

Differential 3' Processing of Specific Transcripts Expands Regulatory and Protein Diversity Across Neuronal Cell Types

Saša Jereb¹, Hun-Way Hwang^{1,5}, Eric Van Otterloo², Eve-Ellen Govek³, John J. Fak¹, Yuan Yuan¹, Mary E. Hatten³, Robert B. Darnell^{1,4}

¹Laboratory of Molecular Neuro-Oncology and Howard Hughes Medical Institute, The Rockefeller University, 1230 York Avenue, New York, New York 10065, USA.

²Department of Craniofacial Biology, University of Colorado Anschutz Medical Campus, Aurora, CO 80045, USA. ³Laboratory of Developmental Neurobiology, The Rockefeller

University, New York, NY 10065, USA. ⁴Corresponding author. ⁵Current address:

Department of Pathology, University of Pittsburgh, School of Medicine, Pittsburgh, PA 15213, USA.

CONTACT INFORMATION:

Robert B. Darnell

Box 226, 1230 York Avenue, New York, New York 10065, USA

Phone: (212) 327-7460

Fax: (212) 327-7109

Email: darnellr@rockefeller.edu (R.B.D.)

ABSTRACT

Alternative polyadenylation (APA) regulates mRNA translation, stability, and protein localization. However, it is unclear to what extent APA regulates these processes uniquely in specific cell types. Using a new technique, cTag-PAPERCLIP, we discovered significant differences in APA between the principal types of mouse cerebellar neurons, the Purkinje and granule cells, as well as between proliferating and differentiated granule cells. Transcripts that differed in APA in these comparisons were enriched in key neuronal functions and many differed in coding sequence in addition to 3'UTR length. We characterize *Memo1*, a transcript that shifted from expressing a short 3'UTR isoform to a longer one during granule cell differentiation. We show that *Memo1* regulates granule cell precursor proliferation and that its long 3'UTR isoform is targeted by miR-124, contributing to its downregulation during development. Our findings provide insight into roles for APA in specific cell types and establish a platform for further functional studies.

INTRODUCTION

Alternative polyadenylation (APA) is a process by which different ends to an mRNA transcript are determined. These alternative mRNA isoforms differ in the length of their regulatory 3' untranslated region (3'UTR) and in some cases their coding sequence (CDS). While we have a good understanding of the scope of molecular functions determined by alternative 3'UTR regions – APA has been implicated in the regulation of subcellular protein and mRNA localization, translational regulation and mRNA stability (Berkovits and Mayr, 2015; Tian and Manley, 2016) – we are only beginning to understand the effects of APA on various physiological processes *in vivo*. Genome-wide changes in APA have been observed during neuronal activation (Flavell et al., 2008), T-cell activation (Sandberg et al., 2008), oncogenesis (Mayr and Bartel, 2009) and development (Ji et al., 2009; Shepard et al., 2011; Wang et al., 2013), suggesting diverse biologic roles for APA.

Examination of specific genes has provided evidence for important roles for APA in embryonic development and synaptic plasticity. For example, during *Drosophila* development, the long 3'UTR isoform of mRNA encoding Polo kinase is expressed in abdominal epidermis precursor cells and is translated with much higher efficiency than the short 3'UTR isoform expressed in the adult epidermis. Because high levels of Polo protein are required for the proliferation of epidermis precursor cells, deletion of the *Polo* distal polyadenylation signal leads to death during development (Pinto et al., 2011). Another example is *BDNF* mRNA; its two 3'UTR isoforms each have distinct functions in neurons. The long *BDNF* isoform is localized to dendrites and translated upon neuronal activity, whereas the short isoform is localized to the cell body and is constitutively translated. Mice that lack the long 3'UTR of *BDNF* exhibit altered dendritic spine

morphology and decreased plasticity of dendritic synapses (An et al., 2008; Lau et al., 2010).

A comprehensive functional understanding of APA in the brain, however, is lacking. Recently, it has been found that mammalian and fly brains express particularly long 3'UTR isoforms compared to other tissues (Miura et al., 2013), suggesting that APA may play a particularly important role in neurons. Current methods have not been able to discern the extent of APA diversity across different neuronal types, and how that may contribute to their morphologic and physiologic diversity. Recently, new approaches, like translating ribosome affinity purification (TRAP), have been developed that enable sequencing of mRNA from specific neurons in a cell type-specific manner (Mellén et al., 2012; Sanz et al., 2013), but they lack the resolution to precisely identify 3'UTR ends. To address this limitation, we recently developed cTag-PAPERCLIP (conditionally-tagged poly(A) binding protein-mediated mRNA 3' end retrieval by crosslinking immunoprecipitation). cTag-PAPERCLIP – which is based on PAPERCLIP (Hwang et al., 2016) and CLIP (Licatalosi et al., 2008; Ule et al., 2003) – enables purification and sequencing of 3'UTR ends of polyadenylated transcripts via their interaction with poly-A binding protein cytoplasmic 1 (PABPC1), a protein that binds with high specificity to mRNA poly(A) tails. A major strength of the CLIP approach is that by covalently crosslinking RNA to protein via ultraviolet light, this method captures direct RNA-protein interactions *in situ*, allowing stringent immunopurification of physiological interactions from non-specific interactions. cTag-PAPERCLIP was recently used to identify APA switches after inflammatory stimulation of microglia in the brain (Hwang et al., 2017).

Here we studied APA in the cerebellum, a cortical region of vertebrate brain that is primarily involved in motor coordination and sensory-motor processing (Buckner, 2013), because it is composed of well described cell types that are genetically

accessible through Cre-driver lines (Barski et al., 2000; Matei et al., 2005). Using cTag-PAPERCLIP in combination with the appropriate Cre-driver lines, we studied APA in the two principal types of cerebellar neurons: Purkinje and granule cells, which are functionally and morphologically distinct. Purkinje cells, the sole output neuron of the cerebellar cortex, are large, inhibitory neurons with extensive dendritic arbors and a single axon that projects to the cerebellar nuclei. Granule cells, the most numerous neurons in the mammalian brain, are small interneurons that provide the major excitatory input onto Purkinje cells, and receive mossy fiber inputs onto a small number of dendrites (Butts et al., 2014). We also compared APA between granule cell precursors and differentiated granule cells. Granule cell precursors proliferate postnatally in the external granule layer, then exit the cell cycle, extend parallel fiber axons that form synapses with Purkinje cells, and then migrate inward, guided by Bergmann glia, to form the internal granule layer (Edmondson and Hatten, 1987; Solecki et al., 2009).

Using cTag-PAPERCLIP, we found that Purkinje and granule neurons differentially regulate APA on the same transcripts. We also observed such changes in APA during cerebellar granule cell development, when granule cells transition from mitotic precursors to post-mitotic, differentiated neurons wired within the cerebellar circuitry. Transcripts that differed in APA in these comparisons were enriched in key neuronal functions. Differences in APA between the cell types affected 3'UTR length and in some cases CDS. We analyzed one of the transcripts that changes in 3'UTR length during granule cell development, *Memo1* (mediator of ErbB2-driven cell motility 1), which we show to be involved in regulating the proliferation of granule cell precursors in the developing cerebellum. We demonstrate that, as granule neurons differentiate, *Memo1* acquires a longer 3'UTR harboring a miRNA binding site, providing a mechanism for its downregulation during development. Taken together, our findings demonstrate the

potential of single cell-type resolution of APA regulation to reveal quantitative as well as qualitative control of functional diversity in different neuronal populations.

RESULTS

cTag-PAPERCLIP Maps mRNA 3'UTR Ends in Specific Cells *In Vivo*

To investigate the diversity of 3'UTR ends in distinct neuronal types in the mouse brain we compared 3'UTR isoform expression between cerebellar Purkinje and granule cells using cTag-PAPERCLIP. We previously generated a genetically modified mouse, termed *Pabpc1^{cTag}*, that expresses GFP-tagged PABPC1 in a Cre-dependent manner. After Cre recombination, the GFP CDS is inserted into the endogenous PABPC1 gene locus just upstream of the stop codon, leading to the expression of the PABPC1-GFP fusion protein. GFP antibodies can then be used to immunoprecipitate polyadenylated transcripts from cells expressing GFP-tagged PABPC1 (Hwang et al., 2017) (Figure 1A). We bred *Pabpc1^{cTag}* mice to *Pcp2-Cre* (Barski et al., 2000) or *NeuroD1-Cre* mice (H. J. Li et al., 2012), to isolate polyadenylated transcripts from cerebellar Purkinje or granule neurons, respectively.

We first confirmed by immunostaining of cerebellar sections that PABPC1-GFP was expressed specifically in Purkinje or granule cells in these mice (Figure 1B). We then performed cTag-PAPERCLIP (Figure 1C). We dissected cerebella from 8-week old mice, UV-crosslinked RNA to protein within the tissue, immunoprecipitated PABPC1-GFP covalently bound to RNA, and radiolabeled this bound RNA (Figure 1C, fourth lane). To verify that we specifically co-immunoprecipitated RNA bound to PABPC1-GFP, we digested RNA to completion, leaving only the RNA segment bound to PABPC1 remaining, and on visualizing the radiolabeled RNA-protein complex, observed a single band corresponding to the size of PABPC1-GFP (Figure 1C, third lane).

We sequenced co-immunoprecipitated RNA fragments and identified clusters of reads that were present in at least three biological replicates to demarcate reproducible binding sites. By these criteria, we identified 10,830 and 12,099 clusters belonging to 8,575 and 9,411 genes in Purkinje and granule cells, respectively. 27% of genes in Purkinje cells (2,336) and 27% of genes in granule cells (2,451) had multiple robust PABPC1 binding sites, suggesting multiple polyadenylation sites for these genes. The number of reads in each of these clusters was highly correlated across biological replicates (Figure 1D), indicating that this method is quantitatively reproducible.

We assessed the ability of cTag-PAPERCLIP to measure 3'UTR isoform abundance by comparing the sum of cTag-PAPERCLIP reads per gene with the mRNA abundance estimated from TRAP-Seq, an alternative method for cell-type specific mRNA profiling (Mellén et al., 2012). We found a high correlation between the two methods for both Purkinje cells ($R = 0.68$, Figure 1E) and granule cells ($R=0.7$, data not shown).

Known marker genes for each cell type were among the most highly expressed genes in the Purkinje cell (Figure 1E) and granule cell datasets (data not shown), indicating that cTag-PAPERCLIP targeted each cell type specifically. Markers from non-target cell types were ranked significantly lower in cTag-PAPERCLIP data (Figure 1 – figure supplement 1) compared to TRAP-Seq, indicating that cTag-PAPERCLIP is highly selective at purifying RNA from specific cell types.

Two thirds of cTag-PAPERCLIP clusters mapped to the exact end of 3'UTRs of Ensembl-annotated genes (Aken et al., 2016) (Figure 1F). Interestingly, we also discovered thousands of new 3'UTR ends, most of which (30-31% of clusters) mapped within the 3'UTRs of Ensembl-annotated genes, but not to the exact annotated ends. Importantly, the majority (87%) of clusters in Purkinje and granule cells harbored a

canonical poly(A) signal sequence (AAUAAA or AUUAAA) (Tian, 2005) within the cluster.

Taken together, these data demonstrate that cTag-PAPERCLIP specifically purifies 3'UTR ends from genetically defined cell types, and can reveal a host of new transcript ends in the brain.

Cell-type Specific APA Regulates 3'UTR Length and CDS of Transcripts Expressed in Purkinje and Granule Cells

We then compared 3'UTR isoform expression between Purkinje and granule cells in our cTag-PAPERCLIP data. We analyzed genes that were expressed in both cell types in order to compare the expression of different isoforms of the same gene across cell types. This analysis revealed extensive isoform diversity: 629 genes expressed different 3'UTR isoforms between the two cell types (FDR < 0.05) (Table S1).

Most transcripts differed in 3'UTR length only, without impact on the CDS; among differentially polyadenylated transcripts with two poly(A) sites, 319 or 81% differed in 3'UTR length only (3'UTR-APA, Figure 2A). For example, the gene coding for vezatin (*Vezt*), which is involved in neurite outgrowth (Sanda et al., 2010), predominantly expressed a shorter 3'UTR isoform in Purkinje cells compared to granule cells (Figure 2B). Conversely, *Zfp609*, a neuron-specific transcription factor (van den Berg et al., 2017), predominantly expressed a long 3'UTR isoform in Purkinje cells, whereas in granule cells, it expressed both a short and the long 3'UTR isoform. The short 3'UTR isoforms for both *Vezt* and *Zfp609* have not been previously annotated (by Ensembl), and contain a canonical poly(A) signal just upstream of their 3'UTR ends (Figure 2B).

Interestingly, in comparing cTag-PAPERCLIP data between Purkinje and granule cells, we also identified transcripts that differed in CDS length (CDS-APA). For example,

among transcripts with two poly(A) sites, 19% (76) showed significant differences in CDS-APA between Purkinje and granule (Figure 2C). 7 instances of CDS-APA altered the inclusion of phosphorylation sites and 29 instances altered the inclusion of known protein domains, including those with enzymatic activities and those mediating protein-protein interactions (Table S2).

Among transcripts differing in CDS between Purkinje and granule cells was *NCAM1*, which codes for a neural cell adhesion molecule implicated in neurite outgrowth (Fields and Itoh, 1996); granule cells mostly expressed the full-length isoform of *NCAM1*, whereas Purkinje cells mostly expressed an isoform coding for a truncated protein (Figure 2D). We observed the opposite pattern for *Copg1*, a protein coding for a subunit of the coatamer complex (Hahn et al., 2000); granule cells expressed an isoform of *Copg1* coding for a truncated protein in addition to the full-length isoform, whereas Purkinje cells only expressed the full-length isoform (Figure 2D).

