1 COLT-Viz: Interactive Visualization of Antibody Lineage Trees

2	Chenfeng He ^{1,*} , Ben S. Wendel ^{2,*} , Jun Xiao ³ , Keke Chen ⁴ , Ning Jiang ^{1,5,\$}
3	
4	Authors and Affiliations:
5	¹ Department of Biomedical engineering, Cockrell School of Engineering, The University of
6	Texas at Austin, Austin, TX 78712, USA
7	² McKetta Department of Chemical Engineering, Cockrell School of Engineering, The University
8	of Texas at Austin, Austin, TX 78712, USA
9	³ ImmuDX LLC, Austin, TX 78750, USA
10	⁴ Department of Computer Science and Engineering, Wright State University, Dayton, OH 45431,
11	USA
12	⁵ Institute for Cellular and Molecular Biology, University of Texas at Austin, Austin, TX 78712,
13	USA.
14	
15	
16	
17	[*] These authors contributed equally.
18	
19	^{\$} Correspondence should be addressed to:
20	Ning Jiang, Ph.D.
21	Email: jiang@austin.utexas.edu
22	Phone: 512-471-4860
23	

Interactive antibody lineage structure visualization

25 Abstract

Many tools have been developed to visualize phylogenetic trees, which is a traditional technique 26 for evolutionary tree analysis. However, due to the unique characteristics of antibody lineage 27 trees, the phylogenetic method cannot adequately construct proper tree structures for antibody 28 29 lineages, and many other tools have been developed to address this problem. However, there still lacks of an adequate tool to visualize the resulted antibody lineage structures that are more 30 31 complicated than phylogenetic trees. In addition, high-throughput sequencing-based antibody repertoire profiling enables the counting of the number of transcripts associated with individual 32 33 antibody sequences, thus more dimensions need to be encoded in the tree structure visualization. 34 Further, users may wish to manually adjust the tree structure for a special context. When doing so, they may wish to maintain some biological constraints that are applicable in antibody lineage 35 36 tree structure, such as isotype switching constraints or different sampling constraints. Here, we report an interactive visualization tool (COLT-Viz) designed to display the number of RNA 37 38 copies, number of somatic hypermutations, and sample collection time associated with each antibody sequence as well as the distance to neighboring sequences for each antibody sequence 39 40 in the lineage. COLT-Viz also allows users to interactively visualize and edit antibody lineage structures while giving users the option to automatically check biological constraints on the 41 42 edited structures to ensure accuracy. COLT-Viz takes JSON text format as input files and can easily be used to visualize networks with or without the biological constraints. We believe the 43 amount of information that can be displayed for complex antibody lineages, the interactive 44 interface, and the option of checking for biological constraints make COLT-Viz a versatile tool 45 for antibody lineage tree visualization that will guide further biological discoveries. 46

47

48

Key words: bioinformatics software, antibody lineage structure, biological constraints,
interactive visualization, intuitive user interface

Interactive antibody lineage structure visualization

51 **1. Introduction**

Lineage analysis is frequently used by researchers to analyze the evolution of different species of organisms or to trace cellular development within an organism. Specifically in the immune system, lineage analysis has been adopted to trace the mutational evolution of antibodies (1–3) and understand selection pressure (4,5).

Typically, an antibody lineage is represented as a tree structure, which describes the 56 57 possible development path (i.e., edges in a tree) among all unique antibody sequences (i.e., nodes in a tree). Traditionally, researchers used phylogenetic trees for antibody lineages; however, a 58 59 phylogenetic tree is a binary tree with all nodes presented as leaf nodes, which is not suitable for 60 antibody lineages (3). Much work have been done to construct tree structures specific to antibody lineages (3,6,7); however, there is still a need for a tool to efficiently visualize these 61 62 trees. The visualization of antibody lineage trees has a few unique needs: (a) there are multiple features specific to antibody sequences (e.g., number of transcripts, number of somatic 63 64 hypermutations, isotype, etc), integrating all these properties together for tree visualization will facilitate lineage analysis; (b) interactively visualize the antibody structure by showing antibody 65 66 sequence information and associated properties when users click on a node of interest. This allows user to have access to detailed information of nodes (antibody sequences) without losing 67 68 the global view of a complex antibody lineage structure; and (c) interactively modify antibody lineage structure. Oftentimes, multiple valid lineage tree structures exist for an antibody lineage. 69 70 Based on special considerations, researchers may want to manually alter their tree structures by 71 changing or adding connections between nodes. When doing so, they may want to make sure that 72 these modified connections do not violate certain biological constraints. These constraints can either be biological constraints (8) or constraints determined by experiment design (9). 73

