Seasonal temperature variation influences climate suitability for dengue,

chikungunya, and Zika transmission

John H. Huber[1, 2], Marissa L. Childs[3], Jamie M. Caldwell[2], Erin A. Mordecai[2*]

[1]Department of Applied and Computational Mathematics and Statistics, University

of Notre Dame, Notre Dame, Indiana, USA

[2]Department of Biology, Stanford University, Stanford, California, USA

[3]Emmett Interdisciplinary Program in Environment and Resources, Stanford

University, Stanford, California, USA

*Corresponding Author: emordeca@stanford.edu

Short title: Seasonal climate affects vector transmission

Abstract

Dengue, chikungunya, and Zika virus epidemics transmitted by *Aedes aegypti*
mosquitoes have recently (re)emerged and spread throughout the Americas,
Southeast Asia, the Pacific Islands, and elsewhere. Understanding how
environmental conditions affect epidemic dynamics is critical for predicting and
responding to the geographic and seasonal spread of disease. Specifically, we lack a
mechanistic understanding of how seasonal variation in temperature affects
epidemic magnitude and duration. Here, we develop a dynamic disease
transmission model for dengue virus and *Aedes aegypti* mosquitoes that integrates
mechanistic, empirically parameterized, and independently validated mosquito and
virus trait thermal responses under seasonally varying temperatures. We examine
the influence of seasonal temperature mean, variation, and temperature at the start
of the epidemic on disease dynamics. We find that at both constant and seasonally
varying temperatures, warmer temperatures at the start of epidemics promote
more rapid epidemics due to faster burnout of the susceptible population. By
contrast, intermediate temperatures (24-25°C) at epidemic onset produced the
largest epidemics in both constant and seasonally varying temperature regimes.
When seasonal temperature variation was low, 25-35°C annual average
temperatures produced the largest epidemics, but this range shifted to cooler
temperatures as seasonal temperature variation increased (analogous to previous
results for diurnal temperature variation). Tropical and sub-tropical cities such as
Rio de Janeiro, Fortaleza, and Salvador, Brazil; Cali, Cartagena, and Barranquilla,
Colombia; Delhi, India; Guangzhou, China; and Manila, Philippines have mean annual

temperatures and seasonal temperature ranges that produced the largest

epidemics. However, more temperate cities like Shanghai, China had high epidemic

suitability because large seasonal variation offset moderate annual average

temperatures. By accounting for seasonal variation in temperature, the model

provides a baseline for mechanistically understanding environmental suitability for

virus transmission by *Aedes aegypti*. Overlaying the impact of human activities and

socioeconomic factors onto this mechanistic temperature-dependent framework is

critical for understanding likelihood and magnitude of outbreaks.

Non-Technical Summary (150-200 Words)

Mosquito-borne viruses like dengue, Zika, and chikungunya have recently caused

large epidemics that are partly driven by temperature. Using a mathematical model

built from laboratory experimental data for *Aedes aegypti* mosquitoes and dengue

virus, we examine the impact of variation in seasonal temperature regimes on

epidemic size and duration. At constant temperatures, both low and high

temperatures (20°C and 35°C) produce small epidemics, while intermediate

temperatures like 25°C and 30°C produce much larger epidemics. In seasonally

varying temperature environments, epidemics peak more rapidly at higher starting

temperatures, while intermediate starting temperatures produce the largest

epidemics. Seasonal mean temperatures of 25-35°C are most suitable for large

epidemics when seasonality is low, but in more variable seasonal environments

epidemic suitability peaks at lower annual average temperatures. Tropical and sub-

tropical cities have the highest temperature suitability for epidemics, but more

temperate cities with high seasonal variation also have the potential for very large

epidemics.

Introduction

Over the last 30-40 years, arboviral outbreaks have dominated the public health

landscape globally [1]. These viruses, most notably dengue (DENV), chikungunya

(CHIKV), and Zika (ZIKV), can cause symptoms ranging from rash, arthralgia, and

fever to hemorrhagic fever (DENV), long-term arthritis (CHIKV), Guillain-Barré

syndrome and microcephaly (ZIKV) [2–4]. DENV, which historically spread

worldwide along shipping routes [5], places 3.97 billion individuals at risk

worldwide [6] and causes an estimated 390 million cases annually, including 96

million symptomatic cases [7]. CHIKV was introduced into the Americas in

December 2013 after an outbreak in St. Martin Island [8]. Since then, autochthonous

transmission has been reported in 45 countries [9], and 1.3 billion people

worldwide are at risk of contracting CHIKV [10]. More recently, the ZIKV epidemic

in the Americas captured global attention after the World Health Organization

(WHO) designated it a Public Health Emergency of International Concern in

February 2016 in response to its association with neurological disorders. Following

the first reported case in Brazil in May 2015, ZIKV has spread to 48 countries and

territories where it is transmitted autochthonously [11]. Because DENV, CHIKV, and

ZIKV are mostly transmitted by *Aedes aegypti* mosquitoes, they may have similar

geographic distributions and risk factors.

Informed public health decisions to limit the spread and magnitude of these

arboviral epidemics depend on a robust understanding of transmission dynamics.

One mechanistic modeling framework, the Susceptible – Infected – Recovered (SIR)

model, has been implemented successfully to model the dynamics of outbreaks of

influenza, measles, and vector-borne diseases such as CHIKV and ZIKV [12–14]. This

approach tracks virus population dynamics by compartmentalizing individuals by

their state in an epidemic (i.e., Susceptible (S), Infected (I), Recovered (R)). This

framework can be extended to include additional compartments, such as a latency

stage, or to incorporate the dynamics of the mosquito population for vector

transmission.

Arbovirus dynamics are strikingly seasonal and geographically restricted to

relatively warm climates [6,7]. This arises because several life history traits of the

mosquitoes that transmit DENV, CHIKV, and ZIKV are strongly influenced by

temperature and seasonality [15–22]. For simplicity, many existing models assume

static life history traits [14], and those that address seasonal forcing tend to

incorporate sinusoidal variation as a single transmission parameter, β [23]. Treating

seasonal temperature variation as a sinusoidal forcing function on the transmission

parameter implies a monotonic relationship between temperature and

transmission, such that transmission is maximized at high temperatures and

decreases at low temperatures. However, decades of experimental work have

demonstrated strongly nonlinear (often unimodal) relationships between mosquito

and pathogen traits and temperature that are not well captured in a single

sinusoidal forcing function [24]. Efforts by Yang et al. [25,26] addressed the need to

include seasonal variation by adopting an SEI-SEIR compartmental framework with

time-varying entomological parameters and fitting the model to DENV incidence

data in Campinas, Brazil. Other previous work has integrated the effects of

temperature on mosquito and parasite traits into temperature-dependent

transmission models for DENV, CHIKV, and/or ZIKV, and revealing a strong,

nonlinear influence of temperature with peak transmission between 29 – 35 °C [27–

34]. However, we do not yet have a mechanistic estimate for the relationship

between seasonal temperature regimes and transmission potential, incorporating

the full suite of transmission-relevant, nonlinear thermal responses of mosquito and

parasite traits.

