo task used a sensory

memory strategy. Why were animals using a motor strategy in the Ego task and a sensory strategy in the Allo task? We posited that animals make a tradeoff when considering a mnemonic strategy in order to reduce overall costs[6,55]. On the one hand is the costs/bits of the representation and the other is the load (in bits) of a particular WM format (FIG. 6b). The total cognitive effort is the product of both quantities. If a trade-off across regions with different cost-per-bit's and mnemonic strategies can arise.

To explain our results we needed to make two assumptions. First, motor planning in a self-centered reference frame is the preferred reference frame and, as such, is assigned a relatively low cost-per-bit (compared to auditory sensory memory). Second, the FOF can only support planning in an egocentric reference frame. To the first point, motor planning also comes with the benefit of quicker movement times and better preparedness for the sensory consequences of movement[56]. Also, Grillner and colleagues have convincingly shown that the forebrain architecture supporting motor planning predates the emergence of high-level associative cortex: suggesting that planning in a self-centered, premotor coordinate frame is an evolutionarily ancient form of cognition[57,58]. To the second point, we have demonstrated here that planning in the FOF is in a self-centered coordinate system: decoding upcoming choice in the Ego task from one start-position generalizes to the others (FIG. 3i,k,m). This is consistent with results from a delayed-visually guided task, where we also found planning in the FOF to be in a self-centered frame[28].

To be more concrete: in both the Ego and Allo tasks, a sound category mnemonic strategy would require only 1 bit (Sound A vs. B). In the Ego task, a motor planning strategy also requires 1 bit (left vs. right). However, using a self-centered strategy in the Allo task requires at least $\approx 3.6$ bits, since there are 12 possible movement vectors (FIG. S1a). A sound-based memory is less preferred (has higher cost/bit) than a motor strategy, so in the Ego task where both strategies have the same load (1 bit), animals choose the more efficient representation. But in the Allo task, the increased load of representing the task in the self-centered frame, means that the sound-based frame is overall cheaper.

We instantiated this hypothesis in a dynamical model with three modules: one representing the FOF, one representing AC, and finally a module that provides selective excitation. We instantiated the FOF module with a bistable attractor in order to replicate the results from the optogenetics experiments. The AC module was a fixed performance module (did not have any dynamics). We set the cost of the motor planning in the FOF to be low (1.0/bit) and the cost of a sound-based memory to be higher (3.0/bit;FIG. 6a, see Methods). The selective excitation module drove either the FOF or the AC module based on the cost. The overall cost, $C$, of each memory system was defined as the 'memory cost per bit' (Biological Preparedness, $b$) $\times$ 'number of bits' (Load, $r$). The load for both tasks in the sensory auditory domain is 1 bit, since there are only two sounds. So, for both tasks the cost of using auditory memory is $C_{sound} = b \times r = 3 \times 1 = 3$. Using the FOF in the Ego task yields a cost of 1, $C_{FOF,Ego} = b \times r = 1 \times 1 = 1$ (FIG. S1a, Left). So, for the Ego task, using a self-centered FOF dependent strategy has lower cost. However, performing the Allo task in self-centered FOF dependent frame has a load of 3.6 bits (since there are 12 possible movement vectors, FIG. S1a, Right). Thus, $C_{FOF,Allo} = b \times r = 1 \times 3.6 = 3.6$, which is higher than $C_{sound} = 3$. Based on the current costs, the relative excitation ($w_{\text{FOF}}$), shifts to support the more efficient memory strategy, subject to noise, $\sigma_w$ and a time-constant, $\alpha$. Based on this task-dependent cost design, the relative excitation $w_{\text{FOF}}$ shifts to support the most efficient memory strategy following noisy gradient descent dynamics.

By design, in the dynamical simulations, the model heavily favors the FOF ($w_{\text{FOF}} \approx 1$) for the Ego task but favors the AC for the Allo task (FIG. 6c). In the Ego task, the FOF module received excitation and showed selective activity for upcoming left/right choice (FIG. 6d left). In the Allo task, there was no selective activity (FIG. 6d right). This

simulation result recapitulated our FOF recordings (FIG. 2). We were able to decode choice from the FOF module during the delay period, for the Ego task but not the Allo task (FIG. 6e). This replicated our findings from FOF recording in real animals during the delay period (FIG. 3b). Moreover, silencing the FOF module during the delay period inhibited the selective activity (FIG. 6f) and impaired task performance for the model in the Ego task but not the Allo task (FIG. 6g) similar to our optogenetic results (FIG. 4).

In the dynamical model, the selective excitation starts at the time of the sound and ramps until go cue (which is the end of the trial) consistent with observations of ramping dynamics in frontal cortex[59,60]. This generated selective activity in FOF module that qualitatively matched the time course of real FOF activity. We realized that the presence of ramping excitation in Ego but not Allo, would predict that decoding of time in the trial should be stronger in Ego vs. Allo. We quantified the time decoding performance as $R^2$ between estimated time and actual time during the trial: the FOF module had higher time-in-trial decoding for the Ego task than the Allo task (FIG. 6h,i). This finding was then confirmed in real data (FIG. 6j), providing evidence for selective excitation as a mechanism for switching mnemonic strategies.

## Rapid reinstatement of FOF dependence following the Allo-to-2AFC task transformation

In the dynamical model, the selection of mnemonic strategy is dynamically based on cost. An alternative model is that the selection of mnemonic strategy is a consequence of learning trajectory: since the Allo and Ego animals were trained on different tasks, they may have adopted different strategies through learning. Our dynamical model should rapidly switch to a FOF dependent motor planning strategy if the task was simplified so that the egocentric strategy fell below the cost of using the auditory mnemonic strategy. In contrast, the learning hypothesis would predict that Allo animals would continue to use an FOF independent strategy in the case of a simplified task. We simulated this idea in our model by removing all start positions except the center port (FIG. 7a). In this reduced task, the egocentric strategy has a cost of 1, and the selective excitation to FOF, $w_{FOF}$, increased within hundreds of trials (FIG. 7b). An increase in $w_{FOF}$ predicts several behavioral and neural consequences: faster movement times, performance impairments with silencing FOF, and appearance of delay period activity encoding upcoming choice.

We tested these predictions using a within-subject experiment. Animals, who had trained for months on the Allo task, were transferred to a reduced version of the task, 2AFC, where all trials were center-start trials (FIG. 7a, see FIG. S9a for details). We compared center-start trials between Allo and 2AFC. After task transformation, movement times began to speed up, with a clear reduction emerging by day 2 (FIG. 7c, TABLE S6). Overall, movement times were 22.3 ms faster after the rapid task reduction for correct trials (FIG. S9g from n=11 rats, Allo task: Median MT = 243.9 ms [240.8 247.5] 95% CI, 1226 trials; reduced 2AFC task: 221.6 ms [219.9 222.8] 95% CI, 4554 trials, $p < .001$, LRT). These faster responses provided behavioral evidence that animals were rapidly shifting to a motor-planning strategy: moving the sensory-motor transformation to earlier in the trial.

We then compared the causal role of FOF in the Allo task with the 2AFC task, in the same subjects. We followed the same experimental and analysis methods used to compare the causal contribution of FOF to the Ego and Allo tasks FIG. 4, but now for Allo vs. 2AFC task. For comparison, we re-plotted results from the Allo task where silencing FOF was not significantly different from Sham illumination (FIG. 4d,f-right). Amazingly, in 2AFC, we observed a significant impairment (FIG. 7d,e-right). Directly comparing the impairment between tasks, we found that silencing the FOF in the 2AFC task resulted in

**Figure 7.** Rats shift to an FOF dependent strategy after task transformation. **a.** We took expert Allo rats and transformed the task into a 2AFC task by removing all start positions except for center-start trials. **b.** Time course of $w_{\mathrm{FOF}}$ after the Allo-to-2AFC transformation. The solid line and shaded area represent the mean $w_{\mathrm{FOF}}$ and standard deviation (SD) of $w_{\mathrm{FOF}}$ across 800 simulated trials. **c.** The thin lines show each rat's median MT from center-start trials on each day (normalized to the baseline MT for each subject), aligned to the day of task transformation. We plotted the pooled median MT from the 7 days preceding the transformation at day=-1. The black line shows the mean $\pm$ SEM across rats. The stars mark days with a significant MT reduction relative to baseline (LRT, $p < .01$). **d.** Performance modulation for the delay period laser effect. Each point represents a single session one FOF session is one point. **e.** Comparison of delay period silencing between Sham and FOF sessions. Y axis: fraction of correct choices for laser-on-trials - laser-off-trials. Dot and error bar reflect mean and 95% bootstrapped CI of the mean across sessions. **f.** Across-time cross-validated choice decoding accuracy averaged across 100 pseudopopulations for the Allo (left) and 2AFC (right) tasks. The decoders were trained at one time window (x-axis) and tested at another (y-axis). Contours, $p < .01$ (extreme pixel-based test). The white and red box highlight the fixation and delay period, respectively. Compare with Fig. 3D. **g.** Cross-validated decoding accuracy for choice, across the trial, averaged over 100 pseudo populations. Shade is 99% CI. The orange and blue thick bars at the top indicate above-chance accuracy for the Allo and 2AFC task. Grey bar indicates significant difference between Allo vs. 2AFC tasks ($p < .05$, permutation test with Bonferroni correction for number of time points). Compare with Fig. 3b.

a significantly larger impairment than in the Allo task (p=0.0121, LRT, FIG. 7e). This confirmed our model prediction: within 2-6 days after simplifying the task, strategies shifted to be FOF dependent. Detailed statistical results can be found in TABLE S4 and the statistical appendix.

We then compared choice decoding performance before and after task simplification for center-start trials. Within days of task transformation, the decoding of upcoming choice emerged earlier in the trial compared to the Allo task (FIG. 7f). We found a significant plateau emerged after task transformation (FIG. 7f-right). During the second half of the delay period, decoding accuracy in the reduced 2AFC task was significantly higher than the original Allo task (FIG. 7g, thick grey bar for $p < .01$, from permutation test). While coding in the 2AFC task was weaker than the coding in the Ego task (FIG. 3b), we note that this activity appeared within days of reducing the task from Allo to 2AFC. Together, the optogenetic and electrophysiological experiments support our model predictions and provide evidence for cost-sensitive rapid switching of working memory strategies .

## Discussion

Here, for the first time, we have shown that non-human animals choose a working-memory (WM) format that minimizes cognitive load. Concretely, we show that frontal motor-planning is not universally adopted by animals to solve delay-response tasks. Instead, the reliability of finding frontal cortical WM in past literature may reflect similarity of task design: subjects are head-fixed or fixating and must execute brief, precise movements like licks, gaze shifts or reaches. Our findings creates a path for a circuit level mechanistic understanding of the diversity of WM strategies and the metacognitive[61] control of WM format.

The present study demonstrates that the frontal orienting fields (FOF) play a crucial role in maintaining prospective memory for the Ego task but not the Allo task. In the Ego task, FOF activity during the delay period was essential for performance, maintaining motor plans in self-centered coordinates (Hypothesis 1 in FIG. 1). In contrast, Allo task performance did not depended on FOF, and delay-period activity shifted to the auditory cortex (AC), suggesting that the animals relied on a sensory form of working memory (Hypothesis 2 in FIG. 1). A cost-sensitive model of working memory allocation accurately predicted these behavioral and neural changes across task transitions. When we simplified the Allo task, both the neural engagement of FOF and the behavioral response speed returned to the pattern observed in the Ego task, validating the model's prediction that animals flexibly allocate mnemonic resources based on computational cost. Together, these findings indicate that FOF supports planning in self-centered coordinates and that rats can rapidly switch to lower-cost mnemonic strategies when task demands change.

Although our analyses focused on choice-related delay-period activity, we also observed representations of start position, a world-centered variable, in the FOF. Start-position coding in FOF was notably stronger in the Allo task, where world-centered reference frames are behaviorally relevant. In the Allo context, the target ports are in world coordinates, so animals must subtract their start position from the target port associated with the sound cue to determine the correct movement plan, which may explain the stronger representation of positional information. Despite the presence of positional information in FOF, animals in the Allo tasks did not use a FOF-dependent location based strategy (Hypothesis 3 in FIG. 1), suggesting that this information cannot be used for planning, but instead only reflects the animals current position. The source of this signal remains uncertain, but FOF is strongly interconnected with posterior parietal cortex (PPC) and retrosple-

nial cortex (RSP), regions previously implicated in spatial navigation and position-based computations[62,63]. In our task, animals quickly move to the reward port after indicating their choice. For these kinds of movement sequences, position can could provide an internal error signal through efference copy[56]. Future studies should examine how world- and self-centered representations coexist and interact across these regions to support flexible spatial behavior.

Our findings speak to broader questions about where and how working memory is maintained. In delayed-response tasks, persistent delay activity has been consistently observed in frontal regions[14,16,18,21,23,64]. Frontal cortex therefore appears to be a part of a canonical circuit for working memory, especially when animals plan future actions. There are a few notable exceptions. For example, in a delayed-response task, in mice that adopted a passive strategy, waiting rather than actively moving, delay activity was encoded by posterior cortexGilad et al.[8], Gallero-Salas et al.[65]. In active mice, however, persistent activity was frontal, encoding future actions rather than past stimuli. These results, in the context of the broader literature, underscore that frontal motor-planning is the default mode of working memory in delayed-response tasks, but posterior representations can emerge as an alternative strategy. Our results extend this by providing a normative reason for this switch, picking a memory format that trades off costs and benefits: the active animals had higher performance but may have incurred a higher effort cost.

The mechanisms of working memory differ strikingly between FOF in the Ego task and AC in the Allo task. In the Ego task frontal cortex maintains self-centered, prospective plans, whereas in the Allo task AC maintains retrospective sensory traces. We also saw that in AC working memory occupies a representational subspace distinct from the sensory subspace during the cue period (FIG. 5h). This was also true, although, less dramatic for the motor planning in FOF: decoders trained at the beginning of the delay ($t = .75\,\mathrm{s}$) generalized up until the go cue ($t = 1.5\,\mathrm{s}$) and then decayed (FIG. 3d). This is consistent with previous literature: both sensory and motor WM have been shown to occupy distinct subspaces from there respective inputs or outputs[20,39,50]. Notably, stability of sensory vs. motor memory subspaces differ sharply. Frontal working memory is robust, showing ramping dynamics consistent with attractor dynamics (FIG. 6d)[17,66,67]. In contrast, sensory traces in AC are decaying (FIG. 5)[49]. This dissociation mirrors findings in human studies, where sensory working memory is more vulnerable to distraction than action-linked memory[68,69]. Our results offer a potential neural account of why action-related WM is 'prioritized'[70,71]: motor-planning codes are inherently more stable. This helps explain why animals normally rely on a frontal, action-oriented strategy in delayed-response tasks, switching to a sensory code only when the cost of motor planning becomes high. These findings raise broader questions about how general this sensory–motor contrast is across modalities. We examined auditory working memory, but visual and olfactory tasks may reveal different balances of sensory and frontal contributions[31]. They also highlight the need to understand how sensory and motor cortices interact during memory maintenance and transformation, and whether similar cost-driven strategy selection operates across these different sensory systems.

