| 1 | Don't throw out the sympatric species with the crater lake water: |
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| 2 | fine-scale investigation of introgression provides weak support for |
| 3 | functional role of secondary gene flow in one of the clearest |
| 4 | examples of sympatric speciation |
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23 Abstract

24 Genomic data has revealed complex histories of colonization and repeated gene flow previously 25 unrecognized in some of the most celebrated examples of sympatric speciation and radiation. 26 However, much of the evidence for secondary gene flow into these radiations comes from genome-wide tests, which tells us little about how gene flow potentially influenced sympatric 27 28 diversification. Here we investigated whole genomes of Barombi Mbo crater lake cichlids for 29 fine-scale patterns of introgression between species with neighboring riverine cichlid 30 populations. We did find evidence of secondary gene flow into the radiation scattered across < 31 0.24% of the genome; however, the functional and genetic diversity in these regions paint no 32 clear picture of how that variation could have contributed to the ecological and morphological 33 diversity found in the lake. Our results suggest that either variation in novel genetic pathways 34 introduced during secondary gene flow contributed to the radiation, or that secondary gene flow 35 was predominantly neutral with respect to the diversification processes. We also found evidence 36 for differential assortment of ancestral polymorphism found in riverine populations between 37 sympatric sister species, suggesting the presence of a hybrid swarm in the past. While the history 38 of gene flow and colonization appears to be more complicated than once thought, the lack of 39 compelling evidence for secondary gene flow influencing diversification suggests that we should 40 not yet rule out one of the most celebrated examples of sympatric speciation in nature.

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46 Introduction

Sympatric speciation, the extreme endpoint on the speciation-with-gene-flow continuum, is 47 48 traditionally defined as the evolution of reproductive isolation without the aid of geographic 49 barriers (Coyne and Orr 2004). Sympatric speciation has fascinated evolutionary biologists since Darwin for its illustration of the power of complex interactions between natural and sexual 50 51 selection to create new species. Despite intense searches, very few case studies have been able to 52 meet the rigorous criteria for demonstrating sympatric speciation in nature (Coyne and Orr 2004; 53 Bolnick and Fitzpatrick 2007). Even in some of the more convincing examples that do meet 54 these criteria, genomic data have revealed more complex evolutionary histories of multiple 55 colonizations and repeated gene flow than previously thought (Papadopulos et al. 2011; The 56 Heliconius Genome Consortium et al. 2012; Geiger et al. 2013; Alcaide et al. 2014; Igea et al. 57 2015; Malinsky et al. 2015; Martin et al. 2015a; Kautt et al. 2016). 58 However, much of the support for complicated histories involving repeated gene flow 59 events into radiations comes from genome-wide tests for gene flow (e.g. (Lamichhaney et al. 2015; Martin et al. 2015a; Meier et al. 2017)). One prediction of models of speciation with gene 60 61 flow is that divergence between incipient species should be heterogeneous across the genome 62 (Turner et al. 2005; Harr 2006; Feder et al. 2012; Nosil and Feder 2012a,b). Indeed, high 63 heterogeneity in genomic differentiation has been found across the genomes of many recent or 64 incipient sister species (e.g. Jones et al. 2012; Martin et al. 2013; Poelstra et al. 2014; Soria-65 Carrasco et al. 2014; Malinsky et al. 2015; McGirr and Martin 2016), although other processes 66 besides differential gene flow across the genome can produce similar heterogeneous patterns 67 (Noor and Bennett 2009; Nachman and Payseur 2012; Cutter and Payseur 2013; Cruickshank 68 and Hahn 2014; Guerrero and Hahn 2017; Ravinet et al. 2017). Only a handful of genes may

directly contribute to the speciation process whereas the rest of the genome is porous to gene flow while reproductive isolation is incomplete (Wu 2001; Wu and Ting 2004). Therefore, gene flow detected at the genome-wide level from populations outside the sympatric radiation does not by itself constitute evidence that secondary gene flow was involved in the divergence process among incipient species and shaped the radiation.

74 The Cameroon crater lake cichlid radiations are some of the most compelling cases for 75 sympatric speciation in the wild (Coyne and Orr 2004). The most speciose of these radiations is 76 found in the isolated 2.3 km-wide volcanic crater lake Barombi Mbo (Trewavas et al. 1972; 77 Schliewen et al. 1994; Schliewen and Klee 2004). Barombi Mbo hosts a radiation of 11 endemic cichlid species, many of which have clear morphological and ecological separation from other 78 79 sympatric species (Schliewen et al. 1994). Some endemics have evolved unique specializations, 80 such as the spongivore *Pungu maclareni* and deep-water hypoxia specialist *Konia dikume* 81 (Trewavas et al. 1972). Other endemics, such as *Stomatepia mariae* and *S. pindu*, appear to be 82 incipient or stalled species complexes with only slight morphological and ecological divergence 83 at the extremes of a unimodal distribution of phenotypes (Martin 2012). However, evidence of 84 differential introgression, weak support for Barombi Mbo monophyly, and differences in levels 85 of shared ancestry with outgroup riverine populations from genome-wide RAD-seq data suggest 86 additional secondary gene flow into the radiation after the initial colonization, casting doubt on 87 one of the best examples of sympatric speciation in the wild (Martin et al. 2015a).

88 Here we dissect those signals of repeated gene flow to investigate their role in the 89 radiation using whole-genome sequences. We performed exhaustive searches for all genetic 90 patterns consistent with secondary gene flow into the ancestral Barombi Mbo population or into 91 subclades after their initial divergence using machine learning to finely dissect phylogenetic

| 92 | signal across the genome and genomic scans to test for differential introgression. We find | | | | | |
|---|--|--|--|--|--|--|
| 93 | evidence of both shared introgression between sister species and across subclades in the radiation | | | | | |
| 94 | as well as differential introgression among sister species across small regions of the genome. | | | | | |
| 95 | However, functional and genetic diversity in these regions do not paint a clear picture of how | | | | | |
| 96 | introgressed variants may have contributed to speciation in these groups. Our results suggest that | | | | | |
| 97 | either 1) rare introgression of variants in novel genetic pathways contributed to the | | | | | |
| 98 | morphological and ecological diversity of the radiation (speciation with an allopatric phase), 2) | | | | | |
| 99 | secondary gene flow was predominantly or completely neutral and did not contribute to | | | | | |
| 100 | diversification in Barombi Mbo (sympatric speciation with gene flow), or 3) multiple | | | | | |
| 101 | colonizations of the lake before diversification brought in genetic variation that was then | | | | | |
| 102 | differentially sorted among incipient species (sympatric speciation from a hybrid swarm). | | | | | |
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| 103 104 | Methods | | | | | |
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115 Characterization of introgression patterns across the genome

116 First, we exhaustively searched the genomes for patterns of non-monophyletic Barombi Mbo 117 relationships using the machine learning program SAGUARO (Zamani et al. 2013) to identify 118 regions of the genome that contained relationships consistent with expectations from multiple 119 colorizations and secondary gene flow into the radiation (i.e. paraphyletic/polyphyletic Barombi 120 Mbo radiations). This method infers relationships among individuals in the form of genetic 121 distance matrices and assigns segments across the genomes to different topologies without a 122 priori hypotheses about these relationships. We partitioned the genome into a total of 75 unique 123 topologies (well past the inflection point at 30 topologies where the percent of genome explained 124 by each additional topology plateaus; Fig S1) to exhaustively search for relationships where 125 subclades or individual Barombi Mbo species were more closely related to riverine populations 126 than other species in the crater lake, suggesting sympatric speciation after a hybrid swarm (i.e. 127 differential sorting of ancestral polymorphism) or secondary gene flow into this subclade 128 (introgression). Details on the SAGUARO analysis and filtering strategies for calculating 129 proportions are provided in the supplementary methods.

130 We also looked for evidence of differential introgression within subclades of the radiation 131 on both a genome-wide and local level using f_4 statistics (Reich et al. 2009; Patterson et al. 2012; 132 Pickrell and Pritchard 2012). The f_4 statistic tests if branches on a four-taxon tree lack residual 133 genotypic covariance (as expected in the presence of incomplete lineage sorting and no 134 introgression) by comparing allele frequencies among the three possible unrooted trees. 135 We focused on tests of introgression with the two outgroup clades from our sample that came 136 from two main clusters: riverine populations of Sarotherodon galilaeus in the Mungo and Meme 137 rivers (MM) and riverine populations of S. galilaeus from the more distant Cross River (CR).

| 138 | Based on the tree ((P1, P2), (S. galilaeus MM, S. galilaeus CR)), f_4 statistics were calculated for |
|-----|--|
| 139 | combinations of species among a) Stomatepia, b) the Konia + Pungu subclade, and c) Myaka |
| 140 | myaka with S. linnelli as a representative of its sister Sarotherodon group. This subset of |
| 141 | groupings was chosen to make these analyses more tractable by focusing on species with unique |
| 142 | trophic ecologies within the radiation. Genome-wide f_4 statistics were calculated using the |
| 143 | fourpop function in Treemix (Pickrell and Pritchard 2012). Standard error was estimated by |
| 144 | jackknifing in windows of 1,000 adjacent SNPs to account for linkage disequilibrium. |
| 145 | We characterized heterogeneity in introgression across the genome among these same |
| 146 | combinations and investigated whether differential introgression contributed variation potentially |
| 147 | important in the divergence between species by calculating f_4 statistics in 10-kb sliding windows. |
| 148 | We did this with a modified version of the ABBABABA.py and genomics.py scripts that use |
| 149 | population allele frequencies of biallelic SNPs |
| 150 | (https://github.com/simonhmartin/genomics_general;(Martin et al. 2015b); our modified version |
| 151 | is provided in the supplementary materials). Significance of f_4 values in sliding windows across |
| 152 | the genome were evaluated using the 1% tails of a null distribution generated from permutations |
| 153 | of the f_4 test. For more details on the sliding window calculations of f_4 , see supplementary |
| 154 | methods. |
| 155 | For each of these regions, we looked for annotated genes using the well annotated NCBI |

156 *Oreochromis* Annotation Release 102 and searched their gene ontology in the phenotype

157 database 'Phenoscape' (Mabee et al. 2012; Midford et al. 2013; Manda et al. 2015; Edmunds et

al. 2016) and AmiGO2 (Balsa-Canto et al. 2016) for pertinent functions to the specializations

and observed morphological differences among species, such as skeletal system or pigmentation.