We also found that previously published TRAP-Seq data from granule and Purkinje cells (Mellén et al., 2012) was consistent with the data from cTag-PAPERCLIP for the above mentioned genes (*Vezt*, *Zfp609*, *NCAM1* and *Copg1*) (Figure 2 – figure supplement 1A and B). However, TRAP-Seq was not able to pinpoint 3'UTR ends accurately (Figure 2 – figure supplement 1A), underscoring the additional resolution of RNA regulation conferred by cTag-PAPERCLIP.

We observed that genes involved in the regulation of cell morphology and ion transport were significantly enriched among genes expressing different 3'UTR isoforms in the two cell types (Table S3 and Figure 2E, which shows the first 6 most enriched relevant functional categories). The genes included *Sept11*, which has been shown to regulate dendritic branching (X. Li et al., 2009); *Trio*, which has been shown to regulate dendritic branching and synapse function (Ba et al., 2016) and *Ube3a*, which has been

shown to regulate neuronal excitability (Judson et al., 2016). This observation suggests that the striking morphological and physiological differences between granule (excitatory, sparse dendrites) and Purkinje (inhibitory, extensive dendrites) cells may at least in part be mediated by fine-tuning gene expression via APA.

APA Alters 3'UTR Length and CDS of Specific Transcripts During Granule Cell Development

To determine if polyadenylation sites are dynamically selected within a given neuronal type as a cell transitions from a mitotic precursor to a post-mitotic, differentiated neuron, we profiled APA in granule cells during development. We analyzed APA at two time points: P0, when granule cell precursors are proliferating in the external granule layer, and P21, when granule cells have completed their migration to the internal granule layer and have made axonal synaptic connections with Purkinje cells, as well as dendritic synaptic connections with mossy fiber and Golgi cell inputs (Hatten and Heintz, 1995). We bred cTag-PABP mice with mice expressing Cre under the control of the *Math1* promoter (Matei et al., 2005) to purify PABP-bound transcripts from both granule cell precursors and differentiated granule neurons. Although *Math1* is only expressed in granule cell precursors, their progeny – the differentiated granule cells – inherit the recombined *PABPC1-GFP* locus.

We first confirmed that PABPC1-GFP is expressed in precursor and differentiated granule neurons by immunostaining on P0 and P21 cerebella (Figure 3A). We then performed cTag-PAPERCLIP on dissected cerebellar cortices (discarding deep cerebellar nuclei, which contain non-granule cell *Math1*-expressing neurons). We obtained 12,014 and 14,491 clusters of reads, which belonged to 9,698 and 10,716 genes expressed in granule cell precursors (from P0 mice) and differentiated granule

cells (from P21 mice), respectively. 32% of genes in granule cell precursors (3,104) and 32% of genes in differentiated granule cells (3,436) had multiple clusters.

Comparing cTag-PAPERCLIP data between precursor and differentiated granule cells from P0 and P21 mice, we found 737 genes that significantly changed in APA (FDR < 0.05) (Table S4). 3'UTRs tended to lengthen during granule cell development, which is consistent with RNA expression data from whole developing mouse brains (Ji et al., 2009). Specifically, among differentially polyadenylated transcripts with two poly(A) sites in 3'UTRs, 288 out of 354 got longer during granule cell development (Figure 3B). For example, *Kpnb1*, which codes for importin subunit $\beta 1$, whose long 3'UTR isoform has been shown to localize the mRNA to axons (Fainzilber et al., 2012), and *Cdk7*, which has been shown to be required for cell cycle progression of neurons (Abdullah et al., 2016), both shifted to predominantly expressing a long 3'UTR isoform in differentiated granule cells (Figure 3C).

In assessing 3'UTR regulation during granule cell development, we also discovered transcripts where APA altered their CDS. Among differentially polyadenylated transcripts with two poly(A) sites, 98 (22%) displayed changes in CDS. In the majority of cases (69/98), CDS lengthened during granule cell differentiation (Figure 3D). Such changes in CDS content indicate that APA can act to modify protein function during granule cell development (Figure 3D). For example, 10 of these APA instances altered the inclusion of sites for posttranscriptional modifications and 42 APA instances altered protein domains, such as those with enzymatic, DNA-binding, and protein-binding activity (Table S5).

An example of the effect of APA change during granule cell development is illustrated by examining cTag PAPERCLIP results for the *Sin3b* transcript, which codes for a scaffold subunit of the Sin3-histone deacetylase (HDAC) transcriptional repression

complex (Silverstein and Ekwall, 2004), and which has been shown to be necessary for cell cycle exit and differentiation (David et al., 2008). Differentiated granule cells express the long isoform of *Sin3b*, whereas granule cell precursors express both the long isoform and a short isoform lacking the HDAC-interaction domain (Figure 3E). These findings suggest that changes in APA of *Sin3b* mRNA during granule cell differentiation may affect the abundance of the Sin3-HDAC complex, which may in turn affect the exit of granule cells from the cell cycle during development. Another example is evident in cTag PAPERCLIP analysis of APA in the *Rbmx* gene, which encodes a protein required for proper brain development in zebrafish (Tsend Ayush et al., 2005). Whereas both the short and the long isoform of *Rbmx* are expressed in granule cell progenitors, only the long isoform is expressed in differentiated granule cells (Figure 3E). The long isoform is predicted to be degraded via nonsense-mediated decay (Aken et al., 2016), which may contribute to a decrease in abundance of the *Rbmx* transcript that we observed in differentiated granule cells (Figure 3E).

To more systematically corroborate these cTag-PAPERCLIP data, we sequenced RNA isolated from FACS-sorted GFP-positive granule cells from P0 and P21 *Math1-Cre; PABP^{cTag}* mice. The total number of cTag-PAPERCLIP reads per gene (i.e. a measure of the total abundance of 3'UTR isoforms) was highly correlated to the density of RNA-seq reads per gene (RPKM, averaged over all exons of a gene) ($R = 0.56$, Figure 3 – figure supplement 1). Moreover, 73% of genes that significantly changed 3'UTR isoform expression during granule cell development as measured by cTag-PAPERCLIP data also changed in the same direction as measured by RNA-seq. RNA sequencing data for the genes mentioned above (*Kpnb1*, *Cdk7*, *Sin3b* and *Rbmx*) are shown in Figure 3 – figure supplement 2A and B. While the pattern of RNA sequencing

reads covering the 3'UTRs was consistent with the cTag-PAPERCLIP data, RNA sequencing cannot pinpoint the exact 3'UTR ends.

Genes that expressed different 3'UTR isoforms in progenitor and differentiated granule cells were significantly enriched for genes involved in regulating neuritogenesis and dendritic branching (Table S6 and Figure 3F, which shows the first 6 most enriched relevant functional categories). These genes include *Nrcam* (Sakurai, 2012), which has been shown to regulate axon growth; *Pds5b*, which has been shown to regulate neuronal migration and axon growth (B. Zhang et al., 2007) and *Dclk1*, which has been shown to regulate dendritic vesicle transport and dendrite development (Lipka et al., 2016). That these functions are enriched suggests that APA plays a role in defining the morphological changes between precursor and differentiated granule cells.

Regulatory elements in non-coding 3'UTRs regulate mRNA stability and its association with ribosomes (Tian and Manley, 2016). For example, it has been shown that 3'UTR extensions acquired during brain development harbor functional, Argonaute-bound miRNA target sites (Hwang et al., 2016; Miura et al., 2013). We therefore asked whether changes in APA affected mRNA abundance and its association with ribosomes during granule cell development. We analyzed the abundance of ribosome-associated mRNAs from a recent TRAP study on developing cerebellar granule cells (Zhu et al., 2016). Acquiring a longer or shorter 3'UTR during development was not globally correlated to a change in ribosome-associated mRNA abundance (Figure 3 – figure supplement 3), which is broadly consistent with previous work (Gruber et al., 2014; Gupta et al., 2014). The lack of a global trend suggests that if APA affects mRNA abundance, it does so in either direction, depending on the gene.

Finally, we asked whether changes in APA during granule cell development contribute to the differences in APA between mature Purkinje and granule cells. ~15% of

transcripts with two poly(A) sites that significantly increased distal poly(A) site usage during granule cell development were significantly shorter in Purkinje cells (overlap $p=6.85 \times 10^{-8}$, hypergeometric test, Figure 4A). This significant overlap suggests the increase in the use of distal poly(A) sites is a developmentally controlled process to derive neuron-type-specific 3'UTR regulation of mRNA transcripts in the cerebellar granule cells. Examples of genes that showed greater expression of the long 3'UTR isoform in differentiated granule cells compared to proliferating granule cell precursors and Purkinje cells include (Figure 4B) *Mbd2*, which codes for methyl CpG-binding domain protein 2 and has been shown to regulate the proliferation of olfactory receptor neuron precursors (J. L. MacDonald et al., 2010), *Tmem57*, which codes for macoilin, and has been shown to regulate neuronal excitability (Arellano-Carbajal et al., 2011), and *Memo1*, which we describe in more detail below.

***Memo1* Regulates Granule Cell Proliferation and its Expression is Developmentally Regulated by APA and miR-124**

Little is known about the physiological consequences of altered 3'UTR length during development (Boutet et al., 2012; Pinto et al., 2011). To better understand these functional consequences, we focused on APA in *Memo1*, a gene that changed 3'UTR isoform expression during granule cell development. *Memo1* has been implicated in promoting cell motility (G. MacDonald et al., 2014) and in the proliferation of breast cancer cells (Sorokin and Chen, 2012), processes which are analogous to events regulated during granule cell development. Granule cell precursors express only the short 3'UTR isoform of *Memo1*, whereas differentiated granule cells express both the short and a long, previously unannotated, isoform ending 14 kb farther downstream (Figure 5A). Both the proximal and the distal poly(A) signals of *Memo1*, ending the short

and the long 3'UTRs, respectively, are highly conserved across placental mammals, and both isoforms are detectable in human brain RNA (data from (Jaffe et al., 2015)). Although they are most salient in cTag-PAPERCLIP data, the presence of both isoforms is consistent with RNA-sequencing data from FACS-sorted granule cells as well as with published TRAP-Seq data (Mellén et al., 2012) (Figure 5 – figure supplement 1A). Interestingly, the overall expression of *Memo1* – as measured by cTag-PAPERCLIP – significantly decreased as granule cells differentiate (Figure 5B). We therefore hypothesized that *Memo1* plays a role in proliferating granule cell precursors, when its expression is highest. We addressed this question in the cerebellum of E18.5 *Memo1* knockout mice (Van Otterloo et al., 2016), given that these mice die after birth. We observed that the cerebella of *Memo1* knockout mice lacked the typical pattern of fissures, i.e. furrows between which the cerebellar lobes will form. At E18.5, control cerebella had five fissures, whereas *Memo1* knockout cerebella had only one fissure (Figure 5 – figure supplement 1B). Given that the formation of cerebellar fissures is thought to be driven by the proliferation of granule cells (Sudarov and Joyner, 2007), we quantified the number of mitotic cells in the proliferating external granule layer (using anti-phospho-H3 antibody) and found significantly fewer mitotic cells in the external granule layer of *Memo1* knockout mice (Figure 5C and S4B). These data suggest that *Memo1* is required for the proliferation of granule cells, consistent with its role in the proliferation of breast cancer cells (Sorokin and Chen, 2012).

To identify a potential functional role for the long *Memo1* 3'UTR acquired during granule cell development, we analyzed the short and the long *Memo1* 3'UTRs for known regulatory sequence motifs (using RegRNA software (Chang et al., 2013)). From this analysis, we identified a highly conserved miR-124 target site on the long 3'UTR that was absent in the short 3'UTR (Figure 5A). This miR-124 target site was also bound by

Argonaute in the mouse cortex, as evidenced by Argonaute CLIP (Moore et al., 2015) (Figure 5A), suggesting that this target site is functional in the mouse brain. miR-124 is a neuron-specific miRNA that has been shown to be upregulated during differentiation of neural stem cells in the subventricular zone and is required for cell cycle exit and differentiation of neuroblasts (Cheng et al., 2009). Thus, we asked whether miR-124 expression also increases during granule cell differentiation. Indeed, we found that miR-124 expression increased between purified granule cell precursors and these same precursors differentiated *in vitro* for 2 weeks, when they morphologically resemble differentiated granule cells (Figure 5D). In addition, *in situ* hybridization experiments have shown high levels of miR-124 in the granule layer of adult mouse cerebella (Pena et al., 2009). These results demonstrate that as the expression of miR-124 increases, *Memo1* expression decreases, suggesting a potential role for miR-124 in downregulating *Memo1* expression during granule cell development.

To test whether miR-124 regulates the stability of *Memo1* via its target site on the 3'UTR long isoform, we blocked miR-124 from binding to its targets by transfecting cultured primary granule cells with an antisense oligonucleotide against miR-124. As expected if miR-124 decreases *Memo1* abundance, the *Memo1* long isoform was upregulated under these conditions compared to a scrambled control (Figure 5E, antagomir). We further corroborated this miR-124-*Memo1* interaction by blocking the miR-124 target site on *Memo1* with an antisense oligonucleotide (Figure 5F, target blocker). These data demonstrate that miR-124 downregulates the *Memo1* long isoform.

Together, these results show that *Memo1* regulates the proliferation of granule cell precursors and cerebellar foliation. During granule cell development, *Memo1* acquires a 3'UTR extension that harbors a miR-124 target site. This extension of the *Memo1* 3'UTR provides a mechanism for *Memo1* downregulation during development.

