A note on measuring natural selection on principal component scores

Veronica K. Chong[1], Hannah F. Fung[1], John R. Stinchcombe[1,2]

1. Department of Ecology and Evolutionary Biology, University of Toronto, Toronto, ON, Canada
2. Koffler Scientific Reserve, University of Toronto, Toronto, ON, Canada

Author for correspondence: John Stinchcombe <john.stinchcombe@utoronto.ca>

Abstract

Measuring natural selection through the use of multiple regression has transformed our understanding of selection, although the methods used remain sensitive to the effects of multicollinearity due to highly correlated traits. While measuring selection on principal component scores is an apparent solution to this challenge, this approach has been heavily criticized due to difficulties in interpretation and relating PC axes back to the original traits. We describe and illustrate how to transform selection gradients for PC scores back into selection gradients for the original traits, addressing issues of multicollinearity and biological interpretation. We demonstrate this approach with empirical data and examples from the literature, highlighting how selection estimates for PC scores can be interpreted while reducing the consequences of multicollinearity.

"Unfortunately, the biological meaning of principal components or discriminant functions is not
always obvious."—Endler (1986, p 192)

"… it is difficult to translate from selection on principal components to selection on the original
traits."—Brodie et al. (1995)

Understanding how natural selection acts on correlated traits remains a fundamental challenge in
evolutionary biology (Lande and Arnold 1983). Most ecologically important traits that influence an
organism's fitness are correlated with other features of their morphology, life history, and behavior. The
canonical approach to estimating selection on correlated traits is selection gradient analysis (Lande and
Arnold 1983). By regressing estimates of relative fitness on multiple traits, the direct effects of individual
traits can be distinguished from the effects of selection on correlated traits. Widespread application of the
Lande-Arnold framework has revolutionized our understanding of natural selection, and has given
empiricists a straightforward tool for evaluating the fitness consequences of phenotypic variation.

One challenge to the Lande-Arnold approach, however, has remained especially stubborn:
regressing fitness estimates on correlated traits can become problematic when traits exhibit
multicollinearity (Mitchell-Olds and Shaw 1987). If the correlation between traits is high enough, it
becomes impossible to reliably estimate their separate contributions to relative fitness. Multicollinearity is
frequently diagnosed with a combination of variance inflation factors (VIF) or condition indices produced
by most regression packages, with a frequent rule of thumb that a VIF greater than 10 indicates serious
multicollinearity (Belsley et al. 1980; Neter 1989; see also O'Brien 2007). Typical recommendations for
multicollinearity include larger sample sizes, dropping or summing variables (Lande and Arnold 1983;
Mitchell-Olds and Shaw 1987), principal component analysis (Lande and Arnold 1983; see below), or
alternative approaches like partial least squares. While these methods have valuable statistical
properties, their widespread adoption has been limited. In many instances estimating selection on highly
correlated traits simultaneously is the object of study, and thus traits cannot be dropped. For partial least
squares, interpretability of the coefficients or the factors identified can remain problematic, as does their
lack of correspondence to selection gradients that can be used to predict evolutionary responses.

The merits of measuring selection on principal component scores, in particular, has been debated
since the advent of selection gradient analysis. Lande and Arnold (1983) themselves measured selection
on PC scores when analyzing selection on the Bumpus (1899) house sparrows dataset. Their use of
principal components was for data reduction when analyzing 9 traits with 49 female birds. They noted that
while directional selection was detected on some of the original traits, they failed to detect directional
selection on PC1 scores. One of the major advantages of selection gradient analysis—measuring and
comparing the magnitude of direct selection on correlated traits—can be lost, as one obtains an estimate
of selection on a composite trait that may not be the same in sign or statistical significance as the original
traits. PC scores are constructed based on their patterns of variation and covariation, regardless of their
importance for fitness, and can be hard to interpret biologically, especially when they encompass many

traits (see Mitchell-Olds and Shaw 1987 for an extended discussion). For these interpretive reasons,

selection estimates on PC scores have been criticized (e.g., Endler 1986; Mitchell-Olds and Shaw 1987;

Conner 2007; Hunt et al. 2007).

In this Comment and Opinion, we present and illustrate a simple approach that circumvents the

interpretation problem of measuring selection on principal components. By projecting selection estimates

for PCs back into the original trait space, their interpretability is preserved. We illustrate this with an

empirical example in the mouse ear cress, *Arabidopsis thaliana,* as well as examples from the literature

where previous authors have presented selection estimates for PC scores.

