All is not decline across global vertebrate populations

Gergana N. Daskalova[1]*, Isla H. Myers-Smith[1] and John L. Godlee[1]

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Author affiliation:

1. School of GeoSciences, University of Edinburgh, Edinburgh EH9 3FF, UK

gndaskalova@gmail.com

johngodlee@gmail.com

isla.myers-smith@ed.ac.uk

* Corresponding author:

Gergana N. Daskalova

gndaskalova@gmail.com

Crew Building, King's Buildings

Edinburgh EH9 1UU Scotland

Tel: +44 7946274522

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Abstract

The populations of Earth's species are changing over time in complex ways, creating a mixture

of winners and losers in a time of accelerating global change. A critical research challenge is to

test if there are specific biomes, taxa or types of species that are experiencing rapid alterations in

abundance over time. We conducted an analysis of the Living Planet Database using a state-

space modelling approach including nearly 10,000 abundance time-series from over 2,000

vertebrate species. We integrated the population abundance data with information on geographic

range, habitat preference, taxonomic and phylogenetic relationships, conservation status and

threats from occurrence, phylogenetic and conservation assessment data. We found that 15% of

populations were declining, 18% were increasing, and 67% showed no net changes in abundance

over time. Against a backdrop of no biogeographic and phylogenetic patterning in population

change, we uncovered a distinct taxonomic signal. Amphibians were the only taxa that

experienced net declines in the analyzed data, while birds, mammals and reptiles on average

became more abundant over time. The continuum of abundance changes over time was poorly

captured by species' rarity and global-scale threats. Capturing the full spectrum of abundance

change of species around the planet will inform conservation efforts and improve projections of

biodiversity and ecosystem function change under the accelerating influence of the

Anthropocene.

Main text

Alterations to ecosystems currently unfolding all around the globe are modifying the abundances

of the different species forming Earth's biota. As global change continues to accelerate[1,2], the

need for comprehensive assessments of the factors explaining the variety of ecological changes

observed across taxa and biomes is also growing[3]. However, existing empirical studies of the

predictors of population change mostly focus on either specific taxa[4] or on population declines

alone[2,5]. A critical research challenge is to disentangle the sources of heterogeneity across the

full spectrum of population change. Recent compilations of long-term population time-series,

extensive occurrence, phylogenetic, habitat preference and conservation status data[6–8] provide a

unique opportunity to quantify population trends and fluctuations among the world's well-

monitored vertebrate species and test which species- and population-level attributes explain

variation in population change. Population change is the underlying process leading to community

reassembly[9] and the resulting changes to biodiversity are vitally important for ecosystem functions

and services[10].

The distribution of global change drivers such as such as land-use change, habitat change,

pollution, invasion by non-native species and climate change[11–13] shows distinct clustering across

space. Thus, we predicted biogeographic patterning of population trends and fluctuations across

the planet's realms and biomes, in line with particular regions of the world experiencing high rates

of environmental change (e.g., tropical forests[14]). Spatial clustering has also been documented

for biodiversity trends derived from assemblage time-series, with the marine realm emerging as

a hotspot for rapid changes in community composition[15]. Since assemblages are made up of

populations, the biogeographic patterns at the assemblage level suggest similar clustering might

occur at the population level as well. In addition to geographic patterns in exposure to

anthropogenic activities, species' vulnerability can also influence population abundance over

time[16]. Species traits can moderate population responses to natural and anthropogenic

environmental change[16], and across evolutionary time, certain clades have been more vulnerable

to declines and extinctions[6–8]. Building on known variability in species' vulnerability[16–18], we

expected taxonomic and phylogenetic signals in population trends and fluctuations (e.g. greater

declines, increases or fluctuations in abundance for specific taxa and among specific clades).

Understanding which biomes and taxa are experiencing the most acute changes in abundance

over time could provide key insights for conservation prioritization.

Conservation efforts often focus on protecting rare species, those with restricted geographic

extents, small population sizes or high habitat specificity, as they are assumed to be more likely

to decline and ultimately go extinct[19–21]. Thus, we predicted that the populations of rare species

are more likely to decline and fluctuate than the populations of common species. Species with a

smaller geographic range might have more concentrated exposure to environmental change, with

less opportunities to find refugia or disperse, thus increasing the likelihood of declines[1,9]. As per

population dynamics theory[22,23] and Taylor's power law[24], species with small populations are more

likely to undergo stochastic fluctuations that could lead to pronounced declines, local extinction

and eventually global species extinction[5]. Small populations are also more likely to decline due to

inbreeding[25,26]. Allee effects, the relationship between individual fitness and population density,

further increase the likelihood of declines due to lack of potential mates and low reproductive

output once populations reach a critically low density[27,28]. Furthermore, a high habitat specificity

means that environmental change might have disproportionately large effects on the populations

of rare species, as their persistence and colonization of new areas are limited by strict habitat

preferences[1,29]. The fossil record indicates that on millennial time scales, rare species are more

likely to decline and ultimately go extinct[30], but human actions have pushed Earth away from

traditional geological trajectories[31], and the relationships between rarity and population change

across the planet have yet to be tested across the Anthropocene.

On a global scale, species are exposed to a variety of threats, among which habitat change,

resource exploitation and hunting dominate as key predictors of extinction risk across the planet's

species[32]. We predicted that the populations of species with more threatened conservation status

and more species-level threats are more likely to decline and fluctuate than the populations of

least concern species and those exposed to a smaller number of threats. Species' IUCN

conservation status levels are often used in conservation prioritization and threatened species

tend to be the focus of conservation initiatives, but on the local scale, there might be variation in

how species abundance is changing over time in different locations[4,33]. Testing population change

across species' IUCN conservation status allows us to link contemporary changes in abundance

with long-term probability of extinction[34]. Determining how local-scale population trends vary

across species' IUCN conservation status has practical applications for assessing species'

recovery which is useful for the proposed IUCN Green List of Species[35].

Here, we asked how the trends and fluctuations of vertebrate populations vary with biogeography,

taxa, phylogenetic relationships and across species' rarity metrics and IUCN conservation and

threat categories. We quantified differences in population trends and fluctuations across latitudes

and biomes within the freshwater, marine and terrestrial realms to test the presence of distinct

hotspots of declines and increases. Additionally, we assessed taxonomic and phylogenetic

signals in the patterns of population change to determine if specific groups experience

pronounced changes over time. We investigated whether the heterogeneity in population change

globally is explained by differences in species' rarity and IUCN conservation status. We measured

rarity using three separate metrics – geographic range, mean population size (number of animals

that were recorded by monitoring) and habitat specificity. In a *post-hoc* analysis, we categorized

each population based on the global IUCN threat categories to determine how the number and

type of threats influences local-scale population change. Using the largest currently available

compilation of population records over time, we conducted a global synthesis of population trends

and fluctuations to provide key empirical evidence for the management, conservation and

prediction of ecological changes across the Anthropocene.

We analyzed vertebrate population time-series from the Living Planet Database (133,092

records) covering the period between 1970 and 2014. These time-series represent repeated

monitoring surveys of the number of individuals in a given area (species' abundance over time),

hereafter called "populations". We focus on two aspects of population change – overall changes

in abundance over time (population trend) and abundance variability over time (population

fluctuations). In the first stage of our analyses, we quantified trends and fluctuations for each

population using state-space models that account for observation error and random fluctuations[36]

(Figure S1). In the second stage, we modelled the population trend and fluctuation estimates from

the first stage across latitude, realm, biome, taxa, rarity metrics, phylogenetic relatedness,

species' conservation status and threat type using a Bayesian modelling framework (Figure S2).

We included a species random intercept effect to account for the possible correlation between

the trends of populations from the same species (see table Table S1 for sample sizes). As

sensitivity analyses, we additionally used variance weighting of the population trend estimates (μ)

by the observation/measurement error around them (τ) and population trend estimates from linear

model fits (slopes instead of μ) as the input variables in the second stage models, as well as

several different fluctuations estimates. We also repeated our analyses on a single-country scale,

using only populations within the United Kingdom, where monitoring efforts are particularly

rigorous and extensive. All different analytical approaches yielded very similar results and are

described in further detail in the methods.

