Brain, immune system and selenium: a starting point for a new diagnostic marker for

Alzheimer's disease?

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**Highlights** 

MsrB1 is one of the 25 selenoenzymes expressed in the humans

MsrB1 is highly expressed in human circulating neutrophils

The diagnostic markers for Alzheimer's disease are still insufficiently validated

The impairment of some selenoenzymes is associated with Alzheimer's disease

Neutrophil MsrB1 can be a peripheral marker for the diagnosis of Alzheimer's disease

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**Abstract** 

The clinical diagnosis of Alzheimer's disease (AD) is based primarily on neuropsychological tests, which assess the involutive damage, and imaging techniques that evaluate morphologic changes in the brain. The currently available diagnostic tests do not show complete specificity and do not permit an accurate differentiation between AD and other forms of senile dementia. The correlation of these tests with laboratory investigations based on biochemical parameters could increase the certainty of the diagnosis. In recent years, several biochemical markers for the diagnosis of AD have been proposed, but in most cases they show a limited specificity and their application is invasive, because it generally requires the sampling of cerebrospinal fluid. Therefore, the use of a peripheral biochemical marker could represent a valuable complement for the diagnosis of this disease.

Several studies have shown a relationship between the neurodegenerative disorders typical of the ageing process, the weakening of the immune system, alterations in the levels of selenium and of the antioxidant selenoenzymes in brain tissues and blood cells, particularly in neutrophil granulocytes. The levels of peripheral selenoenzymes may reveal a promising clinical parameter for helping in the assessment of the pathological condition in AD.

Background

Selenium is an essential element involved in several cellular processes such as immune response, signal transduction, metabolism of thyroid hormones and defence against oxidative damage [1]. Selenium is contained in proteins as selenocysteine, which can be considered as the 21<sup>st</sup> proteinogenic amino acid. When selenocysteine is present in the active site of the protein, it confers to the enzyme a higher catalytic efficiency compared to cysteine. The most recent studies have revealed that 25 selenoproteins are expressed in humans [2, 3]. Many of them, such as glutathione peroxidase (GPx) and methionine sulfoxide reductase B1 (MsrB1), are oxidoreductase enzymes that contain selenocysteine in the catalytic site and are involved in the regulation of cellular redox processes [4].

Members of the GPx family perform the detoxification of reactive oxygen species (ROS) produced during the aerobic metabolism, through the reduction of organic peroxides using glutathione as the electron donor, and are characterised by different substrate specificity, cellular localization and tissue distribution [5].

During ageing, and in association with neurodegenerative diseases such as Alzheimer's disease (AD), a significant decrease of GPx activity in whole blood [6], erythrocytes [7-12], neutrophils [7, 13] and plasma [7, 14, 15] was observed. Quite remarkably neutrophils from patients affected by neurodegenerative diseases displayed less than 30% of the GPx activity of neutrophils from healthy individuals [7]. This phenomenon was correlated with the decrease in selenium levels that was observed to occur in plasma [7, 16, 17, 9, 18] and in erythrocytes [7, 18] during ageing and in individuals affected by various neurodegenerative disorders. In a sample of geriatric population, a 25% decrease in selenium plasma levels was observed, along with a 20% decrease in erythrocyte GPx activity [19]. The changes in selenium status that occur during ageing, which can be attributed to defects of assimilation and/or metabolism of this element, may lead to an increased susceptibility to various degenerative diseases typical of the elderly, including AD, which is characterised by