Given that sensory memory in AC decays well before the go cue, the question arises: how do animals in the Allo task maintain their choice? The most parsimonious explanation is that information is transferred to FOF or another downstream structure, such as striatum[72]. Consistent with this view, we observed choice-selective activity appearing at the end of the delay in FOF, suggesting late transfer of a stored representation from AC. Other possibilities cannot be excluded: unrecorded areas may maintain the memory through persistent or activity-silent mechanisms[73,74].

Simplifying the Allo task shifted working memory engagement back to frontal cortex,

resulting in faster responses, confirming a key prediction of the cost-sensitive model. In the model, cost is proportional to representational precision, measured in bits, but the Allo task also differs in other ways, including movement complexity and planning horizon. Longer or more complex movements may introduce additional motor-planning costs that influence strategy selection. These findings resonate with theories of resource-rational cognition[6,9,75,76] and metacognitive control[61], which propose that animals and humans allocate limited cognitive resources to maximize performance efficiency. In our model, cost sensitivity controls WM format through selective excitation of neural populations, but the biological mechanism implementing this control remains unknown. One likely candidate is the anterior cingulate cortex (ACC), which has been implicated in performance monitoring and cost–benefit computation[77]. Future work should incorporate within-session task switches to probe neural correlates of cost evaluation and strategy selection at finer temporal resolution.

In conclusion, our findings challenge the notion of a universal locus of working memory in frontal cortex. The FOF supports memory only when planning in self-centered coordinates, and rats can flexibly engage alternative strategies when environmental demands shift. This flexibility reflects a form of resource-rational cognition, where animals learn multiple strategies and select the most efficient WM format for a given context. These results open new directions for investigating the neural basis of metacognition, adaptive control, and the dynamic allocation of working memory resources across distributed cortical circuits.

# Supplemental Figures

**Supplementary Figure 1.** Task and behavioral details. **a.** Trial types for each task. The Ego task had 5 different start ports with 8 different start-port × auditory cue combinations. There were only two possible movement vectors in the Ego task: down to the left or right. The Allo task had 7 different start ports with 12 different start-port × auditory cue combinations. Each trial type in the Allo task is associated with a unique movement vector (12 total). **b.** Choice contingency table for each trial-type for the Ego task (Left) and the Allo task (Right). Rows: trial-type, sort by start-position (colored dots) and auditory cue (colored speaker), columns: chosen port. Color indicates the proportion of choices to the given port in each trial-type. The colors in each row sum to 1. Note that there are very few out-of-task errors: choices are either correct or to the port that would have been indicated by the unplayed sound. **c.** Distribution of movement times (MT) for each individual rat for center-start trials. MT is defined as the time from center port withdrawal to choice port entry, hit trials only. Orange: Allo animals; Blue: Ego animals. Allo animals are generally slower than Ego animals. **d.** Decoding accuracy for choice from postural features (at $t = 1.3\,\mathrm{s}$ after cue onset) for each individual rat (colored dot) for center-start trials. Dot size indicates the number of trials. Decoding accuracy was generally higher for the subjects in the Ego task. **e.** Scatter plot of overall hit rate vs. pose decoding accuracy on center-start trials, for Ego task (Left) or Allo task (Right). Lines are TLS fit. Shades are for 95% CI. There is no evidence that task performance improves with a more overt postural strategy. **f.** Scatter plot of MT vs. posture decoding accuracy on center-start trials. Each dot is a subject, for Ego task (Left) or Allo task (Right). Lines are TLS fit. Shades are for 95% CI. The correlation between MT and overt postural strategies across subjects is weak. **g.** Performance. Grey dots are individual rats and the line indicates the mean for each task.

**Supplementary Figure 2.** Learning curves over training. **a.** Learning curve for Ego task animals. Correct rate and timing (fixation duration and memory period duration) development over the number of sessions. **b.** Learning curve for Allo task animals. As in a.

**Supplementary Figure 3. Distribution of single-neuron selectivity indices across subjects. a.** Scatter plot showing the distribution of coefficients of the start port against choice from the mixed-effects linear models for individual neurons recorded from each rat performing the Ego task. Each dot represents a single neuron, with its position reflecting selectivity for upcoming choice (x-axis) and start port position (y-axis), corresponding to analyses in FIG. 2d. **b.** Same as (a), but for the Allo task, corresponding to FIG. 2h. Pie plots next to each panel summarize the proportions of neurons classified as non-selective (gray), choice-selective only (blue), start port-selective only (orange), or selective for both (purple) at the $p < .01$ level.

**Supplementary Figure 4.** Session-wise decoding performance for start position, choice, and time across subjects. **a.** Example session of start port position decoding. Each dot indicates a trial, performance was quantified as $R^2$ between actual and predicted X position. **b-i.** The map from subjects to colors of points is shown in the lower left corner of the figure. For all panels, Ego task results are shown on the left and the Allo task results on the right. **b.** Heatmaps of decoding performance for start position in the Ego (left panel) and Allo (right panel) task. Each row represents a session, with color reflecting decoding performance (cross-validated (CV) $R^2$). Sessions are ordered based on the center of mass of decoding performance (calculated as the dot product between time and decoding performance). The color bar adjacent to each heatmap indicates the subject identity for each session, consistent with the color legend shown below. **c.** Scatter plots showing the relationship between start decoding $R^2$ and behavioral performance for each session. Each point represents a session, color-coded by subject (Ego-left, Allo-right). **d.** Decoding performance as a function of the number of good cells recorded per session, as determined by quality metrics (see *Methods*). **e.** Like **b** but for choice decoding on center-start trials, with the color indicating the CV decoding accuracy. Note, that choice decoding in the Allo task emerges after the go cue. **f.** Like **c** but for choice decoding. **g.** Like **d** but for choice decoding. **h.** Relationship between choice and start position decoding performance across sessions. The dashed lines represent chance-level performance thresholds (50% for choice decoding). **i.** Time decoding performance (CV $R^2$) against the number of good cells for Ego and Allo tasks. Each point in the scatter plots corresponds to an individual session, color-coded by subject. The dashed lines represents the inclusion threshold based on cell count for a comparison in FIG. 6j.

**Supplementary Figure 5.** Trial type details for decoding generalization analysis **a.** for the Ego task, center-start trials were selected for the training set (Left) and other trials started from other ports but moved to the same direction were selected as test set (Right). Decoding results were shown in Fig. 3k and Fig. 3m. **b.** for the Allo task, center-start trials were selected for the training set (Left), trials started from other ports but moved to the same choice port were selected as test set (Right), decoding results were shown in Fig. 3l and Fig. 3n.

**Supplementary Figure 6. a.** Synthetic data that simulated optogenetic modulation for some animals and sessions across two tasks. Y axis: fraction of correct choices for laser on trials - laser off trials for each session (dots, colored by different animals). Shade: density estimation with Gaussian kernel, black dot: mean over all sessions, error bar: 95% CI (bootstrap). **b.** Performance of 8 potential GLMM models on the synthetic data. The best model, GLM7, was used for the main results. **c.** Performance modulation for the cue period laser effect for each animal. Y-axis: individual animals, X-axis: change in performance, (laser-on trials) - (laser-off trials), averaged across sessions for each subject. Error bars are 95% CI (bootstrapped over sessions). Purple: FOF sessions; Grey: Sham sessions. **d.** As in **c**, but for the delay period. **e,f.** Like FIG. 4E,F but restricting analyses to sessions with 60-67% performance on laser-off trials. Dots are sessions colored by different animals. Violins show kernel density estimation.

**Supplementary Figure 7.** Influence of FOF silencing on reaction times (RT) and fixation violations. RT was defined as the time from go cue (at 1.5s after the auditory cue onset) to withdrawal from the start port. A fixation violation was defined as exiting from the start port before the go cue. We quantified fixation completion rate as the fraction of trials without fixation violations. **a.** The distributions of RT for cue period laser-ON trials. The grey and purple lines indicated the cumulative distribution function (CDF) sham fiber trials and FOF fiber trials, respectively. p-values were from bootstrap. **b.** Same as **a** but for the delay period. Silencing FOF during the delay significantly slowed RT, compared to sham trials. **c.** Influence of cue period laser on RT for each animal. Y-axis: individual animals, X-axis: median of ΔRT across sessions for (laser-on trials - laser-off trials). Error bars are 95% CI (bootstrapped across sessions). Purple: FOF sessions; Grey: Sham sessions. **d.** Same as **c** but for the delay period. **e.** Effects of cue period laser on fixation. Y axis: change in fixation completion rate for (laser-on trials - laser-off trials) for each session (dots, colored by different animals). **f.** Same as **e** but for delay period laser effect. **g.** Effect of cue period laser on fixation for each animal. Y-axis: individual animals, X-axis: Change in fixation completion rate for (laser-on trials - laser-off trials) for each session. Error bars are 95% CI (bootstrapped across sessions). Purple: FOF sessions; Grey: Sham sessions. **h.** Same as **g** but for the delay period.

**Supplementary Figure 8.** Detailed results for AC, FOF, SC, and mPFC recording experiment. **a.** Scatter plot showing the distribution of coefficients of the sound coding in the cue period against the delay period from the mixed-effects linear models for individual neurons recorded from each rat performing the Allo task. Corresponding to analyses in FIG. 5b. **b.** Heatmaps showing session-wise decoding performance for choice on correct trials in the Allo task from AC, FOF, SC, mPFC sessions. Each row represents a session, with blue-yellow color reflecting cross-validated decoding accuracy. Sessions are ordered based on the center of mass of decoding performance (calculated as the dot product between time and decoding performance). The color bar to the right of each heatmap indicates the subject identity for each session, consistent with the color legend shown below each panel.

**Supplementary Figure 9. Details for task transformation experiments. a.** Experimental timeline for each subject, showing the sequence of sessions during the Allo and reduced 2AFC tasks with optogenetic manipulations and electrophysiological recording experiments. Each row represents an individual subject. **b.** Pseudo-population decoding accuracy for choice as a function of the number of cells in the pseudo-population. Decoding was performed at two key time points, during the delay period (t = 1.4 s from cue onset) and during the movement period (t = 1.8 s from cue onset). Error bars indicate SEM over pseudopopulations. Orange line represents data from animals in the Allo task. Blue line is from the same animals after the task transformation to 2AFC. The horizontal dashed line represents chance-level decoding. **c.** Session-wise heatmaps of choice decoding accuracy from electrophysiological data for the Allo (left) and reduced 2AFC (right) task. Each row represents a session, and color intensity reflects cross-validated decoding accuracy over time, aligned to cue onset. Sessions are sorted by the center of mass of decoding accuracy. The color bar to the right indicates the subject identity for each session, consistent with the color legend shown below. **d.** Optogenetic silencing effects on task performance ($\Delta P$(correct)) for individual subjects during the cue (left) and delay period (right), for the reduced 2AFC task. Each row represents a subject, and error bars indicate 95% CI estimated from bootstrap resampling. **e.** Comparison of movement times (MT) on center-start trials for each subject before and after task transformation from the Allo to the reduced 2AFC task. Each dot and error bar represents the median and 95%CI of the median (bootstrapped across trials) for each each subject. **f.** Comparison of posture decoding accuracy at t=1.3s after cue onset (during late-delay period) for each subject before and after task transformation from the Allo to the reduced 2AFC task on center-start trials. Each dot represents an individual subject. The horizontal and vertical dashed lines represent chance-level decoding thresholds. **g.** Movement time (MT) distribution for correct (Top) and error (Bottom) trials in the Allo (orange) and 2AFC (blue) tasks. Vertical lines: medians of MT.

**Supplementary Figure 10.** Histology results for FOF experiment. Note, that in the nomenclature of Paxinos and Watson[78] the area that we describe as the FOF is considered to be part of M2. **a.** Reconstructed probe track for each rat that implanted Cambridge Neurotech silicon probes in FOF. Solid line for the Allo task, dash line for the Ego task. **b.** Reconstructed probe track for each rat that implanted Neuropixel 1.0 probes in FOF. Solid line for the Allo task, dash line for the Ego task. **c.** Reconstructed optic fiber track for each rat with optogenetics surgery in bilateral FOF. **d.** Histology image for each rat from optogenetics experiment, in the Ego task. **e.** Histology image for each rat from optogenetics experiment, in the Allo task.

**Supplementary Figure 11.** Histology results for AC/SC/mPFC recordings. **a.** Reconstructed probe track for each AC recording rat. The grey bar indicates the channel selection positions for `bank0`. **b.** Reconstructed probe track for each SC recording rat. The grey bar indicates the channel selection positions for `bank0`. **c.** Reconstructed probe track for one single mPFC implanted rat with Cambridge Neurotech probe. Other rats that have both FOF and mPFC recordings using Neuropixel probes are shown in Fig. S10b)

.

**Supplementary Table 1.** Experiment type for all subjects

| subjid | strain | sex | task | experiment |
|---|---|---|---|---|
| JLI-R-2209 | Sprague-Dawley | M | Allo | Electrophysiology in FOF, Cambridge NeuroTech |
| JLI-R-2234 | Sprague-Dawley | M | Ego | Electrophysiology in FOF, Cambridge NeuroTech |
| JLI-R-2257 | Sprague-Dawley | M | Ego | Electrophysiology in FOF, Cambridge NeuroTech |
| JLI-R-0004 | Lister-Hooded | M | Allo | Electrophysiology in FOF, Cambridge NeuroTech |
| JLI-R-0011 | Lister-Hooded | M | Ego | Optogenetics in FOF |
| JLI-R-0015 | Lister-Hooded | M | Allo | Electrophysiology in mPFC, Cambridge NeuroTech |
| JLI-R-0016 | Lister-Hooded | M | Allo | Electrophysiology in FOF, Cambridge NeuroTech |
| JLI-R-0017 | Lister-Hooded | M | Ego | Optogenetics in FOF |
| JLI-R-0020 | Lister-Hooded | M | Ego | Optogenetics in FOF |
| JLI-R-0022 | Lister-Hooded | M | Ego | Optogenetics in FOF |
| JLI-R-0023 | Lister-Hooded | M | Allo | Electrophysiology in FOF, Cambridge NeuroTech |
| JLI-R-0024 | Lister-Hooded | M | Allo | Electrophysiology in FOF, Cambridge NeuroTech |
| JLI-R-0026 | Lister-Hooded | M | Allo | Optogenetics in FOF |
| JLI-R-0032 | Lister-Hooded | M | Allo | Optogenetics in FOF |
| JLI-R-0033 | Lister-Hooded | M | Allo | Optogenetics in FOF |
| JLI-R-0034 | Lister-Hooded | M | Allo | Optogenetics in FOF |
| JLI-R-0035 | Lister-Hooded | M | Allo | Electrophysiology in FOF and mPFC, Neuropixel 1.0 |
| JLI-R-0036 | Lister-Hooded | M | Allo | Optogenetics in FOF |
| JLI-R-0037 | Lister-Hooded | M | Allo | Electrophysiology in FOF and mPFC, Neuropixel 1.0 |
| JLI-R-0038 | Lister-Hooded | F | Allo | Electrophysiology in SC, Neuropixel 1.0 |
| JLI-R-0041 | Lister-Hooded | M | Allo | Electrophysiology in FOF, Cambridge NeuroTech |
| JLI-R-0042 | Lister-Hooded | M | Allo | Electrophysiology in FOF and mPFC, Neuropixel 1.0 |
| JLI-R-0044 | Lister-Hooded | M | Allo | Electrophysiology in FOF and mPFC, Neuropixel 1.0 |
| JLI-R-0046 | Lister-Hooded | F | Ego | Electrophysiology in FOF, Cambridge NeuroTech |
| JLI-R-0059 | Lister-Hooded | M | Ego | Electrophysiology in FOF and mPFC, Neuropixel 1.0 |
| JLI-R-0060 | Lister-Hooded | M | Ego | Electrophysiology in FOF and mPFC, Neuropixel 1.0 |
| JLI-R-0070 | Lister-Hooded | M | Allo | Electrophysiology in AC, Neuropixel 2.0 |
| JLI-R-0073 | Lister-Hooded | M | Allo | Electrophysiology in AC, Neuropixel 2.0 |
| JLI-R-0074 | Lister-Hooded | M | Allo | Electrophysiology in AC, Neuropixel 2.0 |

