Molecular identification of stone loaches of Choman River system, Tigris Basin, based on the Cytochrome *b* gene with an overview of the Cobitoidea Superfamily

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Abstract

Molecular data and phylogenetic relationships of four Choman loaches species (O*xynoemacheiluschomanicus*, *O. zagrosensis*, *O. kurdistanicus*and*Turcinoemacheiluskosswigi*)recentlymorphologically described from western Iran were evaluated with 64 species from the Cobitoidea superfamily based on their cytochrome *b* sequences to exhibit the placement of the Choman loaches species within the Cobitoidea superfamily. A comparative analysis of Kimura-2-parameter (K2P) distances was accomplished using sequence divergences of Cobitoidea to calculate intra and interspecific in superfamily, family and genus taxa. The average intraspecific K2P genetic distances of Choman loaches species was 0.005 whereas this value was 0.016 for the Cobitoidea superfamily. Molecular phylogenetic relationships were assessed using Maximum likelihood and Bayesian methods. Dendrograms obtained by these methods revealed all four Choman loaches species as distinct species among other reported Nemacheilidae Spp. These species were clustered with*Oxynoemacheilus* and*Turcinoemacheilus*genera withinotherspecies in the Nemacheilidae family.The phylogenetic analysis revealed that Cobitoideasuperfamily consistsof nine families((Gyrinocheilidae + Botiidae) + ((Catostomidae + Vaillentellidae) + ((Nemacheilidae + Cobitidae) + ((Ellopostomidae+Gastromyzontidae) +Balitoridae)))) and indicated Nemacheilidae is a valid and distinct family from Balitoridae.

Key words: Nemacheilidae, Oxynoemacheilus, Choman loaches, Kurdistan, K2P distance

Introduction

Loaches of the family Nemacheilidae represent a major lineage of the superfamily Cobitoidea that are diverse in freshwaters of Asia, Europe and north-east Africa with more than 45 genera and 570 known species (Kottelat, 2012). Eight genera and more than 40 speciesof nemacheilid are widely distributed in Iran's drainage basins (Coad, 1998; Nalbant&Bianco, 1998; Golzarianpouret al., 2009, 2011 and 2013; Abdoliet al., 2011; Kamangaret al., 2014; Freyhofet al., 2016). Although most of the new species are described morphologically, these approaches are faced with challenges (Robertson et al., 2001). Molecular procedures have been

applied in many studies for new speciesidentification and it is clear that these methods have great potential in resolving identification and phylogenic relationship of cryptic species and could be highly effective in classification of confusing species (Teletchea, 2009). Cytochrome *b* gene (Cytb) is frequently utilized for fish identification and phylogenetic study (Song et al., 1998; Biswaset al.,2001; Fariaset al., 2001; Perkins &Schall, 2002; Perdiceset al.,2004; Baharum&Nurdalila, 2011).

Choman watershed is a sub-basin of Tigris and located in the west of Iran. Recently, four stone loach species from Nemacheilidae family; *Turcinoemacheiluskosswigi* (Banarescu&Nalbant, 1964) and three new species from *Oxynoemacheilus* (*O. chomanicus, O. zagrosensis* and *O. kurdistanicus*), have been reported from this watershed based on morphological and some osteological characters (Kamangaret al., 2014). However, molecular data on these species have not as yet been reported. In order to bring molecular data to complete introducing and demonstrating the taxonomic status of these loach species from the Choman River sub-basin, molecular tools based on Cytochrome *b* gene was used complementary to morphological approaches in this study.

The taxonomy of Cobitoidea superfamily has been a controversialsubject for ichthyologistsin the last century. For instance, Nemacheilidloaches were initially classified in the Cobitidae family (Regan, 19111; Hora, 1932) and thereafter transferred to the Balitoridaefamily (Sawada, 1982; Siebert, 1987). For the first time, Nalbant (2002) changed this classification by treating the Nemacheilinae as the Nemacheilidae family. The results of the first molecular phylogenetic study of Cobitoidea by Liu et al. (2002) in which mtDNA control region sequences was used suggested once again the Nemacheilinaerank for this group withinthe Cobitoidea. Nevertheless, this classification was not approved by ichthyologists due to inadequate testing of

the phylogenetic relationship of loachessensustricto (Tang et al., 2006). In following molecular studies, the Nalbant (2002) classification was confirmed and nemacheilid loach fishes were classified in a separate family from the Balitoridae as the Nemacheilidae family (Šlechtováet al., 2007; Bohlen &Šlechtová, 2009; Chen et al., 2009). Finally, Kottelat (2012) identified 10 families (Gyrinocheilidae, Botiidae, Vaillantellidae, Cobitidae, Ellopostomatidae, Barbuccidae, Balitoridae, Gastromyzontidae, Serpenticobitidae and Nemacheilidae)in the Cobitoidea superfamily and Nemacheilidae was introduced as a valid family in this group. However, in some systematic and taxonomic reviews, the “Nemacheilinae” is still used (Wang et al., 2012; Du et al., 2012; Tang et al.,2013; Prokofiev &Golubstove, 2013; Li et al., 2015; Queet al., 2016) and using different expressions for this group has led to confusion.

The objectives of this study were firstly to evaluate cytochrome *b* sequence variation and the morphological tools associated by morphometric truss network analysis in grouping and identifying Choman loach species. Secondly, the aim was to examine the phylogenetic relationships in the superfamily Cobitoidea based on Cytochrome*b* sequences using new taxa and to note the placement of Choman loach species.

Materials and methods

Fish sampling

One hundred twenty five loachspecimens from the Choman sub-basinincluding *O. chomanicus* (38.1- 56.2 mm SL, n=40), *O. zagrosensis* (48.4- 58.4 mm SL, n=20), *O. kurdistanicus* (32.2- 60.5 mm SL, n=35) and *T. kosswigi* (25- 49.4 mm SL, n=30) were sampled from the same four locations as reported by Kamangar et al. (2014) between August and

September 2011 (Fig.1). Five specimens of muscle tissue were taken from each species for molecular study.

Morphological analysis

Truss network is a quantitative method representing the complete shape of fish and is an effective method for obtaining information regarding the great similarity of morphological variation (Cavalcantiet al., 2011). In the truss network system, 16 homologous landmarks (Fig.2) delineating 38 distance characters (Table 1) were measured on the body with calipers following methods outlined by Strauss and Bookstein (1982). An allometric method (Elliott et al., 1995) was used to remove size-dependent variation in morphometric characters:

$$M_{adj} = M\,(L_s / L_0)^{b}$$

In this equation M is the original measurement, M_{adj} is the size adjusted measurement, $L_0$is the standard length of the fish, L_s is the overall mean of the standard length for all fish from all samples in each analysis. The coefficient of b was estimated for each character from the observed data as the slope of the regression of $log\ M$ on $log\ L_0$ using all fish in the groups. Analysis of variance (ANOVA) followed by Duncan's multiple range test was applied to test any significant differences oflandmarks betweenthe four Chomanloach species. The size-adjusted data were submitted to a stepwise discriminant function analysis (DFA). SquaredMahalanobis distances matrix was obtained from the cluster analysis and used to draw a dendrogram based on the unweighted pair group method analysis (UPGMA) (Bektaz&Belduz, 2009). Statistical analysis was performed using SPSS ver. 16 and STATISTICA ver. 6.0 software.

