# Kinetics of winter deacclimation in response to temperature determines dormancy status and quantifies chilling requirement in grapevines (*Vitis* spp.)

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**Title**

Kinetics of winter deacclimation in response to temperature determines dormancy status and quantifies chilling requirement in grapevines (*Vitis* spp.)

**Running title**

Kinetics of deacclimation resolves dormancy status

**Highlight**

Deacclimation rates in *Vitis* species appear to follow enzyme kinetics; and using deacclimation rates, dormancy transitions can be studied in a quantitative way.

**Abstract**

Dormancy and cold hardiness are critical adaptations for surviving winter cold stress for temperate perennial plant species, with shifting temperature-based responses during the winter. The objective of this study was to uncover the relationship between dormancy transition (chilling requirement) and temperature on the loss of cold hardiness. Dormant cuttings of European grapevine cultivars (*Vitis vinifera*; 'Cabernet Franc', 'Cabernet Sauvignon', 'Riesling', and 'Sauvignon blanc') and wild grapevine species (*V. aestivalis*, *V. amurensis*, and *V. riparia)* were examined to determine the relationship between chilling requirement and temperature on rate of deacclimation ($k_{deacc}$). Differential thermal analysis was used to determine $k_{deacc}$ using mean low temperature exotherms. Effect of chill was evaluated as the deacclimation potential ($\Psi_{deacc}$), which was the change in $k_{deacc}$ due to chill accumulation. Results indicate that $\Psi_{deacc}$ varies dependent on dormancy state, following a logarithmic response to chill accumulation. The effect of temperature on $k_{deacc}$ was exponential at low temperatures and logarithmic at high temperatures. The combination of $\Psi_{deacc}$ and $k_{deacc}$ resulted in good prediction of deacclimation, demonstrating that $k_{deacc}$ can be used as a quantitative determinant of dormancy transition. This information can be used to refine models predicting effects of climate change on dormancy and cold hardiness in grapevine.

**Key words**

Bud, cold hardiness, deacclimation, dormancy, grapevines, rates, supercooling, *Vitis*.

**Abbreviations**

DTA – Differential thermal analysis

HTE – High temperature exotherm

LTE – Low temperature exotherm

$k_{deacc}$ – Deacclimation rate

$\Psi_{deacc}$ – Deacclimation potential

Chill – Chill accumulation

t – time

T – temperature

## Introduction

Due to their general stationary habit, plants have evolved many different coping mechanisms to survive stressful environments such as drought and temperature. With regard to winter, annual plants typically survive as dehydrated seeds while perennial plants develop dormancy and cold hardiness. Dormancy is the temporary cessation of visible growth in meristem containing structures, such as buds, and is divided in three types: para-, endo-, and ecodormancy (Lang et al. 1987). Paradormancy is the suspension due to physiological factors within the plant but outside of the dormant structure, such as the suppression of lateral growth by the apical meristem (i.e. apical dominance). Paradormant buds transition into endodormancy as daylength and temperatures decrease, a state suspension of growth due to unknown endogenous factors within the dormant structure, preventing growth during times when environmental conditions fluctuate between conducive and inhibitory (Horvath et al., 2003). During endodormancy, it is generally understood that exposure to low, non-freezing temperatures is necessary (chilling requirement) to transition to the third dormancy state, ecodormancy. Tissues maintain a dormant state during ecodormancy due to unsuitable environmental conditions.

Concomitantly with the onset of dormancy, plants from areas where temperatures drop below freezing develop cold hardiness through different mechanisms. Some plants, or plant

tissues, tolerate intracellular ice formation, in which cells are extremely dehydrated by the formation of extracellular ice, and the remaining water is bound to proteins (Burke et al., 1976) likely solidifying in a glass-like state. Other plants present a freezing resistance mechanism, where water supercools in the intracellular spaces (Burke et al., 1976). Supercooling is a process through which water can remain in liquid state below its equilibrium freezing point, down to a maximum of ~ -40 °C (Bigg, 1953). These plants are typical of mid-latitude temperate climates, where temperatures do not drop below -40 °C during the winter. In buds that present supercooling ability, such as those of grapevines (*Vitis* spp.), cold hardiness may be measured through differential thermal analysis (DTA). This method uses thermoelectric modules that record voltage changes associated with the release of heat due to phase change of water into ice (Mills et al., 2006). In grapevine buds, two peaks can be identified in DTA: a high temperature exotherm (HTE), typically around – 5 °C; and a low temperature exotherm (LTE), which varies depending on the conditions experienced by the bud. The HTE represents the freezing of extracellular water (Andrews et al., 1984), which is normal to the process of resisting cold. The LTE, however, is the freezing of intracellular water, and it happens when the supercooling point has been exceeded (Mills et al., 2006). The LTE represents the lethal temperature for a given bud, is correlated with manual assessments of bud death (Wolf and Cook, 1994), and mean LTE has been adopted as a consistent measurement of bud cold hardiness.

A general U-shaped pattern of changing cold hardiness is followed during the winter through three different stages: acclimation, maintenance, and deacclimation. This pattern is thought to be primarily driven by air temperature (Ferguson et al., 2011, 2014; Londo and Kovaleski, 2017), although other climatic aspects may also be important (Antivilo et al., 2017). Acclimation appears to occur primarily during endodormancy, while deacclimation is enhanced during ecodormancy (Ferguson et al., 2011, 2014; Londo and Kovaleski, 2017). Depending on the climate, the transition between endo- and ecodormancy may occur toward the end of the acclimation stage, or during the maintenance stage of cold hardiness. This transition is typically evaluated by collecting dormant buds from the field and placing them in growth permissive conditions, followed by monitoring the time needed to reach budbreak (Weinbaum et al., 1989; Lloyd and Firth, 1990; Cook and Jacobs, 2000; Fan et al., 2010; Zhang and Taylor, 2011). The chilling requirement has been met when 50% of buds are capable of

breaking within a time threshold {e.g. 3 weeks for peaches [*Prunus persica* (L.) Batsch; Lloyd and Firth, 1990]; 4 weeks for grapevines (Londo and Johnson, 2014)}. This method represents a subjective threshold for the change in endo- to ecodormancy transition. As a result, determining the molecular and metabolic cues and the consequences of this transition, such as budbreak, are very difficult to determine and poorly understood (Penfield, 2008).