Here, we expand on previous work with three main advances: (1) we

incorporate the full suite of empirically-derived, unimodal thermal responses for all

known transmission-relevant mosquito and parasite traits; (2) we examine the

influence of seasonal temperature mean and variation (in contrast to constant

temperatures or daily temperature variation); and (3) we use a dynamic

transmission framework to explore the impact of different seasonal temperature

regimes on the epidemiologically-relevant outcomes of epidemic size, duration, and

peak incidence (in contrast to R_0, or vectorial capacity, which are difficult to

measure directly). To do so, we incorporate previously estimated and independently

validated thermal response functions for all vector and parasite traits [24] into a

dynamic SEI-SEIR model [25,26]. We explore field-relevant temperature regimes by

simulating epidemics across temperature means (10 – 38°C) and seasonal ranges (0

– 17°C) from across the predicted suitable range for transmission. Specifically, we

use the model to ask: (1) How does final epidemic size vary across constant

temperatures? (2) Under seasonally varying temperatures, how does the

temperature at the start of the epidemic affect the final epidemic size and duration?

(3) How do temperature mean and seasonal range interact to determine epidemic

size? (4) Which geographic locations have high epidemic suitability based on

climate?

Methods

Model

Model Framework

We adopted an SEI-SEIR compartmental modeling framework to simulate arboviral

transmission by the *Aedes aegypti* vector (Fig. 1). We introduced temperature-

dependence into the model by using fitted thermal response curves for the

mosquito life history traits provided by Mordecai et al. [24]. The full model is:

142
$$\frac{dS_V}{dt} = EFD(T) * pEA(T) * MDR(T) * \mu(T)^{-1} * N_V * \left(1 - \frac{N_V}{K(T)}\right) - \left(a(T) * pMI(T) * \frac{I_H}{N_H} + \mu(T)\right) * S_V, \quad (1)$$

144
$$\frac{dE_V}{dt} = a(T) * pMI(T) * \frac{I_H}{N_H} * S_V - \left(PDR(T) + \mu(T)\right) * E_V, \quad (2)$$

146
$$\frac{dI_V}{dt} = PDR(T) * E_V - \mu(T) * I_V, \quad (3)$$

148
$$\frac{dS_H}{dt} = -a(T) * b(T) * \frac{I_V}{N_H} * S_H, \quad (4)$$

150
$$\frac{dE_H}{dt} = a(T) * b(T) * \frac{I_V}{N_H} * S_H - \delta * E_H, \quad (5)$$

152
$$\frac{dI_H}{dt} = \delta * E_H - \eta * I_H, \quad (6)$$

154
$$\frac{dR_H}{dt} = \eta * I_H, \quad (7)$$

Fig 1. Compartmental model of transmission. S_H, E_H, I_H, and R_H represent the

susceptible, exposed (or latent), infectious, and recovered segments of the human

population, respectively. Likewise, S_V, E_V, and I_V represent the susceptible, exposed

(or latent), and infectious segments of the mosquito population. Solid arrows signify

the directionality of transition from one compartment to the next, and dashed

arrows indicate the directionality of transmission.

The SEI portion of the model describes the vector population, where S_V

represents the number of susceptible mosquitoes, E_V is the number of mosquitoes in

the latency stage, and I_V is the number of infectious mosquitoes. We assumed that

Aedes aegypti mosquitoes remain infectious until they die. In equations 1-3, *(T)*

indicates temperature-dependent functions, *EFD(T)* is the number of eggs laid per

female per day, *pEA(T)* is the probability of mosquito egg-to-adult survival, *MDR(T)*

is the mosquito egg-to-adult development rate, N_V is the total mosquito population

at time t (i.e., $S_v + E_v + I_v$), *K(T)* is the carrying capacity for the mosquito population,

a(T) is the per mosquito biting rate, *pMI(T)* is the probability of mosquito infection

per bite on an infectious host, *μ(T)* is the adult mosquito mortality rate, and *PDR(T)*

is the parasite development rate. Each life history and pathogen transmission trait

of the *Aedes aegypti* mosquito is a unimodal, temperature-dependent function fit

from experimental laboratory data in previous work [15–22,24] (Table 1; Appendix;

"Functional Forms of Life History Traits").

Table 1. Fitted thermal responses for *Aedes aegypti* life history traits. Traits were fit

to a Brière $[cT(T - T_0)(T_m - T)^{\frac{1}{2}}]$ or a quadratic $[c(T - T_m)(T - T_0)]$ function where

T represents temperature. T_0 and T_m are the critical thermal minimum and

maximum, respectively, and c is the rate constant. Thermal responses were fit by

[24].

Trait	Definition	Function	Fitted Parameters		
a	Biting rate (day⁻¹)	Brière	$c = 2.02\text{e-}04$	$T_{min} = 13.35$	$T_{max} = 40.08$
EFD	Eggs laid per female per day	Brière	$c = 8.56\text{e-}03$	$T_{min} = 14.58$	$T_{max} = 34.61$
pEA	Probability of mosquito egg-to-adult survival	Quadratic	$c = -5.99\text{e-}03$	$T_{min} = 13.56$	$T_{max} = 38.29$
MDR	Mosquito egg-to-adult development rate (day⁻¹)	Brière	$c = 7.86\text{e-}05$	$T_{min} = 11.36$	$T_{max} = 39.17$
lf	Adult mosquito lifespan (days)	Quadratic	$c = -1.48\text{e-}01$	$T_{min} = 9.16$	$T_{max} = 37.73$
b	Probability of mosquito infectiousness	Brière	$c = 8.49\text{e-}04$	$T_{min} = 17.05$	$T_{max} = 35.83$
pMI	Probability of mosquito infection	Brière	$c = 4.91\text{e-}04$	$T_{min} = 12.22$	$T_{max} = 37.46$
PDR	Virus extrinsic incubation rate (day⁻¹)	Brière	$c = 6.65\text{e-}05$	$T_{min} = 10.68$	$T_{max} = 45.90$

The SEIR portion of the model describes the human population, where S_H

represents the number of susceptible individuals, E_H the number of latent (or

exposed) individuals, I_H the number of infectious individuals, and R_H the number of

recovered individuals. We assumed a static population size, N_H, that was neither

subject to births nor deaths because the human lifespan far exceeds the duration of

an epidemic. Further, we binned asymptomatic and symptomatic individuals into a

single infectious class since asymptomatic infections have been shown to transmit

DENV [35] and exhibit similar viremic profiles as symptomatic patients in CHIKV

[36]. Based on previous arboviral outbreaks [37,38], we assumed that an infection

conferred long-term immunity to an individual. Thus, a previously infectious

individual entering the recovered class is protected from subsequent re-infection

for the remainder of the epidemic. In the case of dengue, where there are four

unique serotypes, we consider single-season epidemics of a single serotype. In

equations 4-7, *b(T)* is the probability of human infection per bite by an infectious

mosquito (Table 1), δ^{-1} is the intrinsic incubation period, and η^{-1} is the human

infectivity period. Since human components of the transmission cycle are not

seasonal, we used constants of 5.9 days for the intrinsic incubation period, $1/\delta$, and

5.0 days for the infectious period, $1/\eta$ [14]. All temperature-independent parameter

values are given in Table 2.

Table 2. Values of temperature-independent parameters used in the model, and

their sources.