DNA extraction and Cytochrome *b*sequencing

DNA was extracted using salting out protocol from muscle tissue (Cawthornet al., 2011). To amplify a 1140 bp of Cytochrome *b*, DNA extracts were subjected to PCR amplification,

using primer set of L14724 (5'-GAC TTG AAA AAC CAC CGT TG-3') and H15915 (5'-CTC CGA TCT CCG GAT TAC AAG AC-3') (Tang *et al.*, 2006). PCR was performed at an initial denaturation step at 94 ˚C for 3 min followed by 35 cycles at 94 ˚C for 30 s, 52-58 ˚C for 45 s, 72 ˚C for 1 min and a final extension at 72 ˚C for 8 min. The amplified fragments were purified with Promega DNA purification kit following the manufacturer's instructions. The purified fragments were sequenced bidirectional by BIONEER Company. All sequences are available from the GenBank database.

Phylogenetic analysis

For phylogenetic analysis, sequences from Choman loaches species (20 sequences) were evaluated with sequences from 180 individuals representing a total of 64 species from Cobitoidea. These sequences were downloaded from GenBank. Cytochrome *b* sequence of *Capoetatrutta*(JF798333) and *Cyprinuscarpio*(AB158807) were used as out-grouping in this study. We followed the classification of Cobitoidea proposed by Kottelat (2012).Moreover, we evaluated the Catostomidae relationship with Cobitoidea as proposed by Šlechtováet al.(2007) as a family member of Cobitoidea. There wasno reported Cytochrome *b*sequences from Barbuccidae and Serpenticobitidaefamilies and thus sequences from Gyrinocheilidae,Botiidae, Vaillentellidae, Nemacheilidae, Cobitidae, Gastromyzontidae,Catostomidae, Ellopostomidae and Balitoridae families onlywere used in this analysis.

Kimura two parameter (K2P) distance model (Kimura, 1980) was used to calculate sequence divergences in the Cobitoidea superfamily. Several categories of K2P were calculated to test the first objective of this study: intra and interspecificdistances inthe Cobitoidea superfamily,intra and interspecific distances in the Balitoridae, Nemacheilidae, Catostomidae, Cobitidae, Botiidae and Gastromyzontidae family, intra and interspecific distances for

Oxynoemacheilus genus and finallyintraspecific distance in Choman loachspecies.The averages of each category were compared statistically to test any possible overlapping between these distances. K2P genetic distances were calculated using MEGA 4.0 software package (Tamura et al., 2007). The comparisons between the average genetic distances werecarried out in XLSTAT software trial version. The Mann-Whitney test was used to statistically compare all intraspecific and interspecific comparisons. To calculate conspecific (intraspecific)distances, the species used were ones that were represented at least by two or more specimens in the data bank.Confamilial(intraspecific distance in each family) distanceswere also calculated when at least two genera sequences were available.

Phylogenetic trees were reconstructed for the gene data set using Maximum Likelihood (ML) and Bayesian Analyses (BA). Sequences were aligned using Clustal X ver. 1.85 (Thompsonet al., 1997). Phylogenetic trees using ML were estimated as implemented in RAxML (Randomized Axelerated Maximum Likelihood, version 7.0.4) (Stamatakis, 2006). The Akaike information criterion (AIC) implemented in MODELTEST v3.4 (Posada and Crandall, 1998) was used to identify the optimal molecular evolutionary model for each partition on the sequence data set. GTR+I+G model were utilizedto obtainthe optimal ML trees and bootstrap support. Robustness of the inferred trees was evaluated using bootstrap analysis on 1,000 pseudoreplications using RAxML 7.0.4 (Felsenstein, 1985; Stamatakiset al., 2008). BA analyses were conducted using Mr. Bayes v3.1.2 (Huelsenbeck and Ronquist, 2001). For BA, 5,000,000 cycles were implemented for four simultaneous Monte Carlo Markov chains,sampling the Markov chain at intervals of 100 generations. Log-likelihood stability was attained after 100,000 generations; the first 1,000 trees were discarded as “burn-in” in each analysis. Support for BI tree

nodes was determined based on values of Bayesian posterior probabilities. Final trees were visualized in the FigTree v.1.4.2 program (Rambaut, 2009).

Results

Morphological traits

The Choman loach species were well distinguished morphometricallybased ontruss network analysis. The ANOVA results showed significant differences between species for all truss distances ($p < 0.01$).The effect of sex was not significant within the same species for all examined measurements. The comparison of mean truss distances between the four loach species revealed significant differences between them (Table 1).Most landmarks were significantly longer in *O. zagrosensis* than other Choman loach species (Table 1). Fourteen landmarks (ADPA, PAFUL, OCPP, OCAD, SPG, FULEUL, PVAA, OCPG, PPAV, PDPA, ADPD, EULELL, OCAV, and FLLELL) were revealed by stepwise DFAs as the most discriminative landmarks to differentiate the loach species. The canonical discriminant function analysis produced three functions; the first two canonical discriminant functions explained 95% of variability between species (Table 2). The Wilks' lambda statistics showed the power ofdiscriminant functions in separating species (Table 3). The group centroid of each species was comprehensivelydisparate based on visual analysis of scattering diagrams (Fig. 3).

Cross validation results showed that 99.2% of individuals were correctly classified in their species groups based on morphological differences (Table 4) and only one specimen from *O. zagrosensis* was incorrectly classified in the *O. chomanicus* group (Fig. 3). The dendrogramderived from cluster analysis of groups of centroids showed two separate groups: one included the *Oxynoemacheilus spp.*and the other included the*Turcinoemacheilus* (Fig. 4). These groupings confirm the results obtained from the discriminant analysis for these species.

