Short title: Using Bayesian networks to association metadata and the vaginal microbiome

Title: Associations between sexual habits, menstrual hygiene practices, demographics and

the vaginal microbiome as revealed by Bayesian network analysis

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Abstract

The vaginal microbiome plays an influential role in several disease states in reproductive age

women, including bacterial vaginosis (BV). While demographic characteristics are associated

with differences in vaginal microbiome community structure, little is known about the influence

of sexual and hygiene habits. Furthermore, associations between the vaginal microbiome and risk

symptoms of bacterial vaginosis have not been fully elucidated. Using Bayesian network (BN)

analysis of 16S rRNA gene sequence results, demographic and extensive questionnaire data, we

describe both novel and previously documented associations between habits of women and their

vaginal microbiome. The BN analysis approach shows promise in uncovering complex

associations between disparate data types. Our findings based on this approach support published

associations between specific microbiome members (e.g., *Eggerthella*, *Gardnerella*, *Dialister*,

Sneathia and *Ruminococcaceae*), the Nugent score (a BV diagnostic) and vaginal pH (a risk

symptom of BV). Additionally, we found that several microbiome members were directly

connected to other risk symptoms of BV (such as vaginal discharge, odor, itch, irritation, and

yeast infection) including *L. jensenii*, *Corynebacteria*, and *Proteobacteria*. No direct

connections were found between the Nugent Score and risk symptoms of BV other than pH,

indicating that the Nugent Score may not be the most useful criteria for assessment of clinical

BV. We also found that demographics (i.e., age, ethnicity, previous pregnancy) were associated

with the presence/absence of specific vaginal microbes. The resulting BN revealed several as-yet

undocumented associations between birth control usage, menstrual hygiene practices and

specific microbiome members. Many of these complex relationships were not identified using

common analytical methods, i.e., ordination and PERMANOVA. While these associations

require confirmatory follow-up study, our findings strongly suggest that future studies of the

vaginal microbiome and vaginal pathologies should include detailed surveys of participants'

sanitary, sexual and birth control habits, as these can act as confounders in the relationship

between the microbiome and disease. Although the BN approach is powerful in revealing

complex associations within multidimensional datasets, the need in some cases to discretize the

data for use in BN analysis can result in loss of information. Future research is required to

alleviate such limitations in constructing BN networks. Large sample sizes are also required in

order to allow for the incorporation of a large number of variables (nodes) into the BN,

particularly when studying associations between metadata and the microbiome. We believe that

this approach is of great value, complementing other methods, to further our understanding of

complex associations characteristic of microbiome research.

Introduction

The microbiome plays a critical role in human health, and the vaginal microbiome has been linked to urogential diseases of reproductive age women, including bacterial vaginosis (BV) (1–3). BV affects nearly one-third of women in the United States (4) and has been implicated in poorer pregnancy outcomes, acquisition of sexually-transmitted infections and other vaginal disorders (5,6). Specific changes in the vaginal microflora are associated with BV, including a depletion of *Lactobacillus* species and an increased abundance of strictly anaerobic bacteria (7). However, no single bacterial taxon has been shown to cause BV and the condition can be found in women with widely varying vaginal microbiomes (8–11). BV is characterized clinically by itching, pain, burning, odor and/or discharge, and is often diagnosed based on a combination of symptoms, vaginal pH and cytological findings (12). BV is also diagnosed using the Nugent score, which is the most commonly used diagnostic test for BV within the research community (13).

Just as the microbial underpinnings of BV are complex and varied (14), so too are the influences of a woman's sexual, sanitary and other practices. A wide range of factors have been shown to increase BV risk, including smoking, douching, menstruation, and new sexual partners (15–18). Different ethnic groups exhibit varying rates of BV, leading to conclusions that intrinsic host factors may contribute to the condition (4), although the effect of confounders on this association has been questioned (19). Further complicating this picture is evidence that women from different ethnic groups tend to harbor different vaginal microflora, independent of BV status (8,20).

BV is therefore a multifactorial disease mediated by a complex interplay of host, microbial and

environmental factors. Fortunately, all three of these influences can be measured and evaluated

using a combination of 16S rRNA gene sequencing of vaginal samples and detailed

questionnaires of study participants. However, the analytical methods for identifying associations

between multivariate, often categorical metadata with counts of microbial taxa are either ill-

suited, contested or opaque (21,22). The most widely used analytical method for community-

level data involves ordination of normalized microbiome counts (e.g., principal coordinates

analysis [PCoA] and nonmetric multidimensional scaling [NMDS]) followed by statistical

significance testing of explanatory/metadata variables (e.g., ANOSIM, PERMANOVA).

However, these methods reduce the dimensionality of the microbial community structure to two

or three dimensions, obscuring intra-microbiome interactions. In addition, interactions between

explanatory (metadata) variables are difficult or impossible to uncover because statistical testing

usually occurs on a variable-by-variable basis (23). Therefore, these methods do not allow for

discovery of more nuanced and complex dynamics between and within the microbiome and host

and environmental factors. Identifying such dynamics can be crucial for understanding and thus

combatting multifactorial conditions such as bacterial vaginosis – particularly when such

conditions lack an accurate diagnostic description and/or test (24). Bayesian network (BN)

analysis offers potential advantages in handling mixed datasets for complex conditions, and use

of this approach may help to elucidate some of the nuanced, yet important, associations between

host, environmental and microbial factors in diseases such as BV. For instance, BN's have been

proposed as diagnostic tools for such multifactorial diseases as breast cancer and cardioembolic

stroke (25–27). While the heavy computational burden of building BNs has impeded their

widespread adoption, recent advances in algorithms have removed this barrier (28). In addition,

software implementations now offer BN analysis within user-friendly packages (29,30). Finally,

unlike other machine learning approaches, BN's possess an inherently intuitive interpretation and

allow for a wide range of data input types, both continuous and categorical (31).

The primary goal of this study was to uncover associations between host behavioral

characteristics, vaginal microbiome composition, risk symptoms and diagnostic criteria of BV

utilizing BN's. Using this approach, we have demonstrated associations between women's sexual

and menstrual habits, demographics, vaginal microbiome composition, risk symptoms of BV and

the Nugent Score (a BV diagnostic). Our findings support previously-documented associations

between microbiome members (e.g., *Eggerthella*, *Gardnerella*, *Dialister*, *Sneathia* and

Ruminococcaceae), the Nugent Score and vaginal pH (risk symptom) (8,13,32,33). However,

we found no connections between the Nugent Score and other risk symptoms of BV such as

vaginal discharge, itch, irritation, abdominal or pelvic pain, yeast infection and underwear

staining within 60 days prior to sampling and any type of current vaginal odor. This suggests that

the Nugent Score may not provide an accurate diagnostic of BV in some women. Additionally,

we found that several microbiome members were directly connected to risk symptoms of BV,

including *L. jensenii*, *Corynebacteria*, and *Proteobacteria*. Demographics (i.e., age, ethnicity,

previous pregnancy) also influenced the presence/absence of specific vaginal microbes. The

resulting network also revealed several as-yet undocumented associations between birth control

usage, menstrual hygiene practices and microbiome members. These associations require

confirmatory follow-up study, though strongly suggest that future studies of the vaginal

microbiome and vaginal pathologies include detailed surveys of participants' sanitary, sexual and

birth control habits, as these can act as confounders in the relationship between the microbiome

and disease.

Secondary objectives of this study included demonstrating the hypothesis-generating power of

BN's through confirmation of previously described microbiome-BV associations, as well as the

wide accessibility of BN's within a readily available, user-friendly environment. In addition, we

aimed to illustrate the capacity of the BN method to directly associate component taxa of the

microbiome with metadata. This capacity contrasts with common approaches used to analyze

these data types (e.g., PCoA, NMDS and clustering), which rely on reducing the dimensionality

of the total microbial community to two or three dimensions, followed by clustering or

correlation of this reduced community with metadata (such as demographics). The BN approach

also circumvents the limits on the number of interactions that can be tested between metadata

and microbiome members that are imposed by tools such as PERMANOVA (34) or

metagenomeSeq (35). This advantage of the BN approach is especially true when the number of

variables observed in association with the microbiome is large. In such cases, network analysis

identifies the hierarchy of relationships between the various metadata and microbiome taxa (i.e.,

the nodes of the network), and then represents these relationships using the arcs (or edges) of the

network. When possible, we demonstrated these differences by providing direct comparisons of

the BN approach with results obtained from Nonmetric Multidimensional Scaling Ordination

(NMDS)-Analysis of Similarity (ANOSIM) and PERMANOVA. These comparisons should be

interpreted with caution, as particularly the NMDS/ANOSIM approach is limited by the degrees

of freedom available for significance testing and for assessing interactions. In addition -- and

unlike the BN approach -- NMDS/ANOSIM and PERMANOVA analyses do not allow for the

possibility of hierarchical relationships between variables observed within the data.

It is important to note that the BN approach presented in this work was aimed at inference, rather

than prediction; and that we focused on evaluating associations within the data that could

indicate possible associations in the target population. Use of the BN approach in this way does

not attempt to classify the state of a new individual within the target population, i.e., prediction.