# Statistical appendix table for optogenetics experiment

**Supplementary Table 2.** $\Delta$p(Correct) and 95% CI between laser on/off trials for each rat on each experiment

| task | laser epoch | subjid | region | n_trials(control) | $\Delta$p(Correct) | 95% CI | p-Val |
|---|---|---|---|---|---|---|---|
| Allo | cue | JLI-R-0026 | FOF | 312(866) | -0.049516 | [-0.113069, 0.011119] | 0.221140 |
| | | | Sham | 233(647) | -0.015860 | [-0.089044, 0.055656] | 0.761538 |
| | | JLI-R-0032 | FOF | 104(340) | -0.148998 | [-0.257173, -0.041749] | 0.011512(*) |
| | | | Sham | 253(813) | -0.011448 | [-0.078686, 0.058607] | 0.955173 |
| | | JLI-R-0033 | FOF | 143(479) | -0.015701 | [-0.102754, 0.073388] | 0.714417 |
| | | | Sham | 300(1004) | -0.018699 | [-0.078678, 0.041389] | 0.727454 |
| | | JLI-R-0034 | FOF | 234(685) | -0.012718 | [-0.088049, 0.063466] | 0.901838 |
| | | | Sham | 320(989) | -0.032733 | [-0.095131, 0.025790] | 0.233662 |
| | | JLI-R-0036 | FOF | 209(709) | -0.023468 | [-0.098372, 0.055393] | 0.609981 |
| | | | Sham | 235(883) | 0.009029 | [-0.061000, 0.081170] | 0.924860 |
| | delay | JLI-R-0026 | FOF | 301(866) | -0.046011 | [-0.110711, 0.018818] | 0.250633 |
| | | | Sham | 234(647) | -0.071389 | [-0.141296, 0.001482] | 0.061751(.) |
| | | JLI-R-0032 | FOF | 111(340) | -0.019925 | [-0.122683, 0.083049] | 0.883920 |
| | | | Sham | 251(813) | -0.002208 | [-0.072446, 0.067380] | 0.886522 |
| | | JLI-R-0033 | FOF | 169(479) | -0.049413 | [-0.133725, 0.033767] | 0.278868 |
| | | | Sham | 346(1004) | -0.044993 | [-0.103051, 0.014320] | 0.100101 |
| | | JLI-R-0034 | FOF | 248(685) | -0.021971 | [-0.094171, 0.048545] | 0.472752 |
| | | | Sham | 287(989) | 0.019272 | [-0.044661, 0.082145] | 0.528069 |
| | | JLI-R-0036 | FOF | 218(709) | -0.015497 | [-0.091778, 0.056947] | 1.000000 |
| | | | Sham | 260(883) | -0.019940 | [-0.089332, 0.048482] | 0.406609 |
| 2AFC | cue | JLI-R-0026 | FOF | 77(184) | -0.018099 | [-0.122681, 0.084177] | 0.905438 |
| | | | Sham | 70(204) | 0.042915 | [-0.063032, 0.140240] | 0.780706 |
| | | JLI-R-0034 | FOF | 78(242) | -0.044555 | [-0.165318, 0.072558] | 0.460535 |
| | | | Sham | 70(191) | 0.015268 | [-0.099314, 0.126261] | 0.913880 |
| | | JLI-R-0036 | FOF | 62(233) | -0.045457 | [-0.164811, 0.058616] | 0.582136 |
| | | | Sham | 127(313) | 0.023439 | [-0.060807, 0.104975] | 0.784986 |
| | delay | JLI-R-0026 | FOF | 66(184) | -0.247064 | [-0.385157, -0.111442] | 0.000342(***) |
| | | | Sham | 71(204) | -0.032731 | [-0.153193, 0.082593] | 0.763122 |
| | | JLI-R-0034 | FOF | 88(242) | -0.036953 | [-0.148029, 0.070772] | 0.619477 |
| | | | Sham | 73(191) | -0.008731 | [-0.125786, 0.108667] | 0.907986 |
| | | JLI-R-0036 | FOF | 85(233) | -0.113605 | [-0.220949, -0.005856] | 0.050654(.) |
| | | | Sham | 123(313) | -0.009374 | [-0.098751, 0.075917] | 1.000000 |
| Ego | cue | JLI-R-0011 | FOF | 114(613) | -0.007365 | [-0.108978, 0.100506] | 0.893395 |
| | | | Sham | 191(761) | 0.017805 | [-0.064862, 0.093795] | 1.000000 |
| | | JLI-R-0017 | FOF | 109(753) | -0.116380 | [-0.220056, -0.016131] | 0.006164(**) |
| | | | Sham | 184(853) | -0.115612 | [-0.203061, -0.033190] | 0.011894(*) |
| | | JLI-R-0020 | FOF | 269(892) | -0.041586 | [-0.111688, 0.030269] | 0.306069 |
| | | | Sham | 683(1869) | -0.014617 | [-0.060357, 0.030059] | 0.513458 |
| | | JLI-R-0022 | FOF | 139(727) | -0.028962 | [-0.116409, 0.060175] | 0.438151 |
| | | | Sham | 356(1001) | 0.037580 | [-0.018565, 0.090620] | 0.168160 |
| | delay | JLI-R-0011 | FOF | 175(613) | -0.093055 | [-0.173223, -0.011734] | 0.039708(*) |
| | | | Sham | 222(761) | 0.005589 | [-0.070261, 0.079575] | 1.000000 |
| | | JLI-R-0017 | FOF | 178(753) | -0.106289 | [-0.188384, -0.025089] | 0.009753(**) |
| | | | Sham | 205(853) | -0.063690 | [-0.140026, 0.014211] | 0.105367 |
| | | JLI-R-0020 | FOF | 317(892) | -0.107753 | [-0.173502, -0.042469] | 0.005681(**) |
| | | | Sham | 677(1869) | -0.040938 | [-0.085944, 0.004302] | 0.048089(*) |
| | | JLI-R-0022 | FOF | 219(727) | -0.072842 | [-0.145957, -0.001361] | 0.055825(.) |
| | | | Sham | 362(1001) | -0.016802 | [-0.075513, 0.039530] | 0.557584 |

**Supplementary Table 3.** Effect size for $\Delta p$(correct) between laser on/off trials in Allo/Ego task

| task | n_subj | Laser period | Fiber location | n_sess | n_trials(control) | effect size | 95% CI | p-Val |
|---|---|---|---|---|---|---|---|---|
| Allo | 5 | cue | FOF | 29 | 1002(3079) | -0.040009 | [-0.072301 -0.008449] | 0.044528(*) |
| | | | Sham | 40 | 1341(4336) | -0.014955 | [-0.042427 0.014051] | |
| | | delay | FOF | 29 | 1047(3079) | -0.030626 | [-0.065043 0.002573] | 0.213779(n.s.) |
| | | | Sham | 40 | 1378(4336) | -0.020750 | [-0.053316 0.011577] | |
| Ego | 4 | cue | FOF | 26 | 631(2985) | -0.046469 | [-0.091577 -0.001443] | 0.030966(*) |
| | | | Sham | 35 | 1414(4484) | -0.007554 | [-0.039761 0.026461] | |
| | | delay | FOF | 26 | 889(2985) | -0.092011 | [-0.127527 -0.056639] | 0.002010(**) |
| | | | Sham | 35 | 1466(4484) | -0.028008 | [-0.057153 0.001234] | |

**Supplementary Table 4.** Effect size for $\Delta p$(correct) between laser on/off trials before and after task transformation (n=3 rats)

| task | n_subj | Laser period | Fiber location | n_sess | n_trials(control) | effect size | 95% CI | p-Val |
|---|---|---|---|---|---|---|---|---|
| Allo | 3 | cue | FOF | 22 | 755(2260) | -0.029123 | [-0.061580 0.003018] | 0.202307(n.s.) |
| | | | Sham | 24 | 788(2519) | -0.014743 | [-0.049044 0.020055] | |
| | | delay | FOF | 22 | 767(2260) | -0.029118 | [-0.068147 0.008136] | 0.231717(n.s.) |
| | | | Sham | 24 | 781(2519) | -0.017182 | [-0.062346 0.028509] | |
| 2AFC | 3 | cue | FOF | 6 | 217(659) | -0.034595 | [-0.064793 0.000592] | 0.150659(n.s.) |
| | | | Sham | 7 | 267(708) | 0.026149 | [-0.050796 0.110651] | |
| | | delay | FOF | 6 | 239(659) | -0.129932 | [-0.215686 -0.053892] | 0.013355(*) |
| | | | Sham | 7 | 267(708) | -0.015467 | [-0.034758 0.002421] | |

## Code and data sharing

Code and data are available at `https://gitlab.com/sainsbury-wellcome-centre/delab/publications/soundmap-2025`.

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## Contributions

J.L. and J.C.E. designed the behavioral task and conceptualized the study. J.L. designed and setup the systems for optogenetics and electrophysiology recording and developed enhancements to the automated training facility. J.L. implemented and programmed the task and training pipeline. A.X and J.L. trained the SLEAP model for video analysis and J.L. analyzed and quantified the video data. J.L. analyzed the behavioral data. C.B. supervised the development of surgical techniques. J.L. and C.B. performed the surgeries with A.X. and L.L. assisting. J.L., A.X., C.B., L.L. collected the electrophysiology and optogenetics data. J.L. analyzed the data from electrophysiology and optogenetics experiment. A.A. and J.L. implemented the modeling part under the supervision from J.C.E. and C.C. The paper was written by J.L. and J.C.E. with comments from the other authors. C.C. supervised the modeling part of the work and J.C.E. supervised all aspects of the work.

## Methods

### Subjects

A total of 3 male Sprague Dawley rats (Vital River, Beijing, China) and 26 male and female Lister Hooded rats (#603, Charles River, London, UK) were used for this study. Details of each animals role in the study can be found in TABLE S1. Animals were paired housed during the training period and then single housed after electrode or fiber implantation. Of these 29 animals, 9 Lister Hooded rats were used for FOF optogenetics experiment, another 20 rats were used for *in vivo* electrophysiology recording experiment in FOF, mPFC, SC and AC. Animals were kept on controlled-water-access in order to motivate them to work

for water rewards. Body weight was closely monitored daily to ensure their health according to the relevant animal welfare guidelines. All rats were kept on a reversed 12 hour light-dark cycle and were trained during their dark cycle. Animal use procedures for 3 Sprague Dawley rats were approved by New York University Shanghai International Animal Care and Use Committee follows both US and Chinese regulations. For 26 Lister Hooded rats, animal use procedures were approved by the Home Office of the United Kingdom (Project License: PE4FA53CB).

## Behavioral apparatus

Animal training took place in custom behavioral chambers inside boxed to attenuate sound and light. Each chamber ($23 \times 23 \times 23$ cm for training rigs and $23 \times 23 \times 45$ cm for recording rigs) was fit with 8 nose ports arranged in four rows (Fig. 1a), with stereo speakers located on the left and right side. Each nose port contained a pair of blue and a pair of yellow light emitting diodes (LED) for delivering visual cue, as well as two infrared LED/phototransistor pairs for detecting rats' interactions with the port. The port in the bottom row contained a stainless steel tube and a valve for dispensing precisely measured liquid rewards (the reward port). The LEDs, speakers, water valves and the IR pairs were controlled by Bpod), with some design improvements: more I/O to control more LEDs; used a hysteresis comparator array to avoid jittering of IR beam breaks during detection of pokes; and added better ground/isolation layout to prevent ground loops. Additionally, we integrated all of the LEDs and IR pairs into a single PCB (the port wall, as in Fig. 1a) which was powered by 5 V input and connected to the bpod main hardware with a single ribbon cable. Speaker volume was calibrated to 78 dB peak amplitude using uniform white noise. Those designs greatly improved the stability and reliability of the behavioral system. Hardware designs and related software are freely available (bpod-auto).

## Behavioral task and training pipeline

We developed two task variations of auditory memory-guided orienting task[16] for two groups of water-deprived rats. Both task variations followed the same timeline. At the beginning of each trial, one of the operant ports was randomly illuminated as the start port. Rats were required to fixate in the start port for 1.5 s until a go cue, constituting the fixation period. During the first 0.75 s of fixation, one of two auditory cues was presented: 30 Hz or 68 Hz click trains. Each click was an 8 KHz pure tone lasting 0.003 s. This was followed by a 0.75 s silent delay period. A go cue, 0.2 s 15 KHz pure tone, was played at the end of the delay. After the go cue, rats withdrew from the start port and moved to a target port. A correct response resulted in the illumination and water reward availability at the reward port. Incorrect choices triggered an error sound (1 s white noise), and a new trial became available after a short inter-trial interval (2 s to 10 s; Fig. 1a). Early withdrawal from the start port before the go cue, required animals to restart the fixation from the beginning. These trials (about 20%) were excluded from analyses except when calculating early withdrawal rates (Fig. S7).

The mapping from sound cue to target port depended on the task context and start port:

- **Egocentric (Ego) Task:** The correct target port was either the left or right port adjacent to the start port, determined by the sound cue (blue/red for 30 Hz/68 Hz clicks, respectively; Fig. 1b, left). This resulted in eight trial types spanning two movement directions (Fig. S1a, left).

- **Allocentric (Allo) Task:** The correct target port was fixed in physical space, with the bottom-left/bottom-right port corresponding to the blue/red sound cues, respectively (FIG. 1b, right). This allocentric spatial coding yielded 12 trial types (12 unique movement vectors) from the combination of different start ports and target ports(FIG. S1a, right).

We classified errors in the task in two ways. The first, in-task errors, were errors where the subject's choice port was associated with the unplayed sound. The second, out-of-task errors, were errors where the subject's choice port was not associated with either sound. These later choices were rare and were excluded from all analyses except FIG. S1B.

## Training pipeline

Rats underwent a structured training pipeline towards achieving mastery of the final task. Training comprised three phases:

1. **Operant Conditioning Phase:** In this phase, rats became familiar with the training apparatus and learned to associate illuminated ports with water rewards. Training progressed through distinct stages:

   - **Stage 1:** The reward port was illuminated, and rats received water upon entry. Advancement to the next stage required six consecutive correct trials.
   - **Stage 2:** All operant ports were illuminated, and poking in any lit port activated the reward port. The number of illuminated ports was gradually reduced from eight to one. Progression required six consecutive successful trials without errors (e.g., pokes in unlit ports) or timeouts (no poke within 60 s of trial initiation).