Results

Biogeographic patterns of population trends and fluctuations

Globally, population increases, declines and fluctuations over time occurred across all latitudes

and biomes within the freshwater, marine and terrestrial realms, with no strong biogeographic

patterning and no specific hotspots of population declines (Figure 1, Table S2). Across all time-

series, 15% of populations were declining, 18% were increasing, and 67% showed no net

changes in abundance over time. Across realms, monitored vertebrate populations experienced

net population increases (freshwater slope = 0.005, CI = 0.002 to 0.01; marine slope = 0.004, CI

= 0.002 to 0.01; terrestrial slope = 0.003, CI = 0.001 to 0.005, Figure 1d-e). In the freshwater and

terrestrial realms, there was a bimodal distribution of population trends (Hartigans' dip test, D =

0.04, p < 0.01). Across biomes, populations in Mediterranean forests, montane grasslands, polar

freshwaters, temperate wetlands, tropical forests and tropical coral biomes were more likely to

increase, whereas populations from the remaining studied biomes experienced no net changes

(Figure 1h, Table S2). Population fluctuations were less pronounced in the terrestrial realm (slope

= 0.02, CI = 0.018 to 0.021, Figure 1f-g), but those populations were also monitored for the longest

duration across systems (average duration – 28 years for terrestrial, 18 years for marine and 21

years for freshwater populations, Figure S1, Table S2).

Figure 1. Population declines, increases and fluctuations over time occur across all latitudes and biomes within the freshwater, marine and terrestrial realms. Results include 9286 populations from 2084 species. The lack of biogeographic patterning in vertebrate population trends was also apparent on a UK scale (Figure S6 and Table S2). The numbers in

the legend for plots **d-g** and on the x axis in plot **c** show the sample sizes for realms and biomes, respectively. The μ values of population trend (plots **a-b**, **d-e**, **h**) and the σ^2 values of population fluctuation (plots **c**, **f-g**) were derived from state-space model fits of changes in abundance over the monitoring duration for each population. Plots **d** and **f** show the density distribution of population trends across realms, the raw values (points) and boxplots with the mean, first and third quartiles and boxplot whiskers that indicate the distance that covers 1.5 times the interquartile range. Plots **e**, **g** and **h** show the effect sizes and the 95% credible intervals of population trends (**e**, **h**) across realms and biomes, and fluctuations across realms (**g**). For variation in fluctuations across biomes, see SI Figure S9. The three estimates for each realm/system in plots **e** and **h** refer to different analytical approaches: population trends calculated using linear models (circles), state-space models (μ, triangles), and population trends (μ) weighted by τ^2, the observation error estimate from the state-space models (squares). The five estimates in plot **g** refer to different analytical approaches, where the response variables in the models were: 1) the standard error around the slope estimates of the linear model fits of abundance versus year (circles), 2) half of the 95% confidence interval around the μ value of population change (triangles), 3) half of the 95% confidence interval around μ weighted by τ^2, (full squares), 4) the process noise (σ^2) from the state-space models, and 5) the standard deviation of the raw data for each population time-series (empty squares). Effect sizes were standardized by dividing the effect size by the standard deviation of the corresponding input data. The process noise is the total variance around the population trend minus the variance attributed to observation error. Error bars in plots **e**, **g** and **h** show 95% credible intervals. See Table S2 for model outputs.

Taxonomic and phylogenetic patterns of population trends and fluctuations

We found taxonomic, but not phylogenetic patterns, in population trends and fluctuations over time among nearly 10,000 populations from over 2,000 vertebrate species, with amphibians

emerging as the taxa experiencing pronounced declines (Figure 2, Table S2). Amphibians

experienced net declines over time (slope = -0.01, CI = -0.02 to -0.005), whereas birds, mammals

and reptiles experienced net increases (slope = 0.004, CI = 0.003 to 0.01; slope = 0.01, CI = 0.01

to 0.01; slope = 0.02, CI = 0.01 to 0.02), with birds having a bimodal trend distribution (Hartigans'

dip test, D = 0.04, p < 0.01, Figure 1a). Bony fish population trends were centered on zero (slope

= -0.001, CI = -0.004 to 0.002, Figure 1a-b) and sharks and rays showed net declines, but the

credible intervals overlapped zero (slope = -0.01, CI = -0.02 to 0.01). Fluctuations were most

common for amphibian populations (slope = 0.04, CI = 0.036 to 0.049, Figure 2d), which were

monitored for the shortest time period on average (11 years, Figure S1, Table S2). We did not

detect finer scale species-level phylogenetic clustering of population change (both trends and

fluctuations) within amphibian, bird and reptile classes (Figures 2 and S16, Table S4). Similarly,

species identity within amphibian, bird and reptile classes did not explain variation in population

trends or fluctuations (Figures 2 and S16, Table S4). There were no distinct clusters of specific

clades that were more likely undergo increases, decreases or fluctuations in population

abundance (Figure 2).

Figure 2. Population trends and fluctuations varied more among, rather than within, taxa, with amphibians being the only group showing pronounced declines over time. There were no distinct phylogenetic patterns in population trends and fluctuations (plots **e-j**). For details on phylogenetic models, see methods. Grey color in the heatmap in plot h shows species for which

no population trend data were available. The numbers in the legend for plots **a-d** show sample

size for each taxon. The μ values of population trend (plots **a-b**, **e-g**) and the σ^2 values of

population fluctuation (plots **c-d**, **h-j**) were derived from state-space model fits of changes in

abundance over the monitoring duration for each population. Plots **a** and **c** show the density

distribution of population trends across taxa, the raw values (points) and boxplots with the mean,

first and third quartiles and boxplot whiskers that indicate the distance that covers 1.5 times the

interquartile range. Plots **b** and **d** show the effect sizes and the 95% credible intervals of

population trends (**b**) and fluctuations (**d**) across the five studied taxa. The three estimates for

each metric in plot **b** refer to different analytical approaches: population trends calculated using

linear models (circles), state-space models (μ, triangles), and population trends (μ) weighted by

τ^2, the observation error estimate from the state-space models (squares). The five estimates in

plot **d** refer to different analytical approaches, where the response variables in the models were:

1) the standard error around the slope estimates of the linear model fits of abundance versus year

(circles), 2) half of the 95% confidence interval around the μ value of population change

(triangles), 3) half of the 95% confidence interval around μ weighted by τ^2, (full squares), 4) the

process noise (σ^2) from the state-space models, and 5) the standard deviation of the raw data for

each population time-series (empty squares). Effect sizes (plots **b** and **d**) were standardized by

dividing the effect size by the standard deviation of the corresponding input data. The process

noise is the total variance around the population trend minus the variance attributed to observation

error. Error bars in plots **b** and **d** show 95% credible intervals. See Tables S2 and S4 for model

outputs.

Population trends and fluctuations across rarity metrics, conservation status and threats

Species-level metrics, such as rarity and global conservation status, did not explain the

heterogeneity in trends of monitored populations in the Living Planet Database, and both rare and

common species experienced declines, increases and fluctuations in population abundance over

time (Figures 3 and 4). Across monitored vertebrate populations globally, species with smaller

ranges, smaller population sizes, or narrower habitat specificity (i.e., rare species) were not more

prone to population declines than common species (Figure 3, Table S2). Populations that

experienced more fluctuations had smaller mean population sizes on average (slope = -0.001, CI

= -0.001 to -0.001, Figure 3f). We found increasing, decreasing and stable populations across all

Red List categories of conservation status (Figure 4a). For example, a population of the Least

concern species red deer (*Cervus elaphus*) in Canada declined by 68% over seven years going

from 606 to 194 individuals and a population of the Critically-endangered Hawksbill sea turtle

(*Eretmochelys imbricate*) from Barbados increased by 269% over seven years going from 89 to

328 individuals. We found more fluctuations (Least concern: slope = 0.022, CI = 0.021 to 0.023;

Critically endangered: slope = 0.035, CI = 0.028 to 0.041), but not more population declines, with

increasing conservation status (Figure 4, Table S2). Populations from species that were exposed

to a larger number of threats on a global scale based on the species' IUCN threat categorizations

did not experience greater declines when compared to those categorized with a smaller number

of threats (Figure 4f). There were no distinct signatures of global species threats which were

associated with predominantly declining local trends of monitored populations (Figure 4e) and

there were increasing, decreasing and stable trends across all threat types.