Hence our network was fitted using all data and we did not attempt to perform cross validation to

assess the ability of the network to predict or classify.

Materials and Methods

Study Population and Sampling

Questionnaire responses (see S1 File) and the relative abundance of 16S rRNA gene sequences

(see S1 File) were obtained from a study of 396 healthy, sexually active, non-pregnant women of

reproductive age (range 12-45 years); details of the study population have been described

previously (8). Briefly, study participants submitted 2 self-collected vaginal swabs; one swab

was used for 16S sequencing and the other was used to obtain a Nugent score, which is one of

several diagnostic assays for BV and is the most commonly used within the research community

(13). High Nugent scores are considered to be diagnostic of BV. Since all study participants

were considered to be healthy, those with high Nugent scores were assumed to have

asymptomatic BV. At the same time as vaginal swab collection, study participants were

administered a detailed questionnaire on their sexual and sanitary habits and health histories (8,

S2 File).

Ethics Statement

167   The institutional review boards at Emory University School of Medicine, Grady Memorial

168   Hospital and the University of Maryland School of Medicine approved the protocol. Guidelines

169   of the universities were followed in the conduct of the clinical research. Participants provided

170   written informed consent to participate in the study and written informed consent for use of the

171   data for future studies. The study was registered at clinicaltrials.gov under ID NCT00576797.

Data Preparation

Several filtering steps were taken with the goal of retaining only those variables (i.e., taxa,

survey responses and demographic information) that would provide robust information during

BN construction. In order to reach this goal, we removed taxa and survey response variables that

were sparsely represented within the dataset, as described below.

Ribosomal 16S sequencing data were processed using the Ribosomal Database Project (RDP)

Classifier (36) as described previously (8). In order to allow for robust BN construction, we

removed samples that contained <1,000 16S counts. In addition, subjects who were not in overall

general health or who had experienced toxic shock syndrome were removed from analysis. 16S

counts were then normalized to the sample with the lowest number of counts, and taxa with

counts <0.1% of total 16S counts were removed from further analysis to provide robust inputs

into BN analysis. Finally, 16S counts were converted into presence/absence (i.e., 0/1) data (S3

File). While the implementation of BN analysis used in this study, i.e., bnlearn (29), is able to

incorporate continuous variables with a Gaussian distribution, unfortunately the sparseness and

skewness of the 16S rRNA count distributions in this dataset were such that we could not justify

using abundances without dichotimization (see abundance distributions of used taxa in S8 File).

Furthermore, categorizing these distributions to include more than presence/absence categories

would have inflated the number of parameters that needed to be estimated, resulting in

diminished stability of the BN network given the available sample size.

From the survey data, we removed questions that were related solely to study design factors

(e.g., enrollment information, location of survey administration, survey administrator ID). Binary

questions about specific vaginal odors (i.e., fishy, musty, foul and other) were collapsed into a

single, binary "any odor" variable in order to decrease sparseness. In addition, we dropped

questions about past birth control use in order to focus on current birth control habits, which we

hypothesized would exhibit a stronger effect on the vaginal microbiome. Survey questions for

which >5% of respondents did not provide an answer were excluded from BN analysis so that

variables with a significant proportion of missing data did not exert undue influence over the

graph. For the same reason, and after this variable-level exclusion, we removed any participants

with missing data, keeping only those subjects with complete data in conjunction with the

variables under study. After these filtering steps, we discretized any remaining continuous

variables. Respondents' ages were converted from a continuous into a categorical variable with 3

levels: less than or equal to 30 (young adults), 31 – 40, and greater than 40 years of age (close to

menopause). Nugent scores were categorized into low (0-3), medium (4-6) and high (7-10). Age

at menstrual onset was discretized into <11, 11-15 and >15 years, and number of sexual partners

in the last 60 days was categorized into 0, 1 and >1. Number of vaginal births and duration of

menstruation were treated as categorical variables with 4 levels each (0 – 3 births and 1 – 4 days,

respectively). Finally, we excluded survey questions with sparse outcomes, defined as having

<5% of respondents within any one outcome level (i.e., for binary questions, at least 5% of

respondents had to be in the "yes" and "no" outcome levels). This was also done such that stable

BN's could be constructed. The resulting data are presented in S3 File and a key is available from

S4 File.

BN Construction

Filtered, dichotomized 16S rRNA gene counts as well as filtered and discretized demographic

and survey responses were provided as input to BN construction, which was performed using

bnlearn (29) implemented within the statistical software R (37). Age and ethnicity were

specified as roots of the network, and Nugent score was specified as a leaf. Directionality

between all other nodes was not specified in order to allow for the complex, likely

multidirectional interplay between the microbiome, host factors, vaginal microenvironment and

risk symptoms that might be associated with BV (12,14,38). The hill-climbing algorithm was

used for BN construction with scoring based on the logarithm of the Bayesian Dirichlet

equivalence (BDe) score; an optimal imaginary sample size was estimated using the alpha.star

function in bnlearn (29,39). To obtain a consensus network, 1,000,000 BN's were constructed by

first generating 1,000 random number seeds and, then, generating 1000 bootstraps of the input

data. The aim of varying the seeds was to alleviate the possibility of being stuck in a local

optimum in the space of all possible networks. The 1000 bootstraps per different starting point

were used to assess stability of the network given each starting point. All 1,000,000 networks

were then combined to construct a consensus BN by first using the mean function to pool the

bootstrapped networks and then by using the averaged.network function to construct the

consensus. Both of these functions are part of the bnlearn package. The empirical frequency for

every arc within all BN's was determined, and arcs that met an empirical threshold frequency of

0.30 or more were used to produce the consensus BN. Network analysis and inference was

performed using both this threshold and a 0.50 majority rule threshold. The resulting DAG was

used in maximum likelihood estimation parameter fitting through the bn.fit function, as well as

network analysis (29).

BN Analysis

Prior to BN analysis, nodes were categorized as either "demographic", "microbiome",

"sexual/sanitary habit", "BV risk symptom", or "BV diagnostic criteria" (S4 File). Variables

that were categorized as BV risk symptoms included "staining of underwear", "vaginal

discharge", "vaginal odor", "vaginal itch", "vaginal irritation", "abdominal or pelvic pain" and

"yeast infection" within the past 60 days prior to sampling, and pH and any type of current

vaginal odor (any odor). These variables where chosen based on previously observed

associations with increased risk of BV (6,12,40). Vaginal pH, vaginal discharge and staining,

and odor could be thought of as proxies for the Amsel symptoms used in BV diagnosis in clinical

settings (32). Given that all included subjects were considered healthy at the time of this study,

our intention was to highlight possible increased risk of BV due to presence of these risk

symptoms. The Nugent Score was the only variable designated as a "BV diagnostic criteria".

After *a priori* categorization, the nodes and edges obtained from bnlearn were used to conduct

network analysis in order to understand the structure of the overall network, as well as the

connections within it. Average Markov blanket size and overall graph characteristics were

calculated and reported. Network density was defined as the proportion of all possible edges that

occurred in the final network. Node degree was defined as the number of incoming and outgoing

edges, node closeness centrality was defined as the average shortest path from a node to all other

nodes, and node betweenness centrality was defined as the frequency with which a node was

included in the shortest path between nodes in the network. Node closeness and betweenness

centralities were normalized for comparison purposes. Groupings of nodes (i.e., groupings not

related to the *a priori* node category, but rather agnostic groupings) were detected using a

modularity optimizing algorithm (41) as implemented in Gephi (42), using a resolution of 1.0

and randomization without edge weights. Networks were visualized using Gephi (42), using the

Force Atlas layout unless otherwise specified.

261 **Ordination and Clustering**

Filtered, normalized, non-dichotomized, Hellinger-transformed (43) 16S rRNA gene counts were

used to construct a Euclidean distance matrix, which was then ordinated using NMDS with

multiple random starts and two dimensions (44). Use of two dimensions resulted in a stress value

of 0.19, indicating less-than-ideal fit; however, increasing the number of dimensions resulted in

failure of convergence, and therefore a better fit could not be achieved. Associations between the

ordinated microbiome and metadata variables (i.e., host demographics and survey data) were

tested using analysis of similarities (ANOSIM) (45). A *P*-value of less than 0.05 was considered

statistically significant.

PERMANOVA

Permutation based multivariate analysis of variance (PERMANOVA) was also used to assess the

influence of metadata on the vaginal microbiome. The process of assessing significance of

association followed a forward elimination, stepwise model selection approach based on

Akaike's Information Criteria (AIC) and using the step function as implemented in R.

PERMANOVA was performed onHellinger-transformed microbiome counts. All survey and

demographic response variables that passed the stepwise selection criteria (see above) and

minimized the AIC score were included in the final PERMANOVA model (implemented using

rda function in vegan). A permutation test using 1000 permutations was used to compute *P*-

values to assess significance of associations between the metadata and the observed microbiome.

