

1 **Ant nurse workers exhibit behavioral and transcriptomic signatures of specialization on**

2 **larval stage**

3 Justin T. Walsh<sup>1</sup>, Michael R. Warner<sup>1</sup>, Adrian Kase<sup>1</sup>, Benjamin J. Cushing<sup>1</sup>, Timothy A.

4 Linksvayer<sup>1</sup>

5 <sup>1</sup>Department of Biology, University of Pennsylvania, Philadelphia, PA 19104, USA

6 \*corresponding author, email: [juswalsh@sas.upenn.edu](mailto:juswalsh@sas.upenn.edu)

7

## 8 Abstract

9 Division of labor within and between the worker and queen castes is thought to underlie  
10 the tremendous success of social insects. Colonies might benefit if subsets of nurse workers  
11 specialize further in caring for larvae of a certain stage or caste, given that larval nutritional  
12 requirements depend on stage and caste. We used short- (<1 hr) and long-term (ten days)  
13 behavioral observations to determine whether nurses of the pharaoh ant (*Monomorium*  
14 *pharaonis*) exhibit such specialization. We found that nurses were behaviorally specialized based  
15 on larval instar but not on larval caste. This specialization was widespread, with 56% of nurses in  
16 the short-term and between 22-27% in the long-term showing significant specialization.  
17 Additionally, we identified ~200 genes that were differentially expressed in nurse head and  
18 abdominal tissues between nurses feeding young versus old larvae. These included 18 genes  
19 predicted to code for secreted proteins, which may be passed from nurses to larvae via  
20 trophallaxis, as well as vitellogenin and major royal jelly protein-1, which have previously been  
21 implicated in the transfer of nutrition from nurse to larvae and the regulation of larval  
22 development and caste in social insects. Altogether, our results provide the first evidence in any  
23 social insect for a division of labor among nurse workers based on larval stage, and our study  
24 begins to elucidate the molecular mechanisms underlying this specialization.

25

26 Keywords: eusociality, division of labor, brood care, behavioral specialization, transcriptomic

## 27 **Introduction**

28           Division of labor, one of the defining characteristics of eusociality, is believed to be the  
29 primary reason for the tremendous success of social insects (Wilson 1971; Oster and Wilson  
30 1978; Wilson 1987). Within this system of division of labor, queens specialize on reproduction  
31 while workers specialize on tasks including brood care, foraging, and nest defense (Oster and  
32 Wilson 1978; Wilson 1987; Beshers and Fewell 2001). Increased worker efficiency within  
33 colonies is thought to be the main colony-level benefit of division of labor. Behavioral  
34 specialists, through learning or physiological differences, are expected to be more efficient than  
35 generalists (Oster and Wilson 1978; Robinson 1992; Wahl 2002), but see (Dornhaus 2008;  
36 Muscedere et al. 2009). Indeed, social insect behavioral specialists demonstrate increased  
37 efficiency in nest emigration (Langridge et al. 2008), nest excavation (Jeanson et al. 2008),  
38 undertaking (Trumbo and Robinson 1997; Julian and Cahan 1999), and response to sucrose  
39 (Perez et al. 2013).

40           Worker specialization is widespread, and is driven by a diversity of factors and proximate  
41 mechanisms. In many species, worker specialization depends on age, with younger workers  
42 generally performing tasks inside the nest (e.g. brood care) and older workers performing tasks  
43 outside the nest (e.g. foraging) (Oster and Wilson 1978; Robinson 1992; Beshers and Fewell  
44 2001; Mikheyev and Linksvayer 2015). Alternatively, worker tasks can be allocated based on  
45 body size and shape, as many species exhibit morphologically distinct worker sub-castes that  
46 perform different roles within the colony (Oster and Wilson 1978; Beshers and Fewell 2001).  
47 Worker variation in behavioral specialization can also occur independently of age and  
48 morphology (Gordon 1989; Jeanson and Weidenmuller 2014). This interindividual variability  
49 can be the result of genetic diversity among workers (Oldroyd and Fewell 2007), environmental

50 differences during early development (Tautz et al. 2003; Weidenmuller et al. 2009), variation in  
51 adult nutritional state (Blanchard et al. 2000; Ament et al. 2011; Charbonneau et al. 2017), prior  
52 experience (Theraulaz et al. 1998), and the social environment (Webster and Ward 2011).

53 Cooperative brood care, which includes feeding, grooming, and carrying brood, is one of  
54 the most important suites of tasks performed by adult workers (Oster and Wilson 1978; Wilson  
55 1987). Different larvae have different nutritional requirements depending upon their caste and  
56 developmental stage (Cassill and Tschinkel 1996). For example, young larvae of many ant  
57 species are fed exclusively liquid food via nurse-larva trophallaxis while older larvae are also fed  
58 solid protein (Petralia et al. 1980; Tschinkel 1988; Cassill et al. 2005). Furthermore, old larvae  
59 require more frequent and longer feedings than young larvae (Cassill and Tschinkel 1996, 1999).

60 The caste fate of developing larvae in social insects is socially regulated by nurse workers  
61 (Linksvayer et al. 2011; Linksvayer 2015; Vojvodic et al. 2015), often based on the quantity and  
62 quality of nutrition provided to larvae (Wheeler 1986; Hunt and Nalepa 1994; Tribble and  
63 Kronauer 2017). In ants, adult queens tend to have higher fat and protein content relative to  
64 workers, and it is usually assumed that queen-destined larvae are fed different quantities and  
65 qualities of food compared to worker-destined larvae (Hunt and Nalepa 1994; Smith and Suarez  
66 2010; Amor et al. 2016; Warner et al. 2016). Furthermore, recent research in the Florida  
67 carpenter ant (*Camponotus floridanus*) found that nurse workers transfer juvenile hormone,  
68 microRNAs, hydrocarbons, various peptides, and other compounds to larvae during feeding  
69 (LeBoeuf et al. 2016), providing a potential further mechanism for nurses to provide stage- and  
70 caste-specific nutrition to larvae that may regulate larval development.

71 Recent research in honey bees (*Apis mellifera*) suggests that nurse workers exhibit both  
72 behavioral and transcriptomic specialization on larval caste (He et al. 2014; Vojvodic et al.

73 2015). However, these studies did not test for specialization on larval stage and, to the best of our  
74 knowledge, no previous study has investigated the potential for nurse specialization on caste or  
75 larval stage in ants. In this study, we tested whether individual pharaoh ant (*Monomorium*  
76 *pharaonis*) nurse workers exhibit behavioral specialization on different larval stages or castes, as  
77 measured on both short (< 1 hr) and long (10 days) timescales. We estimated how widespread  
78 such specialization is and the contribution of specialists to colony-level brood care. Building on  
79 our behavioral results, we used an existing transcriptomic data set to identify genes with  
80 expression patterns that may be associated with nurse specialization. Overall, we sought to  
81 elucidate whether nurse specialization exists in ants, how it contributes to colony-level brood  
82 care, and what gene expression patterns might be associated with such specialization.

83

## 84 METHODS

### 85 *Background and Overall Design*

86 All colonies used in this study were reared in the lab and were derived from stock  
87 colonies that have been systematically interbred for the past 10 years. We fed the colonies twice  
88 per week with an agar-based synthetic diet (Dussutour and Simpson 2008) and mealworms, and  
89 we maintained all colonies at  $27 \pm 1$  °C, 50% relative humidity, and a 12:12 light:dark cycle. We  
90 conducted all behavioral observations manually using a dissecting microscope and red light. To  
91 keep the temperature constant during behavioral observations, we kept the colonies on a heating  
92 pad set to 27 °C.

93 *M. pharaonis* larvae have three instars (Alvares et al. 1993) that are distinguishable by  
94 body size, body shape, hair abundance, and hair morphology (Berndt and Kremer 1986).

95 Although reproductive-destined larvae (males and gynes) cannot be distinguished from worker-

96 destined larvae as eggs or 1st instar larvae, they can be readily distinguished after the 1st instar  
97 (Berndt and Kremer 1986; Edwards 1991). Since colonies usually only produce new gynes and  
98 males in the absence of fertile queens (Peacock et al. 1955; Edwards 1987), we set up queen-  
99 absent colonies, which rear both worker- and reproductive-destined larvae, when testing for  
100 specialization on larval caste. For both the behavioral observations and transcriptomic analyses,  
101 we initially classified the larvae into five stages based on size and hair morphology: 1st instar,  
102 2nd instar, and small, medium, and large 3rd instar (see Berndt and Kremer 1986; Warner et al.  
103 2016 for details). However, for subsequent behavioral analyses, we only considered larval instar.

