An ancient transcription factor functions as the master regulator of primary cilia formation

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Abstract

How ancient transcription factors are repurposed during evolution to drive the functional diversification of conserved organelles remains a fundamental question in biology. Although displaying a high degree of conservation in basic morphology, eukaryotic cilia vary extensively in their sizes and functions. Previously, we showed that an evolutionarily ancient transcription factor, X chromosome–associated protein 5 (Xap5), is required for motile ciliogenesis in unicellular and multicellular organisms. Here we show that Xap5 functions as the master regulator of primary ciliogenesis. Xap5 associates with the nuclear protein Non-POU domain–containing octamer-binding protein (Nono) to form a regulatory module that directly binds and activates the transcription factors Sox5 and Sox9. Genetic ablation of *Xap5* or *Nono* impairs primary ciliogenesis, and loss of Sox5 disrupts the downstream ciliogenic program, consistent with the established role of Sox9. Collectively, our results provide new insight into how complex ciliary transcription factor networks determine ciliary diversity during evolution, and will have a profound impact on our understanding of human ciliopathies.

Introduction

A central challenge in evolutionary biology is to understand how a limited number of ancient conserved genes generate tremendous cellular diversity through functional redeployment[1-3]. This challenge is particularly evident in the roles of master transcription factors, which must be precisely regulated across different cellular contexts to control specific developmental programs[4-7]. Such functional plasticity is thought to arise through evolutionary co-option, the process by which existing regulators are recruited into new regulatory modules to drive lineage- or organelle-specific innovations[8-10]. Elucidating the molecular strategies underlying this repurposing is essential for understanding how complex biological systems evolve. The biogenesis of the cilium—an evolutionarily ancient organelle that plays key roles in signaling and motility[11-14], provides an important model for investigating this fundamental question, and its dysfunction can lead to a class of genetic disorders known as ciliopathies[15].

The assembly of the cilium, or ciliogenesis, requires the precise transcriptional coordination of hundreds of genes[16-18]. In metazoans, the regulatory factor X (Rfx) and Forkhead box protein J1 (Foxj1) transcription factor families have been identified as key regulators of ciliogenesis[19-22]. Rfx factors, such as DAF-19 in *C. elegans*, regulate the expression of genes essential for building the core ciliary structure[23], while Foxj1 directs the motile ciliogenic program[24,25]. However, this well-established regulatory network primarily applies to motile, multiciliated cells. How the upstream, cell-type-specific transcriptional programs are initiated, particularly for the non-motile primary cilium found on most somatic cells, remains poorly understood[12,26].

The conserved transcription factor Xap5 serves as a prime example of this challenge, presenting a compelling biological paradox. Our previous work showed that Xap5 is a critical regulator for the assembly of motile appendages—dynamic structures that generate movement by beating in

69 fluid environments—a role conserved from the cilia of the unicellular alga *Chlamydomonas* to the
70 flagella of mammalian sperm[27,28]. These findings cemented Xap5's role in an ancient program for
71 motility but, in doing so, created a profound biological enigma: why is this regulator, seemingly
72 dedicated to motility, highly expressed in somatic cells that exclusively form non-motile primary
73 cilia for sensory functions? Whether Xap5 plays a role in primary ciliogenesis in these cells, and
74 the mechanism by which it might be co-opted for this distinct function, remains to be determined[29].
75 In this study, we specifically address these questions by characterizing the function of Xap5 in
76 primary ciliogenesis using NIH/3T3 mouse fibroblasts, a standard model for primary cilium
77 formation upon serum starvation[30]. Here, we identify the Non-POU domain-containing octamer-
78 binding protein (Nono) as a critical interacting partner of Xap5. Our integrated analyses
79 demonstrate that Xap5 recruits its cofactor Nono to regulate primary cilia formation by modulating
80 a downstream transcriptional axis involving Sox5 and Sox9. While Sox9 has been previously
81 implicated in primary ciliogenesis[31-33], our findings for the first time identify Xap5 as a critical
82 upstream activator of these transcription factors in somatic cells. Together, these findings define
83 the complete Xap5-Nono-Sox5/9 axis for primary ciliogenesis. Moreover, they offer a powerful
84 illustration of evolutionary co-option in action, establishing a mechanistic framework for how
85 context-specific partnerships empower ancient transcription factors to drive cellular innovation.
86
87 **Results**
88 **Xap5 is essential for primary cilia formation**
89 To investigate the role of Xap5 in primary cilia formation, we first determined its subcellular
90 localization in NIH/3T3 cells. Immunofluorescence staining with a specific anti-Xap5 antibody
91 revealed that endogenous Xap5 is predominantly nuclear (Fig. 1a), consistent with our previous

92 work. We confirmed this finding via ectopic expression of a Flag-tagged Xap5 construct (Fig. 1b).
93 Consistent with its nuclear localization, bioinformatic analysis predicted a putative nuclear
94 localization signal (NLS) within the Xap5 protein (Supplementary Fig. 1a). To functionally
95 validate this sequence, we generated a site-directed mutant (Flag-Xap5-ΔNLS) (Supplementary
96 Fig. 1a). Crucially, this mutation abolished the nuclear import of Xap5, leading to its
97 mislocalization in the cytoplasm and confirming that this NLS is essential for its function (Fig.
98 1b).

99 To investigate the function of Xap5, we generated *Xap5* knockout (KO) NIH/3T3 cell lines
100 using the CRISPR/Cas9 system[34] (Supplementary Fig. 1b). The complete depletion of Xap5
101 protein product was confirmed by western blotting (Fig. 1c). Upon inducing ciliogenesis via serum
102 starvation (Fig. 1d), we first observed a striking defect in ciliary length (Fig. 1e). Although
103 comparable to wild-type (WT) cells before starvation, *Xap5* KO cells exhibited a significant
104 shortening of their primary cilia post-starvation (Fig. 1e, f). To determine if this length defect
105 stemmed from an overall failure in ciliogenesis, we next quantified the ciliation frequency.
106 Interestingly, both WT and *Xap5* KO cells responded robustly to the starvation stimulus, showing
107 a comparable increase in the percentage of ciliated cells over 48 h (Fig. 1g). These results
108 collectively demonstrate that Xap5 is dispensable for the initiation of ciliogenesis but plays an
109 essential role in the subsequent elongation and/or maintenance of the ciliary axoneme.

110 **Xap5 forms a nuclear regulatory module with its cofactor Nono**

111 Having established the essential role of Xap5 in primary ciliogenesis, we next sought to uncover
112 its underlying regulatory mechanism by identifying its synergistic cofactors. An unbiased
113 immunoprecipitation-mass spectrometry (IP-MS) screen for Xap5 interactors identified several
114 nuclear proteins (Supplementary Fig. 2), among which we prioritized Non-POU domain-

115 containing octamer-binding protein (Nono), given its well-established, pleiotropic roles in
116 transcriptional regulation and RNA processing[35-37].

117 As a prerequisite for their interaction, we first confirmed that Nono is also a predominantly
118 nuclear protein in NIH/3T3 cells, a localization we validated for both the endogenous protein via
119 immunofluorescence and an ectopically expressed Nono-GFP fusion protein (Fig. 2a). To
120 biochemically validate their physical association, we next performed endogenous co-
121 immunoprecipitation (Co-IP) assays. These results confirmed a robust and specific interaction, as
122 immunoprecipitation of Xap5 specifically co-precipitated Nono (Fig. 2b). Taken together, these
123 localization and co-immunoprecipitation data provide strong evidence that Xap5 and Nono form
124 a bona fide regulatory complex within the nucleus.

125 **The Xap5-Nono module is functionally required for ciliogenesis**

126 Given that Nono forms a complex with the essential ciliogenesis factor Xap5, we hypothesized
127 that the entire module is functionally indispensable. To test this, we generated *Nono* KO NIH/3T3
128 cells using the CRISPR/Cas9 system (Supplementary Fig. 3a). Indeed, upon serum starvation,
129 *Nono* KO cells phenocopied the *Xap5* KO. They exhibited significantly shorter primary cilia
130 compared to wild-type (WT) controls, yet displayed a comparable increase in ciliation frequency
131 (Fig. 3b-d). Notably, in addition to the ciliary defects, we observed a significant increase in nuclear
132 size in *Nono* KO cells (Fig. 3b; Supplementary Fig. 3b). Together, these findings demonstrate that
133 Nono, like its partner Xap5, is dispensable for ciliary initiation but essential for ciliary elongation
134 and maintenance, and may also have a broader role in maintaining nuclear architecture.

135 **The Xap5-Nono module co-regulates a downstream ciliogenic transcriptional cascade**

136 To determine if the Xap5-Nono module cooperates at the transcriptional level, we performed
137 RNA-sequencing (RNA-seq) on WT, *Xap5* KO, and *Nono* KO cells following 24 h of serum

138 starvation, a time point where the phenotype was robustly established. This analysis revealed
139 widespread transcriptional changes; *Xap5* KO cells showed 1,116 upregulated and 1,071
140 downregulated genes, while *Nono* KO cells had 1,425 upregulated and 1,042 downregulated genes
141 relative to WT (fold change ≥ 1.5; Supplementary Fig. 4a, b; Supplementary Data 1, 2).

142 Given that loss of either Xap5 or Nono impairs ciliogenesis, we hypothesized that they function
143 together as transcriptional activators of a shared ciliogenic program. We therefore focused our
144 subsequent analysis on the genes commonly downregulated in both knockouts. Notably, Gene
145 Ontology (GO) analysis of these genes revealed a highly significant enrichment for ciliary
146 categories (Fig. 4a, b; Supplementary Fig. 4c, d). To investigate this further, we compared this
147 downregulated gene set against a comprehensive ciliary gene list (Supplementary Data 3), which
148 was assembled from the SYSCILIA gold standard (SCGSv2)[18] and manual literature curation. This
149 comparison confirmed a significant overlap between the two gene sets (Fig. 4c). We validated
150 several of these shared targets—including the transcription factors *Sox5* and *Sox9*, and the ciliary
151 component *Pkd2*—by qPCR (Fig. 4d, e).

