Fine-grained habitat-associated genetic connectivity in an admixed population of mussels in the small isolated Kerguelen Islands

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Running title: Admixture and genetic-environment association in a small island

Keywords: *Mytilus* mussels, Fine-grained connectivity, , Admixture, Introgression, Genetic - environment association.

<u>Abstract</u>

Reticulated evolution -i.e. secondary introgression / admixture between sister taxa- is increasingly

recognized as playing a key role in structuring infra-specific genetic variation and revealing cryptic

connectivity patterns. When admixture zones coincide with ecological transitions, the connectivity

patterns often follow environmental variations better than distance and introgression clines may

easily be confounded with local adaptation. The Kerguelen mussels is an ideal system to investigate

the potential role of admixture in enhancing micro-geographic structure, as they inhabit a small

isolated island in the Southern Ocean characterized by a highly heterogeneous environment.

Furthermore, genomic reticulation between Northern species (*M. edulis, M. galloprovincialis* and

M. trossulus) and Southern species (*M. platensis*: South America and the Kerguelen Islands; and

M. planulatus: Australasia) has been suspected. Here, we extended a previous analysis by using

targeted-sequencing data (51,878 SNPs) across the three Northern species and the Kerguelen

population. Spatial structure in the Kerguelen was then analyzed with a panel of 33 SNPs, including

SNPs that were more differentiated than the genomic average between Northern species (i.e.,

ancestry-informative SNPs). We first showed that the Kerguelen lineage splitted very shortly after

M. edulis and *M. galloprovincialis* initiated speciation, and it subsequently experienced admixture

with the three Northern taxa. We then demonstrated that the Kerguelen mussels were significantly

differentiated over small spatial distance, and that this local genetic structure was associated with

environmental variations and mostly revealed by ancestry-informative markers. Simulations of

admixture in the island highlight that genetic-environment associations can be better explained by

introgression clines between heterogeneously differentiated genomes than by adaptation from

admixture variation.

Introduction

Marine species with planktonic larvae are high-dispersal organisms, with large effective population

sizes and living in a highly connected environment (Cowen & Sponaugle 2009), therefore they

generally show low levels of genetic differentiation across their species ranges (Palumbi 1992).

Nevertheless, there is some evidence that micro-geographic genetic-environment associations occur

at specific loci in marine species, such as barnacles (Schmidt & Rand 1999), mussels (Koehn *et al.*

1980) or Atlantic killifishes (Reid *et al.* 2017), despite genome-wide genetic homogeneity. These

locus-specific associations are traditionally expected in regions of the species range where

environmental gradients promote local adaptation of specific populations (Schmidt *et al.* 2008).

Alternatively, genetic-environment associations can be observed where semi-permeable genetic

backgrounds form a contact zone that coincides with an ecological boundary (Bierne *et al.* 2011). In

this case, reproductive isolation genes become locally coupled with local adaptation genes and

linked neutral markers, and the two genetic backgrounds become associated with different habitats.

In addition, introgression creates a departure from equilibrium situations in which the influx of

heterospecific alleles from one genetic background generates a gradient in allele frequencies within

the other genetic background, revealing cryptic connectivity patterns (Gagnaire *et al.* 2015).

Gradients of introgression may easily be confounded with local adaptation signatures, especially

when environmental variables are better proxy of the connectivity matrix than geographic distance.

There are two reasons for that: (i) the environment can have a direct effect on connectivity (e.g.

landscape resistance model, Zeller *et al.* 2012), or (ii) the effect of a barrier to dispersal can be

better described by one of the many scored environmental variables that can incidentally or truly

coincide with it, than by distance. In the sea, regions of little water mixing are inevitably both

ecotones and barrier to larval dispersal. Most importantly, the reduction in gene flow between

genetic backgrounds is expected to be visible only on a subset of markers either because localized at

an intermediate linkage map distance to reproductive isolation genes (Gagnaire *et al.* 2015) or

because of heterogeneous differentiation produced by linked selection (Simon and Duranton 2018).

This potential effect of introgression on our capacity to detect connectivity breaks in apparently

well-mixed populations is of central concern to conservation and species management (Gagnaire *et*

al. 2015).

Mytilus mussels are an excellent system to address these issues, because they are subdivided into

partially reproductively isolated species. Moreover, a recent study based on F_{ST} genome scans and

gene genealogies demonstrated that population-specific introgression is widespread, and it is the

primary cause of outlying levels of genetic differentiation between conspecific populations (Fraïsse

et al. 2016). *Mytilus* mussels have an antitropical distribution, i.e. they occur in high latitudes of the

Northern and Southern Hemispheres, as a result of transequatorial migration during the Pleistocene

(Hilbish *et al.* 2000; Gérard *et al.* 2008). In the North, *M. edulis* and *M. galloprovincialis* are

closely-related species which started to diverge about 2.5 mya (Roux *et al.* 2014), while *M.*

trossulus is an outgroup to them with a divergence dated at 3.5 mya (Rawson & Hilbish 1995). The

three species have experienced a complex history of divergence punctuated by periods of gene flow

(Roux *et al.* 2014; Fraïsse et al. 2018); and nowadays they display hybrid zones where their ranges

overlap (Skibinski *et al.* 1983; Väinölä & Hvilsom 1991; Bierne *et al.* 2003). In the South, a re-

evaluation of allozyme data and a review of the results obtained with mtDNA and two nuclear DNA

markers (Borsa *et al.* 2012) encouraged to group Southern mussels in two different taxa, namely *M.*

platensis for those related to *M. edulis* (the South American and Kerguelen mussels), and *M.*

planulatus for those related to *M. galloprovincialis* (the Australasian mussels). The presence of a

mitochondrial clade endemic to the Southern Ocean further suggests Southern mussels are native

rather than introduced by human-mediated activities (Hilbish *et al.* 2000; Gaitán-Espitia *et al.*

2016).

In the Southern Indian ocean, the isolated Kerguelen Islands harbor *M. platensis* mussels which

are polymorphic for allozyme alleles characteristic of all three Northern species (Blot *et al.* 1988), although they are most similar to *M. edulis* at a few allozyme loci (McDonald *et al.* 1991). Further analyses with nuclear markers strengthened the view of a mixed genome ancestry of the Kerguelen mussels (Borsa *et al.* 2007): at Glu-5', a Northern diagnostic marker, mussels carry a heterospecific polymorphism (*M. edulis* / *M. galloprovincialis*). These preliminary results suggest either that reproductive isolation genes responsible of the interspecific barrier in the North were not yet evolved at the time of admixture (if any) in the Kerguelen, or that isolation is not as strong in the demographic, ecological and genetic context of the Kerguelen Islands as it is in the Northern Hemisphere hybrid zones.

The geomorphology of the Kerguelen Islands has been shaped by volcanic activity and glacial erosion which resulted in a carved coast with sheltered bays and fjords (Gérard *et al.* 2015). Recently, Gérard *et al.* (2015) have investigated the genetic-environment associations in the island with four nuclear markers (mac-1, Glu-5', EFbis and EFprem's) and a mitochondrial gene (COI). Only Glu-5' revealed significant genetic differentiation among and within geographic regions, and between habitats, despite the very small geographical scale. This pattern is striking as intraspecific genetic variation rarely shows significant differentiation even over large distances in *Mytilus* (e.g. within the Mediterranean Sea, Fraïsse *et al.* 2016). In particular, allele frequencies at Glu-5' were associated with the presence/absence of the kelp *Macrocystis* in the island, which serves as substrate and refuge for many species of molluscs, including mussels (Adami & Gordillo, 1999). As such, local adaptation was invoked to explain the fine-scale maintenance of polymorphism at Glu-5', although alternative interpretations involving admixture could not have been refuted.

Our aim was to investigate whether reticulate evolution actually contributed to micro-geographic structure in the Kerguelen islands. To this end, we ask the following questions: (i) can we detect footprints of introgression from Northern species and disentangle them from incomplete lineage sorting (i.e. shared ancestral polymorphism)? ; (ii) is there genetic differentiation at fine scale in the

island?; (iii) is there association of genetic differentiation with environmental variables and/or geography?; (iv) is the signal observed in (ii) and (iii) genome-wide or concentrated on ancestry-informative loci and (v) does introgression contribute to this fine-scale genetic differentiation? Based on published genotyping-by-sequencing (GBS) data of the three Northern species (Fraïsse *et al.* 2016) and new GBS data of a sample from a single Kerguelen population, we found that the Kerguelen Islands harbor a Southern lineage of mussels, related to *M. edulis,* and that was subsequently admixed with Northern species. Despite ancient incomplete lineage sorting resolved into multifarious topologies among the four taxa, and despite a non-negligible rate of ongoing sorting of lineages (i.e. true shared ancestral polymorphism), we also robustly detected past introgression events between Northern and Southern mussels by: (i) testing for admixture with genome-wide allele frequency data, (ii) reconstructing gene genealogies at a small chromosomal scale and (iii) inferring their divergence history from the joint site-frequency spectrum. Based on a new panel of SNPs genotyped on 695 mussels across 35 sites in the Kerguelen, we then confirmed using redundancy analysis (RDA) a significant fine-scale genetic differentiation between sites associated with environmental variables. Notably, we found that loci with strong genetic-environment association also tended to be among the most ancestry-informative markers in the *Mytilus spp*. While we cannot refute whether local adaptation has played a role here, simulations of admixture in the island highlights the importance of introgression for the current and local genetic structure of the Kerguelen mussels.

Materials and Methods

Genotyping-by-sequencing of the *Mytilus* spp.