As the dynamics of acclimation and deacclimation change with dormancy state (Ferguson et al. 2011, 2014), and dormancy state is determined by accumulation of chilling hours, a future climate may create issues for both cold hardiness and budbreak phenology. Average global temperatures are increasing (Walsh et al., 2014), although episodes of acute cold weather in the northern hemisphere are likely to increase as well through more frequent arctic oscillations (Kolstad et al., 2010). In addition to direct effects to cold hardiness, increasing temperatures will enhance chill accumulation in higher latitudes, while lower latitudes will experience a decrease (Luedeling et al., 2011). Changes in chill accumulation will modify the dynamics of dormancy transitions both in deciduous tree crops and forests, causing excessive responsiveness to warm spells or erratic budbreak and growth. This will ultimately require a shift towards higher latitudes of tree crops and assisted migration of forest species as they lag behind their optimal climate niche (Gray and Hamann, 2013). Therefore, it is important to understand how both temperature and dormancy status affect the loss of cold hardiness. The objective of this study was to better predict and understand the relationship between chill accumulation, the dormancy transition, and the loss of cold hardiness in wild and cultivated grapevines.

## Materials and methods

Buds of four different species were collected multiple times during the dormant seasons of 2014/15, 2015/16, and 2016/17. *V. vinifera* L. ‘Cabernet Franc’, ‘Cabernet Sauvignon’, ‘Riesling’, and ‘Sauvignon blanc’ were collected from local vineyards, while *V. aestivalis* Michx. (483138, 483143), *V. amurensis* Rupr. (588632, 588635, 588641), and *V. riparia* Michx. (588275, 588562, 588653, 588711) were collected from the USDA germplasm in Geneva, NY. For *V. vinifera*, buds from positions 3 to 20 were used, while for the other species, due to constraints regarding the number of clones, buds beyond position 20 were used as in Londo and Kovaleski (2017).

156 Hourly weather data from the closest Network for Environment and Weather
157 Applications station (NEWA; http://www.newa.cornell.edu/) was used to compute chill
158 accumulation using the "North Carolina" model (Shaltout and Unrath, 1983). The start date for
159 each season was chosen based on when chill started to consistently accumulate instead of
160 being negated, as determined by the NC model. These dates were 11 Sept 2014, 19 Sept
161 2015, and 24 Sept 2016.
162 Upon collection, canes were prepared into single- or two-node cuttings and placed in
163 cups of water. The cups were placed into growth chambers at 2, 4, 7, 8, 10, 11, 22, or 30 °C.
164 Not every genotype or temperature was used at all collection points, but this information is
165 available in Supplementary Table S1. Differential thermal analysis (DTA) was performed to
166 determine the cold hardiness of individual buds as their low temperature exotherm [LTE; see
167 Mills *et al.* (2006) for details]. Briefly, buds are excised from the cane and placed on a
168 thermoelectric module in a plate. These plates are placed in a programmable freezer with a
169 cooling rate of -4 °C/hour. Changes in voltage due to heat release in the freezing of water are
170 measured by the thermoelectric module and recorded using a Keithley data logger (Tektronix,
171 Beaverton, OR) attached to a computer. Between 4 and 8 buds were used at any time point to
172 determine mean LTE, and the intervals between measurement days varied between chill
173 accumulation level and temperature treatment, with details provided in Supplemental Table 1.
174 Because lower rates of deacclimation were expected in lower temperatures and lower chill
175 accumulations, lower temperature treatments were typically surveyed with wider separation
176 between time points compared with high temperature treatments (Table S1).
177
178 ***Statistics***
179 Based on sample size across temperatures and across sampled years, data sets for *V.*
180 *vinifera* were analyzed separately for each cultivar, while data for accessions of *V. aestivalis*,
181 *V. amurensis* and *V. riparia* were combined at the species level. Analyses were separated into
182 the effect of chill accumulation and the effect of temperature on rate of deacclimation. The
183 datasets used for each analysis are specified in Supplementary Table S1.
184
185 <u>*Effect of Chill Accumulation.*</u> In order to assess the effect of chill accumulation in the rate of
186 deacclimation ($k_{deacc}$), or the deacclimation potential ($\Psi_{deacc}$), individual rates were calculated

using linear regression in R (ver. 3.3.0, R Foundation for Statistical Computing) for each temperature and chill accumulation as factors. Although every temperature within a chill accumulation had the same data for day 0 (field collection), temperatures were still allowed to have different intercepts in order to reduce the effect of day 0 on $k_{deacc}$. From this regression model, data points that had a studentized residual ≥2.5 were considered outliers and removed from the dataset, and the model was re-fit. The $k_{deacc}$ at each chill accumulation were then transformed to percentage for 4, 7, 10, 11, and 22 °C, standardizing to their $k_{deacc}$ at highest chill accumulation (either 1440 – 7 and 11 °C – or 1580 chill units – 4, 10, and 22 °C). The $\Psi_{deacc}$ was estimated as a logistic regression. Initial estimation used the drc library, but final estimation used the nls() function with the port algorithm, following the equation: $\Psi_{deacc}\ (\%) = {}^{1}/_{1 + \ e^{[b \times (\ln Chill\ - c)]}}$, where *b* and *c* are the estimated parameters, and *Chill* is the chill accumulation at the time of sample collection. The parameter *c* is the inflection point of the logistic and *b* is the slope associated with the logistic regression.

*Effect of Temperature.* Initially, to study the effect of temperature, $k_{deacc}$ obtained from multiple linear regression (temperature and chill accumulation as factors; Table S1) in lower chill accumulations were re-scaled by dividing by the respective $\Psi_{deacc}$ (e.g., because the $\Psi_{deacc}$ was estimated to be at ~30% at 860 chill units for Riesling, the rates obtained from that data set were divided by 0.30). Since the effect of chill accumulation was estimated to be ~ 0 % for 360 chill units, these were excluded from the dataset for estimation of temperature effect. Arrhenius plots were created to evaluate temperature responses and data was then divided into two data sets, low (2 – 11 °C) and high temperatures (10 – 30 °C), and temperature effects on deacclimation rates were evaluated separately. The rationale for doing this is discussed below.

Instead of using the initially calculated rates derived from the above linear models, the complete data sets were used with the effect of temperature as a continuous variable. The values for the different chill accumulation points were normalized to that of full chill by multiplying the $\Psi_{deacc}$ at any given chill accumulation, effectively removing the effect of chill by mathematically reducing the time for deacclimation. Lower chill accumulations were thus included in the calculations instead of using only those where chill requirements were initially assumed to be fulfilled (1440 and 1580 chill accumulations).