161 Directionality of introgression

162 The sign of f_4 does not indicate the directionality of introgression because of the lack of an explicit outgroup. For example, in the tree (P1,P2),(P3,P4)), a positive f_4 value indicates gene 163 164 flow either between P1 and P3 or P2 and P4. We narrowed down the directionality of 165 introgression detected in these regions using the f_d statistic, a modified version of the D-statistic 166 that looks at allele frequencies fitting two allelic patterns referred to as ABBA and BABA based 167 on the tree ((P1,P2),P3,O)), where O is an outgroup species in which no gene flow is thought to 168 occur with the other populations (Martin et al. 2015b). Using two individuals of Coptodon kottae 169 from another Cameroon crater lake as our distantly related outgroup population and the same 170 riverine and Barombi Mbo population combinations described above, f_d values were calculated in 171 10-kb windows across the genome using the same script and window settings as in the f_4 tests of 172 introgression. The variation in f_d values per region was higher than f_4 of similar window sizes, 173 perhaps due to the high variance between windows in number of sites that fit the desired ABBA/BABA patterns given our outgroup, so we used only those f_d outliers (the top 2%) that 174 175 overlapped with significant f_4 outliers as potentially introgressed regions from which we could 176 narrow down the two populations in which gene flow occurred. 177 We also visualized the directionality of genome-wide introgression detected with the f_4 178 statistics using Treemix (v 1.13) (Pickrell and Pritchard 2012). Treemix estimates a maximum 179 likelihood phylogeny of the focal populations and then fits a user-specified number of migration

edges to the tree by comparing genetic covariances between populations. We ran Treemix with *S*. *galilaeus* as root, and with 0 through 20 migration edges. To determine the most likely number

182 of migration events, we performed likelihood-ratio tests comparing each graph to that with one

fewer migration event, starting with 1 versus 0 events, and took as the most likely value the firstnon-significant comparison.

185

186 Comparison of patterns of introgression to patterns of genetic divergence and diversity

- 187 Reduced levels of genetic polymorphism in a population may indicate a strong selective sweep.
- 188 We can look at introgressed regions found in only a single Barombi Mbo species for evidence
- 189 that they have been adaptive, suggesting that secondary gene flow brought in variation
- 190 potentially important for speciation. To examine genetic diversity in candidate introgressed
- 191 regions, we calculated between-population nucleotide divergence (D_{xy}) and within-population
- 192 nucleotide diversity (π) for pairwise species comparisons among the Barombi Mbo focal species
- and the riverine outgroups. D_{xy} and π were calculated over the same 10-kb windows as the f_4 tests
- 194 using the python script popGenWindows.py
- 195 (<u>https://github.com/simonhmartin/genomics_general</u>; (Martin et al. 2015b); see supplementary
- 196 methods for more details on these calculations).

197

198 Results

Widespread polyphyletic relationships in Barombi Mbo are scattered across small regions of the
genome

201 After conservative filtering of segments to remove uninformative regions (see supplementary

202 methods and Table S1), the Barombi Mbo cichlid radiation was a monophyletic group across

203 53% of the genome and only 0.6% was assigned to topologies indicating a polyphyletic Barombi

- 204 Mbo. These polyphyletic relationships are consistent with many patterns, including secondary
- 205 gene flow, incomplete lineage sorting, divergent selection, and ancestral population structure.
- 206 The most prevalent topology spanned 38.2% of the genome and featured the expected species

207 phylogeny for this group, in which all Barombi Mbo individuals form a single clade with distant 208 relationships to outgroup riverine Sarotherodon populations in Cameroon (Fig. 1A). The second 209 most prevalent topology (spanning 11.8% of the genome) featured identical evolutionary 210 relationships, except for a much shorter branch leading to S. galilaeus Mungo and Meme River 211 populations (Fig. 1B). Branch lengths produced by SAGUARO have no direct interpretation as 212 an evolutionary distance (analogous to a neighbor-joining tree), but may be useful for 213 comparison to similar topologies with different branch lengths, e.g. regions with higher 214 divergence rates (Zamani et al. 2013).

215 In 0.6% of the genome indicating polyphyletic Barombi Mbo relationships, we found 216 evidence consistent with multiple colonizations of the lake. Since we were looking for patterns 217 consistent with secondary gene flow or a hybrid swarm for subclades of the radiation, we 218 focused on topologies where single species or entire subclades were more closely related to 219 outgroups than other Barombi Mbo species, which represented only 0.24% of the genome. Some 220 topologies featured an entire subclade (e.g. Stomatepia) as monophyletic, but more closely 221 related to the riverine populations than other Barombi Mbo species, consistent with a hybrid 222 swarm scenario before the diversification of the Stomatepia subclade. Other topologies featured 223 individual species more closely related to outgroup riverine populations than sister species, 224 consistent with secondary gene flow into that lineage after the initiation of divergence. For 225 example, in *Stomatepia* we found topologies that group multiple species with riverine 226 populations (Fig. 2A-B), but we also found topologies where individual *Stomatepia* species (S. 227 *mariae* and *S. pindu*; Fig. 2C-D) were more closely related to riverine outgroups than other 228 Stomatepia. In the Konia + Pungu subclade, we saw a similar pattern with topologies for the 229 hypoxia and sponge-eating specialists (K. dikume and P. maclareni, respectively; Fig. 3A-B) but

| 230 | also a topology where the entire subclade was sister to the riverine outgroup populations (Fig. |
|-----|---|
| 231 | 3C). In the zooplanktivore M . $myaka$, we found topologies in which M . $myaka$ was sister to the |
| 232 | riverine populations (Fig. 4A-B), but also topologies where <i>M. myaka</i> , along with all the |
| 233 | Barombi Mbo Sarotherodon species, were sister to the riverine outgroup populations (Fig. 4C- |
| 234 | D). |
| 235 | |
| 236 | Genome-wide evidence for differential introgression into the radiation |
| 237 | Consistent with evidence of differential introgression from RAD-seq data (Martin et al. 2015a), |
| 238 | genome-wide f_4 tests provided evidence of genome-wide differential gene flow between some |
| 239 | Barombi Mbo sister species and the outgroup riverine species (Table 1). There was significant |
| 240 | evidence of genome-wide introgression in tests involving both S. pindu in the Stomatepia species |
| 241 | complex and the hypoxia specialist K . dikume in the Konia + Pungu subclade. Some species pair |
| 242 | combinations within these subclades did not show evidence of differential gene flow, suggesting |
| 243 | that there may still be sympatric speciation occurring for some species, if not entire subclades. |
| 244 | For example, there was no significant secondary gene flow detected genome-wide in the tests |
| 245 | involving sister species S. mariae and S. mongo or M. myaka and S. linnelli (Table 1). |
| 246 | We also found evidence for widespread gene flow connecting populations across |
| 247 | Barombi Mbo and neighboring riverine populations in highly interconnected population graphs; |
| 248 | the likelihood of each graph did not plateau until reaching 10 admixture events (Fig S5). On the |
| 249 | Treemix population graph with 10 admixture events, gene flow from the Mungo/Meme River |
| 250 | populations of S. galilaeus occurred directly into individual species S. mongo and K. eisentrauti |
| 251 | rather than the ancestral node of their respective subclades (Fig S6). The proportion of admixture |
| 252 | inferred for these two events (0.1% into S. mongo and 0.4% into K. eisentrauti) was similar to |
| | |

| 253 | the small proportions of the genome assigned to topologies consistent with secondary gene flow |
|-----|---|
| 254 | in the SAGUARO analyses. These admixture events pointing to the tips of the graphs suggest |
| 255 | secondary gene flow events between nearby riverine populations and individual species within |
| 256 | the radiation. In all population graphs allowing up to 21 migration events, any admixture from |
| 257 | outgroup riverine populations appears to be coming from the Mungo and Meme rivers rather |
| 258 | than the Cross River, consistent with the closer geographic proximity of the former drainages. |
| 259 | |
| 260 | Very few genomic regions contain signatures of differential introgression between sister species |
| 261 | Very few regions of the genome introgressed into single species from outgroup riverine |
| 262 | populations (Fig 5A-C). In Stomatepia, only one region introgressed from Mungo/Meme Rivers |
| 263 | into S. pindu and only three regions into S. mariae, respectively, suggesting secondary gene flow |
| 264 | after initial diversification of Stomatepia (Table 2). Similarly, secondary introgression occurred |
| 265 | into the Konia + Pungu subclade (Table 2). However, there was also evidence of shared |
| 266 | introgression signals among sister species across all three subclades, where two subclade sister |
| 267 | species shared introgressed regions from a riverine population. Only 0.000017- 0.0000354% of |
| 268 | the genome appears introgressed into a single species of a Barombi Mbo subclade. This number |
| 269 | is smaller than suggested in the SAGUARO analysis perhaps due to the conservative |
| 270 | significance cut-offs and window size choice for f_4 statistic and that relationships observed in the |
| 271 | polyphyletic topologies are consistent with other patterns besides introgression. |
| 272 | |
| 273 | Evidence for sympatric sorting of ancestral polymorphism within a hybrid swarm |
| 274 | A few of these 10-40 kb regions with peak signals of introgression were also present in multiple |

subclades, indicating differential assortment of introgressed variation shared among clades. For

