The study of animal behaviour as a physical science

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Abstract

Behaviour is the ultimate output of an animal's nervous system and choosing the right action at the right time can be critical for survival. A quantitative understanding of behaviour would therefore advance research in neuroscience as well as ecology and evolution. However, animal posture typically has many degrees of freedom and behavioural dynamics vary on timescales ranging from milliseconds to years, presenting both technical and conceptual challenges. Here we review 1) advances in imaging and computer vision that are making it possible to capture increasingly complete records of animal motion and 2) new approaches to understanding the resulting behavioural data sets. With the right analytical approaches, these data are allowing researchers to revisit longstanding questions about the structure and organisation of animal behaviour and to put unifying principles on a quantitative footing. Contributions from both experimentalists and theorists are leading to the emergence of a physics of behaviour and the prospect of discovering laws and developing theories with broad applicability. We believe that there now exists an opportunity to develop theories of behaviour which can be tested using these data sets leading to a deeper understanding of how and why animals behave.

Keywords: Behaviour, ethology, data, posture dynamics, discretisation, hierarchy, stereotypy, low dimensionality

Introduction

Animals are material things living in a material world governed by physical laws. Research fields where the influence of physics on behaviour is clearest (such as biomechanics, the function of sensory systems, and the physiology of nervous tissues) have well-developed collaboration between biologists and physicists. This is research aimed at the *physical underpinnings* of behaviour. However, we believe that behaviour itself, at the level of the organism, can be fruitfully studied from a physics perspective. The principled, quantitative approach to animal dynamics represents a nascent *physics of behaviour* that is being explored by both experimentalists and theorists.

The conceptual basis for the study of animal behaviour was laid by scientists interested in *Tierpsychology* (animal psychology) or ethology in the early twentieth century[1]. They were fascinated by the bewildering variety of behaviours in nature and wanted to systematically characterise this variety and to understand why animals behaved in the way they did. A modern variant of this approach is the study of an animal's location over time, a field sometimes called movement ecology[2], which has led to fascinating advances on efficient search strategies[3,4] and the study of collective behaviour[5–7]. However, what interested the early ethologists, and our focus in this article, is the study of behavioural repertoires that include not just where an animal is in its environment, but, in Tinbergen's words, the "total movements made by the intact animal"[8]. In early ethology, behaviour was typically categorised by trained observers taking advantage of their experience and intuition to identify relevant elements such as feeding, fighting, mating, or fleeing, but

Tinbergen's definition leaves open the question of how behaviour should be measured and represented. It does not prescribe any particular partitioning or even whether partitioning behaviour into discretely labelled actions is appropriate.

Similar problems arise in physics, where we often have the freedom to measure a vast number of quantities and where the best representation is not always clear. For example, the development of a microscopic theory of superconductivity (BCS) relied not just on experimental observations but also a body of phenomenological theories that each captured an aspect of the problem from a different perspective[9]. By analogy, we believe that alternative representations of animal movement will reveal different phenomena. This approach is well-suited to physicists, and its findings will be helpful in identifying lower level mechanisms underpinning behaviour.

To make use of a given representation, the relevant variables must be measurable in an experiment, preferably in large quantities at low cost. Improved cameras and computer vision algorithms have greatly expanded the range of accessible spatial and temporal scales for behavioural quantification as well as the range of environments in which recording is possible. We begin by reviewing how current experimental techniques enable the quantification of behaviour and then consider how measurements are revealing principles of behaviour.

Techniques

Posture tracking

Behavioural repertoires are composed of posture changes over time. Quantifying the posture of an organism typically begins with an imaging experiment that records a direct representation of the animal, such as an image of it in a video. From this input, increasingly abstract representations of the animal can be computed using a variety of approaches (Figure 1). Historically, this required the mind-numbing manual annotation of individual video frames. This meant postural quantification for long videos at high framerates was essentially impossible. Stopgap techniques drew inspiration from human motion capture techniques, and placed small dye spots on limbs which, by virtue of their contrast or fluorescence, could be isolated from the image background and automatically tracked[10–12]. Gluing on tracking markers introduces potential confounds, particularly the continuous, unnatural mechanosensory stimulation of subject animals.