For low temperatures, the effect was assumed to be an exponential, and therefore the rates were calculated as $k_{\text{deacc}} = m \times e^{(n \times T)}$, where *m* and *n* are the parameters estimated, and *T* is the temperature in °C. For high temperatures, a logarithmic curve was used: $k_{\text{deacc}} = q \times \ln(T - r)$, where *q* and *r* are the parameters estimated, and *T* is the temperature in °C. For both low and high temperatures, parameters were estimated using nls(), with the port algorithm.

The fits of all non-linear curves (effect of chill accumulation, low and high temperatures) were tested using Effron's pseudo-$R^2$ with the Rsq function in the soilphysics library, which is calculated using the sum of squares of ordinary residuals from the models. The final model for loss of cold hardiness from an initial point is then $\Delta_{LTE}(T, Chill) = k_{\text{deacc}} \times \Psi_{\text{deacc}} \times t$, where the change in LTE ($\Delta_{LTE}$) is a function of the temperature (*T*) it was exposed to, how much chill accumulation (*Chill*) had passed before exposed to deacclimation temperatures, and the time (*t*) of exposure. A linear model was used to evaluate the accuracy of the predictions using the complete data set. For this, instead of temperatures, $k_{\text{deacc}} = \begin{cases} m \times e^{(n \times T)}, T < 12\ ℃ \\ q \times \ln(T - r), T \geq 12\ ℃ \end{cases}$ was used, and instead of accumulated chill, the $\Psi_{\text{deacc}}$ was used. From this model ($\Delta_{\text{LTE}} = \beta \times k_{\text{deacc}} \times \Psi_{\text{deacc}} \times \text{time}$), considering estimations were appropriate, the β associated with the estimated $k_{\text{deacc}}$ and $\Psi_{\text{deacc}}$ was expected to be 1.

## Results

After removing outliers using multiple linear regression, data sets kept ≥ 90% of the observations for all genotypes, with the exception of *V. amurensis*, which had 19.5% of observations removed (Table 1). Linear behavior appears to describe well the effects of temperature and chill accumulation in the deacclimation of grapevine buds (Figure 1). A logistic regression appeared to be a good fit for the $\Psi_{\text{deacc}}$ (Figure 2), with pseudo-$R^2$'s greater than 0.87 for all genotypes except for 'Cabernet Sauvignon' and *amurensis* (0.65 and 0.79, respectively; Table 1). The parameter *e* is the natural logarithm of the inflection point, and there was little difference in this parameter for these genotypes (e.g., 6.77 to 6.88, for 'Sauvignon blanc' and 'Riesling', respectively, which is equivalent to 872 to 972 chill units).

Scaled deacclimation rates based on $\Psi_{\text{deacc}}$ and corresponding Arrhenius plot (Figure 3) demonstrated a clear discontinuity between low and high temperatures. A deceleration of the

increase of $k_{deacc}$ as temperature increased was observed, which justifies the different behaviors used (exponential and logarithmic for low and high temperatures, respectively). The estimation of temperature effects for low and high temperature intersect between 10 and 11 °C because of the uncertainty in the exact temperature where there is a change in behaviors. This is partially due to the lack of data points between 11 and 22 °C, although at 10 and 11 °C the calculated rates through either model would be very similar. All the *V. vinifera* cultivars had similar deacclimation rates at low temperatures, but 'Riesling' and 'Cabernet Franc' had higher deacclimation rates at high temperatures than 'Cabernet Sauvignon' and 'Sauvignon blanc' (Table 1, Figure 4). *V. riparia* and *V. aestivalis* had similar rates to those of *V. vinifera* at low temperatures, but *V. riparia* has estimated rates similar to *V. amurensis* in higher temperatures. *V. aestivalis* was similar to the faster *V. vinifera* ('Cabernet Franc' and 'Riesling'). *V. amurensis* showed the highest deacclimation rates at both low and high temperatures.

Linear models showed a good fit for all genotypes ($R^2 \geq 0.80$). *V. vinifera* cultivars, which were analyzed separately had higher $R^2$'s. The better fit for V. *vinifera* cultivars is also seen in their β coefficients, which are very close to 1, although *V. aestivalis* and *V. riparia* had βs of 0.99 and 0.96 respectively. *V. amurensis* had the lowest β, at 0.83.

## Discussion

Chilling accumulation, dormancy state, and cold hardiness are complex traits driven by the interaction of physiological and climactic attributes. The consistency of winter temperatures is expected to be a major aspect impacting the sustainability and survival of perennial species in a future climate. This study was conducted to gain a better method for predicting dormancy transition and chilling requirement in grapevine, as well as understand how deacclimation processes (loss of cold hardiness) is impacted by dormancy state.

This paper provides a quantitative assessment of the transition from endo- to ecodormancy, as evaluated by the $\Psi_{deacc}$. Unlike the typical dormancy assessment that uses time to budbreak as a measurement of the transition (Londo and Johnson, 2014), using the deacclimation rates does not require subjective assumptions currently used to determine chilling fulfillment; such as having 50% budbreak occur within 28 days. Budbreak occurs only

after the freeze mechanisms associated with cold hardiness are lost (Ferguson et al., 2014; Salazar-Gutiérrez et al., 2014). Thus, the amount of time needed to reach 50% budbreak in forcing assays is a combination of the time needed to fully deacclimate, and time required for deacclimated buds to reach a visible change in phenological stage. This study leverages the ability to assess loss of hardiness through decreasing supercooling ability as a way to quantitatively assess rates of deacclimation and its relation to changes in chilling exposure.

