Title: Brown rat demography reveals pre-commensal structure in eastern Asia prior to expansion

into Southeast Asia during the Song dynasty

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ABSTRACT

Fossil evidence indicates that the globally-distributed brown rat (*Rattus norvegicus*) originated in

northern China and Mongolia. Historical records report the human-mediated invasion of rats into

Europe in the 1500s, followed by global spread due to European imperialist activity during

the1600s-1800s. We analyzed 12 genomes representing seven previously identified evolutionary

clusters and tested alternative demographic models to infer patterns of range expansion,

divergence times, and changes in effective population (N_e) size. We observed three range

expansions from the ancestral population that produced the eastern China (diverged ~326kya),

Pacific (~42.8kya), and Southeast Asia (~0.68kya) lineages. Our model shows a rapid range

expansion from Southeast Asia into Europe 535 years ago (1480AD). We observed declining N_e

within all brown rat lineages from 100-2kya, reflecting population contractions during glacial

cycles. N_e increased since 1kya in Asian and European, but not Pacific, evolutionary clusters.

Our results support the hypothesis that northern Asia was the ancestral range for brown rats. We

suggest that southward human migration across China between 800-1550s AD, resulted in the

introduction of rats to Southeast Asia, from which they rapidly expanded into the Middle East

then Europe via existing maritime trade routes. Finally, we discovered that North America was

colonized separately on both the Atlantic and Pacific seaboards, yet with evolutionary clusters of

vastly different ages and genomic diversity levels. Our hypotheses should stimulate discussions

among historians and zooarcheologists regarding the relationship between humans and rats.

INTRODUCTION

The genus *Rattus* originated and diversified in eastern and central Asia, and fossil evidence (1)
suggests northern China and Mongolia as the likely ancestral range of the cold-hardy brown rat
(*Rattus norvegicus*). Yet, their contemporary distribution includes every continent except
Antarctica. As a human commensal, brown rats occupy urban and agricultural areas using food,
water, and shelter provided by humans. Rats are one of the most destructive invasive mammals,
as they spread zoonotic diseases to humans (2), damage food supplies and infrastructure (3), and
contribute to the extinction of native wildlife (4). As an invasive species, brown rats outcompete
native species for resources and are a primary target of eradication efforts (5). Brown rats have
been domesticated as models for biomedical research with inbreeding leading to disease
phenotypes similar to humans (6). Finally, they are a nascent model to study evolution within
urban landscapes, as they likely experience multiple selection pressures given their global
distribution in a range of conditions (7).

The historical record indicates colonization of Europe occurred in the early 1500s, eastern North
America by the 1750s, and the Aleutian Archipelago by the 1780s (8, 9). Yet, the timing of
historical records may be inconsistent with population divergence times estimated from genomic
data for two reasons. First, genetic population divergence is not an instantaneous process, thus
sufficient time must pass for allele frequencies to change enough that groups of randomly mating
individuals are recognized as unique populations within demographic models. Second, the
historical record may not immediately record a species within a new geographic area. We would
expect that: population sizes of new species must increase to a noticeable level, species are
identifiable (i.e. brown rats can be mistaken for black rats [*R. rattus*]), and records survived
through time. Thus, historical records provide minimum dates for introductions.

Research into the global expansion of brown rats has focused on both the routes and timings of
different invasions; questions of specific interest include the location of the ancestral range, and
when they arrived in Europe, particularly since black rats reached southern Europe by 6kya (10)
and Great Britain by the 300s AD (11). Previous phylogeographic studies with mitochondria
identified China as the ancestral range based on private haplotypes and ancestral state
reconstructions, with multiple expansions into Southeast Asia, Europe, and North America (12-

14). Inference from mitochondria has been limited due to high haplotype diversity observed from

locally intense but globally diffuse sampling strategies. Thus, key geographic regions, especially

around the Indian Ocean basin and the Middle East, are missing data; sampling these areas

would allow us to distinguish clinal versus long-distance expansions, where multiple

introductions occurred, and mito-nuclear discordance. A phylogeographic analysis using nuclear

genomic data inferred hierarchical clustering along five range expansion routes (15). From the

putative ancestral range, brown rats expanded southward into Southeast Asia and eastward into

China and Russia (15). The eastward expansion extended to North America with two

independent colonizations of the Aleutian Archipelago and sites along the Pacific coast of

western North America. From Southeast Asia rats expanded into Europe (15, 16). Given

historical records of brown rats in Europe in the 1500s, the likely route was aboard ships

conducting maritime trade across the Indian Ocean into the Red Sea and Persian Gulf before

moving goods onto land. Although these routes were established by the 200s BC, they intensified

in the 1400-1500s AD (17). The fifth range expansion moved rats to eastern North America, the

Caribbean, South America, western Africa, and Australasia during the age of European

imperialism of the 1600-1800s (15). Ultimately, we inferred the following seven genomic

clusters: *Eastern China*, *SE Asia*, *Aleutian*, *Western North America*, *Northern Europe*, *Western*

Europe, and *(Western Europe) Expansion*.