Markerless techniques emerged exploiting frustrated internal reflection to characterise the contact points between an animal and its substrate[13–15], however these approaches cannot track portions of the animal separated from the substrate. A potential way forward to automatically identify postural landmarks lies in model-based image analysis, in which the known biomechanical degrees of freedom of an animal are used to find the posture that would produce a given image with the highest likelihood[16–18]. The challenges of automatic annotation of subtle behavioural events may be solved by deep learning/ neural network approaches[19,20]. However, as these require human-annotated training sets, the power of the technique may

Figure 1 — A modern pipeline for the analysis of behavioural data — From a raw video recording, machine vision techniques are used to produce a representation of the animal's posture in each frame. This can take the form of coordinates of skeletonised landmarks or modes of pixel intensities. This representation can be analysed directly by the techniques shown at the bottom of the pipeline, or first transformed into a posture-dynamics space using, e.g., time-frequency analysis, and then analysed. w = width of video frame, h = height of video frame, t = length of video recording in frames, n = number of skeletal landmarks quantified, m = number of image or skeletal element modes identified, f = number of frequency bands. Generally in such analyses $m \approx f << w \approx h << t$.

be offset by the inconsistency in human labels, which vary from lab to lab and expert to expert[21,22].

An alternative to using image analysis to extract landmark-based or skeletonised representations is to use the raw pixel luminosities in animal-aligned video frames (Figure 1). Identification of the principal components (eigenvectors of the covariance matrix) of pixel luminosity reveals the regions of the animal that change during behaviour[23,24]. The advantage of this pixel-based representation is that it can be applied to animals of any shape or limb configuration.

Real-time feedback and brain imaging

The methods described above vary greatly in their computational intensity. Some computationally lighter methods can be performed in real time, like fitting an ellipse to the silhouette of an adult *Drosophila melanogaster* fly[25] or segmenting the midline of a *Caenorhabditis elegans* round worm[26]. This enables closed-loop experiments in which the animal is stimulated (either through its sensory systems[25,27] or directly in the nervous system using optogenetics[25,27–29]) in response to behaviours it performs in real time, as soon as they are flagged. This also permits real-time prediction of upcoming behaviours, which can facilitate close tracking of animals, improving the quality of neural activity recordings acquired from freely moving animals[30], which faces substantial challenges both in tracking individual animals for long periods and in registering volumetric neural imaging data.

Bridging behavioural timescales

A conspicuous feature of long behavioural recordings modulation on multiple timescales. The swim bouts of zebrafish last a few tens of milliseconds[31], and flies can turn in response to odour stimuli within 70ms, less than the length of a single running stride[32]. And yet, the behavioural repertoire of both of these organisms is tuned by long timescale processes such as satiety/hunger[33] and the circadian cycle[25]. Behaviour changes over the longest timescale possible in organisms, the lifetime[34], and quantitative variation in behaviour with age is conspicuous in many measurements.

While it remains challenging to acquire high resolution video data over long time periods, there are approaches that permit the measurement of behaviour on timescales of lifetimes. Such longitudinal experiments are naturally most advanced for organisms with short lifespans, such as single-celled microbes where tracking of entire cohorts for multiple generations is possible[35]. Long-term recordings are facilitated by real-time processing of video data to extract and save only compressed representations of behaviour, such as animals' centroids. Such low dimensional representations facilitate not just experiments in depth (over time), but also in breadth (over individuals). As an example, tracking the centroids of many flies in parallel exploring simple arenas has permitted the measurement of individual behaviour from tens of thousands of individuals across hours and days[36,37]. In worms, midline tracking can be done in real time for dozens of individuals[26]. Combining multi-worm midline tracking with custom sample chambers[34,38] would enable cradle-to-grave tracking of an animal's complete movements with sub-second resolution, providing valuable information about behavioural ageing and healthspan that could be combined with genetic and drug perturbation and compared with other physiological changes. In mice, the highest resolution tracking is typically done in for limited periods in fairly featureless 'open fields'[39,40], but longer timescales can be accessible in systems that track mice in their home cages[41–44].