2. **Auditory Cue and Task Rule Phase:** In this phase, rats learned the association between auditory cues and target ports. Fixation was not required; rats could initiate a trial by poking the start port, after which the auditory cue was played until choice. Initial training used blocked trial types, with block switching probabilities increasing after each correct trial (starting at 0.02).

3. **Fixation and Timing Phase:** This phase introduced fixation and timing requirements. In this phase, the sounds were played for a maximum of 1.1 s. Initial fixation times were set to 200 ms and adaptively increased by 20 ms increments based on task performance until reaching 1500 ms. As fixation grew beyond 1.1 s, the task required some short-term memory. Once fixation stabilized at 1.5 s, auditory cue durations were reduced from 1.1 s to 0.75 s using a similar adaptive approach. The progress of animals through this stage is shown in FIG. S2.

Rats were trained daily for 80 minutes. The complete training process typically required 2 to 4 months (FIG. S2).

For 11 animals, after reaching asymptotic performance on the Allo task and collecting either electrophysiological or optogenetics data, we transformed the task to a 'reduced 2AFC' task, where all trials started from the center-port in order to perform within-subject tests of how reducing the working memory load for a motor planning would change animals' strategy (FIG. 7 & FIG. S9).

## Surgery

The rats were anesthetized with isoflurane and placed in a stereotaxic apparatus (David Kopf Instruments). The scalp was shaved, washed with ethanol and iodopovidone and

incised. Then, the skull was cleaned of tissue and blood. The stereotax was used to mark the locations of craniotomies for the FOF+mPFC (+2.5 AP, ±1.4 ML mm from Bregma); AC (−4.3 mm AP, ±5.5 ML); mPFC only (3.2 mm AP, ±1.0 mm ML) or SC (-6.4 mm AP, ±1.3 mm ML), relative to Bregma on the skull. Then a 1.5 mm craniotomy was drilled, followed by an entire dura resection. The craniotomy was then filled with saline saturated Gelfoam to protect the brain tissue while the skull was coated with a thin layer of C&B Metabond (Parkell, Inc; New York) and a 1-3 mm high moat built around the craniotomy using the Absolute Dentin (Parkell, Inc; New York).

For electrophysiology surgery, silicon probes (Cambridge NeuroTech/Neuropixels) were implanted into the target region. The silicon probes were adhered to nano-drives (Cambridge NeuroTech, or R2Drive[79]) with super-glue before implant. Ground wires were soldered to titanium ground screws located above primary visual cortex. The silicon probe was slowly lowered into the brain until the desired depth (FOF recording: 1.3 mm DV for the H3 probes, and 0.5 mm DV for the E probes and F probes; FOF+mPFC recording: 3.6 mm DV, Neuropixel 1.0 probes with 10 degree for ML; AC recording: 4.8 mm DV, Neuropixel 2.0 probes with 10-12 Degree in ML; mPFC recording: 3 mm DV for H3 probe; SC recording: 5.5 mm DV, Neuropixel 1.0 probes).

For optogenetics surgery, we slowly injected 400 uL AAV virus solution (pAAV-CKIIa-stGtACR2-FusionRed, AddGene #105669, about $5 \times 10^{12}$) into FOF bilateraly. The injection system comprised of a custom-made glass pipette (20–30 m O.D. at the tip; Drummond Scientific, Wiretrol II Capillary Microdispenser) back-filled with mineral oil, and a fitted plunger inserted into the pipette to load or dispense viral solution. The plunger was controlled by a hydraulic manipulator (Narashige, MO-10); and the injection pipette was advanced into the brain using stereotax. After each injection, the glass pipette was left in the brain for 5 minutes before it was slowly withdrawn, to prevent backflow. Tapered fibers (`MFC_200/245−0.37_4.5mm_ZF2.5(G)_TP1.5`, Doric Lenses Inc.) were inserted 1.2 mm into the cortex measured from the brain surface for each craniotomy for bilateral FOF.

The craniotomy was filled with Dura-Gel (Cambridge NeuroTech), and then silicon probes or the optic fiber was cemented to the skull with Absolute Dentin. Vetbond (3M, U.S.) was applied to glue the surrounding tissue to Absolute Dentin. The animals were given 5 days to recover on free water before resuming training.

## Electrophysiology recording and spike sorting

For Cambridge Neurotech probes, neural activity signal was digitized at 30 kHz, amplified and bandpass filtered at 0.6-7500Hz using a 64 channel Intan headstage (RHD2164, Intan Technologies), the SPI cable of the Intan headstage was tethered to a commutator to allow free spinning (Shenzhen Moflon Technology, MMC250), and all the raw data was processed using the Open Ephys acquisition board connected to a computer to visualize and store the neural signals. The probes were turned down $\approx 100\,\mu m$ for every 3-5 days until the white matter was reached. For Neuropixel probes, we used acquisition hardware from NI (a PXIe-1071 Chassis) with a Neuropixel recording module (PXIE_1000, IMEC). Data was monitored and recorded with SpikeGLX software.

During the recording, at the end of each trial a serial TTL message encoding the current trial number was sent from our behavioral control hardware to the acquisition system to permit synchronization of the neural signal with the behavior data.

Offline spike sorting was performed by Kilosort v2[80] for Cambridge Neurotech probe recording or Kilosort v4[81] for Neuropixel probe recording with default settings. Spike clusters were manually curated using Phy. The quality metrics and waveform metrics for sorted units were computed using ecephys spike sorting[82]. Specifically, we selected units

with averaged firing rate >1 Hz, signal-to-noise ratio >1.5, and a presence ratio >0.95 over the course of recording sessions.

## Optogenetics experiment

The implanted fibers were connected to one (for the sham fiber) or two (for bilateral silencing of FOF) 0.6 m optic fiber cables (`OPT/PC-FC-FCF-200/230-HD-0.6L-MC`, Plexon Inc.). The other end of cable was connected to a dual fiber rotary joint (`FRJ_1x2i_FC-2FC_0.50`, Doric Lenses Inc.) mounted in the ceiling of the sound attenuation chamber. This was connected to a 473 nm laser (473nm LX 50 mW LASER SYSTEM, OBIS Inc.) with a variable optic attenuator (VOAMMF, Thorlabs Inc.) and an optic patch cable (M123L01, Thorlabs Inc,). The laser power was controlled by Pulse Pal (1102, Sanworks LLC) which was triggered by Bpod with 5 V TTL. The laser power was calibrated by a laser power meter (PM20A, Thorlabs Inc.) before and after every session to be 3mW at fiber end.

## Videography

We used USB web camera (FIT0730, DFROBOT) to record video from top-down view during behavior session. Video were acquired at $640 \times 480$ pixels and 20 frames per second. Rat performed the task in complete darkness, and videos were recorded under infrared 940 nm LED illumination. We developed a custom software controlled the video acquisition for each session (zmq-video-server).

## Histology

We imaged the brains using serial section[83] two-photon[84] microscopy. Our microscope was controlled by ScanImage Basic (MBF Bioscience) using BakingTray, a custom software wrapper for setting up the imaging parameters. Images were assembled using StitchIt.

## Analyses and statistics

In the text, when we use $\pm$ (e.g. $4.2 \pm 0.1$), we are reporting the mean and standard error of the mean, unless otherwise specified. In any case where a parametric statistic resulted in a $p$ value $< 10^{-9}$, we report it as $< 10^{-9}$. Details of all statistical tests are included in the Statistical Appendix.

For mixed-effects models we used either `fitlme` (for linear models) or `fitglme` (for Poisson or logistic models) in MATLAB (2024b; The Mathworks). In some cases, we used the `lme4` library (version 1.1-29)[85] in R.

## Behavioral data analysis

Out-of-task errors were rare and were excluded from all analyses except Fig. S1b. We excluded all timeout trials (no choice made within 2 s from go cue) and fixation violation trials (failure to maintain fixation for 1.5 s in the start port, about 20% of trials). Additionally, we excluded trials where animals accidentally used their paw to break an IR beam in a choice port (port entry was detected within 0.2 s after the go cue, 3.71% of trials removed). The hit rate, $p$(correct), was calculated as the fraction of trials in which the animal reached the correct choice port, relative to all remaining trials after these exclusions (Fig. S1g). Movement time (MT) was defined as the interval between withdrawal from the start port and entry into the choice port. For this analysis, we included only center-start trials. To compare MT differences between the Allo and Ego tasks (Fig. 1e and Fig. S1c), we first

computed the median MT for each animal. We then used the Wilcoxon rank-sum test (`wilcox.test` in R) to assess the statistical significance of differences between the Allo and Ego animal groups. The $p$ values or that test are reported in the respective figure panels.

To statistically assess the MT change after task transformation (FIG. 7c), we performed day-by-day likelihood-ratio tests using log-transformed MT values. For each post-transformation day, $0 \leq d \leq 7$, we compared baseline trials, `pre` (all valid trials from the 7 days before the task transformation) to trials from day $d$, `post` using mixed-effects linear models of the form:

$$\log_{10}(\texttt{MT}) \sim \texttt{1 + pre\_post + (1|subject)} \tag{1}$$

Where `pre_post` indicated whether a trial belonged to baseline or to after task transformation. This full model was compared to a reduced model without the `pre_post` term using a likelihood-ratio test. The resulting $p$-values determined which post-transformation days differed significantly from baseline. The same model was used to compare pooled MT (from all tethered experimental sessions) before and after task transformation (FIG. S4G).

In optogenetics data analysis, we also defined reaction time (RT) as the interval between the end of the delay period (indicated by the go cue) and the withdrawal from the start port and entry into the choice port (FIG. S7a,b).

## Video data analysis

We used SLEAP[86] to track body key points (nose, left ear, right ear, neck, FIG. 1f) from video frames recorded for each session. Separate models were used to track animals that have different implant type (optic fiber, or recording probe). We manually labeled ≈3000 randomly picked frames from different sessions to train each model. Training was performed using the default setting parameters of SLEAP. Then the trained model was used to predict the position of each key point for all frames of all sessions. We then calculated vector angles between the pair of each point using the x,y coordinate of each points.

For each trial, the vector data (3-dims: left-ear, right-ear, neck to nose angle, FIG. 1f) was aligned to the trial onset (using **cdraster** function in Erlich lab utilities[87], FIG. 1g), or choice port (target) arrival. At each timepoint for each individual rat, we used logistic regression with LASSO regularization and 10-folds cross-validation that took the vector angles as input to predict the animals' choice (using **fitclinear** function in MATLAB). We concatenate all data from all sessions for each individual rats. Only center-start trials where the errors were 'in task' were included for the video posture analysis to control for the same up-coming movement after the go-cue. We averaged the choice decoding accuracy of all rats at each time-point for each task. This result was shown in FIG. 1h and FIG. S1d.

## Electrophysiology data analysis

**Characterizing single-neuron selectivity.** For the rasters and peri-stimulus time histogram (PSTH) plots (FIG. 2), spike times were aligned to the sound cue onset within a 2.5 s time window ($-0.5$ s before cue onset to 0.5 s after fixation end). The bin size was set to a resolution of 0.01 s and smoothed with a causal half-Gaussian kernel with a standard deviation of 0.2 s.

To evaluate whether the activity of each cell during the delay period (0.75 s to 1.5 s after cue onset) could predict upcoming choice, `spike_count` was calculated for each trial during this period. To quantify choice tuning of each cell, we ran a mixed-effects Poisson

model with the following formula.

$$\texttt{spike\_count} \sim 1 + \texttt{choice} + (1|\texttt{start\_port}) \qquad (2)$$

where `choice` was -1 or 1 for ipsilateral or contralateral choice (relative to the recorded hemisphere) and `start_port` (-2 to 2) denoted the X-axis location of the start ports relative to the center port. The choice was defined by contra/ipsi target direction for the Ego task or contra/ipsi target port for the Allo task. A $p < .01$ threshold for the `choice` coefficient identified choice selective cells.

A similar mixed-effects linear regression model was implemented to evaluate the contribution of start port position to firing rates in the FOF cells, independent of choice. The formula was:

$$\texttt{spike\_count} \sim 1 + \texttt{start\_port} + (1|\texttt{choice}) \qquad (3)$$

so that we could quantify the contribution of `start_port` independent of `choice`. A $p < .01$ threshold for the `start_port` coefficient identified start-port-selective cells.

To assess choice selectivity over the trial's time course for center-start trials (Figure 3a), we applied a linear regression model:

$$\texttt{firing\_rate} \sim 1 + \texttt{choice} \qquad (4)$$

where `firing_rate` was estimated with a $0.25\,\mathrm{s}$ sliding window ($0.025\,\mathrm{s}$ step size) from spike times aligned to cue onset. A $p < 0.01$ threshold for the `choice_index` coefficient was used to identify choice-selective cells at each time point. The percentage of such choice-selective cells was then calculated across time points.

To assess start-port selectivity over the trial's time course, we used the same model as Eq. 3 using a $0.25\,\mathrm{s}$ window and $0.025\,\mathrm{s}$ step size aligned to cue onset. Cells with $p < 0.01$ for `start_port` were identified as start-port selective, and the percentage of such cells was calculated over time points.

For units recorded from the auditory cortex (FIG. 5), we use the same model as in Eq. 2. However, since this is auditory cortex we refer to it as cue selectivity, choice and cue selectivity are indistinguishable when restricted to correct trials. We only included correct trials in cue response analysis except FIG. 5c. A $p < 0.05$ threshold for the `cue_index` coefficient identified cue selective cells as in FIG. 5b,c.

In order to quantify the confidence interval of the slope between the cue selectivity ($\beta_{cue}$) of hit and error trials (FIG. 5c), we resampled the data for TLS regression 10,000 times to approximate a distribution of the slope and reported the mean and 99% CI.

**Pseudopopulation decoding.** To compare choice decoding performance, we generated 100 pseudosessions for each task: both for Ego vs. Allo comparison and also for Allo vs. 2AFC. For each pseudosession, we randomly sampled 100 cells without replacement. For each selected cell, we selected 10 left-choice and 10 right-choice center-start trials. We included both correct trials and in-task error trials for this analysis. Sessions with fewer than 10 left and 10 right choice trials were excluded. For each selected cell, we estimated the firing rate across each trial with a $0.25\,\mathrm{s}$ causal sliding window with a $0.025\,\mathrm{s}$ step size, aligned to cue onset. This process yielded a $100 \times 20 \times 100$ matrix (cells $\times$ trials $\times$ timepoints), $D_{\mathrm{original}}$, for each pseudosession.