We used samples collected from nine localities in the Northern Hemisphere (Figure 1, Supp. Info. M&M and Table S1) to investigate the patterns of admixture between Northern and Southern

genetic backgrounds in the Kerguelen Islands. The genetic composition of these samples has been analysed in Fraïsse *et al.* (2016) with target enrichment sequencing of bacterial artificial chromosomes (BAC) and cDNA sequences (see Supp. Info. M&M for details). The Northern samples have been shown to be representative of monospecific panmictic patches of the *Mytilus edulis* species complex, which comprise three species that hybridize at several places in the Northern Hemisphere: *M. galloprovincialis*, *M. edulis* and *M. trossulus*. In addition to these previously published samples, eight individuals from the Kerguelen Islands (Baie de la Mouche, Table S1) were included in the target enrichment experiment. These individuals were treated together with the Northern samples following the genotyping-by-sequencing (GBS) method described in Fraïsse *et al.* (2016) (see Supp. Info. M&M for details). The final dataset across the ten localities consisted of 1,269 reference sequences (378 BAC contigs that come from a pool of 224 unlinked clones, and 891 cDNA contigs that correspond to unlinked coding sequences of known-functions or randomly selected) and 129,346 SNPs. DNA sequences and VCF files including GBS genotypes are available on Dryad doi: 10.5061/dryad.6k740 (Fraïsse *et al.* 2016).

KASPar SNP panel

Based on the SNP database generated by GBS, we specifically selected SNPs segregating in the eight GBS-typed Kerguelen individuals to analyse the fine-scale genetic structure in the island, and its relation to the local environment. Moreover, as we wanted to determine if the micro-geographic structure in the Kerguelen was primarily driven by standing variation in the Northern complex of species (i.e. SNPs fixed between Northern species), the selected SNPs were not a random sample of the SNPs detected by GBS, otherwise they would have been mainly private polymorphisms to the Kerguelen (60% of the Kerguelen SNPs were found private). As such, we further enriched our SNP array with ancestry-informative markers, the most differentiated SNPs between pairs of Northern Hemisphere species (representing 33% of the non-private Kerguelen SNPs, 10% of the whole SNP

dataset), namely the West-Mediterranean *M. galloprovincialis* population, the North-Sea *M. edulis*

population and the Baltic-Sea *M. trossulus* population. F_{ST} values (Weir & Cockerham 1984) were

calculated using the R package hierfstat (Goudet 2005) for each SNP between pairs of populations

(Text S1). SNPs in the upper 15% of the empirical F_{ST} distribution were categorized as highly-

differentiated. SNPs with more than 25% of missing data were discarded. SNPs were further

filtered-out based on Illumina Assay Design Tool scores (available on Illumina web page,

http://support.illumina.com) which predicts probes success based on the number of degenerated

sites in the flanking sequences (250 bp on each side of the focal SNP). The final array comprised 58

SNPs out of which 30 were highly differentiated between Northern species (11 *M. trossulus*-

specific, 8 *M. edulis*-specific and 10 *M. galloprovincialis*-specific, Table S2).

KASPar genotyping in the Kerguelen Islands

We used samples collected from 35 sites in the Kerguelen Islands by Gérard *et al.* (2015), totalling

695 individuals (Supp. Info. M&M and Table S3). We additionally included other samples from the

Southern Hemisphere (Gérard *et al.* 2008) to assess the genetic relationships previously proposed

by Borsa *et al.* (2012) with a handful of markers: one sample from Western Australia in Nedlands

(AUS, n=12 individuals); two samples from New Zealand, Dunedin (DUN, n=8) and Wellington

Harbour (WHL, n=10); two samples from Tasmania, the Simpson's Bay (SIM, n=8) and Hobart

(HOB, n=9); one sample from Chile in Maullin (MAU, n=15). Pieces of mantle tissue were

preserved in 95% ethanol, and DNA was extracted with the Macherey-Nagel NucleoSPin 96 Tissue

kit. We used KASPar (Kompetitive Allele Specific PCR, Smith & Maughan 2015), a fluorescence-

based genotyping assay of allele-specific PCR products, to genotype the 58 SNPs, of them, 44 SNPs

were successfully amplified. We removed seven loci which showed significant F_{ST} values between

the eight GBS-typed Kerguelen individuals and the KASPar individuals. These may be due to error

in the genotyping-by-sequencing, typically the assembly of paralogous loci in two alleles of the

same locus, or alternatively to problem of amplification in the KASPar assay as a consequence of

primer design. More generally, these discrepancies call for studies that quantitatively compare the

two genotyping methods. We further eliminated two loci with null alleles (significant F_{IS} values in

most of the sampling sites) and two loci physically linked to one another. The final dataset was

composed of 33 KASPar SNPs (Table S2). Additionally, we included allele frequency data of a

length-polymorphism locus in the adhesive plaque protein gene, Glu-5', previously scored in the

same sampling sites (Gérard *et al.* 2015). Genotypes for all individuals at each KASPar SNP is

available in Text S2, and population allele frequencies are given in Table S4.

Genetic network of the *Mytilus* spp.

Genotypes of the GBS dataset were statistically phased with beagle v3.3.2 (Browning & Browning

2007) using genotype likelihoods provided by bcftools. All individuals were included in the analysis

to maximize linkage disequilibrium, and 20 haplotype pairs were sampled for each individual

during each iteration of the phasing algorithm to increase accuracy. Phased sequences (haplotypes)

were then generated using a custom perl script. An individual genetic network analysis was

conducted with splitstree4 v4.12.6 (Hureson & Bryant 2006) to get insight into the population

relationships across the three Northern Hemisphere species and the eight individuals sampled in the

Kerguelen Islands. All haplotype loci were compiled to create a composite chromosome of 51,878

high-quality SNPs and analysed using the neighbour-net method.

Analyses of admixture in the *Mytilus* spp.

An estimation of the historical relationships among the nine Northern populations and the GBS-

typed Kerguelen population was performed with *TreeMix* v.1.1 (Pickrell & Pritchard 2012).

TreeMix jointly estimates population splits and subsequent admixture events based on the F-

statistics introduced by Reich *et al.* (2009), and commonly recognised as a valid support for

admixture (e.g., in humans: Pickrell & Pritchard 2012, Wong *et al.* 2017). A maximum-likelihood

population tree was estimated based on the matrix of GBS allele frequency covariance between

population pairs, and admixture events were sequentially added. To account for linkage

disequilibrium, variants were grouped together in windows of size k=100 SNPs. Trees were rooted

with the two *M. trossulus* populations and no sample size correction (option "-noss") was applied.

We tested for a range of migration events from m=0 to m=12, and looked for an asymptotic value of

the log-likelihood. The number of significant migration events was assessed by stepwise

comparison of Akaike information criterion. Finally, we made 100 bootstrap replicates (option "–

bootstrap") of the maximum-likelihood tree to assess statistical support of migration events.

Based on the GBS genotypes, we additionally performed a model-based clustering analysis of

these populations, which assumes admixed individuals with contributions from *K* panmictic

reference populations. Ancestry of each individual was estimated using the maximum-likelihood

approach implemented in ADMIXTURE v1.23 (Alexander *et al.* 2009). We ran 50 replicates for a

number of clusters from K=2 to K=8 and chose the maximum log-likelihood run for each K. We

also performed a supervised ADMIXTURE analysis (in which the reference populations were

provided) on the Kerguelen individuals with the KASPar SNPs (K=2 clusters and 50 replicates). We

defined *M. edulis* and the *M. platensis* Chilean mussels as ancestral populations from which the

Kerguelen individuals derive their ancestry. Individual ancestries are provided in Text S3 for the

GBS analysis and Text S4 for the KASPar analysis.

In complement to the *TreeMix* analysis, we used a model-based approach implemented in ∂a∂i

v1.6.3 (Gutenkunst *et al.* 2009) to explicitly test for the presence of gene flow. This method has

proven useful for distinguishing ancestral shared polymorphism between recently-diverged species

evolving under strict isolation from a scenario including migration between them (e.g., in

Arabidopsis: Hubert *et al.* 2014; sea bass: Tine *et al.* 2014; poplars: Christe *et al.* 2017; whitefish:

Rougeux *et al.* 2017). We assessed the divergence history between Kerguelen mussels and Northern

species in a pairwise manner based on their folded joint site frequency spectrum at the cDNA

contigs (provided in Text S5 in ∂a∂i format, and plotted in Figure S1): Kerguelen vs. *M. edulis* (represented by the European sample "EU – peripheral"); Kerguelen vs. *M. galloprovincialis* (Mediterranean sample "MED – west"); Kerguelen vs. *M. trossulus* (European sample "EU"). We defined fourteen demographic models following previous studies (Tine *et al.* 2014; Christe *et al.* 2017) to test: (i) the timing of gene flow during divergence (absence of gene flow "SI", continuous migration "IM", secondary contact "SC", ancient migration "AM", periodic secondary contact "PSC" with two periods of current gene flow, periodic ancient migration "PAM" with two periods of ancient migration, and a three-period model of ancient migration, isolation and current gene flow "AMSISC"); (ii) the genomic heterogeneity in gene flow (presence "2M" or absence of interspecific genomic barriers); (iii) the genomic heterogeneity in effective population size (presence "2N" or absence of Hill-Robertson effects). All models began with the split of the ancestral population in two daughter populations, and then were followed by divergence in the absence or presence of gene flow. Each model was fitted to the observed joint site frequency spectrum (singletons were masked) using three successive optimization steps: "hot" simulated annealing, "cold" simulated annealing and BFGS -Broyden–Fletcher–Goldfarb–Shanno- algorithm (Tine *et al.* 2014). Model comparisons were made using Akaike information criterion. A summary of the models is given in Table S5 and the script that defines the models in ∂a∂i is given in Text S6.

Topology weighting of the *Mytilus* spp.