Inexpensive and field imaging

At the same time that new instrumentation has permitted deep and broad behavioural recordings, the affordability and versatility of behaviour rigs has improved. There are now open source designs for behaviour recording devices that capture behaviour from many individuals simultaneously while also permitting closed-loop stimulation of animals[25,45]. These technologies are flourishing in part because of the proliferation of inexpensive hardware for data acquisition, such as single-board computers (e.g., Raspberry Pi) and convenient break-out microcontroller boards (e.g., Ardunio and Teensy). This open hardware community thrives in the groundwork laid by earlier open source animal tracking algorithms[46–48]. We believe there is great potential to bring these technologies out of the lab and into the field to capture behaviour in circumstances closer to that in which it evolved, its proper ethological context[49].

Principles

The growing technical sophistication of behavioural studies has correlated with growing conceptual insights. Several principles have been identified to help guide exploration of animal behaviour (Figure 2). Some of the principles have close parallels in traditional physics, while others appear to be uniquely biological. Some are more tentative, while others have been found in diverse species in diverse contexts. Overall, they highlight regularities in behaviour that may lead to novel theory.

Low postural dimensionality

In principle, animals have many postural degrees of freedom. However, during normal behaviour, correlations between body parts make it possible to derive lower dimensional representations of posture that reveal aspects of coordination and also provide a useful basis for other behavioural analyses. In *C. elegans*, the length-wise midline predicts with high fidelity the position of the entire body. This was exploited to determine the dimensionality of worm posture during spontaneous exploration and it was found that four dimensions explain 95% of the postural variance[50]. Two of these "eigenworms" correspond approximately to a quadrature pair that supports travelling waves with a third mode capturing overall curvature. Related methods agree on the dimensionality but highlight different aspects of locomotion[51,52]. Scrutiny of dynamics in worm postural space showed that direction changes can be modelled as stochastic switches between attractors[53] and revealed a new behaviour (the delta turn) that previous investigators had missed[54]. Related analyses have been applied in larval zebrafish where the midline also captures much of the postural information[55].

Low dimensionality compared with the degree of postural freedom also holds for animals with more complex body plans. The dimensionality of reaching in humans and monkeys has been extensively studied and has led to work on muscle synergies and investigations into the neural control of reaching[56,57]. Interestingly, even octopuses, whose flexible limbs allow for still higher degrees of freedom seem to create 'joints' when reaching, potentially as a way of addressing the otherwise high dimensionality of the task[58].

That behaviour lies in low dimensional subspaces holds in both posture-spaces and posture-dynamics spaces. In the former, data points represent an animal's posture at a specific time[50]. In the latter, data points represent posture and its change

DISCRETISATION

behaviour is organised
into distinct modes

LOW DIMENSIONALITY

trajectories lie on
low-dimensional manifolds

STEREOTYPY

trajectories in behaviour-space
repeat precisely

HIERARCHY

behaviours are organised
on multiple scales

Figure 2 — Principles of the organisation of behaviour — Analyses of behaviour in multiple species using multiple approaches have pointed to four principles organising behaviour. Discretisation: behaviour occurs in distinct patterns that appear as clusters in representations of behaviour space. Low dimensionality: behaviour can be specified with fewer values than the morphological degrees of freedom available to the animal. Stereotypy: instances of behaviour within a distinct cluster can be strikingly similar in implementation. Hierarchy: behaviour is organised on multiple scales, and is naturally categorisable into behavioral states with sub-states (and sub-sub-states, etc.).

over time, as computed by time-frequency analysis[23,59] (Figure 2), or slicing recordings into segments of a particular length[60]. Interestingly, all of the major principles of the organisation of behaviour—low dimensionality, discretisation/stereotypy and hierarchy—emerge in analyses of both post-space and posture-dynamics space.

Discretisation and stereotypy

Behaviour is often thought of as a sequence of discrete actions that can be represented in an ethogram. An assumption at the heart of this approach is that animals perform some actions in ways that are similar enough to previous instances (and different enough from instances of other actions) to be discretely classifiable. Similarity across instances of an action is called stereotypy and it is the basis for several recent methods for segmenting behaviour into actions without the requirement of human-annotated sequences for training. An advantage of these unsupervised segmentations is that it is possible to discover actions that may have been difficult to detect initially by human observations.