276 example, two significant f_4 outliers on linkage group 20 out of the 35 found across the genome 277 appear within the Stomatepia, Konia, and Pungu, suggesting that some of this introgression may 278 have occurred in the ancestral stages of the radiation and differentially sorted among species. 279 We also found 11 regions across the sister species pairs in which one species was more 280 similar to one outgroup riverine population while its sister species was more similar to the other 281 riverine population (Table 2). This signal is consistent with a hybrid swarm scenario due to 282 multiple colonizations by riverine populations before diversification of some of the sister species 283 and the sorting of polymorphisms brought in by these populations among incipient Barombi Mbo 284 species. For example, two regions that appear to be differentially sorted between S. galilaeus and 285 S. mariae and S. pindu from S. galilaeus CR versus S. mongo from S. galilaeus MM. Similar 286 patterns were found scattered across the genomes for K. eisentrauti and K. dikume versus P. 287 maclareni and M. myaka versus S. linelli.

288

289 Weak support for functional importance of introgressed regions for species diversification 290 Although we did find evidence of differential introgression among sister species scattered across 291 a small proportion of the genome, the types of genes found in these regions painted no clear 292 picture of how introgressed variation may have contributed to speciation (Table 2). For example, 293 differential introgression in *Stomatepia* occurred in regions with genes involved in a large range 294 of biological processes, including intracellular signal transduction, immune system response, and 295 motor neuron axon development (Table 2), with no obvious links to the highly divergent 296 morphological, ecological, or patterning traits observed between these species (Martin 2012) nor 297 to those traits normally associated with adaptive radiation in cichlid fishes such as body shape, 298 pharyngeal jaw morphology, retinal pigments, or male coloration (Kocher 2004; Barluenga et al.

2006; Wagner et al. 2012; Brawand et al. 2014; Malinsky et al. 2015; Meier et al. 2017).

300 Similarly, in both the Konia + Pungu and Myaka + Sarotherodon subclades, introgressed regions 301 were near genes involved in a large range of biological processes not clearly associated with 302 adaptive ecological traits in these species, such as K. dikume's hypoxia tolerance, P. maclareni's 303 spongivory, and *M. myaka's* zooplanktivory. For example, while there appears to be differential 304 introgression in *Konia* in a region containing pafahlb3, a gene involved in platelet activation 305 activity, and K. dikume's deep water specialization includes higher blood volume with higher 306 concentrations of hemoglobin (Green et al. 1973), it is not obvious how introgressed variation in 307 *pafah1b3* would have played a role in the evolution of these traits from studies of its function in 308 model organisms, which includes spermatogenesis and sterility in mice (Prescott et al. 2000;

309 Koizumi et al. 2003; Yan et al. 2003).

310 Similarly, the amount of genetic diversity in introgressed regions does not suggest strong 311 divergent selection on introgressed genetic variation due to hard selective sweeps. In line with 312 the presence of peaks in f_4 values in these regions, between-population diversity (D_{yy}) was 313 typically high between one of the species and its sister species (Fig. 6). However, within-314 population diversity across many of these regions was often greater or comparable to scaffold 315 and genome-wide averages (Table S2-4), suggesting these regions may not have experienced 316 hard selective sweeps that would support their role in adaptive divergence among species (Fig. 317 6). In summary, although we found evidence for differential secondary gene flow between sister 318 species in the radiation, we did not find strong functional support from gene ontology terms nor 319 signatures of selection that the introgressed alleles were important for sympatric species 320 diversification.

322 Discussion

323 Little evidence that secondary gene flow promoted the diversification of Barombi Mbo cichlids 324 Our fine-scale investigations of introgression across the genomes of a celebrated putative 325 example of sympatric speciation are consistent with two possible scenarios: 1) sympatric 326 speciation in the presence of continuous neutral secondary gene flow into the radiation, or 2) 327 speciation initiated by secondary gene flow. We found little support for the latter allopatric 328 scenario from both a learning machine and sliding-window approach. From the SAGUARO 329 analyses, our most conservative estimate of introgression into single species of the radiation 330 ranges from 0.013 -0.019% of the genome. Estimates are similarly small from the f_4 statistics, 331 ranging from 0.000017- 0.0000354% of the genome (Fig 7). Furthermore, even these significant 332 outliers may represent false positives. First, our method of selecting introgressed regions from 333 the 1% tails of a null distribution can always find outliers, even in the absence of introgression. 334 Second, it is also difficult to distinguish signatures of differential introgression from the biased 335 assortment of ancestral polymorphism into modern lineages, e.g. a hybrid swarm scenario that 336 would still result in sympatric divergence entirely within the crater lake. Finally, even if our 337 statistical outliers represent differentially introgressed regions, their importance to the speciation 338 process is equivocal. We found no evidence of selective sweeps in these regions that would 339 suggest they aided in divergence between species and they contain mainly housekeeping genes 340 that do not clearly suggest how introgressed variation would have contributed to the radiation. 341 This contrasts studies on other systems using similar approaches which found compelling 342 cases for adaptive introgression contributing to diversification (e.g. Abi-Rached et al. 2011; The 343 Heliconius Genome Consortium et al. 2012; Huerta-Sánchez et al. 2014; Lamichhaney et al. 344 2015; Stankowski and Streisfeld 2015; Arnold et al. 2016; Meier et al. 2017), including our own

345 previous work (Richards and Martin 2017). For example, several studies have found convincing 346 candidate genes/variants in introgressed regions to suggest that adaptive introgression played a 347 role in shaping ecological and morphological diversity. These include the detection of 348 introgressed alleles linked to wing-color patterning involved in mimicry and mate selection in 349 Heliconius butterflies (The Heliconius Genome Consortium et al. 2012), flower coloration 350 involved in pollinator preferences for Mimulus species (Stankowski and Streisfeld 2015), and 351 oral jaw size variation involved in scale-eating trophic specialization in *Cyprinodon* pupfishes 352 (Richards and Martin 2017).

353

Evidence for a hybrid swarm further complicates the role of gene flow in the speciation process
in Barombi Mbo cichlids

356 Beyond speciation scenarios involving secondary gene flow, our findings also suggest another 357 scenario for sympatric speciation in this system: sympatric speciation from a hybrid swarm 358 involving the differential sorting of ancestral polymorphism among incipient species. A hybrid 359 swarm is not easily detectable using the f_4 statistic because introgressed variation could be shared 360 among diverging sister species, leading to an f_4 value of zero (Reich et al. 2009; Patterson et al. 361 2012). However, many of the f_4 peaks appear to be shared across at least two of the sister species 362 in a subclade, shared between species of different subclades, or contain variation from both 363 riverine populations (Mungo/Meme and Cross Rivers) that has been differentially sorted among 364 sister species. All three of these patterns are consistent with an ancestral hybrid swarm before 365 divergence between sister species occurred. This pattern of differential sorting of variation from 366 a hybrid swarm from f_d analyses could also result from a lack of power in the statistic to 367 distinguish the directionality of the introgression detected in those regions when using biallelic

368 patterns and four populations (e.g. when two populations share similar allele patterns, the other 369 two populations can share the opposite allele pattern by default). However, we also found 370 evidence that entire subclades (e.g. *Stomatepia*) were more closely related to riverine populations 371 than other Barombi Mbo subclades from the SAGUARO analyses that are also consistent with a 372 hybrid swarm (e.g. Fig 2).

373 There are some caveats to our interpretations of secondary gene flow and its weak 374 functional role in the ecological and morphological diversity observed within the lake. 375 Recombination rate varies across genomes and determines the scale over which patterns of 376 admixture and differentiation vary (Smukowski and Noor 2011). In our fixed sliding window 377 size of 10-kb, we may have missed important patterns of introgression in regions of 378 recombination hotspots, where such patterns are expected to be very fine-scale. Shared variation 379 among species may reflect unsorted polymorphism from structured ancestral populations rather 380 than hybridization. Introgression events can also be hard to distinguish from ongoing balancing 381 selection of ancestral polymorphism that is sieved between species (Guerrero and Hahn 2017). 382 While we focused on searching for genetic signatures of hard selective sweeps in introgressed 383 regions, some of them with intermediate to high nucleotide diversity may have undergone soft 384 selective sweeps, when selection drives multiple adaptive haplotypes to fixation. Some of these 385 introgressed regions may have been adaptive and undergone soft selective sweeps, although the 386 relative contributions of hard sweeps versus soft sweeps during adaptation and speciation is still 387 the subject of much debate (Hermisson and Pennings 2005, 2017; Pritchard et al. 2010; Jensen 388 2014; Schrider et al. 2015).