Although many network visualization tools with powerful functions have been developed (e.g., Cytoscape (10), Graphviz (11)), they were not designed for antibody lineage structure visualization and thus lack the consideration of biological properties and constraints. To our knowledge, there is no tool specifically designed to visualize antibody lineage tree structures with the functionalities mentioned above. For example, LymAnalyzer (12) incorporates FigTree (13) to visualize antibody lineage tree structures. However, FigTree is only designed for phylogenetic tree which is not suitable for antibody lineage structure and does not have an

Interactive antibody lineage structure visualization

interactive user interface. Another program, Change-O (6), modifies phylogenetic tree analysis
for antibody lineage application, but its visualization function can only generate non-interactive
figures. In addition, none of these tools can easily be used to visualize the multiple features of
antibody lineages, which are unique to antibody lineage tree visualization.

Bearing the drawbacks of current visualization tools in mind, we developed COLT-Viz for antibody lineage tree structure visualization. COLT-Viz enables researchers to interactively explore lineage trees with multiple antibody sequence properties encoded and automatically checked when the tree structure is manually altered. At the same time, researchers have the option to override error messages and proceed with the desired changes. This tool enables users to discover interesting patterns of antibody evolution process and select interesting nodes (antibody sequences) for further evaluation.

92 Specifically, COLT-Viz projects antibody sampling time, total mutations for a sequence, mutation distances between two neighboring sequences, and antibody RNA copies of unique 93 94 sequences onto different elements of each node in the lineage visualization, which allows users to intuitively understand the lineage evolution over time, mutation distances from a germline 95 sequence, mutation distances between neighboring nodes, and the clonal size associated with 96 each unique antibody sequence. Furthermore, users can fine tune the lineage by removing edges 97 and reconnecting nodes with all the constraints automatically checked by the system with an 98 option to either override the error message or make other alternative changes. When users edit 99 100 the connections, COLT-Viz checks two types of constraints: (a) isotype constraint: as described in previous studies (8,14-16), antibody isotype switch can only proceed according to the order of 101 102 IGH constant genes located on the chromosome (17), thus users may want to maintain this 103 constraint while editing the lineage; (b) time constraint: as described in Schramm et al. (9), 104 longitudinal information offers another dimension in some biological experiments (18). 105 Although in many cases, sequences from an earlier time point may not necessarily be ancestors of sequences from later time point, in some cases, such as transplantation, where donor samples 106 107 and recipient samples are experimentally separated, this constraint is critical. Thus, users may 108 want to re-enforce the time constraint when manually changing node connections. Therefore, 109 COLT-Viz also checks if any manual changes on the tree structure comply with the time constraint that sequences from an earlier time point should be ancestors of sequences from a later 110

Interactive antibody lineage structure visualization

time point while give users the option of override this error message and proceed with their intended changes.

113 **2. Implementation**

114 COLT-Viz takes the lineage tree from tree-generating tools (e.g., COLT (7), Change-O (6)) as input and outputs the tree visualization. The input of COLT-Viz contains one node file 115 and one edge file. The node and edge files are encoded in JSON (JavaScript Object Notation) 116 117 format (http://json.org/), which is a lightweight and human-readable text format. JSON expresses 118 data objects in attribute-value pairs, which makes it an ideal data format for visualizing antibody 119 sequences with multiple attributes. Outputs from tree-generating tools/algorithms can be 120 converted to the JSON text format to be compatible with COLT-Viz. Users can also manually edit the JSON text files to change the lineage tree structure. The node file contains a list of nodes, 121 122 or unique antibody sequences. Each node element includes the node identity, node description, associated sequence abundance (NRNAs, number of RNA copies, and NREADs, number of 123 124 sequencing reads), antibody isotype (i.e., antibody heavy chain families, IgM, IgD, IgG, IgA, and IgE), timestamp (i.e., sampling time point, for example, when doing research on vaccine, 125 126 sequences obtained before vaccination are all assigned to a smaller number (e.g. 20) while sequences obtained 7 days post vaccination are all assigned to a larger number(e.g. 40)), and the 127 number of somatic hypermutations from the germline sequence. Each edge in the edge file is 128 defined as a 3-tuple (parent node identity, weight, child node identity), which are also encoded in 129 130 the JSON format.