Projecting selection gradients for PCs back into trait space

For most incarnations, traits will have separate units, and we assume that PCA has been

performed on the correlation rather than the covariance matrix. A vector of selection gradients on PC

scores (i.e., regression coefficients of relative fitness on PC scores) can be projected into the original trait

space by:

$$\boldsymbol{\beta} = \mathbf{E}\,\mathbf{A} \tag{1}$$

Where $\boldsymbol{\beta}$ is a vector of reconstituted selection gradients, $\mathbf{E}$ is a matrix with eigenvectors from the original

PCA as columns, and $\mathbf{A}$ is the vector of regression coefficients obtained from regressing relative fitness

on PC scores (Jolliffe 2002, pg 169). The coefficients in $\mathbf{A}$ describe the relationships between relative

fitness and PC scores, while the elements of $\mathbf{E}$ describe the relationships between the PC scores and the

original traits; combining them yields the relationship between traits and relative fitness. While an

analogous equation for non-linear selection gradients would clearly be useful, we are unaware of any

similar expression in the statistical literature.

Using all the eigenvectors of a PCA for $\mathbf{E}$ and regression with all PC scores as traits for $\mathbf{A}$ with (1)

returns ordinary least squares regression coefficients (Jolliffe 2002; Wilks 2005), and does not eliminate

multicollinearity. Using a subset of PCs can reduce multicollinearity, and estimate selection on the original

traits. To estimate uncertainty in the elements of $\boldsymbol{\beta}$, we estimate their variance using equation 6 of Lafi

and Kaneene (1992), and from that standard errors. If one wishes to perform hypothesis testing on these

reconstituted selection gradients, parameter estimates and standard errors could be used to estimate t-

statistics and P-values. Prior to illustrating the use of (1) with our own data and literature examples, we

note two important items that we return to below. First, using (1) necessarily involves a decision about

how many PC axes to include, with attendant costs and benefits. Second, the elements of $\boldsymbol{\beta}$ estimated by

(1) represent selection within the sub-space represented by the columns of $\mathbf{E}$—e.g., if the columns of $\mathbf{E}$

represent 70% of the phenotypic variation, the elements of **β** represent selection on the traits in the sub-

space describing 70% of the phenotypic variance.

MATERIALS AND METHODS

Size and phenology in *Arabidopsis thaliana*

The mouse ear cress, *Arabidopsis thaliana* (Brassicaceaae) is a selfing annual rosette plant. Our

goal was to test the hypothesis that natural selection favoring early flowering, which is commonly

observed in experiments even with low mortality (Austen et al. 2017), could be explained as selection on

correlated traits, such as flowering duration and branch number. We reasoned that early flowering plants

could potentially have longer flowering durations, and more opportunity to produce branches which could

themselves support flowers and fruit. Stock (2015) crossed two parental lines collected from a single

population in Doylestown, PA, (USA) that had alternative alleles at the flowering time genes *FRIGIDA*

(FRI) and *FLOWERING LOCUS C (FLC;* Caicedo et al. 2004; Shindo et al. 2005) to produce an F1. She

allowed this F1 to self-fertilize to produce F2 progeny, which have all allelic combinations at these two

genes. Stock (2015) scored flowering time and allowed ~500 F2 individuals to self-fertilize, producing F3

seed that we used for our experiment. We used five replicates from 50 F3 lines chosen to evenly span the

F2 flowering time distribution.

We synchronized germination by stratifying seeds in 0.15 mg / 100 mL agar solution for six days

at 4°C (Stock et al. 2015). We planted each seed in a standard conetainer with saturated Sunshine Mix

#1 soil (Sun Gro Horticulture, Agawam, Massachusetts, USA). We arranged plants into conetainer racks

in a randomized, blocked design, where the two shelves of a growth chamber represented experimental

blocks, with plants of each line evenly divided between shelves. We maintained a light-temperature

regime of 16 hour days (22°C) followed by nights of 20° C (Suter and Widmer 2013; Stock et al. 2015).

We bottom-watered every two days by placing conetainers in trays of water for approximately three hours.

After three months, we gradually reduced watering to simulate a terminal end of season drought, and all

plants senesced after four months.

We scored bolting date, first and last flowering date throughout the experiment; at harvest, we

measured branch number, rosette diameter, and counted the number of rosette leaves. We collected and

counted fruits as a proxy for fitness (Stinchcombe et al. 2004). We measured flowering time as the

number of days between planting and the first flowering date, and flowering duration as the number of

days between first and last flowering dates. Analyses were performed using R v3.1.2 (R Foundation

2014) and SAS v9.4 (Cary, NC, USA).

Trait correlations and selection

We calculated line means prior to analysis of correlations between traits and selection (Rausher

1992). We then used Pearson correlation coefficients as estimates of the line mean correlations among

the traits. We first estimated selection differentials using simple univariate regressions regressing relative

fruit number on individual traits, using line means and with traits standardized to a mean of zero and

variance of one. We then estimated selection gradients for all traits, by regressing relative fruit number on

them in a multiple regression. Strong multicollinearity between flowering time and duration lead to high

variance inflation factors. To circumvent this, we used equation (1) above with four PCs, which explained

99% of the variation among line means.