Figure 3. Rarity metrics do not explain heterogeneity in local population trends, and both rare and common species experienced declines and increases over time, whereas smaller populations fluctuated more over time. Numbers on plots show sample size for each metric. Rarity metrics were calculated for all species for which information was available and cover all taxa represented in the Living Planet Database, with the exception of geographic range, which refers to the global range of only bird and mammal species in the global Living Planet Database (plots **a-e**). The μ values of population trend (plots **a-d**) and the σ^2 values of population fluctuation (plots **e-h**) were derived from state-space model fits of changes in abundance over the monitoring duration for each population. Plots **d** and **h** show the effect sizes and the 95% credible intervals of three rarity metrics on population trends (**d**) and fluctuations (**h**). The three estimates for each metric in plot **d** refer to different analytical approaches: population trends calculated using linear models (circles), state-space models (μ, triangles), and population trends (μ) weighted by τ^2, the observation error estimate from the state-space models (squares). The five estimates in plot **h** refer to different analytical approaches, where the response variables in the models were: 1) the standard error around the slope estimates of the linear model fits of abundance versus year

(circles), 2) half of the 95% confidence interval around the μ value of population change

(triangles), 3) half of the 95% confidence interval around μ weighted by τ^2, (full squares), 4) the

process noise (σ^2) from the state-space models, and 5) the standard deviation of the raw data for

each population time-series (empty squares). Effect sizes (plots **d** and **h**) were standardized by

dividing the effect size by the standard deviation of the corresponding input data. The process

noise is the total variance around the population trend minus the variance attributed to observation

error. Error bars in plots **d** and **h** show 95% credible intervals. Lines on **a-c** and **e-g** show model

fits and 95% credible intervals. See Table S2 for model outputs.

Figure 4. On local scales, there are increasing, decreasing and stable populations across

the full spectrum of the globally-determined species' conservation status and

anthropogenic threats (IUCN Red List Index). Numbers in the legend for plots **a-d** and in plots

e-f show sample size for each metric. Plots **a** and **c** show the density distribution of population

trends across Red List status, the raw values (points) and boxplots with the mean, first and third

quartiles and boxplot whiskers that indicate the distance that covers 1.5 times the interquartile

range. Plots **b** and **d** show the effect sizes and the 95% credible intervals of population trends (**b**)

and fluctuations (**d**) across Red List status categories. The μ values of population trend (plots **a**,

e-f) and the σ^2 values of population fluctuation (plots **c**) were derived from state-space model fits

of changes in abundance over the monitoring duration for each population. For the relationships

between type and number of threats and population fluctuations, see Figure S19. Plots **b** and **d**

show the effect sizes and the 95% credible intervals for population trends (**b**) and fluctuations (**d**)

across conservation status across the IUCN Red List categories. The three estimates for each

metric in plot **b** refer to different analytical approaches: population trends calculated using linear

models (circles), state-space models (μ, triangles), and population trends (μ) weighted by τ^2, the

observation error estimate from the state-space models (squares). The five estimates in plot **d**

refer to different analytical approaches, where the response variables in the models were: 1) the

standard error around the slope estimates of the linear model fits of abundance versus year

(circles), 2) half of the 95% confidence interval around the μ value of population change

(triangles), 3) half of the 95% confidence interval around μ weighted by τ^2, (full squares), 4) the

process noise (σ^2) from the state-space models, and 5) the standard deviation of the raw data for

each population time-series (empty squares). The process noise is the total variance around the

population trend minus the variance attributed to observation error. Effect sizes (plots **b** and **d**)

were standardized by dividing the effect size by the standard deviation of the corresponding input

data. Error bars in plots **b** and **d** show 95% credible intervals. Plot **e** shows the distributions of

population trends across different threats that the species face globally, with the central

tendencies of all distributions overlapping with zero. Lines in plot **f** show model fit and 95%

credible intervals, where "number of threats" refers to the number of different threats that each

species, whose populations are locally monitored, are exposed to on a global scale. See Methods

for details on deriving types of threats and Table S2 for model outputs.

Discussion

Taken together, our analysis of the Living Planet Database using a state-space modelling approach demonstrated ubiquitous alterations in vertebrate abundance of monitored populations over time across all biomes on Earth. We revealed that population change includes both increasing and decreasing populations and spans a wide spectrum of magnitudes, and while anthropogenic impacts have accelerated in recent decades, our results highlight that all is not declines across vertebrate species. Against a backdrop of no biogeographic patterning of population trends and fluctuations (Figure 1), we uncovered distinct taxonomic signals, with amphibians representing the only taxa that exhibited pronounced net declines, while birds, mammals and reptiles on average became more abundant over time (Figure 2). Within amphibian, bird and reptile taxa, there was no distinct phylogenetic clustering of closely-related species experiencing similar population trends or fluctuations (Figure 2). We found that both rare and common species experienced the full spectrum of population change, from declines to no net changes and increases. Species' geographic range, mean population size and habitat specificity did not explain variation in population trends, but species with smaller population sizes were nevertheless more likely to fluctuate, potentially increasing their extinction risk (Figure 3). There was no consistent pattern of greater population declines with increasing conservation status (Figure 4). On a global scale, the vertebrate species in the Living Planet Database are exposed to a variety of threats, but on more local scales, none of the threats were characterized by predominantly declining populations (Figure 4), testifying to the diverse ways in which populations are changing across the Anthropocene.

Contrary to our initial predictions, we did not find a distinct geographic patterning of population change around the world, nor a consistent trend of increasing declines in population abundance

with increasing threat level (Figures 1 and 4). Similar lack of biogeographic signal has been

documented in regional studies of population change from the Netherlands[33] and in temperate

North America and Europe[37]. Coarsely represented biogeographic regions and global-scale

species' IUCN status and threat categories might not capture the drivers acting in the locations of

the specific populations we studied[32,38–40]. Furthermore, the same driver can have opposing

effects on population abundance at different sites[41]. A lack of of biome-specific directional trends

in population change, despite a spatial clustering of human pressure around the world[12], can also

arise due to differences in species traits and vulnerability to environmental change within

biomes[16–18]. Accounting for divergent responses of species to global change is key when

translating global policy, such as the upcoming post-2020 planetary biodiversity strategy[42], into

conservation actions implemented on scales much finer than biogeographic realms.

Our results highlight variation in population change among taxa, with amphibians emerging as

the taxa experiencing the most pronounced declines in the Living Planet Database. The remaining

taxa showed either stable or increasing net changes in abundance over time (Figure 2). Such

taxonomic patterns could be driven by different taxon-specific factors including reproductive

strategy, trophic level, generation time and life history traits[43,44]. For amphibians, population

declines have been linked to the spread of a fungal disease (chytrid fungus, *Batrachochytrium*

dendrobatidis), facilitated by global warming[45], as well as habitat loss and Allee effects in small

populations[46]. Within bird, amphibian and reptile taxa, phylogenetic relatedness and species-level

taxonomic classification did not explain variation in population trends and fluctuations. A similar

lack of phylogenetic dependencies has been detected for the population growth rates of migratory

birds[47]. While phylogenetic clustering might be lacking in contemporary trends, there is evidence

that phylogenetic relatedness predicts extinction, a process occurring over much longer time

scales[6,7]. Over shorter time periods, species' traits and ability to persist, reproduce and disperse

in ever changing landscapes might be influencing local abundance[16], which has created a mix of

winners and losers across all taxa[48]. We demonstrate ongoing alterations in the abundances of

six vertebrate taxa which over time, may lead to shifts in community composition and ultimately

alter ecosystem function as some species become locally extinct whilst others become more

abundant[9,10].