Results and Discussion

Data filtering and descriptive statistics

Of the 396 participants included in the original study, 2 were removed *a priori* due to a lack of

any metadata from the questionnaire. Among the remaining 394 participants, 247 genus- and

species-level taxa were identified through 16S sequencing of vaginal swabs. Total 16S counts

per sample ranged from 693 to 7,392 with a median of 2,149. Five samples with <1,000 total 16S

counts were removed from the analysis, for a remaining 389 samples; all 247 taxa were

represented within these 389 samples. 16S counts were normalized to a count of 1,006, which

was the total number of 16S reads in the sample with the lowest total count. The distribution of

16S counts was strongly right-skewed, and therefore 220 taxa were present at <0.1% abundance

(i.e., fewer than an average of 2 reads per sample) and were removed from further analysis.

From the 389 study participants with 16S data that passed filtering, a further 4 participants were

removed because they answered "no" or did not answer the question of whether they were in

overall good health; and a further 9 were removed because they either answered "yes" to

previous toxic shock syndrome or did not answer this question. A further 91 respondents were

excluded from analysis due to missing data for survey questions. The decision to remove

subjects with missing data was not made lightly and was based on two reasons: 1) data

imputation, although useful, could be faulty or could reduce the variability of the estimates of the

network parameters (46) resulting in overconfidence in the network and 2) adding the missing

data as a category by itself would substantially hamper interpretability of results. Accordingly,

the decision was made to err on the conservative side (i.e., increased variability), rather than on

the side of overconfidence or weak interpretability. This left a total of 285 healthy study

participants for inclusion in BN analysis, encompassing a total of 27 taxa with a median

prevalence of 89/285 samples (31%, range 24 - 228 out of 285 samples, S3 File). Among the

285 participants included in the network, 58 self-reported to be Hispanic (20.4%), 71 Asian

(24.9%), 78 White (27.4%) and 78 African-American (27.4%). Nearly half of respondents were

age 30 or under (49.5%, 141/285), and only 25 reported being over age 40 (8.8%).

After removing study-specific questions, redundant questions, past birth control use questions,

questions about frequency of sexual habits and menstrual hygiene, as well as questions related to

study exclusion criteria, a total of 263 survey questions were included in the initial survey

response data. After removing questions with >5% missing responses, we were left with 56

study questions. A further 27 questions were removed due to sparseness, resulting in a total of 29

study questions available for input into the BN (S3 File). A majority of the 285 survey

respondents included in the Bayesian network reported having been pregnant at least once

(170/285, 59.6%). Sixty-three reported having had no sex partners in the 60 days preceding

sampling (22.1%), a further 199 were monogamous in this same time period (69.8%), and 23

respondents had multiple sex partners (8.1%). The majority of women reported an absence of

vaginal itching, irritation, discharge, odor or uro-abdominal pain in the 60 days preceding the

survey (177/285, 62.1%). Vaginal yeast infections in the 60 days preceding sampling were

reported by 19 of the 285 respondents. In addition to the 29 survey questions, we also had data

on the vaginal pH and Nugent score of the 285 women in the study. The majority of participants

had a low Nugent score (188/285, 66.0%), while a smaller proportion were categorized as

intermediate and high (34/285 and 63/285, 11.9% and 22.1%, respectively). Around half of the

285 women had a vaginal pH of less than or equal to 4.5 (152/285, 53.3%), while about half had

a higher vaginal pH (133/285, 46.7%).

Network construction

327 A total of 60 variables were included in BN construction: the 29 survey response variables and

328 27 bacterial taxa that passed filtering criteria, as well as age, ethnicity, vaginal pH and Nugent

329 score. The consensus BN had 152 directed edges with an average Markov blanket size of 9.4.

BN Characteristics

The final consensus graph contained 60 nodes within a single sparse network with a density of

0.043. The majority of nodes contained between 1 and 6 connections, while several were more

highly connected (Fig. 1). *Eggerthella* represented the most highly-connected node with 11

edges, followed by *Sneathia*, previous pregnancy status and current use of any type of birth

control with 10 edges each.

Figure 1. Histogram of node degree for the final BN. The majority of nodes had fewer than 8

connections (i.e., degrees), while a few were more highly connected.

Betweenness centrality measures indicated that *Parvimonas*, report of vaginal odor in the 60

days prior to the survey, as well as presence/absence of *L. vaginalis* within the vaginal

microbiome, were situated along highly-connected paths in the network [S4 File, see column

"Betweenness Centrality"]. Node closeness centrality metrics suggested that presence/absence

of *Lactobacillus iners*, *Mycoplasmataceae, Lachnospiraceae* and *Streptococcus* were all nodes at

the geographic center of the network, along with vaginal irritation in the 60 days preceding the

survey [S4 File, see column "Closeness Centrality"]. Interestingly, *Sneathia*, *Eggerthella*,

Parvimonas and *Streptococcus* have been described as key members of a vaginal microbiome

characterized by a diversity of non-*Lactobacillus* bacteria found more commonly in black and

Hispanic women (8,47). The fact that these bacteria are centrally situated in the BN developed in

this study (as defined by closeness, betweenness and degree metrics) lends support to the idea

that these bacteria could play an important role in differentiating the vaginal microbiome in

different women.

Ordination, Clustering and PERMANOVA

Univariable clustering analysis with ANOSIM significance testing revealed that six of the 33

metadata variables (including host demographics and survey responses) were statistically

significantly associated with the microbiome ordination results. This list comprised ethnicity,

vaginal pH, Nugent score, tampon use, as well as previous pregnancy and vaginal birth statuses

[S5 File]. S6 File provides the associated NMDS plots for these six variables. These analyses

suggested that demographic and behavioral traits of participants were associated with differences

in the microbiome; however, given the nature of these analyses, it was unclear specifically which

microbiome members were being affected, and whether such associations were direct or indirect.

PERMANOVA analysis, alternatively, revealed a best-fit model that also included vaginal pH,

Nugent score and previous pregnancy, as well as participant age, use of pads during

menstruation, staining of underwear in the previous 60 days, and use of male condoms as current

birth control method, which together explained 25% of the variability in the data (Table 1). This

type of analysis provided more information highlighting associations between high Nugent

Score, *Sneathia* and *Megasphaera* and possibly *Atopobium*, *Dialister*, *Eggerthella*,

Ruminococcaceae, and *Lachnospiraceae* (S7 Figure). It also showed putative association

between intermediate Nugent Score and *L. gasseri* (S7 Figure). Other relationships were tenuous

and not clearly defined. Furthermore, the above analyses precluded the testing of more

complicated interactions between multiple variables of diverse types. BN analysis was thus used

to identify such relationships within all of the data, including between individual microbiome

members; we point to the results of NMDS/ANOSIM and PERMANOVA where relevant.

Associations between population demographics, the microbiome and survey responses

There were 58 edges that bridged nodes of different types, i.e., demographics, sexual and

menstrual hygiene habits, risk symptoms of BV, diagnostic criteria and microbiome (see S4 File

for *a priori* classification of nodes); compared to 94 edges that connected nodes of like types. Of

these 58 bridging edges, 16 were identified in 50% of the bootstrapped BN networks (Fig. 2).

Many of these connections confirmed previously documented relationships between

demographic factors and sexual and menstrual habits. For instance, ethnicity was directly related

to Pap testing status, with Asian women exhibiting a much higher likelihood of never having had

a pap smear compared to Hispanic, White and Black respondents (20.8% versus 4.6%, 2.9%, and

1.2%, respectively), a disparity that has been described previously (48,49). At a threshold of at

least 50% bootstrap support, ethnicity also influenced tampon and pad use during menstruation

(50), while at 30% support ethnicity was associated with age at menstrual onset (51,52) and the

likelihood of a woman ever having self-treated a vaginal infection.

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Figure 2. Inter-category bridging edges with >50% bootstrap. Network displaying all nodes,

but depicting only inter-category bridging edges with >50% bootstrap support (arrows). Node

size is proportional to closeness centrality, arrow thickness is proportional to bootstrap support,

and node color signifies category type (orange = demographic, green = microbiome, turquoise =

BV risk symptom, purple = sexual/menstrual habits, and pink = diagnostic criteria of BV). Node

label abbreviations and categorization by type can be found in S4 File.

At 50% bootstrap support, there were no edges between demographic variables (i.e., age,

ethnicity, age at menstrual onset, previous pregnancy and vaginal birth) and microbiome nodes

(Fig. 2), suggesting that these factors may not exert robust influence over the presence of

common microbiome members. However, decreasing bootstrap support to 30% revealed four

such edges, namely associations between participant age and *Lactobacillus iners*, ethnicity and

Eggerthella, age at menstrual onset and *Ureaplasma*, and *Mycoplasmataceae* and period length

(Fig. 3). PERMANOVA testing supported the associations between the vaginal microbiome and

age, while NMDS/ANOSIM analysis supported the association between the microbiome and

ethnicity and marginally (ANOSIM $P = 0.06$) between the microbiome and age at menarch

(Table 1, S5-S7 Files). Additionally, PERMANOVA and NMDS/ANOSIM analyses revealed

associations between the vaginal microbiome and previous pregnancy and/or previous live birth

(Table 1, S5-S7 Files). Given the constraints of PERMANOVA and NMDS/ANOSIM analysis,

we were unable to identify which specific microbiome members may be involved in these

associations.