DISCUSSION

Previous studies have shown that APA can differ between tissues (Lianoglou et al., 2013; Miura et al., 2013; Smibert et al., 2012; H. Zhang et al., 2005) and that 3'UTRs tend to lengthen during embryonic development (Ji et al., 2009; Ulitsky et al., 2012). Here, we studied differences in APA between specific cell types *in vivo* using cTag-PAPERCLIP. Cellular properties have typically been thought of as being controlled at the level of transcription (Jessell, 2000), but our results suggest that post-transcriptional RNA regulation, such as APA, may also contribute to the distinct morphological and physiological properties of cerebellar Purkinje and granule cells, as well as to the different properties of precursor and differentiated granule cells.

We found many differences in APA between precursor and differentiated granule neurons, particularly in genes involved in neuronal development (Figure 3). These APA switches may regulate the developmental transition between proliferating granule cell precursors to differentiated granule neurons. For example, *Memo1* is required for granule cell proliferation. As granule cell precursors differentiate, *Memo1* increases the relative expression of its long 3'UTR isoform, which harbors a functional miR-124 site (Figure 5), that in turn contributes to the downregulation of *Memo1*.

In addition, APA switches during development may contribute to the properties of different mature cell types. We found numerous differences in APA between Purkinje and granule neurons of the cerebellum (Figure 2), many of which arose during granule cell differentiation (Figure 4). Therefore, we hypothesize that previously observed transcript lengthening during brain development (Ji et al., 2009; Miura et al., 2013) occurs in different transcripts across cell types to enable cell type specific post-transcriptional regulation in the adult nervous system, which contributes to the functional diversity between neurons. In particular, we found that *Memo1* undergoes APA in a cell

type specific way: Purkinje cells predominantly express the short 3'UTR isoform of *Memo1*, whereas granule neurons express a higher proportion of the miR-124-sensitive long 3'UTR isoform. Given that miR-124 is expressed in both Purkinje and granule cells (He et al., 2012; Pena et al., 2009), the selective use of the distal *Memo1* poly(A) signal allows for *Memo1* mRNA to be specifically downregulated in granule cells (~3-fold lower compared to Purkinje cells). *Memo1* remains expressed at higher levels in Purkinje cells, where it may play a post-mitotic role. This mechanism of cell type-specific post-transcriptional regulation of *Memo1* is similar to the regulation of *Pax3* by miR-206 in conjunction with APA in muscle stem cells. *Pax3* expresses alternative 3'UTR isoforms based on the location of the muscle (limb vs. diaphragm muscle), which leads to a difference in the abundance of *Pax3* mRNA between these locations (Boutet et al., 2012). Interestingly, among genes that expressed more of a longer 3'UTR isoform in granule cells compared to Purkinje cells, 150 genes were expressed at lower levels in granule cells compared to Purkinje cells and thus are potentially downregulated by miRNAs. Therefore, there are potentially more genes like *Memo1* and *Pax3* that exhibit APA-dependent miRNA-mediated downregulation in specific cell types.

The factors that drive the differences in APA between the cell types profiled here remain unidentified. One possibility is that the overall levels of polyadenylation factors affect polyadenylation site selection. The overall expression levels of polyadenylation factors have been shown to decrease during cell differentiation, as 3'UTRs lengthen, and increase during dedifferentiation, as 3'UTRs shorten (Elkon et al., 2012; Ji and Tian, 2009). Indeed, polyadenylation factors as a group (Shi et al., 2009) were slightly (but not significantly) downregulated (17% median decrease) between granule cell precursors and differentiated granule cells. These observations suggest that an overall decrease in the levels of the core polyadenylation machinery may contribute to the overall

lengthening of 3'UTRs during granule cell development. Whether additional levels of APA regulation, other than the overall levels of polyadenylation factors, exist to modulate APA in the genes discussed here will need to be investigated in the future.

Finally, an especially interesting group of transcripts that show differential APA across cell types are those that differ in CDS. We found a number of APA instances that control the inclusion of known protein domains and protein modifications both during *in vivo* neuronal development and across neuronal classes. As with 3'UTR-APA, these transcripts tend to be longer in differentiated granule cells compared to proliferating precursors, which is consistent with data from MCF10A cell line (Elkon et al., 2012), where proliferating cells tend to express isoforms with shorter CDS compared to non-proliferating, serum-starved cells. This finding suggests that there are common regulatory mechanisms affecting both 3'UTR and CDS length during neuronal differentiation, which result in fine-tuning of both the quantity and quality of cell-specific transcripts produced in different kinds of neurons.

METHODS

Mice

Animals were maintained in an AAALAC-approved animal facility and all procedures were performed in accordance with IACUC guidelines. *Pcp2/L7-Cre*, *Neurod1-Cre* and *Math1-Cre* mice were obtained from The Jackson Laboratory (catalog numbers: 004146, 028364 and 011104). *Memo1* mutant mice were generated using ES-cells from the European Conditional Mouse Mutagenesis (EUCOMM) repository as described previously (Van Otterloo et al., 2016). Mouse studies were allocated by genotype, and were not blinded.

Immunohistochemistry

Adult mice used for immunostaining (*Neurod1-Cre; Pabpc1^{cTag}*, *Pcp2-Cre; Pabpc1^{cTag}* and *Math1-Cre; Pabpc1^{cTag}* mice) were perfused with PBS and 4% paraformaldehyde (PFA) and fixed in 4% PFA/PBS at 4°C overnight. Heads from E18.5 (*Memo1* mutant mice and WT littermates) and cerebella from P0 mice (*Math1-Cre; Pabpc1^{cTag}* mice) were fixed in 4% PFA/PBS at 4°C overnight without perfusion. Fixed heads/cerebella were then incubated in 15% sucrose/PBS followed by 30% sucrose/PBS at 4°C and embedded in O.C.T. compound (Sakura Finetek). Adult cerebella were sliced into 30 µm thick sections; E18.5 and P0 cerebella were sliced to 80 µm thick sections on a cryostat (CM3050S, LEICA). The sections were then washed three times with PBS at room temperature (RT), incubated with 0.2% Triton X-100/PBS at RT, blocked with 1.5% normal donkey serum (NDS)/PBS at RT, and incubated overnight at 4°C with primary antibodies in 1.5% NDS/PBS followed by incubation with Alexa 488 and 555 conjugated donkey secondary antibodies (Thermo Fisher, 1:1000 dilution) in 1.5% NDS/PBS.

Images of immunostained sections were taken using BZ-X700 (KEYENCE) microscope. We used the following primary antibodies for immunohistochemistry: anti-Calb1 (Santa Cruz, catalog number: sc-7691, 1:250 dilution) anti-GFP (AVES, catalog number: GFP-1020, 1:1000 dilution), anti-phospho histone H3 (Cell signaling, catalog number: 9701, 1:100 dilution). Validation of *Pabpc1*-GFP expression was done on one mouse for each genotype, in at least two technical replicates. The number of phospho-histone H3 positive cells per area of external granule layer was quantified using Photoshop by manually counting phospho-histone H3 positive cells (using Count Tool) and dividing the number with the total external granule layer area. Quantification of phospho-histone H3 positive cells was performed on two *Memo1* knockout mice and two WT mice (from two different litters, knockout and WT mice were littermates), each in four technical replicates (i.e. four sections from the middle of the cerebellum).

cTag-PAPERCLIP

The PAPERCLIP procedure was performed as previously described (Hwang et al., 2016). Mouse monoclonal anti-GFP clones 19F7 and 19C8 (Heiman et al., 2008) were used for immunoprecipitation. Individual cTag-PAPERCLIP libraries were multiplexed and sequenced on MiSeq (Illumina) to obtain 75-nt single-end reads. cTag-PAPERCLIP on granule cells was performed in 4 replicates, on Purkinje cells in 3 replicates and developing granule cells in 3 replicates. Due to low amount of RNA obtained by cTag-PAPERCLIP, we pooled 2 mice per replicate for cTag-PAPERCLIP on adult granule cells (*Neurod1-Cre; Pabpc1^{cTag}* and *Math1-Cre; Pabpc1^{cTag}* mice), 7 mice per replicate for cTag-PAPERCLIP on Purkinje cells (*Pcp2-Cre; Pabpc1^{cTag}* mice) and 10 mice per replicate for cTag-PAPERCLIP on granule cell precursors (*Math1-Cre; Pabpc1^{cTag}* mice).

Analysis of cTag-PAPERCLIP data

The processing of raw reads was performed using the CIMS software package as previously described (Moore et al., 2014). Raw reads were filtered based on quality score. Filtered reads with the exact sequence were collapsed into one. Poly(A) sequence at the 3' end was then trimmed using CutAdapt (Martin, 2011). Only reads that are at least 25-nt in length were mapped to mm10 reference genome. Mapping was performed using Novoalign (Novocraft) without trimming. Reads mapping to the same genomic positions without distinct barcodes were further collapsed into a single read as previously described (Moore et al., 2014). CIMS software package was then used to cluster overlapping collapsed reads from all biological replicates (for each condition) and to determine the number of reads in each cluster. To define a list of high confidence clusters we used the following criteria: clusters had to contain reads from all replicates in each condition, the number of reads in a cluster had to be at least 10% of gene total, clusters had to be located within 20 kb from 3' ends annotated by RefSeq, clusters that were located upstream of a stretch of 6 or more adenines were excluded due to potential internal priming of the reverse transcription primer. To identify differences in APA between different conditions, Fisher's exact test was performed comparing the ratio between reads in a particular cluster and the sum of reads in all other clusters in a gene in the two conditions. The output p-values were adjusted for multiple hypotheses testing using the Benjamini-Hochberg method. To determine which genes are displaying 3'UTR vs. CDS-APA, we assigned cTag-PAPERCLIP clusters to the nearest Ensembl-annotated 3'UTR using closestBed from the BEDtools suite (Quinlan and Hall, 2010). If the start sites of the assigned 3'UTRs for the same gene differed, we assigned the gene as displaying CDS-APA. We manually inspected the CDS-APA gene list and removed a small number of genes that we were not able to unequivocally assign to either 3'UTR-

APA or CDS-APA.

To determine overlap between Ensembl 3'UTR ends and cTag-PAPERCLIP clusters, the 3'UTR ends of Ensembl transcripts (Ensembl 89) were extended 10 nucleotides upstream from the end and overlapped with cTag-PAPERCLIP clusters using IRanges and GenomicRanges packages (Lawrence et al., 2013).

To compare cTag-PAPERCLIP data with TRAP-Seq data, TRAP-Seq data were mapped to mm10 with Novoalign using default parameters. RPKM values of Ensembl transcripts from TRAP-Seq were then compared to the total number of reads from cTag-PAPERCLIP for the same transcripts.

Protein domains for CDS-APA analysis were obtained manually from Ensembl genome browser (www.ensembl.org). For each alternative transcript, we inspected whether they harbor any known protein domain from protein domain databases (Superfamily, SMART, Pfam and Prosite). Protein modifications were downloaded from Uniprot (The UniProt Consortium, 2017) and mapped to mm10 using backlocate software (<http://lindenb.github.io/jvarkit/>). Genomic coordinates of protein modifications were then overlapped with alternative 3'UTR isoforms to find those that are unique to the long isoform.

To analyze the relationship between 3'UTR isoform expression changes and the abundance of ribosome-associated RNA between P0 and P21 granule cells, microarray data from P0 and P21 granule cells (Zhu et al., 2016) were processed using gcrma (Irizarry and Wu, 2017) and affy (Gautier et al., 2004) software packages, and logFC values were calculated using limma (Ritchie et al., 2015) package.

Gene ontology analysis

Gene ontology analysis was performed on genes showing significant changes in 3'UTR isoform expression using Ingenuity Pathway Analysis software (Qiagen) Core Analysis tool. Background was determined as all genes with cTag-PAPERCLIP clusters expressed in Purkinje and granule cells or P0 and P21 granule cells. 'User Dataset' was chosen as 'Reference Set.' Genes with significant differences in APA were given 'Expr Other' value of 1, all other genes were given 'Expr Other' value of 0. Cutoff was chosen as 1. We focused on functions related to 'Molecular and Cellular Functions' and 'Physiological System Development and Function' that are overrepresented in the set of genes that show significant changes in 3'UTR isoform expression between Purkinje and granule cells and during development of granule cells. For the Figure 2D and 3E we manually removed categories that were not relevant or contained very similar gene sets (Supplementary Tables contain all significant functional categories).

FACS sorting and RNA sequencing

GFP-positive granule cells from P0 and P21 *Math1-Cre; Pabpc1^{cTag}* mice were sorted using FACS Aria II (BD). Three P0 and two P21 mice were used for this experiment. Post-sort analysis showed that the sorted samples contained 99% of GFP-positive cells. RNA was isolated from sorted cells using Roche High Pure RNA isolation kit as per manufacturer's instructions. RNA sequencing library was prepared using TruSeq kit from Illumina, following manufacturer's instructions. Individual RNA-seq libraries were multiplexed and sequenced on HiSeq (Illumina) to obtain 100-nt paired-end reads. RNA-sequencing reads were mapped to the mouse genome (mm10) using Novoalign (Novocraft), reads overlapping genes were counted using HTSeq-count (Anders et al.,

2015). The total number of cTag-PAPERCLIP reads per transcript was then compared to the total number of RNA-sequencing reads per transcript.