#### 104 *Short-term Observations*

105 First we conducted short-term observations of unmarked workers in both queen-present  
106 ( $n = 8$ ) and queen-absent ( $n = 3$ ) colonies to determine whether nurses exhibited short-term  
107 specialization based on larval instar (using queen-present colonies) or larval caste (using queen-  
108 absent colonies). We observed colonies until we saw a worker feed a larva of any instar or caste,  
109 and then we continuously observed that nurse worker for as long as possible (max = 67 minutes).  
110 We recorded each time the nurse fed a larva, as well as the stage and caste of the larva, using the  
111 event logging software “BORIS” (Friard and Gamba 2016). We defined feeding behavior as a  
112 stereotypical behavioral interaction between the nurse worker and larva in which the mouthparts  
113 of the nurse and larva were in contact for at least three seconds. We defined both the transfer of  
114 solid food particles and liquid food via trophallaxis from nurse to larva as feeding behavior and  
115 did not distinguish between these two feeding behaviors. We restricted subsequent analysis to  
116 nurses we observed feeding at least times.

#### 117 *Long-term Observations*

118           Next, we attempted to test whether individually-marked nurses in queenless colonies  
119 express long-term specialization (across ten days). We wanted to track nurses for at least ten  
120 days because this time scale includes the entire amount of time that *M. pharaonis* workers tend  
121 to perform nursing behaviors (Mikheyev and Linksvayer 2015). In each of five colonies, we  
122 collected a cohort of 63 one-day-old callow workers and we uniquely painted each of these focal  
123 individuals with paint dots on their heads and abdomens using combinations of eight colors.  
124 Specifically, we lightly anesthetized them with carbon dioxide and marked their heads and  
125 abdomens with a dot of paint using Sharpie extra-fine point, oil-based paint pens (Dornhaus  
126 2008; Dornhaus et al. 2008; Charbonneau et al. 2017). To control for potential behavioral effects  
127 of the paint, we painted all remaining adult workers in the colonies with black dots on their heads  
128 and abdomens. Because all 63 focal individuals in each colony were age-matched, we were able  
129 to control for possible effects of nurse age on potential behavioral specialization.

130           We constructed queen-absent colonies with 400 workers and 2.5 mL of brood (i.e.  
131 approximately 500 eggs, larvae, and pupae of different stages; Warner et al. 2016, 2018) and  
132 recorded all observed feeding, grooming, or carrying behaviors performed by all focal  
133 individually-marked workers. We initially used queen absent colonies because such colonies  
134 normally raise new queens and we wanted to test for longer-term specialization for caste.  
135 However, given that we observed no short-term specialization for caste, and our colonies ended  
136 up not producing sexual brood, we only considered potential long-term specialization based on  
137 larval stage. We defined feeding as described above, when an individually-marked worker's  
138 mandibles interacted with a larva's mandibles for at least three seconds. We defined grooming as  
139 an interaction between worker mandibles and a larva for a minimum of three seconds. We  
140 defined carrying as a worker lifting a larva with her mandibles and transporting the larva to

141 another location. We analyzed feeding, grooming, and carrying behavior separately. We  
142 observed all colonies for three hours per day for ten consecutive days and restricted subsequent  
143 analysis to individuals we observed feeding, grooming, or carrying at least three times.

#### 144 *Statistical Analysis of Behavioral Specialization*

145 We performed all statistical analyses in R version 3.4.1 (R Core Team 2014). For both  
146 short and long-term observations, we first used binomial generalized linear mixed models  
147 (GLMMs) to ask whether individual nurses differed significantly in their degree of specialization  
148 on larval stage or caste. To test for nurse specialization on larval instar, we grouped 1st and 2nd  
149 instar larvae as “young” larvae and all 3rd instar larvae as “old” larvae. This grouping is  
150 biologically meaningful as 1st and 2nd instar larvae are fed solely a liquid diet while 3rd instar  
151 larvae are also fed solid food (Petralia et al. 1980; Tschinkel 1988; Cassill et al. 2005).  
152 Specifically, we fit GLMMs with the R package lme4 (Bates et al. 2015) for the proportion of  
153 fed larvae that were young versus old, with the identity of the nurse as a random effect and  
154 colony identity and nurse age as fixed effects when appropriate. Similarly, to test for nurse  
155 specialization on larval caste, we fit GLMMs for the proportion of fed larvae that were  
156 reproductive- versus worker-destined larvae. We evaluated the significance of both fixed and  
157 random effects using likelihood ratio (LR) tests. LR tests are appropriate for evaluating the  
158 significance of random effects in binomial models when the models contain fewer than three  
159 random effects (Bolker et al. 2009). A significant random effect of nurse identity in these models  
160 indicates that there is variation among individual nurses for degree of behavioral specialization,  
161 providing initial evidence for behavioral specialization within colonies.

162 Next, given that we found evidence for behavioral specialization (see Results), we used  
163 binomial tests to ask whether each individual significantly specialized on young versus old



164 larvae, or reproductive versus worker larvae, based on recorded observations. We restricted  
165 analysis to nurses with at least six observations because this is the minimum number of  
166 observations that could potentially identify significant ( $P < 0.05$ ) specialization with a binomial  
167 test. We estimated the expected frequency (i.e. “probability of success” in the binomial test) of  
168 interacting with larvae of one stage or caste relative to another stage or caste based on the  
169 observed proportion of interactions for the two stages or castes (e.g., the number of observed  
170 interactions between nurses and 1st instar larvae relative to 3rd instar larvae). In order to first  
171 determine whether any of the individual nurses we observed could be confidently classified as  
172 specialists, we first used binomial tests with a type I error rate corrected for multiple  
173 comparisons across all tested individuals. Given that some individuals were confidently  
174 identified as specialists with these conservative criteria, we next estimated the overall proportion  
175 of specialist versus non-specialist nurses in our study colonies using a type I error rate of 0.05 for  
176 each binomial test run separately for each individual nurse. This test provides an unbiased  
177 approach to determine one-at-a-time whether each individual displayed significant specialization  
178 or not.

### 179 *Gene Expression Analysis*

180 Warner et al. (2017) performed RNA sequencing on a developmental time series of the  
181 five *M. pharaonis* larval stages as well as nurses collected in the act of feeding each of these  
182 larval stages. This previous study focused on identifying caste-biased genes across development  
183 and studying patterns of molecular evolution of these genes. In the current study, we take  
184 advantage of the fact that nurse samples used in Warner et al. (2017) were collected in the act of  
185 feeding one of the five larval stages, and we use the Warner et al. (2017) data set to compare  
186 transcriptomes of nurses feeding different larval stages. We chose to focus on nurses feeding

187 very young versus very old larvae to maximize our power to detect differential expression based  
188 on the stage of larvae fed. Specifically, we used 11 samples of tissues from nurses collected in  
189 the act of feeding 1st instar larvae (5 head samples and 6 abdomen samples) and 10 samples of  
190 nurses collected in the act of feeding large 3rd instar larvae (5 head and 5 abdomen) to identify  
191 genes differentially expressed between nurses feeding larvae at the extreme young and old end of  
192 the developmental trajectory. See supplemental information for a brief summary of the sample  
193 collection procedure; for details of sample collection, RNA extraction, library preparation,  
194 sequencing, and estimation of per-locus expression, see Warner et al. (2017).

195         After removing lowly expressed genes (FPKM < 1 in ½ the samples), we used the  
196 package EdgeR (Robinson et al. 2010) for differential expression analysis. We constructed a  
197 GLM-like model, including larval stage fed, replicate, and queen presence as additive effects to  
198 identify genes differentially expressed between nurses feeding young versus old larvae (1st instar  
199 versus large 3rd instar; separately for head and abdomens). We calculated gene ontology (GO)  
200 term enrichment of differentially expressed genes using the R package GOSTats, with a cut-off P-  
201 value of 0.05 (Falcon and Gentleman 2007).