Figure 3. Network depicting nodes related to demographics (orange) and microbial taxa (green), as well as all edges with at least 30% bootstrap support. Node size is proportional to node degree (i.e., number of incoming and outgoing edges). Arrow thickness is proportional to bootstrap support. Red arrows bridge demographic and microbiome nodes, while green and orange arrows connect nodes of the same type (i.e., microbiome-to-microbiome or demographic-to-demographic). Node label abbreviations can be found in S4 File.

However, BN analysis revealed that the probability of harboring *L. iners* given age >40 years was 93%, compared to 72% and 85% for women ages 30-40 and <30 years, respectively. To date, longitudinal studies of the vaginal microbiome have been restricted to relatively short time periods, with subsequently little knowledge about how the microbiome may change with age. However, studies suggest that variations in estrogen levels influence presence/abundance of

Lactobacilli (53,54), and age has a significant influence on estrogen levels (55), which together

present a possible mechanism for the association we found between age and *L. iners*. Existence

of an edge between these two nodes suggests significance of association (affirmed by a Chi-

square test-of-independence, p-value 0.0067). BN analysis also revealed a complex relationship

between age and other demographic variables. Age was associated with previous pregnancy

status (which can also affect estrogen levels), as was ethnicity; indeed, previous pregnancy was a

common effect of age and ethnicity within this dataset (i.e., the three nodes formed a v-structure

within the graph). Only 24% of white respondents reported having ever been pregnant (19/78),

compared to 58%, 77% and 86% of Asians, African-Americans and Hispanics, respectively. The

role of age in likelihood of previous pregnancy was very strong among Asian and white

respondents, and less so for African-Americans and Hispanics. Ethnicity also influenced the

likelihood of identifying *Eggerthella* within the microbiome, as African-Americans were more

likely to harbor this bacterium (45.4% versus 18.8%, 16.5% and 3.8% for Hispanics, Caucasians

and Asians, respectively). Interestingly, *Eggerthella* was one of 28 taxa that were previously

found to exhibit a significant interaction with race and BV status (47), and was also a key

member of a certain vaginal microbiome composition that was more commonly found in black

and Hispanic women than white and Asian women (8). Finally, previous pregnancy decreased

the likelihood of a woman harboring *L. crispatus* and age at menarch was associated with

Ureasplasma presence. *Ureaplasma* presence was also influenced by pap smear status.

Ureaplasma in the upper genital tract has been associated with poor pregnancy outcomes, and

Ureaplasma colonization in the lower genital tract has been found to be associated with a variety

of socioeconomic and demographic factors including educational, income, ethnicity and marital

status (56–58). Interestingly, age at menarch has also been associated with socioeconomic-

related factors such as body mass index (BMI) and ethnicity (59,60), as has pap smear status

(48,49). Results such as these demonstrate the complexity of interactions between microbiome

composition and demographic and behavioral factors, and strongly suggest that any attempts to

associate microbial composition with clinical BV should include co-analysis of potential

confounders. The results presented here indicate that age, ethnicity and previous pregnancy

status could potentially be used as portmanteau variables for such confounders.

Upon modularity analysis, nodes within the network tended to segregate into groupings of nodes

of like type (called "communities" in network analysis), with some exceptions (Fig. 4). Nearly

all of the microbiome nodes fell into two largely bacteria-specific groupings, one of which

contained the nodes for Nugent score and vaginal pH. All of the *Lactobacilli* except *L. gasseri*

segregated along with nodes related to demographics (including study participant age, ethnicity,

pap smear status, age at menstrual onset and previous pregnancy), menstrual hygiene habits and

douching. All of the nodes related to risk symptoms associated with increased risk of BV, except

for pH, fell into one grouping that also included use of the withdrawal method for birth control,

and all of the other birth control variables were clustered into a fifth grouping that also included

the number of sexual partners in the previous 60 days (Fig. 4).

These modularity-based grouping results highlight the dissociation of the Nugent score (a BV

diagnostic) and vaginal pH from all other risk symptoms of BV, a finding which supports recent

studies suggesting that these may not be the most useful criteria for assessment of clinical BV

disease (8,33). None of the other risk symptom-based nodes fell into the same modularity

grouping as the Nugent score and vaginal pH, and indeed the BN contained no directed paths of

any length between vaginal pH or Nugent score and risk symptoms of vaginal irritation, itch,

odor, pain or staining. Furthermore, contingency table propagation showed that women

reporting vaginal odor, itch and irritation had an 11.6% chance of a high Nugent score, compared

to 11.0% for women who did not report vaginal odor, itch and irritation, again suggesting that

the Nugent score might not be reliably associated with these risk symptoms. In addition, while

bacteria traditionally thought to be determinants of the Nugent score did belong to the same

modularity grouping as the node for Nugent score (e.g., *Dialister* and *Gardnerella*), these

bacteria were not associated with BV risk symptoms other than pH (Fig. 4).

Figure 4: Network depicting modularity-based communities (node colors) as determined by

a modularity optimization algorithm (41). Size of node is proportional to node's betweenness

centrality. Node label abbreviations can be found in S4 File.

Associations between pH, Nugent Score and the microbiome

Several microbial taxa nodes were directly associated with vaginal pH and the Nugent Score.

Vaginal pH is one of the four Amsel criteria used to diagnose BV (32). A vaginal pH of less

than or equal to 4.5 is considered normal, while >4.5 is considered to contribute to BV.

Although all women in this dataset were healthy and did not have clinical BV as determined by

the Amsel criteria, nearly half had an "abnormal" vaginal pH >4.5 (133/285, 46.7%), a finding

which calls into question the use of vaginal pH as an indicator of clinical BV. As with previous

studies (61), NMDS/ANOSIM and PERMANOVA analyses both showed association between

the vaginal microbiome and vaginal pH (Table 1, S5-S7 Files). Using BN analysis, we were able

to reveal that vaginal pH was influenced by 4 bacteria: *Atopobium*, *Dialister, Gardnerella* and

Peptoniphilus (Fig. 5). These bacteria are all obligate or facultative anaerobes and the presence

of each of these bacteria in the vaginal microbiome increased the likelihood of a vaginal pH >

4.5. These four bacteria have been described as key members of a vaginal microbiome

community type characterized by a diversity of non-*Lactobacilli* bacteria that is found more

commonly in women with a high vaginal pH. It has been hypothesized that the higher vaginal pH

in such women is due to a comparatively low number of lactic acid producing bacteria (8), and

the results of this study support this hypothesis. As with previous research, however, the

presence of these bacteria and the subsequent increase in vaginal pH were reported in healthy

women without clinical BV.

Figure

5. Network depicting vaginal pH and Nugent Score (pink), and bacteria (green). Size of

circle is proportional to degree (i.e., number of incoming and outgoing edges), and edge

thickness is proportional to bootstrap support. Node label abbreviations can be found in S4 File.

The Nugent score is a diagnostic test used primarily in research studies of BV (13). It is rarely

used in practice because it involves time-consuming microscopic examination of bacterial

morphology from vaginal swabs. The Nugent score is based on the relative presence of Gram-

positive versus Gram-negative straight and curved rods, and therefore it is unsurprising that

NMDS/ANOSIM and PERMANOVA testing both uncovered associations between the vaginal

microbiome and the Nugent score (Table 1, S5-S7 Files). Using BN analysis, we were able to

assess this association more closely in order to uncover which specific microbiome members

were driving differences in the Nugent score. We found that the Nugent score for the 285

women in this dataset was influenced by bacteria from two Gram-positive bacteria (*Eggerthella*

and *Ruminococcaceae*), two Gram-negative bacteria (*Dialister* and *Sneathia*) and one Gram-

variable bacterium (*Gardnerella*) group. Vaginal pH was also connected to Nugent score;

women with a pH of >4.5 were much more likely to have a high Nugent score, a finding

consistent with previous literature (8,62). Amongst the 64 combinations of parent variable states

that directly influenced the Nugent Score, there were 6 combinations that conferred 100%

conditional probability of a high Nugent score; all 6 combinations included *Dialister*,

Ruminococcaceae and *Sneathia*, which have been found in higher abundance in women with a

high Nugent score (8, Fig. 6). Overall, these results confirm previous findings concerning the

robust association between certain vaginal microbiome members and vaginal pH and Nugent

score. However, the women in this study were BV-negative, and therefore the presence of high

vaginal pH, a high Nugent score and/or microbes associated with these indicators does not

always correlate with clinical BV (8). It is worth noting that the *Lactobacillus* species and the

Nugent Score were only conditionally independent given the presence of vaginal pH and

Sneathia—which were directly associated with *L. vaginalis* and indirectly with both *L. jensenii*

and *L. crispatus*—and *Ruminococcacea*, which shared a direct edge with *L. gasseri*. This

indicates a weaker association between the Nugent Score and the *Lactobacilli*, which was

masked by the presence of stronger associations, e.g., with vaginal pH. Removing some of these

directly-linked nodes from the network resulted in a direct association of the Nugent score with

some *Lactobacillus* nodes (data not shown).