Parameter	Definition	Value	Source
δ^{-1}	Intrinsic incubation period (days)	5.9	[14]
η^{-1}	Human infectivity period (days)	5.0	[14]
I_0^H / N	Proportion of initially infectious humans	0.0001	
I_0^V / M	Proportion of initially infectious mosquitoes	0.015	[14]

M/N Ratio of mosquitoes-to-humans at 29°C   2.0   [39]

Since the lifespan of an adult mosquito is short relative to the timespan of an

epidemic, we allowed mosquito birth and death rates to drive population dynamics.

Additionally, the birth rate of susceptible mosquitoes was regulated by a

temperature-dependent carrying capacity, K (equation 8), which we modeled as a

modified Arrhenius equation that is a unimodal function of temperature [40]:

$$K(T) = \frac{EFD(T_0) * pEA(T_0) * MDR(T_0) * \mu(T_0)^{-1} - \mu(T_0)}{EFD(T_0) * pEA(T_0) * MDR(T_0) * \mu(T_0)^{-1}} * N_m * e^{\frac{-E_A*(T-T_0)^2}{\kappa_B*(T+273)*(T_0+273)}}, \quad (8)$$

Here, T_0 is defined as the reference temperature (i.e., the temperature at

which the carrying capacity is greatest) in Celsius, N_m is the maximum carrying

capacity, and κ_B is Boltzmann constant (8.617 x 10^{-5} eV/K). EFD is the number of

eggs laid per female per day, pEA is the probability of egg-to-adult mosquito

survival, MDR is the mosquito egg-to-adult development rate, and μ is the adult

mosquito mortality rate. We calculated these values for the reference temperature.

E_A is the activation energy, which we set to 0.5 and represents the temperature

dependence of the carrying capacity, a conservative estimate as we lacked sufficient

data on estimates of the carrying capacity of *Aedes aegypti* and its underlying

temperature dependence. To convert from Celsius to Kelvin, we incremented the

temperature T and the reference temperature T_0 by 273. Equation (8) was adopted

from [40] and modified to allow the distribution to be unimodal. We set the

reference temperature, T_0, to 29°C, which is consistent with optimal temperatures

for *Aedes aegypti* transmission [24,29].

We included a temperature-dependent carrying capacity in the model to

constrain the growth of the mosquito population. As described in the Appendix, all

simulations begin with the mosquito population at its (temperature-dependent)

carrying capacity. As the temperature changes seasonally, the mosquito population

does not necessarily remain at carrying capacity if one or more of the life history

traits that determine the production of new mosquitoes in equation (1)—*EFD*, *pEA*,

and *MDR*—is equal to zero. This occurs below 14.58°C (the highest T_{min} of *EFD*, *pEA*,

and *MDR*) or above 34.61°C (the lowest T_{max} of *EFD*, *pEA*, and *MDR*).

It should be noted that the transmission parameters are only related to the

current temperature at each time point in the simulation. Time lags for each life

history trait were not explicitly built into the model.

Seasonal Forcing

To address seasonality in the model, we allowed temperature to vary over

time. We modeled temperature as a sinusoidal curve with a period of 365 days of

the form:

$$T(t) = \frac{T_{max} - T_{min}}{2} * \sin\left(\frac{2\pi}{365}t\right) + T_{mean}, \quad (9)$$

Here, T_{max}, T_{mean}, and T_{min} represent the average monthly maximum, mean, and

minimum temperatures across a calendar year, respectively, and t is measured in

days. By modeling temperature as a function of time, we allowed the life history

traits of the *Aedes aegypti* vector to vary across time for the duration of the

epidemic. In the absence of a specific focal location we modeled seasonal

temperature as a sinusoidal function for simplicity.

Data

Life History Traits

To incorporate seasonal forcing into the compartmental modeling framework, we

used fitted mechanistic thermal response curves [24]. Mordecai et al. [24] examined

published data on thermal responses for life history traits of the *Aedes aegypti*

vector and DENV and adopted a Bayesian approach for fitting quadratic ($Q(T)$; Eq.

10) or Brière ($B(T)$; Eq. 11) curves (see Appendix for details).

$$Q(T) = c * (T - T_{min}) * (T - T_{max}), \quad (10)$$

$$B(T) = c * T * (T - T_{min}) * \sqrt{T_{max} - T}, \quad (11)$$

Here, c is a rate constant, T_{min} is the critical temperature minimum, and T_{max} is the

critical temperature maximum (Table 1). Following Mordecai et al. [24], we

assumed values above the critical thermal maxima and below the minima were

equal to zero.

273   Mordecai et al. [24] fit the thermal response for adult mosquito lifespan

274   (Table 1), the inverse of the adult mosquito mortality rate (μ, in days^{-1}), used in our

275   model. We set the mortality rate at temperatures outside the critical thermal

276   minimum and maximum to 24 days^{-1} (i.e., mosquitoes survive for one hour at

277   temperatures outside of the T_{min} to T_{max} range).

279   Historical Weather Data

280   To identify areas of epidemic suitability across the globe, we extracted

monthly mean temperatures for 2016 from Weather Underground

(wunderground.com) for twenty different cities (Table 3). For each city, we

calculated the mean, minimum, and maximum from the average monthly mean

temperatures, to estimate temperature seasonality. This provided a range of the

average monthly temperature over the span of a calendar year. We chose this time

period because it provided the most recent full calendar year to demonstrate

seasonal variation in temperature.

Table 3. Temperature regimes for major cities during the 2016 calendar year.

Monthly mean temperatures during 2016 were extracted from Weather

Underground.

City	Annual Mean Temperature (°C)	Annual Temperature Amplitude (°C)
Buenos Aires, Argentina	16.5	8.0
Sao Paulo, Brazil	20.6	5.0

Rio de Janeiro, Brazil	24.3	4.0
Salvador, Brazil	26.3	2.0
Fortaleza, Brazil	27.8	0.50
Belo Horizonte, Brazil	21.9	3.0
Recife, Brazil	27.2	1.5
Shanghai, China	17.6	12.5
Beijing, China	12.8	16
Guangzhou, China	22.9	8.0
Bogotá, Colombia	14.7	1.0
Medellin, Colombia	17.9	1.0
Cali, Colombia	25.1	1.5
Barranquilla, Colombia	28.8	1.0
Cartagena, Colombia	28.6	1.0
Delhi, India	26.3	9.5
Tokyo, Japan	17.0	10.5
Kobe, Japan	17.4	11
Manila, Philippines	29.0	1.5
New York, USA	13.8	12

Variability in Epidemic Dynamics with Constant Temperature

We first examined how epidemic dynamics varied across different constant

temperatures. Here, we did not introduce seasonal forcing into the model but rather

assumed static life history traits for *Aedes aegypti* for the simulation period. We

simulated the model under default starting conditions (see Appendix) at four

different constant temperatures: 20°C, 25°C, 30°C, and 35°C. These temperatures

were chosen to span the range of temperatures at which arbovirus transmission is

likely to be possible [24].