389

390 Best remaining cases for sympatric speciation within Barombi Mbo cichlid radiation

391 While the radiation as a whole may not have entirely arisen from sympatric speciation, some 392 sister species within Barombi Mbo are better case studies of the process than others. Within the 393 three-species Stomatepia subclade, there is little evidence that secondary gene flow played an 394 important role in diversification. On a genome-wide level, we detected secondary gene flow in f_4 395 tests involving S. pindu. However, on a finer scale the one introgressed region unique to S. pindu 396 is unannotated and the three introgressed regions unique to S. mariae contain four housekeeping 397 genes involved in extracellular exosome activity and plasma membranes 398 (*jmjd8*, *prss1*, *cldn4*, *muc19*). Shared signals of introgression among the three species represent a 399 larger proportion of the genome than differentially introgressed regions, although both types of 400 introgressed material appear to be rare in the genome (< 0.045%; Fig 7). The high ecological and 401 morphological overlap among *Stomatepia* species suggests that this species complex may be 402 stalled in the earliest stages of divergence. For example, S. pindu and S. mariae appear be at the 403 extremes of a unimodal distribution of phenotypes; this is one major prediction of sympatric 404 speciation models in the presence of only weak disruptive selection on ecological traits (e.g. 405 (Matessi et al. 2002; Burger et al. 2006)), as measured in this species pair (Martin 2012). 406 Even for the two monotypic specialist species *M. myaka* and *P. maclareni*, there is 407 minimal evidence for a role of secondary gene flow in the evolution of their trophic 408 specializations. On a genome-wide level, the tests for differential introgression with one of the 409 specialists and another species from the radiation were not significant. For the two regions which 410 appear to be differentially introgressed between riverine populations and the zooplantivore M. 411 *myaka*, one has no annotations, suggesting it may be neutral gene flow while the other is an 412 introgressed region in other subclades, a signal of differential sorting of variation after a hybrid 413 swarm. For spongivore specialist *Pungu maclareni*, we found only a single region differentially

414 introgressed with riverine populations. This region contains ddn1, a gene involved in serine-type 415 peptidase activity, proteins which are found ubiquitously in prokaryotes and eukaryotes. 416 Among all the ecologically divergent species pairs focused on in this study, K. eisentrauti 417 and K. dikume are the least convincing as a putative case for sympatric speciation between sister 418 species. Similar to *Stomatepia*, there is more evidence for shared introgression in regions of the 419 Konia genome than differentially introgressed regions (Fig 8). However, differential 420 introgression between the two Konia species occurs in regions with the best potential candidates 421 in this study for contributing to diversification (although these regions are still not as strong as 422 the 'smoking guns' observed in our past study of introgressed variation, e.g. Richards and Martin 423 2017). The gene *pafah1b3*, which is involved in platelet activation activity, is differentially 424 introgressed between Konia species and may function in K. dikume's deep water specialization 425 of higher blood volume with higher concentrations of hemoglobin (Green et al. 1973). However, 426 studies of the phenotypic effects of *pafah1b3* in model organisms do not hint at a role in 427 hemoglobin concentration. Instead, mutations in this gene have been suggested to play a role in 428 male spermatogenesis and fertility in mice (Koizumi et al. 2003; Yan et al. 2003). We also see 429 differential introgression in a region containing the gene *ehmt2*, which is involved in 430 neurogenesis and retinal cell differentiation. While it is not as directly clear how introgressed 431 variation in this gene would have contributed to divergence among the *Konia* sister species, 432 variation in visual traits such as color perception are important axes of diversification in other 433 cichlid radiations, particularly along a depth gradient (Terai et al. 2002, 2006; Seehausen et al. 434 2008). These two species also exist in microallopatry; K. eisentrauti is an abundant detritivore 435 along the shallow littoral region of the lake while K. dikume is a deep-water specialist on

436 *Chaoborus* midge larvae which is only collected in deep-water gill nets (Trewavas et al. 1972;

437 Schliewen 1994). Both species are mouthbrooders and likely breed in non-overlapping habitats.

438

439 Conclusion

The complex history of colonization in the crater evidenced in this and a previous genome-wide 440 441 study suggests allopatric phases of the speciation process in the radiation, which violates one of 442 the strict criteria for demonstrating sympatric speciation in the wild (Coyne and Orr 2004). 443 Nonetheless, from our fine-scale dissection of where in the genome these signals are coming 444 from, we cannot point to a functional role for secondary gene flow in the speciation process 445 across any of the subclades. This suggests that either variation in genes with undiscovered 446 functional effects underlies the divergent ecologies and morphologies seen in the lake or that any 447 secondary gene flow was neutral with regard to its role in the speciation process. We also find 448 evidence to support sympatric speciation after a hybrid swarm that resulted from multiple 449 colonizations of the lake, still consistent with a scenario of sympatric speciation through 450 differential sorting of ancestral polymorphism. Disentangling the effects of a putative hybrid 451 swarm from secondary contact events on the speciation process will require a better 452 understanding of the timing of gene flow events compared to the diversification times of 453 Barombi Mbo species. We found evidence for gene flow into the radiation both before and after 454 initial diversification of subclades within the lake. Even without this information, weak support 455 for a functional role of secondary gene flow in the radiation of Barombi Mbo cichlids suggests 456 that we should not rule out this example of a sympatric radiation just yet.

457

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| 470 | contributed to the development of ideas presented in the study and revised the manuscript. | | | | |
| 471 | | | | | |
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| 473 | All datasets used for this study will be deposited in Dryad and the NCBI Short Read Archive | | | | |
| 474 | (SRA). | | | | |
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482 **References**

- 483 Abi-Rached, L., M. J. Jobin, S. Kulkarni, A. McWhinnie, K. Dalva, L. Gragert, F. Babrzadeh, B.
- 484 Gharizadeh, M. Luo, F. A. Plummer, J. Kimani, M. Carrington, D. Middleton, R.
- 485 Rajalingam, M. Beksac, S. G. E. Marsh, M. Maiers, L. A. Guethlein, S. Tavoularis, A.-M.
- 486 Little, R. E. Green, P. J. Norman, and P. Parham. 2011. The Shaping of Modern Human
- 487 Immune Systems by Multiregional Admixture with Archaic Humans. Science (80-.).
- 488 334:89 LP-94.
- 489 Alcaide, M., E. S. C. Scordato, T. D. Price, and D. E. Irwin. 2014. Genomic divergence in a ring
- 490 species complex. Nature 511:83–85. Nature Publishing Group.
- 491 Arnold, B. J., B. Lahner, J. M. DaCosta, C. M. Weisman, J. D. Hollister, D. E. Salt, K. Bomblies,
- 492 and L. Yant. 2016. Borrowed alleles and convergence in serpentine adaptation. Proc. Natl.
 493 Acad. Sci. 113:8320–8325.
- 494 Balsa-Canto, E., D. Henriques, A. Gabor, and J. R. Banga. 2016. AMIGO2, a toolbox for
- 495 dynamic modeling, optimization and control in systems biology. Bioinformatics 32:1–2.
- 496 Barluenga, M., K. N. Stölting, W. Salzburger, M. Muschick, and A. Meyer. 2006. Sympatric

497 speciation in Nicaraguan crater lake cichlid fish. Nature 439:719–723.

498 Bolnick, D. I., and B. M. Fitzpatrick. 2007. Linked references are available on JSTOR for this

499 article : Sympatric Speciation : Models and Empirical Evidence. Annu. Rev. Ecol. Evol.

- 500 Syst. 38:459–487.
- 501 Brawand, D., C. E. Wagner, Y. I. Li, M. Malinsky, I. Keller, S. Fan, O. Simakov, A. Y. Ng, Z.
- 502 W. Lim, E. Bezault, Turner-Maier, J. J. Johnson, R. Alcazar, H. J. Noh, P. Russell, B. Aken,
- 503 J. Alföldi, C. Amemiya, N. Azzouzi, J.-F. Baroiller, F. Barloy-Hubler, A. Berlin, R.
- 504 Bloomquist, K. L. Carleton, M. a. Conte, H. D'Cotta, O. Eshel, L. Gaffney, F. Galibert, H.

| 505 | F. Gante, S. Gnerre, L. Greuter, R. Guyon, N. S. Haddad, W. Haerty, R. M. Harris, H. a. |
|-----|---|
| 506 | Hofmann, T. Hourlier, G. Hulata, D. B. Jaffe, M. Lara, L. A.P., I. MacCallum, S. Mwaiko, |
| 507 | M. Nikaido, H. Nishihara, C. Ozouf-Costaz, D. J. Penman, D. Przybylski, M. Rakotomanga, |
| 508 | S. C. P. Renn, F. J. Ribeiro, M. Ron, W. Salzburger, L. Sanchez-Pulido, M. E. Santos, S. |
| 509 | Searle, T. Sharpe, R. Swofford, F. J. Tan, L. Williams, S. Young, S. Yin, N. Okada, T. D. |
| 510 | Kocher, E. a. Miska, E. S. Lander, B. Venkatesh, R. D. Fernald, A. Meyer, C. P. Ponting, J. |
| 511 | T. Streelman, K. Lindblad-Toh, O. Seehausen, and F. Di Palma. 2014. The genomic |
| 512 | substrate for adaptive radiation in African cichlid fish. Nature 513:375–381. |
| 513 | Burger, R., K. A. Schneider, and M. Willensdorfer. 2006. The Conditions for Speciation through |
| 514 | Intraspecific Competition. Evolution (N. Y). 60:2185–2206. [Society for the Study of |
| 515 | Evolution, Wiley]. |
| 516 | Coyne, J. A., and H. A. Orr. 2004. Speciation. Sutherland, MA. |
| 517 | Cruickshank, T. E., and M. W. Hahn. 2014. Reanalysis suggests that genomic islands of |
| 518 | speciation are due to reduced diversity, not reduced gene flow. Mol. Ecol. 23:3133-3157. |
| 519 | Cutter, A. D., and B. A. Payseur. 2013. Genomic signatures of selection at linked sites: unifying |
| 520 | the disparity among species. Nat. Rev. Genet. 14:262–274. Nature Publishing Group. |
| 521 | Edmunds, R. C., B. Su, J. P. Balhoff, B. F. Eames, W. M. Dahdul, H. Lapp, J. G. Lundberg, T. J. |
| 522 | Vision, R. A. Dunham, P. M. Mabee, and M. Westerfield. 2016. Phenoscape: Identifying |
| 523 | candidate genes for evolutionary phenotypes. Mol. Biol. Evol. 33:13-24. |
| 524 | Feder, J. L., S. P. Egan, and P. Nosil. 2012. The genomics of speciation-with-gene-flow. Trends |
| 525 | Genet. 28:342–350. |
| 526 | Geiger, M. F., J. K. McCrary, and U. K. Schliewen. 2013. Crater Lake Apoyo Revisited - |
| 527 | Population Genetics of an Emerging Species Flock. PLoS One 8:1–17. |
| | |