Users can explore the tree visualization, e.g., checking the detailed information of each node, and fine tune the tree structure by editing and moving the edges. Before making changes, COLT-Viz automatically checks the constraints and displays a warning message if either the timepoint or isotype constraint is violated. However, user can choose to override this warning message and proceed with the move. Once old edges are removed and new edges are added, the system will automatically update the entire graph. Finally, the updated tree structure can be saved back to the edge files in the JSON format for later usage.

138

Interactive antibody lineage structure visualization

140 **3. Results**

141 **3.1** Lineage Tree Visualization

We visually encode multiple properties of nodes and edges into the lineage tree visualization, so 142 143 users can understand these properties intuitively. Figure 1 shows an antibody lineage we found in a young African child who experienced acute malaria twice in two consecutive malaria seasons 144 from our previous study (19). There are a total of 23 unique sequences obtained from 4 145 timepoints in this lineage. The height of each node represents the number of RNA copies 146 147 associated with that unique sequence. Heatmap is used to color nodes – the warmer the color, the greater the number of somatic hypermutations to the germline sequence. The border of each node 148 149 is colored to represent different timestamps, so users can easily identify the evolution of the antibody lineage over time. 150

The tree layout algorithm uses the well-known force-directed graph drawing algorithm 151 (20). It progressively changes the nodes' positions, as if there are physical forces between the 152 153 nodes. Imagine that neighboring nodes are connected by springs: the force of the spring will prevent the nodes from drifting too far apart or collapsing too close together. This algorithm 154 ideally places the root antibody sequence towards the center of the visualization and unfolds the 155 tree outward to maximize the use of the display area. Thus the directionality of the tree is in 156 157 general from center to periphery. Knowing this makes it easier for user to identify the root node, also if user assign the root as a specific ID (e.g., 0, same as in Figure 1), then the root node 158 should be easily identified in the tree. Then, by following the directionality, users can easily 159 160 understand the development of the whole lineage.

Using COLT-Viz, features of the antibody lineages that are not obvious by mining the 161 162 data can be quickly perceived visually. Using the lineage in Figure 1 as an example, users can quickly identify the root sequence and see that, for this lineage, the sequences at later time points 163 164 have accumulated more somatic hypermutations compared to sequences collected at earlier time points, and they lie in the outer leaves of the lineage. In addition, the relatively even distribution 165 166 of node heights indicates that the RNA copies of these unique sequences do not differ widely. These visual features help immunologists quickly glean an overview of the evolution of this 167 168 antibody lineage over time.

Interactive antibody lineage structure visualization

169

170 3.2 Tree Editing and Constraint Maintaining

COLT-Viz allows users to alter the network connections by removing and adding edges between 171 172 nodes. When a new edge is added, the system will check two types of constraints: (a) time constraint, an edge $x \rightarrow y$ is valid only if x's timestamp is earlier (smaller) than y's; and (b) 173 isotype constraint, an edge x->y is valid only if the isotype is either unchanged or following the 174 class-switching rule: a class-switched sequence cannot switch back to IgM. If either of the 175 176 constraints is violated, COLT-Viz will display a warning. However, users can override this warning and proceed to add the new edge or heed the warning and cancel the new edge. The 177 constraint checking helps avoid artificial mistakes and maintain interesting time features when 178 users manually modify antibody lineage trees. 179

180 **3.3** Intuitive graphic visualization user interface

181 COLT-Viz employs an intuitive graphic visualization user interface (Figure 1). Within the 182 antibody lineage structure displaying window, users can drag on any node to reposition the tree. 183 Users can also click on any node to view the full antibody sequence and annotated information in 184 the sequence display window on the right. In addition, users can change the lineage structure by 185 clicking on a node and specifying the new parent node that they wish to establish a link in the 186 edge editing window.

187 **4. Discussion**

COLT-Viz is an interactive tool designed for antibody lineage tree visualization and editing. By 188 189 using COLT-Viz, researchers can understand lineage trees, validate algorithmic results, and fine-190 tune the tree structures with additional domain knowledge that automated algorithms cannot capture. Further, COLT-Viz allows researcher to identify nodes (antibody sequences) that bear 191 specific features that are only obvious in graphic settings. Constraints are automatically checked 192 to avoid manual editing errors. Although COLT-Viz is specifically designed for antibody lineage 193 194 analysis, we believe with small tweaks it can be easily applied to any other type of lineage 195 analysis in biomedical research.