Granule cell purification and culture

Granule cells were prepared as described (Hatten, 1985). Briefly, cerebella were dissected away from the brains of P6 mice. After the pial layer was peeled away, the tissue was treated with trypsin and triturated into a single-cell suspension using fine bore Pasteur pipettes. The suspension was layered on a discontinuous Percoll gradient and separated by centrifugation. The small cell fraction was isolated, and granule cells were further enriched by panning on tissue culture treated plastic dishes. The resulting cultures routinely contain greater than 95% of cells of the granule cell lineage (Hatten, 1985). Granule cells were plated in serum-free granule cell medium on Poly-D lysine coated 6-well plates and transfected with 50 nM miR-124-3p antagomir (Exiqon) or Memo1 target blocker (Exiqon, custom miRCURY LNA Power Inhibitor, final concentration 50 nM for both) and appropriate negative control using Lipofectamine RNAiMAX (Thermo Fisher). Cells were then let to differentiate (4 days) and RNA was isolated using TRIzol (Thermo Fisher) followed by High Pure RNA Isolation Kit (Roche) following manufacturer's instructions. The experiment with miR-124-3p antagomir was performed in two biological replicates (each biological replicate was performed in two technical replicates) and the experiment with Memo1 target blocker was performed in three biological replicates (each biological replicate was performed in two technical replicates). Biological replicate is defined as an independent experiment (an independent isolation of primary granule cells from P6 pups followed by *in vitro* differentiation and RNA isolation). Technical replicate is defined as an independent quantitative PCR reaction.

Reverse transcription and quantitative PCR

Quantification of Memo1 long isoform in granule cell culture was performed by first isolating RNA using High Pure RNA Isolation Kit (Roche), followed by reverse transcription using SuperScript III reverse transcriptase (Thermo Fisher) and quantitative PCR using FastStart SYBR Green Master mix (Roche). Primers to amplify long Memo1 isoform were: TCCCCTACTCTCCCAGTAGTC and AGGACTTGTTGCTTCCAGGAC. We used the following cycling parameters: 95°C for 10 min. followed by 40 cycles of 95°C for 15 sec., 58°C for 30 sec., 72°C for 20 sec. Fold changes were calculated using Bio-Rad CFX Manager Software. To quantify miR-124 we isolated RNA from granule cells using High Pure RNA Isolation Kit (Roche) and reverse transcribed it using miScript II RT Kit (Qiagen) following manufacturer's instructions. We then performed quantitative PCR using FastStart SYBR Green Master mix (Roche) using the following parameters: 94°C for 15 min, 40 cycles (15 sec at 94°C, 30 sec at 55°C and 30 sec at 70°C) and primers: TAAGGCACGCGGTGAATGCC (aligns to mature miR-124) and GAATCGAGCACCAGTTACGC (aligns to the 'universal tag' added to miRNA using miScript II RT Kit).

ACKNOWLEDGEMENTS

We thank Trevor Williams at the University of Colorado for assistance with the *Memo1* null mice and Xiaodong Zhu and Yin Fang from the Hatten lab for help with the experiments related to granule cell development. We thank Chris Park for advice on data analysis and other members of the Darnell and Hatten labs for helpful discussions. We also thank the Rockefeller University Flow Cytometry Facility. This work was in part supported by grants from the National Institutes of Health (NS034389 and NS081706) and Simons Foundation (SFARI 240432) to R.B.D and by grant K99DE026823 from the NIDCR to E.V.O. R.B.D is an Investigator of the Howard Hughes Medical Institute.

COMPETING INTERESTS STATEMENT

The authors declare no competing interests.

REFERENCES

- Abdullah, A.I., Zhang, H., Nie, Y., Tang, W., Sun, T., 2016. CDK7 and miR-210 Co-regulate Cell-Cycle Progression of Neural Progenitors in the Developing Neocortex. *Stem Cell Reports* 7, 69–79. doi:10.1016/j.stemcr.2016.06.005
- Aken, B.L., Ayling, S., Barrell, D., Clarke, L., Curwen, V., Fairley, S., Fernandez Banet, J., Billis, K., García Girón, C., Hourlier, T., Howe, K., Kähäri, A., Kokocinski, F., Martin, F.J., Murphy, D.N., Nag, R., Ruffier, M., Schuster, M., Tang, Y.A., Vogel, J.-H., White, S., Zadissa, A., Flicek, P., Searle, S.M.J., 2016. The Ensembl gene annotation system. *Database (Oxford)* 2016, baw093. doi:10.1093/database/baw093
- An, J.J., Gharami, K., Liao, G.-Y., Woo, N.H., Lau, A.G., Vanevski, F., Torre, E.R., Jones, K.R., Feng, Y., Lu, B., Xu, B., 2008. Distinct role of long 3' UTR BDNF mRNA in spine morphology and synaptic plasticity in hippocampal neurons. *Cell* 134, 175–187. doi:10.1016/j.cell.2008.05.045
- Anders, S., Pyl, P.T., Huber, W., 2015. HTSeq—a Python framework to work with high-throughput sequencing data. *Bioinformatics* 31, 166–169. doi:10.1093/bioinformatics/btu638
- Arellano-Carbajal, F., Briseño-Roa, L., Couto, A., Cheung, B.H.H., Labouesse, M., de Bono, M., 2011. Macoilin, a Conserved Nervous System-Specific ER Membrane Protein That Regulates Neuronal Excitability. *PLoS Genet* 7, e1001341. doi:10.1371/journal.pgen.1001341
- Ba, W., Yan, Y., Reijnders, M.R.F., Schuurs-Hoeijmakers, J.H.M., Feenstra, I., Bongers, E.M.H.F., Bosch, D.G.M., De Leeuw, N., Pfundt, R., Gilissen, C., De Vries, P.F., Veltman, J.A., Hoischen, A., Mefford, H.C., Eichler, E.E., Vissers, L.E.L.M., Nadif Kasri, N., De Vries, B.B.A., 2016. TRIO loss of function is associated with mild intellectual disability and affects dendritic branching and synapse function. *Human Molecular Genetics* 25, 892–902. doi:10.1093/hmg/ddv618
- Barski, J.J., Dethleffsen, K., Meyer, M., 2000. Cre recombinase expression in cerebellar Purkinje cells. *genesis* 28, 93–98.
- Berkovits, B.D., Mayr, C., 2015. Alternative 3' UTRs act as scaffolds to regulate membrane protein localization. *Nature* 522, 363–367. doi:10.1038/nature14321
- Boutet, S.C., Cheung, T.H., Quach, N.L., Liu, L., Prescott, S.L., Edalati, A., Iori, K., Rando, T.A., 2012. Alternative Polyadenylation Mediates MicroRNA Regulation of Muscle Stem Cell Function. *Stem Cell* 10, 327–336. doi:10.1016/j.stem.2012.01.017
- Buckner, R.L., 2013. The Cerebellum and Cognitive Function: 25 Years of Insight from Anatomy and Neuroimaging. *Neuron* 80, 807–815. doi:10.1016/j.neuron.2013.10.044
- Butts, T., Green, M.J., Wingate, R.J.T., 2014. Development of the cerebellum: simple steps to make a 'little brain'. *Development* 141, 4031–4041. doi:10.1242/dev.106559
- Chang, T.-H., Huang, H.-Y., Hsu, J.B.-K., Weng, S.-L., Horng, J.-T., Huang, H.-D., 2013. An enhanced computational platform for investigating the roles of regulatory RNA and for identifying functional RNA motifs. *BMC Bioinformatics* 14 Suppl 2, S4. doi:10.1186/1471-2105-14-S2-S4
- Cheng, L.-C., Pastrana, E., Tavazoie, M., Doetsch, F., 2009. miR-124 regulates adult neurogenesis in the subventricular zone stem cell niche. *Nat Neurosci* 12, 399–408. doi:10.1038/nn.2294
- David, G., Grandinetti, K.B., Finnerty, P.M., Simpson, N., Chu, G.C., Depinho, R.A., 2008. Specific requirement of the chromatin modifier mSin3B in cell cycle exit and

- cellular differentiation. *Proceedings of the National Academy of Sciences* 105, 4168–4172. doi:10.1073/pnas.0710285105
- Edmondson, J.C., Hatten, M.E., 1987. Glial-guided granule neuron migration in vitro: a high-resolution time-lapse video microscopic study. *J. Neurosci.* 7, 1928–1934.
- Elkon, R., Drost, J., van Haaften, G., Jenal, M., Schrier, M., Vrielink, J.A.O., Agami, R., 2012. E2F mediates enhanced alternative polyadenylation in proliferation. *Genome Biol* 13, R59. doi:10.1186/gb-2012-13-7-r59
- Fainzilber, R.-T.P.E.D.-M.E.I.I.R.S.D.M.T.G.C.M.M.C.G.D.G.J.T.A.Y.M., Doron-Mandel, E., Iavnilovitch, E., Rishal, I., Dagan, S.Y., Tsoory, M., Coppola, G., McDonald, M.K., Gomes, C., Geschwind, D.H., Twiss, J.L., Yaron, A., Fainzilber, M., 2012. Subcellular Knockout of Importin β 1 Perturbs Axonal Retrograde Signaling. *Neuron* 75, 294–305. doi:10.1016/j.neuron.2012.05.033
- Fields, R.D., Itoh, K., 1996. Neural cell adhesion molecules in activity-dependent development and synaptic plasticity. *Trends in Neurosciences* 19, 473–480. doi:10.1016/S0166-2236(96)30013-1
- Flavell, S.W., Kim, T.-K., Gray, J.M., Harmin, D.A., Hemberg, M., Hong, E.J., Markenscoff-Papadimitriou, E., Bear, D.M., Greenberg, M.E., 2008. Genome-wide analysis of MEF2 transcriptional program reveals synaptic target genes and neuronal activity-dependent polyadenylation site selection. *Neuron* 60, 1022–1038. doi:10.1016/j.neuron.2008.11.029
- Gautier, L., Cope, L., Bolstad, B.M., Irizarry, R.A., 2004. affy--analysis of Affymetrix GeneChip data at the probe level. *Bioinformatics* 20, 307–315. doi:10.1093/bioinformatics/btg405
- Gruber, A.R., Martin, G., Müller, P., Schmidt, A., Gruber, A.J., Gumienny, R., Mittal, N., Jayachandran, R., Pieters, J., Keller, W., van Nimwegen, E., Zavolan, M., 2014. Global 3' UTR shortening has a limited effect on protein abundance in proliferating T cells. *Nat Commun* 5, 5465. doi:10.1038/ncomms6465
- Gupta, I., Clauder-Münster, S., Klaus, B., Järvelin, A.I., Aiyar, R.S., Benes, V., Wilkening, S., Huber, W., Pelechano, V., Steinmetz, L.M., 2014. Alternative polyadenylation diversifies post-transcriptional regulation by selective RNA-protein interactions. *Mol. Syst. Biol.* 10, 719. doi:10.1002/msb.135068
- Hahn, Y., Lee, Y.J., Yun, J.H., Yang, S.K., Park, C.W., Mita, K., Huh, T.-L., Rhee, M., Chung, J.H., 2000. Duplication of genes encoding non-clathrin coat protein γ -COP in vertebrate, insect and plant evolution. *FEBS Letters* 482, 31–36. doi:10.1016/S0014-5793(00)02033-0
- Hatten, M.E., 1985. Neuronal regulation of astroglial morphology and proliferation in vitro. *The Journal of Cell Biology* 100, 384–396.
- Hatten, M.E., Heintz, N., 1995. Mechanisms of neural patterning and specification in the developing cerebellum. *Annu. Rev. Neurosci.* 18, 385–408. doi:10.1146/annurev.ne.18.030195.002125
- He, M., Liu, Y., Wang, X., Zhang, M.Q., Hannon, G.J., Huang, Z.J., 2012. Cell-type-based analysis of microRNA profiles in the mouse brain. *Neuron* 73, 35–48. doi:10.1016/j.neuron.2011.11.010
- Heiman, M., Schaefer, A., Gong, S., Peterson, J.D., Day, M., Ramsey, K.E., Suárez-Fariñas, M., Schwarz, C., Stephan, D.A., Surmeier, D.J., Greengard, P., Heintz, N., 2008. A Translational Profiling Approach for the Molecular Characterization of CNS Cell Types. *Cell* 135, 738–748. doi:10.1016/j.cell.2008.10.028
- Hwang, H.-W., Park, C.Y., Goodarzi, H., Fak, J.J., Mele, A., Moore, M.J., Saito, Y., Darnell, R.B., 2016. PAPERCLIP Identifies MicroRNA Targets and a Role of