Molecular study

The average sequence length was 1097 bp (range= 1011 to 1131 bp); all read lengths were greater than 1000 bp. No internal indels were observed in any sequences generated herein or downloaded from the Genbank.There were 1157 nucleotide sites,of which 685were parsimony informative sites (55%). The average nucleotide composition of *O. chomanicus*, *O. zagrosensis*, *O. kurdistanicus* and *T. kosswigi*sequenceswere Adenine (A) 25%, 24.9%, 25.4% and 28.7%, Thymine (T) 31%, 30.4%, 30.2% and 31.5%, Cytosine (C) 27.5%, 27.2%, 27.8% and 24.4% and Guanine (G) 16.5%, 17.4%, 16.6% and 15.4%, respectively. In all four species, the content of A+T was higher than that of C+G. Transitions (Ti) / transversions (Tv) ratios for *O. chomanicus*, *O. zagrosensis*, *O. kurdistanicus* and *T. kosswigi*were 2.54, 1.28, 0.79 and 2.19, respectively. The average nucleotide composition in Cobitoidea superfamily was Adenine (A) 26.3%, Thymine (T) 30.7%, Cytosine (C) 27.2% and Guanine (G) 15.8%, and Transitions (Ti) / transversions (Tv) ratios in this superfamily was estimated as 1.82.

The K2P distances were calculated in the Cobitoidea superfamily and Nemacheilidae, Balitoridae, Gastromyzontidae, Cobitidae,Botiidae, Catostomidaefamilies and the*Oxynoemacheilus* genus in order to reveal any possible overlapping and statistically significant differences between these distances (Table 5).Significant differences were observed between intraspecific and interspecific K2P distances in all comparisons ($p < 0.001$). There was no consequential overlap between all intra and interspecific K2P distance comparisons (Table 5).The mean interspecific K2P distances were higher by at least 10-fold than their intraspecific genetic distances except those for Balitoridae, Gastromyzontidae and Catostomidae families (Table 5). The average interspecies of *Oxynoemacheilus* genus were32.50-fold higher than that

amongst individuals within (intraspecific) the *Oxynoemacheilus*species (Table 5). This ratio was 32.50, 21.67 and 43.33 higher for *O. chomanicus*, *O. zagrosensis* and *O. kurdistanicus*, respectively. The average interspecific from different species but within the Nemacheilidae family (0.216) was 54.00 fold higher than that amongst individuals within the*Oxynoemacheilus*species (0.004) (Table 5).

The same topologies were achieved when the Cobitoidea superfamily data was analyzed with BA (Fig. 5) and ML (Fig. 6) methods. In both analyses,the Choman loach species were correctly classified within their family and genus clades. *O. chomanicus* and *O. zagrosensis* formed monophyletic groups and then gathered in one clade with *O. kurdistanicus*.Therecently described Choman loach species were distinguished from other *Oxynoemacheilus* spp. which had their Cytochrome*b* gene sequencesreported.

The reconstructed phylogenetic treesrevealed the Cobitoideasuperfamily as a monophyletic group (Fig 5 and 6).At the species level, all individuals were clustered in their monophyletic groups except one specimen of *Paracobitisanguillioides* that was grouped within the cluster of *Paracobitisacuticephala*. At the genus level,paraphyletic clusters were found for *Schistura*, *Nemacheilus*, *Cobitis*, *Iksookimia* and *Kichulchoia* genera(Fig. 5, 6).At the family level, all studied families were clustered in their monophyletic clades.Gyrinocheilidae and Botiidaerepresent the most-basal clades of the Cobitoideasuperfamily and formed the sister group to the clade formed by Catostomidae, Vaillentellidae, Nemacheilidae, Cobitidae, Ellopostomidae, Gastromyzontidae and Balitoridae families. These trees showed monophyletic relationship between Nemacheilidae and Cobitidae families. The phylogenetic relationships indicated Balitoridae and Nemacheilidae astwo distinctive families which hadmistakenly been classified as onefamily.

Discussion

Identification of loachsensustricto species hasalways been difficult and their classification not stable (Siebert, 1987; Sawada, 1982; Berg, 1940; Hora, 1932; Regan, 1911). This presented a challenge for ichthyologist toundertake researchon these species.The main problem to considerin studying loach speciesis identification and classification ofcryptic and sibling species. Most problems appeared in two modes: differentiation between specimens from the same species and sometimes high similarity between specimens from different species. Molecular tools in combinationwith morphological methodswereintroduced to resolve these problems(Purryet al., 2016; Kelehearet al., 2011).The present study aimed to obtain molecular data for Choman loach species recently introduced based on morphological methods.

Results of univariate and multivariate analysesof truss network datagreatly distinguished the four Choman loach species based on their truss distances.Univariate analysis showed that *O. zagrosensis* is more rotundthan other Choman loach species. This specie is distinguished from other Choman loach species by having longer truss distances in the head, the trunk and the caudal peduncle parts of body.Multivariate analysis using DFAs well differentiatedthe four loach species based on their truss network data. Based on this analysis, the most discriminant landmarks were the distances located between fins, inthe caudal fin, in the head region, in the trunk and in the caudal peduncle.Canonical scatter plot of DFA and dendrogram based on the cluster analysis using the most discriminative landmarks showed that *O. chomanicus* and *O. zagrosensis* are more similar but*O. zagrosensis* can be more easily distinguished through having

longer landmarksthan their corresponding landmarks in *O. chomanicus*.These results correspond to those reported for these two species using morphometric analysis (Kamangaret al., 2014).

The molecular analysis based on Cyt-b sequences better identified the new loach species from the Chomansub-basin of Kurdistan, Iran. The topology of dendrogramsobtained from the Cyt-b sequences of the three new loach species were in accordance with the dendrogram obtained from the cluster analysis based on truss network measurements. Molecular data as well as morphometric analysis revealed three distinct new described species.We were unable to find similar sequences from *Oxynoemacheilus* in phylogenetic relationships by BA and ML trees which could be very close to the *Oxynoemacheilus*species from the Chomansub-basin. These results agree with the morphological classification of these species by Kamangar et al. (2014). *O. chomanicus* was originally described as a more similar species to *O. zagrosensis* based on morphological features (Kamangaret al., 2014). Two methods which have been broadly used for species identification based on DNA barcoding are genetic distance and monophyly based methods (Meyer *et al.*, 2005; Zang, 2011; Zou*et al.*, 2011). Two distinct monophyletic clades reconstructed based on Cyt-b sequences of this new specieswithin the Cobitoidea superfamily tree, did not show any overlapping between individuals from *O. zagrosensis* and *O. chomanicus*.Significant differences were also obtained in comparisonsof K2P genetic distances between the mean intraspecific and the mean interspecies within the *Oxynoemacheilus* genus. The average K2P distances were 54.00-fold higher within the Nemacheilidae than that betweenindividuals within the *Oxynoemacheilus* genus.Thesefindingsindicate the reliability of the Choman loach identificationmethod based on Cyt-b sequences and whichwere validated by Hebert et al.(2003) and Barrett and Hebert,(2005). The Cytochrome*b* sequences of

Turcinoemacheiluskosswigi from the Choman river shows some similarity with *Turcinoemacheilus sp.* sequences as reported by Jamshidi et al. (2013) (Accessions; GQ 338826, GQ 338827 and GQ 338828). Jamshidi et al. (2013) later declared that these sequences belong to the*Turcinoemacheiluskosswigi*. Our findings did not confirm their declaration. Our reconstructed trees using BA and ML methods revealed that the*Turcinoemacheiluskosswigi*and *Turcinoemacheilus sp.* are two distinct species (Figs. 5 and6). *Turcinoemacheilushafezi*was also reported (Golzarianpouret al., 2013) more recently from the same area by Jamshidi et al. (2013) and it seems that these sequences are related to *T. hafezi*.