Principal component analysis (PCA; using `pca` in MATLAB, which automatically centers data) was then applied to $D_{\mathrm{original}}$, with distinct approaches depending on the analysis type: comparing decoding across tasks (FIG. 3b, FIG. 7g) or cross-time decoding (FIG. 3d, FIG. 7f). For comparing decoding across tasks, PCA was performed independently for each

timepoint (each timepoint giving a $100 \times 20$ (cells × trials) matrix), and the top five principal components (PCs) were used. Projecting the data onto the top 5 PCs resulted in a matrix $D_{\text{independent}}$ of dimensions $5 \times 20 \times 100$ (PC dims × trials × timepoints) for decoding analysis. For cross-time decoding, $D_{\text{original}}$ was reshaped, creating a 2D matrix of dimension $100 \times 2000$ (cells × (timepoints · trials)), denoted as $D_{\text{concatenated}}$. PCA was applied to $D_{\text{concatenated}}$, we used top five principal components and reversing the trial concatenation, to create a $5 \times 20 \times 100$ (PC dims × trials × timepoints) matrix, $D_{\text{shared}}$, for decoding analysis. Decoding was performed using multivariate linear classification (`fitclinear` in MATLAB with L1 regularization) with leave-one-trial-out cross-validation over the 20 trials selected in each pseudosession.

To generate a $p$-value at each time for the comparison of decoding (FIG. 3b, FIG. 7g), we generated a 100 timepoints vector, $\vec{A}_{\text{data}}$, by averaging the decoding accuracy over pseudosessions. We then generated a shuffled decoding distributions for each of the 100 pseudosessions by permuting the label of left/right for the 20 trials 5000 times and doing the decoding analysis for each shuffle, producing a 100 pseudosessions × 5000 shuffles for each time point and averaging over pseudosessions to get a $5000 \times 100$ (pseudosessions × timepoint), $A_{\text{shuffle}}$. We considered a timepoint in $A_{\text{data}}$ to be significantly above chance if it was higher than 0.9995 (p=0.05 / 100 using Bonferroni correction) of the corresponding shuffle distribution for that timepoint. To test if two tasks were different from each other we did a permutation test between the $\vec{A}_{\text{data}}$ with 10,000 shuffles of the task label. For visualizing the timepoints where the tasks has significantly different, we required at least 5 consecutive timepoints to be significant in order to be included. For comparison between the Allo vs. 2AFC task, we required at least 8 consecutive time points to be significant (to account for a greater degree of noisiness in the smaller data set).

For cross-time decoding analyses (FIG. 3d, FIG. 7f), we trained a classifier for each pseudosession at every timepoint in $D_{\text{shared}}$ using the label vector $L$. The classifier predicted the labels across all timepoints of $D_{\text{shared}}$ with leave-one-out cross-validation, yielding a $100 \times 100$ (timepoint × timepoint) matrix of decoding accuracy that captured the stability of choice coding[88]. To assess statistical significance, we generated 1,000 null matrices by shuffling the label vector $L$ for each pseudosession. An extreme pixel-based permutation test[89] was applied to compute $p$-values for each matrix element and a contour was plotted to indicate significant regions. Significant regions smaller than $7 \times 7$ timepoints were excluded.

To compare start position decoding we used the same methods and code as for choice decoding with the following key differences. Instead of using only center-start trials we sampled 10 left and 10 right choice trials (from correct trials and in-task error trials) for each of the top-left, top-right, and center-start trials without replacement, yielding 60 trials for each pseudosession. We used only these three start positions because they are matched across the two tasks (FIG. S1A). For each trial, we coded the X-axis position of each port (-1 for top-left port, 0 for center port and 1 for top-right port). Decoding was performed using multivariate linear regression models (`fitrlinear` in MATLAB) with 10-fold cross-validation. The decoding performance was quantified as the $R^2$ between the true start position and the cross-validated model prediction for each trial.

**Single-trial decoding of choice.** To perform single-trial decoding of choice for each session (FIG. 3c; FIG. S4E-H), we selected center-start trials that were correct or were in-task errors to generate an $N \times I \times T$ data matrix (neurons × trials × timepoints), $D_{\text{sess}}$. We also had a corresponding length $I$ vector for each session: $\vec{L}$, which was set to 1 if the animal chose right and set to 0 for chose-left on each trial. PCA was performed independently at each timepoint to perform PCA whitening. This resulted in a matrix

$Z_{\mathrm{session},t}$ of dimensions $P \times I$ (post PCA dims $\times$ trials) for each timepoint, $t$. Then, we performed binary classification of $\vec{L}$ using $Z_{\mathrm{session},t}$ (`fitclinear` in MATLAB with L1 regularization) with leave-one-out cross-validation. This resulted in a length T decoding accuracy vector. We performed this for each session. For Ego task FOF recording sessions (n=59), we had on average $184 \pm 6$ completed trials per session, with $\approx 29.47\%$ center-start trials per session. In Allo task FOF recording sessions (n=65), we had on average $251 \pm 12$ completed trials per session, with $\approx 20\%$ center-start trials per session.

To compute the latency of the rise in choice decoding performance, we evaluated the time, for each session, at which the decoding accuracy reached 0.7 for at least 7 consecutive timepoints, based on a previously reported method[48,90]. Latency comparison plot was generated with DataViz (FIG. 3C).

**Comparison of single-trial decoding of choice across regions**  For the comparison of decoding accuracy of choice across regions (FIG. 5F,G,H), we only included correct trials to simplify comparison of the auditory cortex with other regions where error trials may be in different reference frames. That is, errors in AC are generally in a sensory reference frame, and errors the in frontal cortex are in a choice reference frame. On correct trials, decoding the choice is equivalent to decoding the sensory cue. We also excluded trials that started from the bottom-left or bottom-right ports.

In contrast to the causal sliding window used elsewhere (e.g. FIG. 3), here we used a 250 ms non-causal sliding window (25 ms step) to make it easier to estimate the decay of cue-coding during the delay. We generated an $N \times I \times T$ (neurons $\times$ trials $\times$ timepoints) matrix, $D_{\mathrm{sess}}$, together with the length I trial label vector $\vec{L}$ (0 for sound A, 1 for sound B). PCA whitening was applied to at each timepoint, giving $Z_{\mathrm{sess}}$ of size $N \times I \times T$. We then added the start-port as a dummy variable to $Z_{\mathrm{sess}}$ to allow the decoder to find start-port dependent coding of choice. Decoding was performed at each time point using logistic classification (`fitclinear` in MATLAB with L1 regularization) with 10-fold cross-validation, resulting in decoding accuracy curves for each session.

To compare regions, we averaged decoding accuracy during the delay period (0.75–1.5 s), ranked sessions by accuracy within each region, and selected the top half for group analysis. Trial labels were shuffled 10,000 times for each session, and the same ranking and selection were repeated for each shuffled set. To balance different numbers of sessions across regions, we randomly resampled seven of the top-half sessions 200 times and compared their average accuracy against shuffled controls. We picked 7 because we only had 15 recording sessions from auditory cortex, thus the top half gave us 7 sessions. A timepoint was considered significant (FIG. 5F, purple and red bars) if more than 50% of resampled sets gave $p < .01$; significance periods shorter than 0.075 s were excluded (3 timepoints).

To test whether AC decoding exceeded that of other regions (FIG. 5F,G ) we compared session-wise decoding accuracy between AC and each non-AC region (FOF, SC, or mPFC) using a Wilcoxon rank-sum test. For FIG. 5F (grey bar), we show the comparison between AC and FOF, applied FDR correction at $p < .05$, and significance periods shorter than 75 ms were removed.

For single-session cross-time decoding (FIG. 5H), we only included the top half of sessions for each region (described above). We generated a trial-concatenated matrix (neurons $\times$ [timepoints $\cdot$ trials]), fit a single PCA, for whitening, for each session on this concatenated matrix, and kept all components. The data were then reshaped back into a representation of $P \times$ trials $\times$ timepoints, so that the same PCA basis was used for all timepoints within a session. Decoding was performed with logistic classifiers (`fitclinear` in MATLAB, 10-fold cross-validation with L1 regularization), training at each timepoint and testing at every

timepoint, which produced a cross-time decoding heatmap for each session. For each region, we randomly resampled 7 sessions and averaged their heatmaps, repeating this 200 times to obtain 200 resampled heatmaps. Each resample was compared against 1,000 trial-label–shuffled dummy heatmaps processed with the same selection method using the extreme pixel-based permutation test[89], yielding a pixelwise $p$-value map. A pixel was considered significant if more than 50% of resamples reached $p < .01$ at that pixel. Small significant clusters ($< 5 \times 5$ timepoints) were removed.

To compare decoding between hit and error trials (FIG. 5D), we constructed two data matrices of dimensions $N \times I$ (neurons $\times$ trials) for the spike count from the cue period (0–0.75 s) and the delay period (0.85–1.5 s; the 0.1 s gap was to reduce contamination from the sound encoding during the cue period). For each session, decoding was performed separately for cue and delay periods and for hit and error trials, using logistic classification classifier (`fitclinear` in MATLAB, 10-fold cross-validation with L1 regularization). For cue/delay period, decoding accuracies from hit and error trials were paired across sessions, and differences were assessed using the Wilcoxon signed-rank test.

**Single-trial decoding of time.** To decode the time in trial during the fixation period (FIG. 6H,I). For each session, we estimated the firing rate with a 400 ms sliding window (100 ms step size) from spike times aligned to cue onset, this gave an $N \times J$ matrix, $D_{\text{time}}$, for each session, where $N$ is the number of neurons and $J$ is the number of samples for time decoding. With our sliding window over 1.5 s fixation period this gave 16 windows per trial, so $J$ was $16 \cdot I$, where $I$ is number of trials in a session. PCA was then performed on $D_{\text{time}}$ for whitening, all PCs were kept. We also had a corresponding length 16 time label vector, $\vec{t}$, which was the time relative to the trial onset. Then, we used multivariate linear regression (`fitrlinear` in MATLAB) to predict $\vec{t}$ with 10-fold cross-validation, resulting in $\hat{t}$. We then calculated $R^2$ between $\vec{t}$ and $\hat{t}$ for each session to quantify time decoding performance.

**Coding direction analysis** To explore how choice coding could generalize across start-ports we found the 'choice-coding direction' in neural population space from center-start trials (FIG. 3i-l)[14].

To find the coding direction for each session, we generated an $N \times I$ (neurons $\times$ trials) matrix, $F$, of the average firing rate of each cell, on center-start trials, from 0.75 to 1.5 s relative to cue onset. We then train a logistic classifier to predict choice from $F$, using the default $L1$ regularization (`fitclinear` in MATLAB). This resulted in a weight vector, $CD$, the coding direction for center-start trials. We repeated this procedure using 10-fold cross-validation to quantify the projection of each center-start trial onto the $CD$. We used all center-start trials to find a $CD$ for out-of-sample prediction of choice from other start positions.

To project each trial onto the CD, we estimated the firing rate using a 400 ms sliding window (10 ms step size) to get a length $N$ vector at each timepoint, $G_{i,t}$ representing the population activity on trial $i$ at time $t$. We then project the activity onto the $CD$. Projecting the activity for all trials and all times, $G$, gives a $I \times T$ (trials $\times$ timepoints) matrix of the projected activity $Q = CD^T G$. We then remove the grand mean of $Q$ to center the projection to allow combining data across sessions.

**Decoding generalization analysis** To statistically quantify the generalizability of choice coding across different start positions, we trained a multivariate linear classification model (`fitclinear` in MATLAB with L1 regularization) for center-start trials for each session

to predict left/right choice label vector $L$ from spike counts matrix $D_{raw}$ during the delay period. Here $D_{raw}$ is a $N \times T$ matrix for N cells and T trials in that session, $L$ is the choice label vector which was 0 for left choice and 1 for right choice.

We firstly predicted the choice label vector $\hat{L}$ for all center-start trials with 10-fold cross-validation and quantified the decoding accuracy for each session. For each session, we also predicted the choice label vector $\hat{L}$ for other trial types started from other ports, using the classification model trained with all center-started trials of that session as an out-of-sample prediction accuracy.

To quantify the confidence interval of the slope between the decoding accuracy of center-started trials and other trials (Fig. 3m,n), we resampled the data for TLS regression 10,000 times to approximate a distribution of the slope and reported the mean and 95% CI.

**Optogenetics data analysis**

For optogenetic analyses, we defined control trials as the no-laser trials from the same sessions as a corresponding laser trial. Sham sessions where the sessions when optic fiber was plugged into a fiber which did not enter the brain.

For Fig. 4e,f, the error bars are just for visualization, since all the $p$-values are based on mixed-effects models (described below). To generate the error bars, we sampled (with replacement) the laser-on and laser-off trials for each session, took the difference in performance ($laser_{ON} - laser_{OFF} = \Delta p(correct)$), and took the mean across sessions. We repeated this process 10,000 times, and show the 95% CI of this distribution as the error bars.

To disentangle the effects of the laser light from its optogenetic effect, we used logistic mixed-effect models (`lme4`, version 1.1-29)[85] in R (4.4.3) and plotted the results using `ggplot2` (version 3.5.1)[91]. In general, we estimated significance by performing likelihood ratio tests (`lrtest` in the `lmtest` package, version 0.9.40)[92] between each model with a reduced model that removed the variable of interest.

To test whether bilateral FOF silencing for during different task epochs had an effect on performance, we specified a mixed-effects model where `hit` was a logistic function of laser on/off (`isopto`) , fiber location (bilateral FOF or sham, `isFOF`) and their interaction as fixed effects. Session level variability were modeled as random effect in this GLMM (using Wilkonsin notation)[93]:

$$\text{hit} \sim \text{isopto} + \text{isFOF} + \text{isopto:isFOF} + (1|\text{session}) \tag{5}$$

We compared that model with a reduced model

$$\text{hit} \sim \text{isopto} + \text{isFOF} + (1|\text{session}) \tag{6}$$

using a likelihood ratio test and reported the $p$-value of that test. By dropping the `isopto:isFOF` we are effectively testing whether knowing if the fiber was connected to the sham or FOF fibers significantly improves the model fit. the This was done 4 times: for (Ego / Allo) $\times$ (cue / delay). The $p$-values are reported in the relevant panels of Fig. 4e,f.

To directly compare whether the optogenetic effect was significantly different in the two tasks, we used the following model:

$$\text{hit} \sim \text{task} + \text{isopto} + \text{isopto:isFOF} + \text{isopto:isFOF:task} + (1|\text{subject/session}) \tag{7}$$

where `task` specified whether a trial was from the Ego or Allo task. We compared this model to a reduced model

$$\text{hit} \sim \text{task} + \text{isopto} + \text{isopto:isFOF} + (1|\text{subject}/\text{session}) \tag{8}$$

and performed a likelihood ratio test. We used this model to compare the tasks in the cue and delay period separately. By dropping the `isopto:isFOF:task` interaction term we are effectively testing whether allowing the model to have a differential effect of FOF silencing relative to sham in each task significantly improves the model fit.

Analyses of the optogenetic effects after task transformation were the same, but rather than comparing Ego and Allo tasks, we were comparing within-subject effects between Allo and the reduced 2AFC task. Note, some small differences between the Allo results in FIG. 4D,F and FIG. 7D,E are due to two animals being included in the comparison with Ego, but not experiencing the task transformation (because of lost implants).

**Synthetic data for optogenetics experiments**  The GLMM model specification for task comparison (Eqs. 7,8) were picked from a set of potential models, based on their ability to discover task differences in synthetic data. We generated synthetic data where there was an effect of silencing FOF in one task but not the other, and included variability across subjects and across sessions. We also matched the trials number and type with our real data (FIG. S6a).