- CstF64/64tau in Promoting Non-canonical poly(A) Site Usage. *CellReports* 15, 423–435. doi:10.1016/j.celrep.2016.03.023
- Hwang, H.-W., Saito, Y., Park, C.Y., Blachère, N.E., Tajima, Y., Fak, J.J., Zucker-Scharff, I., Darnell, R.B., 2017. cTag-PAPERCLIP Reveals Alternative Polyadenylation Promotes Cell-Type Specific Protein Diversity and Shifts Araf Isoforms with Microglia Activation. *Neuron* 95, 1334–1349.e5. doi:10.1016/j.neuron.2017.08.024
- Irizarry, R., Wu, Z., 2017. *gcrma: Background Adjustment Using Sequence Information* 1–6.
- Jaffe, A.E., Shin, J., Collado-Torres, L., Leek, J.T., Tao, R., Li, C., Gao, Y., Jia, Y., Maher, B.J., Hyde, T.M., Kleinman, J.E., Weinberger, D.R., 2015. Developmental regulation of human cortex transcription and its clinical relevance at single base resolution. *Nat Neurosci* 18, 154–161. doi:10.1038/nn.3898
- Jessell, T.M., 2000. Neuronal specification in the spinal cord: inductive signals and transcriptional codes. *Nat Rev Genet* 1, 20–29. doi:10.1038/35049541
- Ji, Z., Lee, J.Y., Pan, Z., Jiang, B., Tian, B., 2009. Progressive lengthening of 3' untranslated regions of mRNAs by alternative polyadenylation during mouse embryonic development. *Proceedings of the National Academy of Sciences* 106, 7028–7033. doi:10.1073/pnas.0900028106
- Ji, Z., Tian, B., 2009. Reprogramming of 3' Untranslated Regions of mRNAs by Alternative Polyadenylation in Generation of Pluripotent Stem Cells from Different Cell Types. *PLoS ONE* 4, e8419–13. doi:10.1371/journal.pone.0008419
- Judson, M.C., Wallace, M.L., Sidorov, M.S., Burette, A.C., Bin Gu, van Woerden, G.M., King, I.F., Han, J.E., Zylka, M.J., Elgersma, Y., Weinberg, R.J., Philpot, B.D., 2016. GABAergic Neuron-Specific Loss of Ube3a Causes Angelman Syndrome-Like EEG Abnormalities and Enhances Seizure Susceptibility. *Neuron* 90, 56–69. doi:10.1016/j.neuron.2016.02.040
- Lau, A.G., Irier, H.A., Gu, J., Tian, D., Ku, L., Liu, G., Xia, M., Fritsch, B., Zheng, J.Q., Dingledine, R., Xu, B., Lu, B., Feng, Y., 2010. Distinct 3'UTRs differentially regulate activity-dependent translation of brain-derived neurotrophic factor (BDNF). *Proceedings of the National Academy of Sciences* 107, 15945–15950. doi:10.1073/pnas.1002929107
- Lawrence, M., Huber, W., Pagès, H., Aboyoun, P., Carlson, M., Gentleman, R., Morgan, M.T., Carey, V.J., 2013. Software for Computing and Annotating Genomic Ranges. *PLoS Comput Biol* 9, e1003118–10. doi:10.1371/journal.pcbi.1003118
- Li, H.J., Kapoor, A., Giel-Moloney, M., Rindi, G., Leiter, A.B., 2012. Notch signaling differentially regulates the cell fate of early endocrine precursor cells and their maturing descendants in the mouse pancreas and intestine. *Developmental Biology* 371, 156–169. doi:10.1016/j.ydbio.2012.08.023
- Li, X., Serwanski, D.R., Miralles, C.P., Nagata, K.-I., De Blas, A.L., 2009. Septin 11 Is Present in GABAergic Synapses and Plays a Functional Role in the Cytoarchitecture of Neurons and GABAergic Synaptic Connectivity. *J. Biol. Chem.* 284, 17253–17265. doi:10.1074/jbc.M109.008870
- Lianoglou, S., Garg, V., Yang, J.L., Leslie, C.S., Mayr, C., 2013. Ubiquitously transcribed genes use alternative polyadenylation to achieve tissue-specific expression. *Genes & Development* 27, 2380–2396. doi:10.1101/gad.229328.113
- Licatalosi, D.D., Mele, A., Fak, J.J., Ule, J., Kayikci, M., Chi, S.W., Clark, T.A., Schweitzer, A.C., Blume, J.E., Wang, X., Darnell, J.C., Darnell, R.B., 2008. HITS-CLIP yields genome-wide insights into brain alternative RNA processing. *Nature*

- 456, 464–469. doi:10.1038/nature07488
- Lipka, J., Kapitein, L.C., Jaworski, J., Hoogenraad, C.C., 2016. Microtubule-binding protein doublecortin-like kinase 1 (DCLK1) guides kinesin-3-mediated cargo transport to dendrites. *The EMBO Journal* 35, 302–318. doi:10.15252/embj.201592929
- MacDonald, G., Nalvarte, I., Smirnova, T., Vecchi, M., Aceto, N., Dolemeyer, A., Frei, A., Lienhard, S., Wyckoff, J., Hess, D., Seebacher, J., Keusch, J.J., Gut, H., Salaun, D., Mazzarol, G., Disalvatore, D., Bentires-Alj, M., Di Fiore, P.P., Badache, A., Hynes, N.E., 2014. Memo is a copper-dependent redox protein with an essential role in migration and metastasis. *Sci. Signal.* 7, ra56. doi:10.1126/scisignal.2004870
- MacDonald, J.L., Verster, A., Berndt, A., Roskams, A.J., 2010. MBD2 and MeCP2 regulate distinct transitions in the stage-specific differentiation of olfactory receptor neurons. *Molecular and Cellular Neuroscience* 44, 55–67. doi:10.1016/j.mcn.2010.02.003
- Martin, M., 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnet.journal* 17, pp. 10–12. doi:10.14806/ej.17.1.200
- Matei, V., Pauley, S., Kaing, S., Rowitch, D., Beisel, K.W., Morris, K., Feng, F., Jones, K., Lee, J., Fritzsche, B., 2005. Smaller inner ear sensory epithelia in Neurog 1 null mice are related to earlier hair cell cycle exit. *Dev. Dyn.* 234, 633–650. doi:10.1002/dvdy.20551
- Mayr, C., Bartel, D.P., 2009. Widespread Shortening of 3'UTRs by Alternative Cleavage and Polyadenylation Activates Oncogenes in Cancer Cells. *Cell* 138, 673–684. doi:10.1016/j.cell.2009.06.016
- Mellén, M., Ayata, P., Dewell, S., Kriaucionis, S., Heintz, N., 2012. MeCP2 binds to 5hmC enriched within active genes and accessible chromatin in the nervous system. *Cell* 151, 1417–1430. doi:10.1016/j.cell.2012.11.022
- Miura, P., Shenker, S., Andreu-Agullo, C., Westholm, J.O., Lai, E.C., 2013. Widespread and extensive lengthening of 3' UTRs in the mammalian brain. *Genome Research* 23, 812–825. doi:10.1101/gr.146886.112
- Moore, M.J., Scheel, T.K.H., Luna, J.M., Park, C.Y., Fak, J.J., Nishiuchi, E., Rice, C.M., Darnell, R.B., 2015. miRNA-target chimeras reveal miRNA 3'-end pairing as a major determinant of Argonaute target specificity. *Nat Commun* 6, 8864. doi:10.1038/ncomms9864
- Moore, M.J., Zhang, C., Gantman, E.C., Mele, A., Darnell, J.C., Darnell, R.B., 2014. Mapping Argonaute and conventional RNA-binding protein interactions with RNA at single-nucleotide resolution using HITS-CLIP and CIMS analysis. *Nature Protocols* 9, 263–293. doi:10.1038/nprot.2014.012
- Pena, J.T.G., Sohn-Lee, C., Rouhanifard, S.H., Ludwig, J., Hafner, M., Mihailovic, A., Lim, C., Holoch, D., Berninger, P., Zavolan, M., Tuschl, T., 2009. miRNA in situ hybridization in formaldehyde and EDC-fixed tissues. *Nat Meth* 6, 139–141. doi:10.1038/nmeth.1294
- Pinto, P.A.B., Henriques, T., Freitas, M.O., Martins, T., Domingues, R.G., Wyrzykowska, P.S., Coelho, P.A., Carmo, A.M., Sunkel, C.E., Proudfoot, N.J., Moreira, A., 2011. RNA polymerase II kinetics in polo polyadenylation signal selection. *The EMBO Journal* 30, 2431–2444. doi:10.1038/emboj.2011.156
- Quinlan, A.R., Hall, I.M., 2010. BEDTools: a flexible suite of utilities for comparing genomic features. *Bioinformatics* 26, 841–842. doi:10.1093/bioinformatics/btq033
- Ritchie, M.E., Phipson, B., Wu, D., Hu, Y., Law, C.W., Shi, W., Smyth, G.K., 2015. limma powers differential expression analyses for RNA-sequencing and microarray studies.

- Nucleic Acids Research 43, e47–e47. doi:10.1093/nar/gkv007
- Sakurai, T., 2012. The role of NrCAM in neural development and disorders—Beyond a simple glue in the brain. *Molecular and Cellular Neuroscience* 49, 351–363. doi:10.1016/j.mcn.2011.12.002
- Sanda, M., Ohara, N., Kamata, A., Hara, Y., Tamaki, H., Sukegawa, J., Yanagisawa, T., Fukunaga, K., Kondo, H., Sakagami, H., 2010. Vezatin, a potential target for ADP-ribosylation factor 6, regulates the dendritic formation of hippocampal neurons. *Neuroscience Research* 67, 126–136. doi:10.1016/j.neures.2010.02.008
- Sandberg, R., Neilson, J.R., Sarma, A., Sharp, P.A., Burge, C.B., 2008. Proliferating cells express mRNAs with shortened 3' untranslated regions and fewer microRNA target sites. *Science* 320, 1643–1647. doi:10.1126/science.1155390
- Sanz, E., Evanoff, R., Quintana, A., Evans, E., Miller, J.A., Ko, C., Amieux, P.S., Griswold, M.D., McKnight, G.S., 2013. RiboTag Analysis of Actively Translated mRNAs in Sertoli and Leydig Cells In Vivo. *PLoS ONE* 8, e66179. doi:10.1371/journal.pone.0066179
- Shepard, P.J., Choi, E.A., Lu, J., Flanagan, L.A., Hertel, K.J., Shi, Y., 2011. Complex and dynamic landscape of RNA polyadenylation revealed by PAS-Seq. *RNA* 17, 761–772. doi:10.1261/rna.2581711
- Shi, Y., Di Giannamartino, D.C., Taylor, D., Sarkeshik, A., Rice, W.J., Yates, J.R., III, Frank, J., Manley, J.L., 2009. Molecular Architecture of the Human Pre-mRNA 3' Processing Complex. *Molecular Cell* 33, 365–376. doi:10.1016/j.molcel.2008.12.028
- Silverstein, R.A., Ekwall, K., 2004. Sin3: a flexible regulator of global gene expression and genome stability. *Curr Genet* 47, 1–17. doi:10.1007/s00294-004-0541-5
- Smibert, P., Miura, P., Westholm, J.O., Shenker, S., May, G., Duff, M.O., Zhang, D., Eads, B.D., Carlson, J., Brown, J.B., Eisman, R.C., Andrews, J., Kaufman, T., Cherbas, P., Celniker, S.E., Graveley, B.R., Lai, E.C., 2012. Global Patterns of Tissue-Specific Alternative Polyadenylation in *Drosophila*. *CellReports* 1, 277–289. doi:10.1016/j.celrep.2012.01.001
- Solecki, D.J., Trivedi, N., Govek, E.-E., Kerekes, R.A., Gleason, S.S., Hatten, M.E., 2009. Myosin II Motors and F-Actin Dynamics Drive the Coordinated Movement of the Centrosome and Soma during CNS Glial-Guided Neuronal Migration. *Neuron* 63, 63–80. doi:10.1016/j.neuron.2009.05.028
- Sorokin, A.V., Chen, J., 2012. MEMO1, a new IRS1-interacting protein, induces epithelial–mesenchymal transition in mammary epithelial cells 32, 3130–3138. doi:10.1038/onc.2012.327
- Sudarov, A., Joyner, A.L., 2007. Cerebellum morphogenesis: the foliation pattern is orchestrated by multi-cellular anchoring centers. *Neural Dev* 2, 26–22. doi:10.1186/1749-8104-2-26
- The UniProt Consortium, 2017. UniProt: the universal protein knowledgebase. *Nucleic Acids Research* 45, D158–D169. doi:10.1093/nar/gkw1099
- Tian, B., 2005. A large-scale analysis of mRNA polyadenylation of human and mouse genes. *Nucleic Acids Research* 33, 201–212. doi:10.1093/nar/gki158
- Tian, B., Manley, J.L., 2016. Alternative polyadenylation of mRNA precursors. *Nat Rev Mol Cell Biol* 18, 18–30. doi:10.1038/nrm.2016.116
- Tsend Ayush, E., O'Sullivan, L.A., Grützner, F.S., Onnebo, S.M.N., Lewis, R.S., Delbridge, M.L., Marshall Graves, J.A., Ward, A.C., 2005. RBMX gene is essential for brain development in zebrafish. *Dev. Dyn.* 234, 682–688. doi:10.1002/dvdy.20432
- Ule, J., Jensen, K.B., Ruggiu, M., Mele, A., Ule, A., Darnell, R.B., 2003. CLIP identifies

Nova-regulated RNA networks in the brain. *Science* 302, 1212–1215.

doi:10.1126/science.1090095

Ulitsky, I., Shkumatava, A., Jan, C.H., Subtelny, A.O., Koppstein, D., Bell, G.W., Sive, H., Bartel, D.P., 2012. Extensive alternative polyadenylation during zebrafish development. *Genome Research* 22, 2054–2066. doi:10.1101/gr.139733.112

van den Berg, D.L.C., Azzarelli, R., Oishi, K., Martynoga, B., Urbán, N., Dekkers, D.H.W., Demmers, J.A., Guillemot, F., 2017. Nipbl Interacts with Zfp609 and the Integrator Complex to Regulate Cortical Neuron Migration. *Neuron* 93, 348–361. doi:10.1016/j.neuron.2016.11.047

Van Otterloo, E., Feng, W., Jones, K.L., Hynes, N.E., Clouthier, D.E., Niswander, L., Williams, T., 2016. MEMO1 drives cranial endochondral ossification and palatogenesis. *Developmental Biology* 1–18. doi:10.1016/j.ydbio.2015.12.024