Mid-winter warming events are of major concern for temperate crops, yet natural areas are also affected. In native trees within natural ranges, little to no damage occurs during the autumn because the freeze resistance is usually much greater than the minimum temperatures they usually encounter (Vitasse et al., 2014). However, damage can be substantial in late freeze events during the spring or unusual warm events in early spring as the potential for deacclimation increases exponentially (Vitasse et al., 2014). For example, a late spring freeze in 2007 in North America caused damage in native forests, delaying and reducing canopy establishment by 16-34 days (Gu et al., 2008). Bokhorst et al. (2009) report that dwarf shrub vegetation in a sub-Artic region where midwinter temperatures rose to ~7 °C for 2 weeks followed by further cold temperatures resulted in 87% reduction in growth during the summer compared to neighboring undamaged areas. Cannell and Smith (1983) show results from different authors for forest species in which the time to budbreak is reduced in an exponential manner. This is similar to our assessment of $\Psi_{deacc}$, where the upper portion of the logistic is like the exponential decrease in time to budbreak. The logistic behavior of $\Psi_{deacc}$ in relation to chill accumulation explains the continuous increase risk of damage of mid-winter warm spells the later they occur until 100% $\Psi_{deacc}$ is achieved. This is in contrast with the model developed by Ferguson et al. (2011; 2014), where a hard limit for the dormancy boundary is used to model grape cold hardiness. It is likely that, similar to the behavior described in our study, forest species also have increased deacclimation rates with higher chill accumulation, seen as a decrease in the time to budbreak. Additionally, it is important to note that while canopies may be able to recover from midwinter bud kill events, damage to the reproductive buds in the case of crops will reduce yields in the season following damage.

Our assessment of $\Psi_{deacc}$ leading to increases in $k_{deacc}$ agrees with the general observation that warm temperatures in the early winter are negligible for bud growth (Cannell and Willett, 1975) and that increasing chill accumulation reduces the heat requirements for

budbreak in fruit species (Couvillon and Erez, 1985; Citadin et al., 2001), and forest species (Hunter and Lechowicz, 1992; Dantec et al., 2014), considering that loss of hardiness is required for budbreak to occur (Ferguson et al., 2014). Even at full chilling (100% $\Psi_{deacc}$), differences in the $k_{deacc}$ between species and cultivars may be responsible for differences in early phenological development in the spring, but it is important to recognize there is an interaction between both. Planted in the same environment, Taulavuori et al. (2004) observed that deacclimation occurs faster in mountain birch [*Betula pubescens* ssp. *czerepanovii* (Orl.) Hämet-Ahti] ecotypes with origin in areas of low chill accumulation. If the mechanisms are similar to those described here, what may have been observed in their study was not different rates of deacclimation, but plants that were at different stages in their deacclimation potential, or a combination of both. Climate change appears to speed the loss of cold hardiness and promote earlier leaf-out, although the response is not linear due to interactions with chilling and photoperiod (Vitasse et al., 2014). In our case, the predicted increase in chill accumulation (Luedeling et al., 2011) will result in higher $\Psi_{deacc}$ and higher risk of damage should midwinter warm spells increase in a future climate (Vitasse et al., 2014; Walsh et al., 2014). However, earlier budbreak may not lead to higher frequency of damage due to frost events if consistently higher spring temperatures (Chmielewski et al., 2018) accompany future climate changes.

Pagter and Arora (2013) suggest that deacclimation rates are not linear, and that a lag phase may occur during initial exposure to warm temperatures. The initial lag phase might explain the differences in deacclimation rates measured in Cragin et al. (2017) compared to those reported in this study, as they only evaluated deacclimation for 4 days. This may also suggest why they had differences in rates across the years, although this may also be an effect of different chill accumulation between the two years in their study. Ferguson et al. (2011; 2014) used asymptotic bounds added through a logistic component, deviating from a strictly linear rate. In our data, although in some cases an initial curvature of the data points can be observed (e.g., Figure 1, 10 °C – 860 chill units), linear rates were strong descriptors of the deacclimation rates as observed in the fits of our calculated rates of deacclimation and in the early multiple linear regression models with temperature and chill accumulation as factors ($R^2 > 0.83$, $p < 0.001$ for all cultivars; Table S1), similar to Wolf and Cook (1992), also looking at deacclimation rates for grapevines – although they only used 1 temperature. We also stopped measuring LTEs when those were close to -5 °C, where we had difficulties reading

341 LTE peaks due to the merge with the much larger high temperature exotherms, and therefore
342 any curvature in the higher portions of the deacclimation curve would not be observed. Such
343 curvature is observed in Arora et al. (2004) and Kalberer et al. (2007), although hardiness loss
344 stopped at a lower temperature, which was speculated to be due to lack of chill.
345 Ferguson et al. (2011; 2014) used a linear response of the rates of deacclimation in
346 relation to temperature for 21 *V. vinifera* and two *V. labruscana* Bailey cultivars, with a different
347 rate and base temperature for endo- and ecodormant buds. In our experiments, deacclimation
348 occurred in temperatures below those established as the threshold for deacclimation by
349 Ferguson et al. (2014). This suggests that although it may produce a good fit for field data, the
350 current model may be over-simplifying the response to temperature from a physiological
351 standpoint. Takeuchi and Kasuga (2017) have also seen deacclimation through electrolyte
352 leakage of bark and xylem of Japanese white birch [*Betula platyphylla* Sukatchiev var. *japonica*
353 (Miq.)] at low temperatures, including high below-freezing temperatures (-5 to 0°C). In our
354 study the temperature dependency of $k_{deacc}$ appears to follow simple enzymatic kinetics,
355 affected by both the catalytic rate (Arrhenius' law) in lower temperatures, and the denaturation
356 of enzymes in higher temperatures. This behavior suggests that an active temperature
357 sensing, as defined by Ruelland and Zachowski (2010), is not necessary for deacclimation.
358 Although deacclimation is likely controlled by a complex metabolic chain or cycle, the rates of
359 any given process as a whole are defined by the slowest enzyme – which may change
360 depending on the temperature (Ruelland and Zachowski, 2010). Additionally, Farquhar and
361 Richards (1984) proposed that whole plant processes should be analyzed as simple chemical
362 processes in respect to temperature. Therefore, further comparisons are made in this
363 discussion with activities of single enzymes and whole metabolic cycles without distinction.
364 It is notable in the deacclimation process measured in this study that rates are departing
365 from the exponential increase due to temperature in low range (estimated as ≥ 12 °C) as seen
366 in the Arrhenius plot (Fig. 3B). This would suggest that some limitation occurs at higher
367 temperature to the process as a whole: either substrate limitations or enzyme denaturation
368 (Truhlar and Kohen, 2001). It is important to note, however, that there is no drop in the $k_{deacc}$
369 from 22 to 30 °C, but a reduction on the increase of $k_{deacc}$ as compared to from 2 to 10 °C (Fig.
370 3). This differs from other processes in grapevines, such as root respiration, which has an
371 exponential response to temperature up to 32 °C for short-term exposure (Huang et al., 2005).