We included, for a sanity check, models (e.g. GLMM0,1) that we know should not fit the data well. The other models included different degrees of complexity (e.g. in the number of parameters) All models shared a similar design that estimated whether a trial would be correct or an error (`hit`) as logistic function of `task`, laser on/off (*isopto*), fiber location (*isFOF*, bilateral FOF or sham). Different interactions among those three terms were implemented to determine the best way to model those interactions. The variability for each sessions, nested into subjects, were modeled as a random effect for all models.

```
GLMM0: hit ~ task + isopto:isFOF + (1|subject/session)
GLMM1: hit ~ task + isopto + isFOF + (1|subject/session)
GLMM2: hit ~  isFOF + isopto + isopto:task + (1|subject/session)
GLMM3: hit ~  task + isopto + isopto:isFOF + (1|subject/session)
GLMM4: hit ~  task:isopto:isFOF + (1|subject/session)
GLMM5: hit ~  task + isopto + isopto:isFOF + isopto:task + (1|subject/session)
GLMM6: hit ~  task*isopto*isFOF + (1|subject/session)
GLMM7: hit ~  task + isopto + isopto:isFOF + isopto:isFOF:task + (1|subject/session)
```

The best model, `GLMM7`, was picked based on the lowest BIC and AIC quantification (FIG. S6b).

## Dynamical model

We developed a computational model to formalize our hypothesis about how different forms of working memory may be recruited under varying task demands. The model has three parts: a selective excitation module and two decision modules. The first decision module is a bistable attractor modeling the FOF as egocentric working memory, FOF. The second module, is a fixed performance modeling the auditory cortex, AC, which supports performance in a sound reference frame. FOF and AC exhibit persistent selective activity in the presence of external excitation, enabling them hold the relevant information over

a delay period. In the absence of external excitation the activity decays to zero. We instantiated the decision modules as dynamicals system using Euler's method (described below).

The excitation to each decision module was based on net cost.

$$C_{NET}(w_{FOF}, task) = (1 - w_{FOF})C_{task}^{AC} + w_{FOF}C_{task}^{FOF} \tag{9}$$

$$C_{task}^{AC} = B^{AC}N_{task}^{AC} \tag{10}$$

$$C_{task}^{FOF} = B^{FOF}N_{task}^{FOF} \tag{11}$$

where $C_{task}^{Region}$ represents the cost for using a region in a task for short-term memory. $0 \leq w_{FOF} \leq 1$, sets the relative excitation to FOF and, $w_{AC} = 1 - w_{FOF}$ denotes the excitation that region AC received. Note that for the Allo task, an egocentric representation would require the FOF module to store a representation that supports at least 12 different movement vectors ($\log_2(12) \approx 3.6$, FIG. S1a, Right), so we set $N_{ALLO}^{FOF} = 3.6$. Since the auditory cue was binary in both tasks, we set $N^{AC} = 1$ for both tasks.

$B^{region}$ represents the cost-per-bits for each region. We set $B^{FOF} = 1$ and $B^{AC} = 3$ to implement the idea that FOF dependent movement planning has higher biological preparedness than a sensory working memory strategy. This gives $C_{Ego}^{FOF} = 1$ and $C_{Allo}^{FOF} = 3.6$. For both tasks, $C^{AC} = 3$.

For optimization, we defined the loss as:

$$L(w_{FOF}) = C_{NET}(w_{FOF}, task)^2 \tag{12}$$

and the trajectory of $w_{FOF}$ takes the form

$$w_{FOF}^{(t+1)} = w_{FOF}^{(t)} - \alpha\frac{\partial}{\partial w_{FOF}}L(w_{FOF}) + \mathcal{N}(0, \sigma_w) \tag{13}$$

where $\alpha$ is the learning rate and $\sigma_w$ is a parameter controlling noise amplitude. $w_{FOF}$ values are clipped between 0 and 1 after each training iteration. Here, $t$ represents each training iteration step, and we considered there to be 10 trials per step in the simulation(FIG. 7b), reflecting a slow metacognitive process of assessing the cost of a specific WM format. For completeness, we include the expressions for $\frac{\partial}{\partial w_{FOF}}L(w_{FOF})$:

$$\frac{\partial}{\partial w_{FOF}}L(w_{FOF}) = -2\left(C^{AC} - w_{FOF}(C^{AC} - C^{FOF})\right)\left(C^{AC} - C^{FOF}\right) \tag{14}$$

**Network model of FOF** We modeled FOF/M2 as a binary attractor network following previous work[66,94]. We use a Wilson-Cowan model of cortical interactions[95,96] with two competing, mutually inhibiting sub-populations, each coding for a different behavioural decision:

$$\tau\frac{dr_L}{dt} = -r_L + F(w_{FOF} \cdot E_{EXT}(t) + I_L + w_{rec}r_L - w_{inh}r_R + I_{opto}) + \mathcal{N}(0, \tau\sigma_r^2) \tag{15}$$

$$\tau\frac{dr_R}{dt} = -r_R + F(w_{FOF} \cdot E_{EXT}(t) + I_R + w_{rec}r_R - w_{inh}r_L + I_{opto}) + \mathcal{N}(0, \tau\sigma_r^2) \tag{16}$$

where $\tau$ is the population rate time constant, $r_i$ is the firing rate of population $i$, driving choice to the right, $r_R$, or left, $r_L$. $I_i$ represents the sound input: $I_i = 1$, if (trial-type=$i$) $\wedge$ ($0 < t < 0.75$ s) else 0. $w_{rec}$ is the strength of recurrent connectivity within each region, $w_{inh}$ is the strength with which populations $L$ and $R$ inhibit each other, and $\sigma_r$ is a parameter controlling noise amplitude. $I_{opto}$ is the strength of optogenetic inhibition. $F(x)$ is the

activation function, defined as a generalized sigmoid: $F(x) = \frac{1}{1+\exp{(x-4)}}$. $E_{\text{EXT}}(t) = \frac{16}{3}t$ if $t > 0$ else 0. $w_{\text{FOF}}$ is the gain from **Selective Excitation** module. Independent samples of Gaussian noise, $\mathcal{N}(0, \tau\sigma_r^2)$, were added to the nodes at each time. We used Euler integration to solve equation dynamics. All parameters used in simulations during learning for the controller are included in Table 5.

| Symbol | Description | Value |
|--------|-------------|-------|
| $\tau$ | Population rate time constant | 0.1 |
| $I_i$ | cue input | 1 (cue present) otherwise 0 |
| $w_{\text{rec}}$ | Strength of recurrent connectivity | 1.1 |
| $w_{\text{inh}}$ | Inhibitory strength | 6 |
| $I_{\text{opto}}$ | Optogenetic inhibition strength | -3.0 (opto on)/0 (opto off) |
| $\sigma_r^2$ | Noise amplitude parameter | $2.25 \cdot 10^{-6}$ |
| $dt$ | Discretized timestep | 0.01 |

**Supplementary Table 5.** Parameters of network model of FOF.

**Cue decoding from network activity**   To quantify whether the simulated FOF network encoded future choice during the trial, the simulated activity was organized into a three-dimensional data matrix of size $N \times I \times T$ (neurons × trials × timepoints). At each time point, population activity across all neurons and trials was extracted as a two-dimensional matrix ($I \times N$) and used as input to a logistic regression classifier trained to predict the cue label (sound A vs. B) with 10-fold cross-validation.

**Time decoding from network activity**   To assess whether the attractor dynamics in the FOF model represented elapsed time within a trial, we trained linear regression models to predict the true trial time from the instantaneous network state. The same $N \times I \times T$ activity tensor was reshaped so that each time point from every trial contributed one sample to the decoding dataset, yielding a matrix of size $(I \times T) \times N$ as the input of regression. The corresponding timestamp for each sample (0–1.5 s, sampled every 0.01 s) were used as the target of regression. A linear regression model was trained to predict trial time from instantaneous network states with 10-fold cross-validation. Decoding performance was quantified as the cross-validated $R^2$ between the predicted and actual timestamps across all samples.

## Use of large language model(LLM)

Large Language Models (LLMs), including ChatGPT and DeepSeek R1, were used to assist with programming tasks and writing refinement. For programming, we provided clear specifications for input/output data structures and the logic of specific small functions before using LLMs for code generation. All generated code was manually reviewed, tested, and validated.

# References

[1] Allen Newell and Herbert Alexander Simon. *Human Problem Solving.* Prentice-Hall, Englewood Cliffs, NJ, 8. print edition, 1972.

[2] Richard P. Rumelt. *Good Strategy, Bad Strategy: The Difference and Why It Matters.* Profile Books, London, paperback ed edition, 2012.

[3] Herbert A. Simon. A Behavioral Model of Rational Choice. *The Quarterly Journal of Economics*, 69(1):99, February 1955. doi: 10.2307/1884852.

[4] Amos Tversky and Daniel Kahneman. Judgment under Uncertainty: Heuristics and Biases. *Science*, 185(4157):1124–1131, September 1974. doi: 10.1126/science.185.4157. 1124.

[5] Gerd Gigerenzer and Wolfgang Gaissmaier. Heuristic Decision Making. *Annual Review of Psychology*, 62(1):451–482, 2011. doi: 10.1146/annurev-psych-120709-145346.

[6] Mark K. Ho, David Abel, Carlos G. Correa, Michael L. Littman, Jonathan D. Cohen, and Thomas L. Griffiths. People construct simplified mental representations to plan. *Nature*, 606(7912):129–136, June 2022. doi: 10.1038/s41586-022-04743-9.

[7] Daniel Kahneman. *Thinking, Fast and Slow.* Penguin Books, London, reissued edition, 2024.

[8] Ariel Gilad, Yasir Gallero-Salas, Dominik Groos, and Fritjof Helmchen. Behavioral Strategy Determines Frontal or Posterior Location of Short-Term Memory in Neocortex. *Neuron*, 99(4):814–828.e7, August 2018. doi: 10.1016/j.neuron.2018.07.029.

[9] Falk Lieder and Thomas L. Griffiths. Resource-rational analysis: Understanding human cognition as the optimal use of limited computational resources. *Behavioral and Brain Sciences*, 43, 2020. doi: 10.1017/s0140525x1900061x.

[10] Amitai Shenhav, Sebastian Musslick, Falk Lieder, Wouter Kool, Thomas L. Griffiths, Jonathan D. Cohen, and Matthew M. Botvinick. Toward a Rational and Mechanistic Account of Mental Effort. *Annual Review of Neuroscience*, 40(1):99–124, 2017. doi: 10.1146/annurev-neuro-072116-031526.

[11] Rahul Bhui, Lucy Lai, and Samuel J Gershman. Resource-rational decision making. *Current Opinion in Behavioral Sciences*, 41:15–21, October 2021. doi: 10.1016/j. cobeha.2021.02.015.

[12] Sabine Joseph, Sundeep Teki, Sukhbinder Kumar, Masud Husain, and Timothy D. Griffiths. Resource allocation models of auditory working memory. *Brain Research*, 1640:183–192, June 2016. doi: 10.1016/j.brainres.2016.01.044.

[13] Frederick Callaway, Bas Van Opheusden, Sayan Gul, Priyam Das, Paul M. Krueger, Thomas L. Griffiths, and Falk Lieder. Rational use of cognitive resources in human planning. *Nature Human Behaviour*, 6(8):1112–1125, April 2022. doi: 10.1038/ s41562-022-01332-8.

[14] Nuo Li, Kayvon Daie, Karel Svoboda, and Shaul Druckmann. Robust neuronal dynamics in premotor cortex during motor planning. *Nature*, 532(7600):459–464, April 2016. doi: 10.1038/nature17643.

[15] Chunyu A. Duan, Yuxin Pan, Guofen Ma, Taotao Zhou, Siyu Zhang, and Ning-long Xu. A cortico-collicular pathway for motor planning in a memory-dependent perceptual decision task. *Nature Communications*, 12(1):2727, May 2021. doi: 10.1038/s41467-021-22547-9.

[16] Jeffrey C Erlich, Max Bialek, and Carlos D Brody. A cortical substrate for memory-guided orienting in the rat. *Neuron*, 72(2):330–343, October 2011. doi: 10.1016/j.neuron.2011.07.010.

[17] Charles D Kopec, Jeffrey C Erlich, Bingni W Brunton, Karl Deisseroth, and Carlos D Brody. Cortical and Subcortical Contributions to Short- Term Memory for Orienting Movements. *Neuron*, 88(2):367–377, October 2015. doi: 10.1016/j.neuron.2015.08.033.

[18] S. Funahashi, C. J. Bruce, and P. S. Goldman-Rakic. Mnemonic coding of visual space in the monkey's dorsolateral prefrontal cortex. *Journal of Neurophysiology*, 61 (2):331–349, February 1989. doi: 10.1152/jn.1989.61.2.331.

[19] David A. Markowitz, Clayton E. Curtis, and Bijan Pesaran. Multiple component networks support working memory in prefrontal cortex. *Proceedings of the National Academy of Sciences*, 112(35):11084–11089, September 2015. doi: 10.1073/pnas. 1504172112.

[20] Matthew T. Kaufman, Mark M. Churchland, Stephen I. Ryu, and Krishna V. Shenoy. Cortical activity in the null space: Permitting preparation without movement. *Nature Neuroscience*, 17(3):440–448, March 2014. doi: 10.1038/nn.3643.

[21] Clayton E. Curtis and Mark D'Esposito. Persistent activity in the prefrontal cortex during working memory. *Trends in Cognitive Sciences*, 7(9):415–423, September 2003. doi: 10.1016/s1364-6613(03)00197-9.

[22] Kyeong-Jin Tark and Clayton E Curtis. Persistent neural activity in the human frontal cortex when maintaining space that is off the map. *Nature Neuroscience*, 12(11):1463–1468, November 2009. doi: 10.1038/nn.2406.

[23] Joaquín M. Fuster. The Prefrontal Cortex—An Update. *Neuron*, 30(2):319–333, May 2001. doi: 10.1016/S0896-6273(01)00285-9.

[24] Karel Svoboda and Nuo Li. Neural mechanisms of movement planning: Motor cortex and beyond. *Current Opinion in Neurobiology*, 49:33–41, April 2018. doi: 10.1016/j. conb.2017.10.023.

[25] Mark D'Esposito and Bradley R. Postle. The Cognitive Neuroscience of Working Memory. *Annual Review of Psychology*, 66(1):115–142, January 2015. doi: 10.1146/ annurev-psych-010814-015031.

[26] Derek Evan Nee and Mark D'Esposito. The Representational Basis of Working Memory. In Robert E. Clark and Stephen J. Martin, editors, *Behavioral Neuroscience of Learning and Memory*, volume 37, pages 213–230. Springer International Publishing, Cham, 2016. doi: 10.1007/7854_2016_456.

[27] Caterina Trentin, Chris Olivers, and Heleen A. Slagter. Action Planning Renders Objects in Working Memory More Attentionally Salient. *Journal of Cognitive Neuroscience*, 36(10):2166–2183, October 2024. doi: 10.1162/jocn_a_02235.

[28] Liujunli Li, Timo Flesch, Ce Ma, Jingjie Li, Yizhou Chen, Hung-Tu Chen, and Jeffrey C. Erlich. Encoding of 2D self-centered plans and world-centered positions in the rat frontal orienting field. *The Journal of Neuroscience*, page e0018242024, August 2024. doi: 10.1523/JNEUROSCI.0018-24.2024.