Literature examples

We searched for examples measuring selection on PCs to illustrate potential use of equation (1). Our

goal was to find illustrative examples rather than a comprehensive meta-analysis. We describe new

findings obtained through the application of (1).

RESULTS

***Arabidopsis* experiment**

The traits showed a range of correlations, ranging from rosette diameter and flower duration which were

uncorrelated (r_g = 0.06, P = 0.647) to flowering time and duration, which were highly correlated (r_g = -0.95,

P< 0.0001; Figure 1). Flowering time showed strong correlations with branch number (r_g = -0.74,

P<0.001) and rosette leaf number (r_g =0.70, P <0.0001); flowering duration had essentially the opposite

correlation pattern (r_g = 0.72 and r_g = -0.73, respectively, for branch number and rosette leaf number;

Table 1). PCA showed that flowering time and flowering duration always had opposite signs in their

loadings on PC1-PC4; rosette diameter, branch number, and rosette leaf number loaded heavily on PC2,

PC3, and PC4.

Selection on flowering time and flowering duration

Univariate estimates of selection suggest that early flowering, long flowering durations, more

branches, and fewer rosette leaves are favored (Table 2a, Figure 2a-d). Interpreting these selection

differentials is complicated by the strong correlations among traits: positive selection for branch number,

for example, could be due to indirect selection for early flowering time (branch number is negatively

correlated) or selection for longer flowering durations (branch number is positively correlated). Traditional

multiple regression, while presented, is difficult to interpret: the high variance inflation factors for flowering

time (12.54) and flowering duration (11.32) suggesting caution in interpreting these gradients. While both

flowering time and flowering duration have large and significant selection differentials, neither are

significant in the multiple regression, and both have standard errors approximately 3-4 fold larger than the

standard error for the differentials, again because of multicollinearity.

Estimating selection gradients via equation 1 using four PCs showed strong directional selection for
earlier flowering ($\beta \pm$ s.e. = -0.17 ± 0.05), and for extended flowering durations ($\beta = 0.17 \pm 0.06$). The sign
of these selection gradients makes sense relative to their selection differentials and the sign and
magnitude of the trait correlation; standard errors are approximately 1/3 to 1/4 of those observed in the
multiple regression. These data suggest that selection is acting in approximately equal strength on
flowering time and flowering duration, in opposite directions. The quantitative similarity of these selection
gradients is likely due to the strong correlation between them, and reflects the two traits having
approximately equal and opposite loadings on PC1-PC4 (Table 1). These data suggest that accounting
for selection on correlated traits such as duration, branch number, size, and leaf number did not eliminate
the observed selection favoring early flowering time, even if longer flowering durations are favored.

The estimated selection gradients for branch number, rosette diameter, and rosette leaf number are
all approximately equal to their standard errors, suggesting that within the space of PC1-4, direct
selection on these traits was weak. In addition, the reconstituted selection gradients for branch number,
rosette diameter, and rosette leaf number (and their standard errors) are of roughly the same magnitude
as in the original multiple regression, suggesting that calculations from (1) accurately characterized
selection on them.

Literature examples

Size and emergence in damselflies: Anholt (1991) measured selection on body size components in the
damselfly *Enallagma boreale*. Several features of this study stand out. First, it was repeated across two
years, and second, it included selection separately for males and females. Third, sample sizes were
already large, and Anholt reported that he used PCs because of multicollinearity, as diagnosed by
variance inflation factors. Anholt (1991) summarized the date of emergence, wing length, abdomen
length, and mass at emergence with PCA, performing four separate PCAs, corresponding to each year-
sex combination. He estimated selection differentials for PC1 and PC2 (which accounted for ~85% of the
variance, depending on sex-year combination), using both relative survival and relative mate acquisition
as fitness estimates. We focus here on the survival data, as sample sizes were much larger (565 to 993 –
vs- 14 to 40). Because PC1 and PC2 are by definition uncorrelated, separate regressions of fitness on
each PC to estimate selection differentials are equivalent to multiple regression of fitness on both PCs
simultaneously. Consequently, we used the differentials for PC scores to calculate selection gradients.

Anholt interpreted PC1 as a size axis (due to the positive loadings of the size and mass
characters), with the negative loadings of date of emergence on PC1 as reflecting the commonly
observed trade-off between size and emergence date. In contrast, PC2 was interpreted as a date of
emergence axis because date loaded most heavily, although wing length and abdomen showed
moderate loadings in the same direction. As a consequence, PC1 represents a trade-off between date
and size, while PC2 represents moderate positive associations between date and size. Because date of
emergence figures in two PC axes in opposite directions vis a vis three other traits, and because the two

PC axes were under selection in opposite directions in three of four cohorts, calculating selection

gradients allows a straightforward assessment of the net pattern of selection on these traits without

keeping track of the relative sign of multiple eigenvector elements and selection coefficients for PC scores

across years and sexes. Our recalculations for the 1985 cohort suggest that selection in males favors

early emergence dates (β = -0.35 ± 0.15) and large mass (β = 0.3321 ± 0.14), while in females the pattern

is more variable on emergence date (β = -0.27 ± 0.19) and almost absent on mass (β = -0.065 ± 0.17).