196

Interactive antibody lineage structure visualization

198 Availability of data and software

- 199 Project name: COLT-Viz
- 200 Project home page: <u>https://github.com/immudx/paper</u> (sample data is included).
- 201 Operation systems: Platform independent
- 202 Programming language: Java
- 203 Any restrictions to use by non-academics: None
- 204

205 Abbreviations

206 IgM (D,G,A,E): Immunoglobulin M (D,G,A,E)

Funding: This work was supported by NIH grants R00AG040149 (N.J.) and by the Welch
Foundation grant F1785 (N.J.). NJ is a Cancer Prevention and Research Institute of Texas
(CPRIT) Scholar and a Damon Runyon-Rachleff Innovator. BW is a recipient of the Thrust 2000
- George Sawyer Endowed Graduate Fellowship in Engineering.

211

Author contributions: NJ conceived the idea and directed study; CH, BW and NJ participated in the algorithm design. KC and JX wrote the computer software. CH, BW contributed to software testing and optimization. CH, KC, JX and NJ contributed to the writing of the manuscript. All of the authors read and approved the final manuscript.

- 216 **Conflict of Interest Statement**: Ning Jiang is a scientific advisor of ImmuDX LLC.
- 217 Acknowledgements: Not applicable

218

- 220
- 221
- 222

Interactive antibody lineage structure visualization

223 **References**

- 1. Jiang N, He J, Weinstein J a, Penland L, Sasaki S, He X-S, Dekker CL, Zheng N-Y,
- Huang M, Sullivan M, et al. Lineage structure of the human antibody repertoire in
- response to influenza vaccination. *Sci Transl Med* (2013) **5**:171ra19.
- doi:10.1126/scitranslmed.3004794
- 228 2. Liao H-X, Lynch R, Zhou T, Gao F, Alam SM, Boyd SD, Fire AZ, Roskin KM, Schramm
- CA, Zhang Z, et al. Co-evolution of a broadly neutralizing HIV-1 antibody and founder
 virus. *Nature* (2013) **496**:469–76. doi:10.1038/nature12053
- 3. Barak M, Zuckerman NS, Edelman H, Unger R, Mehr R. IgTree[©]: Creating
- Immunoglobulin variable region gene lineage trees. *J Immunol Methods* (2008) 338:67–74.
 doi:10.1016/j.jim.2008.06.006
- Yaari G, Vander Heiden JA, Uduman M, Gadala-Maria D, Gupta N, Joel JN, O'Connor
 KC, Hafler DA, Laserson U, Vigneault F, et al. Models of somatic hypermutation
 targeting and substitution based on synonymous mutations from high-throughput
 immunoglobulin sequencing data. *Front Immunol* (2013) 4:
- 238 doi:10.3389/fimmu.2013.00358
- 5. Hershberg U, Uduman M, Shlomchik MJ, Kleinstein SH. Improved methods for detecting
 selection by mutation analysis of Ig V region sequences. *Int Immunol* (2008) 20:683–694.
 doi:10.1093/intimm/dxn026
- 6. Gupta NT, Vander Heiden JA, Uduman M, Gadala-Maria D, Yaari G, Kleinstein SH.
- 243 Change-O: A toolkit for analyzing large-scale B cell immunoglobulin repertoire
- sequencing data. *Bioinformatics* (2015) **31**:3356–3358. doi:10.1093/bioinformatics/btv359
- 245 7. Chen K, Sai V, Gogu A, Wu D, Ning J. COLT: Constrained Lineage Tree Generation
 246 from Sequence Data. *Proc IEEE Int Conf Bioinforma Biomed 2016* (2016)
- Horns F, Vollmers C, Croote D, Mackey SF, Swan GE, Dekker CL, Davis MM, Quake
 SR. Lineage tracing of human B cells reveals the in vivo landscape of human antibody
 class switching. *Elife* (2016) 5: doi:10.7554/eLife.16578.001
- 250 9. Schramm CA, Sheng Z, Zhang Z, Mascola JR, Kwong PD, Shapiro L. SONAR: A high-