- 528 Green, J., S. A. Corbet, and E. Betney. 1973. Ecological studies on crater lakes in West
- 529 Cameroon The blood of endemic cichlids in Barombi Mbo in relation to stratification and
- 530 their feeding habits. J. Zool. 170:299–308. Blackwell Publishing Ltd.
- 531 Guerrero, R. F., and M. W. Hahn. 2017. Speciation as a sieve for ancestral polymorphism. Mol.
- 532 Ecol. 1–7.
- 533 Harr, B. 2006. Genomic islands of differentiation between house mouse subspecies Genomic
- islands of differentiation between house mouse subspecies. 730–737.
- 535 Hermisson, J., and P. S. Pennings. 2005. Soft Sweeps. Genetics 169:2335 LP-2352.
- 536 Hermisson, J., and P. S. Pennings. 2017. Soft sweeps and beyond: understanding the patterns and
- 537 probabilities of selection footprints under rapid adaptation. Methods Ecol. Evol. 8:700–716.
- 538 Huerta-Sánchez, E., X. Jin, Asan, Z. Bianba, B. M. Peter, N. Vinckenbosch, Y. Liang, X. Yi, M.
- He, M. Somel, P. Ni, B. Wang, X. Ou, Huasang, J. Luosang, Z. X. P. Cuo, K. Li, G. Gao,
- 540 Y. Yin, W. Wang, X. Zhang, X. Xu, H. Yang, Y. Li, J. Wang, J. Wang, and R. Nielsen.
- 541 2014. Altitude adaptation in Tibetans caused by introgression of Denisovan-like DNA.
- 542 Nature 512:194–197.
- 543 Igea, J., D. Bogarín, A. S. T. Papadopulos, and V. Savolainen. 2015. A comparative analysis of
- island floras challenges taxonomy-based biogeographical models of speciation. Evolution
 (N. Y). 69:482–491.
- Jensen, J. D. 2014. On the unfounded enthusiasm for soft selective sweeps. Nat. Commun.
 547 5:5281.
- Jones, F. C., M. G. Grabherr, Y. F. Chan, P. Russell, E. Mauceli, J. Johnson, R. Swofford, M.
- 549 Pirun, M. C. Zody, S. White, E. Birney, S. Searle, J. Schmutz, J. Grimwood, M. C. Dickson,
- 550 R. M. Myers, C. T. Miller, B. R. Summers, A. K. Knecht, S. D. Brady, H. Zhang, A. A.

| 551 | Pollen, T. Howes, C. Amemiya, Broad Institute Genome Sequencing Platform & Whole |
|-----|---|
| 552 | Genome Assembly Team, E. S. Lander, F. Di Palma, K. Lindblad-Toh, and D. M. Kingsley. |
| 553 | 2012. The genomic basis of adaptive evolution in threespine sticklebacks. Nature 484:55– |
| 554 | 61. |
| 555 | Kautt, A. F., G. Machado-Schiaffino, J. Torres-Dowdall, and A. Meyer. 2016. Incipient |
| 556 | sympatric speciation in Midas cichlid fish from the youngest and one of the smallest crater |
| 557 | lakes in Nicaragua due to differential use of the benthic and limnetic habitats? Ecol. Evol. |
| 558 | 6:5342–5357. |
| 559 | Kocher, T. D. 2004. Adaptive evolution and explosive speciation: the cichlid fish model. Nat. |
| 560 | Rev. Genet. 5:288–298. |
| 561 | Koizumi, H., N. Yamaguchi, M. Hattori, T. O. Ishikawa, J. Aoki, M. M. Taketo, K. Inoue, and |
| 562 | H. Arai. 2003. Targeted disruption of intracellular type I platelet activating factor- |
| 563 | acetylhydrolase catalytic subunits causes severe impairment in spermatogenesis. J. Biol. |
| 564 | Chem. 278:12489–12494. |
| 565 | Lamichhaney, S., J. Berglund, M. S. Almén, K. Maqbool, M. Grabherr, A. Martinez-Barrio, M. |
| 566 | Promerová, CJ. Rubin, C. Wang, N. Zamani, B. R. Grant, P. R. Grant, M. T. Webster, and |
| 567 | L. Andersson. 2015. Evolution of Darwin's finches and their beaks revealed by genome |
| 568 | sequencing. Nature 518:371–375. |
| 569 | Mabee, P., J. P. Balhoff, W. M. Dahdul, H. Lapp, P. E. Midford, T. J. Vision, and M. |
| 570 | Westerfield. 2012. 500,000 fish phenotypes: The new informatics landscape for |
| 571 | evolutionary and developmental biology of the vertebrate skeleton. J. Appl. Ichthyol. |
| 572 | 28:300–305. |
| 573 | Malinsky, M., R. J. Challis, A. M. Tyers, S. Schiffels, Y. Terai, B. P. Ngatunga, E. A. Miska, R. |
| | |

| 574 | Durbin, M. J. Genner, and G. F. Turner. 2015. Genomic islands of speciation separate |
|-----|--|
| 575 | cichlid ecomorphs in an East African crater lake. Science (80). 350:1493–1498. |
| 576 | Manda, P., J. P. Balhoff, H. Lapp, P. Mabee, and T. J. Vision. 2015. Using the phenoscape |
| 577 | knowledgebase to relate genetic perturbations to phenotypic evolution. Genesis 53:561- |
| 578 | 571. |
| 579 | Martin, C. H. 2012. Weak Disruptive Selection and Incomplete Phenotypic Divergence in Two |
| 580 | Classic Examples of Sympatric Speciation: Cameroon Crater Lake Cichlids. Am. Nat. |
| 581 | 180:E90–E109. |
| 582 | Martin, C. H., J. S. Cutler, J. P. Friel, C. Dening Touokong, G. Coop, and P. C. Wainwright. |
| 583 | 2015a. Complex histories of repeated gene flow in Cameroon crater lake cichlids cast doubt |
| 584 | on one of the clearest examples of sympatric speciation. Evolution (N. Y). 69:1406–1422. |
| 585 | Martin, S. H., K. K. Dasmahapatra, N. J. Nadeau, C. Slazar, J. R. Walters, F. Simpson, M. |
| 586 | Blaxter, A. Manica, J. Mallet, and C. D. Jiggins. 2013. Genome-wide evidence for |
| 587 | speciation with gene flow in Heliconius butterflies. Genome Res. 23:1817-1828. |
| 588 | Martin, S. H., J. W. Davey, and C. D. Jiggins. 2015b. Evaluating the use of ABBA-BABA |
| 589 | statistics to locate introgressed loci. Mol. Biol. Evol. 32:244-257. |
| 590 | Matessi, C., A. Gimelfarb, and S. Gavrilets. 2002. Long-term Buildup of Reproductive Isolation |
| 591 | Promoted by Disruptive Selection: How Far Does it Go? Selection 2:41-64. Akadémiai |
| 592 | Kiadó. |
| 593 | McGirr, J. A., and C. H. Martin. 2016. Novel candidate genes underlying extreme trophic |
| 594 | specialization in Caribbean pupfishes. Mol. Biol. Evol. msw286. |
| 595 | Meier, J. I., D. A. Marques, S. Mwaiko, C. E. Wagner, L. Excoffier, and O. Seehausen. 2017. |
| 596 | Ancient hybridization fuels rapid cichlid fish adaptive radiations. Nat. Commun. 8:14363. |
| | |

597 Nature Publishing Group.

| 598 | Midford, P. E., T. Dece | cchi, J. P. Balhoff, V | V. M. Dahdul, N. Ibra | him, H. Lapp, J. G. Lundbe | rg. |
|-------|-------------------------|------------------------|-----------------------|----------------------------|------|
| - / - | | | | | - 07 |