The distinct haplotype loci of the GBS dataset were also individually analysed with the neighbour-net method. Allele genealogies were inferred with the R package APE (Paradis 2010) using a neighbour-joining algorithm with F84 distances (Felsenstein & Churchill 1996). Haplotype loci were filtered based on the following excluding criteria: scale < 0.00005 ; 0.00005 =< scale < 0.0005 & length < 10000 bp ; 0.0005 =< scale < 0.001 & length < 5000 bp ; and scale >= 0.001 & length >= 1000 bp, where "scale" is the scale of the gene tree and "length" is the length of the sequence.

Neighbour-joining trees of the 395 retained sequences are available in Text S7 and their length are

indicated in Table S6 (4.5 kb in average, a minimum length of 1 kb and a maximum length of 25

kb).

For each haplotype locus, the relationships between the Northern species and the Kerguelen

population were then quantified using *Twisst*, a tree weighting approach that has been successfully

applied to *Heliconius butterflies* to evaluate the support of different phylogenies around colour

pattern loci (Van Belleghem *et al.* 2017). We tested the three possible unrooted topologies: (A) *M.*

edulis grouped with the Kerguelen population; (B) *M. galloprovincialis* grouped with the Kerguelen

population and (C) *M. trossulus* grouped with the Kerguelen population. Their exact weightings to

the full tree were estimated by considering all subtrees ("complete method"). Only contigs with a

resolved topology were analysed: 67 contigs for which one topology had a weight greater or equal

to 0.75. These topologies were further classified in two categories depending on whether they most

plausibly reflect: (i) ancient divergence of the Kerguelen population (i.e. the Kerguelen and

Northern individuals clustered into distinct monophyletic groups) or, (ii) introgression with one of

the Northern species (i.e., the Kerguelen individuals were distributed within one or more Northern

clades); "na" stands for topologies that we were unable to classify in these two categories due to a

lack of informative sites. Tree topology weightings and classification are available in Table S6.

Analyses of genetic variation in the Kerguelen Islands

For each KASPar SNP, estimation of F_{ST} values (Weir & Clark Cockerham 1984) was calculated

over all sampling sites (Table S2), and in a pairwise manner across all SNPs (Table S7) using

Genetix 4.05 (Belkhir *et al.* 2002). Their significance was tested by a permutation procedure (1000

permutations) and adjusted with the Bonferroni's correction for multiple comparisons (Benjamini &

Hochberg 2000).

Analysis of habitat variables in the Kerguelen Islands

To evaluate how much of the genetic variation among sites was explained by local environmental factors, we used redundancy analysis (RDA), a constrained ordination method implemented in the R package vegan (Oksanen *et al.* 2017). It performs a multiple linear regression between a matrix of response variables (individual genotypic data) and a matrix of explanatory variables (environmental factors). Notably, the effect of partially confounded explanatory variables can be estimated separately. RDA is commonly used to estimate the relative contribution of spatial and environmental components on species communities, and it has been recently applied to analysis of population genetic structure (e.g., Legendre & Fortin 2010). Geographic coordinates and five qualitative factors were measured in each site to describe the local habitat (Table S3): (i) Substrate (rock: R, blocks: B, gravels: G, or sand: S); (ii) Wave Exposure (sheltered: Sh, or exposed: E); (iii) Slope (flat: F, steep: St, or hangover: H); (iv) Salinity (oceanic water: OW, or low-salinity water: LSW); (v) *Macrocystis* (presence: P, or absence: A).

We specifically tested the effect of each of these constrained factors (explanatory variables) on the distribution of genotypes at the 33 KASPar SNPs (response variables). The following initial model was used: Genotypes ~ *Macrocystis* + Salinity + Slope + Exposure + Substrate + Longitude + Latitude. The significance of the global model was first established by permutation test, in which the genotypic data were permuted randomly and the model was refitted (1000 permutations). Marginal effect permutation tests were then performed to assess the significance of each factor by removing each term one by one from the model containing all other terms (1000 permutations). Non-significant factors were removed from the final model. Based on that simplified model, we performed a conditioned RDA analysis for each factor to assess its contribution to the genotypic variance independently from the other explanatory variables. These co-variables were removed from the ordination by using a condition function: Genotypes ~ tested variable + condition(all other variables). Finally, we performed a conditioned RDA on geography to specifically control its confounding effect: Genotypes ~ significant environmental variables + condition(significant

307 geographic variables).

Simulations of secondary contact

Following the methodology of Gagnaire *et al.* 2015, we modelled a secondary contact between two

semi-isolated genetic backgrounds (*M. edulis* vs. *M. platensis* Chilean population) that meet twice

on a circular stepping stone model of 30 demes (between demes n°18 / n°19 and demes n°19 /

n°20), and start to exchange genes. At generation zero, the *M. edulis* background settles in deme

n°19 while the *M. platensis* background is located everywhere else. Their initial allele frequency at

the barrier loci was set-up to 0.10 for the *M. edulis* background and 0.70 for the *M. platensis*

background, which correspond to the average foreign allele frequency at the four most

differentiated loci observed in the *M. edulis* and Chilean *M. platensis* mussels, respectively (see

Figure 4B). The auto-recruitment rate was set to 1-*m*, and migration to adjacent demes was *m*/2

(with *m*=0.5). A barrier to dispersal was set between demes n°18 and n°19 (*m*=0.05), which

corresponds to the genetic break observed between sites PAF and RdA in the Kerguelen Islands.

Selection acts in haploid individuals on a two-locus incompatibility, which is linked to a neutral

marker located 1cM away and unlinked to a second neutral marker. Strong and asymmetric

selection were required to maintain the two genetic backgrounds after contact (s=0.5 against the *M.*

platensis allele in the *M. edulis* background vs. s=0.2 against the *M. edulis* allele in the *M. platensis*

background). Deme size was constant and set to 500 individuals.

Results

The Kerguelen mussels: signal of divergence of a Southern lineage after transoceanic

migration and admixture with Northern lineages

328 An individual genetic network (Figure 1) was built from a subset of 51,878 high-quality SNPs

genotyped in nine Northern populations and eight individuals from the Kerguelen Islands. We observed that the Northern populations formed three distinct clusters, corresponding to the three Northern species: *M. edulis*, *M. galloprovincialis* and *M. trossulus*. Accordingly, the majority of SNPs fixed between populations (295 in total) were species-specific: *M. edulis*=6, *M. galloprovincialis*=62 and *M. trossulus*=224. The Kerguelen individuals clustered together into a single clade. Indeed, the proportion of SNPs which were private to the Kerguelen Islands amounted to 60% (3,805 private for a total of 6,297 SNPs in Kerguelen, after removing singletons). In comparison, the number of private SNPs in *M. trossulus* was 3,070, and it was only 492 in *M. galloprovincialis* and 48 in *M. edulis* (indicative of introgression between the two latter species). Among the 2,492 SNPs shared by the Kerguelen mussels with Northern species, 33% (830) were highly differentiated between at least two Northern species. When considering Northern species-specific SNPs, 83% of those fixed in *M. edulis* were segregating in the Kerguelen (5 for a total of 6 fixed). These numbers were 16% for *M. galloprovincialis* (10 for a total of 62 fixed) and 12% in *M. trossulus* (27 for a total of 224 fixed). A multivariate analysis on KASpar-typed SNPs, including the Northern samples, the Kerguelen Islands and other samples from the Southern Hemisphere that were also genotyped in our SNP assay, is provided as a supplementary figure (Figure S2). The principal component analysis clearly shows that the Chilean mussels (MAU) group with the Kerguelen mussels in accordance with them being both named *M. platensis*; while the Australasian samples (Australia, Tasmania and New-Zealand), usually named *M. planulatus*, cluster with the Northern *M. galloprovincialis*. These findings corroborate previous results based on mitochondrial DNA (Gérard *et al.* 2008) and nuclear markers (Borsa *et al.* 2012).

The species relationships found in the genetic network (Figure 1) were generally supported by the maximum-likelihood population tree inferred by *TreeMix* (Figure 2A), except that the Kerguelen population was inferred as the sister-group of *M. edulis*. The pairwise population residuals in a model without admixture (Figure S3) suggested substantial migration between species. So, we

sequentially allowed from 0 to 12 migration events in the analysis, and assessed their significance

by stepwise comparison of Akaike information criterion (Figure S3). The best fit to the data was

obtained with seven migration events, which significantly improved the log-likelihood of the model

(Figure S3). This population tree was bootstrapped 100 times to assess statistical support of

migration events. They were generally weakly supported, with only four migration events that had

more than 50% bootstrap support (Figure 2A and Table S8). The most robustly inferred migration

event was between the Mediterranean *M. galloprovincialis* and the Kerguelen population (81 % of

bootstrap replicates). The two others included migration among Northern species as expected: the

European populations of *M. edulis* and *M. galloprovincialis* (51%), and the European populations of

M. edulis and *M. trossulus* (68%). Migration events were also inferred between the Mediterranean

M. galloprovincialis and the European *M. trossulus* (55%), as well as between the Kerguelen

population and the American *M. trossulus* (38%). Reticulated evolution with the American *M.*

trossulus was further supported by their excess of privately shared polymorphism with the

Kerguelen population (Figure 2B). Migration between the Kerguelen population and the European

M. edulis was detected, but only in 4% of bootstrap replicates.

Our reconstruction of the divergence history with ∂a∂i consolidated evidence for admixture

between the Kerguelen mussels and each Northern species, although pointed out to a periodic

connectivity with several periods of admixture. In all three comparisons, the models without gene

flow were consistently the worst supported ("Strict Isolation", Table S5). In the *M. edulis* and *M.*

galloprovincialis comparisons, the model of periodic ancient migration with varying rates of

introgression among loci ("*PAM_2M*") received the strongest statistical support (Table S5).

