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10	LS <sup>x</sup> : Automated reduction of gene-specific lineage
11	evolutionary rate heterogeneity for multi-gene phylogeny
12	inference
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## ABSTRACT

Motivation: LS<sup>3</sup> is a recently published algorithm to reduce lineage evolutionary rate heterogeneity,
a condition that can produce inference artifacts in molecular phylogenetics. The LS<sup>3</sup> scripts are
Linux-specific and the criterion to reduce lineage rate heterogeneity can be too stringent in datasets
with both very long and very short branches.

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32 **Results:** LS<sup>X</sup> is a multi-platform user-friendly R script that performs the LS<sup>3</sup> algorithm, and has 33 added features in order to make better lineage rate calculations. In addition, we developed and 34 implemented an alternative version of the algorithm, LS<sup>4</sup>, which reduces lineage rate heterogeneity 35 not only by detecting branches that are too long but also branches that are too short, resulting in less 36 stringent data filtering.

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Availability: The LS<sup>x</sup> script LSx\_v.1.1.R and the user manual are available for download at:
 <a href="https://genev.unige.ch/research/laboratory/Juan-Montoya">https://genev.unige.ch/research/laboratory/Juan-Montoya</a>

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### **INTRODUCTION**

We recently showed that biases emerging from evolutionary rate heterogeneity among lineages in multi-gene phylogenies can be reduced with a sequence data subselection algorithm to the point of uncovering the true phylogenetic signal (Rivera-Rivera and Montoya-Burgos 2016).

In that study, we presented an algorithm called Locus Specific Sequence Subsampling (LS<sup>3</sup>), 44 45 which reduces lineage evolutionary rate heterogeneity gene-by-gene in multi-gene datasets. For 46 each gene alignment, LS<sup>3</sup> implements a likelihood ratio test (LRT) (Felsenstein 1981) between two 47 models. One model assumes equal rates of evolution among all ingroup lineages (single rate model) and the other model assumes that three (or more), user-defined ingroup lineages have their own 48 49 independent rate of evolution (multiple rates model). If the multiple rates model fits the data significantly better than the single rate model, the branch lengths of the phylogeny are estimated, 50 51 the fastest-evolving sequence is removed, and the new reduced gene sequence dataset is tested 52 again with the LRT. This process is iterated until a set of ingroup taxa is found whose lineage 53 evolutionary rates can be explained equally well by the single rate model or the multiple rates 54 model. The fast-evolving sequences that had to be removed from each gene alignment to reach this 55 point are flagged as potentially problematic due to their higher evolutionary rate, and gene datasets 56 which never reached this point are flagged as a whole as potentially problematic (Rivera-Rivera and 57 Montova-Burgos 2016). LS<sup>3</sup> proved to be effective in reducing long branch attraction (LBA) 58 artifacts in simulated nucleotide data and in two biological multi-gene datasets (one in nucleotides, 59 and one in amino acids), and its potential to reduce phylogenetic biases has been recognized by 60 several other authors (Cruaud and Rasplus 2016; Suh 2016; Bleidorn 2017).

61 Concomitantly to the publication of the LS<sup>3</sup> algorithm, a set of bash scripts were made 62 available to perform these tasks automatically (http://genev.unige.ch/en/users/Juan-Montoya/unit). 63 While these bash scripts are effective, they only run in Linux systems, and the layout and 64 programming is not entirely user-friendly. This prompted us to produce a new, re-programmed 65 version of the LS<sup>3</sup> algorithm which is user-friendly , contains important new features, and can be

66	used across platforms. For this new implementation we also developed a second data subselection
67	algorithm based on LS <sup>3</sup> , called "LS <sup>3</sup> supplement" (LS <sup>4</sup> ) which subselects sequences for lineage
68	evolutionary rate homogeneity by not only removing very long branches, but also very short ones.

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# LS<sup>X</sup> DESCRIPTION

71 The new script is called LS<sup>X</sup>, is entirely written in R (R Core Team 2016), and uses PAML 72 (Yang 2007) and the R packages ape (Paradis et al. 2004; Popescu et al. 2012) and adephylo 73 (Jombart et al. 2010). If PAML, R, the R packages and Rscript are installed and functional, the 74 script runs regardless of the platform, with all parameters given in a single raw text control file. As 75 with the bash implementation of LS<sup>3</sup>, LS<sup>X</sup> reads alignments in PHYLIP format, but unlike the 76 former, it produces for each gene a version of the alignment including only the sequences that satisfied the lineage rate homogeneity criterion. In LS<sup>X</sup>, the best model of sequence evolution can be 77 78 given for each gene (in the original implementation only a single model could be selected for all 79 genes), thus improving the accuracy of the branch length estimations and likelihood calculations 80 under PAML. In addition, the users can select more than three lineages of interest for the lineage 81 evolutionary rate heterogeneity test (Fig. 1*a*,*b*), while the bash implementation was hardcoded to 82 work with only three lineages of interest.