Cobitoideais the most complicatedfish group inregards their identification and phylogenetic relationship.It appearsthat to reach an acceptable phylogenic position for this superfamilywill remain elusive. Our results clearly revealthatCobitoidea is a monophyletic superfamily and comprises of ninelineages(Fig 5, 6).These classification results are in accordance with those reported by Šlechtová et al. (2007) in whichGyrinocheilidae, Botiidae, Vaillentellidae, Cobitidae,Catostomidae,Nemacheilidae and Balitoridaeare valid family ranks. Moreover, our results illustratedthat Ellopostomidae is a distinct family as previously described by Bohlen and Šlechtová (2009). Nemacheilidae and Balitoridae representtwo distinctfamilies as alreadydescribed by Tang et al. (2006) and Šlechtová et al (2007).In our phylogenetic analysis, Nemacheilidae was clustered as a sister group with Cobitidae which was in agreement with those reportedby Tang et al (2006)using mtDNA.

There are some disagreementsregarding the Catostomidae phylogenetic relationship with Cobitoidea. Šlechtováet al.(2007) reported the Catostomidae as a distinct family within Cobitoidea based on molecular data using nuclear genes. However, some authors prefers to treat

this groupas a sister group with loachessensustricto (Tang et al., 2006; Chen et al., 2009) or even with other Cypriniformes (Maydenet al.,2009).The present results based on Cyt-b demonstrate with strongstatistical support that Catostomidae is a distinct monophyly family within Cobitoidea. These conflictions might be due to using different genes and their evolutionary histories as described by Mayden et al., 2009.

The Balitoridae cluster was divided into two distinct lineages.Onecluster contains speciesthat are currently classified as Balitoridae by some authors (Chen et al., 2009).Tang et al. (2006) treated this group as the Gastromyzoninae subfamily with Balitorinae in Balitoridae family. Kottelat (2012) classified the member of this cluster in the Gastromyzontidae familybased on morphological analysis. Our results obtained strong statistical support for this lineage. When we included data from the Ellopostomidae family, the cluster of the Gastromyzontidae family showed a distinct lineage with high node support and as a sister group with the latter family. Thus, we consider this group as the Gastromyzontidae family as proposed morphologically by Kottelat (2012).Current analysis revealed that *Ellopostomamystax*is a distinct lineage within the Cobitoidea superfamily and is more related to the Gastromyzontidae family than the Balitoridae family. Bohlen and Šlechtová (2009) introduced the Ellopostomidae family within the Cobitoidea superfamily as a valid family most closely related to the Nemacheilidae and Balitorodaefamilies. However, they did not consider the family Gastromyzontidae in their analysis.Dissociating Gastromyzontidae from Balitoridaeand including Ellopostmidae in one phylogenetic analysis coulddescribe why former studies haddifficulties in describing relationships of families in the Cobitoidea superfamily.

In conclusion, our molecular data shows the reliability of Choman loach species. The phylogentic analysis revealed nine distinct families within the Cobitoidea superfamily. However,

we recommend more sample analysis of Barbuccidae and Serpenticobitidae to clarify complete molecular phylogeny of this superfamily.

Competing interests

The authors declare no competing or financial interests.

Author contributions

E. G., H. F., B. B. K. and M. A. N. designed the project, E. G. and B. B. K. conducted experiments;E. G., H. F. and B. B. K. analyzed the data; E. G. and B. B. K. wrote the manuscript; H. F. and B. B. K. edited the manuscript.