Variability in Epidemic Dynamics with Starting Temperature

Using the model that included seasonal variation in temperature, we examined how

the dynamics of an epidemic varied due to the temperature at which the epidemic

began, under two temperature regimes. First, we set T_{max} = 40.0°C, T_{mean} = 25.0°C,

and T_{min} = 10.0°C in the time-varying seasonal temperature model under default

parameters (see Appendix) and varied the temperature at the start of the epidemic

from 10.0°C to 40.0°C in increments of 0.1°C. We examined the response of final

epidemic size, epidemic length, and maximum instantaneous number of infected

individuals. We then repeated this process for a regime with a lower magnitude of

seasonal temperature variation: T_{max} = 30.0°C, T_{mean} = 25.0°C, and T_{min} = 20.0°C. By

comparing these temperature regimes, we can examine how epidemics respond to

starting temperatures that are outside the range of plausible temperatures of

arbovirus transmission (regime 1) versus restricted to the plausible temperatures

for transmission (regime 2) [24].

Seasonal Variability of Final Epidemic Size

Using the compartmental modeling framework with the default starting conditions,

we examined the variation in final epidemic size as a result of seasonal forcing. To

do so, we simulated over a wide range of temperature mean and seasonal variance

regimes. The mean annual temperature varied from 10.0°C to 38.0°C in increments

of 0.1°C, while the seasonal variation about the mean (i.e., $\frac{T_{max}-T_{min}}{2}$) ranged from

0.0°C to 17.0°C in increments of 0.1°C. Many of these temperature regimes are

unlikely to be observed empirically. However, the simulated temperature regimes

spanned the full range of feasible temperature conditions. We recorded the final

epidemic size, measured as the number of individuals in the recovered

compartment at the end of the simulation, for each unique combination of mean

annual temperature and seasonal variation. In addition, we examined the effect of

epidemic starting temperature on final epidemic size across the same seasonal

temperature regimes. We ran the model under default starting conditions, but

allowed the starting temperature to equal T_{min}, T_{mean}, or T_{max}.

To observe the interaction of population immunity with the seasonal

temperature regime, we simulated the model assuming that 0, 20, 40, 60, or 80% of

the population was initially immune. Each simulation began with the introduction

of the infected individual occurring at the mean seasonal temperature.

We then compared simulated climate regimes with actual climates in major

cities, to measure relative epidemic suitability of the following cities: São Paulo,

Brazil; Rio de Janeiro, Brazil; Salvador, Brazil; Fortaleza, Brazil; Belo Horizonte,

Brazil; Recife, Brazil; Bogotá, Colombia; Medellín, Colombia; Cali, Colombia;

Barranquilla, Colombia; Cartagena, Colombia; Tokyo, Japan; Delhi, India; Manila,

Philippines; Shanghai, China; Beijing, China; New York City, USA; Guangzhou, China;

Kobe, Japan; and Buenos Aires, Argentina, given 0, 20, 40, 60, and 80% population

immunity. These cities were chosen because they represent some of the most

populous urban areas across South America and throughout the world.

Model Sensitivity and Uncertainty Analysis

To characterize uncertainty in the model, we sampled 50 joint posterior estimates

for c, T_{min}, and T_{max} for each life history trait provided by Mordecai et al. [24]. We

examined the variability in epidemic dynamics with starting temperatures under

each parameterization and report the 95% credible interval for the epidemiological

indices. We similarly characterize uncertainty in our estimates of the final epidemic

size as a function of the seasonal temperature regime by simulating under each

parameterization and reporting the 95% credible interval.

Results

Variability in Epidemic Dynamics with Constant Temperature

Holding temperature constant, we examined variability in epidemic dynamics

across four temperatures: 20°C, 25°C, 30°C, and 35°C. As temperature increased

from 20°C to 30°C, the number of susceptible individuals depleted more rapidly

(Fig. 2, S_H). At 20°C and 35°C, the epidemics were small (1.33% and 5.92% of the

population infected, respectively) and burned out rapidly. Although simulations run

at 25°C and 30°C produced final epidemic sizes of 94.73% and 99.98% of the

population infected, respectively (Fig. 2, R_H), the epidemic peaked much faster at

30°C.

Fig 2. Variation in epidemic dynamics by temperature. The model was simulated

under default parameters at four constant temperatures: 20°C, 25°C, 30°C, and 35°C.

Variability in Epidemic Dynamics with Starting Temperature

Next, we examined variability in epidemic dynamics due to the temperature at

which the epidemic began, given two seasonal temperature regimes (25°C mean and

a seasonal range of 10°C to 40°C or 20°C to 30°C, respectively). Given that an

epidemic occurred, epidemic length monotonically decreased as a function of

starting temperature for the first temperature regime (Fig. 3, A): warmer

temperatures at the start of the epidemic produced shorter epidemics, and vice

versa. In the second temperature regime, epidemic length monotonically decreased

as a function of starting temperature until ~29°C. When temperature varied from

10°C to 40°C, the longest epidemic simulated was 137.8 days and occurred at

starting temperatures of 11.2°C, and the shortest epidemic lasted 16.82 days and

occurred when the temperature at the epidemic start was 35.7°C. When the

temperature was 35.8°C or higher or 10.2°C or lower, no epidemic occurred. When

temperature was constrained between 20°C and 30°C, the longest epidemic

simulated was 253.64 days at a starting temperature of 20°C, and the shortest

epidemic lasted 136.1 days at a starting temperature of 28.9°C.

Fig 3. Epidemiological indices as a function of starting temperature, within a

given seasonal temperature regime. The red curve represents the maximum

number of humans in the infected class (I_H) at any given point during the simulation.

The blue curve represents the final (or cumulative) epidemic size (R_H at the final

time step). The green curve represents the length of the epidemic (i.e., the point at

which the number of infected individuals was below one). Here, simulations were

run with the temperature conditions: T_{min} = 10°C, T_{mean} = 25°C, and T_{max} = 40°C (A)

and T_{min} = 20°C, T_{mean} = 25°C, and T_{max} = 30°C (B).

In contrast to epidemic length, the response of final epidemic size and

maximum number of infected individuals to the temperature at epidemic onset

depended on the amount of seasonal temperature variation. When temperature

varied widely, from 10°C to 40°C, both final epidemic size and the maximum

number of infected individuals responded unimodally to starting temperature, with

peaks at 23.9°C and 24.1°C, respectively (Fig. 3, A). By contrast, when temperature

varied more narrowly from 20°C to 30°C, the final epidemic size and the maximum

number of infected individuals were insensitive to starting temperature (Fig. 3, B).

Taken together, these results show that epidemics introduced at different times

within identical seasonal temperature regimes can produce very similar final

epidemic sizes and maximum infection rates, provided that the temperature range is

sufficiently constrained. If temperature variation is large, dramatically different final

epidemic sizes and maximum infection rates may result.

Seasonal Variability of Final Epidemic Size

To address how mean temperature and seasonal variance combined to influence the final epidemic size, we simulated over a wide range of temperature regimes that accounted for variation in the mean and temperature range over a calendar year. We calculated relative epidemic suitability, defined as the final epidemic size as a proportion of the human population, for twenty major cities worldwide (Table 3).

In a low-variation thermal environment, a band of mean temperatures between approximately 25°C and 35°C supports the highest epidemic suitability (Fig. 4). As the seasonal temperature range increases, lower mean temperatures are capable of supporting large epidemics. However, outside this narrow band of temperature regimes, epidemic suitability rapidly diminishes, and most temperature regimes did not produce epidemics.