For the 1986 cohort, our calculations suggest that the dominant pattern in both males and females was

for later emergence dates (β_{males} = 0.472 ± 0.18; $\beta_{females}$ = 0.533 ± 0.26).

Size traits in turtles: Janzen (1993) studied selection on body size in hatchling snapping turtles (*Chelydra*

serpentine). After releasing 112 hatchlings, he recaptured 66 individuals and estimated selection. Janzen

summarized five size-related traits with PCA (initial egg mass, hatchling mass, carapace length, carapace

width, and plastron length), using all of them as predictors of relative survival in a regression with other

traits including locomotor performance, the size of the clutch that eggs came from, and water potential of

the substrate eggs were incubated on (which affects size and hatching times). We used the first four PCs,

which explained 99.1% of the phenotypic variance, and confined our attention to size traits used to

construct the PCs. We note that subsequent work by Janzen (1998) suggests using logistic regression

these cases, but we calculated selection gradients from multiple regression on PCs to illustrate the

differences in interpretation.

When we projected selection estimates for the four PCs back into the original trait space, we

found directional selection against hatchling mass (β = -0.360 ± 0.17), and in favor of greater carapace

width (β = 0.370 ± 0.18) and plastron length (β = 0.309 ± 0.15). Two features of our reanalysis differ from

the original results. First, Janzen reported positive directional selection on PC1, which he logically

interpreted as an axis of size (because all elements of PC1 were positive for size-related traits). Our

findings illustrate selection on the component traits of PC1 is not always in the same direction, even when

all traits load on PC1 with similar sign and PC1 is under selection. The largest gradient we calculated was

for decreased mass. The overall patterns of selection against mass, and in favor of carapace width and

plastron length, come from selection on PC3 and PC4. PC3, which was under positive selection, had

strong loadings of plastron length. PC4 had strong and nearly opposite loadings of hatchling mass and

carapace width, which showed selection gradients of opposite sign when recalculated. While the overall

pattern of selection on the individual traits make sense in light of selection on the PCs and the loadings of

the overall traits, the converse is not true: without calculation, it is difficult if not impossible to infer net

patterns of selection on individual traits from regression coefficients for PCs.

Tadpole morphology: Kraft et al. (2006) raised tadpoles of the striped marsh frog (*Limnodynastes*

peroneii) in the presence and absence of dragonfly predators (*Anax* sp.), and measured the plastic

response of morphological traits and their subsequent effects on survival. They measured nine

morphological traits (total length, tail height, tail muscle height, body length, body height, tail area, tail
muscle area, body width, and tail muscle width), and performed PCA on log transformed data. They found
that PCs 4 and 5 (explaining 1.3% and 0.9% of the phenotypic variance, respectively) differed
significantly between predator-induced and control tadpoles. They measured selection on all nine PCs in
a multiple regression, finding significant negative selection on PC2, and positive selection on PC4 and
PC5 for both the control and predator-induced individuals. Because sign is arbitrary within a PC axis,
negative selection on PC2 is difficult to interpret: PC2 in their study had negative loadings of tail muscle
width and positive loadings of body height. Individuals to the extreme of PC2 thus had either narrow tail
muscles and large body heights, *or, equivalently*, wide tail muscles and small body heights. Which of
these character combinations had lower fitness would make dramatic differences in the interpretation of
selection.

When we projected selection coefficients for PC1-5 (representing 98.4% of the phenotypic
variance), we found that selection was acting most strongly on body width in the control individuals (β = -
0.0395), while most strongly on tail muscle height in the induced individuals (β = 0.0426). (Standard
errors for the regression coefficients for PCs were not presented, precluding estimation of standard errors
of the reconstituted selection gradients). The case of tail muscle width and body height is also clarified: in
both treatments, selection favors wider tail muscles and short body heights.

Flower volatiles and size: Schiestl et al. (2011) measured selection on 11 floral volatiles and inflorescence
size in the orchid *Gymnadenia odoratissima*. They performed several steps to facilitate selection analysis:
first they reduced the number of floral volatiles analyzed from >40 to the 11 most abundant compounds in
addition to inflorescence size. They summarized these variables with four principal components which
were subjected to a varimax rotation, explaining 84% of the variation. The varimax rotation is meant to
facilitate interpretation by creating a set of factors where each factor has predominant loadings from a
reduced set of the original traits.