Several methods have been proposed to detect stereotypy. In *C. elegans*, behavioural motifs were identified as precisely repeated segments in postural time series and their variation across hundreds of mutants revealed some underlying genetic relationships[60]. In *Drosophila* larvae, onset of

optogenetic neuronal stimulation was used to align trajectories and unsupervised structure learning detected distinct behavioural phenotypes[61]. In adult flies, parameterized high resolution video of spontaneous behaviour of adult *Drosophila* using time-frequency analysis of principal components of video frame pixels[23]. This method has since been extended to pairs of socially interacting animals[24].

Unsupervised methods have many methodological choice points and tuning parameters, and navigating these without reverting to supervised analysis is a conceptual challenge. One approach is to favour methods that produce behavioural annotations most consistent with axiomatic assumptions of what behaviour is[59], and this approach could be extended to choosing methods based on their adherence to physical rules or probabilistic models.

Reusing a finite set of well-adapted actions could be an efficient way of organising a behavioural repertoire, but too much stereotypy comes with costs. For example, when circumstances change, animals are able to adapt flexibly, performing rare or even unique bouts of activity as required. Compare, for example, the high stereotypy of a person walking down an empty corridor with the novel sequences of stops, starts, and skips that is observed when moving against the flow of a crowd. This may reflect a trade-off between efficient stereotyped behaviour and relatively inefficient but necessary flexible behaviour. A similar efficiency argument has been advanced to underlie Zipf's law[62], the power-law describing the rank-frequency distributions of word use[63] and heavy-tailed distributions of behaviours have also been observed in spontaneous worm behaviour[64]. However, in some circumstances at least, heavy-tailedness observed at a population level reflects heterogeneity among animals that are individually stereotyped[65,66].

Stereotypy is also intimately connected with predictability which can carry its own costs, especially in the presence of predators. Tentacle snakes take advantage of the predictability of escape responses to capture fish[67]. More generally, behavioural variability across or within individuals may reflect bet-hedging strategies to cope with unpredictability in the environment[68]. Neural circuits that increase noise above the constraints imposed by sensory information have been found to actively increase behavioural variability even in seemingly simple sensory pathways[69]. In humans, higher task variability is associated with faster motor learning[70].

Hierarchical organisation

Much like a language organised into phrases, sentences, paragraphs, and documents, hierarchical organisation has been proposed as a guiding principle in the analysis of behaviour[8,71]. It has been shown that hierarchical organisation can be an efficient means of using limited computation to produce complex adaptive behaviour in robotics[72], and the example of language has long been an important inspiration in the study of sequential behaviour[73], but clear demonstrations of hierarchical organisation in non-human animals have been lacking.

The simplest form of hierarchy is a linear 'peck order' in which higher level behaviours dominate lower level behaviours until they are complete. Such a suppression hierarchy

has been recently observed in grooming behaviour in fruit flies[74]. Nested representations with multiple levels of nesting can be constructed from action sequences using dictionary-based compression algorithms[75]. But, on its own, the compressibility of behavioural data using hierarchical algorithms does not imply that behaviour is organised hierarchically. Starting from the observation that transitions between behavioural states in flies are non-Markovian, Berman *et al.* have used a treeness metric to quantify the degree of hierarchy in flies' behavioural repertoire[76] and showed that a hierarchical representation is optimal for predicting the flies future behaviour with progressively coarser-grained representations predicting behaviours at longer time scales.

Conclusion

We have examined how new techniques are producing large data sets that have recently revealed a few high-level principles in the organisation of behaviour. But some of the most interesting directions for future research may be the codification or derivation of these principles from formal theory, the approach which has been so successful in physics. Given the increasingly precise and diverse datasets that are now available, there are ample opportunities to be inspired from behaving animals and to test models quickly. At the same time that we are learning more about how animals behave, physical ethology is being connected to other disciplines. In neuroscience, there is an increasing awareness that deep understanding of behaviour may be needed to understand brain-scale neural recordings[77–79]. In genetics, efforts to improve the characterisation of phenotypes have focused primarily on morphology but would benefit from improvements in behavioural quantification as well[80]. Ultimately, we hope the impact of physics on ethology will be akin to its impact on molecular biology, where ideas like the genetic code and techniques like X-ray crystallography have been key in understanding life.