525
Figure 6. Conditional probability of a high Nugent Score (y-axis) for all possible

combinations of direct parent variables (x-axis) (binary heatmap, black = present, gray =

absent).

Associations between the microbiome and risk symptoms other than

vaginal pH

While the women in this study did not have clinical BV, 108 of them did report experiencing at

least one risk symptom associated with BV within the 60 days prior to sample collection

(108/285, 38.9%). Vaginal yeast infections were reported by 19 of the 285 respondents during

the same time period. Most risk symptoms clustered together within the BN, with edges

connecting vaginal discharge to both vaginal irritation and vaginal itch; a fishy/musty vaginal

odor to staining of the underwear; any vaginal odor to vaginal itch; vaginal itch to vaginal

infection; and vaginal infection to yeast infection specifically. Staining of the underwear was

also directly connected to uro-abdominal pain in the preceding 60 days, which in turn was

connected to self-treatment of a vaginal infection. While the Nugent score and vaginal pH were

not connected to these risk symptoms, there were direct connections with specific microbial taxa.

Presence of *Lactobacillus jensenii* increased the likelihood of a woman reporting uro-abdominal

pain within the preceding 60 days, from a probability of 3.6% to 10.1% conditional on absence

or presence of *L. jensenii*, respectively. Interestingly, PERMANOVA analysis showed an

association between underwear staining in the previous 60 days and the vaginal microbiome

(Table 1); the results of BN analysis suggest that uro-abdominal pain and *L. jensenii* may be the

specific factors that link underwear staining and the microbiome. Use of pantyliners during

menstruation was also associated with *L. jensenii*, with the conditional probability of *L. jensenii*

being present in the vaginal microbiome increasing from 37.4% to 60.7% with use of panty

liners during menstruation. While the directionality of this relationship is not immediately

intuitive, previous work has shown that use of emollient pads changes the vaginal epithelium and

that some women's vaginal microflora does shift with pad versus tampon use (63,64), although

the evidence on this is mixed (65). NMDS/ANOSIM testing indicated that tampon use was

associated with the vaginal microbiome, lending further support to the hypothesis that menstrual

habits could impact the microbiome (S5 and S6 Files). Alternatively, the support for edge

directionality in the Bayesian network was based on majority rule and therefore directionality of

arcs could potentially be reversed; in such a scenario, presence of *L. jensenii* could increase the

likelihood of uro-abdominal pain, and thus underwear staining and pantyliner use. Furthermore,

ethnicity was a potential confounder in the relationship between the microbiome and menstrual

habits, as the network showed an influence of ethnicity on pad versus tampon use, which in turn

directly affected pantyliner use (Fig. 4). The relationships between uro-abdominal pain,

underwear staining, pantyliner use and *L. jensenii* deserve closer study given the importance of

this microbe in vaginal health.

Other connections between bacteria and risk symptoms of itch, irritation, odor, staining, pain or

yeast infection included use of a pad outside of menstruation increasing the conditional

likelihood of a woman harboring *Proteobacteria* from 2.8% to 6.7%, as extra-menstrual pad use

could be considered a proxy for vaginal discharge, staining or odor. Because *Proteobacteria* is

such a diverse phylum that includes pathogens and non-pathogens, it is difficult to formulate

hypotheses concerning these relationships. *Corynebacterium* seemed to play an important role in

symptomology and sexual behaviors, as it was directly connected to vaginal odor and number of

sexual partners in the 60 days preceding the survey. Women with this bacterium in their vaginal

flora were more likely to have reported both no or multiple sexual partners, and were less likely

to have reported a single sexual partner, while presence of *Corynebacterium* increased the risk of

vaginal odor in the 60 days preceding the survey. The association of *Corynebacterium* with

vaginal pathology is not widely described in the literature, although it has been recognized as an

important vaginal community member for several decades (66). More recently, studies indicate

that this bacterium is more prevalent in women with spinal cord injuries, women with treatment-

refractory BV, and women with urgency urinary incontinence (67–69). Depending on the

species, *Corynebacterium* can be fermentative, and many species produce amino acids, which

can alter pH and the metabolite profile of the surrounding environment, thus providing a

potential mechanism of *Corynebacterium's* role in vaginal odor and pathology. These

associations were not and would not have been easily observed using NMDS/ANOSIM and

PERMANOVA.

Presence of *Bacteroides* had a protective effect against vaginal discharge; the probability of

reporting discharge given the presence of *Bacteroides* was 2.0%, compared to 28.9% given

absence of these bacteria within the vaginal microbiome. This runs counter to current notions

about *Bacteroides*, an anerobic bacteria which is considered a commensal of the gastrointestinal

tract and has been implicated in BV (70,71). However, not all vaginal discharge should be

considered abnormal (72,73), and therefore it would not be a contradiction for *Bacteroides* to be

associated with a decreased prevalence of "healthy" vaginal discharge (such as cervical mucous

released during ovulation) and an increased prevalence of clinical BV. Distinctions such as these

will become increasingly important as the scientific community attempts to attain a more

nuanced understanding of connections between the microbiome and true clinical disease.

Microbiome-Behavior Connections

Sexual behavior, sanitary practices and microbiome composition have all been associated with

BV as separate factors. However, little is known about how a woman's sexual and sanitary

habits may influence the microbiome, or vice versa; and how this interplay may influence the

likelihood of developing BV. The network resulting from this dataset showed several direct

connections between the study participants' habits and the microbiome. Whether or not women

had ever had a pap smear was directly related to presence/absence of *Ureaplasma*. Women who

reported having had a pap demonstrated a 34.1% likelihood of harboring *Ureaplasma*, compared

to 18.0% among women who had never had a pap smear. Having had a Pap test was the only

healthcare access question included in the Bayesian network that could be considered a proxy for

socioeconomic status (48).

Microbiome-Birth Control Interactions

Use of certain types of birth control could influence the microbiome by modulating hormone

levels or by introducing chemical or physical barriers into the vaginal environment. Use of an

intrauterine device (IUD) directly influenced the presence/absence of two different bacteria,

namely *Lactobacillus gasseri* and *Sneathia*; in both cases, use of an IUD decreased the

likelihood of finding the bacterium within the vaginal microbiome. Previous research on the

effects of IUD use on vaginal microflora has been mixed, with both copper and hormone IUDs

associated with higher levels of *Candida* fungi (74) but no associations with any bacterial

alterations (74–76). Given the location of IUD use as a relatively highly-connected intercategory

node in the Bayesian network, as well as its potential association with nonspecific vaginitis and

recurrent BV (32,38,68), the associations with *L. gasseri* and *Sneathia* warrant follow-up study.

Women who reported ever using any form of birth control were more likely to have

Lactobacillus crispatus within their microbiome compared to women who had never used birth

control (conditional probability of 56.6% versus 48.1%, respectively). Women who reported

male condom use in the 60 days preceding the survey were less likely to harbor *Lachnospiraceae*

than women who did not (conditional probability of 1.3% versus 9.9%). Interestingly,

PERMANOVA testing also showed a connection between male condom use in the previous 60

days and the overall composition of the vaginal microbiome (Table 1); the BN analysis allowed

us to pinpoint this relationship to *Lachnospiraceae* specifically. As with other associations, there

were some directed edges that did not immediately fit the paradigm for host-microbe

interactions. For example, IUD use and abstinence were influenced by status of *Parvimonas* and

L. vaginalis within the vagina, respectively. It is difficult to hypothesize a mechanism by which

vaginal bacteria could influence a woman's birth control decisions, even though recent studies

have challenged the paradigm that host-microbiome interactions are largely uni-directional by

suggesting that microbiome composition can directly influence host behavior (77–79). This

highlights the importance of remembering that the directionality of edges in the BN are not

definitive.

Microbiome-Microbiome Interactions

Within the entire network, bacterial nodes fell largely within 2 modularity-based groupings,

with some exceptions (Fig. 4). Upon isolation of bacterial nodes in a separate graph and

subsequent modularity optimization, four communities of bacteria became evident (Fig. 7).

These groupings did not seem to segregate by either Gram stain, cellular metabolism, or any

other known characteristics. These groupings could be driven by as-yet undiscovered microbial

relationships within the vaginal environment and/or by unmeasured, non-microbiome factors in

this dataset.

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Figure 7. Network depicting only microbiome nodes, colored by modularity-based

community type, as determined using a modularity optimizing algorithm (41). Node size is

proportional to betweenness centrality, and edge thickness is proportional to bootstrap support.

Node label abbreviations can be found in S4 File.

With the exception of *L. iners* and *L. gasseri*, the *Lactobacilli* formed a tight cluster to

themselves. The fact that *L. iners* segregated separately from the other *Lactobacilli* is interesting

given that this organism differs substantially in terms of genomic and metabolic characteristics,

and its role in vaginal health has been debated despite its very high prevalence (47,80). One clue

in this debate could be the tight association between *L. iners* and *Ureaplasma*, the latter of which

has been implicated in several female reproductive conditions including chorioamnionitis leading

to premature delivery and pelvic inflammatory disease (81,82). Presence of *L. iners* increased the

likelihood of *Ureaplasma* from 19.4% to 36.3%, a significant increase that could have health

implications and therefore deserves follow-up study.