Projecting their selection coefficients for the factors back into the original trait space revealed
appreciable selection on five measured traits: inflorescence size (β = 0.3208 ± 0.065),
phenylacetaldehyde (β = 0.1580 ± 0.066), eugenol (β = 0.1290 ± 0.061), (z)-isoeugenol (β = -0.1733 ±
0.066), and phenylethyl (β = 0.1669 ± 0.061). Natural selection within the sub-space described by the first
four PCs was approximately twice as strong on inflorescence size as on the volatile traits. Shiestl et al.
(2011) detected negative selection on the third factor (most associated with α-pinene and (z)-isoeugenol
after varimax rotation), yet α-pinene does not appear to be under selection when the coefficients are
projected back into the original trait space.

Flower volatiles, display size, and phenology: Parachnowitsch et al. (2012) also measured selection on
floral volatiles, in their case in the beardtongue *Penstemon digitalis*. They measured 30 traits, and faced

power challenges in estimating selection gradients for volatiles and other traits. Their solution was to only
include traits in a multiple regression if they showed significant or marginally significant selection
differentials. They noted several inherent challenges in measuring selection on floral volatiles: there are
many compounds within a floral bouquet; simply measuring selection differentials cannot distinguish
direct and indirect selection; if more than one trait loads on a PC, it is difficult to determine which trait is
under selection if that factor is significant; and measuring selection on principal components or varimax-
rotated factors inhibits comparisons to previous reports of selection on other floral traits.

We used the eight PCs presented by Parachnowitsch et al. (2012) which explained 72% of the
variance. In addition to volatiles, they included morphological and life history traits, including flower size,
corolla pigment, date of first flower, daily floral display size, number of days of flowering, and flower
number. They observed very strong selection on PC3, which was associated with cis-6-nonenal, linalool,
unknown-1, and daily display. By projecting selection estimates for eight PCs into the original trait space,
we see that while selection was observed for all traits ($\beta = 0.0569 \pm 0.0167$; $\beta = 0.0468 \pm 0.01286$; $\beta =$
0.114 ± 0.0161; $\beta = 0.1708 \pm 0.01918$, respectively), it was by far the strongest on daily display size.
Interestingly, the one compound that was significant in the multiple regression performed by the authors,
linalool, has an intermediate magnitude selection gradient when estimated in this manner.

DISCUSSION

Our use of principal components regression combined with simple algebra offers the prospect of
addressing the challenge of multicollinearity while solving the problems of interpreting selection on PC
axes. Below we evaluate general trends that come from our reanalyses, compare the approach to related
approaches in selection analysis, and close with a discussion of the limitations and judgements inherent
to the approach we advocate.

General lessons

We see three general lessons that have emerged from our approach. First, in numerous
examples, it is much simpler to understand the reconstituted selection gradients on the original traits than
on the PC axes, solving the interpretational challenges raised by Endler (1986), Mitchell-Olds and Shaw
(1987), Brodie et al. (1995), Conner (2007), and Hunt et al. (2007). For example, for the *Arabidopsis*
experiment, the net outcome of selection on flowering time and flowering duration being of approximately
equal magnitude and opposite signs was difficult to see from the PC scores, and completely masked by
the multiple regression. Likewise, in interpreting the Anholt and Kraft et al. studies, the original traits are
straightforward to understand biologically, while selection on the PC axes can only be understood by
considering multiple PC axes that have selection estimates of opposite signs, and are themselves made
up of traits loading on those axes with opposite signs. While in some cases PCs have clear biological
interpretations (e.g., the early flowering / small –vs- late flowering / large axis studied by Colautti et al.

(2010] and Colautti and Barrett (2010]), for more complicated cases like the ones analyzed here
projecting selection estimates on PCs back into the original trait space may be a valuable approach. It is
important to recall, however, that these reconstituted selection gradients estimate how selection is acting
on the portion of multivariate trait space spanned by the PCs that were originally included. Including more
PCs captures more of the variation in the original traits measured, and is closer to the typical goal of
selection gradient analysis.

Second, a natural interpretation of selection on PC axes is that they represent selection on the
trait which loads most heavily on that axis. Several of our examples challenge this interpretation: for
example, in the Schiestl et al. study, α-pinene loaded most heavily on an axis under selection, yet it did
not appear to be under much selection when selection on PCs was projected back into the original trait
space. Lande and Arnold (1983) noted that selection on PC scores can be diluted if traits under selection
show strong phenotypic covariances with those unimportant for fitness. Our literature review revealed the
converse problem: several cases in which seemingly straightforward selection on the PC axes was in fact
a complex pattern of selection on the underlying traits. The reason for this is that traits load on all PC
axes, rather than just one, and associations between these other PC axes and fitness contribute to the
net pattern of selection on the traits.