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References

1. Tinbergen, N. On aims and methods of Ethology. *Zeitschrift für Tierpsychologie* **20,** 410–433 (2010).
2. Nathan, R. *et al.* A movement ecology paradigm for unifying organismal movement research. *Proceedings of the National Academy of Sciences* **105,** 19052–19059 (2008).
3. Vergassola, M., Villermaux, E. & Shraiman, B. I. 'Infotaxis' as a strategy for searching without gradients. *Nature* **445,** 406–409 (2007).
4. Bénichou, O., Loverdo, C., Moreau, M. & Voituriez, R. Intermittent search strategies. *Reviews of Modern Physics* **83,** 81–129 (2011).
5. Vicsek, T., Czirók, A., Ben-Jacob, E., Cohen, I. & Shochet, O. Novel Type of Phase Transition in a System of Self-Driven Particles. *Physical Review Letters* **75,** 1226–1229 (1995).
6. Bialek, W. *et al.* Social interactions dominate speed control in poising natural flocks near criticality. *Proceedings of the National Academy of Sciences* (2014).
7. Attanasi, A. *et al.* Finite-Size Scaling as a Way to Probe Near-Criticality in Natural Swarms. *Physical Review Letters* **113,** (2014).
8. Tinbergen, N. *The study of instinct*. (Clarendon Press; Oxford University Press, 1989).
9. Schrieffer, J. R. Macroscopic Quantum Phenomena from Pairing in Superconductors. *Science* **180,** 1243–1248 (1973).
10. Bender, J. A., Simpson, E. M. & Ritzmann, R. E. Computer-Assisted 3D Kinematic Analysis of All Leg Joints in Walking Insects. *PLoS ONE* **5,** e13617 (2010).
11. Schutz, C. & Durr, V. Active tactile exploration for adaptive locomotion in the stick insect. *Philosophical Transactions of the Royal Society B: Biological Sciences* **366,** 2996–3005 (2011).
12. Kain, J. *et al.* Leg-tracking and automated behavioural classification in Drosophila. *Nature Communications* **4,** 1910 (2013).
13. Risse, B. *et al.* FIM, a Novel FTIR-Based Imaging Method for High Throughput Locomotion Analysis. *PLoS ONE* **8,** e53963 (2013).
14. Mendes, C. S., Bartos, I., Akay, T., Márka, S. & Mann, R. S. Quantification of gait parameters in freely walking wild type and sensory deprived *Drosophila melanogaster*. *eLife* **2,** (2013).
15. Mendes, C. S. *et al.* Quantification of gait parameters in freely walking rodents. *BMC Biology* **13,** (2015).
16. Machado, A. S., Darmohray, D. M., Fayad, J., Marques, H. G. & Carey, M. R. A quantitative framework for whole-body coordination reveals specific deficits in freely walking ataxic mice. *eLife* **4,** (2015).
17. Uhlmann, V., Ramdya, P., Delgado-Gonzalo, R., Benton, R. & Unser, M. FlyLimbTracker: An active contour based approach for leg segment tracking in unmarked, freely behaving Drosophila. *PLOS ONE* **12,** e0173433 (2017).
18. Taylor, J. *et al.* Efficient and precise interactive hand tracking through joint, continuous optimization of pose and correspondences. *ACM Transactions on Graphics* **35,** 1–12 (2016).
19. Wei, S.-E., Ramakrishna, V., Kanade, T. & Sheikh, Y. Convolutional Pose Machines. in 4724–4732 (IEEE, 2016). doi:10.1109/CVPR.2016.511
20. Zimmermann, C. & Brox, T. Learning to Estimate 3D Hand Pose from Single RGB Images. *arXiv:1705.01389* (2017).
21. Burghardt, G. M. *et al.* Perspectives - Minimizing Observer Bias in Behavioral Studies: A Review and Recommendations: Minimizing Observer Bias in Behavioral Research. *Ethology* **118,** 511–517 (2012).
22. Szigeti, B., Stone, T. & Webb, B. Inconsistencies in behavioural annotation. (2016). doi:10.1101/066787
23. Berman, G. J., Choi, D. M., Bialek, W. & Shaevitz, J. W. Mapping the stereotyped behaviour of freely moving fruit flies. *Journal of The Royal Society Interface* **11,** 20140672–20140672 (2014).
24. Klibaite, U., Berman, G. J., Cande, J., Stern, D. L. & Shaevitz, J. W. An unsupervised method for quantifying the behavior of paired animals. *Physical Biology* **14,** 15006 (2017).
25. Geissmann, Q. *et al.* Ethoscopes: An open platform for high-throughput ethomics. *PLOS Biology* **15,** e2003026 (2017).
26. Swierczek, N. A., Giles, A. C., Rankin, C. H. & Kerr, R. A. High-throughput behavioral analysis in C. elegans. *Nature Methods* **8,** 592–598 (2011).
27. Kocabas, A., Shen, C.-H., Guo, Z. V. & Ramanathan, S. Controlling interneuron activity in Caenorhabditis elegans to evoke chemotactic behaviour. *Nature* **490,** 273–277 (2012).
28. Stirman, J. N. *et al.* Real-time multimodal optical control of neurons and muscles in freely behaving Caenorhabditis elegans. *Nature Methods* **8,** 153–158 (2011).
29. Leifer, A. M., Fang-Yen, C., Gershow, M., Alkema, M. J. & Samuel, A. D. T. Optogenetic manipulation of neural activity in freely moving Caenorhabditis elegans. *Nature Methods* **8,** 147–152 (2011).
30. Kim, D. H. *et al.* Pan-neuronal calcium imaging with cellular resolution in freely swimming zebrafish. *Nature Methods* **14,** 1107–1114 (2017).
31. Portugues, R. & Engert, F. Adaptive Locomotor Behavior in Larval Zebrafish. *Frontiers in Systems Neuroscience* **5,** (2011).
32. Gaudry, Q., Hong, E. J., Kain, J., de Bivort, B. L. & Wilson, R. I. Asymmetric neurotransmitter release enables rapid odour lateralization in Drosophila. *Nature* **493,** 424–428 (2012).
33. Corrales-Carvajal, V. M., Faisal, A. A. & Ribeiro, C. Internal states drive nutrient homeostasis by modulating exploration-exploitation trade-off. *eLife* **5,** (2016).