Table 3. Estimates of epidemic suitability for major cities. Epidemic suitability was calculated as the proportion of the population that became infected in simulations run with 0, 20, 40, 60, or 80% initial population immunity. Temperature at simulation onset was set to the mean of the temperature regime. Each city was simulated with its respective temperature regime from the 2016 calendar year.

	Epidemic Suitability				
City	0% Immunity	20% Immunity	40% Immunity	60% Immunity	80% Immunity

Buenos Aires, Argentina	0.03656	0.02169	0.01203	0.005975	0.002295
Sao Paulo, Brazil	0.6056	0.3386	0.1518	0.05351	0.01385
Rio de Janeiro, Brazil	0.9984	0.7962	0.5891	0.3618	0.09862
Salvador, Brazil	0.9990	0.7976	0.5937	0.3804	0.1335
Fortaleza, Brazil	0.9993	0.7982	0.5953	0.3861	0.1535
Belo Horizonte, Brazil	0.5909	0.3344	0.1544	0.05771	0.01633
Recife, Brazil	0.9994	0.7985	0.5959	0.3871	0.1517
Shanghai, China	0.9966	0.7878	0.5507	0.2484	0.03456
Beijing, China	0.5268	0.2526	0.09058	0.02298	0.003587
Guangzhou, China	0.9996	0.7989	0.5965	0.3848	0.1254

Bogotá, Colombia	0.0001000	0.0001000	0.0001000	0.0001000	0.0001000
Medellin, Colombia	0.002544	0.002048	0.001556	0.001068	0.0005820
Cali, Colombia	0.9909	0.7822	0.5617	0.3122	0.07217
Barranquilla, Colombia	0.9997	0.7993	0.5979	0.3928	0.1703
Cartagena, Colombia	0.9997	0.7993	0.5978	0.3923	0.1688
Delhi, India	0.9537	0.7215	0.4759	0.2388	0.06803
Tokyo, Japan	0.7269	0.4149	0.1758	0.05159	0.009489
Kobe, Japan	0.9435	0.6669	0.3522	0.1090	0.01632
Manila, Philippines	0.9998	0.7994	0.5981	0.3933	0.1720
New York, USA	0.04088	0.02159	0.01041	0.004390	0.001425

Table 4. Estimates of epidemic suitability for major cities under different

starting temperatures. Epidemic suitability was calculated as the proportion of the

population that became infected in simulations that began at the minimum, mean, or

maximum temperature of the seasonal temperature regime. Each city was simulated

with its respective temperature regime from the 2016 calendar year with 0%

population immunity.

	Epidemic Suitability		
City	Minimum Starting Temperature	Mean Starting Temperature	Maximum Starting Temperature
Buenos Aires, Argentina	0.0001000	0.03656	0.1166
Sao Paulo, Brazil	0.02026	0.6056	0.3480
Rio de Janeiro, Brazil	0.9978	0.9984	0.9760
Salvador, Brazil	0.9965	0.9990	0.9963
Fortaleza, Brazil	0.9986	0.9993	0.9990
Belo Horizonte, Brazil	0.09404	0.5909	0.3273
Recife, Brazil	0.9973	0.9994	0.9987
Shanghai, China	0.0001000	0.9966	0.8905
Beijing, China	0.0001000	0.5268	0.5792
Guangzhou, China	0.9983	0.9996	0.9912
Bogotá, Colombia	0.0001000	0.0001000	0.0001000
Medellin, Colombia	0.0002177	0.002544	0.004472
Cali, Colombia	0.9858	0.9909	0.9623
Barranquilla, Colombia	0.9994	0.9997	0.9997
Cartagena, Colombia	0.9993	0.9997	0.9997
Delhi, India	0.5615	0.9537	0.6954

Tokyo, Japan	0.0001000	0.7269	0.5121
Kobe, Japan	0.0001000	0.9435	0.6890
Manila, Philippines	0.9994	0.9998	0.9998
New York, USA	0.0001000	0.04088	0.1863

Fig 4. Variation in epidemic suitability across different seasonal temperature regimes. The heat map shows the epidemic suitability (represented as the proportion of the total human population infected during an epidemic) as a function of mean annual temperature and temperature range. Here, temperature range is defined as the seasonal variation about the annual mean temperature. Twenty large, globally important cities are plotted to illustrate their epidemic suitability.

Of the focal 20 major cities, those with high mean temperature and small average temperature variation exhibited the highest epidemic suitability. For instance, Manila, Philippines, which has a monthly mean temperature of 29°C and average seasonal amplitude in mean temperature of 1.50°C, had an epidemic suitability of 0.9998. Cartagena and Barranquilla, Colombia had epidemic suitability of 0.9997. On the other hand, areas with low average temperature and greater temperature variation, such as Beijing and New York, exhibited lower—but still non-zero—epidemic suitabilities of 0.5268 and 0.04088 respectively. Notably, Guangzhou and Shanghai, China have high epidemic suitability (0.9996 and 0.9966, respectively) despite moderate mean temperatures (22.9 and 17.6°C, respectively) due to high seasonal variation in temperature. By contrast, high seasonal variation

reduced suitability to 0.9537 in Delhi, India, which has a high mean temperature of

26.3°C (Fig. 4).

The relationship between epidemic suitability and seasonal temperature

regime was consistent across varying levels of population immunity. Locations with

high mean temperatures and small average temperature variation had higher

epidemic suitability, regardless of the level of population immunity (Figures S2-S5).

However, as the level of immunity increased from 20% to 80%, the epidemic

suitability at given seasonal temperature regime decreased (Table 3).

Epidemic suitability also varied by starting temperature, depending on the

seasonal temperature regime. The epidemic suitability of cities with high mean

temperature and small average temperature variation—such as Manila, Philippines

and Cartagena and Barranquilla, Colombia—did not depend on starting temperature

(Table 4). However, areas with low to moderate mean temperature and large

average temperature variation (e.g., Kobe, Japan and Shanghai, China) exhibited low

epidemic suitability (both 0.0001000) at the minimum starting temperature and

moderate-to-high epidemic suitability at the maximum starting temperature

(0.6890 and 0.8905, respectively) (Fig. 5). The opposite occurred in regimes with

high mean temperature and large temperature variation, though these temperature

regimes are rarer.

Estimated epidemic suitability is close to one in the most suitable

temperature regimes because we assumed that: (i) the population was fully

susceptible at the start of the epidemic; (ii) mixing was homogeneous among

humans and mosquitoes; (iii) all cases of infection are included regardless of

whether or not they are symptomatic; and (iv) no other environmental or social

drivers are limiting transmission. As a result, the epidemic suitability metric should

be considered an upper bound on the proportion of the population that could

become infected based on temperature alone.

Fig 5. Variation in epidemic suitability across different seasonal temperature

regimes averaged across starting temperatures. The heat map shows the

epidemic suitability (represented as the proportion of the total human population

infected during an epidemic) as a function of mean annual temperature and

temperature range averaged across simulations where the initial temperature was

set to the seasonal temperature regime's minimum, mean, or maximum

temperature. Here, temperature range is defined as the seasonal variation about the

annual mean temperature. Twenty large, globally important cities are plotted to

illustrate their epidemic suitability.

