Title: On the importance of scales when studying adaptive evolution

Running title: Time scale of bill length evolution

Authors: Charles PERRIER[1] & Anne CHARMANTIER[1]

Affiliations: [1]CEFE UMR 5175, CNRS, Université de Montpellier, Université Paul-Valery Montpellier,

EPHE 1919, route de Mende - 34293 Montpellier cedex 05 – France

Corresponding authors: Charles.perrier@cefe.cnrs.fr

Abstract:

Long-term individual field studies coupled with quantitative genomics offer a powerful means to

understand the genetic bases underlying quantitative traits and their evolutionary changes. A recent

investigation on the genomic architecture and spatio-temporal variation in great tit bill length

reports on a polygenic control of this trait, and suggests that UK great tits have evolved longer bills

than other birds in Europe, possibly because of their relatively recent use of bird feeders. We discuss

how this case study reveals the importance of considering different temporal scales, since an

alternative analysis leads to drastically different conclusions, with a significant decrease in bill length

over the last twenty years and an absence of clear trend over 150 years, most likely disqualifying the

feeder use as driver of an apparently old bill length variation. This study also illustrates the

cautionary steps that are required when interpreting the genomic architecture of quantitative trait

evolution. Especially, large coldspots of recombination most likely boosted large peaks of genetic

differentiation that should be considered carefully when searching for footprints of selection. This

study overall raises a stimulating challenge to unravel the environmental determinants and genomic

bases underlying short-term and long-term quantitative variation in bill length in great tits both in UK

and across Europe.

Main Text:

Longitudinal field studies have brought invaluable insight for the understanding of

evolutionary processes (Clutton-Brock & Sheldon 2010). Long-term studies notably allowed

characterizing the temporality of traits' variations and the strength and directionality of natural

selection underlying such variations. Some of these long-term examinations of key phenotypic traits

detected strikingly fast phenotypic variations, driven by prompt ecological changes (Grant & Grant

2006). In contrast, many long-term studies failed to reveal micro-evolutionary changes and response

to selection (Merila *et al.* 2001; Pujol *et al.* 2018). Hence, examining longitudinal data looking for

both long-term trends and embedded prompt anomalies has the potential to shed light on traits'

evolution and evolutionary trajectories in natural populations. Coupling long-term studies with

recent genomic developments is presently the most powerful way to understand the genetic bases

underlying evolutionary changes.

Avian bill morphology has played a prominent role in empirical studies of evolution and

natural selection (Lack 1947; Grant 1999), perhaps because the size and shape of bills show large

variations across and within bird species, and are shaped by strong selective forces since they

directly determine foraging efficiency on various food sources. For instance, the emblematic study of

Darwin's finches on the Galapagos island of Daphne Mayor aimed at capturing evolutionary changes

in bill size (Boag & Grant 1981; Grant & Grant 1993). From 1977 to 1978, bill size increased markedly

after a severe drought in 1977. This analysis clearly demonstrated that extreme climatic events such

as an El niño event are strong drivers of bill size evolution in this species. After 30 years of

perspective, however, Grant & Grant (2002) concluded that while evolution of beak length was

predictable as a rapid response to strong selection, it remained unpredictable on a slightly longer

microevolutionary scale. Using recent genomic tools, a handful of genes were found significantly

associated to bill size and shape in the medium ground finch *Geospiza fortis* (Lamichhaney et al.

2016), among which the strongest candidate gene has also been shown influencing bill dimensions in

the Darwin finches' entire radiation (Lamichhaney *et al.* 2015). These genomic analyses hence

cracked the oligogenic nature of this trait variation at both small and large microevolutionary scale.

In a recent study, Bosse et al. (2017) investigated the genetic architecture of bill length in

the Great tit *Parus major*, using a tremendous amount of data from long-term research programs in

Wytham woods and in Netherlands. Applying modern analyses of both population and quantitative

genomics using 500k single nucleotide polymorphisms, the authors provide insight into the

signatures of divergent selection in the studied populations and the genomic architecture of

variation in bill length. In line with the quantitative nature of variation in bill length and with

quantitative genetics theory (Lynch & Walsh 1998), the authors showed that the genetic architecture

of bill length was highly polygenic. Specifically, the authors showed, using a mixture analysis fitting

all the SNPs simultaneously, that 3009, or 0.8% of the SNPs, contributed significantly to bill length

variation, explaining collectively 31% of the phenotypic variation. Interestingly, none of these SNPs

reached genome wide significance in genome wide association tests (GWAS) with bill length,

revealing small effect sizes of individual variants. Accordingly with recent quantitative genomic