Conclusions

Using widely accessible software, we have demonstrated the utility of applying a Bayesian

Network approach to a multi-dimensional microbiome dataset. Using this approach, we have

demonstrated associations between women's sexual and menstrual habits, demographics, vaginal

microbiome composition and symptoms and diagnostics of BV. Many of these associations

suggested intriguing relationships, indicating that the BN approach is able to highlight important

associations within complex datasets, which can then be used for hypothesis generation. While

follow-up studies are needed to investigate the significance of these novel associations, the

validity of the associations was buttressed by the presence of many well-documented and self-

evident connections within the overall BN. For instance, our BN confirmed the importance of

vaginal pH and *Gardnerella* as influencers on the Nugent Score (Fig. 4). In addition, we found a

very strong association between previous pregnancy, vaginal birth and tubal ligation. Indeed, all

17 participants who reported tubal ligation had previously been pregnant. The most recent data

available suggest that nearly half of tubal ligations were performed in the postpartum period and

that younger women were more likely to have tubal ligation performed post-partum (83). As

with the BN produced here, this suggests a tight association between tubal ligation and previous

pregnancy. Another confirmatory association showed that women who reported vaginal itch in

the 60 days preceding the survey were much more likely to also have reported a yeast infection

in the same time period. Vaginal itch has been closely associated with vulvovaginitis candidiasis

(yeast infection) (84). While relationships such as these do not provide novel insight, they can be

used as a "sanity check" in order to gain confidence in the structure of the BN to then move on to

analysis of less well-understood network connections.

An advantage of the BN approach to datasets such as the one presented here is that BN

algorithms are now easily accessible through well-documented and well-supported packages

such as bnlearn (29), which has been integrated with the packages "parallel" (which is included

in R-core) and "snow" (85) to support multi-threading for construction of large networks, and

which now supports both categorical and continuous variables. Numerous GUI-based packages

exist to support network visualization once the BN has been constructed (42,86), allowing for

interactive graph exploration in an intuitive interface. Given the increasing use of multi-level,

multivariate microbiome datasets, this accessibility should pave the way for more scientists to

implement the BN analysis for such data. The need for such an approach has been demonstrated

in the complexity of the relationships that we found within and between the vaginal microbiome,

women's sexual and menstrual habits, demographics and symptoms/diagnostics. While non-

network approaches such as PERMANOVA and ordination/clustering can be used to uncover

associations between individual metadata variables and the microbiome as a whole (as we

demonstrate in this work), there are important limitations to these methods: first, these methods

collapse the microbiome data into a single datapoint projected into two or a maximum of three

dimensions, and therefore preclude assessment of intra-microbiome interactions, as well as

interactions between metadata variables and specific microbiome members; second, these

methods usually approach statistical testing on a variable-by-variable basis, and are thus ill-

suited to understanding complex interactions between subsets of variables and the microbiome;

and third, these approaches fail to show intra-metadata interactionswithin an analysis. BN

analysis overcomes these limitations by testing for all possible associations between every node

within the data – thus uncovering complex interactions within hierarchical, multidimensional

datasets. Such datasets increasingly typify microbiome studies, and the need for rich and

standardized metadata to support microbiome studies has been recently noted and implemented

(87,88); such initiatives would be well-complemented by the data-flexible and intuitive nature of

BN's. It is important to note, however, that the incorporation of more metadata variables (nodes)

into BN analyses will necessitate larger sample sizes in order to support BN construction and

inference. Similarly, the data distributions currently accommodated by available BN analysis

packages for continuous variables are limited to the Normal (Gaussian) distribution. This

constraint results in the need to discretize most microbiome data, which are commonly

characterized by zero-inflated, skewed counts that do not conform to a Gaussian distribution. As

was the case in this report, the need to dichotomize may cause loss of information, resulting in

less-than-optimally refined inference. Fortunately, BN analysis methods form an active area of

research, and extensions to accommodate distributions other than the Gaussian are currently

being developed (89). As these efforts evolve, the BN approach is likely to become more

flexible, and thus even better-suited to the unique characteristics of microbiome datasets.

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References

1.   Donders GG, Bosmans E, Dekeersmaecker A, Vereecken A, Van Bulck B, Spitz B.
Pathogenesis of abnormal vaginal bacterial flora. Am J Obstet Gynecol. 2000
Apr;182(4):872–8.

2.   Pybus V, Onderdonk AB. Microbial interactions in the vaginal ecosystem, with
emphasis on the pathogenesis of bacterial vaginosis. Microbes Infect Inst Pasteur.
1999 Apr;1(4):285–92.

3.   Martin HL, Richardson BA, Nyange PM, Lavreys L, Hillier SL, Chohan B, et al. Vaginal
lactobacilli, microbial flora, and risk of human immunodeficiency virus type 1 and
sexually transmitted disease acquisition. J Infect Dis. 1999 Dec;180(6):1863–8.

4.   Allsworth JE, Peipert JF. Prevalence of bacterial vaginosis: 2001-2004 National Health
and Nutrition Examination Survey data. Obstet Gynecol. 2007 Jan;109(1):114–20.

5.   Zhou X, Brotman RM, Gajer P, Abdo Z, Schüette U, Ma S, et al. Recent advances in
understanding the microbiology of the female reproductive tract and the causes of
premature birth. Infect Dis Obstet Gynecol. 2010;2010:737425.

6.   Bautista CT, Wurapa E, Sateren WB, Morris S, Hollingsworth B, Sanchez JL. Bacterial
vaginosis: a synthesis of the literature on etiology, prevalence, risk factors, and
relationship with chlamydia and gonorrhea infections. Mil Med Res. 2016;3:4.

7.   Eschenbach DA, Davick PR, Williams BL, Klebanoff SJ, Young-Smith K, Critchlow CM, et
al. Prevalence of hydrogen peroxide-producing Lactobacillus species in normal
women and women with bacterial vaginosis. J Clin Microbiol. 1989 Feb;27(2):251–6.

8.   Ravel J, Gajer P, Abdo Z, Schneider GM, Koenig SSK, McCulle SL, et al. Vaginal
microbiome of reproductive-age women. Proc Natl Acad Sci. 2011 Mar
15;108(Supplement 1):4680–7.

9.   Fredricks DN, Fiedler TL, Marrazzo JM. Molecular Identification of Bacteria Associated
with Bacterial Vaginosis. N Engl J Med. 2005 Nov 3;353(18):1899–911.

10.   Ferris MJ, Norori J, Zozaya-Hinchliffe M, Martin DH. Cultivation-Independent Analysis
of Changes in Bacterial Vaginosis Flora Following Metronidazole Treatment. J Clin
Microbiol. 2007 Mar;45(3):1016–8.

11.   Lamont RF, Sobel JD, Akins RA, Hassan SS, Chaiworapongsa T, Kusanovic JP, et al. The
vaginal microbiome: new information about genital tract flora using molecular based
techniques. BJOG Int J Obstet Gynaecol. 2011 Apr;118(5):533–49.

12.   Livengood CH. Bacterial Vaginosis: An Overview for 2009. Rev Obstet Gynecol.
2009;2(1):28–37.

13.   Nugent RP, Krohn MA, Hillier SL. Reliability of diagnosing bacterial vaginosis is
improved by a standardized method of gram stain interpretation. J Clin Microbiol.
1991 Feb;29(2):297–301.

14.   Ma B, Forney LJ, Ravel J. The vaginal microbiome: rethinking health and diseases.
Annu Rev Microbiol. 2012;66:371–89.

15.   Beigi RH, Wiesenfeld HC, Hillier SL, Straw T, Krohn MA. Factors associated with
absence of H2O2-producing Lactobacillus among women with bacterial vaginosis. J
Infect Dis. 2005 Mar 15;191(6):924–9.

16.   Brotman RM, Klebanoff MA, Nansel TR, Andrews WW, Schwebke JR, Zhang J, et al. A
Longitudinal Study of Vaginal Douching and Bacterial Vaginosis—A Marginal
Structural Modeling Analysis. Am J Epidemiol. 2008 Jul 15;168(2):188–96.

17.   Hellberg D, Nilsson S, Mårdh PA. Bacterial vaginosis and smoking. Int J STD AIDS. 2000
Sep;11(9):603–6.

18.   Hay PE, Ugwumadu A, Chowns J. Sex, thrush and bacterial vaginosis. Int J STD AIDS.
1997 Oct;8(10):603–8.

19.   Ness RB, Hillier S, Richter HE, Soper DE, Stamm C, Bass DC, et al. Can known risk
factors explain racial differences in the occurrence of bacterial vaginosis? J Natl Med
Assoc. 2003 Mar;95(3):201–12.

20.   Zhou X, Hansmann MA, Davis CC, Suzuki H, Brown CJ, Schütte U, et al. The vaginal
bacterial communities of Japanese women resemble those of women in other racial
groups. FEMS Immunol Med Microbiol. 2010 Mar;58(2):169–81.