- 599 P. M. Mabee, P. C. Sereno, M. Westerfield, T. J. Vision, D. C. Blackburn, S. Federhen, P.
- Mabee, J. Balhoff, W. Dahdul, H. Lapp, P. Midford, T. Vision, M. Westerfield, W. Dahdul,
- J. Balhoff, J. Engeman, T. Grande, E. Hilton, C. Kothari, H. Lapp, J. Lundberg, P. Midford,
- T. Vision, M. Westerfield, P. Mabee, F. Fang, M. Toledo-Piza, M. Uhen, A. Barnosky, B.
- Bills, J. Blois, M. Carrano, M. Carrasco, G. Erickson, J. Eronen, M. Fortelius, R. Graham,
- E. Grimm, M. O'Leary, A. Mast, W. Piel, P. Polly, L. Saila, J. Balhoff, P. Midford, H.
- 605 Lapp, M. Ghiselin, S. Schulz, H. Stenzhorn, M. Boeker, D. Thau, N. Franz, B. Smith, M.
- Ashburner, C. Rosse, J. Bard, W. Bug, W. Ceusters, L. Goldberg, K. Eilbeck, A. Ireland, C.
- 607 Mungall, O. Consortium, N. Leontis, P. Rocca-Seraa, A. Ruttenberg, S. Sansone, R.
- 608 Scheuermann, N. Shah, P. Whetzel, S. Lewis, J. Beaulieu, R. Ree, J. Cavender-Bares, G.
- 609 Weiblen, and M. Donoghue. 2013. The vertebrate taxonomy ontology: a framework for
- 610 reasoning across model organism and species phenotypes. J. Biomed. Semantics 4:34.
- 611 Nachman, M. W., and B. A. Payseur. 2012. Recombination rate variation and speciation:
- 612 theoretical predictions and empirical results from rabbits and mice. Philos. Trans. R. Soc. B
 613 Biol. Sci. 367:409–421.
- Noor, M., and S. Bennett. 2009. Islands of speciation or mirages in the desert? Examingin the
- role of restricted recombination in maintaining species. Heredity (Edinb). 103:439–444.
- 616 Nosil, P., and J. L. Feder. 2012a. Genomic divergence during speciation: causes and
- 617 consequences. Philos. Trans. R. Soc. B Biol. Sci. 367:332–342.
- 618 Nosil, P., and J. L. Feder. 2012b. Widespread yet heterogeneous genomic divergence. Mol. Ecol.
- 619 21:2829–2832.

- 620 Papadopulos, A. S. T., W. J. Baker, D. Crayn, R. K. Butlin, R. G. Kynast, I. Hutton, and V.
- 621 Savolainen. 2011. Speciation with gene flow on Lord Howe Island. Proc. Natl. Acad. Sci.
 622 108:13188–13193.
- 623 Patterson, N., P. Moorjani, Y. Luo, S. Mallick, N. Rohland, Y. Zhan, T. Genschoreck, T.
- Webster, and D. Reich. 2012. Ancient Admixture in Human History. Genetics 192:1069–
 1093.
- 626 Pickrell, J. K., and J. K. Pritchard. 2012. Inference of Population Splits and Mixtures from
 627 Genome-Wide Allele Frequency Data. PLoS Genet. 8.
- 628 Poelstra, J. W., N. Vijay, C. M. Bossu, H. Lantz, B. Ryll, I. Muller, V. Baglione, P. Unneberg,
- 629 M. Wikelski, M. G. Grabherr, and J. B. W. Wolf. 2014. The genomic landscape underlying
- 630 phenotypic integrity in the face of gene flow in crows. Science (80-.). 344:1410–1414.
- 631 Prescott, S. M., G. a Zimmerman, D. M. Stafforini, and T. M. Mcintyre. 2000. Platelet activating
 632 factor. Annu. Rev. Biochem. 69:419–445.
- 633 Pritchard, J. K., J. K. Pickrell, and G. Coop. 2010. The Genetics of Human Adaptation: Hard
- 634 Sweeps, Soft Sweeps, and Polygenic Adaptation. Curr. Biol. 20:R208–R215.
- 635 Ravinet, M., R. Faria, R. K. Butlin, J. Galindo, N. Bierne, M. Rafajlović, M. A. F. Noor, B.
- Mehlig, and A. M. Westram. 2017. Interpreting the genomic landscape of speciation: a road
 map for finding barriers to gene flow. J. Evol. Biol. 30:1450–1477.
- Reich, D., K. Thangaraj, N. Patterson, A. L. Price, and L. Singh. 2009. Reconstructing Indian
 population history. Nature 461:489–94.
- 640 Richards, E. J., and C. H. Martin. 2017. Adaptive introgression from distant Caribbean islands
- 641 contributed to the diversification of a microendemic adaptive radiation of trophic specialist
- 642 pupfishes.

- 643 Schliewen, U. K., and B. Klee. 2004. Reticulate sympatric speciation in Cameroonian crater lake
 644 cichlids. Front. Zool. 1:5.
- 645 Schliewen, U. K., D. Tautz, and S. Paabo. 1994. Sympatric speciation suggested by monophyly
- of crater lake cichlids. Nature 368:629–631.
- 647 Schrider, D. R., F. K. Mendes, M. W. Hahn, and A. D. Kern. 2015. Soft Shoulders Ahead:
- 648 Spurious Signatures of Soft and Partial Selective Sweeps Result from Linked Hard Sweeps.
- 649 Genetics 200:267 LP-284.
- 650 Seehausen, O., Y. Terai, I. S. Magalhaes, K. L. Carleton, H. D. J. Mrosso, R. Miyagi, I. van der
- 651 Sluijs, M. V. Schneider, M. E. Maan, H. Tachida, H. Imai, and N. Okada. 2008. Speciation
- through sensory drive in cichlid fish. Nature 455:620–626.
- Smukowski, C. S., and M. A. F. Noor. 2011. Recombination rate variation in closely related
 species. Heredity (Edinb). 107:496–508. The Genetics Society.
- 655 Soria-Carrasco, V., Z. Gompert, A. A. Comeault, T. E. Farkas, T. L. Parchman, J. S. Johnston, C.
- A. Buerkle, J. L. Feder, J. Bast, T. Schwander, S. P. Egan, B. J. Crespi, and P. Nosil. 2014.
- 657 Stick Insect Genomes Reveal Natural Selection's Role in Parallel Speciation. Science (80-.
- 658). 344:738 LP-742.
- 659 Stankowski, S., and M. A. Streisfeld. 2015. Introgressive hybridization facilitates adaptive
- divergence in a recent radiation of monkeyflowers. Proc. R. Soc. London B 282:20151666.
- Terai, Y., W. E. Mayer, J. Klein, H. Tichy, and N. Okada. 2002. The effect of selection on a long
- 662 wavelength-sensitive (LWS) opsin gene of Lake Victoria cichlid fishes. Proc. Natl. Acad.
- 663 Sci. 99:15501–15506.
- 664 Terai, Y., O. Seehausen, T. Sasaki, K. Takahashi, S. Mizoiri, T. Sugawara, T. Sato, M.
- 665 Watanabe, N. Konijnendijk, H. D. J. Mrosso, H. Tachida, H. Imai, Y. Shichida, and N.

- 666 Okada. 2006. Divergent selection on opsins drives incipient speciation in Lake Victoria
 667 cichlids. PLoS Biol. 4:2244–2251.
- 668 The Heliconius Genome Consortium, K. K. Dasmahapatra, J. R. Walters, A. D. Briscoe, J. W.
- 669 Davey, A. Whibley, N. J. Nadeau, A. V. Zimin, D. S. T. Hughes, L. C. Ferguson, S. H.
- 670 Martin, C. Salazar, J. J. Lewis, S. Adler, S.-J. Ahn, D. a. Baker, S. W. Baxter, N. L.
- 671 Chamberlain, R. Chauhan, B. a. Counterman, T. Dalmay, L. E. Gilbert, K. Gordon, D. G.
- Heckel, H. M. Hines, K. J. Hoff, P. W. H. Holland, E. Jacquin-Joly, F. M. Jiggins, R. T.
- Jones, D. D. Kapan, P. Kersey, G. Lamas, D. Lawson, D. Mapleson, L. S. Maroja, A.
- Martin, S. Moxon, W. J. Palmer, R. Papa, A. Papanicolaou, Y. Pauchet, D. a. Ray, N.
- 675 Rosser, S. L. Salzberg, M. a. Supple, A. Surridge, A. Tenger-Trolander, H. Vogel, P. a.
- 676 Wilkinson, D. Wilson, J. a. Yorke, F. Yuan, A. L. Balmuth, C. Eland, K. Gharbi, M.
- 677 Thomson, R. a. Gibbs, Y. Han, J. C. Jayaseelan, C. Kovar, T. Mathew, D. M. Muzny, F.
- 678 Ongeri, L.-L. Pu, J. Qu, R. L. Thornton, K. C. Worley, Y.-Q. Wu, M. Linares, M. L.
- Blaxter, R. H. Ffrench-Constant, M. Joron, M. R. Kronforst, S. P. Mullen, R. D. Reed, S. E.
- 680 Scherer, S. Richards, J. Mallet, W. Owen McMillan, and C. D. Jiggins. 2012. Butterfly
- genome reveals promiscuous exchange of mimicry adaptations among species. Nature487:94–98.
- Trewavas, E., J. Green, and S. A. Corbet. 1972. Ecological studies on crater lakes in West
 Cameroon fishes of Barombi Mbo. J. Zool. 41–95.
- Turner, T. L., M. W. Hahn, and S. V. Nuzhdin. 2005. Genomic islands of speciation in
 Anopheles gambiae. PLoS Biol. 3:1572–1578.
- 687 Wagner, C. E., L. J. Harmon, and O. Seehausen. 2012. Ecological opportunity and sexual
- 688 selection together predict adaptive radiation. Nature 487:366–369. Nature Publishing

| 689 | Group. |
|-----|--|
| 690 | Wu, CI., and CT. Ting. 2004. Genes and speciation. Nat. Rev. Genet. 5:114-122. |
| 691 | Wu, C. I. 2001. The genic view of the process of speciation. J. Evol. Biol. 14:851-865. |
| 692 | Yan, W., A. H. Assadi, A. Wynshaw-Boris, G. Eichele, M. M. Matzuk, and G. D. Clark. 2003. |
| 693 | Previously uncharacterized roles of platelet-activating factor acetylhydrolase 1b complex in |
| 694 | mouse spermatogenesis. Proc. Natl. Acad. Sci. U. S. A. 100:7189–94. |
| 695 | Zamani, N., P. Russell, H. Lantz, M. P. Hoeppner, J. R. Meadows, N. Vijay, E. Mauceli, F. Di |
| 696 | Palma, K. Lindblad-Toh, P. Jern, and M. G. Grabherr. 2013. Unsupervised genome-wide |
| 697 | recognition of local relationship patterns. BMC Genomics 14:347. |
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- 715 **Table 1**. Genome-wide f_4 statistics supporting differential introgression within Barombi Mbo
- radiation. Tests with significant evidence for differential introgression are highlighted in bold.
- 717 The f_4 statistic was calculated for pairwise combinations among sister species of Barombi Mbo
- 518 subclades and riverine populations of S. galilaeus from Mungo and Meme Rivers (MM) and