34. Churgin, M. A. *et al.* Longitudinal imaging of Caenorhabditis elegans in a microfabricated device reveals variation in behavioral decline during aging. *eLife* **6,** (2017).

35. Jordan, D., Kuehn, S., Katifori, E. & Leibler, S. Behavioral diversity in microbes and low-dimensional phenotypic spaces. *Proceedings of the National Academy of Sciences* **110,** 14018–14023 (2013).

36. Ayroles, J. F. *et al.* Behavioral idiosyncrasy reveals genetic control of phenotypic variability. *Proceedings of the National Academy of Sciences* **112,** 6706–6711 (2015).

37. Buchanan, S. M., Kain, J. S. & de Bivort, B. L. Neuronal control of locomotor handedness in *Drosophila. Proceedings of the National Academy of Sciences* **112,** 6700–6705 (2015).

38. Pittman, W. E., Sinha, D. B., Zhang, W. B., Kinser, H. E. & Pincus, Z. A simple culture system for long-term imaging of individual C. elegans. *Lab Chip* (2017). doi:10.1039/C7LC00916J

39. Kafkafi, N., Mayo, C. L. & Elmer, G. I. Mining mouse behavior for patterns predicting psychiatric drug classification. *Psychopharmacology* **231,** 231–242 (2014).

40. Wiltschko, A. B. *et al.* Mapping Sub-Second Structure in Mouse Behavior. *Neuron* **88,** 1121–1135 (2015).

41. Goulding, E. H. *et al.* A robust automated system elucidates mouse home cage behavioral structure. *Proceedings of the National Academy of Sciences* **105,** 20575–20582 (2008).