Migration occurred right after the split as well as later in the divergence during a relatively short

period (~15% to 25% of the total divergence time) and it was asymmetric with a substantial fraction

(85% to 90%) of the mussel genome in the Kerguelen permeable to *M. edulis* and *M.*

galloprovincialis gene flow. In the *M. trossulus* comparison, the three-period model with

homogeneous rates of introgression among loci ("AMSISC") was the best supported (Table S5). A

first period of ancient migration lasted ~85% of the divergence time, and was followed by a short

period of isolation and a very short secondary contact (~2% of the total divergence time). Kerguelen

mussels were mostly resistant to *M. trossulus* introgression (Table S5). Overall, these results suggest

that the Kerguelen mussel is a Southern lineage related to *M. edulis* and that it admixed with all

three Northern species (*M. edulis*, *M. galloprovincialis* and to a lesser extent with *M. trossulus*) in

the past, either after secondary contact and/or early in the divergence of the species.

Variation of admixture histories across the genome

To further investigate how genetic relationships varied across the genome, we quantified the

contribution of three unrooted topologies (Figure 3) to the full tree at 395 GBS contigs with *Twisst*.

Only 17% (67) of them showed resolved relationships, i.e. one of the unrooted topology weighted

75% or more, among which 40% (27) were highly resolved (weight >= 90%). The reason for this

low rate of resolved relationship is twofold: (i) ongoing sorting of shared ancestral polymorphism,

and (ii) introgression. In the Northern Hemisphere introgression has been shown to be the main

process thanks to model fitting (Fraïsse *et al.* 2018) and using comparison of introgression

differential between populations within species (Fraïsse *et al.* 2016). It is more difficult to tell apart

ancestral shared polymorphism from introgression for the single Kerguelen population. Above

analyses suggest introgression is contributing although it is difficult to evaluate its contribution. The

most represented resolved topology (39 contigs) put the Kerguelen individuals together with *M.*

edulis, while they were grouped with *M. trossulus* in 19 contigs (i.e. ancestral to the *M. edulis* / *M.*

galloprovincialis subgroup) and with *M. galloprovincialis* in 9 contigs (Table 1). This illustrates a

high rate of ancient incomplete lineage sorting that resolved into alternative topologies after the

consecutive speciation events, which resulted in the three species *M. edulis*, *M. galloprovincialis*

and *M. platensis*. In other words, the transequatorial migration that leads to *M. platensis* occurred

soon after the split between *M. edulis* and *M. galloprovincialis*.

When classifying the topologies in categories, 18 contigs supported the "ancient Kerguelen

divergence" scenario while 19 supported an "introgression" scenario among which 4 were from *M.*

trossulus, 12 from *M. edulis* and 3 from *M. galloprovincialis*; 30 contigs could not be classified. It

should be noted that all these patterns hold when using a minimal weight of 90% (Table 1). Figure 3

illustrates representative cases of the different *Twisst* categories, including candidate loci for

introgression (A2, B2 and C2 in Figure 3). These results suggest that the Kerguelen mussels have a

genome of mixed ancestry, mainly dominated by *M. edulis,* from which they are closely-related, but

with which they also have probably secondarily admixed. This is in accordance with the *TreeMix*

analysis where the Kerguelen population was inferred to be the sister-clade of *M. edulis* and

experienced admixture events with *M. edulis* and *M. galloprovincialis*. However, this seems

quantitatively different from the $\partial a \partial i$ results in which both *M. edulis* and *M. galloprovincialis*

broadly contributed to the genomic composition of the Kerguelen mussels. However, we call for

caution when comparing these two methods as $\partial a \partial i$ provides genome-wide migration rate estimates,

while *Twisst* relies on counting different types of topologies across the genome and many gene trees

could not be classified, and *TreeMix* infer single admixture event under the hypothesis of

homogeneous introgression (while *Mytilus* species barriers are often semi-permeable, Roux *et al.*

2014, Fraïsse *et al.* 2018, and see Table S5).

Substantial genetic structure in the Kerguelen Islands

Mussels were collected from 35 sampling sites all around the Kerguelen Islands (Figure 4A, Table

S3) and successfully genotyped at 33 KASpar SNPs. Pairwise F_{ST} values across all SNPs (Table S7)

revealed significant fine-scale genetic differentiation between sites from different geographic

regions. Remarkably, RdA (North-East) and PCu (West) were significantly differentiated with

nearly all other sites. Sites from the South, especially BdS, and from the North, especially AS, were

differentiated from the Gulf of Morbihan. At a smaller scale within the Gulf of Morbihan, genetic

structure among several sites was detected, but their significance level did not pass the correction

for multiple tests. These results extend the study by Gérard *et al.* (2015) to many SNPs and

substantiate their finding of significant genetic differentiation at different scales in the island.

Global F_{ST} across all sites was calculated for each SNP and tested with 1000 permutations (Table

S2). Values were non-significant after Bonferroni's correction, except at the three most

differentiated loci, which are ancestry-informative: X10, X11 and X57. Their foreign allele, defined

based on its frequency in the Northern species (*M. galloprovincialis* Atlantic population of Iberian

Coast, Table S4), was at low frequency in the North of the island, especially in the North-East sites,

RdA and PMo. In contrast, it was at higher frequency in the Gulf of Morbihan and at intermediate to

low frequency in the South and West. These trends were similar to those at Glu-5' (Table S4), a

nuclear marker suspected to be affected by differential selection in the island (Gérard *et al.* 2015)

and at candidate allozymes although with fewer sampling sites (Blot *et al.* 1989). Across all sites,

the frequencies of the foreign allele at Glu-5' were significantly correlated with those at X10

(r=0.61, p-value < 0.001), X11 (r=0.419, p-value=0.012), and X57 (r=0.49, p-value=0.003), but

they were globally higher at Glu-5' (Figure S4).

The foreign allele frequency at those four loci is represented in Figure S4 and the average over

the four loci in Figure 4B (filled symbols). These clearly show a genetic break between two

geographically close sites, PAF and RdA (40 kms apart), and to a lesser extent between RdA and

Pmo, which are separated by only 8 kms of coasts. The average frequency was the highest in the

Gulf of Morbihan (from HdS to PAF), then it abruptly dropped down (in 40 km) between PAF and

RdA (respectively on the West and East coast of the Prince of Wales' Peninsula), and finally

increased gradually along the coast from North-East to South-West. Notably, the variance in allele

frequency at the four loci was weak (0.0208 in average across localities). This is in sharp contrast

with the pattern observed at the other loci (open symbols) of which the average frequency remained

similar across all sites (Figure 4B), and variance was stronger (0.0938 in average across localities).

An admixture analysis using all KASPar SNPs and defining *M. edulis* and the Chilean mussels as

reference populations (Figure 4B, pie charts), suggests that the Kerguelen Island is occupied by

mussels related to Chilean mussels (*M. platensis*), and that RdA has by far the highest level of *M.*

edulis ancestry (31% compared to <19% elsewhere). We therefore hypothesize that two genetic

backgrounds may be present in the island, one related to *M. edulis* and trapped at site RdA close to a

potential density trough in the the Prince of Wales' Peninsula, and the other related to Chilean

mussels and present everywhere else. This pattern is theoretically expected (Barton 1979, Barton &

Hewitt 1985) as areas of low density or low carrying capacity tend to trap "tension zones" between

incompatible genetic backgrounds (similarly to ecological boundaries).

We illustrated this scenario by simulating a secondary contact between these two backgrounds

(including a physical barrier to dispersal between demes n°18 and n°19), and tracking the frequency

of the foreign allele at two neutral markers (linked and unlinked to the barrier locus) a few hundreds

generations after the contact (Figure 4C). Simulations often fitted well with the observed variation

in allele frequency across sites, as predicted by Gagnaire *et al.*'s model (2015), provided that the

species barrier was asymmetric in order to protect *the small* M. edulis patch to be quickly swamped

by *M. platensis* introgression. This suggests that the genetic break at the boundary of the Gulf of

Morbihan and the North-East region is better revealed by the frequency of foreign alleles at

ancestry-informative loci implying a role of admixture either in the maintenance or in the detection

of the genetic structure.

Environment-associated genetic structure in the Kerguelen Islands

We then tested for genetic-environment associations in the Kerguelen by performing a RDA.

Among the seven constrained factors (five qualitative variables, plus geographic coordinates), three

were not significant in the initial model (Salinity, Exposure and Latitude, Table S9) and were

removed from further analyses. The proportion of total genotypic variance explained by all

constrained factors was highly significant in the global model (p-value=0.001, Table 2A, left panel), but quite low (2.32%). The first RDA axis, which explained 61% of the constrained variance, was mainly contributed by *Macrocystis* presence/absence (Figure S5 and Table S10). Accordingly, it was the only factor whose marginal effect remained significant (p-value=0.032, Table 2B, left panel).

We statistically controlled for the effect of geography by performing a conditioned RDA analysis on Longitude (Table 2A, right panel). The combined effect of the three environmental variables remained significant (p-value=0.041), explaining 1.1% of the total genotypic variance. Individually, *Macrocystis* presence/absence and Substrate still showed significant effects, after removing all other confounding factors (p-value=0.011 and 0.001, respectively, Table 2B, right panel). This suggests a limited effect of geography on the genetic structure of the mussels, although coastal distance would be a better proxy to account for spatial structure in the island. Interestingly, it has been previously shown that the fine-scale genetic variation at Glu-5' was also significantly associated with *Macrocystis* presence/absence (Gérard *et al.* 2015). Accordingly, we found a significant correlation between the average foreign allele frequency at the four most differentiated loci in the Kerguelen Islands and the presence/absence of *Macrocystis* (Figure 5A), whereas there was no correlation with the other loci (Figure 5B).