findings, the proportion of variance in bill length explained by each chromosome amazingly scaled

with its size, showing that the many SNPs additively explaining bill length are distributed throughout

the genome but not clustered in a single chromosome. The polygenic analysis also predicted the

difference in bill length observed between Wytham and Netherlands, further illustrating the

polygenic nature of this trait variation. This evidence for a polygenic control of bill length, with no

major effect SNP, is hence very different from previous examples of oligogenic architecture revealed

for example for Darwin finches' beak. This study might become a textbook case study of the

polygenic architecture of an illustrious quantitative trait in wild populations, uncovered by combining

ambitious long-term field monitoring with extensive genomic analyses in a polygenic framework.

Although such a polygenic investigation required these technical commitments and recent advances

(Wellenreuther & Hansson 2016; Gienapp *et al.* 2017a), it also requires a conceptual revolution

consisting in silencing our hope to find large effect alleles that are theoretically rarely responsible for

quantitative variation (Rockman 2011).

Bosse et al. (2017) however concluded on a different interpretation of their data, proposing

that variation at one gene was associated with use of feeders and that this might have driven the

evolution of larger bills in UK compared to Netherlands. Among the arguments pointing to feeders as

drivers of longer bill lengths in UK great tits, Bosse et al. (2017) reported that COL4A5, the locus with

the highest association with bill length in Wytham, was also highly differentiated between Wytham

and Netherlands, and was associated with greater reproductive success and higher activity at feeding

sites in Wytham. In addition, they reported that bills were longer in UK compared to mainland

Europe (Figure 4A in Bosse et al. 2017) and increased from 1982 to 2007 in UK (Figure 4B in Bosse et

al. 2017). This gene was located in one of several large genomic regions strikingly highly

differentiated between Wytham and Netherlands, overall containing genes that could make them

good candidates for bill morphology. We argue in this comment that this speculation on the role of

feeder on bill length evolution is not well supported by these arguments. We propose instead that

differences in bill length between Wytham and Netherlands might have been evolving on the longer

term than during the contemporary use of feeders and that the role of COL4A5, as well as other

highly differentiated regions, might be tested more comprehensively.

Bill length temporal variation might not be contemporary to the use of feeders. Indeed, we

show that the bill length trend is highly dependent on the time scale considered. Inspired by the

readings of papers such as the one of Grant & Grant (2006) relating the effects of rare and extreme

events on the evolution of phenotypic traits, we carefully inspected the evolution of bill length

during the studied period, looking for particularly prompt changes. We reanalyzed the data using a

breakpoint computations method (coin R-package) aiming at localizing such striking change. The best

cutpoint was found between 1986 and 1987 (maxT = 7.41, p-value < 2.2E-16), which corresponds to

a conspicuous change in bill length. Measures from the five years preceding 1987 differed from

subsequent years (t-test: t = -7.28, df = 674.53, p-value = 9.1E-13). Although a linear regression

though the entire period, from 1982 to 2007, as implemented by Bosse et al, yielded a positive slope,

removing the five measures prior to 1987, bill length significantly decreased from 1987 to 2007

(Figure 1A, Linear model from 1987 to 2007 taking into account or not tarsus length, F = 7.13, p-

value = 8.2E-4; F = 13.2, p-value: 2.9E-4, respectively). A LOESS model confirms the decrease in bill

length during the second part of the record. More generally, slopes of linear models linking bill

length to time, using several combinations of 10 to 25 consecutive years, were often positive when

including one or more data points collected between 1982 and 1987, and negative slopes were often

observed when excluding these years (Figure 2), further illustrating that both time periods yield

opposite patterns. Besides, tarsus length also decreased significantly both over the 1987-2007 and

the 1982-2007 periods (Figure 1B, F = 6.61, p-value = 0.01 & F = 24.07, p-value = 9.9E-07,

respectively). Correspondingly, the bill length / tarsus length ratio did not change significantly from

1987 to 2007 (Figure 1C, F = 1.51, p-value = 0.28), indicating no change in bill length, when scaled to

tarsus length, over this period. The origin of this cutpoint is important to investigate, since it could

constitute an evidence for a response, either genetic or plastic, of both bill and tarsus lengths to a

prompt abiotic or biotic factor or to a methodological change. Yet, it seems at present difficult to

decipher whether bill length or tarsus length or both, if any, were targeted by selection. Indeed, both

traits are commonly phenotypically and genetically positively correlated in passerines (Teplitsky *et*

al. 2014; Poissant *et al.* 2016), hence should often evolve together. Overall, these results much likely

rule out the possibility of a contemporary (1982-2007) positive effect of feeders on bill length.