202         To test whether genes found to be differentially expressed between nurses tended to code  
203 for secreted proteins in *Drosophila melanogaster*, we compiled a list of genes annotated as  
204 coding for secreted proteins according to the online tool GLAD (Hu et al. 2015). From this list,  
205 we identified secreted proteins with orthologs in *M. pharaonis* using a recently created orthology  
206 map between *M. pharaonis*, *Apis mellifera*, and *D. melanogaster* (Warner et al. unpublished  
207 manuscript; orthology map included as Supplementary Data). We estimated the association  
208 between a gene's likelihood to be differentially expressed and secreted, removing all genes for

209 which a *D. melanogaster* ortholog was not detected. We generated plots using the R package  
210 ggplot2 (Wickham 2009).

211

## 212 RESULTS

### 213 *Short-term Specialization on Larval Stage*

214 We observed 52 nurses feed at least three times (mean = 8.8 feeding events) and we  
215 included these nurses in the GLMMs. The random effect of nurse identity was significant,  
216 suggesting that nurses tended to specialize on feeding either young or old larvae (table 1). Next,  
217 to classify each individual nurse as showing significant specialization or not (i.e. to classify  
218 nurses as specialists or non-specialists), we used binomial tests with an expected proportion of  
219 old larvae relative to young plus old larvae of 0.781 (the observed proportion of old larvae fed  
220 across all individuals in long-term feeding observations). We used the observed proportion from  
221 long-term observations, as opposed to short-term observations, because we specifically  
222 attempted to balance the number of recorded old and young larvae feeding events (in terms of  
223 total number of observations, not per individual) during short-term observations. Therefore the  
224 observed short-term proportions are not an accurate representation of the naturally-occurring  
225 proportions. We included the 32 nurses we observed feed at least six times. When using a type I  
226 error rate corrected for multiple comparisons, which should produce a conservative estimate of  
227 the frequency of specialists across the whole study, we classified about 56% (18/32) of nurses as  
228 specialists (Bonferroni-adjusted P). When using a type I error rate of 0.05, which should yield an  
229 unbiased estimate of the frequency of specialists versus generalists within colonies, we again  
230 classified about 56% (18/32) of nurses as specialists. These specialists performed about 65%  
231 (242/375) of the observed feedings.



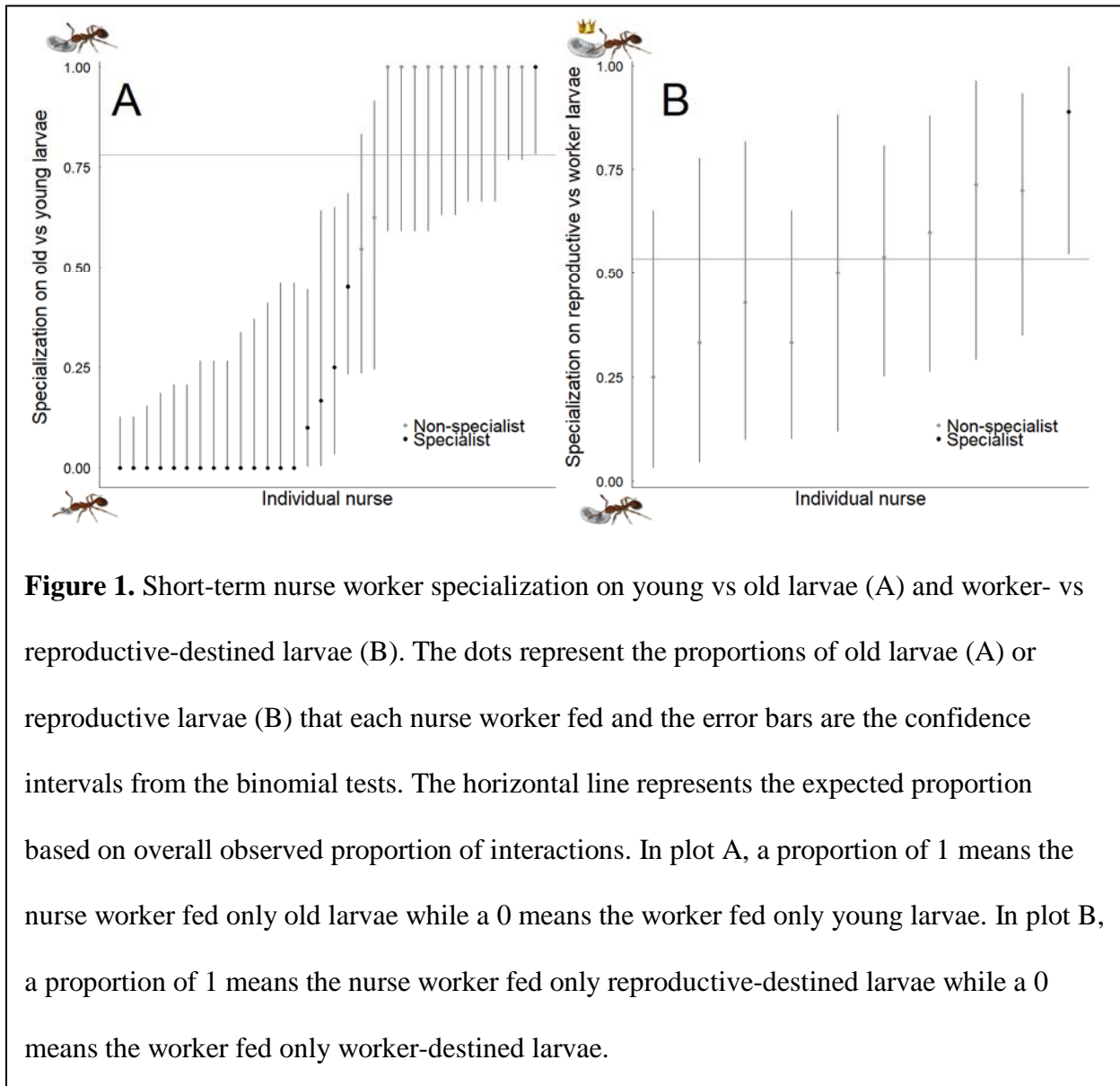
**Table 1.** Summary of effects of factors on short- and long-term nurse behavior on likelihood ratio tests of GLMMs

	$\chi^2$	df	p
<b>Short-term Feeding</b>			
Caste			
Individual Nurse	1.430	1	0.232
Stage			
Individual Nurse	345.56	1	<0.0001
<b>Long-term Feeding</b>			
Individual Nurse	36.934	1	<0.0001
Colony	57.750	4	<0.0001
Age	0.018	1	0.892
<b>Long-term Grooming</b>			
Individual Nurse	21.357	1	<0.0001
Colony	49.576	1	<0.0001
Age	0.022	1	0.882
<b>Long-term Carrying</b>			
Individual Nurse	4.500	1	0.034
Colony	5.048	1	0.025
Age	0.021	1	0.886

235 *Short-term Specialization on Caste*

236           We observed 22 nurses feed at least three times (mean = 5.64 feeding events). The  
237 random effect of nurse identity in the GLMM was not significant (table 1) indicating that nurses  
238 did not specialize on larval caste. In the binomial tests, we included the ten nurses we observed  
239 feed at least six times and used an expected proportion of reproductive-destined larvae of 0.534.  
240 When correcting for multiple comparisons, we classified zero nurses as specialists. When using a  
241 type I error rate of 0.05, we classified 10% (1/10) of nurses as specialists and this specialist  
242 performed about 6% (9/142) of the observed feedings (figure 1).