42. Jhuang, H. *et al.* Automated home-cage behavioural phenotyping of mice. *Nature Communications* **1,** 1–9 (2010).

43. Shemesh, Y. *et al.* High-order social interactions in groups of mice. *eLife* **2,** e00759–e00759 (2013).

44. Hong, W. *et al.* Automated measurement of mouse social behaviors using depth sensing, video tracking, and machine learning. *Proceedings of the National Academy of Sciences* **112,** E5351–E5360 (2015).

45. Maia Chagas, A., Prieto-Godino, L. L., Arrenberg, A. B. & Baden, T. The €100 lab: A 3D-printable open-source platform for fluorescence microscopy, optogenetics, and accurate temperature control during behaviour of zebrafish, Drosophila, and Caenorhabditis elegans. *PLOS Biology* **15,** e2002702 (2017).

46. Branson, K., Robie, A. A., Bender, J., Perona, P. & Dickinson, M. H. High-throughput ethomics in large groups of Drosophila. *Nature Methods* **6,** 451–457 (2009).

47. Kabra, M., Robie, A. A., Rivera-Alba, M., Branson, S. & Branson, K. JAABA: interactive machine learning for automatic annotation of animal behavior. *Nature Methods* **10,** 64–67 (2012).

48. Pérez-Escudero, A., Vicente-Page, J., Hinz, R. C., Arganda, S. & de Polavieja, G. G. idTracker: tracking individuals in a group by automatic identification of unmarked animals. *Nature Methods* **11,** 743–748 (2014).

49. Gattermann, R. *et al.* Golden hamsters are nocturnal in captivity but diurnal in nature. *Biology Letters* **4,** 253–255 (2008).

50. Stephens, G. J., Johnson-Kerner, B., Bialek, W. & Ryu, W. S. Dimensionality and Dynamics in the Behavior of C. elegans. *PLoS Computational Biology* **4,** e1000028 (2008).

51. Hums, I. *et al.* Regulation of two motor patterns enables the gradual adjustment of locomotion strategy in *Caenorhabditis elegans. eLife* **5,** (2016).

52. Gyenes, B. & Brown, A. E. X. Deriving Shape-Based Features for C. elegans Locomotion Using Dimensionality Reduction Methods. *Frontiers in Behavioral Neuroscience* **10,** (2016).

53. Stephens, G. J., Bueno de Mesquita, M., Ryu, W. S. & Bialek, W. Emergence of long timescales and stereotyped behaviors in Caenorhabditis elegans. *Proceedings of the National Academy of Sciences* **108,** 7286–7289 (2011).

54. Broekmans, O. D., Rodgers, J. B., Ryu, W. S. & Stephens, G. J. Resolving coiled shapes reveals new reorientation behaviors in *C. elegans. eLife* **5,** (2016).

55. Girdhar, K., Gruebele, M. & Chemla, Y. R. The Behavioral Space of Zebrafish Locomotion and Its Neural Network Analog. *PLOS ONE* **10,** e0128668 (2015).

56. Wolpert, D. M. & Ghahramani, Z. Computational principles of movement neuroscience. *Nat. Neurosci.* **3 Suppl,** 1212–1217 (2000).

57. Flash, T. & Hochner, B. Motor primitives in vertebrates and invertebrates. *Current Opinion in Neurobiology* **15,** 660–666 (2005).

58. Sumbre, G., Fiorito, G., Flash, T. & Hochner, B. Octopuses Use a Human-like Strategy to Control Precise Point-to-Point Arm Movements. *Current Biology* **16,** 767–772 (2006).

59. Todd, J. G., Kain, J. S. & de Bivort, B. L. Systematic exploration of unsupervised methods for mapping behavior. *Physical Biology* **14,** 15002 (2017).

60. Brown, A. E. X., Yemini, E. I., Grundy, L. J., Jucikas, T. & Schafer, W. R. A dictionary of behavioral motifs reveals clusters of genes affecting Caenorhabditis elegans locomotion. *Proc. Natl. Acad. Sci. U.S.A.* **110,** 791–796 (2013).