Demographic histories of domesticated and commensal species are uniquely compelling as they

infer both the species history and how humans interacted with each species over time.

Demographies can infer domestication history (18, 19), population declines and recoveries (20,

21), human mediated range expansions (22), and factors influencing extinction (23). Although

the demographic history of humans has been extensively researched, inference from domestics

and commensal species can elucidate patterns of human expansion and exploration (24). Thus,

we tested alternative demographic models to identify patterns of population divergence, infer the

timing of range expansions, and estimate effective population size (N_e) changes through time.

From the geographic and temporal patterns, we were able to distinguish natural versus human-

mediated movement of brown rats.

RESULTS

We sequenced two genomes each from *SE Asia*, *Northern Europe*, *Western Europe*, and the *Western Europe-Expansion* (hereafter- *Expansion*) evolutionary clusters, and one genome each from the *Aleutian* and *Western North America* clusters (NCBI SRA PRJNA344413; Table S1). Average sequencing depth was 28. 2X (range 24-38). We estimated heterozygosity for each individual on the 20 autosomes separately. Samples from *Eastern China* had the highest average chromosomal heterozygosity (0.244) where the *Aleutians* and *Western North America* had the lowest heterozygosity (0.143 and 0.148, respectively; Figure 1B).

Ancestral Gene Flow

We used the ABBA-BABA test to examine historic gene flow between evolutionary clusters and verify the global tree topology by comparing all combinations (n=106) of population tree topologies $D(H_1, H_2, H_3, R. rattus)$. We observed 97 significant *D*-scores with absolute Z-scores greater than 3 (Table S3), where most patterns were congruent with our inferred population tree (see below, Figure 2A). However, both the *Aleutian* and *Western North America* clusters shared more derived alleles with the *Expansion* cluster than either Asian or other European clusters (Table S3), suggestive of movement of rats across North America. Further, population trees of the form *D*(Asian or European Population, Western North America, Aleutian, *R. rattus*) consistently observed increased derived alleles within the Asian or European clusters than the Pacific clusters, thus a sister relationship between *Western North America* and *Aleutian* was not observed (Table S3). This result was unexpected, as we demonstrate here and in a previous analysis (15) that the two Pacific clusters share a population ancestor; unsampled diversity may have influenced these results.

WGS Demographic model

We conducted model selection by first identifying the model that best represented divergence patterns in Asia and Europe separately, then combined those tree topologies into a global model for parameter estimation. The best Asian model had an ancestral unsampled population with independent divergence events for *Eastern China*, *SE Asia*, and the Pacific cluster that diverged into the *Aleutians* and *Western North America* (Figure S1). For the European model, the best supported model included an unsampled population (likely in the Middle East) diverging from

SE Asia, where *Western Europe* diverged from the unsampled population, and *Northern Europe*

and the *Expansion* independently diverged from *Western Europe* (Figure S2).

Using WGS data from 12 genomes representing seven evolutionary clusters, we modeled the

nine-population topology inferred from the sub-models (Figure 2, Table S4). We estimated that

Eastern China diverged from the ancestral population 326kya (90% highest density probably

[HPD]: 308-327kya). The Pacific cluster diverged from the ancestral population 42.8kya (HPD:

30.0-41.3kya), then the *Aleutians* and *Western North America* diverged 20.2kya (HPD: 10.7-

20.9kya). The divergence that led to the global expansion of rats occurred rapidly, where rats

first expanded into *SE Asia* 682 years ago (1333AD; HPD: 997-1406AD), followed by the

introduction of rats into the unsampled population 681 generations (HPD: 1001-1409AD). We

estimated rapid divergence of rats into Europe including the *Western Europe* divergence

535years ago (1480 AD; HPD: 1462-1511AD), and *Northern Europe* divergence 534 years ago

(1481 AD; HPD: 1464-1512AD). Finally, we estimated the *Expansion* cluster diverged 527 years

ago (1488 AD; HPD: 1487-1524AD).