oxidative damage to various cellular components [20]. The increased oxidative stress associated with selenium deficiency has been correlated with defects in the regulation of GPx expression [21]. Low selenium levels increase significantly the risk of mortality during senility [22]. This element is therefore considered as an important factor for maintaining a good state of health in the elderly [23]. Components of both the innate and acquired immune systems deteriorate gradually during ageing [24, 25], and this functional impairment further aggravates in the course of AD [26, 27]. The cellular components that guarantee the efficiency of the innate immune response are the phagocytes: macrophages and neutrophil granulocytes, whose task is to engulf the pathogen into a phagosome and degrade it by means of proteolytic enzymes and various ROS, like superoxide, hydrogen peroxide and hypochlorite, produced during the so-called oxidative burst. As ROS can diffuse from the phagosome into the cytosol during the burst, and thus compromise cell function, neutrophils are endowed with powerful antioxidant systems, both enzymatic and non-enzymatic, that act to prevent or repair the oxidative damage [28]. Selenium is essential for the full efficiency of various components of the immune system, both acquired and innate [29, 30]. Several studies conducted in mice and humans have shown that selenium deficiency, as indicated by the loss of GPx activity, leads to a reduction in the phagocytic capacity and in the intensity of the oxidative burst in neutrophils, resulting in decreased bactericidal activity [31, 32], whereas selenium supplementation in the diet restores GPx activity and cell function [32]. Studies in vitro with HL-60 cells (a human promyelocytic leukaemia cell line that can be induced to differentiate into a neutrophilic phenotype) grown in a selenium-deficient medium indicate the complete absence of GPx, whose expression is restored to values comparable to those measured in neutrophils when the medium is supplemented with appropriate amounts of selenium [33, 34]. Another study indicate that the supplementation of selenium in the diet of humans with selenium deficiency associated with neurodegenerative diseases increases both selenium levels and GPx

activity in plasma and erythrocytes [7]. These data indicate that GPx expression is very sensitive to the levels of selenium in the organism.

The decrease in neutrophil's bactericidal capacity appears to be linked to ROS accumulation and to correlate with the loss of GPx activity, resulting in a redox imbalance in the cells, which become therefore more sensitive to oxidative damages. Neutrophils from AD patients were found to contain high concentrations of intracellular ROS under restingesting conditions, pointing to a defect in antioxidant systems of these cells [35].

Human neutrophils are the cell type with the highest selenium concentration, although it has been shown that the levels of GPx activity are lower than those measured in other cell types, such as in kidney and liver. Moreover, neutrophils of mice subjected to a diet supplemented with selenite display, at low selenium concentrations, an increase in bactericidal activity but not in GPx activity, while only at higher concentrations of selenium the two activities increase in parallel [32]. These data suggest that the expression of GPx is particularly dependent on the selenium status of the organism, and, most importantly, that this protein is not the only selenoenzyme that ensures proper neutrophil function.

Only one other selenoenzyme with anti-oxidant function has been characterised in detail in human neutrophils [36]. It is MsrB1, belonging to the family of methionine sulfoxide reductases (MsrA and MsrB), and the only form of Msr containing a catalytically active selenocysteine [37]. The Msrs catalyse the stereospecific reduction of the two diastereoisomers of methionine sulfoxide, which are generated by the oxidation of the sulphide group of methionine, as the free amino acid or when it is present in proteins: [MsrAs reduce (S<sub>C</sub>)methionine-(S<sub>S</sub>)sulfoxide, while MsrBs reduce (S<sub>C</sub>)methionine-(R<sub>S</sub>)sulfoxide] [38]. Msrs perform three main tasks: (i) the repair of the oxidative damage suffered by methionine residues, which results in the alteration of function of many proteins, such as the beta-amyloid peptide, in which the oxidation of methionine appears critical for aggregation, neurotoxicity, and the generation of ROS induced by the peptide [39]; (ii) the

detoxification of ROS through the cyclic oxidation/reduction of methionine, free or inserted in some

proteins, thus preventing the oxidation of other particularly sensitive cellular components [40]; (iii)

the regulation of cellular metabolism through changes in the redox state of specific methionine

residues present in various enzymes, hormones and plasma proteins [41]. The imbalance in the

methionine/methionine sulfoxide ratio can alter the metabolic functions, contributing to the

development of diseases related to the impairment of the cellular processes affected [42].

The selenoenzyme MsrB1 is the form predominantly expressed in circulating human neutrophils,

but is almost entirely absent in other blood cells [36]. MsrB1, like GPx, would be a component of

the selenium-dependent antioxidant machinery that has evolved in neutrophils to counteract the

ROS produced in high concentrations during the immune response.

Hypothesis

We propose this hypothesis: the selenoenzyme MsrB1 from neutrophil granulocytes could represent

a more specific indicator of the selenium status in the organism and in particular a new peripheral

diagnostic marker for AD, alternative to, or in combination with those proposed, which are typically

invasive since they involve the analysis of cerebrospinal fluid, and display low specificity [43, 44].