719 Cross River (CR).

| | Introgression with riverine outgroups: (A,B) <-> (<i>MM, CR</i>) | f4 statistic | Z-score | P-value |
|-----|---|---|---------|-----------------------------|
| | S. mariae, S. mongo | $-2.04 \times 10^{-7} \pm -5.15 \times 10^{-7}$ | -0.39 | 0.69 |
| | S. mariae, S. pindu | -1.92x10 ⁻⁶ ± -4.48x10 ⁻⁷ | -4.29 | 1.8x10 ⁻⁵ |
| | S. mongo, S. pindu | -1.59x10 ⁻⁶ ± -4.98x10 ⁻⁷ | -3.19 | 0.0014 |
| | K. dikume, K. eisentrauti | -2.4x10 ⁻⁶ ± -6 x10 ⁻⁷ | -4.01 | 6.3x10 ⁻⁵ |
| | K. eisentrauti, P. maclareni | -2.12x10 ⁻⁷ ± -6.15x10 ⁻⁷ | 0.35 | 0.73 |
| | K. dikume, P. maclareni | -2.56x10 ⁻⁶ ± -5.86x10 ⁻⁷ | -4.37 | 1.2x10 ⁻⁵ |
| | M. myaka, S. linnelli | $-4.04 \times 10^{-7} \pm -7.11 \times 10^{-7}$ | 0.56 | 0.57 |
| 720 | | | | |
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| 730 | Table 2 . Candidate introgressed regions in Barombi Mbo cichlid radiation. These regions feature |
|-----|---|
| 731 | significant f_4 values between riverine populations of S. galilaeus (MM: Mungo and Meme River; |
| 732 | CR: Cross River) and the three subclades of the radiation focused on in this study. Directionality |
| 733 | was determined from f_d outlier values (above 98 th percentile) that overlapped with significant f_4 |
| 734 | peaks. Unannotated regions, regions with no GO terms, and regions that were not f_d outliers are |
| 735 | marked with (-). Regions introgressed into a single species within a subclade are highlighted in |
| | |

736 bold.

| Linkage Group | Gene regions | GO Terms | Direction (f_d) | | | |
|------------------|---|---|---|--|--|--|
| | | Stomatepia | | | | |
| LG2 | sirpb1 | intracellular signal transduction, extracellular exosome | - | | | |
| LG3 | NA | - | S. galilaeus MM <-> S. pindu | | | |
| LG3 | uncharacterized protein-coding gene | - | S. galilaeus MM <->S. mongo, mariae | | | |
| LG4 | jmjd8; prss1 | extracellular exosome; proteolysis,peptidase activity | S. galileaus CR <-> S. mariae | | | |
| LG7† | NA | - | S. galileaus CR <-> S. pindu, mariae; S. galilaeus MM <- > S. mongo | | | |
| LG14 | cldn4 | plasma membrane; bicellular tight membrane | S. galilaeus MM <-> S. mariae | | | |
| LG18 | muc19 | hematopoietic progenitor cell differentiation; extracellular exosome | S. galilaeus MM <-> S. mariae | | | |
| LG20† | tprxl; plod3 | pseudogene of <i>tprx1</i> ;motor nueron axon guidance, nueral crest cell migration | S. galileaus CR <-> S. pindu, mariae; S. galilaeus MM<- > S. mongo | | | |
| LG20 | samdh1 | dGTPASe, immune system process, brain hemorrhages | - | | | |
| NT_168079.1† | osbpl5;ptprj | lipid transport ; artery morphogenesis, phosphatase activity | S. galileaus CR<-> S. pindu,mariae; S. galilaeus MM<- > S. mongo | | | |
| | Konia + Pungu | | | | | |
| LG2 | sirpb1 | intracellular signal transduction | - | | | |
| LG4 | anapc2 | ubiquitin dependent catabolic process | S. galilaeus MM <-> P. maclareni, K. dikume | | | |
| LG6† | cd209e | endocytosis, mannose binding | S. galileaus CR<-> K. eisentrauti, P. maclareni; S. galilaeus MM<-> K. dikume | | | |
| LG15 | adgrf4 | g-protien coupled receptor activity, signal transduction | S. galilaeus MM<-> K. dikume | | | |