243



244

245

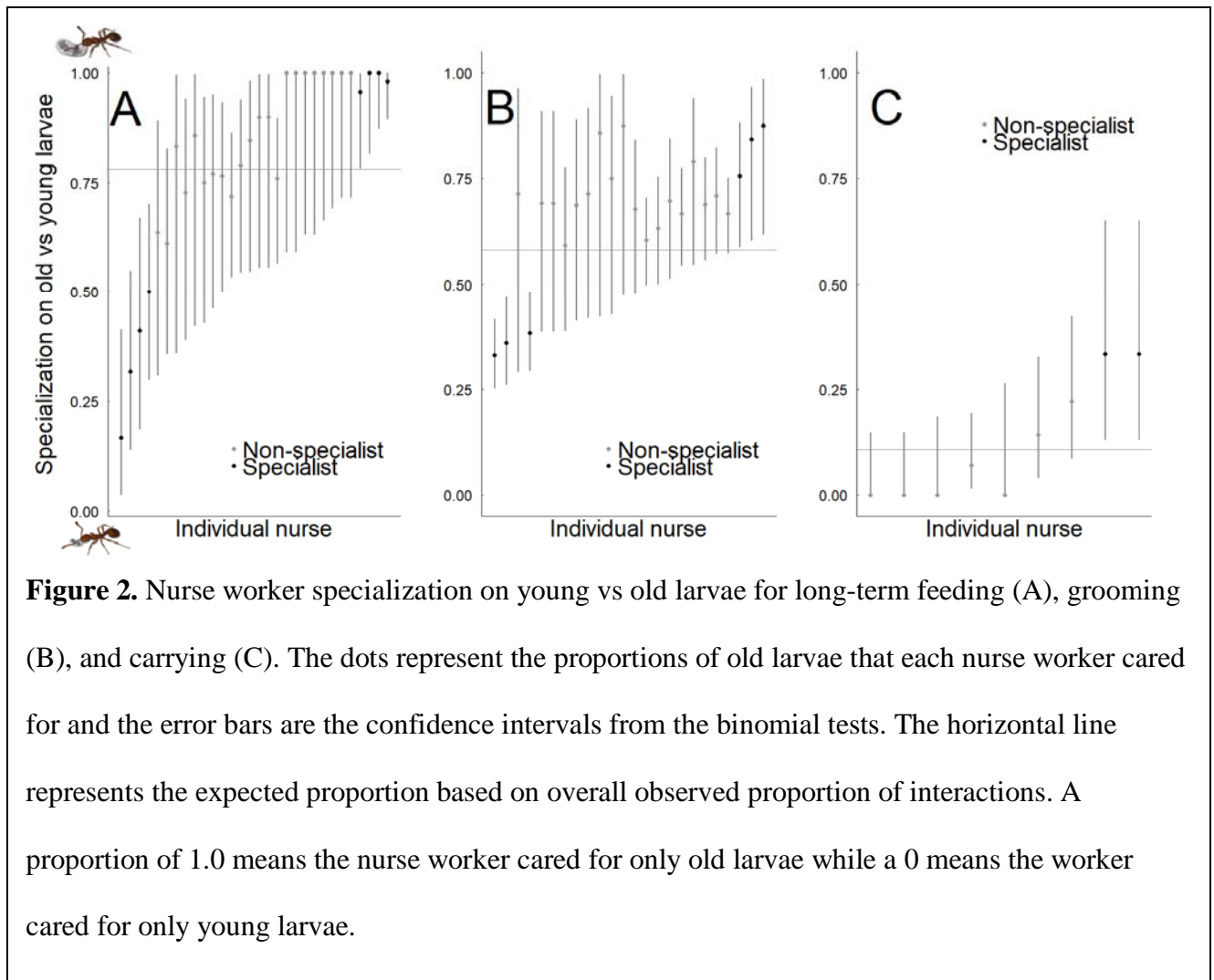
246 *Long-term Feeding Specialization on Larval Stage*

247           We observed 40 nurses feed at least three times (mean = 12.9 feeding events). The effects  
248 of nurse identity and colony identity were significant (table 1), indicating that nurses tended to  
249 specialize on feeding either young or old larvae. The age of the nurse was not significant. In the  
250 binomial tests, we included the 30 nurses we observed feed at least six times and used an  
251 expected proportion of old larvae of 0.781. When correcting for multiple comparisons, we  
252 classified 20% (6/30) of nurses as being long-term specialists on larval stage. When using an  
253 uncorrected type I error rate of 0.05, we classified about 27% (8/30) of nurses as long-term  
254 specialists and these long-term specialists performed about 42% (201/480) of the observed  
255 feedings (figure 2). Long-term specialists performed significantly more feedings than non-  
256 specialists (Mann-Whitney-Wilcoxon test;  $W= 19.5$ ,  $P = 0.0013$ ).

257



258



259

## 260 *Long-term Grooming Specialization on Larval Stage*

261 We observed 32 individuals grooming larvae at least three times (mean = 33.9 grooming  
262 events). The effects of nurse identity and colony identity were significant (table 1), indicating  
263 that nurses tended to specialize on feeding either young or old larvae. The age of the nurse was  
264 not significant. In the binomial tests, we included the 24 nurses we observed groom at least six  
265 times and used an expected proportion of old larvae of 0.581. When correcting for multiple  
266 comparisons, we classified about 13% (3/24) of nurses as specialists. When using an uncorrected  
267 type I error rate of 0.05, we classified 25% (6/24) of nurses as specialists and these specialists  
268 performed about 39% (406/1053) of the observed groomings (figure 2). The number of  
269 groomings performed by specialists and non-specialists was not significantly different ( $W = 29$ ,  
270  $P = 0.1021$ ).

## 271 *Long-term Carrying Specialization on Larval Stage*

272 We observed 17 individuals carrying a larva at least three times (mean = 13.4 carrying  
273 observations). The effects of nurse identity and colony identity were significant (table 1),  
274 indicating that nurses tended to specialize on feeding either young or old larvae. The age of nurse  
275 was not significant. In the binomial tests, we included the nine nurses we observed carrying a  
276 larva at least six times and used an expected ratio of old to young of 0.107. When correcting for  
277 multiple comparisons, we classified zero nurses as specialists. When using an uncorrected type I  
278 error rate of 0.05, we classified about 22% (2/9) of nurses as specialists and these specialists  
279 performed about 12% (24/197) of the carrying observations (figure 2). The number of groomings  
280 performed by specialists and non-specialists was not significantly different ( $W = 13$ ,  $P = 0.100$ ).

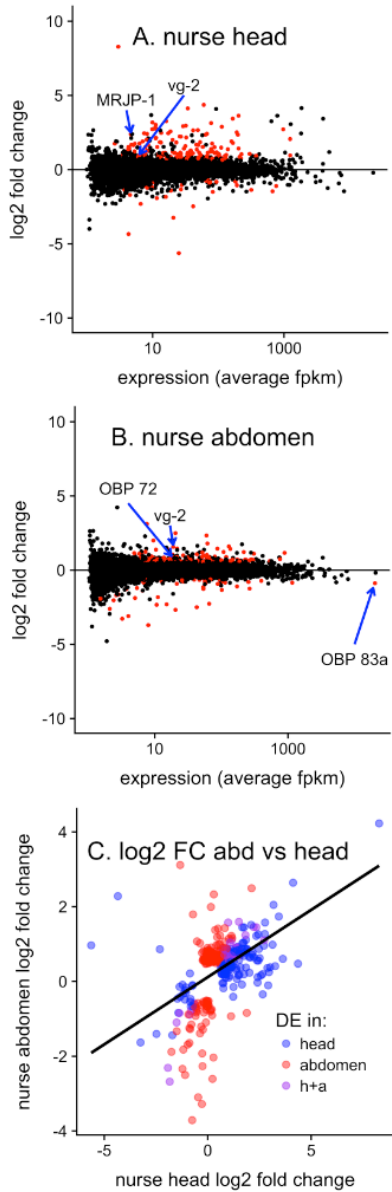
## 281 *Transcriptomic Analysis*

282 We identified 209 and 173 differentially expressed genes (DEGs) in the heads and  
283 abdomens, respectively, of nurses collected while feeding young (i.e. 1st instar) versus old (i.e.  
284 large 3rd instar) worker larvae (FDR < 0.05; figure 3a,b). In both nurse heads and abdomens, we  
285 identified more up-regulated genes in nurses feeding young versus old larvae (two-sided  
286 binomial, null hypothesis of 50% upregulated in nurses feeding young larvae; heads: N = 209, P  
287 < 0.001; abdomens: N = 173, P < 0.001). There was a positive association between genes up-  
288 regulated in the heads and genes upregulated in the abdomens of nurses feeding young larvae ( $\chi^2$   
289 = 312, df = 1, P < 0.001), as well as between genes up-regulated in the heads and abdomens of  
290 nurses feeding old larvae ( $\chi^2$  = 260, df = 1, P < 0.001). Additionally, there was an overall  
291 correlation between expression fold change in nurse heads and abdomens across all differentially  
292 expressed genes between nurses feeding young versus old larvae (figure 3c). For genes  
293 associated with each nurse type, gene ontology was largely dominated by metabolism-related  
294 categories (table A1). Genes up-regulated in the heads of nurses feeding young larvae were also  
295 associated with isoprenoid (a type of hydrocarbon) processing, and genes up-regulated in the  
296 abdomens of nurses feeding young larvae were associated with transport and localization.

