

1 **Reaction of Hydrogen sulfide homeostasis genes under**
2 **biotic and abiotic stress condition in rice – computational**
3 **approach**

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11 **Abstract**

12 **Gaseous molecules are widespread signaling compounds, regulating the cell**
13 **development process in all major plant parts. For many decades, hydrogen sulfide**
14 **molecule is considered mainly for its deleterious effects on plant system. The increasing**
15 **recent experimental evidence and phenomenal concepts on H₂S molecule further**
16 **advance our understanding of H₂S interaction with plant tissues. In addition, the H₂S**
17 **messenger molecule is found to have positive effects on plant growth, in limited**
18 **condition, to maintain the balanced homeostasis. To meet the increasing demand, and**
19 **to sustain the crop yield, various crop improvement programs have been followed.**
20 **However, there is a concern that traditional plant improvement method and increasing**
21 **climate change has a negative impact on crop production. A major approach to**
22 **combating plant stress is to evaluate and explore the alternate source mechanism(s).**

23 **Towards this aim, it will be valuable to characterize the genes involved in H₂S**
24 **homeostasis in the staple food crop rice pan-genome. In this research, we identified 15**
25 **H₂S homeostasis genes in rice and used it for the ~3k rice pan-genome analysis to find**
26 **out the genetic relatedness based on single nucleotide polymorphism data.**
27 **Multidimensional scale plot of 15 H₂S homeostasis genes among the rice cultivars, and**
28 **RNA-seq experimental data analysis under various biotic and abiotic stress shows the**
29 **functional genes involved in biotic and abiotic stress. This study provides new insights**
30 **into plant stress management in crop breeding and suggests how H₂S gene(s) can be**
31 **utilized to improve the agronomic traits in rice and other food crops.**

32

33 **Keywords**

34 Hydrogen sulfide, gaseous homeostasis, plant stress, crop improvement, toxicity.

35

36 **Introduction**

37 Rice, maize, wheat and tapioca are important staple food crop across the world and these
38 crops faces many challenges to attain its full genetic potential. Further, the combinatorial
39 stress and standalone plant stress like salinity and drought are reducing the production and
40 productivity potential of almost every food crop (Suzuki et al., 2014). Research projects with
41 a focus on genetic and molecular analysis of unexplored functional genes will be very useful
42 in crop breeding to combat the plant stress. Hence, the plant stress research has witnessed an
43 increased attention, due to climate change, and continuous evolution of more virulent biotic
44 and abiotic stress factor.

45 H₂S homeostasis is an important source of mechanisms to tolerate the plant stress (Romero et
46 al., 2014; Mostofa et al., 2015b; Dai et al., 2017; Tain et al., 2017). Recently, Mostofa et al.
47 (2015b) reported the physiological implications of hydrogen sulfide in rice tissues under
48 salinity stress. They have analyzed and discussed the effect of exogenous application of H₂S
49 and its effect on plant growth, particularly by maintaining a Na⁺/K⁺ ratio. The physiological
50 functions of H₂S in plants are mediated by sulfur-oxidation pathways (Mishanina et al.,
51 2015), and different molecular targets, such as different ion channels, sulfate transporters and
52 signaling proteins (Wang, 2012). In plants, the exogenous/endogenous hydrogen sulfide is
53 responsible for conferring tolerance to both the biotic and abiotic stress (Bloem et al., 2004;
54 Shi et al., 2015; Mostofa et al., 2015b). In every growth stage, plants do produce H₂S in the
55 cytosol through enzymatic mechanisms, particularly desulfhydrases. However, the H₂S
56 molecule also has the lethal effect on plant tissues at higher concentrations. Therefore, it is
57 important to evaluate the potential of H₂S gene in applied aspects before using it in crop
58 breeding program. To explore the characteristic features and behavioral pattern of H₂S
59 homeostasis genes in biotic and abiotic stress, we have performed this study in rice H₂S
60 genes.

61
62 In this study, we focused to elucidate the role of H₂S homeostasis genes under biotic and
63 abiotic stress through combined computational genomics approach. Based on previous
64 physiological experimental evidence (Mostofa et al., 2015a; Chen et al., 2017; Duan et al.,
65 2015; Mostofa et al., 2015b), we have determined our gene identification and analysis criteria
66 to characterize the H₂S homeostasis genes in rice. The reason for selecting the genes from a
67 different category of hydrogen sulfide activity is mainly to maintain the balanced H₂S

68 content in plants. Since the higher H₂S content will lead to tissue toxicity in rice, it is better
69 to introgress, and/or clone the H₂S homeostasis genes.