372 This is likely an adaptation of the enzymes involved in the process of deacclimation,
373 considering low temperatures in early spring when deacclimation occurs. Rigano et al. (2006)
374 demonstrated similar kinetics when comparing the activity of methyl viologen-dependent nitrate
375 reductase extracts from psychrophilic and mesophilic algae. Arrhenius plots of the
376 psychrophilic alga had an inflection point at 15 °C, whereas the mesophilic alga had an
377 inflection point at 25 °C. We made no attempt in our study to find different inflection points for
378 each species, as we lacked data in the 12-21 °C range. However, future studies could be
379 conducted to examine if deacclimation rate is adaptive in widely distributed *Vitis* species as
380 well. The activity of some enzymes, such as catalases and peroxidases, appears to be
381 correlated with release of dormancy (Farokhzad et al., 2018) and subsequent budbreak (Pérez
382 and Burgos, 2004). It may be interesting to measure the activity of these enzymes at different
383 temperatures to evaluate whether they have a similar response to temperature as that
384 evaluated in the deacclimation here. Other enzymes involved in the synthesis and degradation
385 of plant hormones might also be involved, considering that growth inhibition by abscisic acid or
386 promotion by gibberellins and cytokinins appear to interplay in the control of dormancy
387 (Horvath et al., 2003).
388 In a review of photosynthesis responses to temperature, Berry and Björkman (1980)
389 showed that the phase of exponential increase in photosynthesis rate can end at temperatures
390 as low as 10 °C and as high as 40 °C, and that both the rates and the optimum temperatures
391 are also affected by the temperature regime in which plants are grown. For deacclimation rates
392 in grapevine buds, Cragin et al. (2017) suggested an effect of the field LTE (or the intercept)
393 on the deacclimation rate, which would indicate an effect of the environment. We did not
394 attempt to use field LTE as a factor in our calculations; however, we expect that it would not
395 greatly impact $k_{deacc}$: both 860 and 1580 chill units had similar field LTEs, but very different
396 $k_{deacc}$ (Fig. 1). We believe that most of the environmental effect on the $k_{deacc}$ is well described
397 by the chill accumulation through the $\Psi_{deacc}$.
398 Cultivar or accession differences are seen within species in cold hardiness (Arora et al.,
399 2004; Ferguson et al., 2011, 2014; Salazar-Gutiérrez et al., 2014, 2016; Szalay et al., 2017),
400 although deacclimation rates may or may not differ (Arora et al., 2004; Ferguson et al., 2011,
401 2014; Szalay et al., 2017). The effect of genotypes within species in $k_{deacc}$ is suggested by the
402 better fit of our curves to *V. vinifera* cultivars as compared to the other species, where

403 parameters were estimated by combining different accessions in the wild material. However,
404 our dataset did not allow us to properly analyze each genotype of the wild accessions
405 separately. It has been suggested that there is no correlation among species between the
406 maximum cold hardiness or climate of origin and the rate of deacclimation (Arora et al., 2004;
407 Kalberer et al., 2006). Vitasse et al. (2014), however, suggests that the rate of deacclimation is
408 more related to temperature fluctuations in the area where species evolved, diminishing the
409 potential for deacclimation in locations where frequent temperature fluctuations occur in the
410 winter. In our case, *V. riparia* and *V. amurensis*, two species from colder climates as compared
411 to *V. vinifera*, had higher $k_{deacc}$ at all (*V. amurensis*) or at moderate to high temperatures (*V.*
412 *riparia*) than *V. vinifera*. *V. aestivalis*, however, did not appear to differ from the *V. vinifera*
413 cultivars, in agreement with the assessment by Kalberer et al. (2006). Kalberer et al. (2006)
414 also suggested that field and controlled studies can result in different responses to
415 temperature. In our study, however, we observed higher rates of deacclimation in *V. amurensis*
416 and *V. riparia* compared to *V. aestivalis*, which is comparable and follows the same behavior
417 as the "responsiveness" described by Londo and Kovaleski (2017) using field observations.
418 Cannell and Smith (1983) speculate that average daily temperature is a good descriptor
419 for timing of budbreak, and that daily amplitude and photoperiod may be ignored. Similarly,
420 Antivilo et al. (2017) saw no or little differences in deacclimation due to low and high
421 temperature amplitudes, respectively. Although this was not tested, it appears that a linear
422 approximation may be used for temperature effects on deacclimation, especially in small
423 temperature amplitudes. Therefore, using daily average temperatures may be appropriate for
424 field data. However, long-term averages could result in larger error in assessments, and
425 physiological-based studies should use more precise descriptions of temperature effects such
426 as those reported here.
427 It is possible that the accuracy of models for prediction of budbreak may be increased if
428 they are attached to predictions of cold hardiness and deacclimation kinetics. Rubio et al.
429 (2016) reported both LTE and time to 50% budbreak for *V. vinifera* 'Thompson seedless',
430 which appear to be positively correlated in the acclimation period. Although the authors did not
431 report chill accumulation, it is clear that cold hardiness increases (more negative LTEs) and
432 days to budbreak decreases once low temperatures start occurring. In their case the time to
433 budbreak appears to reach the limit of endodormancy [50% budbreak within 28 days as per

Londo and Johnson (2014)] in early June in Chile, but continue dropping until their last evaluation in August. This is in accordance with our assessment that as chill accumulates, there is an increase in $\Psi_{deacc}$, leading to potentially faster budbreak. Because of this relationship between $\Psi_{deacc}$ (through chill accumulation) and $k_{deacc}$ leading to budbreak, we hypothesize that studies that identify earlier or faster budbreak due to the use of hydrogen cyanamide (Pérez and Burgos, 2004; Pérez and Lira, 2005) would likely find faster rates of deacclimation as well.

Specific studies linking environmental factors to understand plant phenology in order to evaluate impacts of climate change are needed (Cleland et al., 2007; Chmielewski et al., 2018). In this study, we demonstrate an objective method for determining the dormancy transition that could prove useful in other perennial species that utilize supercooling for cold hardiness [e.g. peach, cherry (*Prunus* spp. L.), azalea (*Rhododendron* spp. L.), larch (*Larix kaempferi* Sarg.)] in order to understand how they might respond to the future shifts in chill accumulation and increase in average temperature as a consequence of climate change. Although measurements of cold hardiness may be different for each species, it appears that this phenotype can aid in understanding dormancy transitions and spring phenology of plants. It is clear that simple assessments of deacclimation rates simply using “endo-” and “ecodormant” material is not appropriate, as there is a continuum of response in deacclimation rates as chill is accumulated. Additionally, although we made no attempt to evaluate different chill accumulation models, it is possible that using deacclimation rates may be a more appropriate way of defining chilling temperatures, considering the more quantitative nature of this analysis.