Wang, L., Dowell, R.D., Yi, R., 2013. Genome-wide maps of polyadenylation reveal dynamic mRNA 3'-end formation in mammalian cell lineages. *RNA* 19, 413–425. doi:10.1261/rna.035360.112

Zhang, B., Jain, S., Song, H., Fu, M., Heuckeroth, R.O., Erlich, J.M., Jay, P.Y., Milbrandt, J., 2007. Mice lacking sister chromatid cohesion protein PDS5B exhibit developmental abnormalities reminiscent of Cornelia de Lange syndrome. *Development* 134, 3191–3201. doi:10.1242/dev.005884

Zhang, H., Lee, J.Y., Tian, B., 2005. Biased alternative polyadenylation in human tissues. *Genome Biol* 6, R100–13. doi:10.1186/gb-2005-6-12-r100

Zhu, X., Girardo, D., Govek, E.-E., John, K., Mellén, M., Tamayo, P., Mesirov, J.P., Hatten, M.E., 2016. Role of Tet1/3 Genes and Chromatin Remodeling Genes in Cerebellar Circuit Formation. *Neuron* 89, 100–112. doi:10.1016/j.neuron.2015.11.030

FIGURES

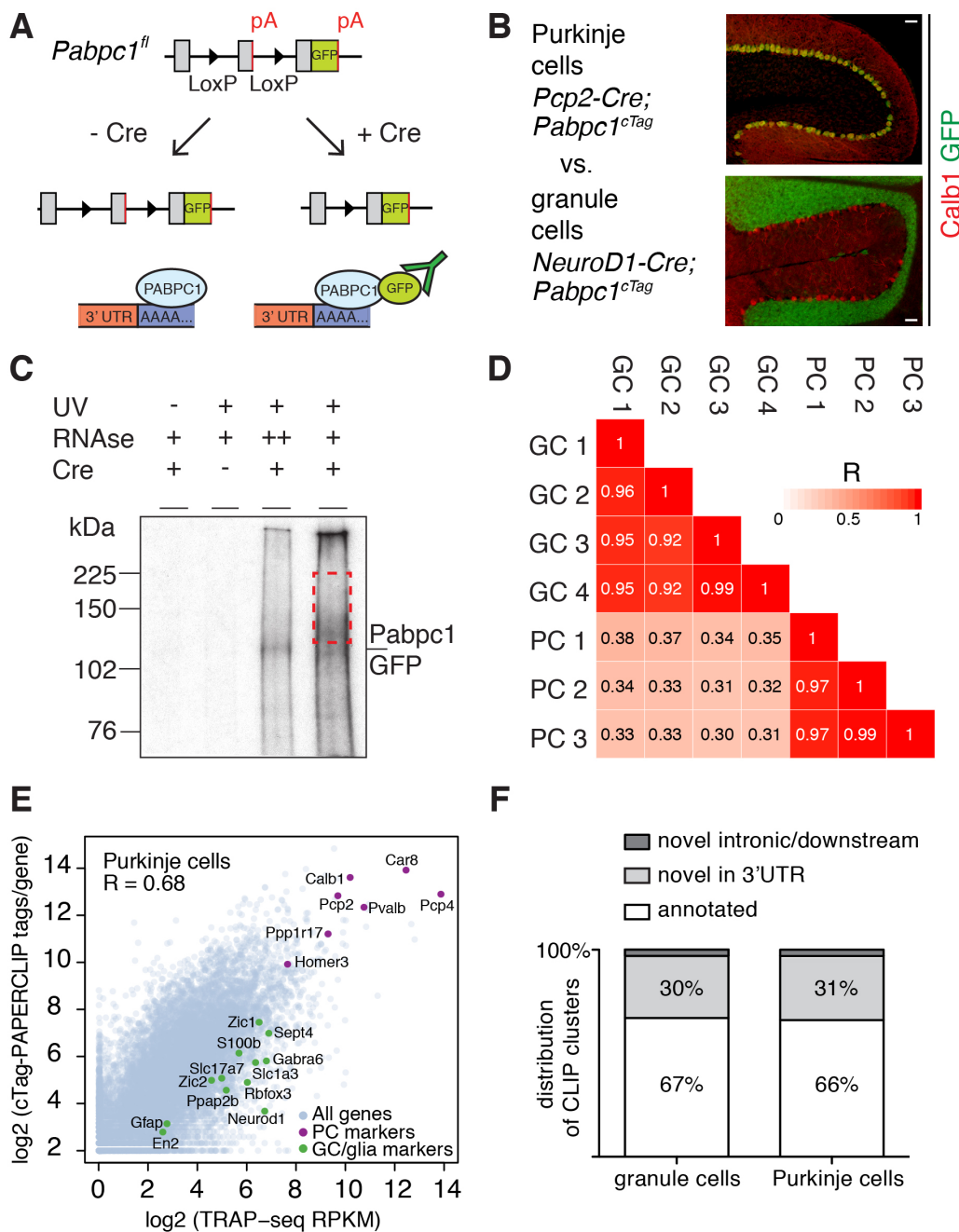


Figure 1. cTag-PAPERCLIP Identifies 3'UTR Isoforms Expressed in Specific Neuronal Types.

(A) Schematic of the cTag-PAPERCLIP approach. Breeding of cTag-PABP mice with Cre-expressing mice restricts expression of PABPC1-GFP to the cells of interest (B) Immunostaining of cerebella from mice expressing PABPC1-GFP in Purkinje cells (top) and granule cells (bottom). Calb1: Purkinje cell marker. Scale bars: 50 μ m. (C) Autoradiogram of radiolabelled RNA cross-linked to PABPC1-GFP purified by

immunoprecipitation from granule cells. Red dashed rectangle shows the area of the membrane from which RNA was eluted and sequenced. **(D)** Correlation between the total number of cTag-PAPERCLIP reads per cluster in four biological replicates from granule cells and three biological replicates from Purkinje cells. R: Pearson correlation coefficient. **(E)** Comparison between total uniquely mapped cTag-PAPERCLIP reads per gene and TRAP-Seq Reads Per Kilobase per Million mapped reads (RPKM) per gene from Purkinje cells. R: Pearson correlation coefficient. Purkinje cell markers are highlighted in red and non-target cell markers are highlighted in green. **(F)** Overlap of cTag-PAPERCLIP clusters that contain reads from three biological replicates from Purkinje cells and at least three biological replicates from granule cells with 3'UTR ends of Ensembl-annotated genes (annotated – clusters that overlap with annotated 3'UTR ends, novel – clusters that do not overlap with annotated 3'UTR ends).

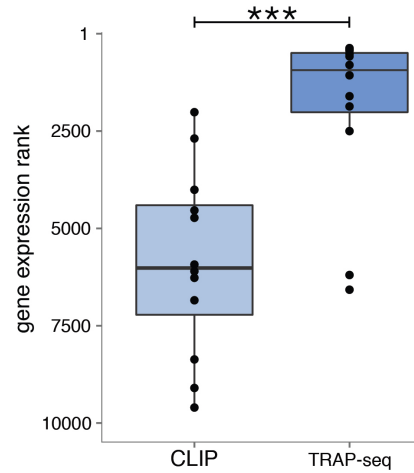


Figure 1 – figure supplement 1. Comparison of marker gene ranks from non-target cell types in cTag-PAPERCLIP data and TRAP-Seq data in Purkinje cells. We used the markers of granule cells and Bergmann glia shown in green in Figure 1E. Mann-Whitney test. P-value < 0.001. The highest expressed gene has a rank of 1.

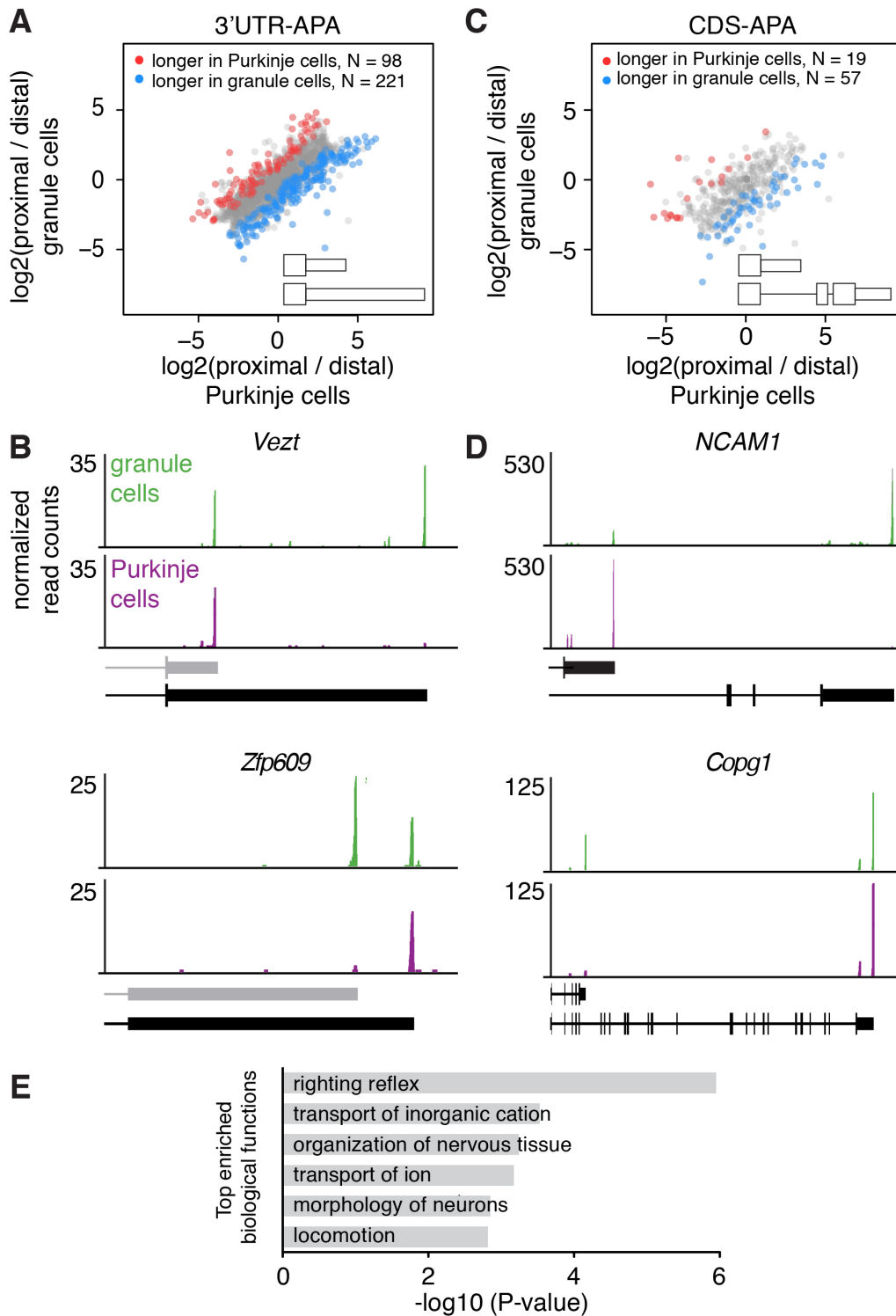


Figure 2. Differences in APA Between Purkinje and Granule Cells.

(A) Scatterplot representing the ratio between the number of cTag-PAPERCLIP reads at the end of the proximal 3'UTR isoform and the number of cTag-PAPERCLIP reads at the end of the distal 3'UTR isoform in Purkinje vs. granule cells. Only genes with two tandem

3'UTR isoforms are shown. Genes showing significantly different ratios ($FDR < 0.05$) are highlighted in red or blue. **(B)** Examples of cTag-PAPERCLIP data for genes that show a large difference in 3'UTR-APA between Purkinje and granule cells. Black bars represent isoforms annotated by Ensembl, gray ones are (predicted) novel isoforms discovered by cTag-PAPERCLIP. **(C)** Scatterplot representing the ratio between the number of cTag-PAPERCLIP reads at the end of the proximal 3'UTR isoform and the number of cTag-PAPERCLIP reads at the end of the distal 3'UTR isoform in Purkinje vs. granule cells. Only genes with two 3'UTR isoforms that differ in CDS-APA are shown. Genes showing significantly different ratios ($FDR < 0.05$) are highlighted in red or blue. **(D)** Examples of cTag-PAPERCLIP data for genes that show a large difference in CDS-APA between Purkinje and granule cells. **(E)** Gene ontology analysis of genes showing significant differences in APA between Purkinje and granule cells. Top 6 relevant functional categories are shown.