**Results and Discussion** 

Concerning the hypothesis presented here, MsrB1 activity was analysed in neutrophil samples from

healthy donors and patients affected by AD associated with diversified diseases typical of the

elderly. A limited cohort of patients was studied, not allowing statistical elaboration, therefore the

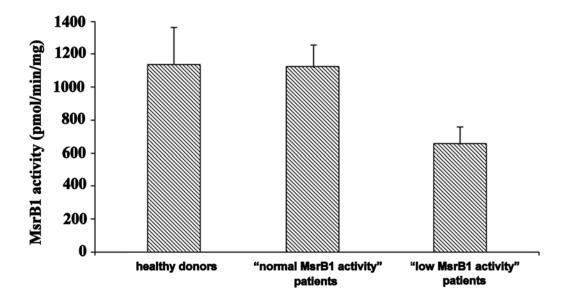
present study should be considered as a preliminary investigation. Yet, this pilot study showed that

the patients could be grouped in two populations: one group with MsrB1 specific activity similar to

that of healthy donors, and one group with approximately the 50% of the specific activity of healthy

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donors (Figure 1).



**Figure 1.** Specific activity of MsrB1 (pmol of dabsyl-methionine produced per min per mg total protein) detected in neutrophils from: healthy donors (n=8), "normal MsrB1 activity" patients (n=8), and "low MsrB1 activity" patients (n=5).

The novel finding of decreased MsrB1 activity with ageing presented here is consistent with literature data about the decrease in antioxidant defence, and particularly of selenoenzymes, associated with age-related neurodegenerative pathologies typical of the elderly. Moreover, particularly remarkable is a post-mortem observation that showed a significant decrease in Msr activity (MsrA plus MsrB) in various areas of the brain in subjects affected by AD [45]. The alterations observed in the immune system of patients suffering from neurodegenerative disorders, particularly AD, suggest a correlation between the two events, supporting the hypothesis of a systemic nature of this disease [26, 46]. The observation of a concomitant impairment of Msr activity in brain tissues and peripheral neutrophils, in association with AD, seem to corroborate this hypothesis.

Moreover, the increase in the intracellular content of oxidized proteins, especially at the level of

methionine residues, which occurs during ageing, has been correlated with the onset of degenerative

diseases typical of advanced age [42, 47, 48]. The limited capacity of reduction of methionine

sulfoxide by Msr enzymes, including the selenoenzyme MsrB1, renders the defence systems against

oxidative stress inadequate, thus favouring the onset of degenerative diseases, like the Alzheimer's,

which are characterised by accumulation of oxidized proteins [42, 47].

In light of the preliminary evidence presented here, the idea of a correlation between AD, partial

impairment of the immune system and decreased MsrB1 activity should be further investigated. It is

also to be clarified whether the decline MsrB1 activity observed in some AD patients was due to a

decrease in protein expression or to an impairment of the catalytic activity. Although MsrB1 was

discovered in 1999 [38], it is still poorly studied and it has not yet been considered as a possible

indicator of the selenium status of the organism [49]. Therefore, we deem it of particular interest to

explore the potential use of MsrB1 as a selective and reliable diagnostic, peripheral marker for AD,

a very invaliding neurodegenerative condition that afflicts millions of people in the world, and for

which none of the imaging or neurochemical diagnostic markers proposed to date can be considered

to be sufficiently validated [50].

Methods

Purification of human neutrophils

Fresh human blood was from the Institute of Geriatric Rehabilitation "Santa Margherita" (Pavia,

Italy) and from the Immunohematology and Transfusion Medicine Department of the "San Matteo"

Hospital (Pavia, Italy) as approved by the respective internal Institution Review Boards.

Neutrophils were obtained by Dextran-70 sedimentation of blood followed by Ficoll-Hypaque

gradient centrifugation, essentially as described [51] with minor modifications [52], washed in PBS

supplemented with 2 mM EDTA and 0.5% (w/v) BSA, and centrifuged at 500g for 10 min at 4 °C.

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The residual erythrocytes in the neutrophils-rich fraction were eliminated by differential hypotonic

lysis with ice cold 0.2% (w/v) NaCl for 30 s under agitation. The isotonicity was restored by adding

an equal volume of 1.6% (w/v) NaCl. The cells were sedimented under the same conditions, and

then washed three times in supplemented PBS. The number of neutrophils was determined by

microscope-count, and the samples were then stored frozen at -80 °C.