297 Genes that were differentially expressed in nurses based on larval stage were more likely  
298 to code for proteins known to be cellularly secreted in *Drosophila melanogaster* ( $\chi^2$  = 29.1, df =  
299 1, P < 0.001; 18 DGEs coding for secreted proteins out of 148 total DGEs with orthologs in *D.*  
300 *melanogaster*. 178 genes had orthologs that code for secreted proteins in *D. melanogaster*, out of  
301 5391 genes in the analysis). Nearly all of the DEGs that are predicted to code for secreted  
302 proteins were upregulated in nurses feeding young larvae (14/14 in heads, 9/10 in abdomens, see  
303 table A2 for complete list of DEGs based on larval stage fed).

304

305



**Figure 3.** Differential expression between nurses feeding young (1st-instar) and old (large 3rd instar) larvae in A) nurse heads and B) nurse abdomens. Genes colored red are differentially expressed (FDR < 0.05). C) Correlation of log<sub>2</sub> fold change of differentially expressed genes as measured in nurse abdomens and heads (Spearman rho = 0.345, P < 0.001). Black line represents trendline of linear model. Genes are colored by tissue differentially expressed in (FDR < 0.05). In all plots, genes with positive “log<sub>2</sub> fold change” are upregulated in nurses feeding large 1st vs 3rd instar larvae (i.e feeding young vs old larvae).

## 306 DISCUSSION

307           The tremendous ecological success of social insects is thought to be primarily due to  
308 efficient division of labor within colonies (Wilson 1971; Oster and Wilson 1978; Wilson 1987).  
309 Here we provide to the best of our knowledge the first evidence for the existence of a division of  
310 labor within nurse workers based on the instar of larvae they care for. We found evidence for  
311 behavioral specialization in the short-term (less than an hour) and the long-term (over 10 days).  
312 Nurses specialized on either old (3rd instar) or young (1st and 2nd instar) larvae, and this  
313 specialization was consistent across feeding, grooming, and carrying behaviors. In the short-  
314 term, we classified 56% of nurses as specialists in terms of feeding, and in the long-term, we  
315 classified 27%, 25%, and 22% of workers as specialists in feeding, grooming, and carrying  
316 respectively.

317           Specialists are predicted to increase colony efficiency (Oster and Wilson 1978; Robinson  
318 1992; Wahl 2002). Although we cannot say whether specialist nurses increase *M. pharaonis*  
319 colony efficiency, our data suggest that specialists do play an important role in the colony as they  
320 performed more per capita feedings than non-specialists. The specialization of nurse workers on  
321 old or young larvae might be explained by specialization on trophallaxis (i.e. feeding liquids)  
322 versus feeding solid food particles since young larvae are fed only a liquid diet while old larvae  
323 are also fed solid protein (Petralia et al. 1980; Tschinkel 1988; Cassill et al. 2005). If so, the  
324 nurses specialized for trophallaxis may play a disproportionately large role in regulating larval  
325 development since trophallactic fluid contains not only nutrition but also juvenile hormone,  
326 microRNAs, hydrocarbons, various peptides, and other compounds (LeBoeuf et al. 2016).

327           Given that we found evidence for the behavioral specialization of nurses on young versus  
328 old larvae, we also tested for differential gene expression in the head and abdominal tissues of

329 nurses feeding young versus old larvae as a first step in identifying transcriptomic signatures of  
330 specialization. We expect that differentially expressed genes (DEGs) might be functionally  
331 associated with different types of care provided by nurse workers to differently-aged larvae,  
332 which might actively contribute to the social regulation of larval development (see Vojvodic et  
333 al. 2015). We identified 209 and 173 differentially expressed genes in nurse heads and  
334 abdomens, respectively, between nurses feeding young versus old worker-destined larvae. Based  
335 on our behavioral analyses, we expect approximately one half of the individuals used in our gene  
336 expression samples to be specialized based on larval age, rendering our differential expression  
337 analysis effectively conservative (i.e. non-specialists included in our sample would weaken the  
338 transcriptomic signature of specialists). Interestingly, while the majority of DEGs were tissue  
339 specific, a significant proportion of genes were differentially expressed in the same direction in  
340 heads and abdomens, and there was an overall correlation between log fold expression change  
341 from young to old nurses in both heads and abdomens. Together, these results indicate that some  
342 transcriptomic changes associated with nurse specialization occur throughout nurse bodies.

343 Intriguingly, genes with *D. melanogaster* orthologs that are known to code for cellularly-  
344 secreted proteins were overrepresented among the DEGs between *M. pharaonis* nurses feeding  
345 young versus old larvae. The DEGs we detected in nurse tissues could directly affect larval  
346 development if the proteins were secreted by nurses and transferred to larvae via trophallaxis  
347 (Linksvayer 2015). Many of these DEGs which are predicted to code for secreted proteins have  
348 metabolic functions, suggesting they may play a role in the the breakdown of food before it is  
349 passed to larvae.

350 The DEGs between nurses may also play a role in responding to larval signals. Two  
351 odorant binding proteins (OBP) were differentially expressed in nurse abdomens (figure 3a).

352 These OBPs potentially play a role in communication between nurses and larvae (Zhou et al.  
353 2015, McKenzie et al. 2016). Although OBPs are predicted to be primarily expressed in the  
354 antennae, previous studies found that OBPs are frequently expressed in non-chemosensory  
355 tissues (McKenzie et al. 2014) and can exhibit various functions beyond olfaction (Nomura et al.  
356 1992, Maleszka et al. 2007, Dani et al. 2011, Zhang et al. 2016). For example, the Gp-9 gene  
357 encodes for the odorant binding protein SiOBP3 and has been linked to colony organization in  
358 the fire ant *Solenopsis invicta* (Wang et al. 2013). Expression of SiOBP3 is found throughout the  
359 bodies of workers, gynes, and males and is actually lowest in the antennae (Zhang et al. 2016).

360 Interestingly, in both measured tissues (heads and abdomens), we identified more genes  
361 upregulated in nurses feeding 1st-instar larvae than those feeding 3rd-instar larvae, and all DEGs  
362 that code for proteins secreted in *D. melanogaster* were upregulated in nurses feeding 1st-instar  
363 larvae. These genes might be involved in regulating early larval development, or perhaps  
364 regulation of larval caste fate, given that caste determination occurs at least by the end of the 1st  
365 instar (Berndt and Kremer 1986; Alvares et al. 1993; Khila et al. 2010; Warner et al. 2016;  
366 Warner et al. 2018). Genes upregulated in nurses feeding 1st-instar larvae included genes such as  
367 vitellogenin (Vg2) (Libbrecht et al. 2013) and a member of the major royal jelly protein family  
368 (MRJP-1) (Schonleben et al. 2007), both of which have been implicated in the production and  
369 transfer of proteinaceous food to honey bee larvae, which then shapes larval development and  
370 caste fate (Amdam et al. 2003) (figure 3a). Interestingly, LeBoeuf et al (2016) found both a  
371 MRJP homolog and vitellogenin in the trophallactic fluid of ant nurses fed to developing larvae.  
372 Therefore, it is possible that *M. pharaonis* nurses feeding young larvae are passing on these  
373 compounds directly to larvae as a means to regulate larval development.

374 Contrary to findings in honey bees (He et al. 2014; Vojvodic et al. 2015), we found no  
375 behavioral evidence for nurse specialization on larval caste. This lack of specialization in *M.*  
376 *pharaonis* is somewhat surprising, given that worker- and reproductive-destined larvae likely  
377 have different nutritional needs (Hunt and Nalepa 1994; Smith and Suarez 2010; Amor et al.  
378 2016; Warner et al. 2016). However, this difference may be attributable to differences in timing  
379 of caste determination. In honey bees, caste determination occurs relatively late in development  
380 and over a period of time, as queen-worker inter-castes can be produced by experimental  
381 manipulation of diet late in development (Linksvayer et al. 2011; Dedej et al. 1998; Wang et al.  
382 2014). Therefore, in honey bees, continued nurse-larval interactions are likely essential to fine-  
383 tune caste dimorphism (Linksvayer et al. 2011).