We ran the cross-coalescence analysis within MSMC2 to estimate the rate of divergence between

the seven clusters. We observed that divergence was complete between both the *Aleutians* or

Western North America and all other populations (Figure S3). The European clusters showed

similar patterns of divergence with *Eastern China* with approximately 60% divergence complete

(Figure S3A). Cross-coalescence between the *Aleutians* and *Western North America* increased

approximately 200 generations ago before decreasing to 50% (Figure S3C). The four clusters

making up the most recent expansions (*SE Asia*, *Northern Europe*, *Western Europe*, and

Expansion) had signatures of increasing cross-coalescence over the past 1,000 generations

(Figure S3 B, D-F). We believe this result was due to a population version of incomplete lineage

sorting, where the rapid global range expansion resulted in high rates of coalescence events

between instead of within evolutionary clusters.

ddRAD Demographic Models

We used a ddRAD-Seq dataset (Table S2) to further investigate the within-cluster population

tree topologies. Given the hypothesis that the ancestral populations for western North America

were in eastern Asia, we first compared these clusters and observed that eastern China and

eastern Russia were sister populations (Figure S5). Further, our modeling supported an

admixture pulse of 30% 215 generations ago from eastern Russia into rats on Adak Island within

the Aleutian Archipelago (Figure S5). Next, we modeled relationships between four populations

within Southeast Asia and observed that rats first expanded into central Southeast Asia. There

were two independent expansions eastward: one into Cambodia and Vietnam which formed

sister populations, and a second to the Philippines (Figure S5).

We split European populations between the *Western* and *Northern* evolutionary clusters. The

best supported topology for *Northern Europe* had the continental Netherlands diverging from a

sister group including Norway and Sweden (Figure S6). In *Western Europe*, we observed a split

between continental Europe (populations in France and Spain) and the British Isles (Figure S7).

The Pacific coast of North America contains populations with *Aleutian*, *Western North America*,

and *Expansion* genomic signatures (15). To better understand admixed rat populations we

modeled two North American populations (Vancouver, Canada and Berkeley, USA). We

observed that both populations were best described by admixture between the *Western North*

America and *Expansion* clusters (Figure S8).

Effective population size through time

We inferred the change in N_e over time using the multiple sequentially Markovian coalescent

(MSMC) model (25) and scaled the estimates to years and N_e using the estimated mutation rate

(μ) from the coalescent modeling analysis of 9.90×10^{-8} and 3 generations per year. As Deinum *et*

al. (26) estimated μ of 2.96×10^{-9} and the precise generation time for rats is unknown, we

present alternative estimates of the MSMC model in Figure S9. We observed two distinct

patterns; first, the *Aleutian* and *Western North America* clusters declined sharply in N_e

approximately 50kya (Figure 1A); this was consistent with the estimated divergence of the

Pacific clusters at 42.8kya. Following divergence, N_e steadily declined until 500 years ago when

it stabilized to approximately 1,400 and 1,500 effective individuals in *Aleutian* and *Western*

North America, respectively (Figure 1A).

The second pattern was concordant between the *Eastern China*, *SE Asia*, *Northern Europe*,

Western Europe, and *Expansion* clusters. Similar to the Pacific clusters, N_e steadily declined

from approximately 100 – 1kya before increasing in the most recent time periods (Figure 1A).

Approximately 200 years ago (the first reliable time step), N_e was: 12,000 in *Eastern China*,

36,000 in *SE Asia*, 45,000 in *Northern Europe*, 28,000 in *Western Europe*, and 25,000 in

Expansion (Figure 1A).

DISCUSSION

Our demographic model showed that from an unsampled ancestral population, brown rats

diverged into three genomic clusters that we sampled in eastern China, two locations in North

America, and Southeast Asia. These results were congruent with a hypothesized ancestral range

in northern China and Mongolia where brown rat fossils were identified (1). Divergence of

brown rats from their sister species *R. nitidus* was estimated at 633kya (27). We estimated

divergence of the *Eastern China* cluster at 326kya (Figure 2A, Table S4), which would be

consistent with the establishment of ancient population structure within glacial refugia near the

end of the interglacial period of the Middle Pleistocene (28). This estimate contrasts with the

observed mitogenome diversity, where the five clades that occur within the samples genotyped

for the *Eastern China* nuclear cluster diverged since 96kya (HPD: 70-128kya) (13). We

previously hypothesized that the incongruence between a single nuclear cluster and multiple

deeply diverged mitochondrial clades represented ancestral population structure and later

admixture. The discrepancies in the divergence timing estimation between the nuclear (326kya;

307-327kya) and mitochondrial (96kya; 70-128kya) genomes could be do either to extirpation or

incomplete sampling of mitogenome diversity. Alternatively, our coalescence modeling may

have overestimated divergence age between *Eastern China* and the ancestral range, which was

suggested by the upper limit of the MSMC analysis (Figure 1A) at 100kya.

We observed that the Pacific cluster diverged from the ancestral population 42.8kya (Figure 2A,

Table S4), where this result was supported by our MSMC analysis (Figure 1A). Divergence of

the *Aleutian* and *Western North America* clusters occurred 20.2kya, although MSMC did not

identify this signal. The similarity in N_e over time between these two clusters may have been due

to either true recent divergence as suggested by cross-coalescence (Figure S4C) or similar

climatic conditions through the Pleistocene when population size decreased for many vertebrates.