The sharp genetic break between RdA and PAF further indicates that two genetic backgrounds may have been locally trapped by an ecological boundary or a region of reduced dispersal (Bierne *et al.* 2011). Accordingly, there is an oceanic threshold at the entrance of the Gulf of Morbihan that impedes exchanges with water masses from outside; and at a larger scale, the Antarctic circumpolar current moves the water masses from West to East causing gyres and turbulences on the North-Eastern coast and pushing water masses far to the East (Karin Gerard, pers. comm.). Thus, the water masses between the Gulf of Morbihan and the North Coast do not mix well, suggesting that exchanges between the two sites are limited. Moreover, these two sites differ at all five ecological variables (Table S3), but not in the direction predicted by their geographical origin: RdA shows an

 habitat characteristic of the Gulf of Morbihan while being located on the East coast and having the

 lowest foreign allele frequency (and the reverse is true for PAF). This imperfect association between

 genotypes and habitats supports the hypothesis that enhanced genetic drift and intense gene flow in

 the island grambled the signal of genetic-environment association at our markers rather than

 environmental differential selection.

507 **Most differentiated SNPs in the Kerguelen Islands are primarily ancestry-informative in the**

508 **Northern Hemisphere**

509 In the total sample, the average allele frequency of the foreign allele was 0.417 at Glu-5', 0.503 at

510 X10, 0.619 at X11 and 0.480 at X57. These polymorphisms were well-balanced in the island (i.e.

511 close to 0.5 frequency), despite being species-specific in the Northern species (Table S2, also see

512 Gérard *et al.* 2015 for Glu-5'). To investigate whether micro-geographic structure in the island was

513 primarily depending on Northern ancestry-informative loci, we compared the degree of

514 differentiation between sites in the Kerguelen Islands and that of the Northern species, *M. edulis*

515 and *M. galloprovincialis*, at the 33 KASPar SNPs (Figure 6). Figure 6A shows that the level of

516 genetic differentiation among sites in the Kerguelen (global F_{ST}, Table S2) was significantly higher

517 (p-value=0.021) for the ancestry-informative loci (mean=0.015, in orange) compared to the

518 background loci (mean=0.007, in grey). Importantly, the difference between the two categories was

519 also significant when considering the genetic-by-environment association across all variables

520 (Figure 6B: mean_orange=0.112, mean_grey=0.048, p-value=0.006), which was measured by the

521 locus coordinates on the first axis of the conditioned RDA (Table S11); or only including

522 *Macrocystis* presence/absence (Table S12: mean_orange=0.089, mean_grey=0.041, p-value=0.016).

523 Moreover, these patterns hold when adding the locus Glu-5' (Figure S6) in the case of genetic

524 differentiation (Figure S6A, p-value=0.01), and genetic-environment associations (Figure S6B, p-

525 value=0.004) measured by the F_{CT} from an AMOVA analysis performed by grouping sites according

526 to the presence/absence of *Macrocystis* (see Gérard *et al.* 2015).

Discussion

In this work, we first analyzed the genetic relationship of the Southern Hemisphere Kerguelen

mussels with the three Northern species (*M. edulis*, *M. galloprovincialis* and *M. trossulus*) at 1,269

contigs (51,878 SNPs), and we confirmed with this large dataset that mussels in the Kerguelen

Islands belong to a Southern lineage (*M. platensis*). We further showed that the Kerguelen

population is a sister clade of *M. edulis*.

Our population tree inference provides evidence of genetic exchanges with Northern mussels that

occurred after the first establishment in the Southern Hemisphere. This was confirmed by

reconstructing their divergence history, which further suggested that introgression occured at

variable rates across the genome with some genomic regions resistant to gene-flow while others are

essentially permeable. The resulting genome-wide ancestry variation was estimated by applying a

new topology weighting method to each GBS sequence (Martin & Van Belleghem 2016), which

weighted the contribution of three topologies to the full tree. The majority of the genome showed

evidence for both ancient incomplete lineage sorting, which resolved into alternative topologies

among the four species studied *(M. trossulus*, *M. edulis*, *M. galloprovincialis* and *M. platensis*), and

shared polymorphisms due to ongoing incomplete lineage sorting and introgression among North

Hemisphere species and with the Southern lineage, and this resulted in only 17% of the regions with

a well-resolved topology. In 51% of these resolved regions, we found clear evidence of admixture,

i.e., the Kerguelen haplotypes were all (or part of) nested within a Northern clade. We thus

confirmed that the reticulated evolution of the Southern Hemisphere Kerguelen mussels, which was

suggested by a handful of nuclear markers (Borsa *et al.* 2007) and mitochondrial DNA (Hilbish *et*

al. 2000; Gérard *et al.* 2008), holds at a genome-wide scale. This is in line with recent genomic

studies (e.g., *Anopheles gambiae* mosquitoes : Fontaine *et al.* 2015 ; *Xiphophorus* fishes : Cui *et al.*

2013 ; African lake cichlids : Meier *et al.* 2017 ; Caribbean *Cyprinodon* pupfishes: Richards &

Martin 2017 ; *Heliconius* butterflies : Martin *et al.* 2013) that recognized a prominent role of

introgressive hybridization as a source of reticulate phylogenies, besides ancestral shared

polymorphism.

At some GBS loci, Kerguelen mussels possessed alleles characteristic of both *M. edulis* and *M.*

galloprovincialis or *M. trossulus* indicating polymorphism for Northern species-specific alleles in

the Kerguelen. Importantly, these loci did not depart from Hardy-Weinberg and linkage equilibrium

as shown by an ADMIXTURE analysis (Figure S7) in which the Kerguelen mussels appeared as a

well demarcated panmictic cluster. Therefore, contrary to what is known in Northern hybrid zones

(Bierne *et al.* 2003), there is no evidence of reproductive barriers impeding admixture in the

Kerguelen Islands. Several hypotheses can be proposed: (i) a weaker reproductive barrier between

Northern backgrounds at the time of contact in the South; or (ii) an insufficient barrier to gene flow

under the demographic and environmental conditions, specifically strong genetic drift, high-

potential for hybridization in this small isolated island, or a strong demographic asymmetry between

the native and the introduced populations. The first hypothesis relies on the importance of

Dobzhansky-Müller incompatibilities for reproductive isolation (Coyne & Orr 2004), and may

explain how *M. edulis*, *M. galloprovincialis* and *M. trossulus* alleles at different loci can co-exist

into a single Southern population that did not evolve their incompatible interactors, as opposed to

the Northern populations. The second hypothesis is based on the fact that the outcome of

hybridization can be highly dependent on the demographic context.

Finally, by collecting mussels from contrasted habitats, we demonstrated significant

differentiation across 35 sampling sites at the scale of the island, and at a smaller scale between

geographically close sites, especially within the Gulf of Morbihan. As found by Gérard *et al.* (2015)

at Glu-5', and previously at allozyme loci (Blot *et al.* 1989), the strongest structure was observed

between the North-South coasts and the Gulf of Morbihan, with a genetic break between RdA and

PAF at the three most differentiated loci. The fine-scale genetic structure observed in such a high-

dispersal species as mussels is at first sight at odds with selective neutrality. So, we explicitly tested

the role of habitat heterogeneity in explaining this differentiation. Our RDA analysis shows that

genetic variation was associated with habitats, even after controlling for spatial effects; and the most

important factors were the presence of *Macrocystis* kelps, substrate type and slope. Despite being

low, this significant habitat-driven genetic differentiation could suggest a role of selection. Another

caveat is that spatial distance between sampling sites may be better described by oceanographic

distance than geographic coordinates, and so we may be overestimating the influence of habitats.

Firstly, genetic-environment associations could be due to local adaptation of the mussels opposed

by gene flow between habitats. Accordingly, we observed a significant correlation between the

presence/absence of *Macrocystis* and the average foreign allele frequency at the four most

differentiated loci. This points toward a primary effect of *Macrocystis* presence/absence, which is a

keystone species in marine ecosystems that forms kelp forest serving as substrate and refuge for

many molluscs species, including *Mytilus* (Adami & Gordillo, 1999), in areas exposed to wave

action. Nevertheless, the RdA site, which has the lowest foreign allele frequency, is not occupied by

Macrocystis kelp, weakening this local adaptation hypothesis.

Alternatively, the consistent genetic patterns observed across several physically unlinked loci

indicate the possible existence of two genetic backgrounds maintained at the scale of the island. We

propose, and illustrate by simulations, that a genetic background related to *M. edulis* is trapped at

RdA and surrounded by another genetic background related to Chilean *M. platensis* mussels and

present everywhere else. The enclosed location of the genetic background at RdA explains that it is

strongly introgressed at most markers and thus hard to detect. However, the physical barrier to

dispersal between sites PAF and RdA produces a clear genetic break on the West side of the contact.

Introgression between the two backgrounds generates gradients in allele frequencies, which are

better correlated with habitat variation than geographical distance. The foreign allele (as defined by its frequency in the *M. galloprovincialis* Atlantic population of Iberian Coast) tended to be at higher frequency in shallow sites sheltered from the influence of open marine waters with a low salinity and flat-sandy bottoms, mainly in the inner part of the Gulf of Morbihan. These sites are characterized by an absence of *Macrocystis* kelp beds, as opposed to exposed rocky shores. Port aux Français (PAF) is also the harbour where ships arrive and it is the best place for the arrival of non-native genetic backgrounds. Interestingly, *M. galloprovincialis* alleles are found more frequent in exposed, rather than sheltered sites in the hybrid zone between *M. edulis* and *M. galloprovincialis* in Europe, which would suggest inverted genetic-environment associations between hemispheres. Inverted genetic-environment associations are predicted by the coupling hypothesis (Bierne *et al.* 2011), which proposes that genetic-environment associations can easily be revealed by intrinsically maintained genetic backgrounds in linkage disequilibrium with local adaptation genes, and that the phase of the disequilibrium can reverse when contacts are replicated as could have happened in Southern Hemisphere mussels. Overall, these findings reinforce the idea that genetic variation can be maintained at fine geographical scales in high-dispersal organisms, as recently shown in Chilean mussels (Araneda *et al.* 2016) or in passerine birds (Szulkin *et al.* 2016, Perrier *et al.* 2017). In these examples however the link with a possible history of admixture has not been investigated.