384 In contrast to honey bees where each larva develops in an isolated brood cell, worker-  
385 and reproductive-destined larvae are not spatially separated in *M. pharaonis*. This lack of  
386 separation could also help explain the lack of specialization on larval caste in *M. pharaonis*  
387 compared to honey bees. Additionally, many ants (including *M. pharaonis*) spatially arrange  
388 their brood such that younger larvae and eggs tend be in the center and older larvae and pupae  
389 towards the edge of brood piles (Franks & Sendova-Franks 1992; Lim & Lee 2005). This spatial  
390 arrangement could potentially contribute to the observed short-term specialization if nurses  
391 spend most of their time in one area of the nest and feed larvae close to them. However, we  
392 observed nurses frequently moving around the nest during our short-term observations,  
393 interacting with other workers or collecting food in between subsequent feedings, so that each  
394 individual nurse had the potential to interact with all brood stages.

395 Further research is necessary to characterize the implications of nurse specialization,  
396 elucidate the detailed molecular and physiological underpinnings, and to determine how



397 widespread specialization is across ants and other social insects. Interestingly, we found  
398 significant effects of the colony identity for long-term nursing, grooming, and carrying.  
399 Although outside the scope of this study, it is possible that different colonies exhibit different  
400 levels of specialization in either the number of specialists or the proportion of brood care  
401 behaviors performed by specialists. Future studies should test for colony-level variation in nurse  
402 specialists.

### 403 *Conclusion*

404 This study describes a previously undocumented form of division of labor within ant  
405 nurse workers: specialization based on larval instar. We found evidence for this specialization in  
406 three different brood care behaviors. Additionally, we found ~200 differentially expressed genes  
407 between nurses feeding young versus old larvae. Contrary to findings in honey bees, we found  
408 no evidence for specialization of nurse workers on larval caste. Further research is necessary to  
409 characterize the implications of nurse specialization, elucidate the detailed molecular and  
410 physiological underpinnings, and to determine how widespread specialization is across ants and  
411 other social insects.

### 412 *Data Accessibility*

413 Data supporting this paper are included as supplemental files.

414

### 415 REFERENCES

416 Alvares, L. E., Bueno, O. C., & Fowler, H. G. (1993). Larval instars and immature development  
417 of a Brazilian population of pharaohs ant, *Monomorium pharaonis*. [Article]. *Journal of*  
418 *Applied Entomology-Zeitschrift Fur Angewandte Entomologie*, 116(1), 90-93.

- 419 Amdam, G. V., Norberg, K., Hagen, A., & Omholt, S. W. (2003). Social exploitation of  
420 vitellogenin. [Article]. *Proceedings of the National Academy of Sciences of the United*  
421 *States of America*, 100(4), 1799-1802. doi: 10.1073/pnas.0333979100
- 422 Ament, S. A., Chan, Q. W., Wheeler, M. M., Nixon, S. E., Johnson, S. P., Rodriguez-Zas, S. L., .  
423 . . Robinson, G. E. (2011). Mechanisms of stable lipid loss in a social insect. [Article].  
424 *Journal of Experimental Biology*, 214(22), 3808-3821. doi: 10.1242/jeb.060244
- 425 Amor, F., Villalta, I., Doums, C., Angulo, E., Caut, S., Castro, S., . . . Boulay, R. (2016).  
426 Nutritional versus genetic correlates of caste differentiation in a desert ant. [Article].  
427 *Ecological Entomology*, 41(6), 660-667. doi: 10.1111/een.12337
- 428 Bates, D., Machler, M., Bolker, B. M., & Walker, S. C. (2015). Fitting Linear Mixed-Effects  
429 Models Using lme4. [Article]. *Journal of Statistical Software*, 67(1), 1-48.
- 430 Berndt, K. P., & Kremer, G. (1986). Larvenmorphologie der Pharoameise *Monomorium*  
431 *pharaonis*. *Zoologischer Anzeiger*.
- 432 Beshers, S. N., & Fewell, J. H. (2001). Models of division of labor in social insects. [Review].  
433 *Annual Review of Entomology*, 46, 413-440. doi: 10.1146/annurev.ento.46.1.413
- 434 Blanchard, G. B., Orledge, G. M., Reynolds, S. E., & Franks, N. R. (2000). Division of labour  
435 and seasonality in the ant *Leptothorax albipennis*: worker corpulence and its influence on  
436 behaviour. [Article]. *Animal Behaviour*, 59, 723-738. doi: 10.1006/anbe.1999.1374
- 437 Bolker, B. M., Brooks, M. E., Clark, C. J., Geange, S. W., Poulsen, J. R., Stevens, M. H. H., &  
438 White, J. S. S. (2009). Generalized linear mixed models: a practical guide for ecology  
439 and evolution. [Review]. *Trends in Ecology & Evolution*, 24(3), 127-135. doi:  
440 10.1016/j.tree.2008.10.008

- 441 Cassill, D. L., Butler, J., Vinson, S. B., & Wheeler, D. E. (2005). Cooperation during prey  
442 digestion between workers and larvae in the ant, *Pheidole spadonia*. [Article]. *Insectes*  
443 *Sociaux*, 52(4), 339-343. doi: 10.1007/s00040-005-0817-x
- 444 Cassill, D. L., & Tschinkel, W. R. (1996). A duration constant for worker-to-larva trophallaxis in  
445 fire ants. [Article]. *Insectes Sociaux*, 43(2), 149-166. doi: 10.1007/bf01242567
- 446 Cassill, D. L., & Tschinkel, W. R. (1999). Regulation of diet in the fire ant, *Solenopsis invicta*.  
447 [Article]. *Journal of Insect Behavior*, 12(3), 307-328.
- 448 Charbonneau, D., Poff, C., Nguyen, H., Shin, M. C., Kierstead, K., & Dornhaus, A. (2017). Who  
449 are the "lazy" ants? The function of inactivity in social insects and a possible role of  
450 constraint: inactive ants are corpulent and may be young and/or selfish. *Integrative and*  
451 *Comparative Biology*, 57(3), 649-667.
- 452 Dani, F. R., Michelucci, E., Francese, S., Mastrobuoni, G., Cappellozza, S., La Marca, G., . . .  
453 Pelosi, P. (2011). Odorant-Binding Proteins and Chemosensory Proteins in Pheromone  
454 Detection and Release in the Silkworm *Bombyx mori*. [Article]. *Chemical Senses*, 36(4),  
455 335-344. doi: 10.1093/chemse/bjq137
- 456 Dedej, S., Hartfelder, K., Aumeier, P., Rosenkranz, P., & Engels, W. (1998). Caste determination  
457 is a sequential process: effect of larval age at grafting on ovariole number, hind leg size  
458 and cephalic volatiles in the honey bee (*Apis mellifera carnica*). [Article]. *Journal of*  
459 *Apicultural Research*, 37(3), 183-190.
- 460 Dornhaus, A. (2008). Specialization does not predict individual efficiency in an ant. *PLoS*  
461 *Biology*, 6(11).
- 462 Dornhaus, A., Holley, J. A., Pook, V. G., Worswick, G., & Franks, N. R. (2008). Why do not all  
463 workers work? Colony size and workload during emigrations in the ant *Temnothorax*