Surprisingly, our results indicate that despite decades of conservation focus on rare species[19–21],

both rare and common species experienced declines and increases in population abundance over

the period of monitoring. The lack of rarity effects on population trends can be explained by theory

and empirical evidence demonstrating that small populations do not necessarily have a higher

likelihood of experiencing declines and some species are able to persist in small, but stable

populations[49]. The power of rarity metrics to predict population trends could also be mediated by

whether species are naturally rare, or have become rare due to external drivers in recent

years[50,51]. Naturally rare species might be more likely to persist over time, whereas species that

have more recently become rare might be more likely to decline in response to environmental

disturbance. Furthermore, the timing and magnitude of past and current disturbance events

influence population trends[41,52] and there could be temporal lags in both positive and negative

abundance changes over time[41,53]. However, disentangling the processes leading to rarity over

time remains challenging, and across the 2084 species we studied, there are likely cases of both

natural and human-driven population change. We found that species with small populations were,

nevertheless, more likely to fluctuate (Figure 3f), which may increase their probability of extinction,

a process that could play out over longer time-scales than found for most population monitoring

time-series[22,23,54]. Our results highlight that rarity metrics alone do not capture the heterogeneity

in local population change over time, and common species should not be overlooked in

conservation prioritization decisions as they could be as likely to decrease in abundance over

time as rare species.

Our finding that all is not declines for vertebrate populations monitored for longer than five years

contrasts with reports of the overall decline in the Living Planet Index (LPI)[55], a weighted summary

of population change across all abundance time-series in the Living Planet Database. The LPI

reports[55–57] also document that the numbers of declining and increasing species are similar across

this dataset but that the magnitude of declines is larger than the magnitude of increases. The

calculation of the index used involves differential weighting of population trends derived using

logged abundance data, geometric means and generalized additive models[58]. The LPI is

hierarchically averaged from populations to species, taxa and realm and is also weighted by the

estimated and relative number of species within biomes, which influences the direction and

magnitude of the index[58,59]. Our analysis explores the heterogeneity in local trends and

fluctuations of monitored species from the raw population abundance data, and thus, we did not

use an index with weightings and we did not aggregate population trends to a species-level. We

detected net population declines at local scales over time only in the amphibian taxa, in contrast

with the overall negative trend of the aggregate weightings of the LPI[55]. Our results were similar

when we weighted population trends by the state-space model derived observation error (Figures

1-4 and Tables S2-3). We caution that distilling the heterogeneity of local population change at

sites around the world into a simple metric may hide diverging trends at local scales, where we

found both increases and declines among species.

To attribute population trends and fluctuations to site-specific anthropogenic drivers, we need to

go beyond previous studies that have focused exclusively on declines and extinctions[5,60].

Attribution analyses statistically test the links between observed changes in ecosystems and the

experienced extrinsic pressure[3]. Through attribution studies encompassing the full spectrum of

population change, including positive, negative and stable trends[41,61], we can better understand

the variety of ways in which climate change, land-use change and other drivers are altering

biodiversity. For a subset of the bird populations in the Living Planet Database, higher

temperature warming corresponded with a higher likelihood of population declines over time[61],

which could be caused by worldwide and cross-biome phenological mismatches between

breeding and resource availability[62]. Across terrestrial species represented in the Living Planet

Database, peak forest loss was associated with accelerations in both population increases and

decreases in the period following habitat alteration[41]. The strength of documented relationships

between population alterations and global change could be influenced by how well-monitored

populations capture the full range of variation in driver intensity as well as whether monitoring

began during a population peak or a population trough[63]. Additionally, there is evidence from the

marine realm that when species are simultaneously exposed to multiple drivers, the resulting

biodiversity effects are antagonistic and could produce patterns of no net biodiversity changes[64].

The next critical step is to test how multiple global change drivers together[12] influence populations

across both terrestrial and marine realms and determine how these relationships are mediated

by species' traits and vulnerability to extrinsic threats[65].

In summary, our global analysis reveals the ubiquitous nature of population change over time

across monitored vertebrate species. We show that in a time of accelerating global change, there

were as many increases as there are decreases in population abundance over time. Among this

heterogeneity, we uncover a pronounced net decline in amphibian abundance as well as net

abundance increases for birds, mammals and reptiles. The taxonomic patterning of population

change highlights amphibians as a conservation priority, especially as their declines can have

further cascading effects across trophic levels within ecosystems. Rarity metrics, specifically

geographic range, mean population size and habitat specificity, as well as conservation status

and evolutionary history, did not explain the heterogeneity in population change across the data

analyzed in this study. Our findings caution the use of rarity metrics for the prediction of future

global population trends, but suggest that such metrics, in particular mean population size, are

nevertheless indicators of population fluctuations, which might ultimately link to increased species

extinction risk. On a global scale, both rare and common vertebrate species face numerous

threats due to resource exploitation and habitat change. As human activities continue to

accelerate, the next key step is to determine how intrinsic factors, such as rarity attributes and

threats, interact with extrinsic global change drivers and together influence the persistence of

Earth's biota. The ongoing alterations in the abundances of species around the world may lead

to shifts in community composition and ultimately alter ecosystem functions and services.

Methods

All data syntheses, visualization and statistical analyses were conducted using R version 3.5.1[66].

For conceptual diagrams of our workflow, see Figures S1 and S2.

Data sources

Population data

To quantify vertebrate population change (trends and fluctuations), we extracted the abundance

data for 9284 population time-series from 2084 species from the publicly available Living Planet

Database[67] (http://www.livingplanetindex.org/data_portal) that covered the period between 1970

and 2014 (Table S1). These time-series represent repeated monitoring surveys of the number of

individuals in a given area, hereafter called "populations". Monitoring duration differed among

populations (Figure S1, see SI Figures S6 and S7 for effects of monitoring duration on detected

trends). In our analysis, we retained only populations that had at least five time points of

monitoring data, as that is a considered a long enough period to detect a directional population

trend (if one exists) in 80-90% of cases[68]. Populations were monitored using different metrics of

abundance (e.g., population indices vs. number of individuals). Before analysis, we scaled the

abundance of each population to a common magnitude between zero and one to analyze within-

population relationships to prevent conflating within-population relationships and between-

population relationships[69]. This allowed us to explore trends among populations relative to the

variation experienced across each time series.

Phylogenetic data

We obtained phylogenies for amphibian species from https://vertlife.org[4], for bird species from

https://birdtree.org[8], and for reptile species from https://vertlife.org[6]. For each of the three classes

(*Amphibia*, *Aves* and *Reptilia*), we downloaded 100 trees and randomly chose 10 for analysis (30

trees in total). Species-level phylogenies for the classes *Actinopterygii* and *Mammalia* have not

yet been resolved with high confidence[70,71].

Rarity metrics, conservation status and threat data

We defined rarity following a simplified version of the 'seven forms of rarity' model[72], and thus

consider rarity to be the state in which species exist when they have a small geographic range,

low population size, or narrow habitat specificity. We combined publicly available data from three

sources: 1) population records for vertebrate species from the Living Planet Database to calculate

mean population size, 2) occurrence data from the Global Biodiversity Information Facility[73]

(https://www.gbif.org) and range data from BirdLife[74] (http://datazone.birdlife.org) to estimate

geographic range size, and 3) habitat specificity and Red List classification data for each species

from the International Union for Conservation[75] (https://www.iucnredlist.org). The populations in

the Living Planet Database[67] do not include species that have gone extinct on a global scale, and

we use species' Red List conservation status as a proxy for extinction risk[75], an approach used

recently in other studies[76]. We extracted the number and types of threats that each species is

exposed to globally from their respective IUCN profiles[75].

Quantifying population trends and fluctuations over time

In the first stage of our analysis, we used state-space models that model abundance (scaled to a

common magnitude between zero and one) over time to calculate the amount of overall

abundance change experienced by each population (μ, [36,77]). State-space models account for

process noise (σ^2) and observation error (τ^2) and thus deliver robust estimates of population

change when working with large datasets where records were collected using different

approaches, such as the Living Planet Database[37,78,79]. Previous studies have found that not

accounting for process noise and measurement error could lead to over-estimation of population

declines[80], but in our analyses, we found that population trends derived from state-space models

were similar to those derived from linear models. Positive μ values indicate population increase

and negative μ values indicate population decline. State-space models partition the variance in

abundance estimates into estimated process noise (σ^2) and observation or measurement error

(τ^2) and population trends (μ):

504
$$X_t = X_{t-1} + \mu + \varepsilon_t, (1)$$

505   where X_t and X_{t-1} are the scaled (observed) abundance estimates (between 0 and 1) in the present

506   and past year, with process noise represented by $\varepsilon_t \sim gaussian(0, \sigma^2)$. We included measurement

507   error following:

508
$$Y_t = X_t + F_t,$$

509   where Y_t is the estimate of the true (unobserved) population abundance with measurement error:

510
$$F_t \sim gaussian(0, \tau^2).$$

511 We substituted the estimate of population abundance (Y_t) into equation 1:

$$Y_t = X_{t-1} + \mu + \varepsilon_t + F_t.$$

513 Given $X_{t-1} = Y_{t-1} - F_{t-1}$, then:

$$Y_t = Y_{t-1} + \mu + \varepsilon_t + F_t - F_{t-1}.$$

For comparisons of different approaches to modelling population change, see *Comparison of*

modelling approaches section.