Although we had the hypothesis that admixture events could be an explanation of the micro-structure observed in the Kerguelen (Gérard *et al.* 2015), we could not know which backgrounds were in interaction on the sole basis of the GBS data of a single sample. However, our procedure of identifying SNPs that were both polymorphic in the Kerguelen and highly differentiated between Northern Hemisphere species proved to be an interesting way to enrich for loci able to reveal the micro-geographic structure in the Kerguelen. Luckily the sample we used for the GBS analysis (sample "ker-GBS") was localised in the introgression cline, and this can also explain why the enrichment procedure was successful. This is exemplified with the genealogy around locus X10

(Figure S8), which shows that a SNP that differentiates *M. edulis* from other Northern species, and

was polymorphic in the Kerguelen Islands, is able to reveal the cline of introgressed heterospecific

allele we observed in the island.

In this work, we showed that the most differentiated SNPs in the Kerguelen and those that most

strongly drive the genetic-environment associations are primarily ancestry-informative, suggesting

that maintenance of genetic differentiation at a small spatial scale, and possibly adaptation to fine-

scale environmental variations in the island, may have been facilitated by admixture and

introgression of alleles from Northern species. These foreign alleles may have adaptively

introgressed the Southern background in the Kerguelen, but the signal is probably erasing because

of recombination between adaptive alleles and our neutral markers, and is also probably further

blurred by genetic drift. A central question is whether admixture is a simple source of variation on

which local selection can effectively act or if the initial linkage disequilibria between foreign alleles

in the donor background are required for the successful emergence of micro-geographic adaptation

(or speciation in the case of cichlids, Meier *et al.* 2017) and are maintained rather than built-on. In

the case of Kerguelen mussels, the evidence we gained here for the maintenance of linkage

disequilibrium are limited and indeed rather support extensive recombination. However our markers

have likely lost too much signal to answer the question. Maybe local adaptation is operating at loci

linked to the candidate SNPs, but most probably these markers simply better reveal a genome-wide

signal of habitat constrained connectivity (Gagnaire *et al.* 2015). Overall, our work underlines the

opportunity of using non-equilibrium introgression clines to assess genetic connectivity in natural

populations and warns against systematically interpreting genetic-environment association as signal

of local adaptation.

Acknowledgements

Jean-Pierre Féral and Christian Marschal provided the eight Kerguelen specimens used for GBS during the PROTEKER campaign (Programme IPEV n°1044). Other Kerguelen samples (>600) were collected during scientific program IPEV-MACROBENTHOS n° 195 (1999-2003) by the technical volunteers (VATs) from IPEV missions 49–53 in Kerguelen. This work was funded by the Research Network GDR 3445 cnrs ifremer MarCo and the Agence Nationale de la Recherche (HYSEA project, ANR-12-BSV7-0011). This is article 2015-130 of Institut des Sciences de l'Evolution de Montpellier.

References

Adami ML, Gordillo S (1999) Structure and dynamics of the biota associated with *Macrocystis pyrifera (*Phaeophyta) from the Beagle Channel, Tierra del Fuego. *Sci. Mar.*, 63(S1), 183–191.

Alexander DH, Novembre J, Lange K (2009) Fast model-based estimation of ancestry in unrelated individuals. *Genome Res.*, 19, 1655–1664.

Araneda C, Larraín MA, Hecht B, Narum S (2016) Adaptive genetic variation distinguishes Chilean blue mussels (*Mytilus chilensis*) from different marine environments. *Ecol. Evol.*, 6, 3632–3644.

Barton NH, Hewitt GM (1985) Analysis of hybrid zones. *Annu. Rev. Ecol. Evol. Syst.*, 16(1), 113–148.

Barton NH (1979) The dynamics of hybrid zones. *Heredity*, 43(3), 341.

Belkhir K, Borsa P, Chikhi L, Raufaste N, Bonhomme F (2002) GENETIX 4.03, Logiciel Sous Windows Pour la Génétique Des Populations. Laboratoire Génome, Populations, Interactions: CNRS UMR. 5000. Université Montpellier 2, Montpellier, France.

Benjamini Y, Hochberg Y (2000) On the Adaptive Control of the False Discovery Rate in Multiple Testing with Independent Statistics. *Journal of educational and Behavioral Statistics*, 25, 60–83.

Bierne N, Welch J, Loire E, Bonhomme F, David P (2011) The coupling hypothesis: why genome scans may fail to map local adaptation genes. *Mol. Ecol.*, 20, 2044–2072.

Bierne N, Borsa P, Daguin C, Jollivet D, Viard F, Bonhomme F, David P (2003) Introgression

patterns in the mosaic hybrid zone between *Mytilus edulis* and *M. galloprovincialis*. *Mol.*
Ecol., 12, 447–461.

Blot M, Thiriot-Quiévreux C, Soyer J (1988) Genetic relationships among populations of
Mytilus desolationis from Kerguelen, *M. edulis* from the North Atlantic and *M.*
galloprovincialis from the Mediterranean. *Mar. Ecol. Prog. Ser.*, 44, 239–247.

Blot M, Thiriot-Quivreux C, Soyer J (1989) Genetic differences and environments of mussel
populations in Kerguelen Islands. *Polar Biol.*, 10, 167–174.

Borsa P, Rolland V, Daguin-Thiébaut C (2012) Genetics and taxonomy of Chilean smooth-
shelled mussels, *Mytilus spp.* (Bivalvia: Mytilidae). *Comptes rendus biologies*, *335*(1), 51–
61.

Borsa P, Daguin C, Bierne N (2007) Genomic reticulation indicates mixed ancestry in Southern-
Hemisphere *Mytilus* spp. mussels. *Biol. J. Linn. Soc.*, 92, 747–754.

Browning SR, Browning BL (2007) Rapid and accurate haplotype phasing and missing-data
inference for whole-genome association studies by use of localized haplotype clustering.
Am. J. Hum. Genet., 81, 1084–1097.

Christe C, Stölting KN, Paris M, Fraïsse C, Bierne N, Lexer C (2017) Adaptive evolution and
segregating load contribute to the genomic landscape of divergence in two tree species
connected by episodic gene flow. *Mol Ecol*, 26, 59–76.

Cowen RK, Sponaugle S (2009) Larval Dispersal and Marine Population Connectivity. *Annu.*
Rev. Mar. Sci., 1, 443–466.

Coyne JA, Allen Orr H (2004) Speciation. Sinauer Associates Incorporated.

Cui R, Schumer M, Kruesi K, Walter R, Andolfatto P, Rosenthal GG (2013) Phylogenomics
reveals extensive reticulate evolution in *Xiphophorus* fishes. *Evolution*, 67, 2166–2179.

Felsenstein J, Churchill GA (1996) A Hidden Markov Model approach to variation among sites
in rate of evolution. *Mol. Biol. Evol.*, 13, 93–104.

Fontaine MC, Pease JB, Steele A *et al.* (2015) Mosquito genomics. Extensive introgression in a
malaria vector species complex revealed by phylogenomics. *Science*, 347, 1258524.

Fraïsse C, Roux C, Gagnaire P-A, Romiguier J, Faivre N, Welch JJ, Bierne N (2018) The
divergence history of European blue mussel species reconstructed from Approximate
Bayesian Computation: the effects of sequencing techniques and sampling strategies. *PeerJ*,
6:e5198.

Fraïsse C, Belkhir K, Welch JJ, Bierne N (2016) Local interspecies introgression is the main
cause of extreme levels of intraspecific differentiation in mussels. *Mol. Ecol.*, 25, 269–286.

Gagnaire P-A, Broquet T, Aurelle D, Viard F, Souissi A, Bonhomme F, Arnaud-Haond S, Bierne
N (2015) Using neutral, selected, and hitchhiker loci to assess connectivity of marine
populations in the genomic era. *Evol. App.*, 8, 769–78

Gaitán-Espitia JD, Quintero-Galvis JF, Mesas A, D'Elía G (2016) Mitogenomics of southern hemisphere blue mussels (Bivalvia: Pteriomorphia): Insights into the evolutionary characteristics of the *Mytilus* edulis complex. *Sci. Rep.*, 6, 26853.

Gérard K, Bierne N, Borsa P, Chenuil A, Féral J-P (2008) Pleistocene separation of mitochondrial lineages of *Mytilus* spp. mussels from Northern and Southern Hemispheres and strong genetic differentiation among southern populations. *Mol. Phylogenet. Evol.*, 49, 84–91.

Gérard K, Roby C, Bierne N, Borsa P, Féral JP, Chenuil A (2015) Does natural selection explain the fine scale genetic structure at the nuclear exon Glu-5′ in blue mussels from Kerguelen? *Ecol. Evol.*, 5, 1456–1473.

Goudet J (2005) hierfstat, a package for r to compute and test hierarchical F-statistics. *Mol. Ecol. notes*, 5, 184–186.

Gutenkunst RN, Hernandez RD, Williamson SH, Bustamante CD (2009) Inferring the Joint Demographic History of Multiple Populations from Multidimensional SNP Frequency Data. PLOS Genetics 5(10), e1000695.

Hilbish TJ, Mullinax A, Dolven SI, Meyer A, Koehn RK, Rawson PD (2000) Origin of the antitropical distribution pattern in marine mussels (*Mytilus spp.*): routes and timing of transequatorial migration. *Mar. Biol.*, 136, 69–77.

Huber CD, Nordborg M, Hermisson J, Hellmann I (2014) Keeping it local: evidence for positive selection in Swedish *Arabidopsis thaliana*. *Mol. Biol. Evol.*, 31(11), 3026–3039.