152 Among this cohort of downregulated genes, two transcription factors, Sox5 and Sox9, stood out.
153 Sox9 is a known regulator of primary cilia in other contexts, and both factors are known to
154 cooperate in developmental programs. This made them compelling candidates for further
155 investigation as potential downstream effectors in our system. Consistent with this hypothesis,
156 western blot analysis confirmed that the protein levels of both Sox5 and Sox9 were markedly
157 reduced in *Xap5* and *Nono* KO cells (Fig. 4f). These data strongly indicate that Xap5 and Nono
158 co-regulate a transcriptional network that includes the downstream effectors Sox5 and Sox9,
159 thereby driving primary cilia formation.

160 **The Xap5-Nono module directly co-occupies the regulatory regions of Sox5 and Sox9**

161 To determine if Xap5 and Nono directly regulate their target genes, we mapped their genome-wide
162 chromatin occupancy using CUT&Tag[38] in NIH/3T3 cells (Supplementary Data 4, 5). This
163 analysis revealed a strong enrichment of both factors at promoter regions; approximately 30% of
164 Xap5 peaks and a striking 50% of Nono peaks were located at promoters (Supplementary Fig. 5a,
165 b). Consistent with this, genomic heatmaps demonstrated that both Xap5 and Nono binding signals
166 were highly enriched around the transcription start sites of their target genes (Fig. 5a, b;
167 Supplementary Fig. 5c, d).

168 Integrating these binding profiles with our RNA-seq data revealed a strong correlation between
169 genes bound by Xap5 or Nono and those whose expression was altered upon KO (Fig. 5c, d),
170 supporting a model of direct transcriptional regulation. De novo motif analysis revealed that Xap5
171 and Nono recognize a highly similar DNA-binding motif (Fig. 5e). Importantly, this Xap5 motif
172 is virtually identical to that from our previous work in testicular germ cells (Fig. 5e), demonstrating
173 that Xap5 maintains its DNA-binding specificity across distinct cellular contexts.

174 Crucially, we found a significant overlap between the Xap5/Nono co-occupied genes and our
175 curated ciliary gene set (Fig. 5f). Among these, we observed direct co-occupancy of Xap5 and
176 Nono at the promoter regions of the key downstream regulators Sox5 and Sox9, a finding we
177 validated by CUT&Tag-qPCR (Fig. 5g). Collectively, these genomic data provide strong evidence
178 that Xap5 and Nono form a complex that directly binds to the regulatory regions of a common set
179 of ciliary genes, including Sox5 and Sox9, to orchestrate primary cilia formation.

180 **Sox5, a key downstream effector of the Xap5-Nono pathway, is essential for ciliogenesis**

181 Our finding that Xap5 and Nono directly co-regulate Sox5 and Sox9 prompted us to investigate
182 their respective roles in ciliogenesis. Given that the essential role of Sox9 in maintaining primary
183 cilia length is well-documented[31,33], we focused our functional analysis on the uncharacterized role

184 of Sox5. To this end, we generated *Sox5* KO NIH/3T3 cells using the CRISPR/Cas9 system (Fig.
185 6a; Supplementary Fig. 6).

186 Analysis after serum starvation revealed that *Sox5* KO cells precisely phenocopied the upstream
187 knockouts. *Sox5* KO cells exhibited significantly shorter primary cilia than WT controls (Fig. 6b,
188 c), yet this occurred despite a normal ciliation response, with both genotypes showing a robust and
189 comparable increase in ciliation frequency (Fig. 6d). These results establish Sox5 as a critical
190 effector responsible for ciliary elongation. Collectively, our data on Sox5, combined with the
191 established literature on Sox9, strongly support a model where the Xap5-Nono complex acts
192 through two essential downstream transcription factors, Sox5 and Sox9, to orchestrate the
193 maintenance of primary cilia.

194 **Sox5 executes a shared ciliogenic program downstream of the Xap5-Nono complex**

195 To confirm that Sox5 functions transcriptionally downstream of the Xap5-Nono complex, we
196 performed RNA-seq on *Sox5* KO and WT cells after 24 h of serum starvation. The analysis
197 identified 2,124 upregulated and 2,105 downregulated genes in *Sox5* KO cells (fold change ≥ 1.5;
198 Supplementary Fig. 7a; Supplementary Data 6). To assess the global impact on ciliary pathways,
199 we first performed Gene Set Enrichment Analysis (GSEA)[39] on all ranked differentially expressed
200 genes, which revealed a significant enrichment of the "Cilium organization" pathway (Fig. 7a).
201 Consistent with this, GO analysis of the strictly defined downregulated gene set (fold change ≥
202 1.5) also highlighted terms related to cilium assembly (Fig. 7b; Supplementary Fig. 7b).
203 Furthermore, direct comparison showed a significant overlap between the full set of genes
204 dysregulated by Sox5 loss and our curated ciliary gene list (Supplementary Fig. 7c), reinforcing
205 the strong link between Sox5 and ciliogenesis.

206 Critically, to place Sox5 within the Xap5-Nono pathway, we compared the downregulated gene

207 sets across all three KO lines. A Venn diagram revealed a significant and substantial overlap
208 among them (Fig. 7c). We confirmed this shared transcriptional dependency by qPCR, validating
209 that several key ciliary genes—including *Adcy3*, *Ccdc39*, and *Pkd2*—were indeed downregulated
210 across all three KO models (Fig. 7d). These results provide strong evidence that Sox5 acts as a key
211 transcriptional mediator downstream of Xap5 and Nono, executing a common program of ciliary
212 gene expression required for proper cilia elongation.
213

214 **Discussion**

215 The assembly of the cilium, a highly conserved organelle, requires a sophisticated and hierarchical
216 transcriptional program that is responsive to extracellular cues and precisely adapted to generate
217 enormous functional diversity[13,19-22,26]. The transcriptional program's complexity is evident in its
218 role in regulating flagella in unicellular organisms like *Chlamydomonas reinhardtii* and
219 orchestrating the synchronous formation of hundreds of motile cilia in the mammalian airway[14,40-
220 43]. In mammals, a core ciliogenic program is controlled by the Rfx family, which act as master
221 architects for the core ciliary structure[20,44-46]. Built upon this foundation are multi-tiered regulatory
222 cascades, often initiated by morphogenetic signaling pathways, that deploy cell-type-specific
223 "selector cassettes"[14,20]. For the highly specialized motile cilia, this cascade involves key initiators
224 like GMNC/MCIDAS (antagonized by GMNN)[47-50], integrates context-specific factors like
225 NOTO[51,52], and converges on downstream effectors TAp73/MYB[53-55], which in turn activate the
226 master regulator Foxj1 to execute the motile program[24,25]. While this elegant hierarchy explains
227 the formation of multiciliated cells, how upstream signaling pathways initiate the transcriptional
228 programs for the vast diversity of primary cilia remains a key unanswered question.

229 Central to this complexity is the ancient, evolutionarily conserved transcription factor Xap5.

230 Xap5 regulates flagellar assembly in *Chlamydomonas reinhardtii*, and is widely expressed in
231 mammalian tissues[27,56,57]. Our recent work revealed that during motile ciliogenesis in
232 spermatogenesis, Xap5 functions upstream of Rfx2 and Foxj1 to drive the sperm flagellar
233 program[28]. This finding solidified Xap5's role as a key regulator for motility but also sharpened
234 the central paradox addressed in this study: how does this same factor, which utilizes the
235 established motile cilia pathway in germ cells, function in somatic cells that build exclusively non-
236 motile primary cilia?

237 In this study, we resolve this fundamental paradox by uncovering a regulatory strategy that
238 enables Xap5's functional switch. In our somatic cell model, Xap5 is repurposed through
239 recruitment of the nuclear protein Nono, forming an Xap5–Nono regulatory module that promotes
240 primary ciliogenesis by directly engaging downstream Sox transcriptional effectors. This work
241 provides a molecular blueprint for how a deeply conserved master regulator can achieve functional
242 diversity through context-dependent cofactor assembly, while ultimately converging on core
243 organelle biogenesis programs.

244 Specifically, we find that the Xap5–Nono module directly co-regulates Sox5 and Sox9,
245 positioning them as downstream transcriptional effectors in primary ciliogenesis. The essential
246 role of Sox9 in promoting primary cilia formation is well-established in multiple developmental
247 contexts[31-33]. Significantly, the finding that Xap5-Nono directly co-regulates Sox9 and Sox5
248 nominates Sox5 as a previously unrecognized, critical player in primary ciliogenesis. This
249 organization exemplifies the "selector cassette" concept[20], in which context-specific deployment
250 of transcription factors modifies the execution of a conserved program; we therefore propose that
251 the context-dependent deployment of Sox5 and Sox9 downstream of Xap5–Nono functions as such
252 a cassette. This model is supported by evidence that Sox5 can cooperate with Sox9 in other

253 developmental programs (e.g., chondrogenesis[58]) and that Sox5 has been linked to ciliogenic gene
254 regulation in motile contexts[59,60]. Together, these findings position Xap5–Nono as an upstream
255 integrator that co-opts two related Sox factors to regulate primary ciliogenesis.

256 Our results suggest an evolutionary framework in which the DNA-recognition properties of
257 Xap5 function as a conserved genomic anchor, while biological outputs diversify through context-
258 dependent regulatory logic. Intriguingly, the GC-rich Xap5 binding motif identified here is nearly
259 reverse-complementary to the CT-rich C-box core (5'-CCTCC-3') that functions as a DAF-
260 19/RFX-dependent enhancer in *C. elegans* ciliome promoters[23,45,61]. Although direct binding of
261 Xap5 family members to C-box elements remains to be demonstrated, this sequence-level
262 complementarity, together with the deep conservation of the Xap5 family[27,56,57], supports the
263 hypothesis that Xap5 could contribute to C-box–like regulatory architectures in distinct lineages.
264 If so, it would be consistent with a broader model of evolutionary co-option[1-3,8-10], in which the
265 core DNA-binding interface of Xap5 remains relatively constrained while regulatory outcomes
266 may be reprogrammed via partner switching—for example, potential integration into RFX-driven
267 X-box/C-box promoter modules in nematodes[23,45,61], versus association with Nono in mammalian
268 somatic cells. This "rigid anchor, flexible cofactor" model provides a plausible route to expand
269 regulatory complexity while preserving constraints imposed by DNA-binding specificity.