We emphasize that the divergence of these clusters does not identify the timing of the

introduction to the Aleutian Archipelago or the Pacific coast of North America where samples

were collected. The historic record indicates rats were moved to the Aleutian Archipelago by

Russian fur traders in the 1780s (9). Our regional population model suggested a scenario with

gene flow from a population in eastern Russia into Adak Island (Figure S4). We modeled this

scenario as an admixture pulse from eastern Russia around 1943, suggesting two introductions of

rats onto Adak. Our results suggest that rats from Japan and sites around the Bering Sea in

Russia and the USA may be a dynamic region for rat population genomics. Specifically by

incorporating both more sites and ancient samples, demographic models could estimate

divergence times to differentiate between models of contemporary or historic movement of rats,

particularly as human divergence into the region occurred approximately 36kya (29).

We estimated that the *SE Asia* cluster diverged from the ancestral population 0.68kya (1333 AD,

Figure 2A, Table S4). The timing of this divergence immediately raises the question of why rats

did not expand sooner as overland trade between China and Southeast Asia was established by

the 500s AD (30); and maritime trade between these regions and the Indian Ocean basin was

established before the 900s AD (31). A partial explanation may be due to the intersection of

climate and human demography across eastern Asia. The Medieval Climate Anomaly (850-1250

AD) aided agricultural expansion and human demographic growth in China, specifically

expanding urban centers outwards at a time of human movement from northern arid lands to

more agriculturally productive lands in the south (see references within 30). However, the end of

this climatic period resulted in drought, famine, political instability, and ultimately human

demographic contractions in both China and Southeast Asia; fortunes reversed in the late 1400s

to mid-1500s as the climate improved and populations expanded again (30). We can view rats'

southward expansion from this historical context. As we lack samples between the ancestral

range and Southeast Asia, we were unable to determine population divergence along this axis

and intermediate time points, although Zeng *et al*. (16) observed a clinal pattern of ancestry in

southern China. Thus, the founding of new agrarian communities and increasing inter-

connectedness with urban centers would serve as stepping-stones moving rats from northern

China to Southeast Asia during the two periods of human demographic expansion.

Our results stand in marked contrast to a similar analysis that identified brown rats in Southeast

Asia as the ancestral population with a northward expansion to colonize eastern Asia (16). Both

analyses used similar modeling approaches but with different datasets. We suggest that our

results differ due to the following two factors: first, we included unsampled populations that

allowed for coalescence within that group instead of other contemporary lineages. Inclusion of

unsampled populations influences parameter estimates of N_e and migration rates and has been

shown to improve or at least not harm parameter estimation within the full model (32). Adding

unsampled populations to our model was important given the limited number of chromosomes

genotyped, as large sample sizes decrease the effect of unsampled populations on parameter

estimates (33). Second, Zeng *et al*. included admixed samples (that were geographically located

in southern China) between the SE Asia and northern Asia (our *Eastern China*) genomic clusters

where the admixed samples were exclusively grouped with the northern Asia genomic cluster.

By grouping the mixed ancestry samples preferentially with the northern Asia cluster, the

coalescent model was better supported when the SE Asia population was ancestral. This was

likely do to better model fit given that a small proportion of SE Asian alleles were within the

northern Asia cluster. We believe this was not due to the true history of the populations but

instead to artefacts introduced by inappropriate sample clustering. The cline in ancestry

proportions from the two divergent groups may be from a southern expansion from the north

and/or a northern expansion from SE Asia, thereby creating a regional admixture zone in

southern China where samples were collected (16).

Finally, we show that from Southeast Asia there was a rapid range expansion into Europe

approximately 1480 AD (Table S4); here our phylogeographic pattern but not divergence time

results were congruent with those of Zeng and colleagues (16). While our coalescent models

support the population topology of divergence into *Western Europe* then into *Northern Europe*

(Figure S3), it is important to note that parameter estimates of divergence between those clusters

were one generation apart. Thus, the range expansion into Europe occurred so rapidly that

precise topology patterns and divergence times may be difficult to recover. Further, potential

stepping-stone areas in India, the Middle East, and coastal eastern Africa have been sparsely

sampled; although samples from Iran cluster with *Western Europe* with small ancestry

proportions from *Northern Europe* and *SE Asia* (16). This pattern was consistent with the rapid

range expansion into Europe we observed, thus not allowing sufficient time for drift to establish

unique allele frequency patterns. We estimated that the *Expansion* cluster diverged from *Western*

Europe approximately 1488 AD (HPD: 1464-1512 AD; Table S4). While European seafaring to

North America occurred during this time, this particular estimate may be too young to represent

the actual introduction into New York, USA and instead may represent the timing of divergence

of the ancestral population likely within Great Britain.