| LG20† | klhdc8h | ubiquitin-protein transferase activity cytoplasm | S. galilaeus CR<-> P. maclareni; |
|---|--|--|--|
| | <i>Minuc</i> 00 | uolquiun protein transierase activity, cytopiasin | S. galilaeus MM<-> K. dikume, eisentrauti |
| LG20 | NA | - | S. galilaeus MM <-> K. dikume, eisentrauti |
| LG20 | tprxl; plod3;clec10a | pseudogene of <i>tprx1</i> ; motor nueron axon guidance, nueral crest cell migration ; immune system response | - |
| LG22 | ehmt2 | neurogenesis, camera-type eye photoreceptor cell differentiation | - |
| LG23 | ddn1 | serine-type endopeptidase activity,proteolysis | S. galilaeus MM<-> P. maclareni |
| NT_167508.1† | fam159a | integral component of membrane | S. galileaus CR<-> K. dikume, eisentrauti; S. galilaeus MM<-> P. maclareni |
| NT_167623.1 | терсе | methyltransferase activity | S. galilaeus MM<-> K. eisentrauti |
| NT_167637.1 | NA | - | S. galilaeus MM<-> K. dikume |
| NT_167671.1 | NA | - | S. galileaus CR<-> K. dikume |
| NT_167702.1* | pafah1b3; hmcn1 | platelet-activating factor acetyltransferase activity,brain development;fin morphogenesis, calcium ion binding | S. galileaus CR<-> K. eisentrauti |
| | | respiratory abnormal muscle physiology g-protien | |
| NT_168003.1 | p2ry14 | coupled purinergic nucleotide receptor activity | - |
| NT_168003.1 | p2ry14 | coupled purinergic nucleotide receptor activity Myaka + S. linnelli | - |
| NT_168003.1 LG6† | <i>p2ry14</i> | coupled purinergic nucleotide receptor activity Myaka + S. linnelli complement activation, inflammatory response | - S. galilaeus MM<->S. linnelli; S. galileaus CR<-> M.myaka |
| NT_168003.1 LG6† LG8/LG24† | <i>p2ry14</i> <i>c3</i> NA | coupled purinergic nucleotide receptor activity Myaka + S. linnelli complement activation, inflammatory response | - S. galilaeus MM<>S. linnelli; S. galileaus CR<-> M.myaka S. galileaus CR<->S. linnelli; S. galilaeus MM<-> M.mvaka |
| NT_168003.1 LG6† LG8/LG24† LG11† | p2ry14 c3 NA hfe2;txnip | | - S. galilaeus MM<>S. linnelli; S. galileaus CR<-> M.myaka S. galileaus CR<->S. linnelli; S. galilaeus MM<-> M.myaka S. galilaeus MM<->S. linnelli; S. galileaus CR<-> M.myaka |
| NT_168003.1 LG6† LG8/LG24† LG11† LG11 | p2ry14 c3 NA hfe2;txnip vmo1 | coupled purinergic nucleotide receptor activity Myaka + S. linnelli complement activation, inflammatory response - somite development, BMP signaling pathway; enzyme inhibitor activity extracellular exosome | - S. galilaeus MM<>S. linnelli; S. galileaus CR<-> M.myaka S. galileaus CR<->S. linnelli; S. galilaeus MM<-> M.myaka S. galilaeus MM<->S. linnelli; S. galileaus CR<-> M.myaka S. galilaeus MM<-> M.myaka |
| NT_168003.1 LG6† LG8/LG24† LG11† LG11 LG16/LG21 | p2ry14 c3 NA hfe2;txnip vmo1 parp4 | Image: complement of physiclegy, g protein Myaka + S. linnelli complement activation, inflammatory response - somite development, BMP signaling pathway; enzyme inhibitor activity extracellular exosome ribosyltranferase activity | - S. galilaeus MM<>S. linnelli; S. galileaus CR<-> M.myaka S. galileaus CR<->S. linnelli; S. galilaeus MM<-> M.myaka S. galilaeus CR<-> M.myaka S. galilaeus MM<-> M.myaka S. galilaeus MM<-> M.myaka S. galilaeus CR<-> S. linnelli |
| NT_168003.1 LG6† LG8/LG24† LG11† LG11 LG11 LG19† | <i>p2ry14</i> <i>c3</i> NA <i>hfe2;txnip</i> <i>vmo1</i> <i>parp4</i> NA | coupled purinergic nucleotide receptor activity Myaka + S. linnelli complement activation, inflammatory response - somite development, BMP signaling pathway; enzyme inhibitor activity extracellular exosome ribosyltranferase activity - | - S. galilaeus MM<->S. linnelli; S. galileaus CR<-> M.myaka S. galileaus CR<->S. linnelli; S. galilaeus MM<-> M.myaka S. galilaeus CR<-> M.myaka S. galilaeus MM<-> M.myaka S. galileaus CR<->S. linnelli S. galileaus CR<->S. linnelli; S. galileaus CR<->S. linnelli; S. galilaeus MM<-> M.myaka |
| NT_168003.1 LG6† LG8/LG24† LG11† LG11 LG16/LG21 LG19† LG20 | <i>p2ry14</i> <i>c3</i> NA <i>hfe2;txnip</i> <i>vmo1</i> <i>parp4</i> NA <i>tprx1,plod3</i> | coupled purinergic nucleotide receptor activity Myaka + S. linnelli complement activation, inflammatory response - somite development, BMP signaling pathway; enzyme inhibitor activity extracellular exosome ribosyltranferase activity - psuedogene of tetra-peptide repeat homeobox ; motor nueron axon guidance, nueral crest cell migration | - S. galilaeus MM<>S. linnelli; S. galileaus CR<-> M.myaka S. galileaus CR<->S. linnelli; S. galilaeus MM<-> S. linnelli; S. galilaeus MM<-> S. linnelli; S. galilaeus MM<-> M.myaka S. galilaeus MM<-> M.myaka S. galileaus CR<->S. linnelli S. galileaus CR<->S. linnelli; S. galileaus CR<->S. linnelli; S. galilaeus MM<-> M.myaka S. galilaeus MM<-> M.myaka |
| NT_168003.1 LG6† LG8/LG24† LG11† LG11 LG10/LG21 LG19† LG20 | p2ry14 c3 NA hfe2;txnip vmo1 parp4 NA tprxl.plod3 klhdc8b; cd16311 | coupled purinergic nucleotide receptor activity Myaka + S. linnelli complement activation, inflammatory response - somite development, BMP signaling pathway; enzyme inhibitor activity extracellular exosome ribosyltranferase activity - psuedogene of tetra-peptide repeat homeobox ; motor nueron axon guidance, nueral crest cell migration ubiquitin-protein transferase activity, cytoplasm; receptor mediated endocytosis, scavenger receptor activity | - S. galilaeus MM<>S. linnelli; S. galileaus CR<> M.myaka S. galileaus CR<> S. linnelli; S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galilaeus CR<> S. linnelli S. galilaeus CR<> S. linnelli S. galilaeus CR<> S. linnelli S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka |
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| NT_168003.1 LG6† LG8/LG24† LG11† LG11 LG10† LG19† LG20 LG20† NT_167617.1† NT_168092.1 | p2ry14 c3 NA hfe2;txnip vmo1 parp4 NA tprxl,plod3 klhdc8b; cd16311 ssr1 NA | Proprioticity denominal mascree physiclegy, g protein coupled purinergic nucleotide receptor activity Myaka + S. linnelli complement activation, inflammatory response - somite development, BMP signaling pathway; enzyme inhibitor activity extracellular exosome ribosyltranferase activity - psuedogene of tetra-peptide repeat homeobox ; motor nueron axon guidance, nueral crest cell migration ubiquitin-protein transferase activity, cytoplasm; receptor mediated endocytosis, scavenger receptor activity endoplasmic reticulum membrane; cotranslational protein targeting | - S. galilaeus MM<>S. linnelli; S. galileaus CR<> M.myaka S. galileaus CR<> S. linnelli; S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galilaeus CR<> S. linnelli; S. galilaeus CR<> S. linnelli; S. galilaeus CR<> S. linnelli; S. galilaeus MM<> M.myaka S. galilaeus CR<> S. linnelli; S. galilaeus MM<> M.myaka |
| NT_168003.1 LG6† LG8/LG24† LG11† LG111† LG10/LG21 LG19† LG20 LG20† NT_167617.1† NT_168092.1 | p2ry14 c3 NA hfe2;txnip vmo1 parp4 NA klhdc8b; cd16311 ssr1 NA itgam | <i>Myaka + S. linnelli</i> coupled purinergic nucleotide receptor activity <i>Myaka + S. linnelli</i> complement activation, inflammatory response - somite development, BMP signaling pathway; enzyme inhibitor activity extracellular exosome ribosyltranferase activity - psuedogene of tetra-peptide repeat homeobox ; motor nueron axon guidance, nueral crest cell migration ubiquitin-protein transferase activity, cytoplasm; receptor mediated endocytosis, scavenger receptor activity endoplasmic reticulum membrane; cotranslational protein targeting integrin mediated signalling pathway, cell adhesion | S. galilaeus MM<>S. linnelli; S. galileaus CR<> M.myaka S. galileaus CR<>S. linnelli; S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galileaus CR<>S. linnelli S. galileaus CR<>S. linnelli; S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galileaus CR<>S. linnelli; S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka S. galilaeus MM<> M.myaka |

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*best candidate region for secondary gene flow contributing to diversification † regions with signatures of

738 differential sorting of polymorphism from putative hybrid swarm



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Across 96.2% of the genome Barombi Mbo species (black) are more closely related to each other

than riverine outgroup populations of *S. galilaeus* Mungo and Meme River (green) and *S.*

746 galileaus Cross River (red), or the Lake Ejagham Sarotherodon radiation (blue).





Fig 2. Topologies featuring Barombi Mbo polyphyly with riverine populations involving
the *Stomatepia* three-species complex. Across small and independent proportions of the genome
A-B) the entire *Stomatepia* clade, C) only *S. pindu*, and D) only *S. mariae* are more closely
related to outgroups than other Barombi Mbo species.



Fig 3. Topologies featuring Barombi Mbo polyphyly with riverine populations involving the *Konia* + *Pungu* subclade. Across small and independent proportions of the genome A) only *P. maclareni*, B) only *K. dikume*, and C) the entire *Konia* + *Pungu* subclade are more closely related to outgroups than other Barombi Mbo species.



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Fig 4. Topologies featuring Barombi Mbo polyphyly with riverine populations involving
the *Myaka* + *Sarotherodon* subclade. Across small and independent proportions of the genome
A-B) only *M. myaka*, C) *M. myaka* and two Barombi Mbo *Sarotherdon* species (*S. linelli* and *S. lohbergi*), and D) *M. myaka* and *S. caroli* are more closely related to outgroups than other
Barombi Mbo species.

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| 772 | Figure 5. Manhattan plots of f_4 values between riverine populations of S. galilaeus from |
|-----|---|
| 773 | Mungo and Meme river and S. galileaus from Cross River and A) combinations of the three |
| 774 | species of Stomatepia, B) combinations of the three species in the Konia+Pungu subclade, and |
| 775 | C) M. myaka and a representative from its sister Sarotherodon clade: S. linnelli. Alternating |
| 776 | gray/black colors indicate different linkage groups. Dotted red lines mark the permutation-based |
| 777 | significance thresholds for each test ($P = 0.02$). Peaks highlighted in colors represent those |
| 778 | signals of introgression shared across different subclades. Manhattan plots for the scaffolds not |
| 779 | assigned to the 24 linkage groups are presented in Fig S2-4. |



Figure 6. Candidate introgression region in the *Konia* + *Pungu* subclade of Barombi Mbo region containing genes *pafah1b3* and *hmcn1*. Row 1 shows the peak signal of introgression across scaffold NT_167702.1 detected from the f_4 statistic across the three test combinations involving the three species in the *Konia* + *Pungu* subclade and riverine populations of *S*. *galilaeus* from Mungo, Meme River, and Cross River in non-overlapping 10-kb windows. The

- two genes in this peak are shown in red (*pafah1b3* on the left and *hmcn1* on the right). Row 2
- shows between-population divergence (D_{xy}) among the three combinations of sister species in the
- subclade calculated in non-overlapping 10-kb windows. Row 3 shows within-population
- 789 diversity (π) in the same non-overlapping 10-kb windows.
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- 791
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Best candidate for allopatric phase

Konia



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| 798 | with secondary gene flow respectively. Species with no evidence for differential introgression |
|-----|---|
| 799 | are highlighted with an asterick (*). The true sister species in the Stomatepia are unknown, so all |
| 800 | three potential relationships among the complex are shown. Outgroup riverine S. galilaeus |
| 801 | lineages from Mungo/Meme Rivers (MM) and Cross River (CR) are shown with gray branches. |
| 802 | The percentage of introgression consistent with secondary gene flow into single species and a |
| 803 | hybrid swarm from sliding window f_4 statistic tests is show by above arrows. Proportion of |
| 804 | genome from two riverine sources (red: Mungo/Meme River; blue: Cross River) estimated by |
| 805 | SAGUARO is shown below arrows. Introgression patterns not found in the SAGUARO analysis |
| 806 | are represented with dashes (). Blue arrows with no information that are aligned with red |
| 807 | arrows represent patterns found consistent with hybrid swarm. Note that the separation of arrows |
| 808 | along a branch is for the purpose of clarity and doesn't represent known differences in the timing |
| 809 | of introgression. |
| | |