Koehn RK, Bayne BL, Moore MN, Siebenaller JF (1980) Salinity related physiological and genetic differences between populations of *Mytilus edulis*. *Biol. J. Linn. Soc.*, 14, 319–334.

Legendre P, Fortin MJ (2010) Comparison of the Mantel test and alternative approaches for detecting complex multivariate relationships in the spatial analysis of genetic data. *Mol. Ecol. Resour.*, 10(5), 831–844.

Martin SH, Dasmahapatra KK, Nadeau NJ, Salazar C, Walters JR, Simpson F, Blaxter M, Manica A, Mallet J, Jiggins CD (2013) Genome-wide evidence for speciation with gene flow in *Heliconius* butterflies. *Genome Res.*, 23, 1817–1828.

Martin SH, Van Belleghem SM (2016) Exploring evolutionary relationships across the genome using topology weighting. *Genetics*, 206, 429-438.

McDonald JH, Seed R, Koehn RK (1991) Allozymes and morphometric characters of three species of *Mytilus* in the Northern and Southern Hemispheres. *Marine Biol.*, 111, 323–333.

McGuire G, Wright F (2000) TOPAL 2.0: Improved Detection of Mosaic Sequences within Multiple Alignments. *Bioinformatics*, 16(2), 130–134.

Meier JI, Sousa VC, Marques DA, Marques DA, Selz OM, Wagner CE, Excoffier L, Seehausen O (2017) Demographic modelling with whole-genome data reveals parallel origin of similar

Pundamilia cichlid species after hybridization. *Mol. Ecol.*, 26, 123–141.

Oksanen J, *et al.* (2017) vegan: Community Ecology Package. R package version 2.4-2.
https://CRAN.R-project.org/package=vegan

Palumbi SR (1992) Marine speciation on a small planet. *Trends Ecol. Evol.*, 7, 114–118.

Paradis E (2010) pegas: an R package for population genetics with an integrated-modular
approach. *Bioinformatics,* 26, 419–420.

Perrier C, Lozano del Campo A, Szulkin M, Demeyrier V, Gregoire A, Charmantier A (2017)
Great tits and the city: distribution of genomic diversity and gene - environment associations
along an urbanization gradient. *Evol. App.*, doi:10.1111/eva.12580

Pickrell JK, Pritchard JK (2012) Inference of population splits and mixtures from genome-wide
allele frequency data. *PLoS Genet.*, 8, e1002967.

Rawson PD, Hilbish TJ (1995) Evolutionary relationships among the male and female
mitochondrial DNA lineages in the *Mytilus edulis* species complex. *Mol. Biol. Evol.*, 12,
893–901.

Reid NM, Jackson CE, Gilbert D, Minx P, Montague MJ, Hampton TH, Helfrich LW, King BL,
Nacci DE, Aluru N, Karchner SI, Colbourne JK, Hahn ME, Shaw JR, Oleksiak MF,
Crawford DL, Warren WC, Whitehead A (2017) The landscape of extreme genomic
variation in the highly adaptable Atlantic killifish. *Genome Biol. Evol.*, 9(3), 659–676.

Richards EJ, Martin CH (2017) Adaptive introgression from distant Caribbean islands
contributed to the diversification of a microendemic adaptive radiation of trophic specialist
pupfishes. *PLoS Genet.* 13(8), e1006919.

Rougeux C, Bernatchez L, Gagnaire PA (2017) Modeling the multiple facets of speciation-with-
gene-flow toward inferring the divergence history of lake whitefish species pairs
(*Coregonus clupeaformis*). *Genome Biol. Evol.,* 9(8), 2057–2074.

Roux C, Fraïsse C, Castric V, Vekemans X, Pogson GH, Bierne N (2014) Can we continue to
neglect genomic variation in introgression rates when inferring the history of speciation? A
case study in a *Mytilus* hybrid zone. *J. Evol. Biol.*, 27, 1662–1675.

Schmidt PS, Rand DM (1999) Intertidal microhabitat and selection at mpi: interlocus contrasts
in the Northern acorn barnacle, *Semibalanus balanoides*. *Evolution*, 53, 135–146.

Schmidt PS, Serrao EA, Pearson GA, Riginos C, Rawson PD, Hilbish TJ, Brawley SH, Trussell
GC, Carrington E, Wethey DS, Grahame JW, Bonhomme F, Rand DM (2008). Ecological
genetics in the North Atlantic: environmental gradients and adaptation at specific loci.
Ecology, 89, 91–107.

Simon A, Duranton M (2018) Digest: Demographic inferences accounting for selection at linked
sites. *Evolution*, 72(6), 1330–1332.

Skibinski DOF, Beardmore JA, Cross TF (1983) Aspects of the population genetics of *Mytilus*
(Mytilidae; Mollusca) in the British Isles. *Biol. J. Linn. Soc.*, 19, 137–183.

Smith SM, Maughan PJ (2015) SNP genotyping using KASPar assays. *Methods in Mol. Biol.*,
1245, 243–256.

Szulkin M, Gagnaire P-A, Bierne N, Charmantier A (2016) Population genomic footprints of
fine-scale differentiation between habitats in Mediterranean blue tits. *Mol. Ecol.*, 25, 542–
558.

Tine M, Kuhl H, Gagnaire P-A, Louro B, Desmarais E, Martins RST, Hecht J, Knaust F, Belkhir
K, Klages S, Dieterich R, Stueber K, Piferrer F, Guinand B, Bierne N, Volckaert FAM,
Bargelloni L, Power DM, Bonhomme F, Canario AVM, Reinhardt R (2014) European sea
bass genome and its variation provide insights into adaptation to euryhalinity and speciation.
Nat. Comm., 5, 1–10.

Väinölä R, Hvilsom MM (1991) Genetic divergence and a hybrid zone between Baltic and
North Sea *Mytilus* populations (Mytilidae: Mollusca). *Biol. J. Linn. Soc.*, 43, 127–148.

Van Belleghem SM, Rastas P, Papanicolaou A, Martin SH, Arias CF, Supple MA, Hanly JJ,
Mallet J, Lewis JJ, Hines HM, Ruiz M, Salazar C, Linares M, Moreira GRP, Jiggins CD,
Counterman BA, McMillan WO, Papa R (2017) Complex modular architecture around a
simple toolkit of wing pattern genes. *Nat. Ecol. Evol.*, 1, 0052.

Weir BS, Clark Cockerham C (1984) Estimating F-Statistics for the Analysis of Population
Structure. *Evolution*, 38, 1358.

Wong EH, Khrunin A, Nichols L, Pushkarev D, Khokhrin D, Verbenko, D, et al. (2017)
Reconstructing genetic history of Siberian and Northeastern European populations. *Genome
Res.* 27(1), 1–714.

Zeller KA, McGarigal K, Whiteley AR (2012) Estimating landscape resistance to movement: a
review. *Landscape Ecology*, 27(6), 777–797.

Data Accessibility

Data Accessibility

Text S1. Pairwise F_{ST} values between Northern species at the GBS SNPs. (A) F_{ST} between Med and Nor; (B) F_{ST}

between Med and Tva; (C) F_{ST} between Nor and Tva; Nor: North Sea *M. edulis*; Med: West-Mediterranean *M.*

galloprovincialis; Tva: Baltic Sea *M. trossulus*.

Text S2. KASPar genotypes of each individual in the Kerguelen Islands (35 sampling sites), plus those of the additional

individuals from other Southern Hemisphere populations (6 sampling sites).

Text S3. Individual ancestries estimated with ADMIXTURE for the GBS samples. (A) K=2; (B) K=3; (C) K=4; (D)

K=5; (E) K=6; (F) K=7; (G) K=8.

Text S4. Individual ancestries estimated with ADMIXTURE for the KASPar samples in the Kerguelen (K=2; defining

reference populations as *M. edulis* and Chilean mussels).

Text S5. Joint site frequency spectrum (in $\partial a \partial i$ format) between the Kerguelen mussels and (A) *M. edulis;* (B) *M.*

galloprovincialis; (C) *M. trossulus*.

Text S6. Definition of the eight models of divergence used in our inferences with $\partial a \partial i$.

Text S7. Neighbour-joining trees of the 395 retained GBS sequences.

Dryad doi: 10.5061/dryad.6k740. DNA sequences and VCF files including GBS genotypes of each individual in the

ten GBS-typed populations (nine Northern populations and eight Kerguelen mussels).

Supplementary Information

823 **Supplementary Information**

824 Supplementary Information M&M

825 Supplementary Information Tables (S1 – S12)

826 Supplementary Information Figures (S1 – S8)

827 Supplementary Information Texts (S1 – S7)

Author Contributions

828 **Author Contributions**

829 *Data acquisition*: A. Haguenauer, A. Weber and K. Gérard. *Data analysis*: C. Fraïsse, A. Chenuil and N. Bierne.

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Tables

832 **Tables**

Table 1. Counting the different gene topologies for the GBS dataset (10 populations, 67 gene trees)

topology	categories	counts	
		weight >= 0.90	weight >= 0.75
A. *M. edulis* with Kerguelen	A1 ancient Kerguelen divergence	3	5
	A2 introgression	5	12†
	NA	5	22
	total	**13**	**39**
B. *M. galloprovincialis* with Kerguelen	B1 ancient Kerguelen divergence	2	2
	B2 introgression	3	3
	NA	0	4
	total	**5**	**9**
C. *M. trossulus* with Kerguelen	C1 ancient Kerguelen divergence	5	11
	C2 introgression	3	4
	NA	1	4
	total	**9**	**19**
TOTAL		**27**	**67**

833 **topology**: three topologies have been weighted with *Twisst* (among the 395 gene trees, only 67 were topologically resolved); **categories**: "*ancient divergence*": the Kerguelen
834 population branched as a sister-clade to a Northern species; "*introgression*": Kerguelen mussels were distributed within a Northern species; "*na*": topologies that we were unable to
835 classify in these two categories. **weight**: exact weightings of each topology to the full tree.