61. Vogelstein, J. T. *et al.* Discovery of brainwide neural-behavioral maps via multiscale unsupervised structure learning. *Science* **344,** 386–392 (2014).

62. Salge, C., Ay, N., Polani, D. & Prokopenko, M. Zipf's Law: Balancing Signal Usage Cost and Communication Efficiency. *PLOS ONE* **10,** e0139475 (2015).

63. Gerlach, M. & Altmann, E. G. Stochastic Model for the Vocabulary Growth in Natural Languages. *Physical Review X* **3,** (2013).

64. Schwarz, R. F., Branicky, R., Grundy, L. J., Schafer, W. R. & Brown, A. E. X. Changes in Postural Syntax Characterize Sensory Modulation and Natural Variation of C. elegans Locomotion. *PLOS Computational Biology* **11,** e1004322 (2015).

65. Petrovskii, S., Mashanova, A. & Jansen, V. A. A. Variation in individual walking behavior creates the impression of a Levy flight. *Proceedings of the National Academy of Sciences* **108,** 8704–8707 (2011).

66. Kain, J. S., Stokes, C. & de Bivort, B. L. Phototactic personality in fruit flies and its suppression by serotonin and white. *Proceedings of the National Academy of Sciences* **109,** 19834–19839 (2012).

67. Catania, K. C. Born Knowing: Tentacled Snakes Innately Predict Future Prey Behavior. *PLoS ONE* **5,** e10953 (2010).

68. Kain, J. S. *et al.* Variability in thermal and phototactic preferences in *Drosophila* may reflect an adaptive bet-hedging strategy. *Evolution* **69,** 3171–3185 (2015).

69. Gordus, A., Pokala, N., Levy, S., Flavell, S. W. & Bargmann, C. I. Feedback from Network States Generates Variability in a Probabilistic Olfactory Circuit. *Cell* **161,** 215–227 (2015).

70. Wu, H. G., Miyamoto, Y. R., Castro, L. N. G., Ölveczky, B. P. & Smith, M. A. Temporal structure of motor variability is dynamically regulated and predicts motor learning ability. *Nature Neuroscience* **17,** 312–321 (2014).

71. Dawkins, R. Hierarchical organisation: a candidate principle for ethology. in *Growing Points in Ethology* 7–54 (Cambridge University Press, 1976).

72. Barto, A. G. & Mahadevan, S. Recent Advances in Hierarchical Reinforcement Learning. *Discrete Event Dynamic Systems* **13,** 341–379 (2003).

73. Lashley, K. S. The problem of serial order in behavior. in *Cerebral Mechanisms in Behavior* (ed. Jeffress, L. A.) (John Wiley Press, 1951).

74. Seeds, A. M. *et al.* A suppression hierarchy among competing motor programs drives sequential grooming in Drosophila. *Elife* **3,** e02951 (2014).

75. Gomez-Marin, A., Stephens, G. J. & Brown, A. E. X. Hierarchical compression of *Caenorhabditis elegans* locomotion reveals phenotypic differences in the organization of behaviour. *Journal of The Royal Society Interface* **13,** 20160466 (2016).

76. Berman, G. J., Bialek, W. & Shaevitz, J. W. Predictability and hierarchy in *Drosophila* behavior. *Proceedings of the National Academy of Sciences* **113,** 11943–11948 (2016).

77. Krakauer, J. W., Ghazanfar, A. A., Gomez-Marin, A., MacIver, M. A. & Poeppel, D. Neuroscience Needs Behavior: Correcting a Reductionist Bias. *Neuron* **93,** 480–490 (2017).

78. Gomez-Marin, A., Paton, J. J., Kampff, A. R., Costa, R. M. & Mainen, Z. F. Big behavioral data: psychology, ethology and the foundations of neuroscience. *Nature Neuroscience* **17,** 1455–1462 (2014).

79. Calhoun, A. J. & Murthy, M. Quantifying behavior to solve sensorimotor transformations: advances from worms and flies. *Current Opinion in Neurobiology* **46,** 90–98 (2017).

80. Houle, D. Numbering the hairs on our heads: The shared challenge and promise of phenomics. *Proceedings of the National Academy of Sciences* **107,** 1793–1799 (2010).