Third, and perhaps most important, multicollinearity or high-dimensional data (e.g., floral volatiles,
gene expression) do not need to be a stopping point for selection gradient analysis. For these types of
studies, multicollinearity or high dimensionality will force investigators into doing *something*: either
forgoing analysis, changing biological hypotheses, dropping traits, modifying their approach to selection
gradient analysis, or facing complex interpretation challenges. The studies we reviewed were explicit
about their approaches to these challenges. However, it is unclear how many studies in the literature
encountered multicollinearity and dropped traits or changed their analysis approach, especially since the
challenges and criticisms associated with measuring selection on PC axes may have convinced many
authors to forgo PC regression. Measuring selection on PC axes and then projecting into the original
phenotypic space, while not perfect, offers a route forward.

Similarity to other approaches

Our approach is similar to other variants of selection gradient analysis, and it is worth considering
the similarities and differences. At first glance, it appears similar to canonical rotation analysis (Simms
1990; Phillips and Arnold 1989; Blows 2007) because both involve PCA, although there are key
differences. In that approach, the γ matrix of stabilizing, disruptive, and correlational selection gradients is
subject to eigenanalysis, to identify the axes of trait variation under the strongest non-linear selection. In
contrast, our approach is to apply PCA to the raw data, estimate linear selection on the synthetic axes,
and then project back into the original trait space. Janzen and Stern (1998) presented a similar approach
to ours, showing how to convert logistic regression coefficients into selection gradients that can be used
to predict evolutionary change. Finally, our approach is somewhat similar to projection pursuit regression

(Schluter and Nychka 1994). Projection pursuit regression can be applied to identify a single axis (a linear

combination of traits), or multiple axes (multiple or combinations of traits) that explain the most variation in

fitness (see also Morrissey and Sakrejda (2013) and Morrissey (2014)). Morrissey (2014) considered

numerous types of regression methods for selection gradient estimation using both empirical data and

simulations; he strongly supported projection pursuit regression. We see our approach as

complementary, with one benefit being that it involves the typical machinery of least-squares regression

and PCA familiar to many biologists, and can be applied retrospectively to summary data reported in

papers (as we have here).

Prospects and caveats

Our use of PCA in combination with selection gradient analysis for our own data and examples

from the literature entailed costs that are worth considering before wider application. First, the

reconstituted regression coefficients are no longer unbiased estimators like least-squares estimates

(Wilks 2005; Jolliffe 2002). By omitting PCs that explain relatively little variance, one reduces the variance

in the estimated regression coefficients caused by multicollinearity (and their associated variance inflation

factors), but at the expense of statistical bias (Jolliffe 2002). Jolliffe (2002, pages 170-171) shows how

multicollinearity is driven by PCs with small eigenvalues, and accordingly, that regression coefficients for

the predictor traits that load heavily on those axes will be estimated imprecisely in the original regression

(see also Wilks 2005, page 505). Eliminating multicollinearity requires dropping a PC axis; it is important

to note that information about all of the original traits still remains in the model, through the retained PC

axes. In contrast, a trait omitted from a traditional regression could still have important effects on fitness,

and be correlated with included predictors, but it is impossible to evaluate the effects of traits not included

or reported in a selection analysis.

While omitting trailing PCs eliminates multicollinearity, Mitchell-Olds and Shaw (1987) criticized

the assumption that trailing PCs (trait combinations with less variance) were not predictive of relative

fitness and could be safely omitted. Likewise, Joliffe (1982) described several cases outside of

evolutionary biology where PCs explaining relatively little variance were still predictive of the response

variable. In the event that these PCs are associated with relative fitness, omitting them is not an advisable

solution. In contrast, Blows and McGuigan (2015) and Sztepanacz and Blows (2017) show how the

distribution of even *leading* eigenvalues from phenotypic and genetic data can be strikingly difficult to

distinguish from sampling variation, suggesting that trailing principal components might primarily

represent sampling variation. In the statistical literature, numerous approaches have been developed for

determining how many PCs to use in principal component regression; Jolliffe (2002, pages 173-177) and

Hill et al. (1977) provide overviews. One practical way forward would be to evaluate whether trailing PCs,

which are responsible for multicollinearity, are predictive of relative fitness. If they are not (as in the

Arabidopsis example), omitting these PCs and then projecting selection estimates back into the original

trait space seems to be a viable option. Fundamentally, some judgement of the investigator will be

required to decide how many PCs to include without reintroducing multicollinearity while still capturing a

sufficient portion of phenotypic variance to evaluate biological hypotheses about selection.

Acknowledgements. Our work is supported by NSERC Canada (JRS, VKC) and the Department of

Ecology and Evolutionary Biology. We thank Ben Gilbert, Don Jackson, Dave Punzalan, Jessica Forrest,

and Corlett Wood for discussion; comments by Anne Charmantier and anonymous reviewers improved

the manuscript.