70 Despite the availability of multiple completely sequenced rice genomes, little is known
71 on the occurrence of H₂S in rice. Cultivated rice does not have all agriculturally desirable
72 traits, which they might have lost during segmental, and/or tandem duplication events. As the
73 wild rice contains many desirable traits, it is important to mine alleles from wild rice. ~3k
74 rice genome project sequencing project made it possible to use the potential genes(s) from
75 wide range of rice germplasm. Here we present the pan-genome analysis of the H₂S
76 homeostasis genes extended across the largest part of *Oryza* phylogeny using sequencing
77 data from the 3k rice genome project.

78

79 **Materials and methods**

80 **Comparative analysis of H₂S homeostasis genes in rice**

81 The keywords viz., sulfite reductase, cysteine synthase, cyanoalanine synthase and cysteine
82 desulphydrase were searched in the Gramene and multiple rice database to retrieve the full-
83 length gene sequences. Redundant results were filtered out for further downstream analyses.

84 The retrieved gene sequences were manually annotated with FGENESH
85 (<http://www.softberry.com/>). The annotated sequences were cross-checked in the public
86 databases. The identified genes were positioned on their respective chromosome using the
87 Oryzabase database. Number of intron and exons in the H₂S genes were predicted in GSDS
88 (<http://gsds.cbi.pku.edu.cn/>). The FGENESH derived protein sequences were subjected to
89 conserved domain analysis in NCBI-CDD, Pfam identifiers, HMMER to confirm the
90 presence H₂S homeostasis domains. These protein sequences were used for Phylogenetic

91 classification of H₂S homeostasis genes through W-IO-TREE
92 (<http://iqtree.cibiv.univie.ac.at/>) server with default parameters. Protein topology and signal
93 peptides were predicted through Protter. To find the potential miRNA targets, the gene
94 sequences were scanned in psRNATarget (<http://plantgrn.noble.org/psRNATarget/>) server.
95 The association between the potential miRNA expression with agronomic traits was done in
96 RiceATM (<http://syslab3.nchu.edu.tw/rice/>). Rice pan-genome analysis was performed in the
97 rice SNP seek database (<http://snp-seek.irri.org/>) to determine the genetic distance of ~3k rice
98 genotypes based on H₂S genes. For promoter analysis, 2 kb upstream of all gene sequences
99 was subjected to transcription factor analysis in PlantPAN database. Whole genome RNA-
100 seq data were used to determine the expression of fifteen H₂S in various abiotic stress
101 phosphorus stress (PRJEB11899); drought stress and salinity stress (GSE60287). To compare
102 the H₂S gene expression level in biotic stress, bacterial blight (GSE57670) transcriptome data
103 were used to generate the FPKM value. The obtained FPKM value was used to generate the
104 heat map to quantify the transcript abundance.

105

106 **Results**

107 ***In silico* functional characterization of H₂S gene family**

108 To comprehensively investigate and characterize the H₂S gene family in rice, a genome-wide
109 survey covering the entire length of all the 12 rice chromosomes were performed. The hidden
110 Markov model and keyword-based and search in Gramene and rice genome annotation
111 database resulted in the identification of 15 potential full-length H₂S homeostasis related
112 genes. The H₂S homeostasis genes identified in the rice genome database and their features
113 are mentioned in (Table). The chromosomal localization analysis of rice H₂S genes revealed

114 variable distribution of the genes in all chromosomes except for chromosome 7, 8, 9, 10 and
115 11. While a maximum number of four genes were located on chromosome 1 and 6. In
116 contrast, only one gene was identified in both the chromosome number 4 and 12. Of fifteen
117 H₂S genes, three (OsH₂S12, OsH₂S13 and OsH₂S14) were defined as sulfite reductase, ten
118 (OsH₂S2, OsH₂S3, OsH₂S4, OsH₂S5, OsH₂S6, OsH₂S7, OsH₂S8, OsH₂S9, OsH₂S10 and
119 OsH₂S15) were defined as cysteine synthase and one as (OsH₂S1) cyanoalanine synthase and
120 another one as (OsH₂S11) L-cysteine desulfhydrase. The H₂S homeostasis family genes possess
121 a small number of introns in their sequences (Figure 2).