**Supplementary data**

Table S1. Information associated with each deacclimation experiment for each genotype used. Linear models were fit per genotype using chill accumulation and temperature as factors.

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**Tables**

Table 1. Parameter estimates and model fits for deacclimation potential, low and high temperature deacclimation rates, and linear relationship between LTE and the deacclimation rate × deacclimation potential for 7 *Vitis* genotypes.

**Figures**

Figure 1. Deacclimation of *V. vinifera* 'Riesling' buds at three temperatures collected from the field at three different chill accumulations.

Figure 2. Deacclimation potential of *V. vinifera* 'Riesling' buds collected at different chill accumulations in two temperatures. Deacclimation rate at 1580 chill units was used as reference (100 %). The relationship between deacclimation potential and accumulated chill was described by the equation: $\Psi_{deacc}\,(\%) = {}^{1}/_{1 + e^{[-7.23 \times (\ln Chill - 6.88)]}}$, with a pseudo-$R^2$ = 0.87.

Figure 3. Effect of temperature on deacclimation rates of *V. vinifera* 'Riesling'. A) Scaled deacclimation rates (based on deacclimation potential) at three different chill accumulations in response to temperature; B) Arrhenius plot of scaled deacclimation rates. Tick marks in both panels are equivalent and mirrored (e.g. 0 °C = 3.66 × 1000 $K^{-1}$).

Figure 4. Deacclimation rates of *Vitis* buds as a function of temperature. A) Deacclimation rates at >90% deacclimation potential for three different *Vitis* genotypes: *V. riparia* (closed circle – full line), *V. vinifera* 'Riesling' (open circles – dashed line), and *V. vinifera* 'Cabernet Sauvignon' (closed squares – dotted line); B) Deacclimation rates of *V. vinifera* 'Riesling' at different chill accumulations as a function of temperature.

Table 1

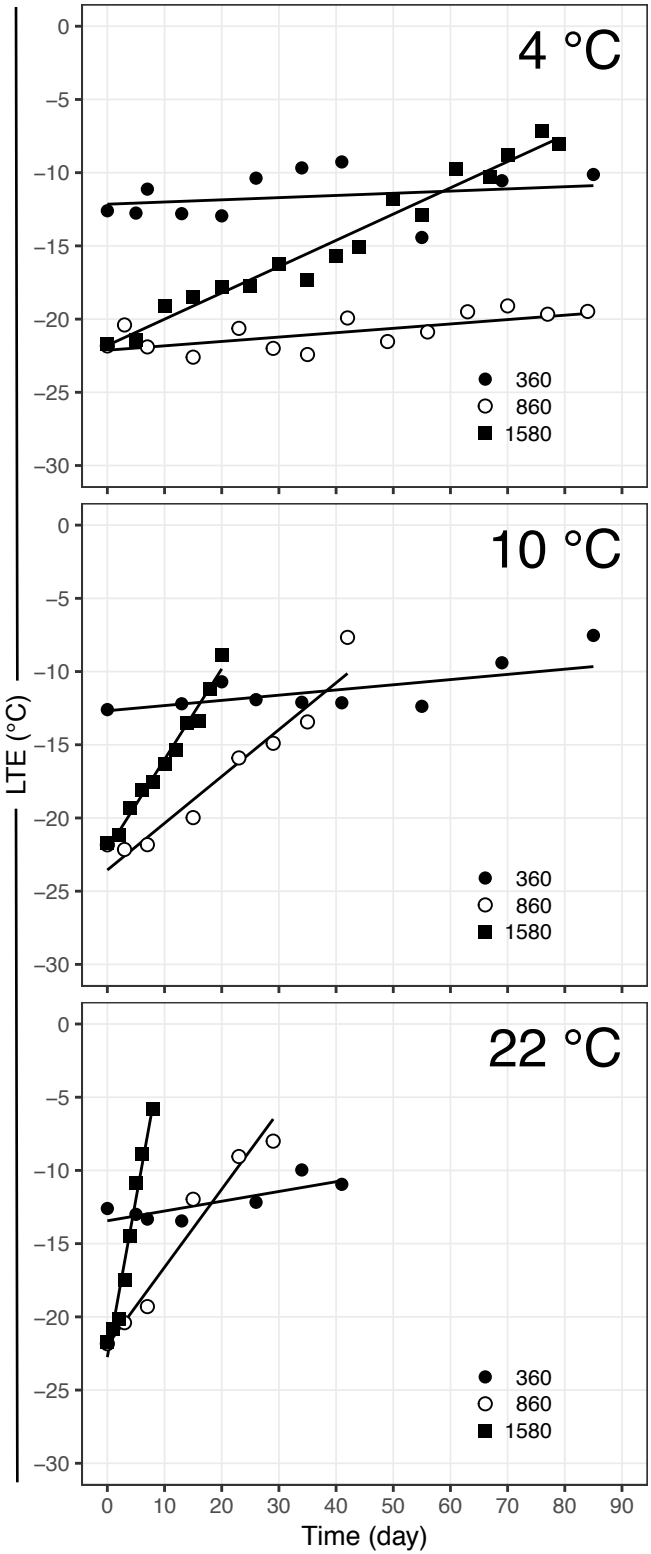
| Species | Cultivar | Deacclimation potential[a] | | | |
|---|---|---|---|---|---|
| | | b | e | Convergence | Pseudo-$R^2$ |
| *V. vinifera* | 'Cab. Franc' | -9.47 | 6.79 | Relative | 0.93 |
| | 'Cab. Sauvignon' | -8.27 | 6.80 | Relative | 0.65 |
| | 'Riesling' | -7.23 | 6.88 | Relative | 0.87 |
| | 'Sauvignon blanc' | -43.66 | 6.77 | Relative | 0.93 |
| *V. aestivalis* | | -22.31 | 6.86 | Relative and x | 0.97 |
| *V. amurensis* | | -12.94 | 6.85 | Relative | 0.79 |
| *V. riparia* | | -9.79 | 6.82 | Relative | 0.96 |
| | | Low temperature[b] | | | |
| | | m | n | Convergence | Pseudo-$R^2$ |
| *V. vinifera* | 'Cab. Franc' | 0.0600 | 0.253 | Relative | 0.89 |
| | 'Cab. Sauvignon' | 0.0620 | 0.218 | Relative | 0.87 |
| | 'Riesling' | 0.0675 | 0.236 | Relative | 0.92 |
| | 'Sauvignon blanc' | 0.0542 | 0.235 | Relative and x | 0.84 |
| *V. aestivalis* | | 0.0965 | 0.187 | Relative and x | 0.80 |
| *V. amurensis* | | 0.1988 | 0.211 | Relative | 0.75 |
| *V. riparia* | | 0.0754 | 0.259 | Relative | 0.81 |
| | | High temperature[c] | | | |
| | | q | r | Convergence | Pseudo-$R^2$ |
| *V. vinifera* | 'Cab. Franc' | 0.762 | 7.52 | Relative | 0.92 |
| | 'Cab. Sauvignon' | 0.593 | 7.73 | Relative | 0.84 |
| | 'Riesling' | 0.732 | 7.39 | Relative | 0.88 |
| | 'Sauvignon blanc' | 0.513 | 6.66 | Relative | 0.83 |
| *V. aestivalis* | | 0.830 | 7.91 | Relative and x | 0.80 |
| *V. amurensis* | | 0.913 | 3.34 | Relative | 0.80 |
| *V. riparia* | | 0.977 | 7.32 | Relative | 0.84 |
| | | Linear model[d] | | | |
| | | β | $R^2$ | N | n (%) |
| *V. vinifera* | 'Cab. Franc' | 0.962 | 0.88 | 853 | 0.96 |
| | 'Cab. Sauvignon' | 0.997 | 0.84 | 1160 | 0.93 |
| | 'Riesling' | 1.018 | 0.90 | 1260 | 0.93 |
| | 'Sauvignon blanc' | 1.002 | 0.86 | 745 | 0.90 |
| *V. aestivalis* | | 0.963 | 0.80 | 700 | 0.92 |
| *V. amurensis* | | 0.828 | 0.83 | 730 | 0.81 |
| *V. riparia* | | 0.987 | 0.82 | 1499 | 0.93 |