Model Sensitivity and Uncertainty Analysis

Final epidemic size was not sensitive to life history trait parameterization (Figs. S8-

S10), using samples from the posterior distribution of thermal response fits for each

temperature-dependent trait.

There was uncertainty in the specific numerical values of the epidemiological

indices across starting temperatures (Fig. S1). However, the overall functional

response of the final epidemic size, maximum number of infected individuals, and

the epidemic length to starting temperature was consistent across the samples from

the joint posterior distribution.

Discussion

Recent outbreaks of DENV, CHIKV, and ZIKV in Latin America and across the

globe have captured the attention of the public health community and underscore

the importance of preparation for future outbreaks. As temperatures rise, the global

landscape suitable for such outbreaks will expand and shift geographically,

potentially placing a larger proportion of the world's population at risk [24,29,31].

Understanding how local temperature regimes govern epidemic dynamics is

increasingly important for determining resource allocation and control

interventions [41]. While previous work has investigated the effects of temperature

on DENV, CHIKV, and/or ZIKV transmission, until now we have lacked

comprehensive, mechanistic, and dynamic understanding of the effects of seasonally

varying temperature on transmission via its (nonlinear) effects on mosquito and

parasite traits [27–34]. With our model, which expands on [24] and [25], we show

that seasonal temperature mean and amplitude interact with the temperature at

epidemic onset to shape the speed and magnitude of epidemics.

At constant temperature, epidemics varied substantially in the rate at which

susceptible individuals were depleted. Epidemics simulated at 25°C and 30°C

reached similar sizes but the epidemic at 25°C proceeded at a much slower rate (Fig.

2). This "slow burn" phenomenon occurs because slower depletion of susceptible

individuals can produce epidemics of similar size to epidemics that infect people

very rapidly. This phenomenon also occurs in more realistic, seasonally varying

temperature regimes.

527   The temperature at which an epidemic started affected dynamics only under

528   large ranges of temperature variation. When temperature ranged from 10°C to 40°C,

529   the final epidemic size peaked at intermediate starting temperatures (24°C; Fig. 3,

530   A). However, in highly suitable seasonal environments, final epidemic size was large

531   regardless of the starting temperature (Fig. 3, B).

532   At mean starting temperatures, epidemic suitability was sensitive to the

interaction between annual temperature mean and seasonal variation. Under low

seasonal temperature variation, a narrow band of annual mean temperatures

(approximately 25-35°C) had the highest epidemic suitability (Figs. 4 & S2-S5).

Outside this band of temperature regimes, suitability diminishes rapidly. Larger

seasonal variation in temperature lowers the range of optimal annual mean

temperatures (i.e., suitability is high in cooler places with larger seasonal variation

in temperature; Fig. 4).

540   The relationship between epidemic suitability and the seasonal temperature

regime also depended on the temperature at the epidemic onset. Three distinct

relationships emerged (Figs. 5 & S6-S7). At intermediate annual mean temperatures

of ~25-35°C and low seasonal temperature variation (~0-10°C), epidemic suitability

is insensitive to starting temperature because temperature is suitable for

transmission year-round. At lower annual mean temperatures (~10-25°C) and

higher seasonal temperature variation (~10-15°C), epidemic suitability is highest

when epidemics start in moderate to warm seasons, and lower when epidemics

start during cooler seasons. Finally, at high annual mean temperatures (> 35°C) and

low seasonal temperature variation (~0-10°C), epidemic suitability is high only

when epidemics start at the coldest period of the year, because otherwise the

temperature is too warm for efficient transmission. The interaction between

temperature mean, annual variation, and starting point sharply illustrates the

unimodal effect of temperature on transmission. Models that do not include

unimodal effects of temperature (e.g., those with sinusoidal forcing on a

transmission parameter) may fail to capture the limits on transmission in warm

environments.

With rising mean annual temperatures and increasing seasonal temperature

variation due to climate change, the landscape of epidemic suitability is likely to

shift. Importantly, areas with previously low epidemic suitability may have

increasing potential for transmission year-round. By contrast, warming

temperatures may drive epidemics in cities with high current suitability (e.g.,

Manila, Philippines, Barranquilla, Colombia, and Fortaleza, Brazil) to shift toward

cooler months. Thus, climate change may alter not only epidemic size and duration

but also seasonal timing globally, as it interacts with other important drivers like

rainfall and human behavior.

It is important to note that model-estimated epidemic suitability should be

treated as an upper bound on the potential for large epidemics because within

highly suitable climate regimes, epidemics can vary in magnitude due to human

population size and movement dynamics [28], effective vector control, and other

mitigating factors. Likewise, our estimates are conditioned on *Aedes aegypti*

presence and virus introduction to support an outbreak.

Although seasonal temperature dynamics provide insight into vector-borne

transmission dynamics, other factors like mosquito abundance, vector control, and

rainfall also determine transmission dynamics. Thus, temperature must be

considered jointly with these factors. Moreover, accurately describing epidemic

dynamics of emerging and established vector-borne pathogens will ultimately

require integrating realistic models of environmental suitability, as presented here,

with demographic, social, and economic factors that promote or limit disease

transmission [42,43]. Conversely, we show that the interaction between

temperature and the availability of susceptible hosts alone can drive epidemic

burnout even in the absence of other limiting factors like vector control and

seasonal precipitation. This suggests that correctly representing the nonlinear

relationship between temperature and epidemic dynamics is critical for accurately

inferring mechanistic drivers of epidemics and, in turn, predicting the efficacy of

control interventions.