270 Collectively, the discovery of this adaptable, Xap5-driven regulatory network offers new
271 perspectives on human ciliopathies[15]. Given that Sox9 dysfunction is linked to cystic diseases with
272 ciliary defects[31,32], disruption of the upstream Xap5–Nono module represents a plausible, yet
273 unexplored, pathogenic mechanism. As upstream regulators of primary ciliogenesis continue to
274 emerge—including the metazoan-specific SP5/SP8 factors[62,63]—our work highlights a distinct
275 regulatory layer rooted in an evolutionarily ancient transcription factor[27,56,57]. Notably, systemic

276 loss of Xap5 causes embryonic lethality prior to E7.5[28], indicating essential functions that likely
277 extend beyond ciliary regulation alone. One possibility is that Xap5 establishes a permissive
278 transcriptional state that enables (or constrains) the execution of ciliogenic gene programs, upon
279 which later-evolved, developmental regulators such as SP5/SP8 impose tissue- and stage-specific
280 logic[62]. Future work should define the upstream cues that govern Xap5–Nono assembly and test
281 how alternative cofactors reprogram Xap5 outputs across tissues. Mapping the context-dependent
282 partner repertoire of Xap5 will be key to explaining how an ancient regulator is repurposed to drive
283 distinct transcriptional outputs across cell types, with direct implications for development and
284 ciliopathy mechanisms.
285

286 **Materials and Methods**

287 **Antibodies**

288 The following primary antibodies were used in this study: from ABclonal Biotechnology, anti-
289 Flag (IF 1:1,000; AE005), anti-Gapdh (IB 1:10,000; AC002), anti-Nono (IB 1:1,000, IF 1:500;
290 A3800), and anti-Sox5 (IB 1:1,000; A23127); from Sigma-Aldrich, anti-Acetyl-α-Tubulin (IF
291 1:500; T7451); from Proteintech, anti-Arl13b (IF 1:500; 17711-1-AP); from HUABIO, anti-Sox9
292 (IB 1:1,000; ET1611-56); and from Prospertech, anti-Xap5 (IB 1:1,000, IF 1:500; HU-412003).
293 The following secondary antibodies were used: Alexa Fluor 488- and 594-conjugated goat anti-
294 mouse/rabbit IgG (IF 1:500; ThermoFisher Scientific); HRP-conjugated goat anti-mouse IgG (IB
295 1:2,000; Bio-Rad, #1706516) and HRP-conjugated goat anti-rabbit IgG (IB 1:2,000; Bio-Rad,
296 #1706515).

297 **Cell culture**

298 The NIH/3T3 mouse embryonic fibroblast cell line (ATCC, CRL-1658) was cultured in

299 Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% calf serum (CS) and 1%
300 penicillin-streptomycin (all from Life Technologies). Cells were maintained in a humidified
301 incubator at 37°C and 5% CO_2.

302 **Plasmid construction, transfection and generation of knockout cell lines**

303 For protein expression, the coding sequences of mouse Xap5 and Nono were amplified from
304 NIH/3T3 cDNA and cloned into the pECMV-MCS-Flag and pCMV-C-GFP vectors, respectively,
305 to generate constructs encoding C-terminally tagged proteins. Site-directed mutagenesis to delete
306 the Xap5 nuclear localization signal (NLS) was performed using the Mut Express II Fast
307 Mutagenesis Kit V2 (Vazyme, C214). For CRISPR/Cas9-mediated gene knockout, the all-in-one
308 plasmid pU6-sgRosa26-1_CBh-Cas9-T2A-BFP (a gift from Ralf Kuehn; Addgene plasmid
309 #64216) was used as the backbone. Gene-specific sgRNA sequences were inserted by replacing
310 the original sgRosa26-1 guide RNA sequence via site-directed mutagenesis. The integrity of all
311 constructs was confirmed by Sanger sequencing. All primer and sgRNA sequences are listed in
312 Supplementary Tables 1 and 2, respectively.

313 For transient expression experiments, cells were transfected with the indicated DNA constructs
314 using Lipofectamine 3000 (Life Technologies) according to the manufacturer's protocol.

315 Stable knockout cell lines were generated by electroporation. Briefly, 1×10^6 NIH/3T3 cells were
316 resuspended in 250 μL of Opti-MEM I Reduced Serum Medium (Gibco) containing 20 μg of the
317 appropriate all-in-one CRISPR-Cas9 plasmids. The cell suspension was transferred to a 0.4 cm
318 gap electroporation cuvette (Bio-Rad) and electroporated using a Gene Pulser Xcell
319 Electroporation System (Bio-Rad) with a single exponential decay pulse (250 V, 950 μF, ∞ Ω
320 resistance). Twenty-four hours post-electroporation, single cells exhibiting bright BFP
321 fluorescence were identified by fluorescence microscopy and manually isolated. Individual cells

322 were seeded into separate wells of a 96-well plate to grow into monoclonal colonies, which were
323 subsequently expanded and validated for successful gene knockout.

324 **Immunofluorescence and microscopy**

325 To induce primary cilia formation, NIH/3T3 cells were seeded at low density (< 30% confluency)
326 onto sterile, poly-D-lysine-coated (Gibco) glass coverslips (CITOTEST). Following 24 h of
327 growth in standard medium, ciliogenesis was induced by switching to serum-free DMEM for an
328 additional 24–48 h.

329 Cells on coverslips were washed three times with PBS and fixed with 4% paraformaldehyde
330 (PFA) in PBS for 10 min at room temperature. Following fixation, residual PFA was quenched
331 with 50 mM NH_4Cl in PBS for 5 min. Cells were then permeabilized with 0.2% Triton X-100 for
332 15 min and blocked with 3% bovine serum albumin (BSA) in PBS for 1 h. Primary antibody
333 incubations were performed for 1 h at room temperature in a humidified chamber. After washing,
334 cells were incubated with Alexa Fluor-conjugated secondary antibodies for 1 h, followed by a 5
335 min counterstain with DAPI (Sigma-Aldrich). Finally, coverslips were washed and mounted onto
336 glass slides using Fluoromount-G mounting medium (SouthernBiotech). This protocol was used
337 for all cilia detection and Xap5 ectopic expression experiments. For the detection of ectopically
338 expressed Nono-GFP, cells were fixed and washed as above, then directly counterstained with
339 DAPI before mounting, omitting the permeabilization and antibody incubation steps.

340 General fluorescence for confirming ectopic protein expression was assessed using a Zeiss Axio
341 Vert. A1 inverted microscope. High-resolution imaging of primary cilia was performed on a Leica
342 SP8 confocal microscope using a 63× oil-immersion objective. The percentage of ciliated cells
343 and cilia length were quantified from confocal images using the Automated Cilia Detection and
344 Counter (ACDC) software (v0.93)[64]. The nuclear area was quantified from DAPI-stained images

345 using Fiji software.

346 **RNA isolation and RT-qPCR**

347 Total RNA was isolated from cells using the FastPure Cell/Tissue Total RNA Isolation Kit
348 (Vazyme, RC112) according to the manufacturer's protocol. For each sample, 500 ng of total RNA
349 was reverse-transcribed into cDNA using the HiScript II 1st Strand cDNA Synthesis Kit (Vazyme,
350 R212). RT-qPCR was then performed with ChamQ SYBR qPCR Master Mix (Vazyme, Q311) on
351 an Applied Biosystems StepOnePlus Real-Time PCR Systems. Relative gene expression was
352 calculated using the $2^{-\Delta\Delta Ct}$ method, with *Gapdh* serving as the internal normalization control. All
353 primer sequences are listed in Supplementary Table 1.

354 **Western blotting**

355 Cells were lysed on ice in RIPA buffer (Beyotime, P0013B) supplemented with 1× protease
356 inhibitor cocktail (Roche, 11873580001) and 1 mM PMSF (Sigma, 93482). The resulting lysates
357 were resolved by SDS-PAGE and transferred to polyvinylidene fluoride membranes (Millipore,
358 IPVH00010) using a wet transfer system (Tanon). Membranes were blocked with 5% non-fat milk
359 in TBST (20 mM Tris-HCl, pH 7.5, 150 mM NaCl, 0.1% Tween-20) for 1 h at room temperature,
360 followed by incubation with primary antibodies overnight at 4 °C. After washing, membranes were
361 incubated with HRP-conjugated secondary antibodies for 1 h at room temperature.
362 Chemiluminescence signals were developed using the ECL Super Kit (ABClonal, RM02867) and
363 visualized with a Tanon 5200 Chemiluminescence Imaging System.

364 **Co-immunoprecipitation (Co-IP) and mass spectrometry**

365 For Co-IP and IP-MS experiments, cultured cells were washed twice with cold 1× PBS and
366 harvested by scraping. Total proteins were extracted on ice using a Co-IP buffer containing 50 mM
367 HEPES (pH 7.5), 200 mM KCl, 1 mM EGTA, 1 mM $MgCl_2$, 10% (v/v) glycerol, 0.5 mM DTT,

368 0.3% (v/v) NP-40, and 1× protease inhibitor cocktail (Roche). For immunoprecipitation,
369 antibodies were first crosslinked to Protein A/G magnetic beads (Invitrogen, 80105G) using 5 mM
370 BS^3. The resulting antibody-conjugated beads were then incubated with the cell lysates for 2 h at
371 4°C with gentle rotation. After extensive washing, bound proteins were eluted with 0.1 M Glycine-
372 HCl (pH 2.5). For western blot analysis, the eluates and input samples were neutralized, mixed
373 with SDS loading buffer, boiled for 5 min, and subsequently analyzed by immunoblotting. For
374 protein identification, the eluates were sent for analysis by liquid chromatography with tandem
375 mass spectrometry (LC-MS/MS) at Novogene (Beijing, China).

376 **RNA-sequencing and analysis**

377 For transcriptomic analysis, cells were plated for 24 h and subsequently serum-starved for 24 h.
378 At the end of the starvation period, cells were immediately washed three times with 1× PBS and
379 then lysed directly in TRIzol Reagent (Invitrogen) for total RNA extraction. Library construction
380 and paired-end (2 × 150 bp) sequencing on an Illumina NovaSeq 6000 platform were performed
381 by BioMarker Technologies (Beijing, China).