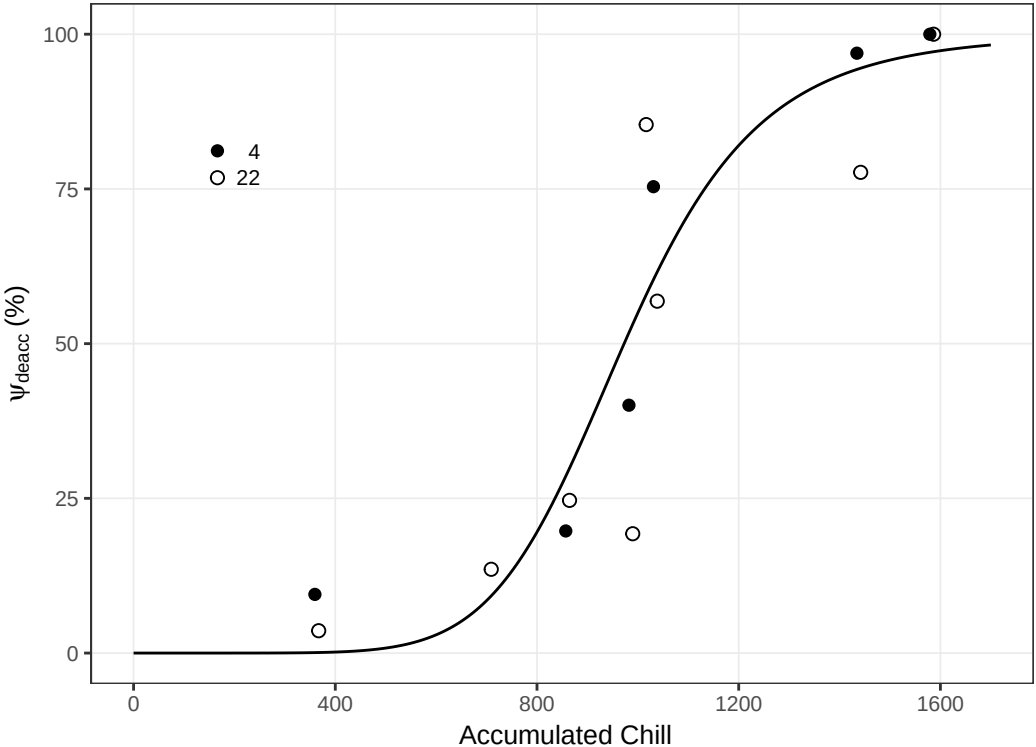
[a] $\Psi_{deacc}\ (\%) = {}^{1}/_{1 + e^{[b \times (\ln Chill - c)]}}$, where *Chill* = chill accumulation

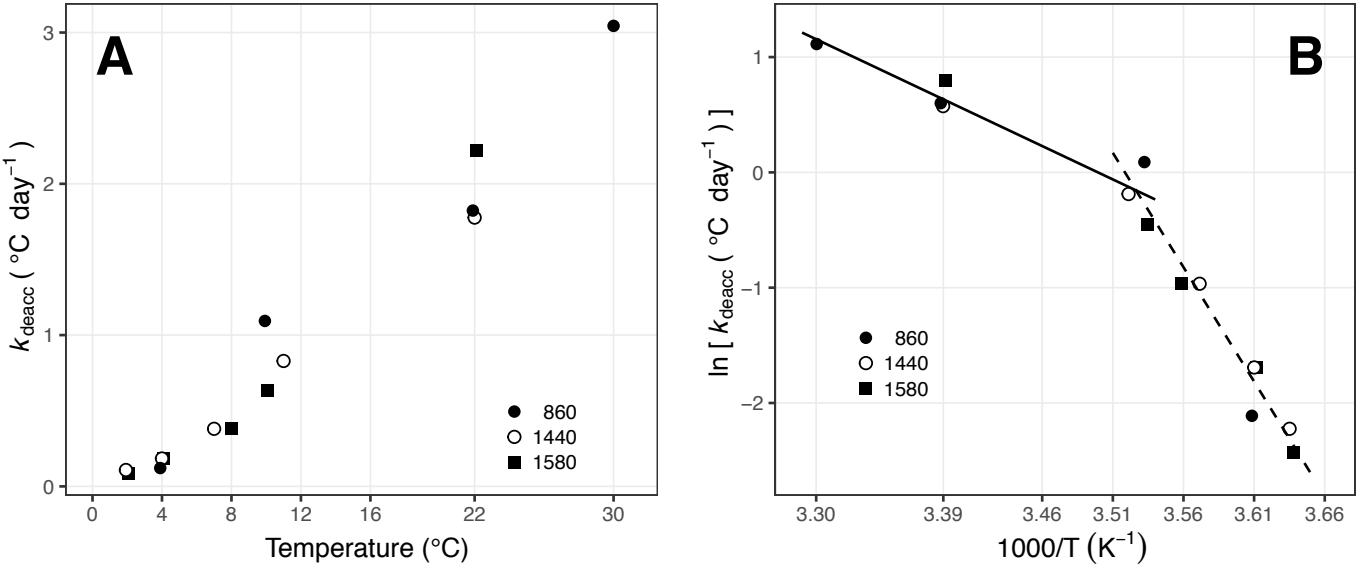
[b] $k_{\mathrm{deacc}} = m \times e^{(n \times T)}$, where *T* = temperature