[29] Elise N. Mangin, Jian Chen, Jing Lin, and Nuo Li. Behavioral measurements of motor readiness in mice. *Current Biology*, 33(17):3610–3624.e4, September 2023. doi: 10.1016/j.cub.2023.07.029.

[30] Alexa Riehle and Jean Requin. The predictive value for performance speed of preparatory changes in neuronal activity of the monkey motor and premotor cortex. *Behavioural Brain Research*, 53(1-2):35–49, February 1993. doi: 10.1016/S0166-4328(05)80264-5.

[31] Ivan Voitov and Thomas D. Mrsic-Flogel. Cortical feedback loops bind distributed representations of working memory. *Nature*, 608(7922):381–389, August 2022. doi: 10.1038/s41586-022-05014-3.

[32] M. Pare and D. P. Munoz. Saccadic reaction time in the monkey: Advanced preparation of oculomotor programs is primarily responsible for express saccade occurrence. *Journal of Neurophysiology*, 76(6):3666–3681, December 1996. doi: 10.1152/jn.1996.76.6.3666.

[33] Mark M. Churchland, Byron M. Yu, Stephen I. Ryu, Gopal Santhanam, and Krishna V. Shenoy. Neural Variability in Premotor Cortex Provides a Signature of Motor Preparation. *The Journal of Neuroscience*, 26(14):3697–3712, April 2006. doi: 10.1523/jneurosci.3762-05.2006.

[34] Munib A. Hasnain, Jaclyn E. Birnbaum, Juan Luis Ugarte Nunez, Emma K. Hartman, Chandramouli Chandrasekaran, and Michael N. Economo. Separating cognitive and motor processes in the behaving mouse. *Nature Neuroscience*, February 2025. doi: 10.1038/s41593-024-01859-1.

[35] Timothy D Hanks, Charles D Kopec, Bingni W Brunton, Chunyu A Duan, Jeffrey C Erlich, and Carlos D Brody. Distinct relationships of parietal and prefrontal cortices to evidence accumulation. *Nature*, 520(7546):220–223, April 2015. doi: 10.1038/nature14066.

[36] Diksha Gupta, Charles D. Kopec, Adrian G. Bondy, Thomas Z. Luo, Verity A. Elliott, and Carlos D. Brody. A multi-region recurrent circuit for evidence accumulation in rats, July 2024.

[37] Zengcai V. Guo, Hidehiko K. Inagaki, Kayvon Daie, Shaul Druckmann, Charles R. Gerfen, and Karel Svoboda. Maintenance of persistent activity in a frontal thalamo-cortical loop. *Nature*, 545(7653):181–186, May 2017. doi: 10.1038/nature22324.

[38] R. Becket Ebitz and Benjamin Y. Hayden. The population doctrine in cognitive neuroscience. *Neuron*, 109(19):3055–3068, October 2021. doi: 10.1016/j.neuron.2021.07.011.

[39] Hidehiko K. Inagaki, Susu Chen, Margreet C. Ridder, Pankaj Sah, Nuo Li, Zidan Yang, Hana Hasanbegovic, Zhenyu Gao, Charles R. Gerfen, and Karel Svoboda. A midbrain-thalamus-cortex circuit reorganizes cortical dynamics to initiate movement. *Cell*, 185(6):1065–1081.e23, March 2022. doi: 10.1016/j.cell.2022.02.006.

[40] Valerio Mante, David Sussillo, Krishna V. Shenoy, and William T. Newsome. Context-dependent computation by recurrent dynamics in prefrontal cortex. *Nature*, 503(7474): 78–84, November 2013. doi: 10.1038/nature12742.

[41] Mathias Mahn, Lihi Gibor, Pritish Patil, Katayun Cohen-Kashi Malina, Shir Oring, Yoav Printz, Rivka Levy, Ilan Lampl, and Ofer Yizhar. High-efficiency optogenetic silencing with soma-targeted anion-conducting channelrhodopsins. *Nature Communications*, 9(1):4125, October 2018. doi: 10.1038/s41467-018-06511-8.

[42] Andrew Gelman and Hal Stern. The Difference Between "Significant" and "Not Significant" is not Itself Statistically Significant. *The American Statistician*, 60(4):328–331, November 2006. doi: 10.1198/000313006X152649.

[43] Chunyu A Duan, Jeffrey C Erlich, and Carlos D Brody. Requirement of Prefrontal and Midbrain Regions for Rapid Executive Control of Behavior in the Rat. *Neuron*, 86(6):1491–1503, June 2015. doi: 10.1016/j.neuron.2015.05.042.

[44] Masayoshi Murakami, M Inês Vicente, Gil M Costa, and Zachary F Mainen. Neural antecedents of self-initiated actions in secondary motor cortex. *Nature Neuroscience*, 17(11):1574–1582, November 2014. doi: 10.1038/nn.3826.

[45] Tsukasa Kamigaki and Yang Dan. Delay activity of specific prefrontal interneuron subtypes modulates memory-guided behavior. *Nature Neuroscience*, 20(6):854–863, June 2017. doi: 10.1038/nn.4554.

[46] David R. Euston, Aaron J. Gruber, and Bruce L. McNaughton. The Role of Medial Prefrontal Cortex in Memory and Decision Making. *Neuron*, 76(6):1057–1070, December 2012. doi: 10.1016/j.neuron.2012.12.002.

[47] Dong-Hyun Lim, Young Ju Yoon, Eunsil Her, Suehee Huh, and Min Whan Jung. Active maintenance of eligibility trace in rodent prefrontal cortex. *Scientific Reports*, 10(1):18860, November 2020. doi: 10.1038/s41598-020-75820-0.

[48] Chunyu A. Duan, Marino Pagan, Alex T. Piet, Charles D. Kopec, Athena Akrami, Alexander J. Riordan, Jeffrey C. Erlich, and Carlos D. Brody. Collicular circuits for flexible sensorimotor routing. *Nature Neuroscience*, 24(8):1110–1120, August 2021. doi: 10.1038/s41593-021-00865-x.

[49] Liping Yu, Jiawei Hu, Chenlin Shi, Li Zhou, Maozhi Tian, Jiping Zhang, and Jinghong Xu. The causal role of auditory cortex in auditory working memory. *eLife*, 10:e64457, April 2021. doi: 10.7554/eLife.64457.

[50] Alexandra Libby and Timothy J. Buschman. Rotational dynamics reduce interference between sensory and memory representations. *Nature Neuroscience*, 24(5):715–726, May 2021. doi: 10.1038/s41593-021-00821-9.

[51] Florent Barthas and Alex C. Kwan. Secondary Motor Cortex: Where 'Sensory' Meets 'Motor' in the Rodent Frontal Cortex. *Trends in Neurosciences*, 40(3):181–193, March 2017. doi: 10.1016/j.tins.2016.11.006.

[52] Hidehiko K. Inagaki, Susu Chen, Kayvon Daie, Arseny Finkelstein, Lorenzo Fontolan, Sandro Romani, and Karel Svoboda. Neural Algorithms and Circuits for Motor Planning. *Annual Review of Neuroscience*, 45(1):249–271, July 2022. doi: 10.1146/annurev-neuro-092021-121730.

[53] Ranulfo Romo, Carlos D. Brody, Adrián Hernández, and Luis Lemus. Neuronal correlates of parametric working memory in the prefrontal cortex. *Nature*, 399(6735): 470–473, June 1999. doi: 10.1038/20939.

[54] Vahid Esmaeili and Mathew E. Diamond. Neuronal Correlates of Tactile Working Memory in Prefrontal and Vibrissal Somatosensory Cortex. *Cell Reports*, 27(11): 3167–3181.e5, June 2019. doi: 10.1016/j.celrep.2019.05.034.

[55] Joseph T. McGuire and Matthew M. Botvinick. Prefrontal cortex, cognitive control, and the registration of decision costs. *Proceedings of the National Academy of Sciences*, 107(17):7922–7926, April 2010. doi: 10.1073/pnas.0910662107.

[56] Trinity B. Crapse and Marc A. Sommer. Corollary discharge across the animal kingdom. *Nature Reviews Neuroscience*, 9(8):587–600, August 2008. doi: 10.1038/nrn2457.

[57] Sten Grillner. Evolution of the vertebrate motor system — from forebrain to spinal cord. *Current Opinion in Neurobiology*, 71:11–18, December 2021. doi: 10.1016/j. conb.2021.07.016.

[58] Sten Grillner. *The Brain in Motion: From Microcircuits to Global Brain Function*. The MIT Press, Cambridge London, 2023.

[59] Rifqi O. Affan, Ian M. Bright, Luke N. Pemberton, Nathanael A. Cruzado, Benjamin B. Scott, and Marc W. Howard. Ramping dynamics in the frontal cortex unfold over multiple timescales during motor planning. *Journal of Neurophysiology*, 133(2):625–637, February 2025. doi: 10.1152/jn.00234.2024.

[60] C. K. Machens, R. Romo, and C. D. Brody. Functional, But Not Anatomical, Separation of "What" and "When" in Prefrontal Cortex. *Journal of Neuroscience*, 30(1): 350–360, January 2010. doi: 10.1523/JNEUROSCI.3276-09.2010.

[61] Stephen M. Fleming. Metacognition and Confidence: A Review and Synthesis. *Annual Review of Psychology*, 75(1):241–268, January 2024. doi: 10.1146/ annurev-psych-022423-032425.

[62] Aaron A. Wilber, Benjamin J. Clark, Tyler C. Forster, Masami Tatsuno, and Bruce L. McNaughton. Interaction of Egocentric and World-Centered Reference Frames in the Rat Posterior Parietal Cortex. *Journal of Neuroscience*, 34(16):5431–5446, April 2014. doi: 10.1523/JNEUROSCI.0511-14.2014.

[63] Dario Campagner, Ruben Vale, Yu Lin Tan, Panagiota Iordanidou, Oriol Pavón Arocas, Federico Claudi, A. Vanessa Stempel, Sepiedeh Keshavarzi, Rasmus S. Petersen, Troy W. Margrie, and Tiago Branco. A cortico-collicular circuit for orienting to shelter during escape. *Nature*, 613(7942):111–119, January 2023. doi: 10.1038/s41586-022-05553-9.

[64] Joaquin M. Fuster and Garrett E. Alexander. Neuron Activity Related to Short-Term Memory. *Science*, 173(3997):652–654, August 1971. doi: 10.1126/science.173.3997.652.

[65] Yasir Gallero-Salas, Shuting Han, Yaroslav Sych, Fabian F. Voigt, Balazs Laurenczy, Ariel Gilad, and Fritjof Helmchen. Sensory and Behavioral Components of Neocortical Signal Flow in Discrimination Tasks with Short-Term Memory. *Neuron*, 109(1):135–148.e6, January 2021. doi: 10.1016/j.neuron.2020.10.017.

[66] Hidehiko K. Inagaki, Lorenzo Fontolan, Sandro Romani, and Karel Svoboda. Discrete attractor dynamics underlies persistent activity in the frontal cortex. *Nature*, 566 (7743):212–217, February 2019. doi: 10.1038/s41586-019-0919-7.

[67] Klaus Wimmer, Duane Q Nykamp, Christos Constantinidis, and Albert Compte. Bump attractor dynamics in prefrontal cortex explains behavioral precision in spatial working memory. *Nature Neuroscience*, 17(3):431–439, March 2014. doi: 10.1038/nn.3645.

[68] Elizabeth S. Lorenc, Remington Mallett, and Jarrod A. Lewis-Peacock. Distraction in Visual Working Memory: Resistance is Not Futile. *Trends in Cognitive Sciences*, 25 (3):228–239, March 2021. doi: 10.1016/j.tics.2020.12.004.

[69] Margaret M Henderson, Rosanne L Rademaker, and John T Serences. Flexible utilization of spatial- and motor-based codes for the storage of visuo-spatial information. *eLife*, 11:e75688, May 2022. doi: 10.7554/eLife.75688.

[70] Freek Van Ede. Visual working memory and action: Functional links and bi-directional influences. *Visual Cognition*, 28(5-8):401–413, September 2020. doi: 10.1080/13506285. 2020.1759744.

[71] Nicholas E. Myers, Mark G. Stokes, and Anna C. Nobre. Prioritizing Information during Working Memory: Beyond Sustained Internal Attention. *Trends in Cognitive Sciences*, 21(6):449–461, June 2017. doi: 10.1016/j.tics.2017.03.010.

[72] P. Znamenskiy and A. M. Zador. Corticostriatal neurons in auditory cortex drive decisions during auditory discrimination. *Nature*, 497:482–485, May 2013.

[73] Joao Barbosa, Heike Stein, Rebecca L. Martinez, Adrià Galan-Gadea, Sihai Li, Josep Dalmau, Kirsten C. S. Adam, Josep Valls-Solé, Christos Constantinidis, and Albert Compte. Interplay between persistent activity and activity-silent dynamics in the prefrontal cortex underlies serial biases in working memory. *Nature Neuroscience*, 23 (8):1016–1024, August 2020. doi: 10.1038/s41593-020-0644-4.

[74] Mark G. Stokes. 'Activity-silent' working memory in prefrontal cortex: A dynamic coding framework. *Trends in Cognitive Sciences*, 19(7):394–405, July 2015. doi: 10. 1016/j.tics.2015.05.004.

[75] Samuel J. Gershman, Eric J. Horvitz, and Joshua B. Tenenbaum. Computational rationality: A converging paradigm for intelligence in brains, minds, and machines. *Science*, 349(6245):273–278, July 2015. doi: 10.1126/science.aac6076.

[76] Shuze Liu, Samuel J. Gershman, and Bilal A. Bari. Quantifying the Cost of Context Sensitivity in Decision-Making. *Topics in Cognitive Science*, page tops.70030, October 2025. doi: 10.1111/tops.70030.

[77] Amitai Shenhav, Matthew M. Botvinick, and Jonathan D. Cohen. The Expected Value of Control: An Integrative Theory of Anterior Cingulate Cortex Function. *Neuron*, 79 (2):217–240, July 2013. doi: 10.1016/j.neuron.2013.07.007.

[78] George Paxinos and Charles Watson. *The Rat Brain in Stereotaxic Coordinates*. Acad. Press, San Diego [u.a., 2004.

[79] Mihály Vöröslakos, Peter C Petersen, Balázs Vöröslakos, and György Buzsáki. Metal microdrive and head cap system for silicon probe recovery in freely moving rodent. *eLife*, 10:e65859, May 2021. doi: 10.7554/eLife.65859.

[80] Marius Pachitariu, Nicholas Steinmetz, Shabnam Kadir, Matteo Carandini, and Harris Kenneth D. Kilosort: Realtime spike-sorting for extracellular electrophysiology with hundreds of channels, June 2016.

[81] Marius Pachitariu, Shashwat Sridhar, Jacob Pennington, and Carsen Stringer. Spike sorting with Kilosort4. *Nature Methods*, 21(5):914–921, May 2024. doi: 10.1038/s41592-024-02232-7.