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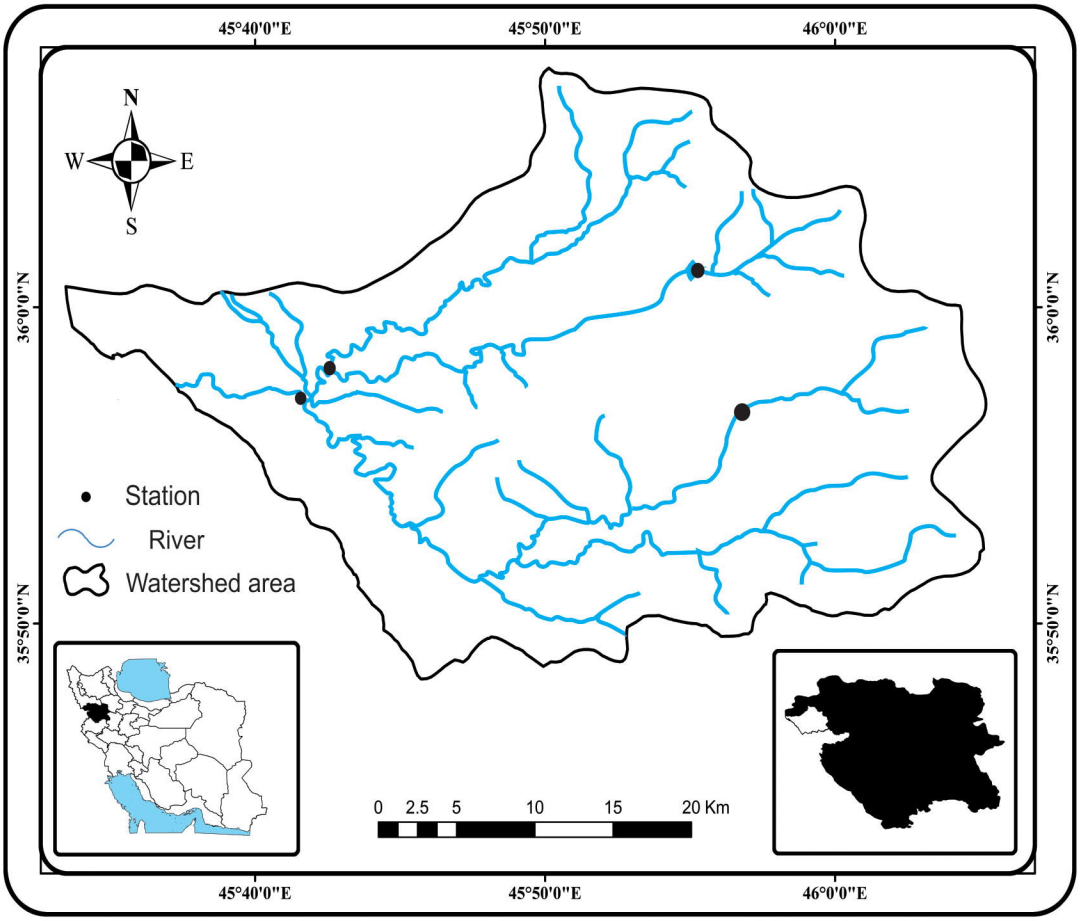
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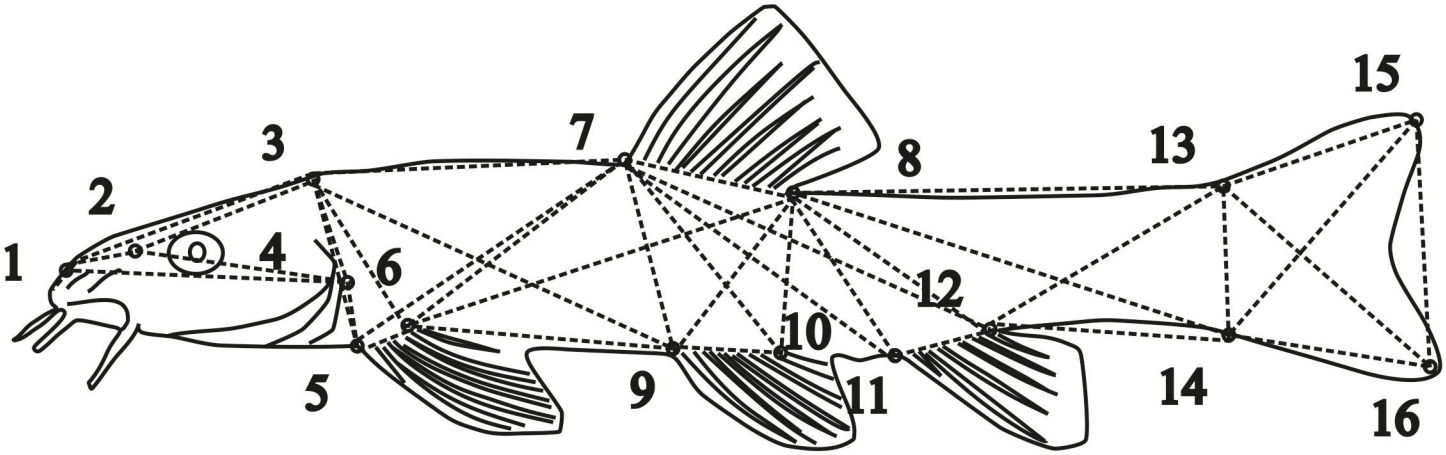
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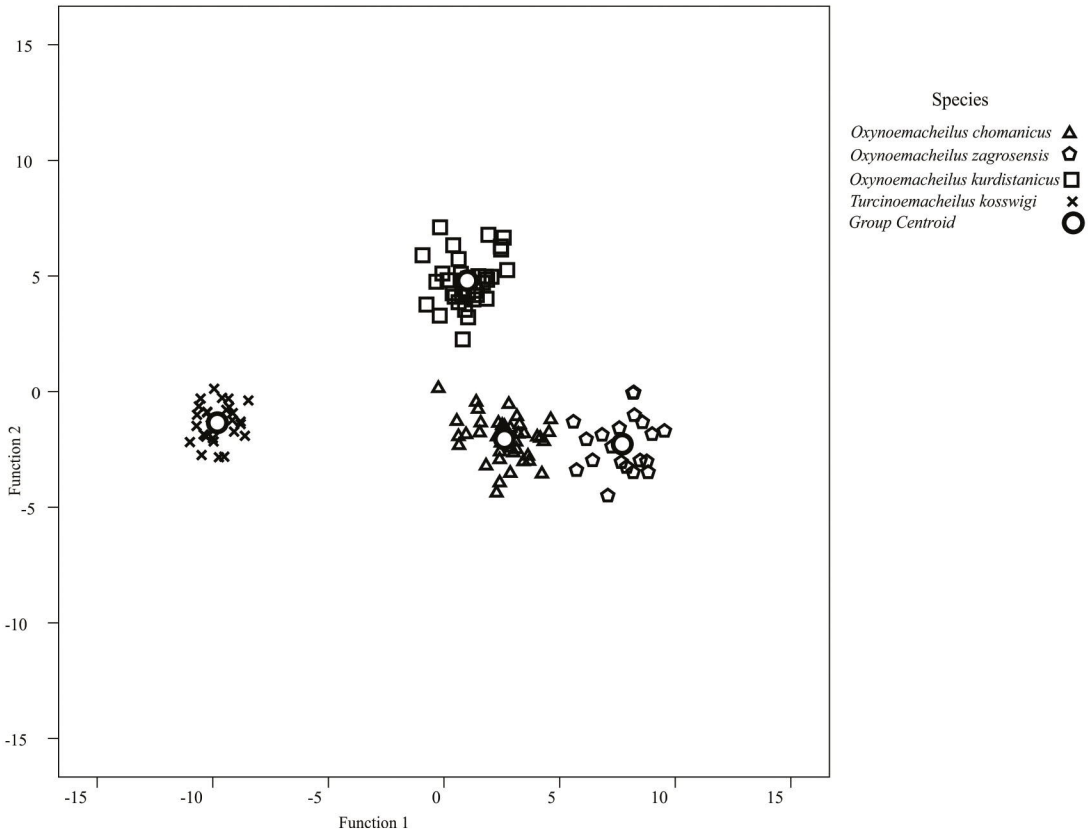
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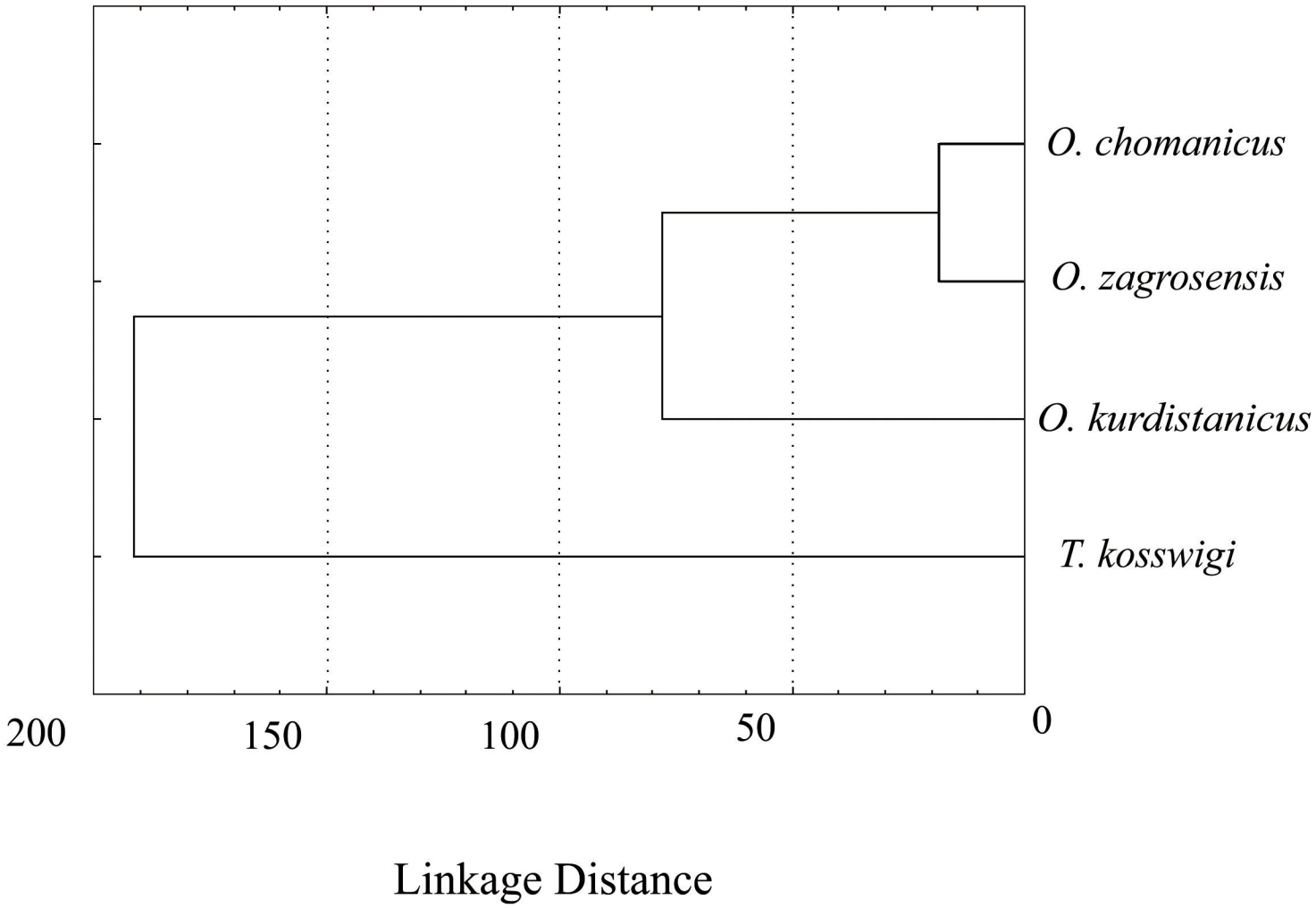
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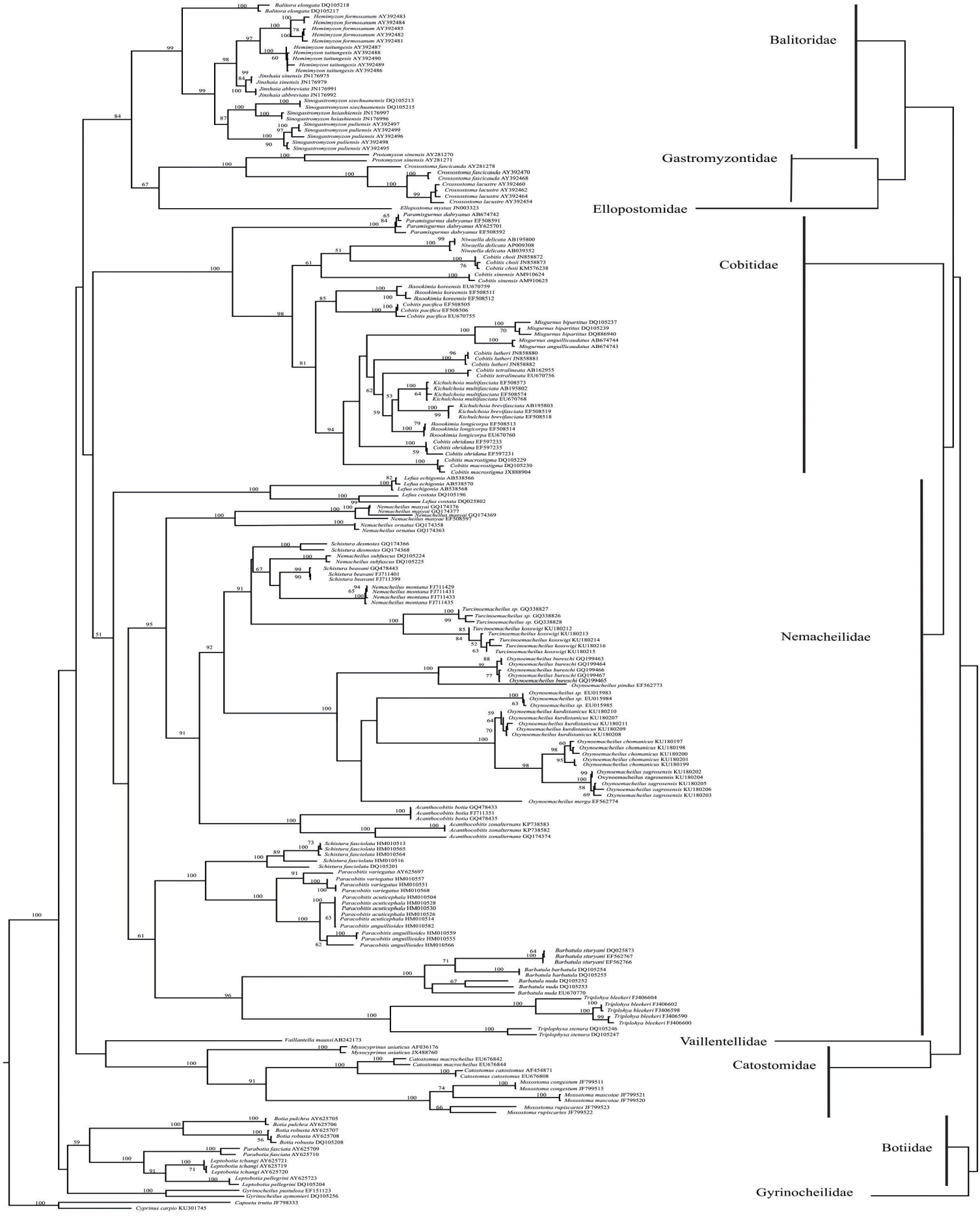
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Catostomus catostomus AF454871
Catostomus catostomus EU676808
Moxostoma congestum JF799511
Moxostoma congestum JF799515
Moxostoma mascotae JF799521
Moxostoma mascotae JF799520
Moxostoma rupiscartes JF799523
Moxostoma rupiscartes JF799522
Botia pulchra AY625705
Botia pulchra AY625706
Botia robusta AY625707
Botia robusta AY625708
Botia robusta DQ105208
Parabotia fasciata AY625709
Parabotia fasciata AY625710
Leptobotia tchangi AY625721
Leptobotia tchangi AY625719
Leptobotia tchangi AY625720
Leptobotia pellegrini AY625723
Leptobotia pellegrini DQ105204
Gyrinocheilus pustulosa EF151123
Gyrinocheilus aymonieri DQ105256
Capoeta trutta JF798333
Cyprinus carpio KU301745