Quantifying rarity metrics

We tested how population change varied across rarity metrics – geographic range, mean

population size and habitat specificity – on two different but complementary scales. In the main

text, we presented the results of our global scale analyses, whereas in the SI we included the

results when using only populations from the UK – a country with high monitoring intensity, Thus,

we quantified rarity metrics for species monitoring globally and in the UK.

Geographic range

To estimate geographic range for bird species monitored globally, we extracted the area of

occurrence in km[2] for all bird species in the Living Planet Database that had records in the BirdLife

Data Zone[74]. For mammal species' geographic range, we used the PanTHERIA database[81]

(http://esapubs.org/archive/ecol/E090/184/). To estimate geographic range for bony fish, birds,

amphibians, mammals and reptiles monitored in the UK (see Table S5 for species list), we

calculated a km[2] occurrence area based on species occurrence data from GBIF[73]. Extracting and

filtering GBIF data and calculating range was computationally-intensive and occurrence data

availability was lower for certain species; thus, we did not estimate geographic range from GBIF

data for all species part of the Living Planet Database. Instead, we focused on analyzing range

effects for birds and mammals globally, as they are a very well-studied taxon and for species

monitored in the UK, a country with intensive and detailed biodiversity monitoring of vertebrate

species. We did not use IUCN range maps, as they were not available for all of our study species,

and previous studies using GBIF occurrences to estimate range have found a positive correlation

between GBIF-derived and IUCN-derived geographic ranges[82].

For the geographic ranges of species monitored in the UK, we calculated range extent using a

minimal convex hull approach based on GBIF[73] occurrence data. We filtered the GBIF data to

remove invalid records and outliers using the *CoordinateCleaner* package[83]. We excluded records

with no decimal places in the decimal latitude or longitude values, with equal latitude or longitude,

within a one-degree radius of the GBIF headquarters in Copenhagen, within 0.0001 degrees of

various biodiversity institutions and within 0.1 degrees of capital cities. For each species, we

excluded the lower 0.02 and upper 0.98 quantile intervals of the latitude and longitude records to

account for outlier points that are records from zoos or other non-wild populations. We drew a

convex hull to most parsimoniously encompass all remaining occurrence records using the *chull*

function, and we calculated the area of the resulting polygon using *areaPolygon* from the

geosphere package .

Mean size of monitored populations

We calculated mean size of the monitored population, referred to as population size, across the

monitoring duration using the raw abundance data, and we excluded populations which were not

monitored using population counts (i.e., we excluded indexes).

Habitat specificity

To create an index of habitat specificity, we extracted the number of distinct habitats a species

occupies based on the IUCN habitat category for each species, accessed through the package

rredlist[85]. We also quantified habitat specificity by surveying the number of breeding and non-

breeding habitats for each species from its online IUCN profile (the 'habitat and ecology' section).

The two approaches yielded similar results (Figure S3, Table S2, key for the profiling method is

presented in Table S4). We obtained global conservation status and threat data for all study

species through the IUCN Red List classification[75].

Testing the sources of variation in population trends and fluctuations

In the second stage of our analyses, we modelled the trend and fluctuation estimates from the

first stage across latitude, realm, biome, taxa, rarity metrics, phylogenetic relatedness, species'

conservation status and threat type using a Bayesian modelling framework through the package

MCMCglmm[86]. We included a species random intercept effect in the Bayesian models to account

for the possible correlation between the trends of populations from the same species (see Table

S1 for sample sizes). The models ran for 120 000 iterations with a thinning factor of ten and a

burn-in period of 20 000 iterations. We assessed model convergence by visually examining trace

plots. We used weakly informative priors for all coefficients (an inverse Wishart prior for the

variances and a normal prior for the fixed effects):

$$Pr(\mu) \sim N(0, 10^8)$$

$$Pr(\sigma^2) \sim Inverse\ Wishart\ (V = 0, nu = 0)$$

Population trends and fluctuations across latitude, biomes, realms and taxa

To investigate the geographic and taxonomic patterns of population trends and fluctuations, we

modelled population trends (μ) and population fluctuations (σ^2), derived from the first stage of our

analyses (state-space models), as a function of 1) latitude, 2) realm (freshwater, marine,

terrestrial), 3) biome (as defined by the 'biome' category in the Living Planet Database, e.g.,

'temperate broadleaf forest'[87] and 4) taxa (*Actinopterygii*, bony fish; *Elasmobranchii*, sharks and

rays; *Amphibia*, amphibians; *Aves*, birds; *Mammalia*, mammals; *Reptilia*, reptiles). We used

separate models for each variable, resulting in four models testing the sources of variation in

trends and four additional models focusing on fluctuations. Each categorical model from this

second stage of our analyses was fitted with a zero intercept to allow us to determine if net

population trends differed from zero for each of the categories under investigation. The model

structures for all models with a categorical fixed effect were identical with the exception of the

identity of the fixed effect, and below we describe the taxa model:

593
$$\mu_{i,j,k} = \beta_0 + \beta_{0j} + \beta_1 * taxa_{i,j,k},$$

594
$$y_{i,j,k} \sim gaussian(\mu_{i,j,k}, \sigma^2),$$

where $taxa_{i,j,k}$ is the taxa of the *ith* time-series from the *jth* species; β_0 and β_1 are the global

intercept (in categorical models, $\beta_0 = 1$) and the slope estimate for the categorical taxa effect (fixed

effect), β_{0j} is the species-level departure from β_0 (species-level random effect); $y_{i,j,k}$ is the estimate

for change in population abundance for the *ith* population time-series from the *jth* species from

the *kth* taxa.

Population trends and fluctuations across amphibian, bird and reptile phylogenies

To determine if there is a phylogenetic signal in the patterning of population change within

amphibian, bird and reptile taxa, we modelled population trends (μ) and fluctuations (σ^2) across

phylogenetic and species-level taxonomic relatedness. We conducted one model per taxa per

population change variable – trends or fluctuations using Bayesian linear mixed effects models

using the package *MCMCglmm*[86]. We included phylogeny and taxa as random effects. The

models did not include fixed effects. We assessed the magnitude of the random effects

(phylogeny and species) by inspecting their posterior distributions, with a distribution pushed up

against zero indicating lack of effect, since these distributions are always bounded by zero and

have only positive values. We used parameter-expanded priors, with a variance-covariance

structure that allows the slopes of population trend (the *mu* values from the first stage analysis

using state-space models) to covary for each random effect. The prior and model structure were

as follows:

614
$$Pr(\mu) \sim N(0, 10^8),$$

615
$$Pr(\sigma^2) \sim Inverse\ Wishart\ (V = 1, nu = 1),$$

616
$$\mu_{i,k,m} = \beta_0 + \beta_{0k} + \beta_{0m},$$

617
$$y_{i,k,m} \sim gaussian(\mu_{i,k,m}, \sigma^2),$$

where β_0 is the global intercept ($\beta_0 = 1$), β_{0l} is the phylogeny-level departure from β_0 (phylogeny

random effect); $y_{i,k,m}$ is the estimate for change in population abundance for the *ith* population

time-series for the *kth* species with the *mth* phylogenetic distance.

To account for phylogenetic uncertainty, for each class, we ran ten models with identical

structures but based on different randomly selected phylogenetic trees. We report the mean

estimates and their range for each class.