382 The raw sequencing reads were mapped to the Mus musculus reference genome
383 (GRCm39/mm39) using HISAT2[65] (v2.1.0). StringTie[66] (v1.3.3b) was subsequently used to
384 assemble the mapped reads. Gene expression levels were quantified as fragments per kilobase of
385 transcript per million fragments mapped (FPKM). Differential expression analysis between
386 experimental groups was performed using DESeq2[67] (v1.12.4). Genes with a false discovery rate
387 (FDR) < 0.01 and an absolute fold change ≥ 1.5 were considered significantly differentially
388 expressed. Gene Ontology (GO) enrichment analysis was performed using the DAVID database.
389 Gene Set Enrichment Analysis (GSEA) was conducted using the OECloud platform
390 (https://cloud.oebiotech.com).

391 **CUT&Tag-sequencing and analysis**

392 For CUT&Tag analysis, NIH/3T3 cells were plated for 24 h and subsequently serum-starved for
393 24 h. Following harvesting, 100,000 cells were collected for each sample. Library construction
394 was performed using the Hyperactive Universal CUT&Tag Assay Kit for Illumina Pro (Vazyme,
395 TD904) according to the manufacturer's protocol. The resulting libraries were sequenced on an
396 Illumina NovaSeq platform (150 bp paired-end reads) at Novogene Co., Ltd. (Beijing, China).

397 For bioinformatic processing, raw reads were assessed for quality with FastQC[68] (v0.11.9) and
398 trimmed with fastp[69] (v0.20.0; parameters: --length_required 15, --n_base_limit 6). Clean reads
399 were aligned to the Mus musculus reference genome (mm39) using BWA-MEM (v0.7.12;
400 parameters: -k 32, -T 30, -M). Peak calling was performed using MACS2[70] (v2.1.0) with a q-value
401 threshold of 0.05 and the following parameters: -f AUTO --call-summits --nomodel --shift -100 -
402 -extsize 200 --keep-dup all. For *de novo* motif discovery, peak summits were extended to 500 bp
403 and analyzed using the HOMER[71] (v4.9.1) findMotifsGenome.pl script. Peak annotation and
404 association with the nearest genes were performed using the ChIPseeker[72] R package. Genome
405 browser tracks were generated using the Integrative Genomics Viewer (IGV, v2.18.1)[73].

406 **CUT&Tag-qPCR**

407 For targeted validation, DNA from 100,000 cells was prepared in parallel using the same
408 CUT&Tag kit according to the manufacturer's protocol. qPCR was performed using the resulting
409 DNA. Immunoprecipitation with IgG sera served as a negative control, and an exogenous DNA
410 spike-in was used as an internal normalization control. Enrichment at specific loci was calculated
411 relative to the spike-in DNA control, and the $2^{-\Delta\Delta Ct}$ method was used to compare experimental
412 samples. All primer sequences used for CUT&Tag-qPCR are listed in Supplementary Table 1.

413 **Statistical analysis**

Statistical analysis was performed using GraphPad Prism v6.01 or v8.0.2. All data were presented as mean ± SEM. Values of $P < 0.05$ were considered statistically significant. Statistical significance between two groups was calculated using an unpaired, parametric, two-sided Student's t-test.

Data availability

The RNA-sequencing data generated in this study have been deposited in the Gene Expression Omnibus (GEO) database under accession code GSE305355. The CUT&Tag-sequencing data reported in this paper are available in the GEO database under accession code GSE304980. All other data supporting the findings of this study are available within the article and its Supplementary Information files. Source data are provided with this paper.

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Author Contributions

W.W., X.Z., Y.Q., X.M., G.T. and Z.H. conceived and designed the research. W.W., X.Z., Y.Q., X.M., S.C., Y.C., S.L., W.C., J.Y., X.Y., H.L., J.X., C.X. and H.J. performed the research. W.W., G.T. and Z.H. contributed new reagents/analytic tools. W.W., H.W., G.T. and Z.H. analyzed the data. W.W. and Z.H. wrote the manuscript. All authors reviewed the manuscript and approved the final version.

637 **Competing interests**

638 The authors declare no competing interests.

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640 **Supplementary information**

641 **Supplementary Data 1**

642 Differentially expressed genes between WT and *Xap5* KO identified by RNA-seq

643 **Supplementary Data 2**

644 Differentially expressed genes between WT and *Nono* KO identified by RNA-seq

645 **Supplementary Data 3**

646 Curated list of cilia-associated genes used in the study

647 **Supplementary Data 4**

648 List of Xap5-bound genes identified by CUT&Tag analysis

649 **Supplementary Data 5**

650 List of Nono-bound genes identified by CUT&Tag analysis

651 **Supplementary Data 6**

652 Differentially expressed genes between WT and *Sox5* KO identified by RNA-seq

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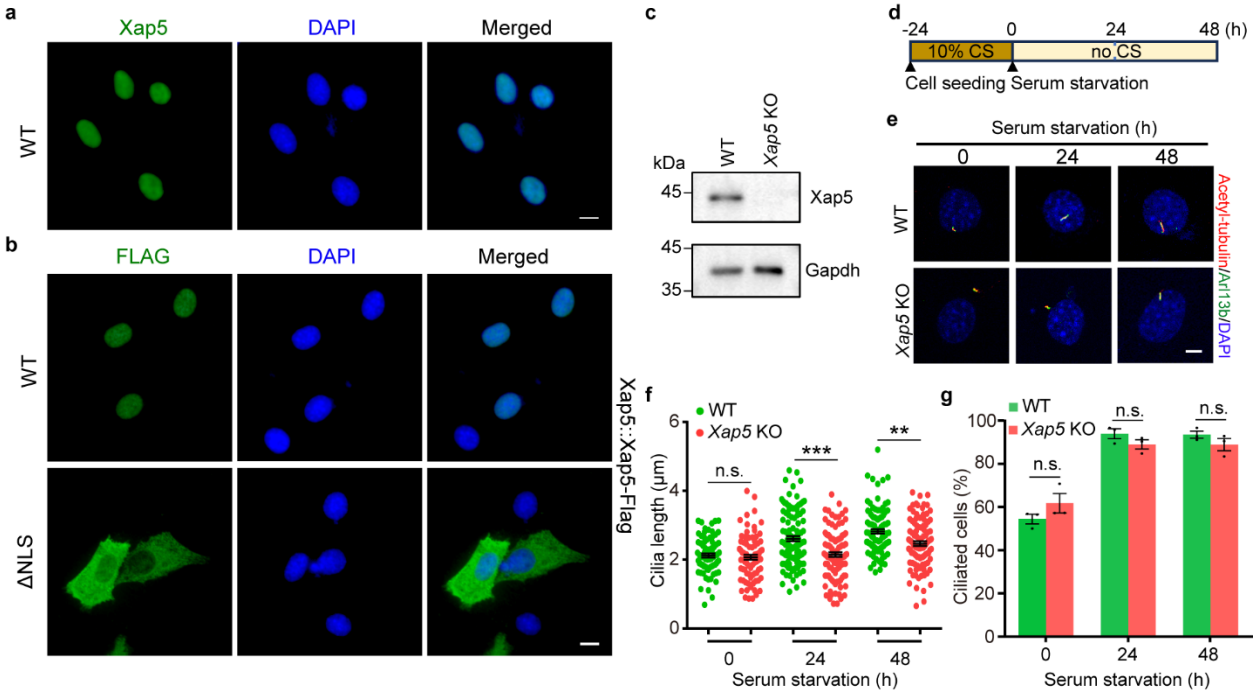
654 **Figures**



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656 **Fig. 1** Xap5 is a nuclear protein essential for primary ciliogenesis. **a** Immunofluorescence analysis
657 of endogenous Xap5 (green) in NIH/3T3 cells. Nuclei were counterstained with DAPI (blue). Scale
658 bar, 10 μm. **b** Validation of the Xap5 nuclear localization signal (NLS). Representative images of
659 NIH/3T3 cells expressing either Flag-tagged wild-type Xap5 (WT) or an NLS-deleted mutant
660 (ΔNLS) (green). Scale bar, 10 μm. **c** Western blot analysis confirming the complete depletion of
661 Xap5 protein in the *Xap5* knockout (KO) clonal cell line. Gapdh serves as a loading control. The
662 blot shown is representative of three independent biological replicates. **d** Schematic illustrating the
663 time-course assay for primary cilia assembly induced by serum starvation. CS, calf serum. **e**
664 Representative immunofluorescence images of cilia from WT and *Xap5* KO cells at the indicated
665 time points. Cilia were co-stained for acetylated tubulin (red) and Arl13b (green). Nuclei were
666 counterstained with DAPI (blue). Scale bar, 5 μm. **f** Quantification of primary cilia length in WT
667 and *Xap5* KO cells over a 48-h time course of serum starvation. Data are presented as means ±
668 SEM (n = 3 independent experiments). **g** Quantification of the percentage of ciliated cells as in **f**.

669 Data are presented as mean ± SEM (n = 3 independent experiments). All statistical significance
670 between WT and KO at each time point was determined by multiple two-tailed unpaired t-tests.
671 n.s., not significant; $^{**}P < 0.01$; $^{***}P < 0.001$. Source data are provided as a Source Data file.
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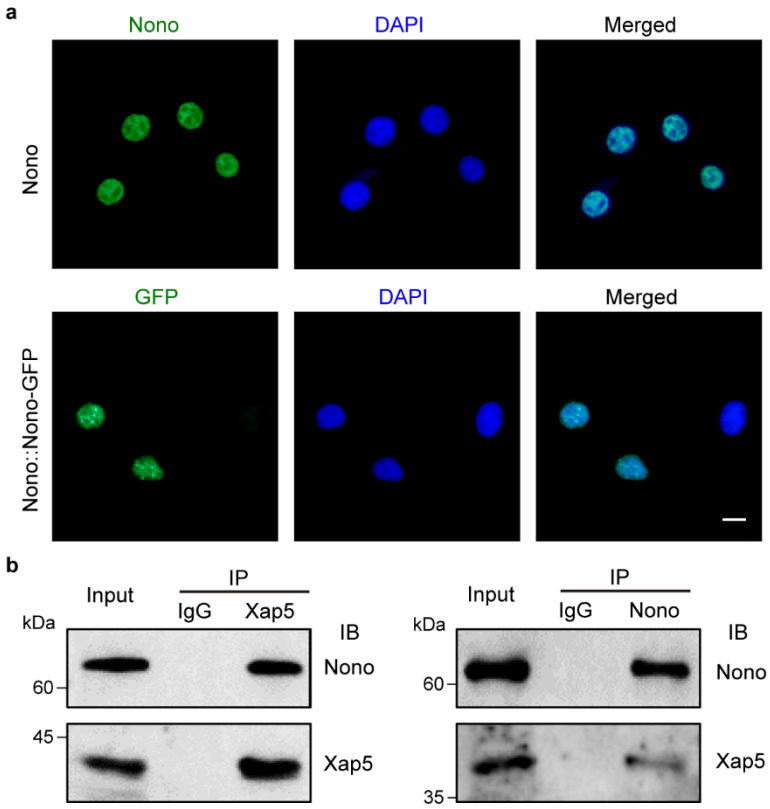