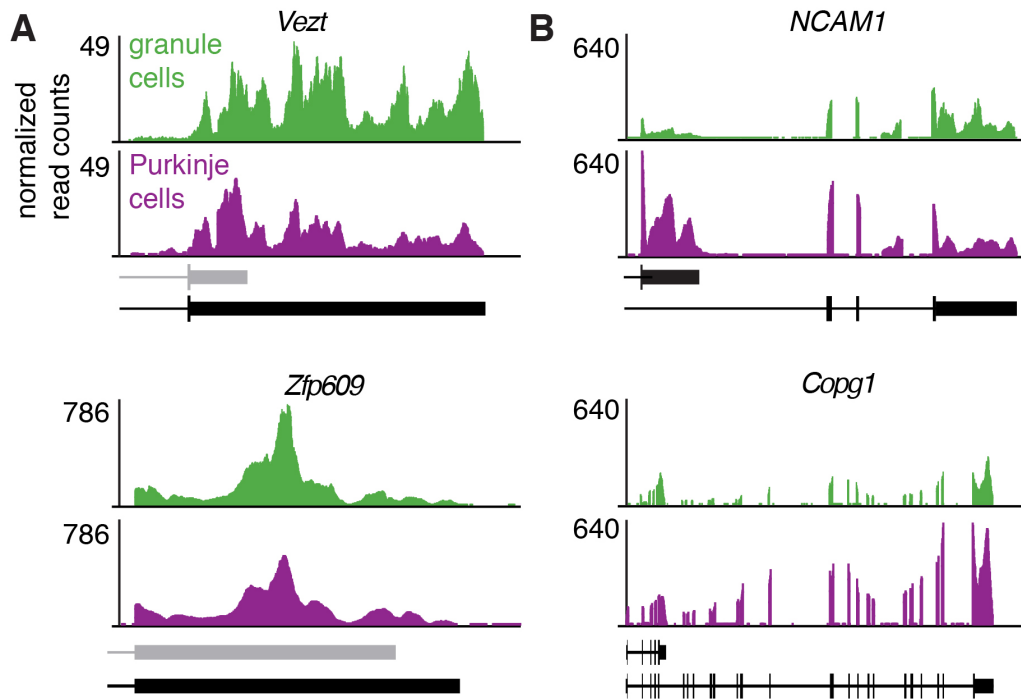


Figure 2 – figure supplement 1. TRAP-Seq data for genes shown in Figure 2B and D. TRAP-Seq data on granule and Purkinje cells from adult mice (Mellén et al., 2012).

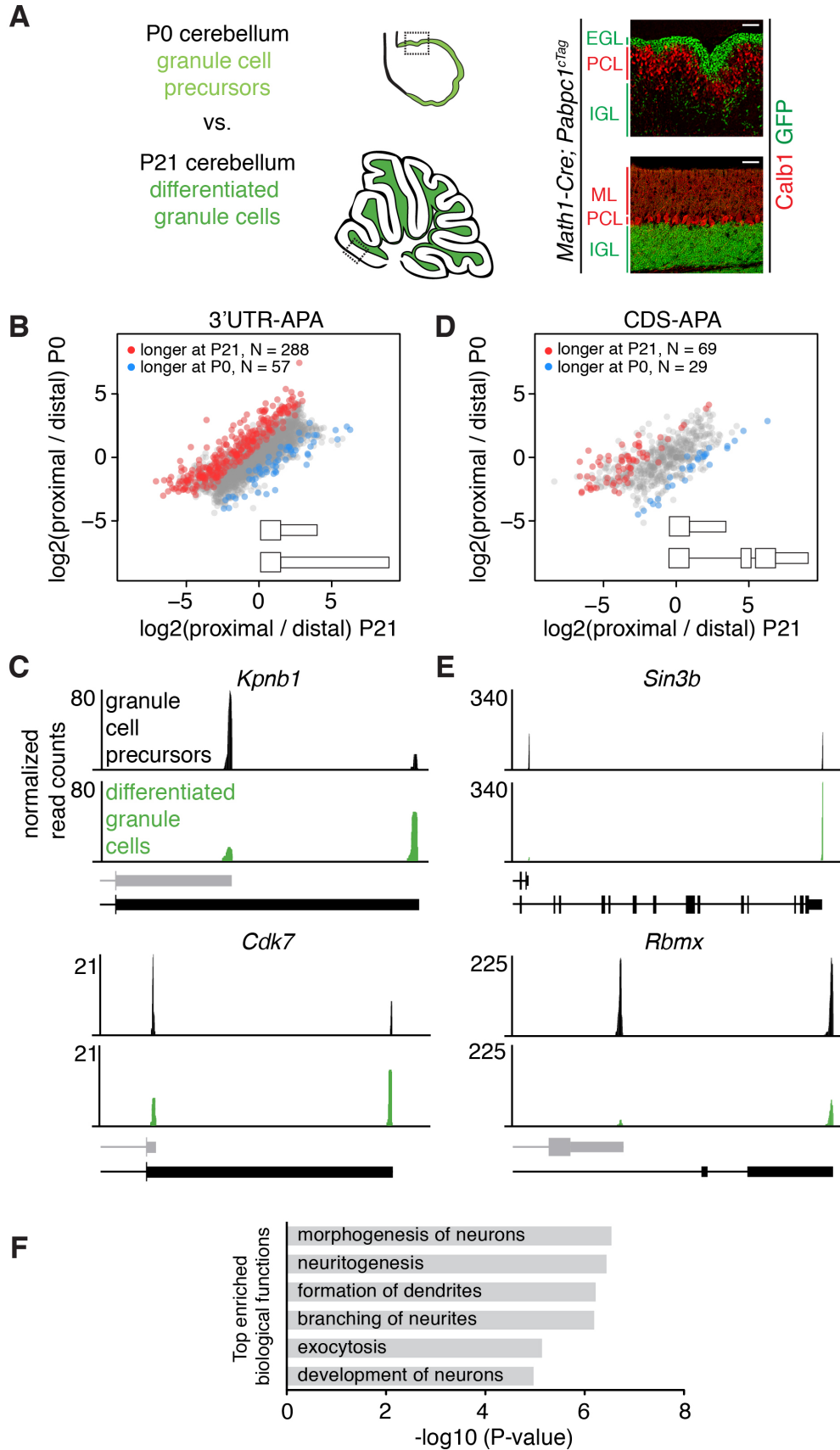


Figure 3. Changes in APA During Granule Cell Development.

(A) Immunostaining showing cell type specific expression of conditionally tagged *Pabpc1* in P0 and P21 granule cells. EGL – external granule layer, PCL – Purkinje cell layer, IGL – internal granule layer, ML – molecular layer. *Calb1*: Purkinje cell marker. Scale bars: 50 μ m. Dotted square on the schematic of cerebellum at P0 and P21 (left side of the panel) shows the location of the immunostaining on the right. (B) Scatterplot representing the ratio between the number of cTag-PAPERCLIP reads at the end of the proximal 3'UTR isoform and the number of cTag-PAPERCLIP reads at the end of the distal 3'UTR isoform in P0 vs. P21 granule cells. Only genes with two tandem 3'UTR isoforms are shown. Genes showing significantly different ratios (FDR < 0.05) are highlighted in red or blue. (C) Examples of cTag-PAPERCLIP data for genes that show a large difference in 3'UTR-APA between P0 and P21 granule cells. Black bars represent isoforms annotated by Ensembl, gray ones are (predicted) novel isoforms discovered by cTag-PAPERCLIP. (D) Scatterplot representing the ratio between the number of cTag-PAPERCLIP reads at the end of the proximal 3'UTR isoform and the number of cTag-PAPERCLIP reads at the end of the distal 3'UTR isoform in P0 vs. P21 granule cells. Only genes with two 3'UTR isoforms that differ in CDS-APA are shown. Genes showing significantly different ratios (FDR < 0.05) are highlighted in red or blue. (E) Examples of cTag-PAPERCLIP data for genes that show a large difference in CDS-APA between P0 and P21 granule cells. Black bars represent isoforms annotated by Ensembl, gray (predicted) isoform of *Rbmx* was discovered by cTag-PAPERCLIP. The CDS portion of the last exon in the predicted *Rbmx* isoform was inferred from an annotated isoform not shown in the picture. (F) Gene ontology analysis of genes showing significant differences in APA between P0 and P21 granule cells. Top 6 relevant functional categories are shown.

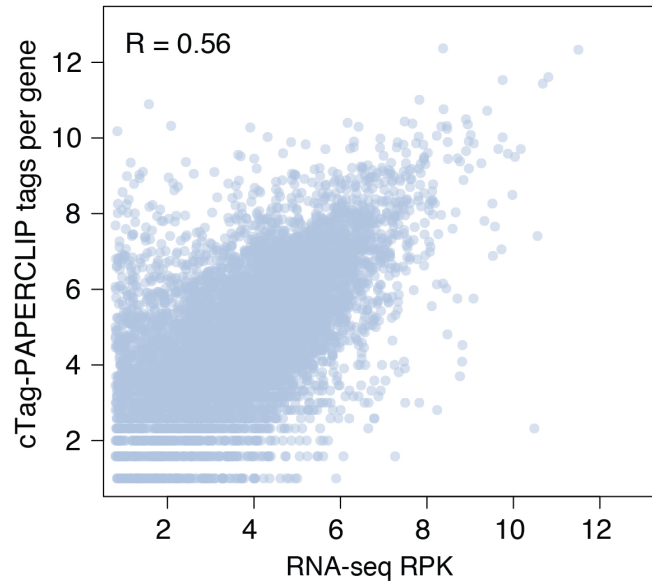


Figure 3 – figure supplement 1. The number of cTag-PAPERCLIP reads per gene is correlated with RNA sequencing reads per kilobase (RPK) per gene.

Comparison between cTag-PAPERCLIP data from granule cell precursors at P0 and RNA-seq data from the same population purified by FACS. R: Pearson correlation coefficient.

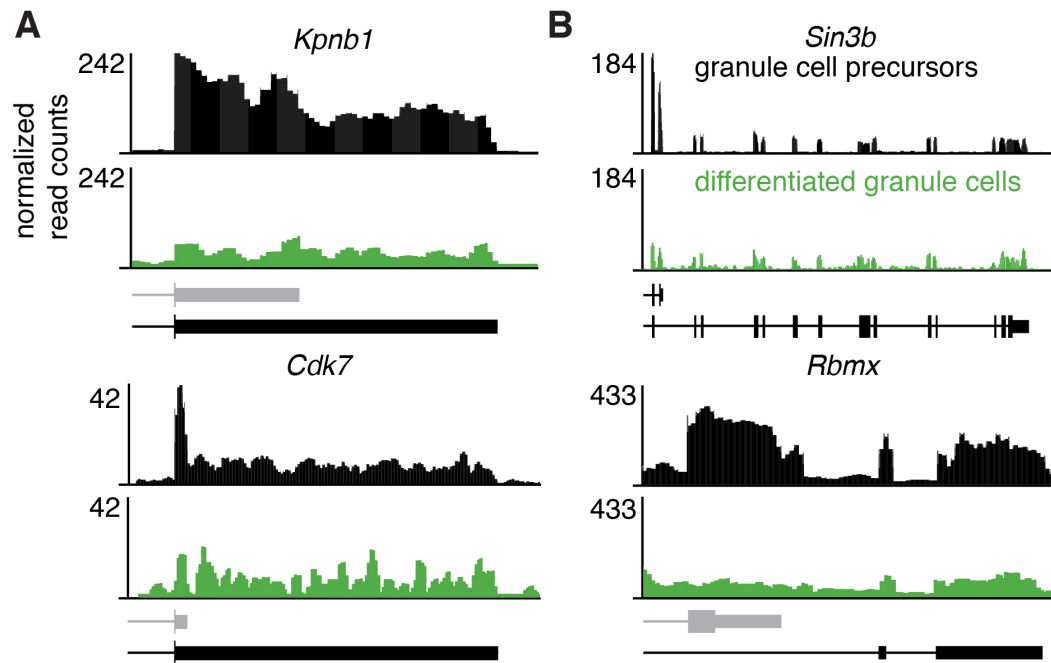


Figure 3 – figure supplement 2. RNA-seq data for genes shown in Figure 3C and E. RNA-Seq data on FACS-sorted granule cell precursors (from P0 mice) and differentiated granule cells (from P21 mice).

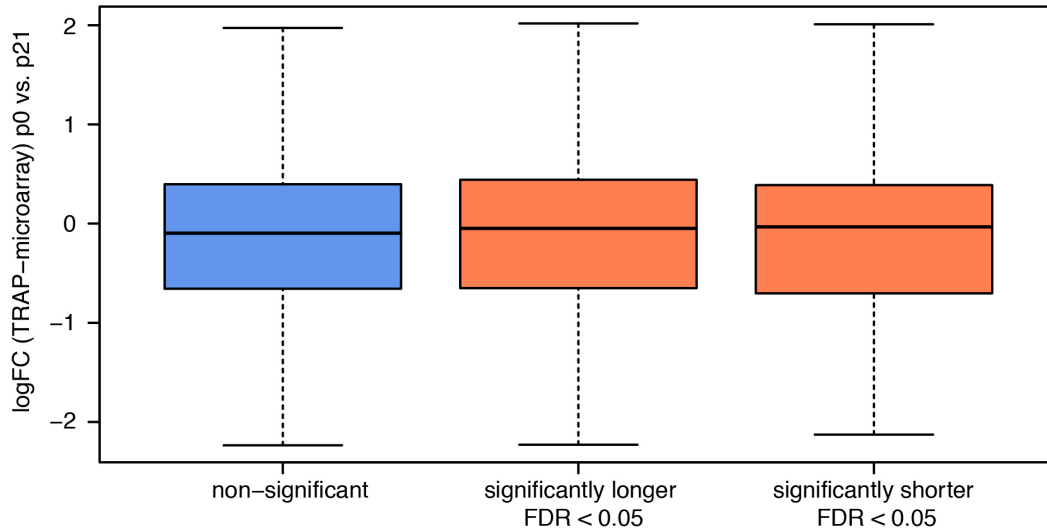


Figure 3 – figure supplement 3. Influence of changes in APA on ribosome-associated mRNA abundance between P0 and P21 in cerebellar granule cells.

Distribution of log₂ fold-changes (logFC) in ribosome-associated mRNA abundance between P0 and P21 for genes with two 3'UTR isoforms that do not show significant changes in APA between the two time points (blue box) and those that do (red boxes). Mann-Whitney test. P-value = NS for both comparisons (non-significant vs. significantly longer and non-significant vs. significantly shorter). LogFC in ribosome-associated mRNA abundance was determined using data from a recently published TRAP microarray study on developing cerebellar granule cells (Zhu et al., 2016).