- 464 *albipennis*. [Article]. *Behavioral Ecology and Sociobiology*, 63(1), 43-51. doi:  
465 10.1007/s00265-008-0634-0
- 466 Dussutour, A., & Simpson, S. J. (2008). Description of a simple synthetic diet for studying  
467 nutritional responses in ants. [Article]. *Insectes Sociaux*, 55(3), 329-333. doi:  
468 10.1007/s00040-008-1008-3
- 469 Edwards, J. P. (1987). Caste regulatin in the pharoahs ant *Monomorium pharaonis*- The  
470 influence of queens on the production of new sexual forms. [Article]. *Physiological*  
471 *Entomology*, 12(1), 31-39. doi: 10.1111/j.1365-3032.1987.tb00721.x
- 472 Edwards, J. P. (1991). Caste regulation in the pharaohs ant *Monomorium pharaonis*- recognition  
473 and cannibalism of sexual brood by workers. *Physiological Entomology*, 16(3), 263-271.
- 474 Falcon, S., & Gentleman, R. (2007). Using GOSTats to test gene lists for GO term association.  
475 [Article]. *Bioinformatics*, 23(2), 257-258. doi: 10.1093/bioinformatics/btl567
- 476 Franks, N. R., & Sendova-Franks, A. B. (1992). Brood sorting by ants- distributing the workload  
477 over the work-surface. [Article]. *Behavioral Ecology and Sociobiology*, 30(2), 109-123.
- 478 Friard, O., & Gamba, M. (2016). BORIS: a free, versatile open-source event-logging software  
479 for video/audio coding and live observations. [Article]. *Methods in Ecology and*  
480 *Evolution*, 7(11), 1325-1330. doi: 10.1111/2041-210x.12584
- 481 Gordon, D. M. (1989). Dynamics of task switching in harvester ants. [Article]. *Animal*  
482 *Behaviour*, 38, 194-204. doi: 10.1016/s0003-3472(89)80082-x
- 483 He, X. J., Tian, L. Q., Barron, A. B., Guan, C., Liu, H., Wu, X. B., & Zeng, Z. J. (2014).  
484 Behavior and molecular physiology of nurses of worker and queen larvae in honey bees  
485 (*Apis mellifera*). [Article]. *Journal of Asia-Pacific Entomology*, 17(4), 911-916. doi:  
486 10.1016/j.aspen.2014.10.006

- 487 Hu, Y., Comjean, A., Perkins, L. A., Perrimon, N., & Mohr, S. E. (2015). GLAD: an Online  
488 Database of Gene List Annotation for *Drosophila*. *J Genomics*, 3, 75-81. doi:  
489 10.7150/jgen.12863
- 490 Hunt, J. H., & Nalepa, C. A. (1994). *Nourishment, evolution, and insect sociality*. Westview,  
491 Boulder: Westview, Boulder.
- 492 Jeanson, R., Clark, R. M., Holbrook, C. T., Bertram, S. M., Fewell, J. H., & Kukuk, P. F. (2008).  
493 Division of labour and socially induced changes in response thresholds in associations of  
494 solitary halictine bees. [Article]. *Animal Behaviour*, 76, 593-602. doi:  
495 10.1016/j.anbehav.2008.04.007
- 496 Jeanson, R., & Weidenmuller, A. (2014). Interindividual variability in social insects - proximate  
497 causes and ultimate consequences. [Review]. *Biological Reviews*, 89(3), 671-687. doi:  
498 10.1111/brv.12074
- 499 Julian, G. E., & Cahan, S. (1999). Undertaking specialization in the desert leaf-cutter ant  
500 *Acromyrmex versicolor*. [Article]. *Animal Behaviour*, 58, 437-442. doi:  
501 10.1006/anbe.1999.1184
- 502 Khila, A., & Abouheif, E. (2010). Evaluating the role of reproductive constraints in ant social  
503 evolution. [Article]. *Philosophical Transactions of the Royal Society B-Biological*  
504 *Sciences*, 365(1540), 617-630. doi: 10.1098/rstb.2009.0257
- 505 Langridge, E. A., Sendova-Franks, A. B., & Franks, N. R. (2008). How experienced individuals  
506 contribute to an improvement in collective performance in ants. [Article]. *Behavioral*  
507 *Ecology and Sociobiology*, 62(3), 447-456. doi: 10.1007/s00265-007-0472-5

- 508 LeBoeuf, A. C., Waridel, P., Brent, C. S., Goncalves, A. N., Menin, L., Ortiz, D., . . . Keller, L.  
509 (2016). Oral transfer of chemical cues, growth proteins and hormones in social insects.  
510 [Article]. *Elife*, 5, 27. doi: 10.7554/eLife.20375
- 511 Libbrecht, R., Corona, M., Wende, F., Azevedo, D. O., Serrao, J. E., & Keller, L. (2013).  
512 Interplay between insulin signaling, juvenile hormone, and vitellogenin regulates  
513 maternal effects on polyphenism in ants. [Article]. *Proceedings of the National Academy  
514 of Sciences of the United States of America*, 110(27), 11050-11055. doi:  
515 10.1073/pnas.1221781110
- 516 Lim, S. P., & Lee, C. Y. (2005). Brood arrangement and food distribution among larvae under  
517 different colony conditions in the Pharaoh's ant, *Monomorium pharaonis* (Hymenoptera :  
518 Formicidae). [Article]. *Sociobiology*, 46(3), 491-503.
- 519 Linksvayer, T. A. (2015). The Molecular and Evolutionary Genetic Implications of Being Truly  
520 Social for the Social Insects. In A. Zayed & C. F. Kent (Eds.), *Genomics, Physiology and  
521 Behaviour of Social Insects* (Vol. 48, pp. 271-292). London: Academic Press Ltd-  
522 Elsevier Science Ltd.
- 523 Linksvayer, T. A., Kaftanoglu, O., Akyol, E., Blatch, S., Amdam, G. V., & Page, R. E. (2011).  
524 Larval and nurse worker control of developmental plasticity and the evolution of honey  
525 bee queen-worker dimorphism. [Article]. *Journal of Evolutionary Biology*, 24(9), 1939-  
526 1948. doi: 10.1111/j.1420-9101.2011.02331.x
- 527 Maleszka, J., Foret, S., Saint, R., & Maleszka, R. (2007). RNAi-induced phenotypes suggest a  
528 novel role for a chemosensory protein CSP5 in the development of embryonic integument  
529 in the honeybee (*Apis mellifera*). [Article]. *Development Genes and Evolution*, 217(3),  
530 189-196. doi: 10.1007/s00427-006-0127-y

- 531 McKenzie, S. K., Fetter-Pruneda, I., Ruta, V., & Kronauer, D. J. C. (2016). Transcriptomics and  
532 neuroanatomy of the clonal raider ant implicate an expanded clade of odorant receptors in  
533 chemical communication. [Article]. *Proceedings of the National Academy of Sciences of*  
534 *the United States of America*, 113(49), 14091-14096. doi: 10.1073/pnas.1610800113
- 535 McKenzie, S. K., Oxley, P. R., & Kronauer, D. J. C. (2014). Comparative genomics and  
536 transcriptomics in ants provide new insights into the evolution and function of odorant  
537 binding and chemosensory proteins. [Article]. *Bmc Genomics*, 15, 14. doi: 10.1186/1471-  
538 2164-15-718
- 539 Mikheyev, A. S., & Linksvayer, T. A. (2015). Genes associated with ant social behavior show  
540 distinct transcriptional and evolutionary patterns. [Article]. *Elife*, 4, 29. doi:  
541 10.7554/eLife.04775
- 542 Muscedere, M. L., Willey, T. A., & Traniello, J. F. A. (2009). Age and task efficiency in the ant  
543 *Pheidole dentata*: young minor workers are not specialist nurses. *Animal Behaviour*, 77,  
544 911-918.
- 545 Nomura, A., Kawasaki, K., Kubo, T., & Natori, S. (1992). Purification and localization of P10, a  
546 novel protein that increases in nymphal regenerating legs of *Periplaneta americana*  
547 (American cockroach). [Article]. *International Journal of Developmental Biology*, 36(3),  
548 391-398.
- 549 Oldroyd, B. P., & Fewell, J. H. (2007). Genetic diversity promotes homeostasis in insect  
550 colonies. [Review]. *Trends in Ecology & Evolution*, 22(8), 408-413. doi:  
551 10.1016/j.tree.2007.06.001
- 552 Oster, G. F., & Wilson, E. O. (1978). *Caste and ecology in the social insects*. Princeton, NJ:  
553 Princeton University Press.