One could nevertheless argue that bill length could have increased at a longer time scale

than the period 1982-2007, but still recent enough to incriminate the use of feeders. Figure 3

however shows the absence of a clear pattern of bill length increase in museum specimens collected

in UK. Although more data might be needed to confirm the pattern, it suggests that larger bill length

in the UK seems to have evolved on a larger time period than the one during which feeders have

been used. Moreover, bill length across Europe is not displaying a clear dichotomy between UK and

mainland Europe but rather smooth spatial variations (Figure 4) with an ANOVA showing a significant

effect of country (p = 3.75e-07) but not of sex (p = 0.345), with indeed UK birds having longer bills

compared to three other countries (France, Italy, and Netherlands). This spatial variation challenges,

once again, the supposition by Bosse et al.'s (2017) of a recent increase of bill length in British great

tits that would have been caused by feeders.

The gene COL4A5 was the corner stone linking bill length and feeders, via the association of

COL4A5 with bill length, activity at feeders, reproductive success, and divergent selection. However,

the association of COL4A5 was non-significant at the genome wide level. The authors acknowledged

that this locus probably has a small effect on the focal trait, which is in line with the demonstrated

polygenic control of bill length discussed above. The authors also propose that the small sample size

could limit the power of the analysis. Indeed, only 89 out of the initial 2322 individuals were used for

this GWAS analysis of quantitative variation of bill length. Furthermore, bill length had a relatively

low repeatability of 0.62, estimated over several thousands of individuals. Moreover, contrary to

Wytham, in Dutch birds the 200kb genomic regions surrounding COL4A5 did not yield significant p-

values in the GWAS with bill length, although the focal SNP was polymorphic in both popualtions

(data not shown). Collectively, although COL4A5 genomic region was linked to bird fitness and

displayed high Fst between Wytham and Netherlands, COL4A5 might only have a minor contribution

to the entire additive genetic variance of bill length.

SNPs in COL4A5 nevertheless showed elevated differentiation between Wytham and

Netherlands, which could be due to positive selection. However, COL4A5 was found in a relatively

large genomic region with very low SNP density (Figure 5), likely due to low recombination, which

might coincide with the centromere of the chromosome 4A. Importantly, although the authors did

not discuss this key fact, nearly all of the regions identified as highly differentiated (Figure 1A in

Bosse et al 2017) seem to coincide with regions of low SNP density (Supplementary figure 1), most

likely corresponding to low recombination regions hence with centromeres (Nordborg *et al.* 2009;

Limborg *et al.* 2015). Consistently, most of these regions were also identified by Laine et al. (2016) as

putative selective sweeps (Figure 2 in Laine et al. 2016), with reduced diversity and increased

divergence over large DNA regions, at the geographic scale of the species entire distribution,

suggesting a common origin of such large-scale patterns of differentiation among populations.

Specifically, the chromosome 4A region containing COL4A5 (Figure 1C in (Burri *et al.* 2015)) has

already been identified as showing elevated differentiation in several lineages and populations of

Ficedula flycatchers (Ellegren *et al.* 2012; Burri *et al.* 2015). Burri et al. (2015) determined that such

high differentiation shared across lineages was due to a reduction in effective population size of this

genomic region in the ancestral population, due to the effect of linked selection in interaction with

low recombination, since it displayed lower absolute divergence (dxy) and larger linkage

disequilibrium than the genomic background. Burri et al. concluded on a crucial warning: "scans are

likely to identify recombination-mediated elevations of differentiation not necessarily attributable to

selective sweeps". More broadly, variation in recombination rate and background selection have

received increasing attention since they are likely to shape larges-scale patterns of diversity and

differentiation along the genome and it has therefore been demonstrated that they should be

considered thoroughly when looking for footprints of divergent selection (Roesti *et al.* 2012; Berner

& Roesti 2017; Burri 2017; Comeron 2017; Delmore *et al.* 2018). Hence, the principal peaks of high

genetic differentiation identified in Bosse et al. 2017, matching the main areas of reduced diversity

displaying signatures of selective sweeps identified in Laine et al. 2016, may not necessarily illustrate

relatively recent divergent selection, but instead ancestral reduction of effective population size due

to low recombination, background selection and drift. Symmetrically to the risk of identifying false

positive loci in coldspots of recombination, it is also expected that neglecting the effects of variation

in recombination results in false negative loci localized outside of recombination coldspots (Berner &

Roesti 2017). Accordingly, both studies could have also missed candidate loci outside of these

recombination coldspots. Unfortunately, the large averaging sliding window (200kb) used in Bosse et

al. relatively to the average short distance linkage disequilibrium ($r2 = 0.1$ at 2kb) might have

worsened the problems raised above, by capturing even more principally large-scale areas of

recombination-elevated-differentiation. Correspondingly, this large window much likely increased

the risk of missing narrow peaks of elevated differentiation actually tagging candidate genes

targeted by relatively recent positive selection, and especially so outside of recombination coldspots.