Ancestral population size

We observed that each evolutionary cluster declined in N_e since 100kya (Figure 1A), consistent

with many studies that observed population declines during the Pleistocene. The decline lasted

longer for the two Pacific clusters, consistent with the lower heterozygosity estimates (Figure

1B). We observed an increasing trend in the most recent time segments for the two Asian and

three European clusters. This result was consistent with our demographic models showing a

rapid and recent global range expansion.

Range expansion via human-mediated movements

Our results identified that the global range expansion of rats occurred recently (1480s AD) and

rapidly from Southeast Asia into Europe. This stands in marked contrast to previous assumptions

that brown rats were transported westward along the Silk Road through central Asia into Europe.

This is counterintuitive as overland trade routes from central China to Persia were established

2.1kya (105 BC) and goods reached Rome by 46 BC (17). Brown rats are native to the

geographic region through which part of the Silk Road connected, unlike black rats which

speciated on the Indian subcontinent (34). Assuming that rats evolved their commensal

relationship with humans prior to their global range expansion, as observed with house mouse

(35), the availability of cities, road networks, and a flow of merchants naturally suggests a way

to expand westward. The Silk Road may have not been the route for expansion due to the limited

distance that merchants traveled along the route, as goods went further than caravans (17).

Further, high aridity and lack of water sources may have limited rat movement through this

region. Yet this does not preclude the idea that brown rats may have expanded westward via Silk

Road cities and were then extirpated due to the collapse of those cities during changing geo-

politics and shifts towards maritime trade (17). We instead suggest that pulses of southward

human demographic expansion from northern China during favorable climatic conditions

enabled the expansion of rats into Southeast Asia from which they expanded into Europe. We

present this historical narrative as a hypothesis supported by our demographic model, but also to

stimulate interest in further study by historians and zooarchaeologists to examine the historical

expansion of this globally important invader.

MATERIALS and METHODS

Whole genome sequencing and datasets

We selected ten individuals for whole genome sequencing: two each representing evolutionary

clusters within *SE Asia* (Philippines and Cambodia), *Northern Europe* (Sweden and

Netherlands), *Western Europe* (England and France), and *Expansion* (New York, USA), and one

sample each from the *Aleutian Islands* and *Western North America* (Table S1). We generated

paired-end reads for each sample (4ng RNase treated genomic DNA) by sequencing on an

Illumina HiSeq 2500 at the New York Genome Center. We also downloaded whole genomes

from 11 brown rats and one black rat (*R. rattus*) collected in Harbin, China (ENA ERP001276),

although to not bias estimates with unequal sample sizes we ran analyses using only Rnor13 and

Rnor14 (26) which were randomly selected. All genomes were mapped to the Rnor_5.0.75

reference (36) using BWA-MEM v0.7.8 (37). We marked duplicates using Picard Tools v1.122

and realigned indels using GATK v3.4.0 (38). Data for 10 genomes are available on the NCBI

SRA BioProject PRJNA344413 (13).

Instead of calling variants, we used the genotype likelihoods implemented in ANGSD v0. 915

(39), using a minimum base quality score of 20, minimum mapQ score of 30, then identified

sites genotyped in all 12 genomes on the 20 autosomes. We estimated genotype likelihoods using

the function implemented in SAMTOOLS (-GL 1) (40). The *R. rattus* individual from China

served as the outgroup allowing for identification of ancestral and derived alleles.

Chromosomal Diversity

We estimated heterozygosity on each chromosome for each individual. We exported the

genotype likelihoods from ANGSD into PLINK v1.9 (41, 42) and estimated heterozygosity (--

het) on each chromosome.

Historic gene flow

The ABBA-BABA test (43) measures the proportion of "ABBA" and "BABA" patterns of

ancestral and derived alleles in a four population tree, where an excess of one pattern indicates

gene flow from the third population. To test for ancient admixture, we ran all possible ABBA-

BABA tests (n = 106) in ANGSD using the multipop version of the analysis that allowed

individuals from the same evolutionary cluster to be analyzed together. We used the black rat as

the outgroup for the test $D(H_1, H_2, H_3, R. rattus)$. We calculated D-statistics in program R (44)

using the estAvgError.R file provided with ANGSD.

WGS Demographic Modeling

We inferred the demographic history of rats by modeling alternative scenarios that compared the

observed and expected site frequency spectra (SFS) for multiple individuals from different global

locations. We estimated the SFS for each evolutionary cluster, then output a joint SFS for each

pair of clusters.