References

Austen, E. J., L. Rowe, J. R. Stinchcombe, and J. R. K. Forrest. Explaining the apparent paradox of persistent selection for early flowering. New Phytologist 215: 929-934.

Anholt, B. R. 1991. Measuring Selection on a Population of Damselflies with a Manipulated Phenotype. Evolution 45:1091-1106.

Belsley, D.A., E. Kuh, and R.E. Welsch. 1980. *Regression Diagnostics: Identifying Influential Data and Sources of Collinearity",* John Wiley and Sons, Hoboken, NJ, USA.

Blows, M.W. 2007. A tale of two matrices: multivariate approaches in evolutionary biology. Journal of Evolutionary Biology 20: 1-8.

Blows, M. W., and K. McGuigan. 2015. The distribution of genetic variance across phenotypic space and the response to selection. Molecular Ecology 24:2056-2072.

Brodie, E. D., A. J. Moore, and F. J. Janzen. 1995. Visualizing and quantifying natural selection. Trends in Ecology & Evolution 10:313-318.

Bumpus, H. 1899. The elimination of the unfit as illustrated by the introduced sparrow, *Passer domesticus*. Biological lectures from the Marine Biological Laboratory of Woods' Hole, Mass. Ginn, Boston.

Caicedo, A. L., J. R. Stinchcombe, K. M. Olsen, J. Schmitt, and M. D. Purugganan. 2004. Epistatic interaction between *Arabidopsis FRI* and *FLC* flowering time genes generates a latitudinal cline in a life history trait. PNAS 101:15670-15675.

Campbell, D. R. 1989. Measurements of selection in a hermaphroditic plant: Variation in male and female pollination success. Evolution 43:318-334.

Colautti, R. I., C. G. Eckert, and S. C. H. Barrett. 2010. Evolutionary constraints on adaptive evolution during range expansion in an invasive plant. Proceedings of the Royal Society B: Biological Sciences 277:1799-1806.

Colautti, R. I., and S. C. H. Barrett. 2010. Natural Selection and Genetic Constraints on Flowering Phenology in an Invasive Plant. International Journal of Plant Sciences 171:960-971.

Conner, J. K. 2007. A tale of two methods: putting biology before statistics in the study of phenotypic evolution. Journal of Evolutionary Biology 20:17-19.

Endler, J. A. 1986. Natural selection in the wild. Princeton University Press, Princeton, NJ, USA.

Hill, R. C., T. B. Fomby, and S. R. Johnson. 1977. Component selection norms for principal components regression. Communications in Statistics - Theory and Methods 6:309-334.

Hunt, J., J. B. Wolf, and A. J. Moore. 2007. The biology of multivariate evolution. Journal of Evolutionary Biology 20:24-27.

Janzen, F. J. 1993. An experimental analysis of natural selection on body size of hatchling turtles. Ecology 74:332-341.

Janzen, F. J., and H. S. Stern. 1998. Logistic regression for empirical studies of multivariate selection. Evolution 52:1564-1571.

Jolliffe, I.T. (1982). A note on the use of principal components in regression. Appl. Statist., 31, 300–303.

Jolliffe, I. T. 2002. Principal Component Analysis. Springer, NY, NY, USA.

Kraft, P. G., C. E. Franklin, and M. W. Blows. 2006. Predator-induced phenotypic plasticity in tadpoles: extension or innovation? Journal of Evolutionary Biology 19:450-458.