In fact, such a large sliding window probably rather represents a neutral local baseline (Roesti 2012;

Burri 2017) that could have been implemented to detect local outlier markers showing locally

elevated divergence (e.g. supplementary figure 2). These caveats challenge the results and

conclusions of Bosse et al, questioning the potential role of divergent selection versus large-scale

variation in recombination and background selection in shaping the few large regions with elevated

differentiation. These considerations cast further doubt on the potential contemporary role of

feeders on bill length evolution.

Opportunely, this context and the available data offer exciting avenues of research to

unravel the determinants of both recent and long-term variation in bill length as well as adaptive

divergence between great tit populations across Europe. Spatial variation in bill length could be

unraveled in a multi-trait genome wide association test, using a polygenic framework, which could

be inspired from a pioneering study on great tit breeding phenology (Gienapp *et al.* 2017b). Besides,

a formal selection analysis relating great tit bill length to over-winter survival when birds are most

likely to benefit from food provided by people, is required, to elucidate the nature of the

evolutionary forces behind bill length variation. Finally, taking into account the effect of variation in

recombination rate and background selection along the genome to draw local neutral envelops of

genomic differentiation will likely help to validate or invalidate the potential footprints of divergent

selection identified almost only in coldspots of recombination, and to identify other footprints

outside of these regions (Roesti *et al.* 2012; 2013; Burri 2017; Comeron 2017; Delmore *et al.* 2018).

Providentially, the genomic analyses performed in other populations across Europe could also help

determining whether elevated population differentiation was driven by population-specific relatively

recent positive selection or by ancestral variation due to physical feature of the species genome (e.g.

Figure Box 3 in Burri 2017).

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References and Notes

Berner D, Roesti M (2017) Genomics of adaptive divergence with chromosome-scale heterogeneity
in crossover rate. *Molecular Ecology*, **26**, 6351–6369.
Boag PT, Grant PR (1981) Intense natural selection in a population of Darwin's finches (Geospizinae)
in the Galapagos. *Science*, **214**, 82–85.
Bosse M, Spurgin LG, Laine VN *et al.* (2017) Recent natural selection causes adaptive evolution of an
avian polygenic trait. *Science (New York, N.Y.)*, **358**, 365–368.
Burri R (2017) Interpreting differentiation landscapes in the light of long-term linked selection.
Evolution Letters, **1**, 118–131.
Burri R, Nater A, Kawakami T *et al.* (2015) Linked selection and recombination rate variation drive the
evolution of the genomic landscape of differentiation across the speciation continuum of
Ficedulaflycatchers. *Genome Research*, **25**, 1656–1665.
Clutton-Brock T, Sheldon BC (2010) Individuals and populations: the role of long-term, individual-
based studies of animals in ecology and evolutionary biology. *Trends in Ecology & Evolution*, **25**,
562–573.
Comeron JM (2017) Background selection as null hypothesis in population genomics: insights and
challenges from Drosophilastudies. *Philosophical Transactions of the Royal Society B: Biological*
Sciences, **372**, 20160471–13.
Delmore KE, Lugo Ramos JS, Van Doren BM *et al.* (2018) Comparative analysis examining patterns of
genomic differentiation across multiple episodes of population divergence in birds. *Evolution*

Letters, **2**, 76–87.

Ellegren H, Smeds L, Burri R *et al.* (2012) The genomic landscape of species divergence in Ficedula
flycatchers. *Nature*, **491**, 756–760.

Gienapp P, Fior S, Guillaume F *et al.* (2017a) Genomic Quantitative Genetics to Study Evolution in the
Wild. *Trends in Ecology & Evolution*, 1–12.

Gienapp P, Laine VN, Mateman AC, Van Oers K, Visser ME (2017b) Environment-Dependent
Genotype-Phenotype Associations in Avian Breeding Time. *Frontiers in Genetics*, **8**, 1392–9.

Grant BR, Grant PR (1993) Evolution of Darwin's Finches Caused by a Rare Climatic Event.
Proceedings of the Royal Society B-Biological Sciences, **251**, 111–117.