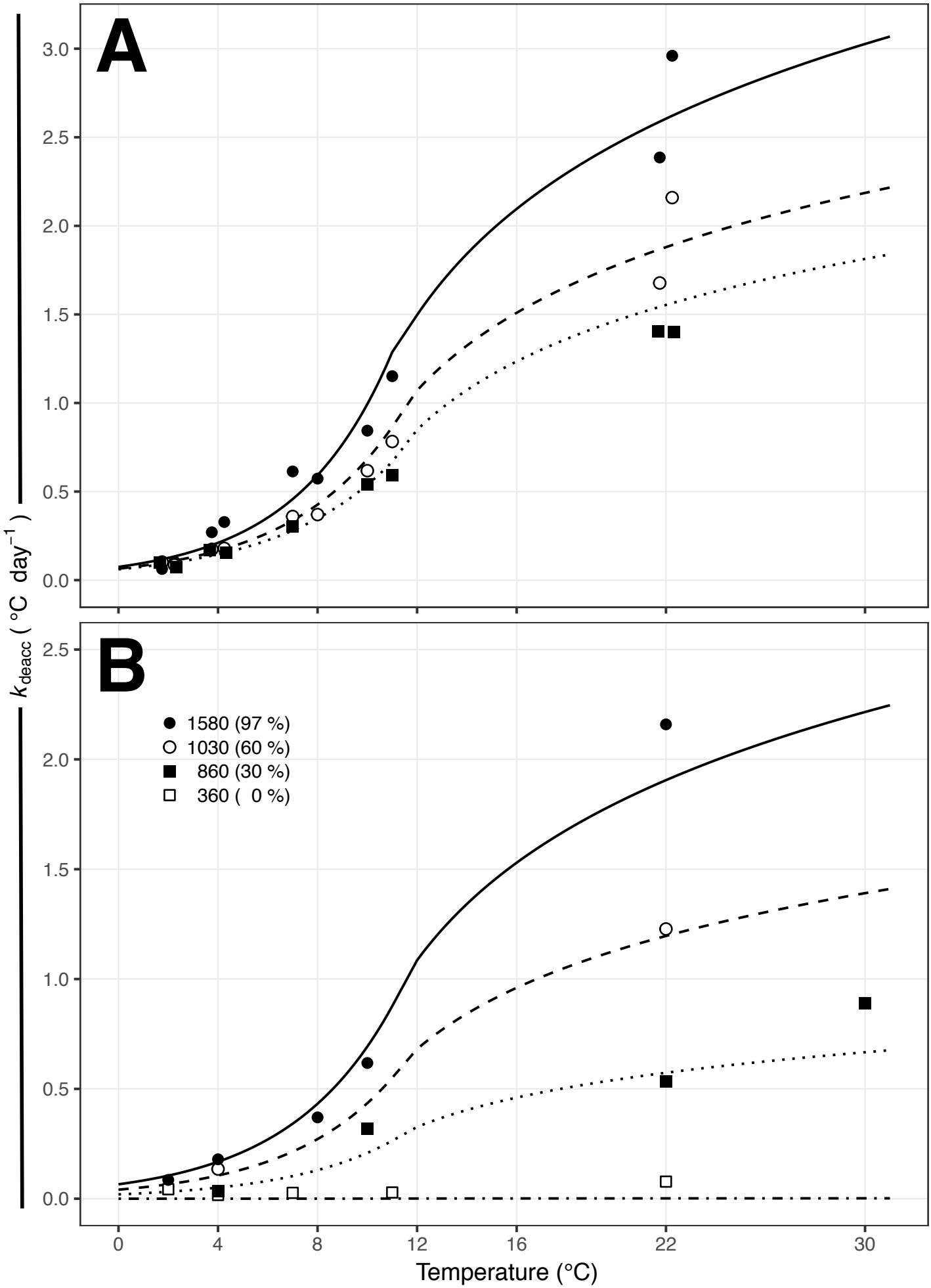
[c] $k_{\mathrm{deacc}} = q \times \ln(T - r)$, where *T* = temperature

[d] $\Delta_{LTE}(T, Chill) = \beta \times k_{\mathrm{deacc}} \times \Psi_{\mathrm{deacc}} \times t$, where *t* = time


4 °C
10 °C
22 °C
360
860
1580
LTE (°C)
Time (day)
0
−5
−10
−15
−20
−25
−30
0
10
20
30
40
50
60
70
80
90


4
22
100
75
50
25
0
$\psi_{deacc}$ (%)
0
400
800
1200
1600
Accumulated Chill


A
B
$k_{deacc}$ ( °C day$^{-1}$ )
Temperature (°C)
0
4
8
12
16
22
30
0
1
2
3
860
1440
1580
ln [ $k_{deacc}$ ( °C day$^{-1}$ ) ]
1000/T (K$^{-1}$)
3.30
3.39
3.46
3.51
3.56
3.61
3.66
1
0
−1
−2


A
B
1580 (97 %)
1030 (60 %)
860 (30 %)
360 ( 0 %)
$k_{deacc}$ ( °C day$^{-1}$ )
Temperature (°C)
0.0
0.5
1.0
1.5
2.0
2.5
3.0
0
4
8
12
16
22
30