21.   Warton DI, Wright ST, Wang Y. Distance-based multivariate analyses confound
location and dispersion effects. Methods Ecol Evol. 2012 Feb 1;3(1):89–101.

22.   Gloor GB, Reid G. Compositional analysis: a valid approach to analyze microbiome
772   high-throughput sequencing data. Can J Microbiol. 2016 Apr 12;62(8):692–703.

773   23.   Xia Y, Sun J. Hypothesis testing and statistical analysis of microbiome. Genes Dis
774   [Internet]. 2017 Jun 23 [cited 2017 Aug 27]; Available from:
775   http://www.sciencedirect.com/science/article/pii/S2352304217300351

776   24.   Shankar J. Insights into study design and statistical analyses in translational
777   microbiome studies. Ann Transl Med [Internet]. 2017 Jun [cited 2017 Aug 27];5(12).
778   Available from: http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5497086/

779   25.   Ramoni RB, Himes BE, Sale MM, Furie KL, Ramoni MF. Predictive Genomics of
780   Cardioembolic Stroke. Stroke J Cereb Circ. 2009 Mar;40(3 Suppl):S67–70.

781   26.   Gevaert O, Smet FD, Timmerman D, Moreau Y, Moor BD. Predicting the prognosis of
782   breast cancer by integrating clinical and microarray data with Bayesian networks.
783   Bioinformatics. 2006 Jul 15;22(14):e184–90.

784   27.   Cruz-Ramírez N, Acosta-Mesa HG, Carrillo-Calvet H, Nava-Fernández LA, Barrientos-
785   Martínez RE. Diagnosis of breast cancer using Bayesian networks: a case study.
786   Comput Biol Med. 2007 Nov;37(11):1553–64.

787   28.   Gámez JA, Mateo JL, Puerta JM. Learning Bayesian networks by hill climbing: efficient
788   methods based on progressive restriction of the neighborhood. Data Min Knowl
789   Discov. 2010 May 11;22(1-2):106–48.

790   29.   Scutari M. Learning Bayesian Networks with the bnlearn R Package. J Stat Softw.
791   2010;35(3):1–22.

792   30.   Højsgaard S. Graphical Independence Networks with the gRain Package for R. J Stat
793   Softw. 2012;46(10):1–26.

794   31.   Uusitalo L. Advantages and challenges of Bayesian networks in environmental
795   modelling. Ecol Model. 2007 May 10;203(3–4):312–8.

796   32.   Amsel R, Totten PA, Spiegel CA, Chen KC, Eschenbach D, Holmes KK. Nonspecific
797   vaginitis. Diagnostic criteria and microbial and epidemiologic associations. Am J Med.
798   1983 Jan;74(1):14–22.

799   33.   Gajer P, Brotman RM, Bai G, Sakamoto J, Schütte UME, Zhong X, et al. Temporal
800   Dynamics of the Human Vaginal Microbiota. Sci Transl Med. 2012 May
801   2;4(132):132ra52–132ra52.

802   34.   Anderson MJ. A new method for non‐parametric multivariate analysis of variance.
803   Austral Ecol. 2001 Feb 1;26(1):32–46.

804   35.   Paulson JN, Stine OC, Bravo HC, Pop M. Differential abundance analysis for microbial
805   marker-gene surveys. Nat Methods. 2013 Dec;10(12):1200–2.

806   36.   Wang Q, Garrity GM, Tiedje JM, Cole JR. Naïve Bayesian Classifier for Rapid Assignment
807   of rRNA Sequences into the New Bacterial Taxonomy. Appl Environ Microbiol. 2007
808   Aug 15;73(16):5261–7.

809   37.   R Core Team. R: A language and environment for statistical computing [Internet].
810   Vienna, Austria: R Foundation for Statistical Computing; 2016. Available from:
811   https://www.R-project.org/

812   38.   Li X-D, Wang C-C, Zhang X-J, Gao G-P, Tong F, Li X, et al. Risk factors for bacterial
813   vaginosis: results from a cross-sectional study having a sample of 53,652 women. Eur J
814   Clin Microbiol Infect Dis Off Publ Eur Soc Clin Microbiol. 2014 Sep;33(9):1525–32.

815   39.   Steck H. Learning the Bayesian Network Structure: Dirichlet Prior versus Data. Proc
816   24th Conf Uncertain Artif Intell. 2008;511–8.

817   40.   Nwankwo TO, Aniebue UU, Umeh UA. Syndromic Diagnosis in Evaluation of Women
818   with Symptoms of Vaginitis. Curr Infect Dis Rep. 2017 Jan 1;19(1):3.

819   41.   Blondel VD, Guillaume J-L, Lambiotte R, Lefebvre E. Fast unfolding of communities in
820   large networks. J Stat Mech Theory Exp. 2008 Oct 9;2008(10):P10008.

821   42.   Bastian M, Heymann S, Jacomy M. Gephi: An Open Source Software for Exploring and
822   Manipulating Networks. In San Jose, California, USA; 2009.

823   43.   Legendre P, Gallagher ED. Ecologically meaningful transformations for ordination of
824   species data. Oecologia. 2001 Oct;129(2):271–80.

825   44.   Oksanen J, Blanchet FG, Kindt R, Legendre P, Minchin PR, O'Hara RB, et al. vegan:
826   Community Ecology Package [Internet]. 2014. Available from: http://CRAN.R-
827   project.org/package=vegan

828   45.   Clarke KR. Non-parametric multivariate analyses of changes in community structure.
829   Aust J Ecol. 1993 Mar 1;18(1):117–43.

830   46.   Gelman A, Hill J. Data analysis using regression and multivariate/hierarchical models.
831   New York, NY: Cambridge University Press; 2007. 625 p. (Analytical Methods for
832   Social Research).

833   47.   Srinivasan S, Hoffman NG, Morgan MT, Matsen FA, Fiedler TL, Hall RW, et al. Bacterial
834   Communities in Women with Bacterial Vaginosis: High Resolution Phylogenetic
835   Analyses Reveal Relationships of Microbiota to Clinical Criteria. PLOS ONE. 2012 Jun
836   18;7(6):e37818.

48. Monnat SM. Race/Ethnicity and the Socioeconomic Status Gradient in Women's Cancer
Screening Utilization: A Case of Diminishing Returns? J Health Care Poor Underserved.
2014 Feb;25(1):332–56.

49. Sauer AG, Jemal A, Simard EP, Fedewa SA. Differential uptake of recent Papanicolaou
testing by HPV vaccination status among young women in the United States, 2008–
2013. Cancer Epidemiol. 2015 Aug;39(4):650–5.

50. Romo LF, Berenson AB. Tampon use in adolescence: differences among European
American, African American and Latina women in practices, concerns, and barriers. J
Pediatr Adolesc Gynecol. 2012 Oct;25(5):328–33.

51. Anderson SE, Dallal GE, Must A. Relative weight and race influence average age at
menarche: results from two nationally representative surveys of US girls studied 25
years apart. Pediatrics. 2003 Apr;111(4 Pt 1):844–50.

52. Chumlea WC, Schubert CM, Roche AF, Kulin HE, Lee PA, Himes JH, et al. Age at
menarche and racial comparisons in US girls. Pediatrics. 2003 Jan;111(1):110–3.

53. Nunn KL, Forney LJ. Unraveling the Dynamics of the Human Vaginal Microbiome. Yale
J Biol Med. 2016 Sep 30;89(3):331–7.

54. Romero R, Hassan SS, Gajer P, Tarca AL, Fadrosh DW, Nikita L, et al. The composition
and stability of the vaginal microbiota of normal pregnant women is different from
that of non-pregnant women. Microbiome. 2014 Feb 3;2(1):4.

55. Horstman AM, Dillon EL, Urban RJ, Sheffield-Moore M. The Role of Androgens and
Estrogens on Healthy Aging and Longevity. J Gerontol A Biol Sci Med Sci. 2012
Nov;67(11):1140–52.

56. Tibaldi C, Cappello N, Latino MA, Polarolo G, Masuelli G, Cavallo F, et al. Maternal risk
factors for abnormal vaginal flora during pregnancy. Int J Gynaecol Obstet Off Organ
Int Fed Gynaecol Obstet. 2016 Apr;133(1):89–93.

57. Carey J, Blackwelder W, Nugent R, Matteson M, Rao A, Eschenbach D, et al. Antepartum
Cultures for Ureaplasma-Urealyticum Are Not Useful in Predicting Pregnancy
Outcome. Am J Obstet Gynecol. 1991 Mar;164(3):728–33.

58. Goldenberg RL, Hauth JC, Andrews WW. Intrauterine Infection and Preterm Delivery.
N Engl J Med. 2000 May 18;342(20):1500–7.

59. Freedman DS, Khan LK, Serdula MK, Dietz WH, Srinivasan SR, Berenson GS. Relation of
age at menarche to race, time period, and anthropometric dimensions: the Bogalusa
Heart Study. Pediatrics. 2002 Oct;110(4):e43.