- 554 Peacock, A. D., Sudd, J. H., & Baxter, A. T. (1955). Studies in pharaohs ant, *Monomorium*  
555 *pharaonis*. *Entomologist's Monthly Magazine*, 91, 130-133.
- 556 Perez, M., Rolland, U., Giurfa, M., & d'Ettorre, P. (2013). Sucrose responsiveness, learning  
557 success, and task specialization in ants. [Article]. *Learning & Memory*, 20(8), 417-420.  
558 doi: 10.1101/lm.031427.113
- 559 Petralia, R. S., Sorensen, A. A., & Vinson, S. B. (1980). Labial gland system of the larvae of the  
560 imported fire ant, *Solenopsis invicta* Buren- Ultrastructure and enzyme analysis [Article].  
561 *Cell and Tissue Research*, 206(1), 145-156.
- 562 Robinson, G. E. (1992). Regulation of division-of-labor in insect societies. [Review]. *Annual*  
563 *Review of Entomology*, 37, 637-665. doi: 10.1146/annurev.en.37.010192.003225
- 564 Robinson, M. D., McCarthy, D. J., & Smyth, G. K. (2010). edgeR: a Bioconductor package for  
565 differential expression analysis of digital gene expression data. [Article]. *Bioinformatics*,  
566 26(1), 139-140. doi: 10.1093/bioinformatics/btp616
- 567 Schonleben, S., Sickmann, A., Mueller, M. J., & Reinders, J. (2007). Proteome analysis of *Apis*  
568 *mellifera* royal jelly. [Article]. *Analytical and Bioanalytical Chemistry*, 389(4), 1087-  
569 1093. doi: 10.1007/s00216-007-1498-2
- 570 Smith, C. R., & Suarez, A. V. (2010). The trophic ecology of castes in harvester ant colonies.  
571 [Article]. *Functional Ecology*, 24(1), 122-130. doi: 10.1111/j.1365-2435.2009.01604.x
- 572 Tautz, J., Maier, S., Groh, C., Rossler, W., & Brockmann, A. (2003). Behavioral performance in  
573 adult honey bees is influenced by the temperature experienced during their pupal  
574 development. [Article]. *Proceedings of the National Academy of Sciences of the United*  
575 *States of America*, 100(12), 7343-7347. doi: 10.1073/pnas.1232346100



- 576 R Core Team. 2014 R: a language and environment for statistical computing. R Foundation for  
577 Statistical Computing, Vienna, Austria. <http://www.R-project.org/>.
- 578 Theraulaz, G., Bonabeau, E., & Deneubourg, J. L. (1998). Response threshold reinforcement and  
579 division of labour in insect societies. [Article]. *Proceedings of the Royal Society B-*  
580 *Biological Sciences*, 265(1393), 327-332.
- 581 Tribble, W., & Kronauer, D. J. C. (2017). Caste development and evolution in ants: it's all about  
582 size. *Journal of Experimental Biology*, 220(1), 53-62. doi: 10.1242/jeb.145292
- 583 Trumbo, S. T., & Robinson, G. E. (1997). Learning and task interference by corpse-removal  
584 specialists in honey bee colonies. [Article]. *Ethology*, 103(11), 966-975.
- 585 Tschinkel, W. R. (1988). Social control of egg laying rate in queens of the fire ant, *Solenopsis*  
586 *invicta*. [Article]. *Physiological Entomology*, 13(3), 327-350. doi: 10.1111/j.1365-  
587 3032.1988.tb00484.x
- 588 Vojvodic, S., Johnson, B. R., Harpur, B. A., Kent, C. F., Zayed, A., Anderson, K. E., &  
589 Linksvayer, T. A. (2015). The transcriptomic and evolutionary signature of social  
590 interactions regulating honey bee caste development. *Ecology and Evolution*, 5(21),  
591 4795-4807. doi: 10.1002/ece3.1720
- 592 Wahl, L. M. (2002). Evolving the division of labour: Generalists, specialists and task allocation.  
593 [Article]. *Journal of Theoretical Biology*, 219(3), 371-388. doi: 10.1006/jtbi.2002.3133
- 594 Wang, J., Wurm, Y., Nipitwattanaphon, M., Riba-Grognuz, O., Huang, Y. C., Shoemaker, D., &  
595 Keller, L. (2013). A Y-like social chromosome causes alternative colony organization in  
596 fire ants. [Article]. *Nature*, 493(7434), 664-668. doi: 10.1038/nature11832

- 597 Wang, Y., Kaftanoglu, O., Fondrk, M. K., & Page, R. E. (2014). Nurse bee behaviour  
598 manipulates worker honeybee (*Apis mellifera* L.) reproductive development. [Article].  
599 *Animal Behaviour*, 92, 253-261. doi: 10.1016/j.anbehav.2014.02.012
- 600 Warner, M. R., Kovaka, K., & Linksvayer, T. A. (2016). Late-instar ant worker larvae play a  
601 prominent role in colony-level caste regulation. [Article]. *Insectes Sociaux*, 63(4), 575-  
602 583. doi: 10.1007/s00040-016-0501-3
- 603 Warner, M. R., Lipponen, J., & Linksvayer, T. A. (2018). Pharaoh ant colonies dynamically  
604 regulate reproductive allocation based on colony demography. [journal article].  
605 *Behavioral Ecology and Sociobiology*, 72(3), 31. doi: 10.1007/s00265-017-2430-1
- 606 Warner, M. R., Mikheyev, A. S., & Linksvayer, T. A. (2017). Genomic signature of kin selection  
607 in an ant with obligately sterile workers. [Article]. *Molecular Biology and Evolution*,  
608 34(7), 1780-1787. doi: 10.1093/molbev/msx123
- 609 Webster, M. M., & Ward, A. J. W. (2011). Personality and social context. [Article]. *Biological*  
610 *Reviews*, 86(4), 759-773. doi: 10.1111/j.1469-185X.2010.00169.x
- 611 Weidenmuller, A., Mayr, C., Kleineidam, C. J., & Roces, F. (2009). Preimaginal and adult  
612 experience modulates the thermal response behavior of ants. [Article]. *Current Biology*,  
613 19(22), 1897-1902. doi: 10.1016/j.cub.2009.08.059
- 614 Wheeler, D. E. (1986). Developmental and physiological determinants of caste in social  
615 hymenoptera- Evolutionary implications. [Article]. *American Naturalist*, 128(1), 13-34.  
616 doi: 10.1086/284536
- 617 Wickham, H. (2009). *ggplot2: Elegant graphics for data analysis*. New York, New York:  
618 Spring-Verlag.
- 619 Wilson, E. O. (1971). *The insect societies*. Cambridge, Massachusetts: Harvard University Press.

- 620 Wilson, E. O. (1987). Causes of ecological success- the case of the ants. [Article]. *Journal of*  
621 *Animal Ecology*, 56(1), 1-9. doi: 10.2307/4795
- 622 Zhang, W., Wanchoo, A., Ortiz-Urquiza, A., Xia, Y. X., & Keyhani, N. O. (2016). Tissue,  
623 developmental, and caste-specific expression of odorant binding proteins in a eusocial  
624 insect, the red imported fire ant, *Solenopsis invicta*. [Article]. *Scientific Reports*, 6, 16.  
625 doi: 10.1038/srep35452
- 626 Zhou, X. F., Rokas, A., Berger, S. L., Liebig, J., Ray, A., & Zwiebel, L. J. (2015).  
627 Chemoreceptor Evolution in Hymenoptera and Its Implications for the Evolution of  
628 Eusociality. [Article]. *Genome Biology and Evolution*, 7(8), 2407-2416. doi:  
629 10.1093/gbe/evv149

**young (head)****old (head)****young (abdomen)****old (abdomen)**

isoprenoid biosynthetic process

single-organism metabolic process

alpha-amino acid metabolic process

single-organism metabolic process

isoprenoid metabolic process

oxidation-reduction process

alpha-amino acid catabolic process

oxidation-reduction process

amino sugar metabolic process

lipid metabolic process

cellular amino acid catabolic process

metabolic process

chitin metabolic process

single-organism process

transmembrane transport

single-organism process

glucosamine-containing compound metabolic process

metabolic process

organic acid catabolic process

proteolysis

aminoglycan metabolic process

NA

carboxylic acid catabolic process

lipid metabolic process

single-organism metabolic process

NA

localization

NA

lipid biosynthetic process

NA

carbohydrate metabolic process

NA

lipid metabolic process

NA

small molecule catabolic process

NA

cellular lipid metabolic process

NA

organonitrogen compound catabolic process

NA