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## LS<sup>3</sup>: AN LS<sup>4</sup> SUPPLEMENT

Within  $LS^{x}$  we also implemented  $LS^{4}$ , a second data subselection algorithm that is optimized for datasets in which not only extremely long branches but also extremely short branches are present in the starting phylogenetic tree. While both  $LS^{3}$  and  $LS^{4}$  follow the same general algorithm, they differ in the criterion for choosing the sequence to be removed in the sequence subselection steps. Under  $LS^{3}$ , the fastest-evolving of the ingroup sequences is removed in each iteration, as determined by its calculated sum of branch length (SBL), starting from the stem of the ingroup (Fig. 2*a*). This can lead to the flagging of too much data in a dataset containing extremely

92 slow-evolving sequences (Fig. 2*b*). In such cases, under LS<sup>3</sup>, not only the fast-evolving sequences 93 will be removed, but also the sequences with intermediate evolutionary rates which are still 94 evolving "too fast" relative to the extremely slow-evolving ones (Fig. 2*b*). While this approach is 95 not incorrect, simply very stringent, it can lead to the flagging of too many sequences in certain 96 datasets and to poorly-resolved phylogenies when concatenating and analyzing the remaining data.

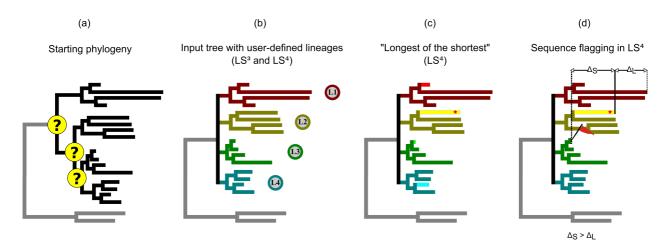


Figure 1. Schematic representation of the procedure for flagging sequences in  $LS^4$ . In (*a*), the general phylogeny for this group of taxa, with the nodes in question highlighted. For for  $LS^3$  and  $LS^4$ , an input tree is given in which the nodes into question are collapsed, and the lineages involved are identified (*b*). In (*c*),  $LS^4$  identifies the shortest branch for each clade (highlighted), and then identifies the longest among them (red star). The sequence to be removed in each iteration of  $LS^4$  is the one with the tip furthest from the tip of the "longest of the shortest" branch (*d*), resulting in the flagging of both extremely long and extremely short branches.

98 In order to allow for the inclusion of more data while still reaching lineage rate homogeneity, LS<sup>4</sup> employs a different criterion which considers both too fast- and too slow-99 100 evolving sequences for removal. Under LS<sup>4</sup>, when the SBLs for all ingroup sequences of a given gene are calculated, they are grouped by the user-defined lineage of interest to which they belong. 101 102 The shortest branch of each lineage of interest is identified, and then the longest among them across 103 all ingroup lineages ("the longest of the shortest", see Fig. 1*c*) is picked as a benchmark. Because in both LS<sup>3</sup> and LS<sup>4</sup> each lineage of interest has to be represented by a minimum of species (defined 104 105 by the user), this "longest of the shortest" branch represents the slowest evolutionary rate at which 106 all lineages could converge. Then, in the sequence subselection steps, the sequence to be removed is 107 the ingroup sequence for which the absolute value of the difference between its SBL and the SBL of 108 the benchmark sequence is the largest. In other words, for a given gene, the sequence removed in each iteration of LS<sup>4</sup> is that which produces the tip furthest from the benchmark, be it faster- or 109

- 110 slower-evolving (Fig. 1*d*). We are confident that the criterion used in LS<sup>4</sup> will result in less genes
- 111 flagged completely (as compared to LS<sup>3</sup>) when extremely slow-evolving sequences are present (Fig.
- 112 2b).

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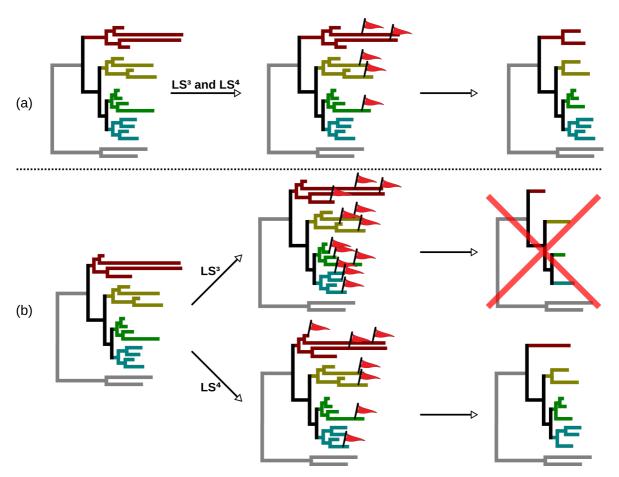
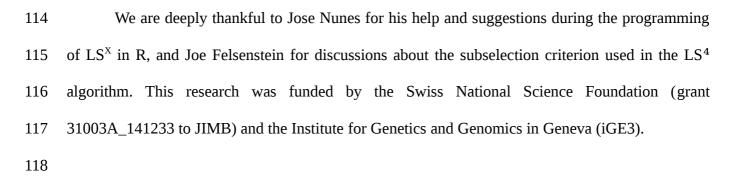


Figure 2. A schematic representation of the different ways that  $LS^3$  and  $LS^4$  reach lineage rate homogeneity for a given gene sequence dataset. In (*a*), a dataset with a general rate homogeneity with the exception of several faster evolving branches. In this case, both methods will flag the faster evolving sequences, and reach the same taxon subset with homogeneous rates of evolution. In (*b*), a dataset with general lineage rate heterogeneity, and with a very short branch (the top branch of that tree). In such a case,  $LS^3$  will remove all of the faster evolving sequences, until only the slowest sequence of each clade of interest remains. At this point, that gene sequence dataset is flagged completely as problematic because lineage rate heterogeneity is still too strong. In contrast,  $LS^4$  will remove the faster evolving sequences and also the slowest one, thus reaching lineage rate homogeneity for this dataset.

#### **ACKNOWLEDGEMENTS**



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