Population trends and fluctuations across rarity metrics

To test the influence of rarity metrics (geographic range, mean population size and habitat

specificity) on variation in population trends and fluctuations, we modelled population trends (μ)

and fluctuations (σ^2) across all rarity metrics. We conducted one Bayesian linear models per rarity

metric per model per scale (for both global and UK analyses) per population change variable –

trends or fluctuations. The response variable was population trend (μ values from state-space

models) or population fluctuation (*σ^2 values from state-space models*), and the fixed effects were

geographic range (log transformed), mean population size (log transformed) and habitat

specificity (number of distinct habitats occupied). The model structures were identical across the

different rarity metrics and below we outline the equations for population trends and geographic

range:

$$\mu_{i,k,n} = \beta_0 + \beta_{0k} + \beta_1 * geographic\ range_{i,k,n},$$

$$y_{i,k,n} \sim gaussian(\mu_{i,k,n}, \sigma^2),$$

where *geographic range$_{i,k,n}$* is the logged geographic range of the *kth* species in the *ith* time-

series; β_0 and β_1 are the global intercept and slope estimate for the geographic range effect (fixed

effect), β_{0j} is the species-level departure from β_0 (species-level random effect); $y_{i,k,n}$ is the

estimate for change in population abundance for the *ith* population time-series from the *jth*

species with the *nth* geographic range.

Population trends across species' IUCN conservation status

To investigate the relationship between population change and species' Red List status, we

modelled population trends (μ) and fluctuations (σ^2) as a function of Red List status (categorical

variable). We conducted one Bayesian linear model per population change metric per scale (for

both global and UK analyses). To test variation in population trends and fluctuations across the

types and number of threats to which species are exposed, we conducted a post-hoc analysis of

trends and fluctuations across threat type (categorical effect, determined by species IUCN threat

categorization) and number of threats that each species is exposed to across its range (in

separate models). The model structures were identical to those presented above, except for the

fixed effect which was a categorical IUCN conservation status variable. The analytical workflow

of our analyses is summarized in conceptual diagrams (Figures S1 and S2) and all code is

available on GitHub (link to public repository to be added at time of publication).

Data limitations

Taxonomic and geographic gaps

Our analysis is based on 9286 monitored populations from 2084 species from the largest currently

available public database of population time-series, the Living Planet Database[67]. Nevertheless,

the data are characterized by both taxonomic and geographic gaps that can influence our findings.

For example, there are very few population records from the Amazon and Siberia (Figure 1b) –

two regions currently undergoing rapid environmental changes due to land-use change and

climate change respectively. Additionally, birds represent 63% of all population time-series in the

Living Planet Database, whilst taxa such as amphibians and sharks where we find declines are

included with fewer records (Figures 2 and S5). On a larger scale, the Living Planet Database

under-represents populations outside of Europe and North America and over-represents common

and well-studied species[59]. We found that for the populations and species represented by current

monitoring, rarity does not explain variation in population trends, but we note that the relationship

between population change and rarity metrics could differ for highly endemic specialist species

that are not included in the Living Planet Database. As ongoing and future monitoring begins to

fill in the taxonomic and geographic gaps in existing datasets, we will be able to re-assess and

test the generality of the patterns of population change across biomes, taxa, phylogenies, species

traits and threats.

Monitoring extent and survey techniques

The Living Planet Database combines population time-series where survey methods were

consistent within time-series but varied among time-series. Thus, among populations, abundance

was measured using different units and over varying spatial extents. There are no estimates of

error around the raw population abundance values available and detection probability likely varies

among species. We used state-space models to estimate trends and fluctuations to account for

these limitations as this modelling framework is particularly appropriate for analyses of data

collected using disparate methods[37,78,79]. Because the precise coordinates of the polygons where

the individual populations were monitored are not available, we were not able to test for the

potential confounding effect of monitoring extent, but our sensitivity analysis indicated that survey

units do not explain variation in the detected trends (Figure S13).

Temporal gaps

The population time-series we studied cover the period between 1970 and 2014, with both

duration of monitoring and the frequency of surveys varying across time-series. We only included

populations with more than five survey points in time in our analyses, because this amount of data

has been found to be enough to detect directional trends in 80-90% of cases[68]. In a separate

analysis, we found significant lags in population change following disturbances (forest loss) and

that population monitoring often begins decades to centuries after peak forest loss has occurred

at a given site[41]. The findings of this related study suggest that the temporal span of the population

monitoring does not always capture the period of intense environmental change and lags suggest

that there might be abundance changes that have not yet manifested themselves. Thus, the

detected trends and the baseline across which trends are compared might be influenced by when

monitoring takes place and at what temporal frequency. Nevertheless, the Living Planet Database

represents the most comprehensive compilation of temporal population records to date, allowing

for the most comprehensive analyses possible into the patterns of vertebrate trends and

fluctuations around the world.

Challenges in estimating geographic range

Estimating geographic range across taxa, specifically species that are not birds or mammals,

remains challenging due to data limitations. We used a static measure of geographic range, which

does not account for changes in species distributions over time. Furthermore, species could

naturally have a small range or the small range size could be due to historic habitat loss[88]. The

UK populations included in the Living Planet Database are predominantly from species with wide

geographic ranges (Table S3), and our global scale analysis of the relationship between

population change and geographic range is based on mammal and bird data. As data availability

improves, future research will allow us to test the effect of geographic range on the trends of other

taxa, such as amphibians and sharks.

Sensitivity analyses

Monitoring duration, sampling methods and site-selection bias

To assess the influence of monitoring duration on population trends, we used a Bayesian linear

model (Figures S6). We modelled population trend (*mu*) as a function of monitoring duration

(years) for each population, fitted with a zero intercept, as when duration is zero, no population

change has occurred. Monitoring duration was weakly positively related to vertebrate population

trends, with slightly greater population increases found for longer duration studies (Figure S9,

Table S2). Variation in population trends was not explained by sampling method across the five

most commonly used abundance metrics (population index, number of individuals, number of

pairs, number of nests and population estimate, Figure S13). Following Fournier *et al.* 2019[63], we

tested the time-series that we analyzed for site-selection bias. Removing the first five survey

points reduces the bias stemming from starting population surveys at points when individual

density is high, whereas removing the last five years reduces the bias of starting surveys when

species are very rare. The distribution of population trend values across time-series was not

sensitive to the omission of the first five (left-truncation) or the last five years (right-truncation) of

population records (Figure S8). Overall, our sensitivity analyses suggest that our findings are

robust to the potential confounding effects of differences in monitoring duration, sampling method

and site-selection.

Comparison of modelling approaches

We conducted the following supplementary analyses: in the second-stage Bayesian models

estimating population trends across systems, biomes, taxa and rarity metrics, 1) we weighed *mu*

values by the square of *tau*, the observation error estimate derived from the state-space models[36],

2) we used slopes of linear model fits of abundance (scaled at the population level, centered on

zero and with a standard deviation of one)[69] instead of the *mu* estimates from state-space models,

3) we modelled the standard error around the slope values of the linear models, the error around

mu (half of the 95% confidence interval) and the standard deviation of the raw population data for

each time-series as additional metrics of population variability. To allow comparison, we scaled

the different metrics of population variability to be centered on zero and with a standard deviation

of one before they were used as response variables in models. All different analytical approaches

yielded very similar results (see main text and Figures S6, S7 and S10, Table S2).

Data availability

Code for all data processing and analyses and summary datasets are publicly available on GitHub

(link to be added at time of publication). Raw data are available from the following websites: for

population time-series[67] - http://www.livingplanetindex.org/data_portal, GBIF occurrences[73] -

https://www.gbif.org, bird geographic ranges[74] - http://datazone.birdlife.org, mammal geographic

ranges[81] - http://esapubs.org/archive/ecol/E090/184/, species' habitat preferences, threats and

conservation status[75] - https://www.iucnredlist.org, and phylogenies[6–8] - https://vertlife.org and

https://birdtree.org.

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Author contributions

GND and IMS conceived the idea and conducted the statistical analyses. JLG contributed to the

calculation of geographic range estimates. All authors contributed to the integration of the LPI,

GBIF and IUCN databases, which GND led. GND created all figures with input from IMS. GND

wrote the first draft of the manuscript and all authors contributed to writing. IMS supervised the

research as a senior author.

Competing interests

The authors declare no competing interests.

References

1. Ehrlén, J. & Morris, W. F. Predicting changes in the distribution and abundance of species under environmental change. *Ecol. Lett.* **18**, 303–314 (2015).