122 Based on the results of Pfam, HMMER, CDD and Phylogenetic classification of conserved
123 domain analysis, the H₂S homeostasis genes were independently grouped as sulfite reductase,
124 cysteine synthase, cyanoalanine synthase and cysteine desulfhydrase (Figure 1). Bzip, NAC,
125 WRKY, MYB and AP2/EREBP are the predominant stress related transcription factor
126 present in all the H₂S genes promoter sequence. Of 15 H₂S genes, only two genes (OsH₂S2
127 and OsH₂S14) had a potential miRNA binding site (Figure 1). In total, seven miRNA (osa-
128 miR818a, osa-miR818b, osa-miR818c, osa-miR818d, osa-miR818e, osa-miR1436 and osa-
129 miR2879) targeting two genes have been identified. The miRNA (osa-miR818a, osa-
130 miR818b, osa-miR818c and osa-miR818e) are found to strongly associated with the plant
131 height and 1000 seed grain weight (osa-miR818b and osa-miR818b). In the surveyed ~3k
132 rice accessions, 2757 accessions have all the 15 H₂S homeostasis genes. The genes OsH₂S3,
133 OsH₂S12 and OsH₂S15 had a number of allelic variations across the surveyed rice pan-
134 genome. The pan-genome analysis revealed a good genetic diversity analysis based on H₂S
135 sequences. This may help in selecting the donor plants with potential H₂S alleles in plant

136 breeding programs. More number of indica type rice possess all the 15 genes, while the
137 japonica rice has the second most number H₂S genes.

138 To check the expression level of all 15 genes under P stress, salinity & drought stress and
139 bacterial blight stress, the RNA-seq were analyzed (Figure 2). The differential expression
140 pattern of H₂S genes strongly supports the potential role of H₂S homeostasis genes under
141 various biotic and abiotic stress. For example, in IR 64 variety the cysteine synthase genes
142 (OsH₂S8 and OsH₂S9) were up-regulated under drought stress. In Pokkali OsH₂S1, OsH₂S6,
143 OsH₂S10, OsH₂S13 were up-regulated and OsH₂S4, OsH₂S5, OsH₂S12 and OsH₂S15 were
144 highly down-regulated under salinity stress. Under biotic stress condition (bacterial blight),
145 the genes OsH₂S1, OsH₂S3, OsH₂S5 and OsH₂S12 had a maximum level of expression.
146 While in P stress, the genes OsH₂S1, OsH₂S3, OsH₂S5, OsH₂S12 and OsH₂S15 had a
147 maximum expression and OsH₂S7 had a negligible/or low expression level. Among biotic
148 and abiotic stress condition, the H₂S genes expression is comparatively higher in P stress
149 condition. Hence the research should be focused more on characterizing the H₂S homeostasis
150 genes under various P treatments in rice.

151

152 **Conclusion**

153 In this paper, an in-depth *insilico* gene characterization of the H₂S family of rice was
154 performed. We identified and characterized 15 H₂S homeostasis genes in rice. The
155 phylogenetic grouping of protein sequences confirmed the presence of conserved domains in
156 the H₂S related gene family. In addition, the presence of H₂S gene family in seven
157 chromosomes reflects the unequal distribution in the rice genome. Analysis of the promoter
158 sequence, transcription factors and quantification of transcript abundance enabled the

159 retrieval of valuable information related to the functional response, diversity in the stress
160 responsive elements and biotic/abiotic stress responsiveness of these genes. Finally, a
161 comparative analysis between rice accessions revealed a high degree of sequence
162 conservation/variation within the H₂S domain as well as in the domain organization of these
163 genes. Furthermore, analysis of the expression profiles of the H₂S genes confirmed that they
164 are differentially regulated in response to several types of stress. These data suggest a
165 potential role for the H₂Ss in plant signaling and defense mechanisms.