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684 **Fig. 2** Xap5 forms a nuclear complex with its cofactor Nono. **a** Representative
685 immunofluorescence images showing the predominantly nuclear localization of Nono in NIH/3T3
686 cells. Top: Endogenous Nono detected with a specific anti-Nono antibody (green). Bottom:
687 Ectopically expressed Nono-GFP fusion protein (green). Nuclei were counterstained with DAPI
688 (blue). Scale bars, 10 µm. **b** Endogenous co-immunoprecipitation (Co-IP) of Xap5 and Nono from
689 NIH/3T3 cell lysates. Left: Immunoprecipitation (IP) with an anti-Xap5 antibody specifically co-
690 precipitated endogenous Nono. Right: Conversely, IP with an anti-Nono antibody co-precipitated
691 endogenous Xap5. Control IgG was used as a negative control for the IP. The resulting precipitates
692 (IP) and input lysates were analyzed by western blotting (IB) with the indicated antibodies. Data
693 are representative of three independent experiments. Source data are provided as a Source Data
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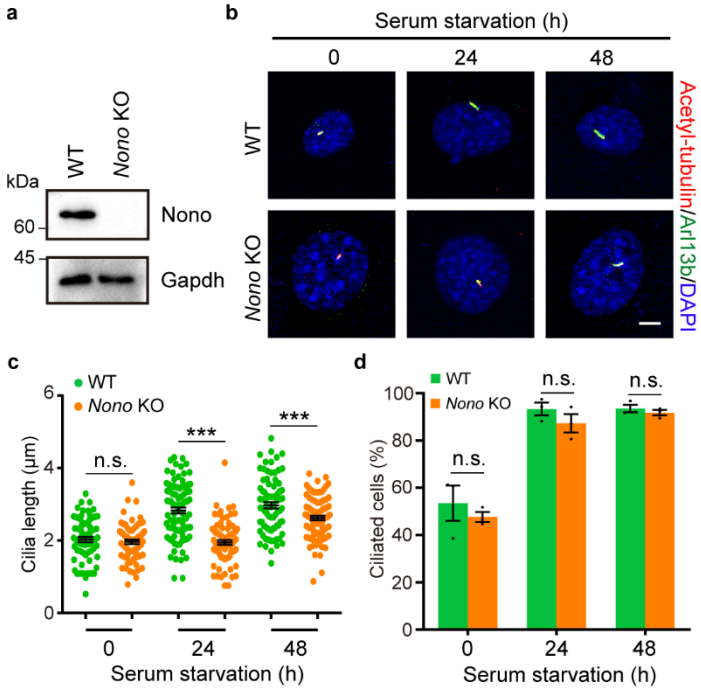

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697 **Fig. 3** Loss of Nono phenocopies Xap5 depletion, impairing primary cilia elongation. **a** Western
698 blot analysis confirming the complete depletion of Nono protein in *Nono* KO cell lysates compared
699 to WT. The blot shown is representative of three independent biological replicates. Gapdh serves
700 as a loading control. **b** Representative immunofluorescence images of primary cilia in WT and
701 *Nono* KO cells after 48 h of serum starvation. Cilia were co-stained for acetyl-tubulin (red) and
702 Arl13b (green). Nuclei were counterstained with DAPI (blue). Scale bar, 5 μm. **c** Quantification
703 of primary cilia length in WT and *Nono* KO cells over a 48-h time course of serum starvation. Data
704 are presented as mean ± SEM (n = 3 independent experiments). **d** Quantification of the percentage
705 of ciliated cells as in **c**. Data are presented as mean ± SEM (n = 3 independent experiments). All
706 statistical significance between WT and KO at each time point was determined by multiple two-
707 tailed unpaired t-tests. n.s., not significant; $^{**}P < 0.01$; $^{***}P < 0.001$. Source data are provided as
708 a Source Data file.

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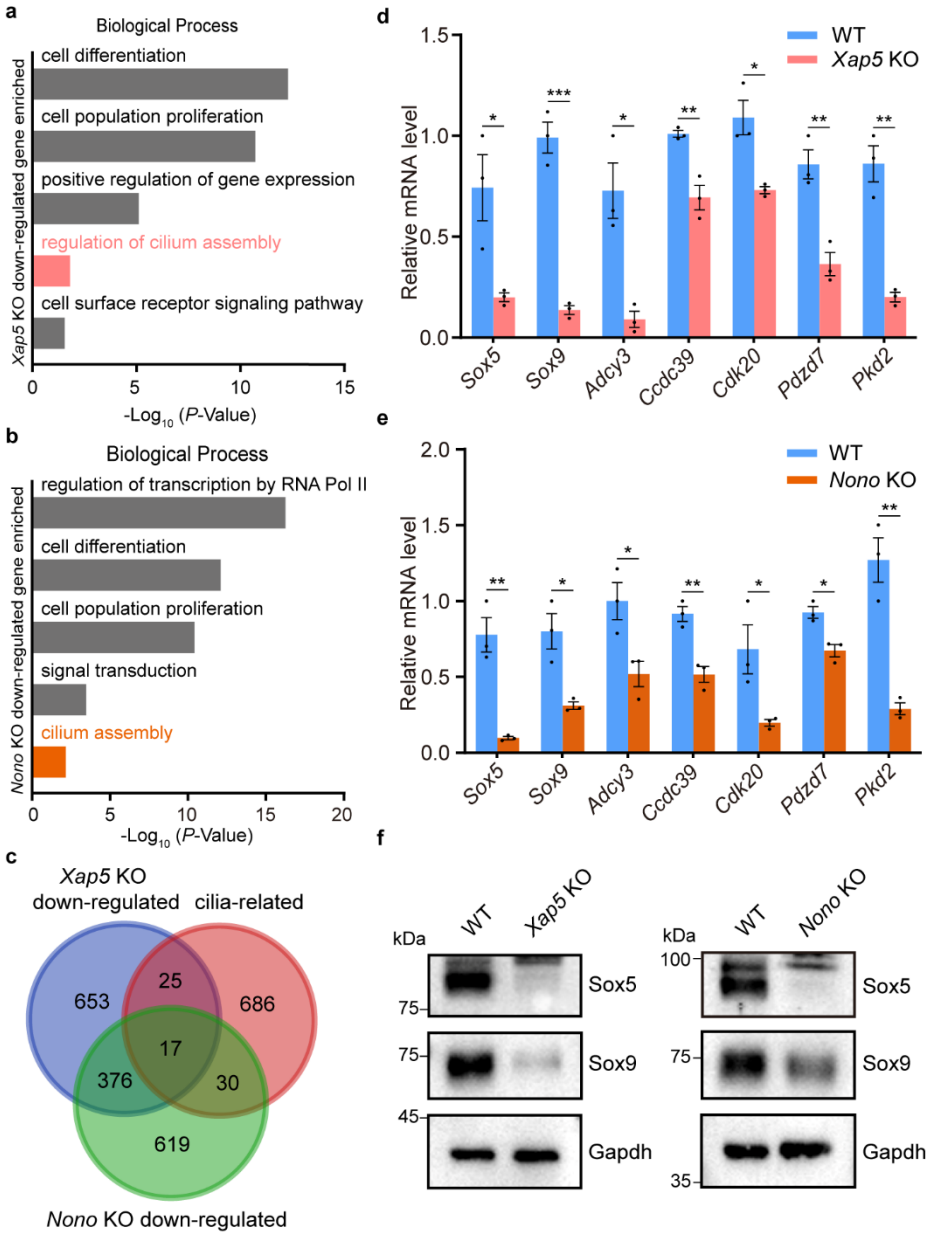

Fig. 4 Xap5 and Nono co-regulate a ciliogenic transcriptional network. All analyses were performed on NIH/3T3 cells following 24 h of serum starvation. **a, b** Representative enriched Gene Ontology (GO) terms (Biological Process) for genes downregulated in *Xap5* KO (**a**) and *Nono* KO (**b**) cells. **c** Venn diagram illustrating the significant overlap between genes downregulated in *Xap5* KO cells, *Nono* KO cells, and a curated list of cilia-associated genes (see also Supplementary Data 3). **d, e** qPCR validation of shared, downregulated ciliary target genes in *Xap5* KO (**d**) and *Nono*