Interactive antibody lineage structure visualization

251 252		throughput pipeline for inferring antibody ontogenies from longitudinal sequencing of B cell transcripts. <i>Front Immunol</i> (2016) 7 : doi:10.3389/fimmu.2016.00372
253 254 255	10.	Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B, Ideker T. Cytoscape: A software Environment for integrated models of biomolecular interaction networks. <i>Genome Res</i> (2003) 13 :2498–2504. doi:10.1101/gr.1239303
256 257 258	11.	Gansner ER, North SC. An open graph visualization system and its applications to software engineering. <i>Softw Pract Exp</i> (2000) 30 :1203–1233. doi:10.1002/1097-024X(200009)30:11<1203::AID-SPE338>3.0.CO;2-N
259 260 261	12.	Yu Y, Ceredig R, Seoighe C. LymAnalyzer: A tool for comprehensive analysis of next generation sequencing data of T cell receptors and immunoglobulins. <i>Nucleic Acids Res</i> (2015) 44 : doi:10.1093/nar/gkv1016
262 263	13.	Drummond AJ, Suchard MA, Xie D, Rambaut A. Bayesian phylogenetics with BEAUti and the BEAST 1.7. <i>Mol Biol Evol</i> (2012) 29 :1969–1973. doi:10.1093/molbev/mss075
264 265 266	14.	Iwasato T, Shimizu A, Honjo T, Yamagishi H. Circular DNA is excised by immunoglobulin class switch recombination. <i>Cell</i> (1990) 62 :143–9. doi:0092-8674(90)90248-D [pii]
267 268	15.	von Schwedler U, Jack HM, Wabl M. Circular DNA is a product of the immunoglobulin class switch rearrangement. <i>Nature</i> (1990) 345 :452–456. doi:10.1038/345452a0
269 270 271 272	16.	Yoshida K, Matsuoka M, Usuda S, Mori A, Ishizaka K, Sakano H. Immunoglobulin switch circular DNA in the mouse infected with Nippostrongylus brasiliensis: evidence for successive class switching from mu to epsilon via gamma 1. <i>Proc Natl Acad Sci U S A</i> (1990) 87 :7829–7833. doi:10.1073/pnas.87.20.7829
273	17.	Murphy, K., & Weaver C. Janeway's immunobiology. Garland Science
274 275 276 277	18.	Wu X, Zhang Z, Schramm CA, Joyce MG, Do Kwon Y, Zhou T, Sheng Z, Zhang B, O'Dell S, McKee K, et al. Maturation and diversity of the VRC01-antibody lineage over 15 years of chronic HIV-1 infection. <i>Cell</i> (2015) 161 :480–485. doi:10.1016/j.cell.2015.03.004

Interactive antibody lineage structure visualization

278	19.	Wendel BS, He C, Qu M, Wu D, Hernandez SM, Ma K-Y, Liu EW, Xiao J, Crompton PD,		
279		Pierce SK, et al. Accurate immune repertoire sequencing reveals malaria infection driven		
280		antibody lineage diversification in young children. Nat Commun (2017) 8:531.		
281		doi:10.1038/s41467-017-00645-x		
282	20.	Kobourov SG. Spring Embedders and Force Directed Graph Drawing Algorithms. arXiv		
283		<i>Prepr arXiv12013011</i> (2012)1–23.		
284				
285	Figur	'es:		
286	Figur	re 1. User interface of COLT-Viz. Within the 'Lineage structure window', height of each		
287	node represents the RNA copy of each sequence, color of each node represents the mutation with			
288	respect to the germline, color of the border of each node represents the time point and the edge			
289	length represents the edit distance between neighboring nodes if user specifies the option. By			
290	clicking on a node, the information of this node will be displayed in the 'sequence display			
291	window'. User can also edit the connections through 'edge editing window'. The 'lineage			
292	structure window' shows an antibody lineage found in a young child who experienced two			
293	conse	cutive malaria infection. 23 sequences were found from 4 time points and visualized using		
294	COL	Γ-viz. Four time points are: year 1 pre-malaria infection (light orange node frame for most		
295	of the nodes displayed), year 1 acute malaria infection (brown node frame for node 16, 13, and 8			
296	year 2 pre-malaria infection (light purple), and year 2 acute malaria infection (dark purple).			
297	Sequence 0 is the root and arrows point to the progeny sequences. Sequences from later time			
298	exist	in the outer leaves of the lineage, which represents their evolution overtime.		