166

167 **Future direction of research**

168 Interestingly, Neale et al., (2017) reported that H₂S signals produced from the plants have the
169 ability to alter pathogenicity of microbes. The interaction between H₂S genes and microbes,
170 whether the triggered plant H₂S genes are race-specific or race non-specific, and the genes
171 and/or QTL controlling the specificity are needs to be clearly addressed. Research describing
172 the reaction of plant H₂S genes with specific microbial receptor protein will reflect the
173 outcome of the interactions between alleles at all avirulence loci in the phyto-pathogen
174 and alleles at all H₂S loci of the plant gene. In addition, delineating the role of H₂S directed
175 regulation of abiotic stress responsive genes/QTLs/transcription factors will provide clues to
176 the mechanisms controlling H₂S homeostasis in plants. Further, the application of next-
177 generation sequencing techniques will explore the presence of genotype specific novel
178 INDEL region/SNPs in the H₂S genes in plants. Some of the H₂S genes have the signal
179 peptide and are predicted to be involved in the secretory pathway. It would be interesting to
180 see whether these signal peptides have any role in protein targeting and what happens if we
181 truncate the signal peptide. It will also be motivating to observe the localization pattern of

182 H₂S proteins. The current evidences on the role of signal peptides suggests that these proteins
183 are secreted in some other cellular components, and being transported to inter-cellular
184 spaces. Meanwhile, H₂S genes have the potential to interact with other stress related genes
185 (Wang, 2012). In addition, hydrogen sulfide has a positive effect on plant growth at various
186 stress condition (Chen et al., 2013; Christou et al., 2013; Li et al., 2012; Suzuki et al., 2014;
187 Wang et al., 2010; Wang et al., 2012; Zhang et al., 2010a; Zhang et al., 2010b; Zhang et al.,
188 2010c). Hence, the similarities of these longer genes should be better studied in order to
189 analyze their significance in altering plant tolerance to stress and/or other important
190 agronomic traits that may bring interesting insights about H₂S evolution or that may be of
191 interest of plant breeders.

192

193 **Author Contributions**

194 GA and JR initiated the project. All the authors have made a substantial, direct, intellectual
195 contribution to the work, and reviewed the final version of the manuscript.

196

197 **Conflict of Interest Statement**

198 The authors declare that the research was conducted in the absence of any commercial or
199 financial relationships that could be construed as a potential conflict of interest.

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204 expression patterns.

205 **References**

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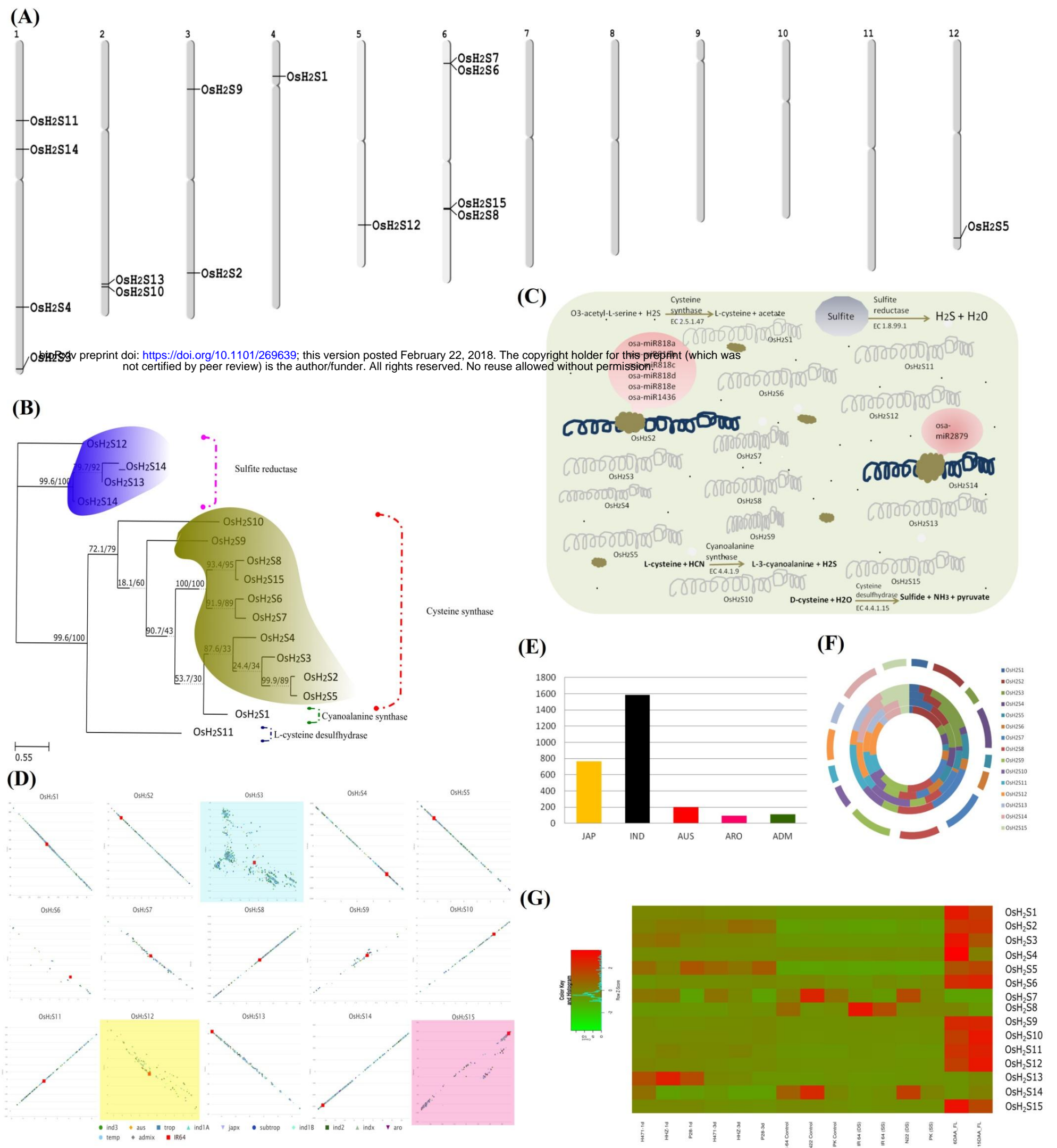