717 KO (**e**) cells. Data are presented as mean ± SEM from three independent experiments. *P* values
718 were determined by two-tailed unpaired t-test. **P* < 0.05, ***P* < 0.01, ****P* < 0.001. **f** Representative
719 western blots showing reduced protein levels of Sox5 and Sox9 in *Xap5* KO (left) and *Nono* KO
720 (right) cells compared to their respective WT controls. Gapdh serves as a loading control. Blots
721 are representative of three independent experiments. Source data are provided as a Source Data
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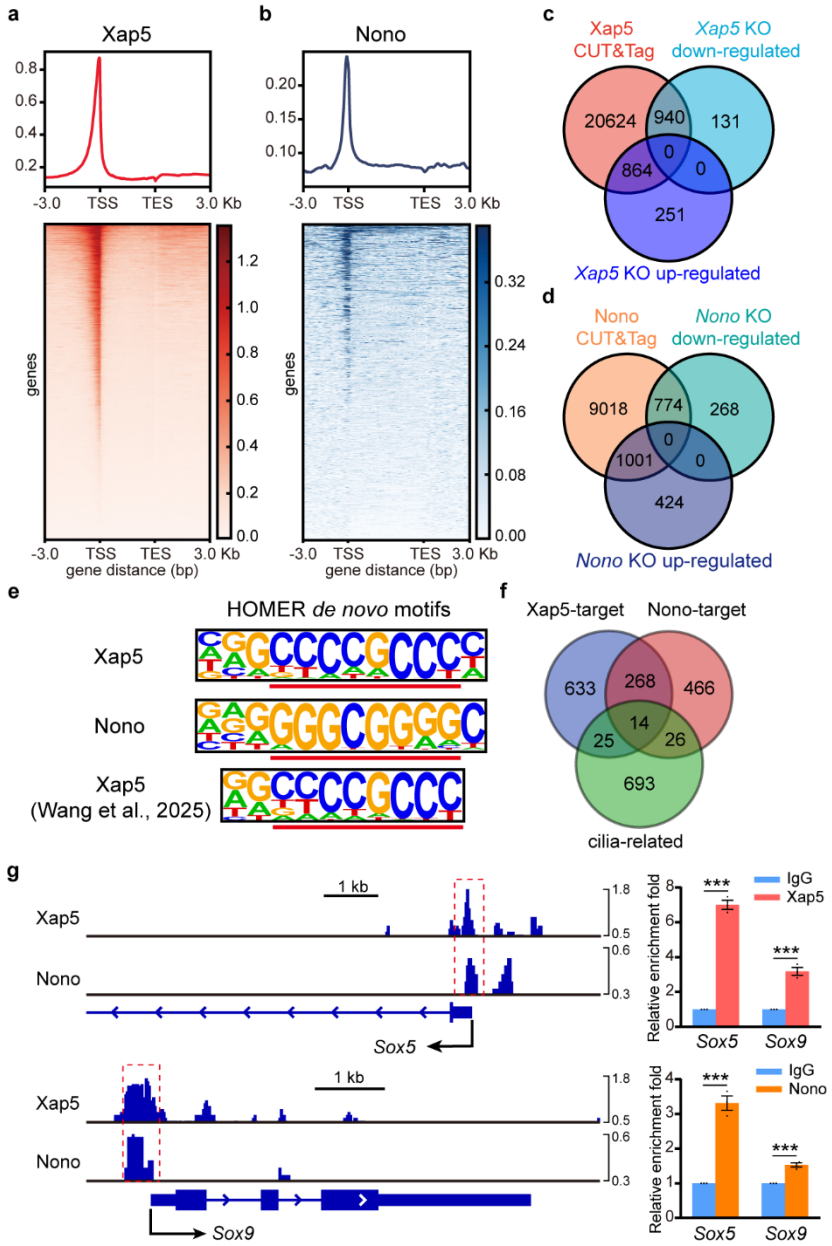


Fig. 5 Xap5 and Nono directly co-occupy the regulatory regions of ciliary genes. **a, b** Genome-wide binding profiles of Xap5 (**a**) and Nono (**b**) in NIH/3T3 cells determined by CUT&Tag. Top panels show the average enrichment profiles over gene bodies (from transcription start site, TSS, to transcription end site, TES). Bottom panels show heatmaps of binding signals centered on the TSS (±3 kb). **c, d** Venn diagrams illustrating the significant overlap between genes bound by Xap5 (**c**) or Nono (**d**) and the respective sets of differentially expressed genes identified upon their

knockout. **e** Comparison of *de novo* DNA-binding motifs derived from Xap5 and Nono CUT&Tag peaks in this study, alongside the Xap5 motif from our previous work in testicular germ cells[28]. The high degree of similarity indicates a conserved DNA-binding preference across different cellular contexts. **f** Venn diagram showing the overlap between direct Xap5 target genes (defined as genes that are both bound by Xap5 and downregulated in *Xap5* KO), direct Nono target genes, and the curated ciliary gene set (from Supplementary Data 3). **g** Direct co-occupancy of Xap5 and Nono at the *Sox5* and *Sox9* gene loci. Left: Integrative Genomics Viewer (IGV) tracks showing CUT&Tag signal enrichment. Right: CUT&Tag-qPCR validating the co-enrichment of Xap5 and Nono at the *Sox5* and *Sox9* promoters. Red dashed box: Xap5 and Nono binding regions. Data are presented as mean ± SEM from three independent experiments. *P* values were determined by two-tailed unpaired t-test. ***$P < 0.001$. Source data are provided as a Source Data file.

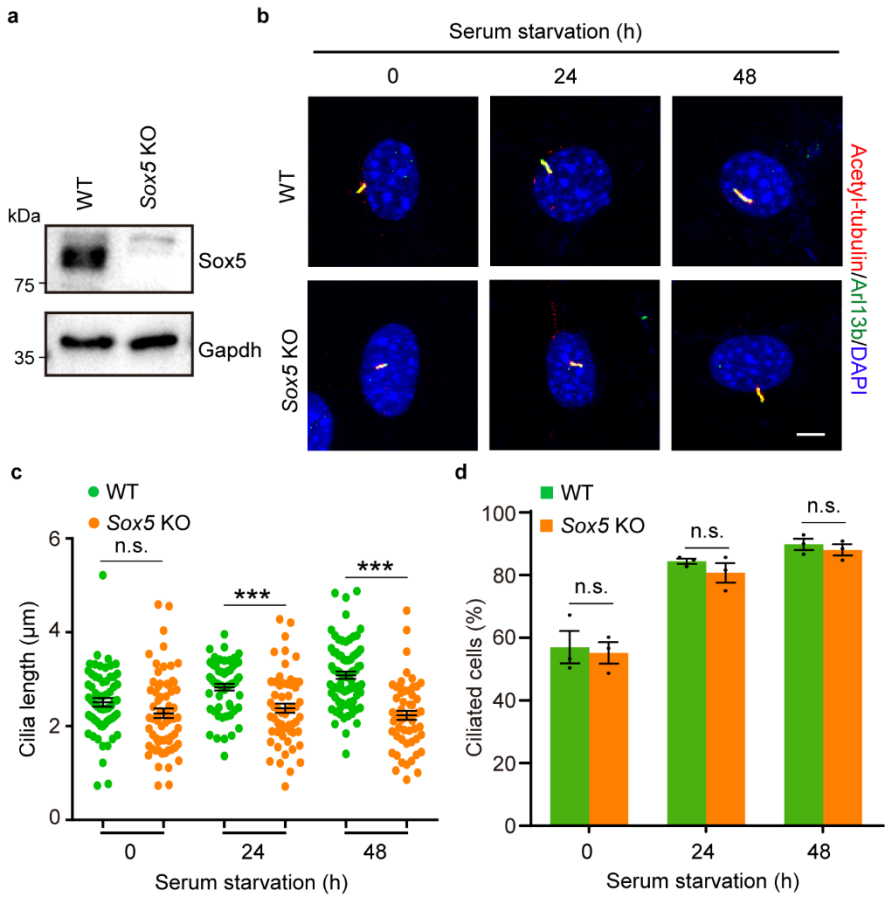


Fig. 6 Downstream effector Sox5 is essential for primary ciliogenesis. **a** Western blot analysis confirming the complete depletion of Sox5 protein in *Sox5* KO cell lysates compared to WT. The blot shown is representative of three independent biological replicates. Gapdh serves as a loading control. **b** Representative immunofluorescence images of primary cilia in WT and *Sox5* KO cells after serum starvation. Cilia were co-stained for acetylated tubulin (red) and Arl13b (green). Nuclei were counterstained with DAPI (blue). Scale bar, 5 μm. **c** Quantification of primary cilia length in WT and *Sox5* KO cells over a 48-h time course of serum starvation. Data are presented as mean ± SEM (n = 3 independent experiments). **d** Quantification of the percentage of ciliated cells as in **c**. Data are presented as mean ± SEM from three independent experiments. *P* values were determined by two-tailed unpaired t-test. n.s., not significant; ***$P < 0.001$. Source data are provided as a Source Data file.

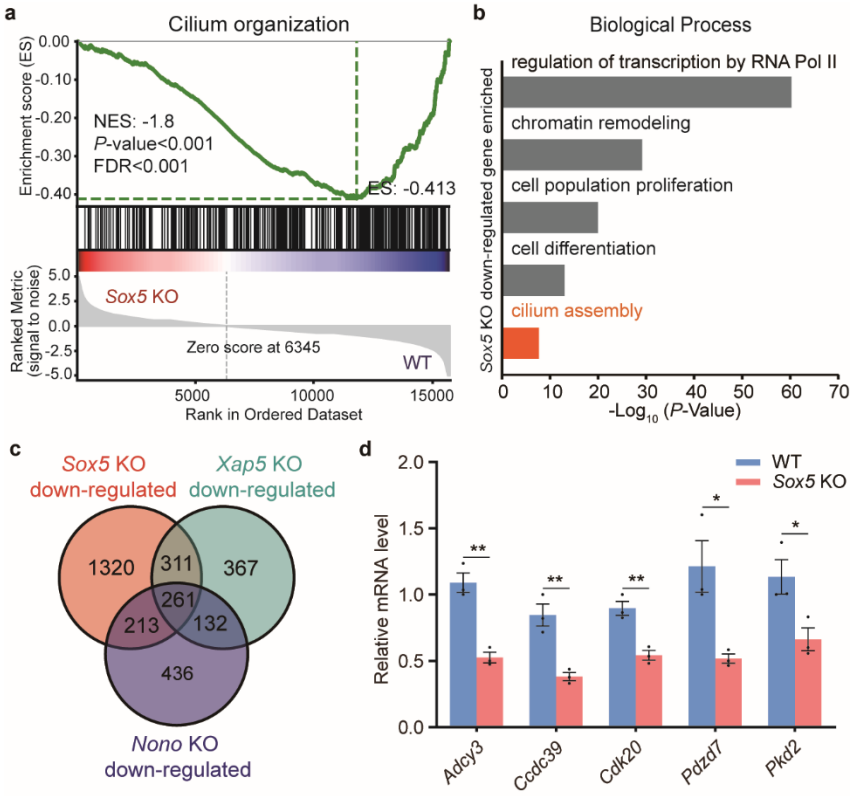
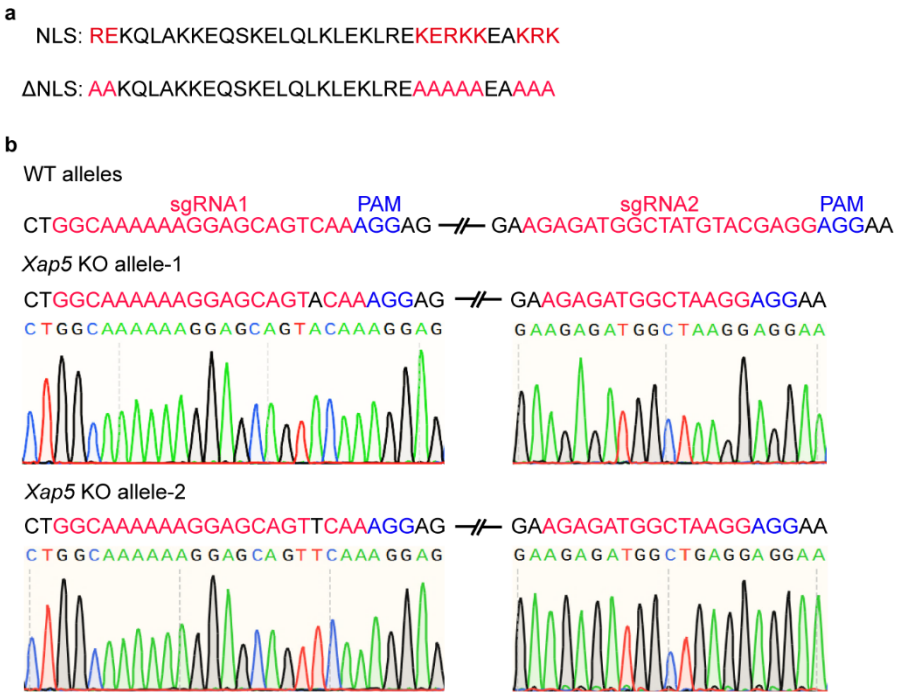