Given the large number of evolutionary clusters to model, we first modeled the relationship

between *Eastern China*, *SE Asia*, *Aleutian*, and *Western North America* by comparing five four-

population models and five five-population models that included an unsampled population

(Figure S1). The best supported scenario (Model 6 in Figure S1) had a topology that included an

ancestral unsampled ghost population with independent divergence of *Eastern China*, *SE Asia*,

and the Pacific clusters. We then modeled four populations of a five-tree topology between an

unsampled population, *SE Asia*, *Northern Europe*, *Western Europe*, and the *Expansion*. Our

previous work on brown rat phylogeography suggested that rats expanded into Europe from SE

Asia (15); thus, we tested the topology between the three European clusters arising from an

unsampled population (likely in the Middle East) which was founded from SE Asia. The best

supported scenario (Figure S2) had an initial divergence of *Western Europe* from the unsampled

population, with *Northern Europe* and the *Expansion* diverging independently. For all testing

models, we did not allow population size to change through time, and we set the mutation rate at

2.5×10^{-8} mutations per generation.

We combined the best supported topologies from both sub-models into a nine-population model.

We ran this model both with and without independent growth rates on each cluster, and log

likelihoods were higher for the model without changes in growth parameters (although that may

have been due to the reduction in parameter space), thus we report that model. Unlike in the sub-

models described above, we allowed the mutation rate parameter to be estimated with the model.

We used three generations per year to convert parameter estimates; all time calculations were

done since 2015.

We ran 50 iterations of the nine-population model in fastsimcoal2, then identified the iteration

with the highest estimated likelihood. Using these point estimates, we generated 500 samples of

pairwise SFS each containing 30,000 markers that served as pseudo-observed data for estimating

parameters ranges under the best supported model. We observed nine outlier iterations based on

the difference between the maximum observed and estimated likelihoods in fastsimcoal2; thus,

we removed those samples before calculating the 90% highest probability density (HPD) using

the *HDInterval* v0.1.3 package (45) in R.

ddRAD-Seq Demographic Modeling

While our WGS had many more loci, there was limited geographic representation, as well as

fewer individuals sampled; therefore, we built regional models from double digest restriction

enzyme associated DNA sequencing (ddRAD-Seq; NCBI SRA BioProject PRJNA344413)) data

to explore additional population tree topologies. We estimated the SFS of each genomic cluster

or population of interest in ANGSD using the reference aligned Illumina reads instead of the

previously called SNPs.

We built regional models within the evolutionary clusters for eastern Asia/Pacific, SE Asia,

Northern Europe, and Western Europe. We used this reductive approach to limit the number of

parameters being estimated. Within each region, we compared topologies between populations

suggested by previous population structure analyses (15). We used the same fastsimcoal2 run

parameters as described above; however, we did not create pseudo-observed datasets for

parameter estimation, unless noted, as our interest was in topology. A secondary reason we did

not further explore population parameters within the regions was that we observed these datasets

tended to overestimate divergence times, likely due to unsorted variation remaining within

populations until coalescence with the unsampled ancestral population. Finally, we investigated

population topology and admixture proportions in Vancouver, Canada and Berkeley, USA since

each site was identified as admixed in our previous analysis (Table S2).

Effective population size through time

We estimated the change in effective population size over time in each evolutionary cluster using

MSMC2 (25). To call variants, we used SAMTOOLS v1.3 (40) mpileup across all samples with

a minimum mapping quality of 18 and the coefficient to downgrade mapping qualities for

excessive mismatches at 50. We then utilized the variant calling in BCFTOOLS v1.3 with the

consensus caller and excluded indels which limited the dataset to bi-allelic SNPs. As there was

not a brown rat reference panel, we phased the 12 individuals plus two inbred lines (SS/Jr and

WKY/NHsd; NCBI SRA accessions ERR224465 and ERR224470, respectively (6)) for each of

the 20 autosomes using fastPHASE v1.4.8 (46).

We estimated change in N_e over time for each of the seven evolutionary clusters using two

haplotypes for the *Aleutian* and *Western North American* clusters and four haplotypes for each

other cluster. We also estimated the proportion of population divergence over time using the

cross-population analysis, and combined results from individual populations with the cross-

population analysis using the combineCrossCoal.py script provided.

DATA ACCESSIBILITY

Data for whole genome sequences from 10 brown rats available on NCBI SRA BioProject

PRJNA344413.

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Washington, and the Museum of Texas Tech University also graciously provided tissue samples.

FIGURE LEGENDS

Figure 1- (A) Plots of change in N_e over time using MSMC where the x-axis is years before the present. (B) Heterozygosity measured on each chromosome (n=20) for each of 12 brown rat samples. A box and whisker plot of mean chromosomal heterozygosity was overlaid the point estimates where colors indicate genomic cluster assignment. Additional details of sample locations available in Table S1. Each evolutionary cluster was represented by a different color: *eastern China-* dark brown; *SE Asia-* light brown; *Aleutian-* orange; *Western North America-* yellow; *Northern Europe-* purple; *Western Europe-* light blue; and *Expansion-* medium blue.