Figure 1: Genome-wide characterization of H₂S homeostasis genes in rice. (A) Chromosomal positioning and distribution of genes. H₂S genes were identified using a combined computational approaches in rice, i.e., key word search, conserved domain identification, Pfam identifier and HMMER search. Chromosomal positioning was based on the physical position (Mb) in 12 rice chromosomes. The chromosome number is indicated at the top of each chromosome. (B) The evolutionary history was inferred by using the Maximum Likelihood method based on W-IQ-TREE. The bootstrap consensus tree inferred from 1000 replicates is taken to represent the evolutionary history of the taxa analyzed (Felsenstein, 1985). The resulting four major clusters were shown in the figure. The analysis involved 15 amino acid sequences. All amino acid sequences in this study have been manually annotated in FGENESH to avoid the redundancy. (C) miRNA scanning and target prediction. The full-length nucleotide sequences were subjected to verify the presence of miRNA targets. Of 15 H₂S gene, only two gene sequence had a potential miRNA binding site. Interestingly, the gene OsH₂S2 has a single binding site for six marinas. (D) Pan-genome analysis in ~3k rice genome sequence data. The pan-genome analysis revealed the possible and potential H₂S alleles across wide rice accessions. The data in this study are obtained from Rice SNP seek database. The genetic relatedness among these accessions is drawn based on the variations in any particular H₂S gene. IR-64 variety is highlighted in gene based genetic diversity analysis (E) The distribution of all 15 H₂S homeostasis genes in wide rice accessions. The graph indicates the number of any particular rice type having all 15 H₂S. The results show that maximum number of *Indica* type rice possess all H₂S homeostasis genes. (F) The distribution of stress related transcription factors in the 2kb upstream nucleotide sequences. The five rings in the figure indicate the five transcription factor. From outer side, ring 1- Bzip, ring 2- NAC, ring 3- WRKY, ring 4- MYB and ring 5- AP2/EREBP. (G) Functional characterization of H₂S homeostasis genes. Heatmap showing the expression of H₂S genes under salinity and drought stress. The FPKM value is calculated from the RNA-seq data derived from the whole genome transcriptome study under salinity and drought stress. The colored bar at the left indicates the relative expression value, wherein, -2.0, 0.0 and 2.0 indicates low, medium and high expression respectively.