[82] Xiaoxuan Jia, Joshua H. Siegle, Corbett Bennett, Samuel D. Gale, Daniel J. Denman, Christof Koch, and Shawn R. Olsen. High-density extracellular probes reveal dendritic backpropagation and facilitate neuron classification. *Journal of Neurophysiology*, 121 (5):1831–1847, May 2019. doi: 10.1152/jn.00680.2018.

[83] D. Mayerich, L. Abbott, and B. McCORMICK. Knife-edge scanning microscopy for imaging and reconstruction of three-dimensional anatomical structures of the mouse brain. *Journal of Microscopy*, 231(1):134–143, July 2008. doi: 10.1111/j.1365-2818.2008.02024.x.

[84] Timothy Ragan, Lolahon R Kadiri, Kannan Umadevi Venkataraju, Karsten Bahlmann, Jason Sutin, Julian Taranda, Ignacio Arganda-Carreras, Yongsoo Kim, H Sebastian Seung, and Pavel Osten. Serial two-photon tomography for automated ex vivo mouse brain imaging. *Nature Methods*, 9(3):255–258, March 2012. doi: 10.1038/nmeth.1854.

[85] Douglas Bates, Martin Mächler, Ben Bolker, and Steve Walker. Fitting Linear Mixed-Effects Models Using lme4. *Journal of Statistical Software*, 67(1):1–48, October 2015. doi: 10.18637/jss.v067.i01.

[86] Talmo D. Pereira, Nathaniel Tabris, Arie Matsliah, David M. Turner, Junyu Li, Shruthi Ravindranath, Eleni S. Papadoyannis, Edna Normand, David S. Deutsch, Z. Yan Wang, Grace C. McKenzie-Smith, Catalin C. Mitelut, Marielisa Diez Castro, John D'Uva, Mikhail Kislin, Dan H. Sanes, Sarah D. Kocher, Samuel S.-H. Wang, Annegret L. Falkner, Joshua W. Shaevitz, and Mala Murthy. SLEAP: A deep learning system for multi-animal pose tracking. *Nature Methods*, 19(4):486–495, April 2022. doi: 10.1038/s41592-022-01426-1.

[87] Jeffrey Erlich, Josh Moller-Mara, and Jingjie Li. Erlichlab/elutils: First Zenodo Release. Zenodo, May 2021.

[88] J-R. King and S. Dehaene. Characterizing the dynamics of mental representations: The temporal generalization method. *Trends in Cognitive Sciences*, 18(4):203–210, April 2014. doi: 10.1016/j.tics.2014.01.002.

[89] Mike X Cohen. *Analyzing Neural Time Series Data: Theory and Practice*. The MIT Press, January 2014. doi: 10.7551/mitpress/9609.001.0001.

[90] M. Pagan and N. C. Rust. Dynamic Target Match Signals in Perirhinal Cortex Can Be Explained by Instantaneous Computations That Act on Dynamic Input from Inferotemporal Cortex. *Journal of Neuroscience*, 34(33):11067–11084, August 2014. doi: 10.1523/JNEUROSCI.4040-13.2014.

[91] Hadley Wickham. *Ggplot2: Elegant Graphics for Data Analysis.* Use R! Springer International Publishing : Imprint: Springer, Cham, 2nd ed. 2016 edition, 2016. doi: 10.1007/978-3-319-24277-4.

[92] Achim Zeileis and Torsten Hothorn. Diagnostic checking in regression relationships. *R News*, 2(3):7–10, 2002.

[93] G. N. Wilkinson and C. E. Rogers. Symbolic Description of Factorial Models for Analysis of Variance. *Applied Statistics*, 22(3):392, 1973. doi: 10.2307/2346786.

[94] Alex T. Piet, Jeffrey C. Erlich, Charles D. Kopec, and Carlos D. Brody. Rat Prefrontal Cortex Inactivations during Decision Making Are Explained by Bistable Attractor Dynamics. *Neural Computation*, 29(11):2861–2886, August 2017. doi: 10.1162/neco_a_01005.

[95] H. R. Wilson and J. D. Cowan. A mathematical theory of the functional dynamics of cortical and thalamic nervous tissue. *Kybernetik*, 13(2):55–80, September 1973. doi: 10.1007/BF00288786.

[96] Alain Destexhe and Terrence J. Sejnowski. The Wilson–Cowan model, 36 years later. *Biological Cybernetics*, 101(1):1–2, July 2009. doi: 10.1007/s00422-009-0328-3.

# 1 Statistical appendix

## 1.1 Figure 2

For choice coding cell count in the Ego/Allo task
=== Chi-square Test Report ===
Observed Table:
```
        864(ego_choice)         1725(ego_nonchoice)
        374(allo_choice)        2812(allo_nonchoice)

Expected Counts:
    555.01      2033.99
    682.99      2503.01
Chi-square statistic: 396.8971
Degrees of freedom:    1
p-value:               0
```

## 1.2 Figure 4

### 1.2.1 Figure 4e, Left Panel, Ego task, Sham/FOF comparison, Cue period

Likelihood ratio test

```
Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
  #Df  LogLik Df  Chisq Pr(>Chisq)
1   3 -472.40
2   4 -470.07  1 4.6549    0.03097 *
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

### 1.2.2 Figure 4e, Right Panel, Allo task, Sham/FOF comparison, Cue period

Likelihood ratio test

```
Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
  #Df  LogLik Df  Chisq Pr(>Chisq)
1   3 -441.03
2   4 -439.01  1 4.0364    0.04453 *
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

### 1.2.3 Figure 4e, Main comparison between Allo/Ego task, Cue period

Likelihood ratio test

```
Model 1: mu ~ 0 + ego_task + isopto + isopto:fof_fiber + isopto:fof_fiber:ego_task +
    (1 | subjid/sessid)
Model 2: mu ~ 0 + ego_task + isopto + isopto:fof_fiber + isopto:fof_fiber +
    (1 | subjid/sessid)
```

```
#Df  LogLik Df Chisq Pr(>Chisq)
1    6 -776.33
2    5 -776.49 -1 0.324       0.5692
```

### 1.2.4  Figure 4f, Left panel, Ego task, Sham/FOF comparison, Delay period

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
  #Df  LogLik Df  Chisq Pr(>Chisq)
1    3 -478.16
2    4 -473.39  1 9.5406     0.00201 **
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

### 1.2.5  Figure 4f, Right panel, Allo task, Sham/FOF comparison, Delay period

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
  #Df  LogLik Df  Chisq Pr(>Chisq)
1    3 -445.28
2    4 -444.51  1 1.5456       0.2138
```

### 1.2.6  Figure 4f, Main comparison Between Allo/Ego task, Delay period

```
Likelihood ratio test

Model 1: mu ~ 0 + ego_task + isopto + isopto:fof_fiber + isopto:fof_fiber:ego_task +
    (1 | subjid/sessid)
Model 2: mu ~ 0 + ego_task + isopto + isopto:fof_fiber + isopto:fof_fiber +
    (1 | subjid/sessid)
  #Df  LogLik Df  Chisq Pr(>Chisq)
1    6 -786.52
2    5 -790.51 -1 7.9718    0.004751 **
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## 1.3  Figure 5

### 1.3.1  Figure 5g

```
for t=0.95 (left panel)
Wilcoxon rank-sum (Mann-Whitney U) test results:
------------------------------------------------------------
Comparison          p-value      U          z       n1  n2  med1   med2
------------------------------------------------------------
AC vs FOF         : 0.0004675    829.000   3.499    15   70  0.648  0.537
AC vs SC          : 0.005677     212.000   2.766    15   18  0.648  0.524
AC vs mPFC        : 0.003669     194.000   2.905    15   16  0.648  0.530
```

```
1565   ----------------------------------------------------------
1566
1567   for t=1.05  (right panel)
1568   Wilcoxon rank-sum (Mann-Whitney U) test results:
1569   ----------------------------------------------------------
1570   Comparison          p-value      U         z       n1  n2  med1  med2
1571   ----------------------------------------------------------
1572   AC vs FOF       :   0.03391   709.500   2.121    15  70  0.597  0.551
1573   AC vs SC        :   0.03762   193.000   2.079    15  18  0.597  0.524
1574   AC vs mPFC      :   0.03796   173.000   2.075    15  16  0.597  0.536
1575   ----------------------------------------------------------
```

## 1.4  Figure 7

### 1.4.1  Figure 7c

**Supplementary Table 6.** Mixed-effects likelihood-ratio test comparing movement time (MT) on each post-transformation day to baseline.

| Day | p-Val | LRStat | $\Delta$DF | $\text{AIC}_{m1}$ | $\text{BIC}_{m1}$ | $\text{LogLik}_{m1}$ | $\text{AIC}_{m0}$ | $\text{BIC}_{m0}$ | $\text{LogLik}_{m0}$ |
|---|---|---|---|---|---|---|---|---|---|
| 0 | 0.16993 | 1.8836 | 1 | -1569.6 | -1554.8 | 787.79 | -1569.5 | -1549.8 | 788.74 |
| 1 | 0.15637 | 2.0090 | 1 | -2678.3 | -2661.6 | 1342.20 | -2678.3 | -2656.0 | 1343.20 |
| 2 | 5.1536e-06 | 20.779 | 1 | -2802.7 | -2786.2 | 1404.30 | -2821.4 | -2799.4 | 1414.70 |
| 3 | 9.0271e-08 | 28.572 | 1 | -2655.8 | -2639.5 | 1330.90 | -2682.3 | -2660.7 | 1345.20 |
| 4 | 3.9951e-10 | 39.115 | 1 | -1971.9 | -1956.1 | 988.97 | -2009.1 | -1988.0 | 1008.50 |
| 5 | 7.7906e-09 | 33.327 | 1 | -1988.7 | -1972.6 | 997.34 | -2020.0 | -1998.5 | 1014.00 |
| 6 | 7.7716e-16 | 64.925 | 1 | -2036.5 | -2020.8 | 1021.30 | -2099.4 | -2078.5 | 1053.70 |
| 7 | 6.3647e-09 | 33.720 | 1 | -2018.8 | -2003.1 | 1012.40 | -2050.6 | -2029.6 | 1029.30 |

### 1.4.2  Figure 7e, Left panel, Allo task, Sham/FOF comparison

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
  #Df  LogLik Df  Chisq Pr(>Chisq)
1   3 -291.09
2   4 -290.38  1 1.4303      0.2317
```

### 1.4.3  Figure 7e, Right panel, 2AFC task, Sham/FOF comparison

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
  #Df  LogLik Df  Chisq Pr(>Chisq)
1   3 -89.993
2   4 -86.932  1 6.1215     0.01335 *
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

### 1.4.4 Figure 7e, Main comparison between Allo/2AFC task

```
Likelihood ratio test

Model 1: mu ~ 0 + full_task + isopto + isopto:fof_fiber + isopto:fof_fiber:full_task +
    (1 | subjid/sessid)
Model 2: mu ~ 0 + full_task + isopto + isopto:fof_fiber + isopto:fof_fiber +
    (1 | subjid/sessid)
   #Df  LogLik Df  Chisq Pr(>Chisq)
1    6 -332.13
2    5 -335.28 -1 6.2956     0.0121 *
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## 1.5 Supplementary figure 6

## 1.6 Supplementary figure 6e, Left panel, Ego task, Sham/FOF comparison

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
   #Df  LogLik Df  Chisq Pr(>Chisq)
1    3 -115.73
2    4 -114.32  1 2.8171    0.09326 .
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## 1.7 Supplementary figure 6e, Right panel, Allo task, Sham/FOF comparison

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
   #Df  LogLik Df  Chisq Pr(>Chisq)
1    3 -236.04
2    4 -234.91  1 2.2586     0.1329
```

### 1.7.1 Supplementary figure 6e, Main comparison between Allo/Ego task

```
Likelihood ratio test

Model 1: mu ~ 0 + ego_task + isopto + isopto:fof_fiber + isopto:fof_fiber:ego_task +
    (1 | sessid)
Model 2: mu ~ 0 + ego_task + isopto + isopto:fof_fiber + (1 | sessid)
   #Df  LogLik Df Chisq Pr(>Chisq)
1    5 -358.34
2    4 -358.35 -1 0.015     0.9025
```

## 1.8 Supplementary Figure 6f, Left Panel, Ego task, Sham/FOF comparison

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
  #Df  LogLik Df  Chisq Pr(>Chisq)
1   3 -118.73
2   4 -116.02  1 5.4103    0.02002 *
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## 1.9 Supplementary Figure 6f, Right panel, Allo task, Sham/FOF comparison

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
  #Df  LogLik Df  Chisq Pr(>Chisq)
1   3 -238.48
2   4 -238.01  1 0.9379    0.3328
```

### 1.9.1 Supplementary figure 6f, Main comparison between Allo/Ego task

```
Likelihood ratio test

Model 1: mu ~ 0 + ego_task + isopto + isopto:fof_fiber + isopto:fof_fiber:ego_task +
    (1 | sessid)
Model 2: mu ~ 0 + ego_task + isopto + isopto:fof_fiber + (1 | sessid)
  #Df  LogLik Df Chisq Pr(>Chisq)
1   5 -363.23
2   4 -365.27 -1 4.079    0.04342 *
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## 1.10 Supplementary figure 7

### 1.10.1 Supplementary figure 7e, left Panel, Ego task, Sham/FOF comparison

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
  #Df  LogLik Df  Chisq Pr(>Chisq)
1   3 -571.04
2   4 -530.34  1 81.403  < 2.2e-16 ***
```

### 1.10.2 Supplementary figure 7e, right Panel, Allo task, Sham/FOF comparison

```
Likelihood ratio test
```

```
1680
1681 Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
1682 Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
1683    #Df  LogLik Df  Chisq Pr(>Chisq)
1684 1    3 -477.31
1685 2    4 -477.25  1 0.1232     0.7256
```

### 1.10.3  Supplementary figure 7f, left Panel, Ego task, Sham/FOF comparison

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
   #Df  LogLik Df  Chisq Pr(>Chisq)
1    3 -496.79
2    4 -483.67  1 26.235  3.023e-07 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

### 1.10.4  Supplementary figure 7f, right Panel, Allo task, Sham/FOF comparison

```
Likelihood ratio test

Model 1: mu ~ 0 + isopto + fof_fiber + (1 | sessid)
Model 2: mu ~ 0 + isopto + fof_fiber + isopto:fof_fiber + (1 | sessid)
   #Df  LogLik Df  Chisq Pr(>Chisq)
1    3 -475.75
2    4 -466.63  1 18.249  1.938e-05 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## 1.11  Supplementary figure 9

### 1.11.1  Supplementary figure 9g hit trials

```
Likelihood ratio test

Model 1: logMT ~ 1 + task + (1 | subjid)
Model 2: logMT ~ 1 + (1 | subjid)
   #Df LogLik Df  Chisq Pr(>Chisq)
1    4 4295.2
2    3 4209.3 -1 171.99  < 2.2e-16 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

### 1.11.2  Supplementary figure 9g error trials

```
Likelihood ratio test

Model 1: logMT ~ 1 + task + (1 | subjid)
Model 2: logMT ~ 1 + (1 | subjid)
   #Df LogLik Df  Chisq Pr(>Chisq)
```

```
1  4 961.48
2  3 942.99 -1 36.982  1.192e-09 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```