References

1.   Gubler DJ. The global emergence/resurgence of arboviral diseases as public
health problems. Arch Med Res. 2002;33: 330–342.
2.   Faria NR, Azevedo R do S da S, Kraemer MUG, Souza R, Cunha MS, Hill SC, et
al. Zika virus in the Americas: Early epidemiological and genetic findings. Science.
2016;352: 345–349. doi:10.1126/science.aaf5036
3.   Staples JE, Breiman RF, Powers AM. Chikungunya Fever: An Epidemiological
Review of a Re-Emerging Infectious Disease. Clin Infect Dis. 2009;49: 942–948.
doi:10.1086/605496
4.   Special Programme for Research and Training in Tropical Diseases, World Health
Organization, editors. Dengue: guidelines for diagnosis, treatment, prevention, and
control. New ed. Geneva: TDR : World Health Organization; 2009.
5.   Gubler DJ. Dengue/dengue haemorrhagic fever: history and current status.
Novartis Found Symp. 2006;277: 3–16; discussion 16-22, 71–73, 251–253.
6.   Brady OJ, Gething PW, Bhatt S, Messina JP, Brownstein JS, Hoen AG, et al.
Refining the global spatial limits of dengue virus transmission by evidence-based
consensus. PLoS Negl Trop Dis. 2012;6: e1760. doi:10.1371/journal.pntd.0001760
7.   Bhatt S, Gething PW, Brady OJ, Messina JP, Farlow AW, Moyes CL, et al. The
global distribution and burden of dengue. Nature. 2013;496: 504–507.
doi:10.1038/nature12060
8.   Khan K, Bogoch I, Brownstein JS, Miniota J, Nicolucci A, Hu W, et al. Assessing
the origin of and potential for international spread of chikungunya virus from the
Caribbean. PLoS Curr. 2014;6.
doi:10.1371/currents.outbreaks.2134a0a7bf37fd8d388181539fea2da5
9.   Perkins TA, Metcalf CJE, Grenfell BT, Tatem AJ. Estimating drivers of
autochthonous transmission of chikungunya virus in its invasion of the americas. PLoS
Curr. 2015;7. doi:10.1371/currents.outbreaks.a4c7b6ac10e0420b1788c9767946d1fc
10.   Nsoesie EO, Kraemer MU, Golding N, Pigott DM, Brady OJ, Moyes CL, et al.
Global distribution and environmental suitability for chikungunya virus, 1952 to 2015.
Euro Surveill Bull Eur Sur Mal Transm Eur Commun Dis Bull. 2016;21.
doi:10.2807/1560-7917.ES.2016.21.20.30234
11.   PAHO. Regional Zika Epidemiological Update (Americas) August 25, 2017. Pan
American Health Organization;
12.   Bjornstad ON, Finkenstadt BF, Grenfell BT. Dynamics of Measles Epidemics:
Estimating Scaling of Transmission Rates Using a Time Series SIR Model. Ecol Monogr.
2002;72: 169. doi:10.2307/3100023
13.   Coburn BJ, Wagner BG, Blower S. Modeling influenza epidemics and
pandemics: insights into the future of swine flu (H1N1). BMC Med. 2009;7.
doi:10.1186/1741-7015-7-30
14.   Kucharski AJ, Funk S, Eggo RM, Mallet H-P, Edmunds WJ, Nilles EJ.
Transmission Dynamics of Zika Virus in Island Populations: A Modelling Analysis of the
2013–14 French Polynesia Outbreak. Barker CM, editor. PLoS Negl Trop Dis. 2016;10:
e0004726. doi:10.1371/journal.pntd.0004726
15.   Eisen L, Monaghan AJ, Lozano-Fuentes S, Steinhoff DF, Hayden MH, Bieringer
PE. The impact of temperature on the bionomics of Aedes (Stegomyia) aegypti, with

special reference to the cool geographic range margins. J Med Entomol. 2014;51: 496–
516.
16.   Rueda LM, Patel KJ, Axtell RC, Stinner RE. Temperature-dependent
development and survival rates of Culex quinquefasciatus and Aedes aegypti (Diptera:
Culicidae). J Med Entomol. 1990;27: 892–898.
17.   Tun-Lin W, Burkot TR, Kay BH. Effects of temperature and larval diet on
development rates and survival of the dengue vector Aedes aegypti in north Queensland,
Australia. Med Vet Entomol. 2000;14: 31–37.
18.   Kamimura K, Matsuse IT, Takahashi H, Komukai J, Fukuda T, Suzuki K, et al.
Effect of temperature on the development of Aedes aegypti and Aedes albopictus. Med
Entomol Zool. 2002;53: 53–58. doi:10.7601/mez.53.53_1
19.   Beserra EB, Fernandes CRM, Silva SA de O, Silva LA da, Santos JW dos. Efeitos
da temperatura no ciclo de vida, exigências térmicas e estimativas do número de gerações
anuais de Aedes aegypti (Diptera, Culicidae). Iheringia Sér Zool. 2009;99: 142–148.
doi:10.1590/S0073-47212009000200004
20.   Couret J, Dotson E, Benedict MQ. Temperature, larval diet, and density effects on
development rate and survival of Aedes aegypti (Diptera: Culicidae). PloS One. 2014;9:
e87468. doi:10.1371/journal.pone.0087468
21.   Watts DM, Burke DS, Harrison BA, Whitmire RE, Nisalak A. Effect of
temperature on the vector efficiency of Aedes aegypti for dengue 2 virus. Am J Trop Med
Hyg. 1987;36: 143–152.
22.   McLean DM, Clarke AM, Coleman JC, Montalbetti CA, Skidmore AG, Walters
TE, et al. Vector capability of Aedes aegypti mosquitoes for California encephalitis and
dengue viruses at various temperatures. Can J Microbiol. 1974;20: 255–262.
23.   Altizer S, Dobson A, Hosseini P, Hudson P, Pascual M, Rohani P. Seasonality
and the dynamics of infectious diseases. Ecol Lett. 2006;9: 467–484. doi:10.1111/j.1461-
0248.2005.00879.x
24.   Mordecai EA, Cohen JM, Evans MV, Gudapati P, Johnson LR, Lippi CA, et al.
Detecting the impact of temperature on transmission of Zika, dengue, and chikungunya
using mechanistic models. PLoS Negl Trop Dis. 2017;11: e0005568.
doi:10.1371/journal.pntd.0005568
25.   Yang HM, Macoris MLG, Galvani KC, Andrighetti MTM, Wanderley DMV.
Assessing the effects of temperature on dengue transmission. Epidemiol Infect. 2009;137:
1179–1187. doi:10.1017/S0950268809002052
26.   Yang HM, Boldrini JL, Fassoni AC, Freitas LFS, Gomez MC, de Lima KKB, et
al. Fitting the Incidence Data from the City of Campinas, Brazil, Based on Dengue
Transmission Modellings Considering Time-Dependent Entomological Parameters. PloS
One. 2016;11: e0152186. doi:10.1371/journal.pone.0152186
27.   Johansson MA, Powers AM, Pesik N, Cohen NJ, Staples JE. Nowcasting the
spread of chikungunya virus in the Americas. PloS One. 2014;9: e104915.
doi:10.1371/journal.pone.0104915
28.   Wesolowski A, Qureshi T, Boni MF, Sundsøy PR, Johansson MA, Rasheed SB,
et al. Impact of human mobility on the emergence of dengue epidemics in Pakistan. Proc
Natl Acad Sci U S A. 2015;112: 11887–11892. doi:10.1073/pnas.1504964112
29.   Liu-Helmersson J, Stenlund H, Wilder-Smith A, Rocklöv J. Vectorial capacity of
Aedes aegypti: effects of temperature and implications for global dengue epidemic

potential. PloS One. 2014;9: e89783. doi:10.1371/journal.pone.0089783

30.   Morin CW, Monaghan AJ, Hayden MH, Barrera R, Ernst K. Meteorologically
Driven Simulations of Dengue Epidemics in San Juan, PR. PLoS Negl Trop Dis. 2015;9:
e0004002. doi:10.1371/journal.pntd.0004002

31.   Caminade C, Turner J, Metelmann S, Hesson JC, Blagrove MSC, Solomon T, et
al. Global risk model for vector-borne transmission of Zika virus reveals the role of El
Niño 2015. Proc Natl Acad Sci U S A. 2017;114: 119–124.
doi:10.1073/pnas.1614303114

32.   Lourenço J, Recker M. The 2012 Madeira dengue outbreak: epidemiological
determinants and future epidemic potential. PLoS Negl Trop Dis. 2014;8: e3083.
doi:10.1371/journal.pntd.0003083

33.   Faria NR, da Costa AC, Lourenço J, Loureiro P, Lopes ME, Ribeiro R, et al.
Genomic and epidemiological characterisation of a dengue virus outbreak among blood
donors in Brazil. Sci Rep. 2017;7: 15216. doi:10.1038/s41598-017-15152-8