Fig. 7 Sox5 executes a ciliogenic transcriptional program downstream of the Xap5-Nono complex. **a** Gene Set Enrichment Analysis (GSEA) plot for the "cilium organization" pathway, performed on all ranked differentially expressed genes from the *Sox5* KO transcriptome. **b** Representative enriched GO terms (Biological Process) for genes significantly downregulated (fold change ≥ 1.5) in *Sox5* KO cells. **c** Venn diagram illustrating the significant overlap among genes downregulated in *Xap5* KO, *Nono* KO, and *Sox5* KO cells, identifying a common set of co-regulated genes. **d** qPCR validation confirming the reduced expression of shared downstream target genes in *Sox5* KO cells compared to WT. Data are presented as mean ± SEM from three independent experiments. *P* values were determined by two-tailed unpaired t-test. $^{*}P < 0.05$, $^{**}P < 0.01$. Source data are provided as a Source Data file.

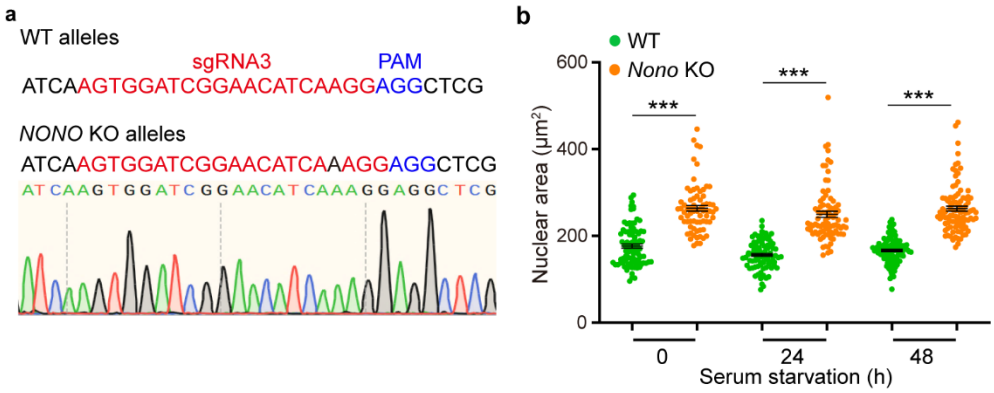
Supplementary Figures



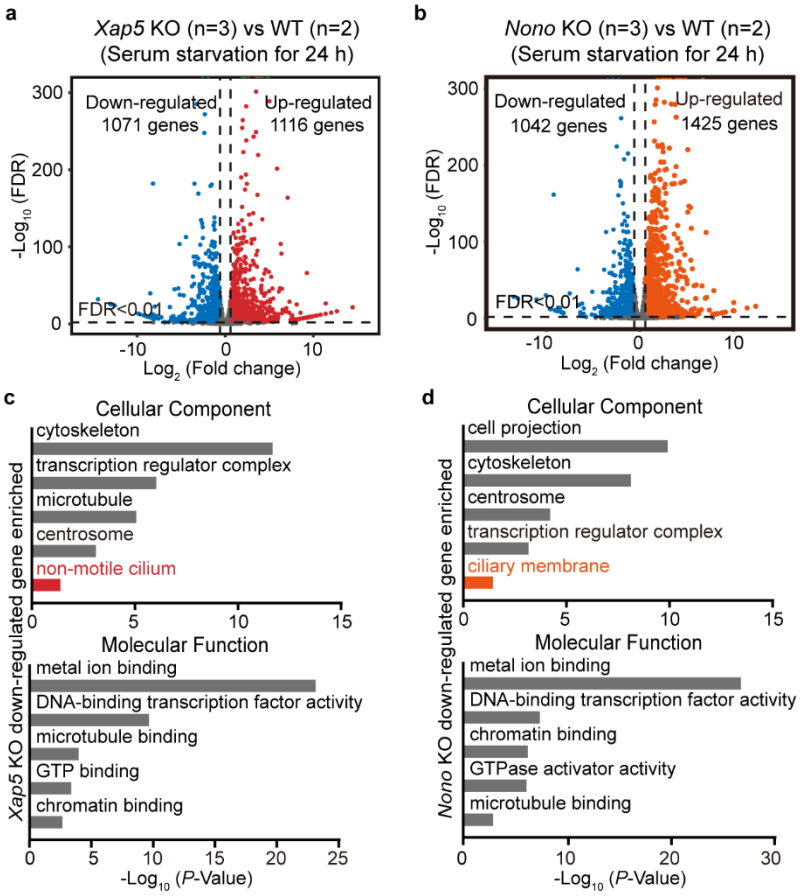
Supplementary Fig. 1 Design and validation of NLS-mutant and CRISPR/Cas9-mediated *Xap5* KO alleles. **a** Amino acid sequences of the wild-type nuclear localization signal (NLS) in Xap5 and the NLS-deleted mutant (ΔNLS) generated for this study. Key basic residues targeted for mutation are highlighted in red. **b** Validation of CRISPR/Cas9-mediated knockout of *Xap5*. Top: Schematic of the wild-type genomic locus showing the target sites for the two guide RNAs (sgRNA1 and sgRNA2, red) and their adjacent PAM sequences (blue). Bottom: Sanger sequencing chromatograms from a selected knockout clone revealing two distinct mutant alleles (allele-1 and allele-2). Both alleles share an identical mutation at the sgRNA2 target site but harbor different frameshift-inducing mutations at the sgRNA1 site, consistent with independent non-homologous end joining (NHEJ) repair events at this locus.

Accession	Description	Gene names
P62806	Histone H4	*Hist1h4a*
Q3TF40	Uncharacterized protein (Fragment)	*Nono*
Q9WV03	Protein Xap5	*Xap5*
P27661	Histone H2ax	*H2afx*
Q921L4	Histone H2b	*LOC665622*
P97350	Plakophilin-1	*Pkp1*
Q8VIJ6	Splicing factor, proline-and glutamine-rich	*Sfpq*
G3UXW9	COP9 signalosome complex subunit 1	*Gps1*

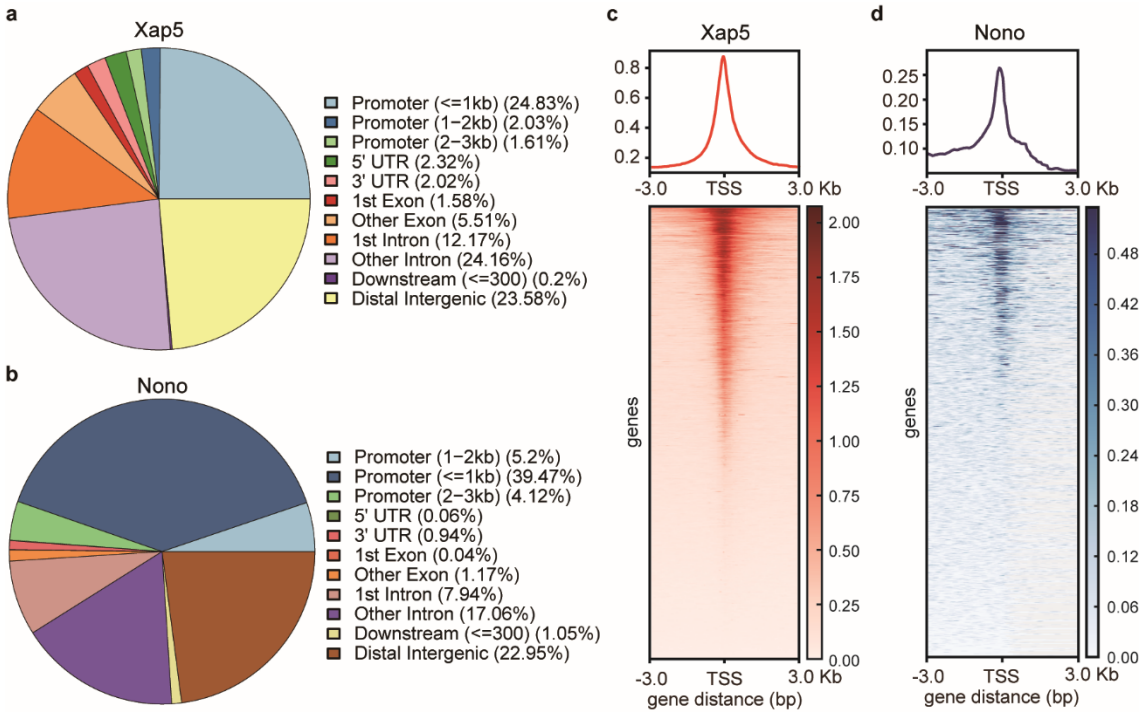
Supplementary Fig. 2 Identification of Xap5-interacting nuclear proteins by mass spectrometry. Table showing selected high-confidence nuclear proteins identified by immunoprecipitation (IP) of endogenous Xap5 from NIH/3T3 cell lysates followed by mass spectrometry (IP-MS). Notable hits include core histones (e.g., Hist1h4a, H2afx) and key RNA-binding proteins such as Nono and Sfpq, suggesting Xap5 may function in complexes involved in chromatin regulation and RNA processing.