Grant PR (1999) *Ecology and evolution of Darwin's finches*. Princeton University Press.

Grant PR, Grant BR (2002) Unpredictable Evolution in a 30-Year Study of Darwin's Finches. *Science*,
296, 707–711.

Grant PR, Grant BR (2006) Evolution of character displacement in Darwin's finches. *Science (New
York, N.Y.)*, **313**, 224–226.

Lack D (1947) *Darwin's finches*. Cambridge University Press, London.

Laine VN, Gossmann TI, Schachtschneider KM *et al.* (2016) Evolutionary signals of selection on
cognition from the great tit genome and methylome. *Nature Communications*, **7**, 1–9.

Lamichhaney S, Berglund J, Almén MS *et al.* (2015) Evolution of Darwin's finches and their beaks
revealed by genome sequencing. 1–16.

Lamichhaney S, Han F, Berglund J, Wang C, 2016 A beak size locus in Darwin's finches facilitated
character displacement during a drought. *science.sciencemag.org*

Limborg MT, McKinney GJ, Seeb LW, Seeb JE (2015) Recombination patterns reveal information
about centromere location on linkage maps. *Molecular Ecology Resources*, **16**, 655–661.

Lynch M, Walsh B (1998) *Genetics and Analysis of Quantitative Traits*. Sinauer Associates
Incorporated.

Merila J, Sheldon BC, Kruuk LE (2001) Explaining stasis: microevolutionary studies in natural
populations. *Genetica*, **112-113**, 199–222.

Nordborg M, Charlesworth B, Charlesworth D (2009) The effect of recombination on background
selection. *Genetical Research*, **67**, 159–16.

Poissant J, Morrissey MB, Gosler AG, Slate J, Sheldon BC (2016) Multivariate selection and intersexual
genetic constraints in a wild bird population. *Journal of Evolutionary Biology*, **29**, 2022–2035.

Pujol B, Blanchet S, Charmantier A *et al.* (2018) The Missing Response to Selection in the Wild. *Trends
in Ecology & Evolution*, 1–10.

Rockman MV (2011) The QTN program and the alleles that matter for evolution: all that's gold does
not glitter. *Evolution*, **66**, 1–17.

Roesti M, Hendry AP, Salzburger W, Berner D (2012) Genome divergence during evolutionary
diversification as revealed in replicate lake–stream stickleback population pairs. *Molecular
Ecology*, **21**,   2852–2862.

Roesti M, Moser D, Berner D (2013) Recombination in the threespine stickleback genome-patterns
and consequences. *Molecular Ecology*, **22**, 3014–3027.

Teplitsky C, Tarka M, Møller AP *et al.* (2014) Assessing Multivariate Constraints to Evolution across
Ten Long-Term Avian Studies. **9**, e90444–15.

Wellenreuther M, Hansson B (2016) Detecting Polygenic Evolution: Problems, Pitfalls, and Promises.
Trends in Genetics, **32**, 155–164.

Figures

Figure 1. Decrease of mean bill length (A) and tarsus length (B) but conservation of the allometry

between both traits (C), from 1987 to 2007 (in red) in Wytham great tits. Regressions from 1982

to 2007 presented by Bosse et al. are shown in blue.

Figure 2. Slopes of linear models of bill length temporal evolution, including every possible

combination of 10 to 25 consecutive years of data, with or without one to several years of data

from 1982 to 1986.

288 Figure 3. Variation of mean bill length in UK from 1850 to 2007, based on museum records used

289 in Bosse et al. 2017.

Figure 4. Variation in bill length across European countries for both female (in blue) and male (in green) great tit specimens from museums. Letters A and B illustrate significant differences between sites and 'ns' refers to non-significant differences (Tukey HSD).

Figure 5. Fst at each SNP from chromosome 4A (blue dots), sliding window (red), and SNP

density over 200 Kb regions.

Supplementary figure 1: Number of SNPs per 200 Kb window in chromosomes 2, 3, 5, 6, 1A &

4A, in which main Fst and eigenGWAS peaks have been identified.

Supplementary figure 2: Illustration on chromosome 2, of the combination of variation in recombination rate and large scale Fst smoothing, on the discovery of loci with elevated Fst between Wytham and Netherlands. SNP density, raw Fst per SNP, large scale Fst smoothing, local Fst residuals from the large-scale smoothing, SNP Fst residuals from the large-scale smoothing. The centromere regions result in a locally elevated peak of divergence that is not found anymore when calculating the residuals between local or SNP Fst and the large scale

307   smoothing. In contrast, peaks in the middle of both chromosome arms likely correspond to true

308   Fst outliers.

309