34.   Lourenço J, Maia de Lima M, Faria NR, Walker A, Kraemer MU, Villabona-
Arenas CJ, et al. Epidemiological and ecological determinants of Zika virus transmission
in an urban setting. eLife. 2017;6. doi:10.7554/eLife.29820

35.   Duong V, Lambrechts L, Paul RE, Ly S, Lay RS, Long KC, et al. Asymptomatic
humans transmit dengue virus to mosquitoes. Proc Natl Acad Sci U S A. 2015;112:
14688–14693. doi:10.1073/pnas.1508114112

36.   Appassakij H, Khuntikij P, Kemapunmanus M, Wutthanarungsan R, Silpapojakul
K. Viremic profiles in asymptomatic and symptomatic chikungunya fever: a blood
transfusion threat? Transfusion (Paris). 2013;53: 2567–2574. doi:10.1111/j.1537-
2995.2012.03960.x

37.   Murphy BR, Whitehead SS. Immune response to dengue virus and prospects for a
vaccine. Annu Rev Immunol. 2011;29: 587–619. doi:10.1146/annurev-immunol-031210-
101315

38.   Weaver SC, Osorio JE, Livengood JA, Chen R, Stinchcomb DT. Chikungunya
virus and prospects for a vaccine. Expert Rev Vaccines. 2012;11: 1087–1101.
doi:10.1586/erv.12.84

39.   Le Menach A, McKenzie FE, Flahault A, Smith DL. The unexpected importance
of mosquito oviposition behaviour for malaria: non-productive larval habitats can be
sources for malaria transmission. Malar J. 2005;4: 23. doi:10.1186/1475-2875-4-23

40.   Palamara GM, Childs DZ, Clements CF, Petchey OL, Plebani M, Smith MJ.
Inferring the temperature dependence of population parameters: the effects of
experimental design and inference algorithm. Ecol Evol. 2014;4: 4736–4750.
doi:10.1002/ece3.1309

41.   Perkins TA. Retracing Zika's footsteps across the Americas with computational
modeling. Proc Natl Acad Sci U S A. 2017;114: 5558–5560.
doi:10.1073/pnas.1705969114

42.   Zhang Q, Sun K, Chinazzi M, Pastore Y Piontti A, Dean NE, Rojas DP, et al.
Spread of Zika virus in the Americas. Proc Natl Acad Sci U S A. 2017;114: E4334–
E4343. doi:10.1073/pnas.1620161114

43.   Alex Perkins T, Siraj AS, Ruktanonchai CW, Kraemer MUG, Tatem AJ. Model-
based projections of Zika virus infections in childbearing women in the Americas. Nat
Microbiol. 2016;1: 16126. doi:10.1038/nmicrobiol.2016.126

Supporting Information Legends

S1 Fig. Sensitivity of epidemiological indices as a function of starting temperature to the parametrization of life history traits. The red curve represents the median maximum number of humans in the infected class (I_H) at any given point during the simulation. The blue curve represents the median final (or cumulative) epidemic size (R_H at the final time step). The green curve represents the median length of the epidemic (i.e., the point at which the number of infected individuals was below one). Each shaded area represents the 95% credible interval for the epidemiological indices ran under 50 different parameterizations of the life history traits. Here, simulations were run with the temperature conditions: T_{min} = 10°C, T_{mean} = 25°C, and T_{max} = 40°C (A) and T_{min} = 20°C, T_{mean} = 25°C, and T_{max} = 30°C (B).

S2 Fig. Variation in epidemic suitability across different seasonal temperature regimes with 20% population immunity. The heat map shows the epidemic suitability (represented as the proportion of the total human population infected during an epidemic) as a function of mean annual temperature and temperature range assuming 20% population immunity. Here, temperature range is defined as the seasonal variation about the annual mean temperature. Twenty large, globally important cities are plotted to illustrate their epidemic suitability.

S3 Fig. Variation in epidemic suitability across different seasonal temperature regimes with 40% population immunity. The heat map shows the epidemic suitability (represented as the proportion of the total human population infected during an epidemic) as a function of mean annual temperature and temperature

range assuming 40% population immunity. Here, temperature range is defined as

the seasonal variation about the annual mean temperature. Twenty large, globally

important cities are plotted to illustrate their epidemic suitability.

S4 Fig. Variation in epidemic suitability across different seasonal temperature

regimes with 60% population immunity. The heat map shows the epidemic

suitability (represented as the proportion of the total human population infected

during an epidemic) as a function of mean annual temperature and temperature

range assuming 60% population immunity. Here, temperature range is defined as

the seasonal variation about the annual mean temperature. Twenty large, globally

important cities are plotted to illustrate their epidemic suitability.

S5 Fig. Variation in epidemic suitability across different seasonal temperature

regimes with 80% population immunity. The heat map shows the epidemic

suitability (represented as the proportion of the total human population infected

during an epidemic) as a function of mean annual temperature and temperature

range assuming 80% population immunity. Here, temperature range is defined as

the seasonal variation about the annual mean temperature. Twenty large, globally

important cities are plotted to illustrate their epidemic suitability.

S6 Fig. Variation in epidemic suitability across different seasonal temperature

regimes with minimum starting temperature. The heat map shows the epidemic

suitability (represented as the proportion of the total human population infected

during an epidemic) as a function of mean annual temperature and temperature

range. Here, temperature range is defined as the seasonal variation about the annual

mean temperature, and the simulation began at the minimum temperature of the

regime. Twenty large, globally important cities are plotted to illustrate their

epidemic suitability.

S7 Fig. Variation in epidemic suitability across different seasonal temperature

regimes with maximum starting temperature. The heat map shows the epidemic

suitability (represented as the proportion of the total human population infected

during an epidemic) as a function of mean annual temperature and temperature

range. Here, temperature range is defined as the seasonal variation about the annual

mean temperature, and the simulation began at the maximum temperature of the

regime. Twenty large, globally important cities are plotted to illustrate their

epidemic suitability.

S8 Fig. The 2.5% quantile of epidemic suitability to the parameterization of

life history traits. Epidemic suitability (represented as the proportion of the total

human population infected during an epidemic) as a function of mean annual

temperature and the temperature range. Temperature varied according to a

seasonal temperature regime, and 50 samples of c, T_{min}, and T_{max} were taken from

the joint posterior distribution of each trait thermal response from Mordecai et al.

[24].

S9 Fig. The 50% quantile of epidemic suitability to the parameterization of life

history traits. Epidemic suitability (represented as the proportion of the total

human population infected during an epidemic) as mean annual temperature and

the temperature range. Temperature varied according to a seasonal temperature

regime, and 50 samples of c, T_{min}, and T_{max} were taken from the joint posterior

distribution of each trait thermal response from Mordecai et al. [24].

S10 Fig. The 97.5% quantile of epidemic suitability to the parameterization of

life history traits. Epidemic suitability (represented as the proportion of the total

human population infected during an epidemic) as mean annual temperature and

the temperature range. Temperature varied according to a seasonal temperature

regime, and 50 samples of c, T_{min}, and T_{max} were taken from the joint posterior

distribution of each trait thermal response from Mordecai et al. [24].

799
800