Figure 2- (A) The best supported demographic model contained nine evolutionary clusters inclusive of two unsampled populations. The divergence times in generations and N_e are listed in Table S4. (B) Map of global sampling locations including WGS (plus sign, +) and ddRAD-Seq (circles) samples, where locations utilizing both datasets had a plus within a circle. Evolutionary clusters were represented different colors: *Eastern China-* dark brown; *SE Asia-* light brown; *Aleutian-* orange; *Western North America-* yellow; *Northern Europe-* purple; *Western Europe-* light blue; and *Expansion-* medium blue.

REFERENCES

1. Smith AT & Xie Y (2008) *A Guide to the Mammals of China* (Princeton University Press, Princeton, NJ) p 544.
2. Himsworth CG, Parsons KL, Jardine C, & Patrick DM (2013) Rats, cities, people, and pathogens: a systematic review and narrative synthesis of literature regarding the ecology of rat-associated zoonoses in urban centers. *Vector borne and zoonotic diseases (Larchmont, N.Y.)* 13(6):349-359.
3. Pimentel D, Lach L, Zuniga R, & Morrison D (2000) Environmental and Economic Costs of Nonindigenous Species in the United States. *BioScience* 50(1):53-65.
4. Harper GA & Bunbury N (2015) Invasive rats on tropical islands: Their population biology and impacts on native species. *Global Ecology and Conservation* 3(Supplement C):607-627.
5. Jones HP*, et al.* (2016) Invasive mammal eradication on islands results in substantial conservation gains. *Proceedings of the National Academy of Sciences* 113(15):4033-4038.
6. Atanur Santosh S*, et al.* (2013) Genome Sequencing Reveals Loci under Artificial Selection that Underlie Disease Phenotypes in the Laboratory Rat. *Cell* 154(3):691-703.
7. Johnson MTJ & Munshi-South J (2017) Evolution of life in urban environments. *Science* 358(6363).
8. Armitage P (1993) Commensal rats in the New World, 1492-1992. *Biologist* 40:174-178.
9. Black L (1983) Record of maritime disasters in Russian America, Part One: 1741-1799. *Proceedings of the Alaska Maritime Archaeology Workshop, May 17-19, 1983*, (University of Alaska- Fairbanks).
10. Ervynck A (2002) Sedentism or urbanism? On the origin of the commensal black rat (*Rattus rattus*). *Bones and the man: Studies in honour of Don Brothwell*, eds Dobney K & O'Connor T (Oxbow Books, Oxford), pp 95-109.
11. Yalden DW (2003) Mammals in Britain - A historical perspective. *British Wildlife* 14(4):243-251.
12. Song Y, Lan Z, & Kohn MH (2014) Mitochondrial DNA phylogeography of the Norway rat. *PLoS ONE* 9(2):e88425.
13. Puckett EE, Micci-Smith O, & Munshi-South J (In Press) Genomic analyses identify multiple Asian origins and deeply diverged mitochondrial clades in inbred brown rats (*Rattus norvegicus*). *Evolutionary Applications*.
14. Lack J, Hamilton M, Braun J, Mares M, & Van Den Bussche R (2013) Comparative phylogeography of invasive *Rattus rattus* and *Rattus norvegicus* in the U.S. reveals distinct colonization histories and dispersal. *Biological Invasions* 15(5):1067-1087.
15. Puckett EE*, et al.* (2016) Global population divergence and admixture of the brown rat (*Rattus norvegicus*). *Proceedings of the Royal Society B: Biological Sciences* 283(1841):1-9.
16. Zeng L*, et al.* (2017) Out of Southern East Asia of the Brown Rat Revealed by Large-Scale Genome Sequencing. *Molecular Biology and Evolution*.
17. Tucker J (2015) *The Silk Road: China and the Karakorum Highway* (I.B.Tauris & Company, New York, USA) p 254.
18. Frantz LAF*, et al.* (2016) Genomic and archaeological evidence suggest a dual origin of domestic dogs. *Science* 352(6290):1228-1231.