2. Hefley, T. J., Hooten, M. B., Drake, J. M., Russell, R. E. & Walsh, D. P. When can the cause of a population decline be determined? *Ecol. Lett.* **19**, 1353–1362 (2016).

3. IPBES. Summary for policymakers of the regional assessment report on biodiversity and ecosystem services for Europe and Central Asia of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services. M. Fischer, M. Rounsevell, A. Torre-Marin Rando, A. Mader, A. Church, M. Elbakidze, V. Elias, T. Hahn. P.A. Harrison, J. Hauck, B. Martín-López, I. Ring, C. Sandström, I. Sousa Pinto, P. Visconti and N.E Zimmermann (eds.). IPBES secretariat, Bonn, Germany. (2018).

4. Gilroy, J. J., Gill, J. A., Butchart, S. H. M., Jones, V. R. & Franco, A. M. A. Migratory diversity predicts population declines in birds. *Ecol. Lett.* **19**, 308–317 (2016).

5. Ceballos, G., Ehrlich, P. R. & Dirzo, R. Biological annihilation via the ongoing sixth mass extinction signaled by vertebrate population losses and declines. *Proc. Natl. Acad. Sci.* 201704949 (2017). doi:10.1073/pnas.1704949114

6. Tonini, J. F. R., Beard, K. H., Ferreira, R. B., Jetz, W. & Pyron, R. A. Fully-sampled phylogenies of squamates reveal evolutionary patterns in threat status. *Biol. Conserv.* **204**, 23–31 (2016).

7. Jetz, W. & Pyron, R. A. The interplay of past diversification and evolutionary isolation with

present imperilment across the amphibian tree of life. *Nat. Ecol. Evol.* (2018).

doi:10.1038/s41559-018-0515-5

8. Jetz, W., Thomas, G. H., Joy, J. B., Hartmann, K. & Mooers, A. O. The global diversity of

birds in space and time. *Nature* **491**, 444–448 (2012).

9. Batt, R. D., Morley, J. W., Selden, R. L., Tingley, M. W. & Pinsky, M. L. Gradual changes in

range size accompany long-term trends in species richness. *Ecol. Lett.* **20**, 1148–1157

(2017).

10. Oliver, T. H. *et al.* Biodiversity and Resilience of Ecosystem Functions. *Trends Ecol. Evol.*

30, 673–684 (2015).

11. Hansen, M. C. *et al.* High-resolution global maps of 21st-century forest cover change.

Science **342**, 850–853 (2013).

12. Bowler, D. *et al.* The geography of the Anthropocene differs between the land and the sea.

(2018). doi:10.1101/432880

13. Halpern, B. S. *et al.* Spatial and temporal changes in cumulative human impacts on the

world's ocean. *Nat. Commun.* **6**, (2015).

14. Barlow, J. *et al.* Quantifying the biodiversity value of tropical primary, secondary, and

plantation forests. *Proc. Natl. Acad. Sci.* **104**, 18555–18560 (2007).

15. Blowes, S., Supp, S. R. *et al.* Biodiversity trends are stronger in marine than terrestrial

assemblages. (2018). doi:10.1101/457424

16. Isaac, N. J. B. & Cowlishaw, G. How species respond to multiple extinction threats. *Proc. R.*

Soc. Lond. B Biol. Sci. **271**, 1135–1141 (2004).

17. Khaliq, I., Hof, C., Prinzinger, R., Bohning-Gaese, K. & Pfenninger, M. Global variation in

thermal tolerances and vulnerability of endotherms to climate change. *Proc. R. Soc. B Biol.*

Sci. **281**, 20141097–20141097 (2014).

18. Morrison, L., Estrada, A. & Early, R. Species traits suggest European mammals facing the

greatest climate change are also least able to colonize new locations. *Divers. Distrib.* **24**,

1321–1332 (2018).

19. Pigott, C. D. & Walters, S. M. The scientific basis of practical conservation: aims and

methods of conservation. in *Proc. R. Soc. Lond. B* **197**, 59–68 (The Royal Society, 1977).

20. Longton, R. E. & Hedderson, T. A. What are rare species and why conserve them?

Lindbergia **25**, 53–61 (2000).

21. Gaston, K. & Fuller, R. Commonness, population depletion and conservation biology.

Trends Ecol. Evol. **23**, 14–19 (2008).

22. Lande, R. Risks of population extinction from demographic and environmental stochasticity

and random catastrophes. *Am. Nat.* **142**, 911–927 (1993).

23. Melbourne, B. A. & Hastings, A. Extinction risk depends strongly on factors contributing to

stochasticity. *Nature* **454**, 100–103 (2008).

24. Kilpatrick, A. M. & Ives, A. R. Species interactions can explain Taylor's power law for

ecological time series. *Nature* **422**, 65–68 (2003).

25. Kareiva, P. Population dynamics in spatially complex environments: theory and data. *Phil*

Trans R Soc Lond B **330**, 175–190 (1990).

26. Hanski, I. Metapopulation dynamics. *Nature* **396**, 41 (1998).

27. Dennis, B., Assas, L., Elaydi, S., Kwessi, E. & Livadiotis, G. Allee effects and resilience in

stochastic populations. *Theor. Ecol.* **9**, 323–335 (2016).

28. Sun, G.-Q. Mathematical modeling of population dynamics with Allee effect. *Nonlinear Dyn.*

85, 1–12 (2016).

29. Bowler, D. E., Heldbjerg, H., Fox, A. D., O'Hara, R. B. & Böhning-Gaese, K. Disentangling

the effects of multiple environmental drivers on population changes within communities. *J.*

Anim. Ecol. **87**, 1034–1045 (2018).

30. Harnik, P. G., Simpson, C. & Payne, J. L. Long-term differences in extinction risk among the

seven forms of rarity. *Proc. R. Soc. B Biol. Sci.* **279**, 4969–4976 (2012).

31. Steffen, W., Crutzen, P. J. & McNeill, J. R. The Anthropocene: Are Humans Now

Overwhelming the Great Forces of Nature. *AMBIO J. Hum. Environ.* **36**, 614–621 (2007).

32. Maxwell, S. L., Fuller, R. A., Brooks, T. M. & Watson, J. E. Biodiversity: The ravages of

guns, nets and bulldozers. *Nature* **536**, 143–145 (2016).

33. van Strien, A. J. *et al.* Modest recovery of biodiversity in a western European country: The

Living Planet Index for the Netherlands. *Biol. Conserv.* **200**, 44–50 (2016).

34. Mace, G. M. *et al.* Quantification of Extinction Risk: IUCN's System for Classifying

Threatened Species. *Conserv. Biol.* **22**, 1424–1442 (2008).

35. Akçakaya, H. R. *et al.* Quantifying species recovery and conservation success to develop an

IUCN Green List of Species: Species Recovery and Conservation Success. *Conserv. Biol.*

32, 1128–1138 (2018).

36. Humbert, J.-Y., Scott Mills, L., Horne, J. S. & Dennis, B. A better way to estimate population

trends. *Oikos* **118**, 1940–1946 (2009).

37. Leung, B., Greenberg, D. A. & Green, D. M. Trends in mean growth and stability in

temperate vertebrate populations. *Divers. Distrib.* **23**, 1372–1380 (2017).

38. Fritz, S. A., Bininda-Emonds, O. R. P. & Purvis, A. Geographical variation in predictors of

mammalian extinction risk: big is bad, but only in the tropics. *Ecol. Lett.* **12**, 538–549 (2009).

39. Brook, B. W., Traill, L. W. & Bradshaw, C. J. A. Minimum viable population sizes and global

extinction risk are unrelated: MVP is unrelated to global extinction risk. *Ecol. Lett.* **9**, 375–

382 (2006).

40. Pereira, H. M., Navarro, L. M. & Martins, I. S. Global Biodiversity Change: The Bad, the

Good, and the Unknown. *Annu. Rev. Environ. Resour.* **37**, 25–50 (2012).

41. Daskalova, G. N. *et al.* Forest loss as a catalyst of population and biodiversity change:

bioRxiv (2018). doi:10.1101/473645

42. CBD. Strategic plan for biodiversity 2011–2020 and the Aichi targets. In Report of the Tenth

Meeting of the Conference of the Parties to the Convention on Biological Diversity. (2010).