Determination of methionine sulfoxide reductase activity

Purified neutrophils were resuspended (10<sup>8</sup> cells/ml) in PBS containing 1% (v/v) Triton X-100 and

0.1% (v/v) diisopropylfluorophosphate as a serine-protease inhibitor, and left on ice for 30 min. The

lysate was then centrifuged at 18000 g for 10 min at 4°C. The supernatant was collected and

analyzed directly. Protein concentration was assayed with the bicinchoninic acid method using BSA

as a standard.

MsrB enzymatic activity was assayed using the R diastereoisomer of methionine sulfoxide

conjugated with the dabsyl group as the substrate, and 1,4-dithioerythritol as the electron donor.

Methionine-R-sulfoxide was separated from its optical antipode as described [53, 54] and

functionalized with the dabsyl group as described [55]. The reaction mixture contained 5 mM

sodium phosphate (pH 7.4), 154.5 mM NaCl, 4.5 mM KCl, 20 mM 1,4-dithioerythritol, 500 μM

dabsyl-methionine-R-sulfoxide and 50 µg of total protein from the cell extract. The reaction was

carried out at 37 °C for 30 min, and the reaction product, dabsyl-methionine, was analyzed by

reverse-phase HPLC, monitoring the chromophoric dabsyl moiety at the wavelength of 436 nm

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[55].

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## References

- 1) Rayman MP. The importance of selenium to human health. Lancet 2000;356:233-241.
- 2) Kryukov GV, Castellano S, Novoselov SV, Lobanov AV, Zehtab O, Guigò R, Gladyshev VN. Characterization of Mammalian Selenoproteomes. Science 2003;300:1439-1443.
- 3) Papp LV, Lu J, Holmgren A, Khanna KK. From selenium to selenoproteins: synthesis, identity, and their role in human health. Antioxid Redox Signal 2007;9:775-806.
- 4) Hawkes WC, Alkan Z. Regulation of redox signaling by selenoproteins. Biol Trace Elem Res 2010;134:235-251.
- 5) Schweizer U, Bräuer AU, Köhrle J, Nitsch R, Savaskan NE. Selenium and brain function: a poorly recognized liaison. Brain Res Brain Res Rev 2004;45:164-178.
- 6) Espinoza SE, Guo H, Fedarko N, DeZern A, Fried LP, Xue QL, Leng S, Beamer B, Walston JD. Glutathione peroxidase enzyme activity in aging. J Gerontol A Biol Sci Med Sci 2008;63:505-509.
- 7) Clausen J, Jensen GE, Nielsen SA. Selenium in chronic neurologic diseases. Multiple sclerosis and Batten's disease. Biol Trace Elem Res 1988;15:179-203.
- 8) Kharrazi H, Vaisi-Raygani A, Rahimi Z, Tavilani H, Aminian M, Pourmotabbed T. Association between enzymatic and non-enzymatic antioxidant defense mechanism with apolipoprotein E genotypes in Alzheimer disease. Clin Biochem 2008;41:932-936.
- 9) Demirin H, Kara Y, Eren I, Delibas N. Alterations of plasma magnesium, copper, zinc, iron and selenium concentrations and some related erythrocyte antioxidant enzyme activities in patients with Alzheimer's disease. J Trace Elem Med Biol 2010;24:169-173.
- 10) Jeandel C, Nicolas MB, Dubois F, Nabet-Belleville F, Penin F, Cuny G. Lipid peroxidation and free radical scavengers in Alzheimer's disease. Gerontology 1989;35:275-282.
- 11) Johannsen P, Velander G, Mai J, Thorling EB, Dupont E. Glutathione peroxidase in early and advanced Parkinson's disease. J Neurol Neurosurg Psychiatry 1991;54:679-682.
- 12) Syburra C, Passi S. Oxidative stress in patients with multiple sclerosis. Ukr Biokhim Zh 1999;71:112-115.
- 13) Ito Y, Kajkenova O, Feuers RJ, Udupa KB, Desai VG, Epstein J, Hart RW, Lipschitz DA. Impaired glutathione peroxidase activity accounts for the age-related accumulation of hydrogen peroxide in activated human neutrophils. J Gerontol A Biol Sci Med Sci 1998;53:169-175.
- 14) Rinaldi P, Polidori MC, Metastasio A, Mariani E, Mattioli P, Cherubini A, Catani M, Cecchetti R, Senin U, Mecocci P. Plasma antioxidants are similarly depleted in mild cognitive impairment and in Alzheimer's disease. Neurobiol Aging 2003;24:915-919.
- 15) Padurariu M, Ciobica A, Hritcu L, Stoica B, Bild W, Stefanescu C. Changes of some oxidative stress markers in the serum of patients with mild cognitive impairment and Alzheimer's disease. Neurosci Lett 2010;469:6-10.
- 16) Smorgon C, Mari E, Atti AR, Dalla Nora E, Zamboni PF, Calzoni F, Passaro A, Fellin R. Trace elements and cognitive impairment: an elderly cohort study. Arch Gerontol Geriatr 2004;Suppl. 9:393-402.
- 17) Akbaraly TN, Hininger-Favier I, Carrière I, Arnaud J, Gourlet V, Roussel AM, Berr C. Plasma selenium over time and cognitive decline in the elderly. Epidemiology 2007;18:52-58.