Figure legends

Fig. 1. Map of Choman river basin showing the Choman loaches sampling sites.

Fig. 2. The truss network, anatomical landmark points (16 points) and distance measured for Choman loaches species. Anatomical landmark points, 1: Tip of snout; 2: End of nostril; 3: Forehead (end of frontal bone); 4: Hind most point of opercle; 5: Insertion of pectoral fin; 6: Insertion of last pectoral fin ray; 7: Insertion of dorsal fin; 8: Insertion of last dorsal fin ray; 9: Insertion of pelvic fin; 10: Insertion of last pelvic fin ray; 11: Insertion of anal fin; 12: Insertion of last anal fin ray; 13: Insertion of first caudal fin ray in upper lobe; 14: Insertion of first caudal fin ray in lower lobe; 15: The most distal point of caudal fin ray in upper lobe; 16:The most distal point of caudal fin ray in lower lobe.

Fig. 3. Scatterplot of individual distances measured based on truss network analysis from the first two canonical functions for Choman loaches species.

Fig. 4. Dendrogram derived from cluster analysis of Mahalanobis distance using UPGMA between Choman loaches species.

Fig. 5. The 50% majority-rule consensus tree resulting from Bayesian analysis of the concatenated cytochrome *b* sequences dataset. Numbers at nodes represent posterior probabilities for Bayesian analysis. An overview of the relationship obtained for Cobitoidea superfamily shown (right).

Fig. 6. Phylogenetic tree relationships among taxa of the Cobitoidea, Branch lengths are proportional to inferred character substitutions under the GTR+G+I model. Numbers on branches are ML bootstrap values; those below 50% are not shown. An overview of the relationship obtained for Cobitoidea superfamily shown (right).

Tables

Table 1. Comparisons of the landmark distance measurements between Choman loaches species. Values are mean ± standard error. The values that followed by different letters indicate a significant difference between species for the same measurements ($P < 0.05$).

Distance measured	Characters abbreviation	*O. chomanicus*	*O. zagrosensis*	*O. kurdistanicus*	*T. kosswigi*
1-2	SPN	2.844 ±0.347 ^{b}	3.519 ± 0.276 ^{c}	3.034 ± 0.516 ^{b}	2.308 ± 0.240 ^{a}
1-3	SOC	8.601 ± 0.618 ^{b}	10.084 ±0.597 ^{c}	8.863 ± 0.547 ^{b}	6.517 ± 0.468 ^{a}
1-4	SPG	10.374 ± 0.509 ^{c}	11.746 ± 0.361 ^{d}	9.815 ± 0.565 ^{b}	7.241 ± 0.541 ^{a}
2-3	PNOC	6.162 ± 0.392 ^{b}	6.954 ± 0.394 ^{c}	6.336 ± 0.379 ^{b}	4.862 ± 0.320 ^{a}
2-4	PNPG	8.170 ± 0.589 ^{c}	9.048 ± 0.469 ^{d}	7.537 ± 0.329 ^{b}	5.460 ± 0.314 ^{a}
3-4	OCPG	5.945 ± 0.590 ^{c}	6.725 ± 0.604 ^{d}	4.923 ± 0.520 ^{b}	3.178 ± 0.383 ^{a}
3-5	OCAP	6.418 ± 0.556 ^{c}	7.246 ± 0.375 ^{d}	5.742 ± 0.484 ^{b}	3.842 ± 0.469 ^{a}
3-6	OCPP	7.324 ± 0.557 ^{c}	8.100 ± 0.326 ^{d}	6.388 ± 0.625 ^{b}	4.368 ± 0.568 ^{a}
3-7	OCAD	15.042 ± 1.080 ^{b}	17.298 ± 1.177 ^{c}	14.251 ± 1.926 ^{a}	16.791 ± 1.385 ^{c}
3-9	OCAV	18.730 ± 0.944 ^{c}	21.488 ± 0.756 ^{d}	17.293 ± 1.076 ^{b}	15.733 ±0.624 ^{a}
4-5	PGAP	1.528 ± 0.394 ^{b}	1.479 ± 0.478 ^{b}	1.463 ± 0.395 ^{b}	1.091 ± 0.352 ^{a}
5-6	APPP	2.243 ± 0.337 ^{b}	2.689 ± 0.301 ^{c}	2.368 ± 0.456 ^{b}	1.524 ± 0.255 ^{a}
5-7	APAD	13.767 ± 1.370 ^{a}	16.149 ± 0.830 ^{c}	13.647 ± 1.628 ^{a}	15.197 ± 0.997 ^{b}
6-7	PPAD	13.106 ± 1.378 ^{a}	14.910 ± 1.071 ^{b}	12.820 ± 1.540 ^{a}	14.786 ± 0.896 ^{b}
6-8	PPPD	18.599 ± 1.494 ^{a}	21.816 ± 0.802 ^{b}	18.365 ± 2.002 ^{a}	18.677 ± 0.694 ^{a}
6-9	PPAV	13.224 ± 1.216 ^{b}	14.998 ± 0969 ^{c}	12.288 ± 1.080 ^{a}	13.075 ± 2.139 ^{b}
7-8	ADPD	6.473 ± 0.551 ^{b}	8.349 ± 0.672 ^{d}	6.837 ± 0.776 ^{c}	4.148 ± 0.513 ^{a}
7-9	ADAV	7.373 ± 0.840 ^{c}	9.012 ± 0.955 ^{d}	6.805 ± 0.652 ^{b}	4.651 ± 0.516 ^{a}
7-10	ADPV	7.605 ± 1.061 ^{c}	8.506 ± 0.812 ^{d}	6.745 ± 0.786 ^{b}	4.658 ± 0.434 ^{a}
7-11	ADAA	15.353 ± 0.844 ^{c}	17.808 ± 0.982 ^{d}	13.336 ± 0.964 ^{b}	10.432 ± 1.071 ^{a}
7-12	ADPA	18.115 ± 0.834 ^{c}	21.346 ± 1.068 ^{d}	16.088 ± 0.933 ^{b}	12.969 ± 0.668 ^{a}
8-9	PDAV	7.891 ± 0.759 ^{b}	9.837 ± 1.011 ^{c}	7.725 ± 1.101 ^{b}	6.857 ± 0.733 ^{a}
8-10	PDPV	7.781 ± 1.030 ^{b}	9.354 ± 0.632 ^{c}	7.387 ± 1.369 ^{b}	6.658 ± 0.886 ^{a}
8-11	PDAA	9.094 ± 0.772 ^{c}	10.618 ± 0.402 ^{d}	6.973 ± 0.465 ^{b}	6.418 ± 0.739 ^{a}
8-12	PDPA	11.347 ± 0.751 ^{c}	13.143 ± 0.828 ^{d}	9.116 ± 0.705 ^{b}	8.378 ± 0.633 ^{a}
8-13	PDFLL	16.639 ± 0.821 ^{b}	19.068 ± 0.924 ^{c}	16.260 ± 0.893 ^{b}	14.436 ± 0.725 ^{a}
8-14	PDFUL	17.670 ± 0.850 ^{c}	20.290 ± 0.876 ^{d}	16.682 ± 0.775 ^{b}	14.879 ± 0.615 ^{a}
9-10	AVPV	1.934 ± 0.216 ^{b}	2.290 ± 0.182 ^{c}	1.989 ± 0.281 ^{b}	1.452 ± 0.352 ^{a}
10-11	PVAA	9.462 ± 0.580 ^{b}	10.837 ± 0.696 ^{c}	7.942 ± 0.721 ^{a}	9.429 ± 1.353 ^{b}
11-12	AAPA	3.935 ± 0.505 ^{b}	4.494 ± 0.428 ^{c}	4.105 ± 0.509 ^{b}	3.152 ± 0.410 ^{a}