60.   Wu T, Mendola P, Buck GM. Ethnic differences in the presence of secondary sex
characteristics and menarche among US girls: the Third National Health and Nutrition
Examination Survey, 1988-1994. Pediatrics. 2002 Oct;110(4):752–7.

61.   Ravel J, Gajer P, Abdo Z, Schneider GM, Koenig SSK, McCulle SL, et al. Vaginal
microbiome of reproductive-age women. Proc Natl Acad Sci. 2011 Mar
15;108(Supplement 1):4680–7.

62.   Hoffman MK, Bellad MB, Charantimath US, Kavi A, Nagmoti JM, Nagmoti MB, et al. A
Comparison of Colorimetric Assessment of Vaginal pH with Nugent Score for the
Detection of Bacterial Vaginosis. Infect Dis Obstet Gynecol. 2017 Feb
15;2017:e1040984.

63.   Farage MA, Miller KW, Azuka CE, Wang B, Lankhof S, Henn D, et al. Skin benefits with a
novel emollient-treated menstrual pad. Rev Recent Clin Trials. 2013 Mar;8(1):29–35.

64.   Hickey RJ, Abdo Z, Zhou X, Nemeth K, Hansmann M, Osborn TW, et al. Effects of
tampons and menses on the composition and diversity of vaginal microbial
communities over time. BJOG Int J Obstet Gynaecol. 2013 May;120(6):695–704;
discussion 704–6.

65.   Klebanoff MA, Nansel TR, Brotman RM, Zhang J, Yu K-F, Schwebke JR, et al. Personal
hygienic behaviors and bacterial vaginosis. Sex Transm Dis. 2010 Feb;37(2):94–9.

66.   McCormack WM, Hayes CH, Rosner B, Evrard JR, Crockett VA, Alpert S, et al. Vaginal
colonization with Corynebacterium vaginale (Haemophilus vaginalis). J Infect Dis.
1977 Dec;136(6):740–5.

67.   Pires CVG, Linhares IM, Serzedello F, Fukazawa EI, Baracat EC, Witkin SS. Alterations
in the Genital Microbiota in Women With Spinal Cord Injury. Obstet Gynecol. 2016
Feb;127(2):273–8.

68.   Wang B, Xiao BB, Shang CG, Wang K, Na RS, Nu XX, et al. Molecular analysis of the
relationship between specific vaginal bacteria and bacterial vaginosis metronidazole
therapy failure. Eur J Clin Microbiol Infect Dis Off Publ Eur Soc Clin Microbiol. 2014
Oct;33(10):1749–56.

69.   Pearce MM, Hilt EE, Rosenfeld AB, Zilliox MJ, Thomas-White K, Fok C, et al. The female
urinary microbiome: a comparison of women with and without urgency urinary
incontinence. mBio. 2014 Jul 8;5(4):e01283–01214.

70.   Verhelst R, Verstraelen H, Claeys G, Verschraegen G, Delanghe J, Van Simaey L, et al.
Cloning of 16S rRNA genes amplified from normal and disturbed vaginal microflora
suggests a strong association between Atopobium vaginae, Gardnerella vaginalis and
bacterial vaginosis. BMC Microbiol. 2004 Apr 21;4:16.

71. Machado D, Castro J, Palmeira-de-Oliveira A, Martinez-de-Oliveira J, Cerca N. Bacterial
Vaginosis Biofilms: Challenges to Current Therapies and Emerging Solutions. Front
Microbiol [Internet]. 2016 [cited 2017 Feb 10];6. Available from:
http://journal.frontiersin.org/article/10.3389/fmicb.2015.01528/abstract

72. Zinaman MJ. Using cervical mucus and other easily observed biomarkers to identify
ovulation in prospective pregnancy trials. Paediatr Perinat Epidemiol. 2006 Nov;20
Suppl 1:26–9.

73. Powell AM, Nyirjesy P. New Perspectives on the Normal Vagina and Noninfectious
Causes of Discharge. Clin Obstet Gynecol. 2015 Sep;58(3):453–63.

74. Erol O, Simavlı S, Derbent AU, Ayrım A, Kafalı H. The impact of copper-containing and
levonorgestrel-releasing intrauterine contraceptives on cervicovaginal cytology and
microbiological flora: a prospective study. Eur J Contracept Reprod Health Care Off J
Eur Soc Contracept. 2014 Jun;19(3):187–93.

75. Jacobson JC, Turok DK, Dermish AI, Nygaard IE, Settles ML. Vaginal microbiome
changes with levonorgestrel intrauterine system placement. Contraception. 2014
Aug;90(2):130–5.

76. Donders G, Bellen G, Janssens D, Van Bulck B, Hinoul P, Verguts J. Influence of
contraceptive choice on vaginal bacterial and fungal microflora. Eur J Clin Microbiol
Infect Dis Off Publ Eur Soc Clin Microbiol. 2017 Jan;36(1):43–8.

77. Fischer CN, Trautman EP, Crawford JM, Stabb EV, Handelsman J, Broderick NA.
Metabolite exchange between microbiome members produces compounds that
influence Drosophila behavior. eLife. 2017 09;6.

78. Bharwani A, Mian MF, Surette MG, Bienenstock J, Forsythe P. Oral treatment with
Lactobacillus rhamnosus attenuates behavioural deficits and immune changes in
chronic social stress. BMC Med. 2017 Jan 11;15(1):7.

79. Yang NJ, Chiu IM. Bacterial Signaling to the Nervous System through Toxins and
Metabolites. J Mol Biol. 2017 Jan 6;

80. Petrova MI, Reid G, Vaneechoutte M, Lebeer S. Lactobacillus iners: Friend or Foe?
Trends Microbiol. 2016 Nov 30;

81. Sweeney EL, Kallapur SG, Gisslen T, Lambers DS, Chougnet CA, Stephenson S-A, et al.
Placental Infection With Ureaplasma species Is Associated With Histologic
Chorioamnionitis and Adverse Outcomes in Moderately Preterm and Late-Preterm
Infants. J Infect Dis. 2016 Apr 15;213(8):1340–7.

82. Haggerty CL, Totten PA, Tang G, Astete SG, Ferris MJ, Norori J, et al. Identification of
novel microbes associated with pelvic inflammatory disease and infertility. Sex
Transm Infect. 2016 Sep;92(6):441–6.

83.   MacKay AP, Kieki BA, Koonin LM, Beattie K. Tubal Sterilization in the United States,
942   1994-1996. Fam Plann Perspect. 2001;33(4):161–5.

943   84.   Gondo F, da Silva MG, Polettini J, Tristao A da R, Peracoli JC, Witkin SS, et al. Vaginal
944   flora alterations and clinical symptoms in low-risk pregnant women. Gynecol Obstet
945   Invest. 2011;71(3):158–62.

946   85.   Tierney L, Rossini A., Li N, Sevcikova H. Simple Network of Workstations. 2016.

947   86.   Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: a
948   software environment for integrated models of biomolecular interaction networks.
949   Genome Res. 2003 Nov;13(11):2498–504.

950   87.   Nayfach S, Pollard KS. Toward Accurate and Quantitative Comparative Metagenomics.
951   Cell. 2016 Aug 25;166(5):1103–16.

952   88.   Bernstein MN, Doan A, Dewey CN. MetaSRA: normalized sample-specific metadata for
953   the Sequence Read Archive. bioRxiv. 2016 Nov 30;090506.

954   89.   Bhadra A, Rao A, Baladandayuthapani V. Inferring network structure in non-normal
955   and mixed discrete-continuous genomic data. ArXiv160400376 Stat [Internet]. 2016
956   Apr 1 [cited 2017 Sep 29]; Available from: http://arxiv.org/abs/1604.00376

Supporting information

959   **S1 File. Unfiltered, non-normalized, non-dichotomized microbiome counts and survey**

960   **responses (upon request)**

961   **S2 File. Survey questions and abbreviations (upon reques)**

962   **S3 File. Filtered, discretized microbiome counts and survey responses for all 60 variables**

963   **used in BN construction and analysis. (upon request)**

964   **S4 File.** *A priori* **classifications, abbreviations, and BN analysis results (degree, closeness**

965   **centrality and betweenness centrality) for all 60 variables used in BN construction and**

966   **analysis. (upon request)**

967   **S5 File. NMDS-ANOSIM testing results. (upon request)**

968   **S6 File. NMDS plots of the six variables found to significantly influence the vaginal**

969   **microbiome using the ANOSIM test. (upon request)**

970   **S7 Figure. 3D ordination plot based on PERMANOVA results.**

971   **S8 File. Figures describing the distribution of the abundances of taxa used in BN analysis.**

972   **Table 1. PERMANOVA permutation test results after stepwise model selection using the**

973   **AIC. PERMANOVA was used to assess the significance of associations between metadata**

974   **and the vaginal microbiome composition. P-values are based on 1000 permutations.**

Variable	Degrees of freedom	p-values
Nugent score	2	0.001
Previous pregnancy	1	0.001
Vaginal pH	1	0.001
Participant age	2	0.012
Use of menstrual pads during menstruation	1	0.023
Staining of underwear in previous 60 days	1	0.027
Use of male condoms as current birth control method	1	0.066