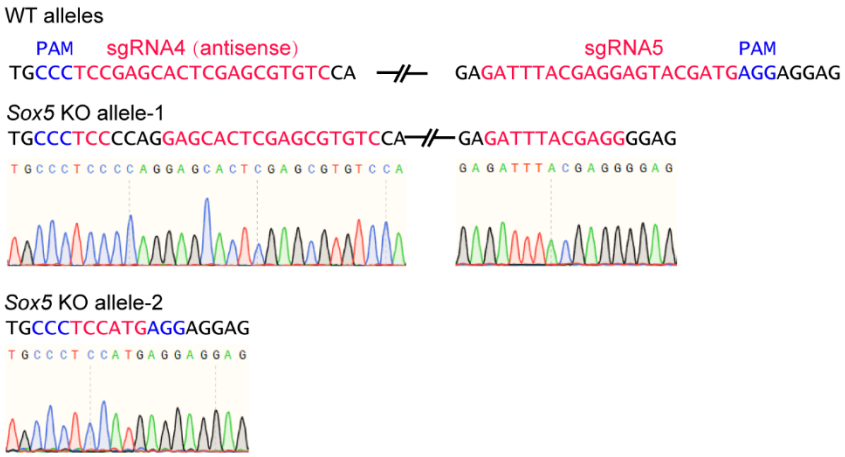
Supplementary Fig. 3 Validation of *Nono* KO and its impact on nuclear architecture. **a** Sanger sequencing chromatogram validating the CRISPR/Cas9-mediated KO of the *Nono* gene in a selected NIH/3T3 clone. The WT genomic sequence, sgRNA3 target site (red), and PAM sequence (blue) are indicated above. The resulting KO allele harbors a frameshift-inducing indel at the target site, consistent with successful gene disruption. **b** Quantification of nuclear area (μm²) in WT and *Nono* KO cells over a 48-h time course of serum starvation. Data are presented as the mean ± SEM (n = 3, independent biological replicates). Statistical significance between WT and KO cells at each time point was determined by multiple two-tailed unpaired t-tests. ***$P < 0.001$. Source data are provided as a Source Data file.



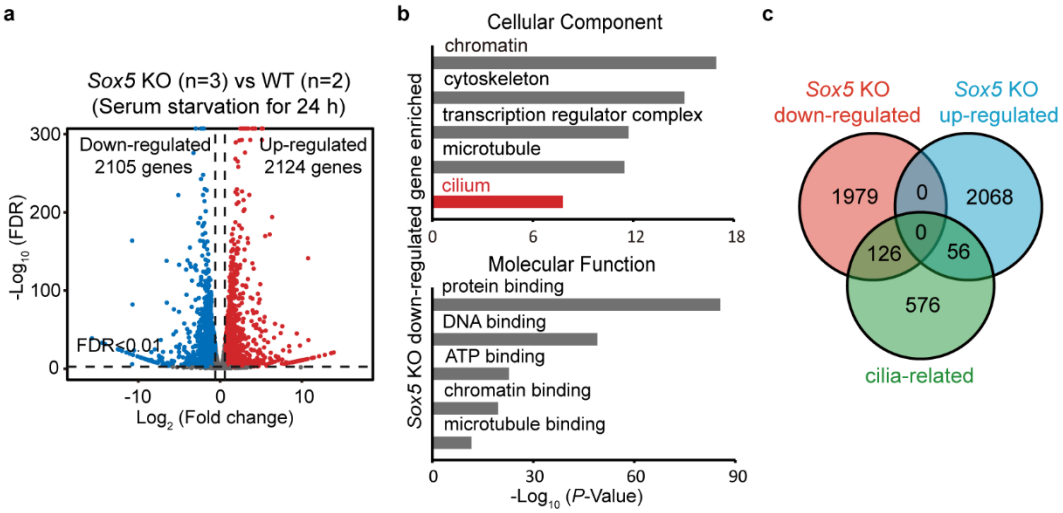
Supplementary Fig. 4 Transcriptomic profiling of *Xap5* and *Nono* KO cells. All analyses were performed on cells following 24 h of serum starvation. **a** Volcano plot of differentially expressed genes in *Xap5* KO (n=3 biological replicates) versus WT (n=2) cells. Using cutoffs of absolute fold change ≥ 1.5 and a false discovery rate (FDR) < 0.01 (highlighted dots), we identified 1,116 upregulated and 1,071 downregulated genes. **b** Volcano plot for *Nono* KO (n=3) versus WT (n=2) cells, analyzed using the same conditions and cutoffs as in **a**. This analysis identified 1,425 upregulated and 1,042 downregulated genes. **c**, **d** Representative enriched Gene Ontology (GO) terms for Cellular Component and Molecular Function categories among genes downregulated in *Xap5* KO (**c**) and *Nono* KO (**d**) cells. Cilia- and transcription-related terms are prominent in both datasets, reinforcing their dual roles in ciliogenesis and nuclear function.



Supplementary Fig. 5 Genomic distribution and metagene profiles of Xap5 and Nono CUT&Tag peaks. **a**, **b** Pie charts showing the genomic distribution of CUT&Tag peaks for Xap5 (**a**) and Nono (**b**). A substantial fraction of peaks for both Xap5 and Nono are located in promoter regions (defined as ≤3 kb upstream of a TSS). **c**, **d** Metagene plots (top) and heatmaps (bottom) showing the enrichment of Xap5 (**c**) and Nono (**d**) CUT&Tag signals centered on transcription start sites (TSS ±3 kb). The strong signal enrichment at the TSS is consistent with their roles as direct transcriptional regulators.



Supplementary Fig. 6 Validation of *Sox5* KO alleles by Sanger sequencing. Sanger sequencing chromatograms confirming CRISPR/Cas9-mediated deletion of *Sox5*. The wild-type genomic sequence, positions of the sgRNA4 (antisense strand) and sgRNA5 target sites (red), and the PAM sequences (blue) are indicated. Two independent *Sox5* KO alleles (KO allele-1 and KO allele-2) from the same clone display distinct mutations between the sgRNA cut sites. KO allele-1 harbors distinct frameshift-inducing mutations at each of the two sgRNA target sites, while KO allele-2 contains a large deletion spanning the entire region between the sgRNA sites. These results confirm successful biallelic disruption of the *Sox5* locus.



Supplementary Fig. 7 Transcriptomic analysis of *Sox5* KO cells. All analyses were performed on cells following 24 h of serum starvation. **a** Volcano plot of differentially expressed genes in *Sox5* KO (n=3 biological replicates) versus WT (n=2) cells. Using cutoffs of absolute fold change ≥ 1.5 and FDR < 0.01 (highlighted dots), we identified 2,124 upregulated and 2,105 downregulated genes. **b** Representative enriched GO terms for Cellular Component and Molecular Function categories among genes downregulated in *Sox5* KO cells. The significant enrichment of cilia-related terms reinforces the link between Sox5 and ciliogenesis. **c** Venn diagram illustrating the overlap between the curated ciliary gene set (from Supplementary Data 3) and genes either upregulated or downregulated in *Sox5* KO cells. The analysis reveals that a total of 182 ciliary genes were dysregulated, consisting of 126 downregulated and 56 upregulated genes, indicating a broad impact of Sox5 on the ciliary transcriptome.

910 **Supplementary Table 1. Primers used in the study**

Name	Sequences (5' to 3')	Usage
Xap5-Flag-F	GGGAATTCGTCGACTGGATCCATGGCCCAGTACAAGGGCG	Cloning primer
Xap5-Flag-R	TGAGATGAGTTTCTGCTCGAGCCGGATCGTGTACTTGTCCCA	
Nono-GFP-F	TCCAAGCTTCTGCAGGAATTCATGCAGAGCAATAAAGCCTTTAACT	
Nono-GFP-R	GCCCTTGCTCACCATTCTAGAATATCGGCGGCGTTTATTTG	
Sox5-RT-R	CCAGCCTTGTAGCTGAAACCA	qPCR primer
Sox9-RT-F	GAGCCGGATCTGAAGAGGGA	
Sox9-RT-R	GCTTGACGTGTGGCTTGTTC	
Adcy3-RT-F	AATGGCCTCAACGGCTCGGC	
Adcy3-RT-R	ATGAGGGCAGGGGAGCTGGG	
Ccdc39-RT-F	GAGCGAAGAACATTTTAAGGCCA	
Ccdc39-RT-R	TTCAAGTCATCCAGCTTTTGAGT	
Cdk20-RT-F	CAAAGCCAAGCACGTAGAGAC	
Cdk20-RT-R	TGCACCACATATTGACTGTCC	
Pdzd7-RT-F	TCTCTGGGGGCATCGAGTC	
Pdzd7-RT-R	GCTCAAAGCCAGCCTGTAGAT	
Pkd2-RT-F	GGGGAACAAGACTCATGGAAG	
Pkd2-RT-R	GCCGTAGGTCAAGATGCACAA	
Gapdh-RT-F	ATGGTGAAGGTCGGTGTGAAC	

Gapdh-RT-R	GCCTTGACTGTGCCGTTGAAT	
Sox5-CUT&Tag-F	TAGTACACTTCCAGCAGCGGT	
Sox5-CUT&Tag-R	GCCTGGAGGTTGTCTAATGCC	
Sox9-CUT&Tag-F	AAATCCGGTCCAATCAGCGA	
Sox9-CUT&Tag-R	ACGTTAGATACCTCGGGCCA	
DNA Spike in-CUT&Tag-F	GCCTTCTTCCCATTTCTGATCC	
DNA Spike in-CUT&Tag-R	CACGAATCAGCGGTAAAGGT	

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927 **Supplementary Table 2. sgRNAs used in the study**

Name	Sequences (5' to 3')
Xap5-sgRNA1	GGCAAAAAAGGAGCAGTCAA
Xap5-sgRNA2	AGAGATGGCTATGTACGAGG
Nono-sgRNA3	AGTGGATCGGAACATCAAGG
Sox5-sgRNA4	GACACGCTCGAGTGCTCGGA
Sox5-sgRNA5	GATTTACGAGGAGTACGATG

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