508   19.   Shannon LM, *et al.* (2015) Genetic structure in village dogs reveals a Central Asian
509   domestication origin. *Proceedings of the National Academy of Sciences* 112(44):13639-
510   13644.
511   20.   Vernesi C, *et al.* (2003) The genetic impact of demographic decline and reintroduction in
512   the wild boar (Sus scrofa): A microsatellite analysis. *Molecular Ecology* 12(3):585-595.
513   21.   Gautier M, *et al.* (2016) Deciphering the Wisent Demographic and Adaptive Histories
514   from Individual Whole-Genome Sequences. *Molecular Biology and Evolution*
515   33(11):2801-2814.
516   22.   Estoup A, Beaumont M, Sennedot F, Moritz C, & Cornuet JM (2004) Genetic analysis of
517   complex demographic scenarios: Spatially expanding populations of the cane toad, *Bufo*
518   *marinus. Evolution* 58(9):2021-2036.
519   23.   Murray GGR, *et al.* (2017) Natural selection shaped the rise and fall of passenger pigeon
520   genomic diversity. *Science* 358(6365):951.
521   24.   Jones EP & Searle JB (2015) Differing Y chromosome versus mitochondrial DNA
522   ancestry, phylogeography, and introgression in the house mouse. *Biological Journal of*
523   *the Linnean Society* 115(2):348-361.
524   25.   Schiffels S & Durbin R (2014) Inferring human population size and separation history
525   from multiple genome sequences. *Nature Genetics* 46(8):919-925.
526   26.   Deinum EE, *et al.* (2015) Recent evolution in *Rattus norvegicus* is shaped by declining
527   effective population size. *Molecular Biology and Evolution* 32(10):2547-2558.
528   27.   Teng H, *et al.* (2017) Population Genomics Reveals Speciation and Introgression
529   between Brown Norway Rats and Their Sibling Species. *Molecular Biology and*
530   *Evolution* 34(9):2214-2228.
531   28.   Zheng B, Xu Q, & Shen Y (2002) The relationship between climate change and
532   Quaternary glacial cycles on the Qinghai–Tibetan Plateau: review and speculation.
533   *Quaternary International* 97-98(Supplement C):93-101.
534   29.   Moreno-Mayar JV, *et al.* (2018) Terminal Pleistocene Alaskan genome reveals first
535   founding population of Native Americans. *Nature* 553:203.
536   30.   Lieberman V (2009) *Strange Parallels: Southeast Asia in Global Context, c. 800-1830*
537   (Cambridge University Press, Cambridge, Great Britain) p 947.
538   31.   Heng D (2009) *Sino-Malay Trade and Diplomacy from the Tenth through the Fourteenth*
539   *Century* (Ohio University Press, Athens, USA) p 286.
540   32.   Beerli P (2004) Effect of unsampled populations on the estimation of population sizes
541   and migration rates between sampled populations. *Molecular Ecology* 13(4):827-836.
542   33.   Slatkin M (2005) Seeing ghosts: the effect of unsampled populations on migration rates
543   estimated for sampled populations. *Molecular Ecology* 14(1):67-73.
544   34.   Aplin KP, *et al.* (2011) Multiple geographic origins of commensalism and complex
545   dispersal history of black rats. *PLoS ONE* 6(11):e26357.
546   35.   Suzuki H, *et al.* (2013) Evolutionary and dispersal history of Eurasian house mice *Mus*
547   *musculus* clarified by more extensive geographic sampling of mitochondrial DNA.
548   *Heredity* 111(5):375-390.
549   36.   Gibbs RA, *et al.* (2004) Genome sequence of the Brown Norway rat yields insights into
550   mammalian evolution. *Nature* 428(6982):493-521.
551   37.   Li H & Durbin R (2010) Fast and accurate long-read alignment with Burrows-Wheeler
552   transform. *Bioinformatics* 26(5):589-595.

553   38.   McKenna A*, et al.* (2010) The Genome Analysis Toolkit: A MapReduce framework for
554   analyzing next-generation DNA sequencing data. *Genome Research* 20(9):1297-1303.
555   39.   Korneliussen TS, Albrechtsen A, & Nielsen R (2014) ANGSD: Analysis of Next
556   Generation Sequencing Data. *BMC Bioinformatics* 15(1):1-13.
557   40.   Li H*, et al.* (2009) The Sequence Alignment/Map format and SAMtools. *Bioinformatics*
558   25(16):2078-2079.
559   41.   Purcell S*, et al.* (2007) PLINK: A tool set for whole-genome association and population-
560   based linkage analyses. *The American Journal of Human Genetics* 81(3):559-575.
561   42.   Chang CC*, et al.* (2015) Second-generation PLINK: rising to the challenge of larger and
562   richer datasets. *GigaScience* 4(1):7.
563   43.   Green RE*, et al.* (2010) A draft sequence of the Neandertal genome. *Science*
564   328(5979):710-722.
565   44.   R Core Team (2013) R: A language and environment for statistical computing (R
566   Foundation for Statistical Computing, Vienna, Austria).
567   45.   Meredith M & Kruschke J (2016) Highest (Posterior) Density Intervals (CRAN, CRAN),
568   0.1.3.
569   46.   Scheet P & Stephens M (2006) A fast and flexible statistical model for large-scale
570   population genotype data: Applications to inferring missing genotypes and haplotypic
571   phase. *American Journal of Human Genetics* 78(4):629-644.
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