- 18) Cardoso BR, Ong TP, Jacob-Filho W, Jaluul O, Freitas MI, Cozzolino SM. Nutritional status of selenium in Alzheimer's disease patients. Br J Nutr 2010;103:803-806.
- 19) Olivieri O, Stanzial AM, Girelli D, Trevisan MT, Guarini P, Terzi M, Caffi S, Fontana F, Casaril M, Ferrari S, Corrocher R. Selenium status, fatty acids, vitamins A and E, and aging: the Nove Study. Am J Clin Nutr 1994;60:510-517.
- 20) Stadtman ER, Berlett BS. Reactive oxygen-mediated protein oxidation in aging and disease. Chem Res Toxicol 1997;10:485-494.
- 21) Lei XG, Cheng WH, McClung JP. Metabolic regulation and function of glutathione peroxidase-1. Annu Rev Nutr 2007;27:41-61.
- 22) Walston J, Xue Q, Semba RD, Ferrucci L, Cappola AR, Ricks M, Guralnik J, Fried LP. Serum antioxidants, inflammation, and total mortality in older women. Am J Epidemiol 2006;163:18-26.
- 23) González S, Huerta JM, Fernández S, Patterson AM, Lasheras C. Life-quality indicators in elderly people are influenced by selenium status. Aging Clin Exp Res 2007;19:10-15.
- 24) Gomez CR, Nomellini V, Faunce DE, Kovacs EJ. Innate immunity and aging. Exp Gerontol 2008;43:718-728.
- 25) Fulop T, Larbi A, Douziech N, Fortin C, Guérard KP, Lesur O, Khalil A, Dupuis G. Signal transduction and functional changes in neutrophils with aging. Aging Cell 2004;3:217-226.
- 26) Shalit F, Sredni B, Brodie C, Kott E, Huberman M. T lymphocyte subpopulations and activation markers correlate with severity of Alzheimer's disease. Clin Immunol Immunopathol 1995;75:246-250.
- 27) Song C, Vandewoude M, Stevens W, De Clerck L, Van der Planken M, Whelan A, Anisman H, Dossche A, Maes M. Alterations in immune functions during normal aging and Alzheimer's disease. Psychiatry Res 1999;85:71-80.
- 28) Splettstoesser WD, Schuff-Werner P. Oxidative stress in phagocytes "the enemy within". Microsc Res Tech 2002;57:441-455.
- 29) Hoffmann PR. Mechanisms by which selenium influences immune responses. Arch Immunol Ther Exp (Warsz) 2007;55:289-297.
- 30) Hoffmann PR, Berry MJ. The influence of selenium on immune responses. Mol Nutr Food Res 2008;52:1273-1280.
- 31) Serfass RE, Ganther HE. Defective microbicidal activity in glutathione peroxidase-deficient neutrophils of selenium-deficient rats. Nature 1975;255:640-641.
- 32) Arthur JR, McKenzie RC, Beckett GJ. Selenium in the immune system. J Nutr 2003;133:1457-1459.
- 33) Takahashi K, Newburger PE, Cohen HJ. Glutathione peroxidase protein. Absence in selenium deficiency states and correlation with enzymatic activity. J Clin Invest 1986;77:1402-1404.
- 34) Chada S, Whitney C, Newburger PE. Post-transcriptional regulation of glutathione peroxidase gene expression by selenium in the HL-60 human myeloid cell line. Blood 1989;74:2535-2341.
- 35) Vitte J, Michel BF, Bongrand P, Gastaut JL. Oxidative stress level in circulating neutrophils is linked to neurodegenerative diseases. J Clin Immunol 2004;24:683-692.
- 36) Achilli C, Ciana A, Rossi A, Balduini C, Minetti G. Neutrophil granulocytes uniquely express, among human blood cells, high levels of Methionine-sulfoxide-reductase enzymes. J Leukoc Biol 2008;83:181-189.