12-13	PAFLL	8.758 ± 0.607 [b]	10.584 ± 0.509 [d]	9.790 ± 0.597 [c]	7.508 ± 0.448 [a]
12-14	PAFUL	7.334 ± 0.687 [b]	8.429 ± 0.792 [c]	9.355 ± 0.695 [d]	6.944 ± 0.457 [a]
13-14	FLLFUL	4.966 ± 0.589 [c]	6.172 ± 0.487 [d]	3.997 ± 0.288 [b]	3.195 ± 0.237 [a]
13-15	FLLEUL	8.970 ± 0.670 [b]	10.161 ± 0.653 [c]	10.217 ± 0.565 [c]	6.321 ± 0.432 [a]
13-16	FLLELL	8.585 ± 0.462 [b]	9.583 ± 0.423 [c]	9.570 ± 0.787 [c]	6.121 ± 0.422 [a]
14-15	FULEUL	8.504 ± 0.447 [b]	9.348 ± 0.515 [c]	9.698 ± 0.736 [d]	6.237 ± 0.448 [a]
14-16	FULELL	9.752 ± 0.742 [b]	11.071 ± 0.722 [d]	10.230 ±0.849 [c]	6.564 ± 0.425 [a]
15-16	EULELL	4.158 ± 0.860 [c]	4.609 ± 1.121 [c]	3.606 ± 1.075 [b]	2.853 ± 0.520 [a]

Table 2. Eigenvalue, percentage of variance and cumulative variance explained by three discriminant functions in the truss network analysis of Choman loaches species.

Function	Eigenvalue	% of Variance	Cumulative %
1	36.198	75.5	75.5
2	9.345	19.5	95.0
3	2.420	5.0	100.0

Table 3. Test of significance of discriminant functions for verifying differences of Choman loach species.

Functions	Wilks' Lambda	Chi-square	df	P value
1 through 3	0.001	825.962	42	0.000
2 through 3	0.028	410.093	26	0.000
3	0.292	141.393	12	0.000

Table 4. Percentage of specimens classified in each species and after cross validation for truss network distance measurements for Choman loaches species.

species	*O. chomanicus*	*O. zagrosensis*	*kurdistanicus*	*osswigi*
Original				
O. chomanicus	100	00.0	00.0	00.0
O. zagrosensis	00.0	100.0	00.0	00.0
O. kurdistanicus	00.0	00.0	100.0	00.0
T. kosswigi	00.0	00.0	00.0	100.0
Cross validated				
O. chomanicus	100	00.00	00.0	00.0
O. zagrosensis	5.0	95.0	00.0	00.0
O. kurdistanicus	00.0	00.0	100.0	00.0
T. kosswigi	00.0	00.0	00.0	100.0

Table 5. Summary of intra and interspecific K2P distances in the Cobitoidea superfamily, 6 families, *Oxynoemacheilus* genus and Choman loaches species.

Taxa	Comparisons	Number of comparisons	Median	Minmum	Maximum	Mean	ratio	Standard deviation	Z, P-value
Cobitoidea	intraspecific	306	0.006	0.000	0.108	0.016		0.024	
	interspecific	19664	0.239	0.002	0.339	0.231	14.44	0.044	-25.96, 0.000
Nemacheilidae	intraspecific	131	0.006	0.000	0.108	0.018		0.028	
	interspecific	4240	0.223	0.002	0.317	0.216	12.00	0.049	-19.28, 0.000
Balitoridae	intraspecific	35	0.006	0.000	0.028	0.010		0.009	
	interspecific	265	0.091	0.016	0.137	0.091	9.10	0.025	-9.56, 0.000
Gastromyzontidae	intraspecific	10	0.015	0.005	0.082	0.032		0.034	
	interspecific	26	0.157	0.041	0.182	0.119	3.72	0.058	-3.64, 0.000
Cobitidae	intraspecific	45	0.005	0.000	0.075	0.012		0.018	
	interspecific	889	0.151	0.050	0.218	0.150	12.50	0.032	-11.30, 0.000
Bottidae	intraspecific	9	0.005	0.000	0.030	0.007		0.009	
	interspecific	57	0.153	0.073	0.190	0.144	20.57	0.034	-4.79, 0.000
Catostomidae	intraspecific	6	0.004	0.000	0.090	0.021		0.035	
	interspecific	60	0.184	0.093	0.237	0.168	8.00	0.041	-4.02, 0.000
Oxynoemacheilus	intraspecific	43	0.003	0.000	0.010	0.004		0.004	
	interspecific	257	0.164	0.011	0.201	0.130	32.50	0.063	-10.50, 0.000
O. chomanicus	intraspecific	10	0.005	0.000	0.008	0.004	32.50	0.003	-5.370, 0.000
O. zagrosensis	intraspecific	10	0.006	0.000	0.008	0.006	21.67	0.003	-5.370, 0.000
O. kurdistanicus	intraspecific	10	0.003	0.000	0.006	0.003	43.33	0.002	-5.370, 0.000
T. kosswigi	intraspecific	10	0.0097	0.006	0.016	0.009		0.003	