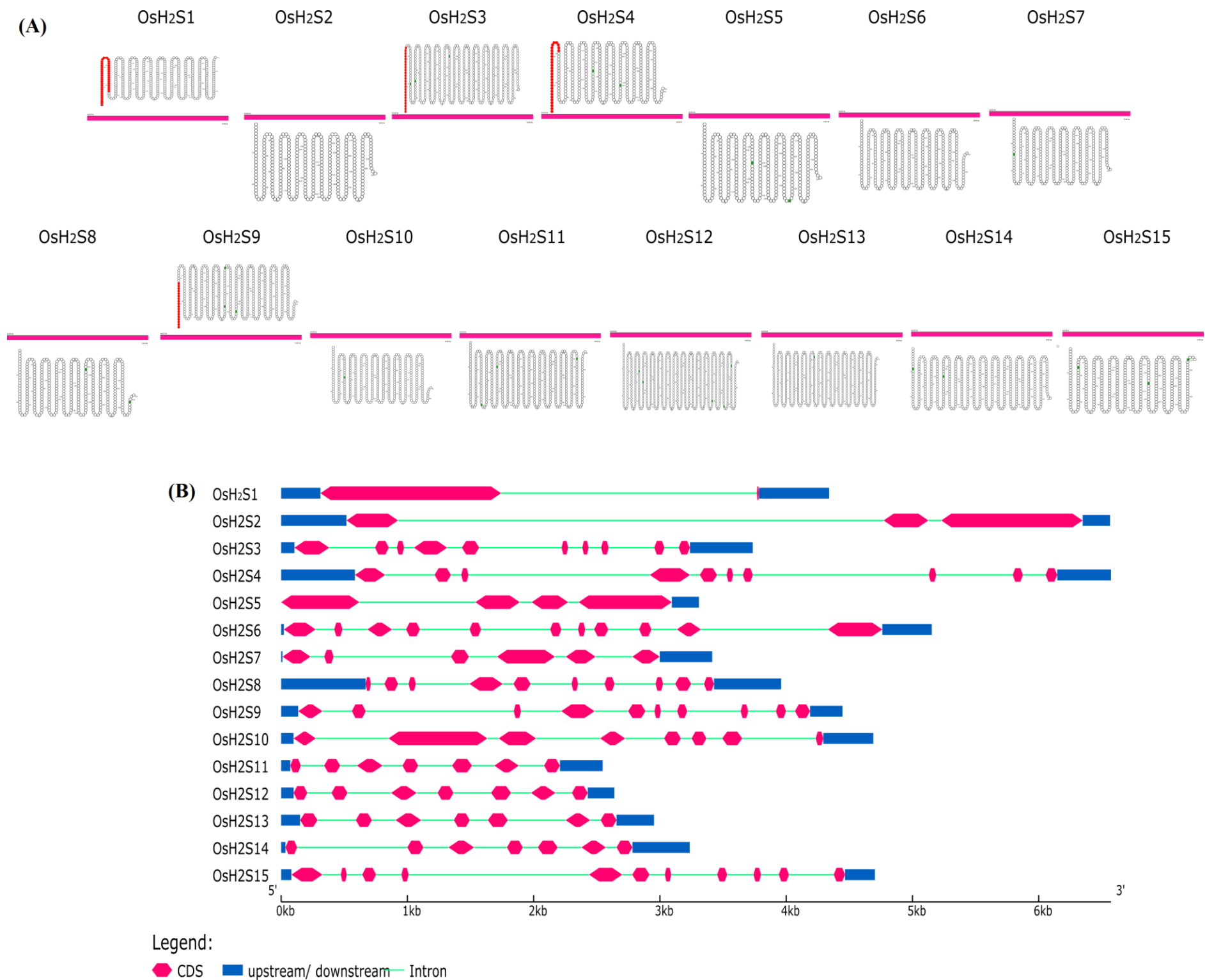


Figure 2: H₂S protein membrane topology and gene structure. (A) Signal peptide prediction and orientations of membrane-spanning segments with respect to the inner and outer sides of the plant cell plasma membrane. The pink colored peptide chain at the end of N-terminus indicates the presence of signal-peptide in the H₂S homeostasis gene (B) representation of the presence and arrangements of number of introns/exons in the genes.