- 37) Kim HY, Gladyshev VN. Methionine sulfoxide reductases: selenoprotein forms and roles in antioxidant protein repair in mammals. Biochem J 2007;407:321-329.
- 38) Achilli C, Ciana A, Minetti G. The discovery of methionine sulfoxide reductase enzymes: An historical account and future perspectives. Biofactors 2015;41:135-152.
- 39) Varadarajan S, Kanski J, Aksenova M, Lauderback C, Butterfield DA. Different mechanisms of oxidative stress and neurotoxicity for Alzheimer's A beta(1-42) and A beta(25-35). J Am Chem Soc 2001;123:5625-5631.
- 40) Weissbach H, Resnick L, Brot N. Methionine sulfoxide reductases: history and cellular role in protecting against oxidative damage. Biochim Biophys Acta 2005;1703:203-212.
- 41) Hansel A, Heinemann SH, Hoshi T. Heterogeneity and function of mammalian MSRs: enzymes for repair, protection and regulation. Biochim Biophys Acta 2005;1703:239-247.
- 42) Stadtman ER, Van Remmen H, Richardson A, Wehr NB, Levine RL. Methionine oxidation and aging. Biochim Biophys Acta 2005;1703:135-140.
- 43) Bailey P. Biological markers in Alzheimer's disease. Can J Neurol Sci 2007;34:72-76.
- 44) Galasko D, Montine TJ. Biomarkers of oxidative damage and inflammation in Alzheimer's disease. Biomark Med 2010;4:27-36.
- 45) Gabbita SP, Aksenov MY, Lovell MA, Markesbery WR. Decrease in peptide methionine sulfoxide reductase in Alzheimer's disease brain. J Neurochem 1999;73:1660-1666.
- 46) Fortin CF, McDonald PP, Lesur O, Fülöp T Jr. Aging and neutrophils: there is still much to do. Rejuvenation Res 2008;11:873-882.
- 47) Stadtman ER. Protein oxidation and aging. Free Radic Res 2006;40:1250-1258.
- 48) Radak Z, Zhao Z, Goto S, Koltai E. Age-associated neurodegeneration and oxidative damage to lipids, proteins and DNA. Mol Aspects Med 2011;32:305-315.
- 49) Combs GF. Biomarkers of Selenium Status. Nutrients 2015;7:2209-2236.
- 50) Hampel H, Broich K, Hoessler Y, Pantel J. Biological markers for early detection and pharmacological treatment of Alzheimer's disease. Dialogues Clin Neurosci 2009;11:141–157.
- 51) Böyum A. Isolation of mononuclear cells and granulocytes from human blood. Isolation of monuclear cells by one centrifugation, and of granulocytes by combining centrifugation and sedimentation at 1 g. Scand J Clin Lab Invest (Suppl) 1968;97: 77-89.
- 52) Melloni E, Pontremoli S, Michetti M, Sacco O, Sparatore B, Salamino F, Horecker BL. Binding of protein kinase C to neutrophil membranes in the presence of Ca2+ and its activation by a Ca2+-requiring proteinase. Proc Natl Acad Sci U S A 1985;82:6435-6439.
- 53) Lavine TF. The formation, resolution, and optical properties of the diastereoisomeric sulfoxides derived from L-methionine. J Biol Chem 1947;169:477-491.
- 54) Greenstein JP, Winitz M. Chemistry of the Amino Acids, Krieger Publishing Company, Malabar, vol. 3, 2125-2155 (1984).
- 55) Minetti G, Balduini C, Brovelli A. Reduction of DABS-L-methionine-dl-sulfoxide by protein methionine sulfoxide reductase from polymorphonuclear leukocytes: stereospecificity towards the l-sulfoxide. Ital J Biochem 1994;43:273-283.