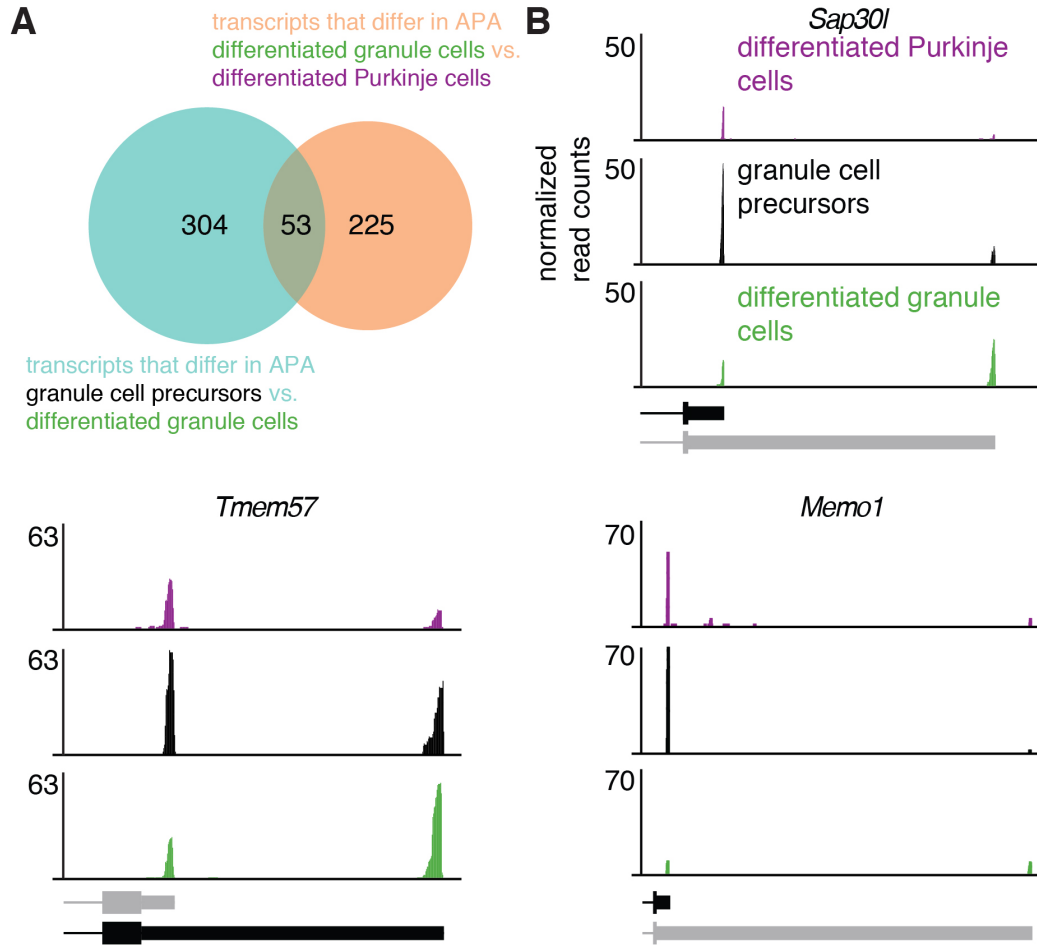


Figure 4. Increase in Expression of Distal 3'UTR Isoforms During Development Is Cell-type Specific

(A) Overlap between genes that exhibit a significant shift towards distal 3'UTR isoform expression during granule cell development (blue circle) and genes that express significantly more of the distal 3'UTR isoforms in granule cells compared to Purkinje cells (orange circle). $p=6.85 \times 10^{-8}$, hypergeometric test. (B) cTag-PAPERCLIP data for three genes that show a significant shift towards distal 3'UTR isoform expression during granule cell development and a difference in 3'UTR isoform expression between Purkinje and granule cells. Black bars represent isoforms annotated by Ensembl, gray ones are (predicted) novel isoforms discovered by cTag-PAPERCLIP.

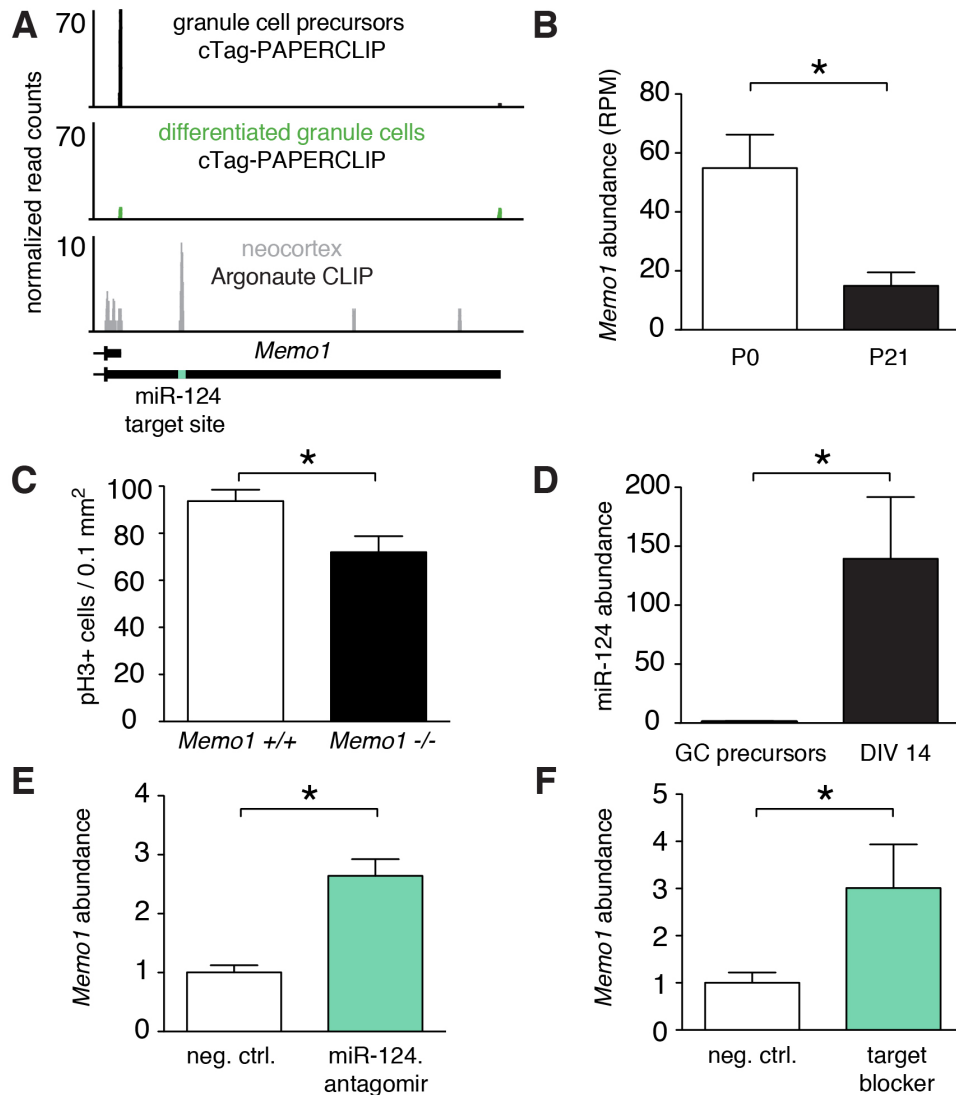


Figure 5. *Memo1* Expression is Developmentally Regulated by miR-124 and APA.

(A) cTag-PAPERCLIP reads on *Memo1* 3'UTR in P0 and P21 granule cells. The location of miR-124 target site is indicated by the green bar. (B) Expression of *Memo1* gene as determined by cTag-PAPERCLIP. The y-axis represents the number of total unique reads per gene normalized to sequencing depth (RPM – reads per million). Two-tailed t-test. P-value < 0.05 Error bars: standard error. (C) Density of phospho-H3 positive (i.e. mitotic) cells in the external granule layer of E18.5 cerebella of wild-type and *Memo1* knockout mice. Two-tailed t-test, P-value < 0.05. Error bars: standard error. (D) Relative abundance of miR-124 in purified granule cell precursors (GC precursors), after 14 days in culture (DIV 14, two-tailed t-test, P-value < 0.05). Error bars: standard error. (E) Relative abundance of long *Memo1* isoform in cultured primary granule cells after treatment with scrambled control (neg. ctrl.) and miR-124 antagomir. Two-tailed t-test. P-value < 0.05. Error bars: standard error. (F) Relative abundance of long *Memo1* isoform in cultured primary granule cells after treatment with scrambled control (neg. ctrl.) and *Memo1* miR-124 target site blocker oligonucleotide (target blocker). t-test. P-value < 0.05. Error bars: standard error.

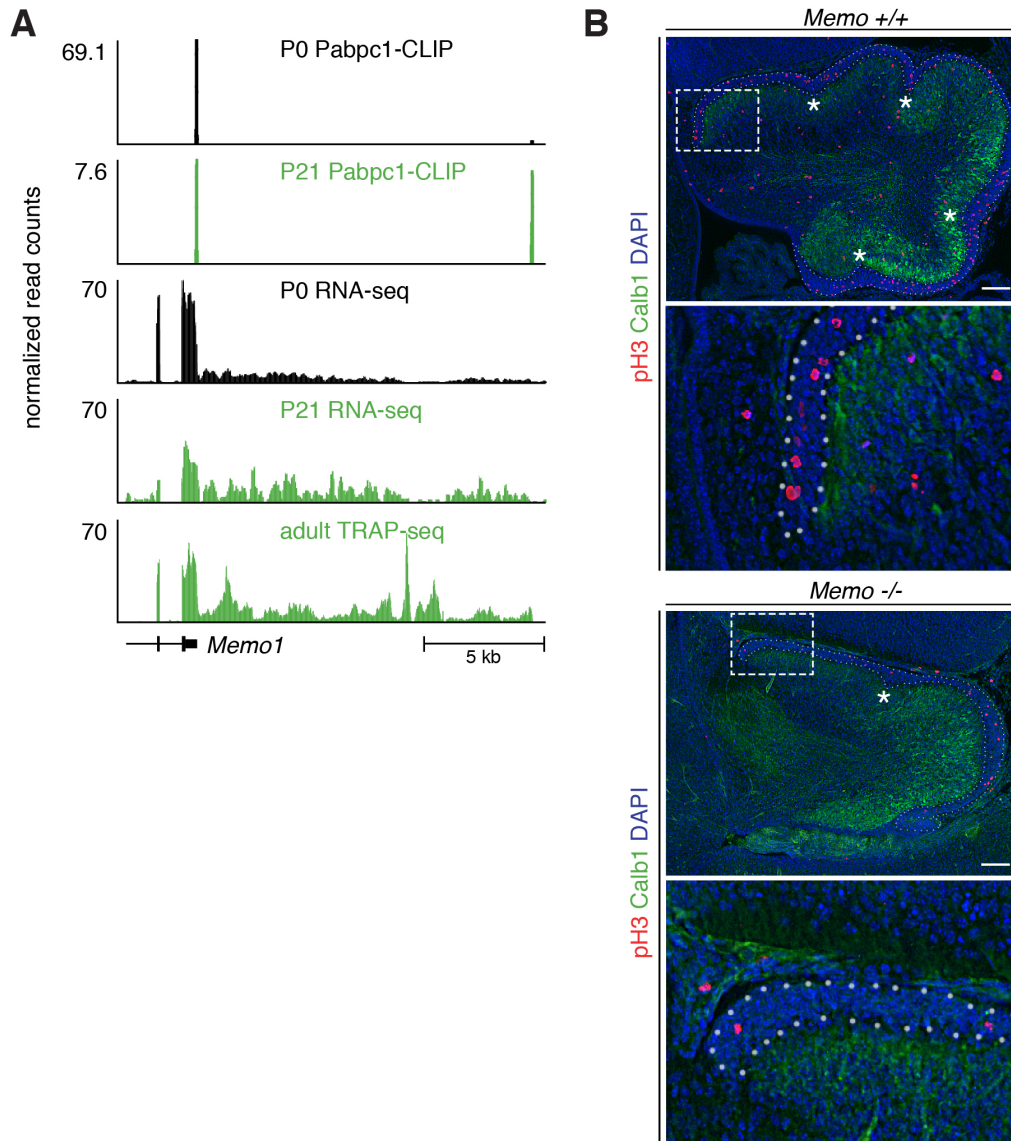


Figure 5 – figure supplement 1. *Memo1* Changes APA During Granule Cell Development and Regulates Granule Cell Precursor Proliferation

(A) Additional data showing *Memo1* APA change during granule cell development. The first two tracks from above represent cTag-PAPERCLIP data from P0 and P21 granule cells. The tracks below cTag-PAPERCLIP represent RNA-sequencing data from FACS sorted P0 and P21 granule cells and TRAP-Seq data from mature granule cells (from Mellén et al., 2012). In the lower left corner, Ensembl annotation for *Memo1* is shown. (B) Immunofluorescence of *Memo1* *+/+* and *-/-* mouse cerebella at E18.5. Purkinje cell layer was stained with anti-Calb1 antibody (to visualize the border between external granule layer and developing Purkinje cell layer) and anti-phospho-H3 antibody was used to stain all cells undergoing mitosis. In the first picture for each genotype, external granule layer is marked with a white dotted line and white asterisks denote developing fissures. The second picture for each genotype is a close-up marked with white rectangle in the first picture. Scale bars: 50 μ m.