43. Purvis, A., Gittleman, J. L., Cowlishaw, G. & Mace, G. M. Predicting extinction risk in

declining species. *Proc. R. Soc. B Biol. Sci.* **267**, 1947–1952 (2000).

44. Cardillo, M. *et al.* Human Population Density and Extinction Risk in the World's Carnivores.

PLoS Biol. **2**, e197 (2004).

45. Pounds, A. J. *et al.* Widespread amphibian extinctions from epidemic disease driven by

global warming. *Nature* **439**, 161–167 (2006).

46. Green, D. M. The ecology of extinction: population fluctuation and decline in amphibians.

Biol. Conserv. **111**, 331–343 (2003).

47. Moller, A. P., Rubolini, D. & Lehikoinen, E. Populations of migratory bird species that did not

show a phenological response to climate change are declining. *Proc. Natl. Acad. Sci.* **105**,

16195–16200 (2008).

48. Dornelas, M. *et al.* A balance of winners and losers in the Anthropocene. *Ecol. Lett.* [in

press] (2019).

49. Caughley, G. Directions in Conservation Biology. *J. Anim. Ecol.* **63**, 215 (1994).

50. Robbirt, K. M., Roberts, D. L. & Hawkins, J. A. Comparing IUCN and Probabilistic

Assessments of Threat: Do IUCN Red List Criteria Conflate Rarity and Threat? *Biodivers.*

Conserv. **15**, 1903–1912 (2006).

51. Harrison, S., Viers, J. H., Thorne, J. H. & Grace, J. B. Favorable environments and the

persistence of naturally rare species. *Conserv. Lett.* **1**, 65–74 (2008).

52. Mihoub, J.-B. *et al.* Setting temporal baselines for biodiversity: the limits of available

monitoring data for capturing the full impact of anthropogenic pressures. *Sci. Rep.* **7**, 41591

(2017).

53. Vellend, M. *et al.* Extinction debt of forest plants persists for more than a century following

habitat fragmentation. *Ecology* **87**, 542–548 (2006).

54. Fagan, W. F. & Holmes, E. E. Quantifying the extinction vortex. *Ecol. Lett.* **0**,

051109031307004 (2005).

55. WWF. Living Planet Report - 2018: Aiming Higher. Grooten, M. and Almond, R.E.A.(Eds).

WWF, Gland, Switzerland. (2018).

56. McRae, L., Freeman, R. & Marconi, V. 'The Living Planet Index' in: Living Planet Report

2016: Risk and resilience in a new era (ed. Oerlemans N). WWF International, Gland,

Switzerland. (2016).

57. McRae, L. *et al.* The Living Planet Index in: Living Planet Report 2012 (ed. Almond R).

WWF, Gland, Switzerland. (2012).

58. WWF. Living Planet Report 2018 Technical Supplement: Living Planet Index. (2018).

59. McRae, L., Deinet, S. & Freeman, R. The Diversity-Weighted Living Planet Index:

Controlling for Taxonomic Bias in a Global Biodiversity Indicator. *PLOS ONE* **12**, e0169156

(2017).

60. Davidson, A. D. *et al.* Geography of current and future global mammal extinction risk. *PLOS

ONE* **12**, e0186934 (2017).

61. Spooner, F. E. B., Pearson, R. G. & Freeman, R. Rapid warming is associated with

population decline among terrestrial birds and mammals globally. *Glob. Change Biol.*

(2018). doi:10.1111/gcb.14361

62. Keogan, K. *et al.* Global phenological insensitivity to shifting ocean temperatures among

seabirds. *Nat. Clim. Change* **8**, 313–318 (2018).

63. Fournier, A. M. V., White, E. R. & Heard, S. B. Site-selection bias can drive apparent

population declines in long-term studies. doi:10.7287/peerj.preprints.27507v1

64. Dunic, J. C. *et al.* Attributing the variability in direction and magnitude of local-scale marine

biodiversity change to human activities. *bioRxiv* (2017). doi:10.1101/162362

65. D. Vinebrooke, R. *et al.* Impacts of multiple stressors on biodiversity and ecosystem

functioning: the role of species co-tolerance. *Oikos* **104**, 451–457 (2004).

66. R Core Team. R: A language and environment for statistical computing. R Foundation for

Statistical Computing, Vienna, Austria. (2017).

67. LPI. Living Planet Index database, 2016. www.livingplanetindex.org/ Downloaded February

2016. (2016).

68. Wauchope, H. S., Johnston, A., Amano, T. & Sutherland, W. J. When can we trust

population trends? A method for quantifying the effects of sampling interval and duration.

bioRxiv (2019). doi:10.1101/498170

69. van de Pol, M. & Wright, J. A simple method for distinguishing within- versus between-

subject effects using mixed models. *Anim. Behav.* **77**, 753–758 (2009).

70. Foley, N. M., Springer, M. S. & Teeling, E. C. Mammal madness: is the mammal tree of life

not yet resolved? *Philos. Trans. R. Soc. B Biol. Sci.* **371**, 20150140 (2016).

71. Tarver, J. E. *et al.* The Interrelationships of Placental Mammals and the Limits of

Phylogenetic Inference. *Genome Biol. Evol.* **8**, 330–344 (2016).

72. Rabinowitz, D. Seven forms of rarity. in *The biological aspects of rare plants conservation*

205–217 (1981).

73. GBIF. Biodiversity occurrence data. http://www.gbif.org/ Downloaded Feb 2016 - Apr 2017.

(2017).

74. BirdLife International. IUCN Red List for birds. Downloaded from http://www.birdlife.org in

March 2018. (2018).

75. IUCN. The IUCN Red List of Threatened Species. Version 2017-3. (2017).

76. Di Marco, M., Venter, O., Possingham, H. P. & Watson, J. E. M. Changes in human footprint

drive changes in species extinction risk. *Nat. Commun.* **9**, (2018).

77. Dennis, B., Ponciano, J. M., Lele, S. R., Taper, M. L. & Staples, D. F. Estimating density

dependence, process noise, and observation error. *Ecol. Monogr.* **76**, 323–341 (2006).

78. Knape, J., Jonzén, N. & Sköld, M. On observation distributions for state space models of

population survey data: Observation models for population data. *J. Anim. Ecol.* **80**, 1269–

1277 (2011).

79. Pedersen, M. W., Berg, C. W., Thygesen, U. H., Nielsen, A. & Madsen, H. Estimation

methods for nonlinear state-space models in ecology. *Ecol. Model.* **222**, 1394–1400 (2011).

80. Rueda-Cediel, P., Anderson, K. E., Regan, T. J. & Regan, H. M. Effects of uncertainty and

variability on population declines and IUCN Red List classifications. *Conserv. Biol.* (2018).

doi:10.1111/cobi.13081

81. Jones, K. E. *et al.* PanTHERIA: a species-level database of life history, ecology, and

geography of extant and recently extinct mammals: *Ecological Archives* E090-184. *Ecology*

90, 2648–2648 (2009).

82. Phillips, H. R. P., Newbold, T. & Purvis, A. Land-use effects on local biodiversity in tropical

forests vary between continents. *Biodivers. Conserv.* **26**, 2251–2270 (2017).

83. Zizka, A. *et al.* CoordinateCleaner: Standardized cleaning of occurrence records from

biological collection databases. *Methods Ecol. Evol.* (2019). doi:10.1111/2041-210X.13152

84. Hijmans, R. J. geosphere: Spherical Trigonometry. R package version 1.5-5.

https://CRAN.R-project.org/package=geosphere. (2016).

85. Chamberlain, S. rredlist: 'IUCN' Red List Client. R package version 0.4.0.   https://CRAN.R-

project.org/package=rredlist. (2017).

86. Hadfield, J. D. MCMC Methods for Multi-Response Generalized Linear Mixed Models: The

MCMCglmm R Package. *J. Stat. Softw.* **33**, 1–22 (2010).

87. Olson, D. M. & Dinerstein, E. The Global 200: Priority Ecoregions for Global Conservation.

Ann. Mo. Bot. Gard. **89**, 199 (2002).

88. Magurran, A. E. & Henderson, P. A. Explaining the excess of rare species in natural species

abundance distributions. *Nature* **422**, 714 (2003).