Lande, R., and S. J. Arnold. 1983. The measurement of selection on correlated characters.
Evolution 37:1210-1226.
Lafi, S.Q. and J.B. Kaneene. 1992. An explanation of the use of principal components analysis to
detect and correct for multicollinearity. Preventive Veterinary Medicine 13: 261-275.
Mitchell-Olds, T., and R. G. Shaw. 1987. Regression analysis of natural selection: statistical
inference and biological interpretation. Evolution 41:1149-1161.
Morrissey, M.B. and K. Sakrejda. 2013. Unification of regression-based methods for the analysis
of natural selection. Evolution 67: 2094-2100.
Morrissey, M.B. 2014. In search of the best methods for multivariate selection analysis.
Methods in Ecology and Evolution 5: 1095-1109.
Neter, J., Wasserman, W. & Kutner, M. H. (1989). Applied Linear Regression Models.
Homewood, IL: Irwin
O'Brien, R.M. 2007. A caution regarding rules of thumb for variance inflation factors. Quality
and Quantity 41: 673-690.
Parachnowitsch, A. L., R. A. Raguso, and A. Kessler. 2012. Phenotypic selection to increase floral
scent emission, but not flower size or colour in bee-pollinated Penstemon digitalis. New
Phytologist 195:667-675.
Phillips PC, Arnold SJ. 1989. Visualizing multivariate selection. Evolution 43: 1209–1222.
R Core Team. 2014. R: a language and environment for statistical computing. Vienna: R
Foundation for Statistical Computing.
Rausher, M. D. 1992. The measurement of selection on quantitative traits: Biases due to
environmental covariances between traits and fitness. Evolution 46:616-626.
Schiestl, F. P., F. K. Huber, and J. M. Gomez. 2011. Phenotypic selection on floral scent: trade-off
between attraction and deterrence? Evolutionary Ecology 25:237-248.
Schluter, D. and D. Nychka. 1994. Exploring fitness surfaces. American Naturalist 143: 597-616.
Shindo, C., M. J. Aranzana, C. Lister, C. Baxter, C. Nicholls, M. Nordborg, and C. Dean. 2005. Role
of *FRIGIDA* and *FLOWERING LOCUS C* in determining variation in flowering time of
Arabidopsis. Plant Physiology 138:1163-1173.
Simms EL. 1990. Examining selection on the multivariate phenotype: plant resistance to
herbivores. Evolution 44: 1177–1188.
Stinchcombe, J.R., L.A. Dorn, and J. Schmitt. 2004. Flowering time plasticity in Arabidopsis
thaliana: a reanalysis of Westerman & Lawrence (1970). Journal of Evolutionary Biology
17:197-207.
Stock, A. J. 2015. Evolution of Arabidopsis thaliana flowering time in response to water
availability post-introduction. MSc Thesis. University of Toronto, Toronto, ON, Canada.
Stock, A. J., B. V. McGoey, and J. R. Stinchcombe. 2015. Water availability as an agent of
selection in introduced populations of Arabidopsis thaliana: impacts on flowering time
evolution. PeerJ 3:e898.
Sztepanacz, J. L., and M. W. Blows. 2017. Accounting for Sampling Error in Genetic Eigenvalues
Using Random Matrix Theory. Genetics 206: 1271-1284.
Suter, L., and A. Widmer. 2013. Environmental Heat and Salt Stress Induce Transgenerational
Phenotypic Changes in Arabidopsis thaliana. PLoS ONE 8:e60364.
Wilks, D.S. 2005. Statistical methods in the atmospheric sciences. Elsevier, NY, NY, USA.
468

TABLES

Table 1. Line mean correlations for the *Arabidopsis* study, with eigenvector loadings for PC1-4.

	Flowering Time	Flowering Duration	Branch Number	Rosette Diameter	PC1	PC2	PC3	PC4
Flowering Time	1				0.522	-0.215	0.154	-0.352
Flowering Duration	-0.95 P<0.0001	1			-0.523	0.157	-0.244	0.426
Branch Number	-0.74 P<0.0001	0.72 P<0.0001	1		-0.476	-0.018	0.856	-0.189
Rosette Diameter	-0.12 P=0.39	0.06 P = 0.67	-0.09 P= 0.54	1	0.045	0.913	-0.047	-0.397
Rosette Leaf Number	0.70 P<0.0001	-0.73 P<0.0001	-0.64 P<0.0001	0.33 P = 0.02	0.473	0.306	0.425	0.708
Percent Variance Explained					65.17%	23%	6.94%	3.5%

Table 2. Selection estimates for the *Arabidopsis* experiment. Univariate estimates of selection are from regressions of relative fruit production on each trait alone. Selection gradients are estimated from a multiple regression of relative fitness on all traits, although the estimates for flowering time and duration are uncertain because of multicollinearity (indicated by variance inflation factors (VIF)). Principal component regression estimates are regression coefficients for PC1-PC4 projected back into the original trait space.

	Univariate Regressions			Multiple Regression				Principal Component Regression
Trait	s ± se	t	P	β ± se	t	P	VIF	β ± se
Flowering time	**-0.37 ± 0.05**	**6.96**	**<0.0001**	-0.30 ± 0.19	1.57	0.12	12.54	-0.17 ± 0.05
Flowering Duration	**0.35 ± 0.06**	**6.29**	**<0.0001**	0.05 ± 0.18	0.27	0.78	11.32	0.17 ± 0.06
Branch Number	**0.295 ± 0.06**	**4.73**	**<0.0001**	0.07 ± 0.08	0.95	0.35	2.45	0.09 ± 0.08
Rosette Diameter	0.11 ± 0.07	1.52	0.136	0.059 ± 0.07	0.87	0.39	1.60	0.07 ± 0.07
Rosette Leaf number	**-0.218 ± 0.07**	**3.18**	**0.0026**	0.061 ± 0.09	0.63	0.53	3.21	-0.06 ± 0.10

FIGURES

Figure 1. Line mean correlation between flowering time and flowering duration in the *Arabidopsis* chamber experiment. The inbred line mean correlation is -0.95.

Figure 2. Selection differentials for: (A) flowering time, (B) flowering duration, (C) rosette leaf number, and (D) branch number in the *Arabidopsis* experiment. Differentials are portrayed in the original trait units.