Table 2. RDA analysis for the KASPar dataset (695 individuals, 33 SNPs)

	RDA (N=695)					conditioned RDA (N=695)			
	variance	%variance	d.f.	p-value		variance	%variance	d.f.	p-value
A. Global effect					**A. Environmental effect**				
Model	0.5288	2.324	7	0.001 ***	Substrate+*Macrocystis*+Slope	0.1213	1.0929	6	0.041 *
Residual	22.2183	97.676	687	-	Residual	10.9621	98.7588	687	-
B. Marginal effect					**B. Individual effect**				
Longitude	0.0128	0.0563	1	0.664	Longitude	0.1424	0.626	1	0.001 ***
Substrate	0.0594	0.261	3	0.133	Substrate	0.2322	1.021	3	0.001 ***
Macrocystis	0.0275	0.121	1	0.032 *	*Macrocystis*	0.0691	0.304	1	0.011 *
Slope	0.0379	0.167	2	0.217	Slope	0.0912	0.401	2	0.072 .
Residual	10.9621	48.191	687	-	Residual	22.2183	97.676	687	-

836 RDA significance tests are shown on the left for: (**A**) the global model with non-significant terms removed; and (**B**) the marginal effect of each constraining variable, which was
837 tested through permutation tests by removing each term one by one from the model containing all other terms. Conditioned RDA significance tests are shown on the right for: (**A**) the
838 combined effect of environmental variables after conditioning on Longitude; and (**B**) for each term after conditioning on other constraining variables to remove their confounding
839 effects. **N**: number of individual genotypes; **variance**: genotypic variance explained by each factor; **%variance**: percent of genotypic variance; **d.f.**: degrees of freedom; **p-value**: p-
840 values obtained through 1000 permutations (* ≤ 0.05; ** ≤ 0.01; *** ≤ 0.001).

Figures

Figure 1. Localities and genetic network of the Northern and Southern *Mytilus* samples (ten GBS-typed populations). The genetic network was produced with the neighbour-net method based on 51,878 high-quality GBS SNPs. The *Mytilus platensis* sample is 'Baie de la Mouche' (KER, purple) in the Kerguelen Islands (Southern Ocean). *Mytilus trossulus* samples are 'Tvarminne' (EU, light green) in the European population of the Baltic Sea and 'Tadoussac' (AM, dark green) in the American population of the Saint Lawrence River. *Mytilus galloprovincialis* samples are 'Faro' (ATL – peripheral, red) in the Atlantic population of Iberian Coast, 'Guillec' (ATL – embedded, orange) in the Atlantic population of Brittany, 'Sete' (MED – west, yellow) in the Occidental Mediterranean basin and 'Crete' (MED – east, black) in the Oriental Mediterranean basin. *Mytilus edulis* samples are 'Wadden Sea' (EU – peripheral, light blue) in the European population of the North Sea, 'Lupin/Fouras' (EU – embedded, cyan) in the European population of the Bay of Biscay and 'Quonochontaug/Old Saybrook Town' (AM, slate blue) in the American population of Rhode Island. Estimate of divergence times between Northern species are indicated with braces (Rawson & Hilbish 1995 , Roux *et al.* 2014), and contact zones between them are depicted by grey stripes.

Figure 2. Evidence of admixture in the ten GBS-typed *Mytilus* spp. populations.
A. Maximum-likelihood population tree inferred by *TreeMix* showing 7 migration
events. ML estimation was made by using 32,162 GBS SNPs grouped in windows
of 100 SNPs. Terminal nodes are labelled by locality abbreviation (Figure 1 and
Table S1) and colors match Figure 1. The drift parameter is shown on the x-axis.
Horizontal branch lengths are proportional to the amount of genetic drift that has
occurred in each branch. Admixture arrows are coloured according to the migration
weight, which represents the fraction of ancestry derived from the migration event.
Migration events with bootstrap support less than 50% are shown with dotted lines,
and those with bootstrap support less than 20% are not shown (Table S8). **B.**
Pairwise privately shared SNPs between a subset of populations of similar sample
size (Table S1). The width of the lines is proportional to the number of SNPs shared
(*M. trossulus:* KER-AM=1,394 (America); *M. galloprovincialis*: KER-ATL=254
(Atlantic - peripheral) and KER-MED=392 (Occidental Mediterranean basin); *M.*
edulis: KER-EU=421 (Europe - peripheral) and KER-AM=532 (America)). The
size of the circles is proportional to the number of non-shared private SNPs
(KER=6,022; *M. trossulus*: AM=13,095; *M. galloprovincialis*: ATL=1,948,
MED=3,927; *M. edulis*: EU=3,251, AM=2,731). Singletons were removed.

Figure 3. Summary of the different gene topologies obtained with *Twisst* for the GBS dataset. Three topologies have been weighed with *Twisst* for each of the 395 contigs (of which only 67 were topologically resolved), and classified in different categories depending whether the Kerguelen mussels branched as a sister-clade to a Northern species ("ancient divergence"), or were distributed within a Northern species ("introgression"). **A.** Kerguelen clustered with *Mytilus trossulus*: A1 "ancient Kerguelen divergence" and A2: "introgression"; **B.** Kerguelen clustered with *Mytilus edulis*: B1 "ancient Kerguelen divergence" and B2: "introgression"; **C.** Kerguelen clustered with *Mytilus galloprovincialis*: C1 "ancient Kerguelen divergence" and C2: "introgression". For each category, a typical neighbour-joining tree computed on the longest non-recombining block of the contig is shown (defined with the Difference of Sums of Squares method of McGuire & Wright 2000). In panel A2, the *Mytilus edulis* haplotypes have totally replaced their Southern counterparts in the Kerguelen. Panel B2 suggests a more ancient introgression of *Mytilus galloprovincialis* haplotypes given that all haplotypes sampled in the Kerguelen form a distinct cluster within the *Mytilus galloprovincialis* clade. And panel C2 represents a complete introgression of *Mytilus trossulus* haplotypes into the Kerguelen. Contig IDs correspond to Table S6, and sizes of non-recombining blocks are given in brackets. Colors match Figure 1.

Figure 4. Geographic variation of the average foreign allele frequency across the four most differentiated loci in the Kerguelen Islands (filled symbols, X10, X11, X57 and Glu-5' from Gérard *et al. (*2015)) and the other loci (open symbols). Alleles were labelled based on their frequencies in the *M. galloprovincialis* Atlantic population of Iberian Coast (Table S4). Red points indicate sites located in the Gulf of Morbihan. **A.** Map of the Kerguelen Islands (150 km East to West; 120 km North to South) together with a world map indicating its location in the Southern Ocean (surrounded in red), and an enlarged map of the Henri Bossière Fjord. Sites PAF (the main harbour of the island) and RdA are separated by 40 kms of coasts, RdA and Pmo by 8 kms, and they show a genetic break on Panel B. Sampling details are provided in Table S3. **B.** Frequency of the average foreign allele across sampling sites in the Kerguelen Islands (ordered by geography), in the three reference Northern species and in the Chilean mussels. See Figure S4 for the detailed pattern at each locus. On top, the average *Mytilus edulis* vs. Chilean ancestries from an ADMIXTURE analysis (K=2) are shown for each site. **C.** Simulation results after 300 generations of a secondary contact between *Mytilus edulis* and Chilean *Mytilus platensis* mussels that meet twice on a circular stepping-stone model (between demes n°18 / n°19 and demes n°19 / n°20). A physical barrier to dispersal was modelled between the *Mytilus edulis* deme (n°19) and the Chilean deme n°18 (dashed vertical bar, migration rate=0.05 instead of 0.5 as everywhere else). Deme size was fixed to 500 individuals, and the species barrier was asymmetric (at barrier loci, the selection coefficient against *Mytilus edulis* in the *Mytilus platensis* background was set to 0.2 while it was set to 0.5 against the *Mytilus platensis* allele in the *Mytilus edulis* background).

Figure 5. Correlation between the average foreign allele frequency of the four most differentiated loci (**A**) and
other loci (**B**) at each sampling site in the Kerguelen Islands (y-axis, Table S4), and the presence/absence of
Macrocystis (x-axis, this is the average sampling site coordinates on the first axis of the *Macrocystis* RDA, see
Table S12). Pearson correlation coefficient: on the left panel, 0.494 (p-value=0.003); on the right panel, 0.107 (p-
value=0.541). Red points indicate sites located in the Gulf of Morbihan (see Figure 4A).

Figure 6. Correlation between the level of differentiation among the Kerguelen Islands (x-axis) and the Northern
species (F_{ST}, y-axis) at each KASPar SNP. Panel (**A**) shows the genetic differentiation between Kerguelen
populations (global F_{ST}); and panel (**B**) shows the genetic-by-environment association (locus coordinates on the
first axis of the conditioned RDA (absolute values), see Table S11). Northern species are *Mytilus edulis* (EU –
peripheral, European population of the North Sea) and *Mytilus galloprovincialis* (MED - west, Mediterranean
population of the West basin). Ancestry-informative loci, i.e., F_{ST} *M.edulis_M.galloprovincialis* > 0.120
(horizontal dashed line, see Table S2), are depicted in orange. Wilcoxon's test between the ancestry-informative
loci (orange) and the background loci (grey): (**A**) mean_orange=0.015, mean_grey=0.007 (p-value=0.021); (**B**)
mean_orange=0.112, mean_grey=0.048 (p-value=0.006). Their respective means are depicted by vertical dashed
lines. Pearson correlation coefficient: (**A**) r=0.540 (p-value=0.001); (**B**) r=0.439 (p-value=0.011).