**Spatial and Temporal Changes in Habitat Heterogeneity Promote Partition of**

**Resources of Small Mammals in a Midlatitude Temperate Forest**

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26                                    **Abstract**

One of the basics and fundamentals problems in ecology is understand the factors that

shape the spatial patterns in the distribution of the species and the coexistence of close

relatives species. Among the most important factors governing the distributions and the

coexistence of species are the spatiotemporal changes occurring in the microhabitat

heterogeneity. Here, we assessed the heterogeneity of microhabitats and how they have

an effect in the spatial segregation of two species of small mammals (*i. e., Peromyscus*

*difficilis* and *P. melanotis*), which coexist in a temperate, mixed forest. We evaluated the

microhabitat heterogeneity through multivariate statistics, using onto 23 habitat variables

for vertical-horizontal habitat structure along pluvial seasons. To detect specific

microdistribution changes and habitat preferences by two species of small mammals, we

used second order spatial statistics and general linear models. According to their

respective morphology and locomotive adaptations, the middle sized, midscansorial *P.*

*difficilis* was resident all year long and preferred microhabitats with a high log ground

cover, while the opportunistic, small sized, cursorial *P. melanotis* changed its occupancy

area, depending on density of herbaceous and woody plants cover. Under the more benign

microhabitat conditions of rainy season (denser plant coverage, milder temperature), both

species showed closer microdistribution patterns; while these became repulsive at the less

benign conditions of dry season (scarcer plant cover, colder temperature). Thus, we could

confirm that seasonal changes of microhabitat heterogeneity promoted *Peromyscus*

coexistence, through dispersion patterns reflecting partition of microhabitat resources.

**Key words:** Microhabitat selection, Use of space, Small mammals, Dispersion, Resource

partition.

**Introduction**

Spatial heterogeneity has strong impact on coexistence among species, since they become

spatially segregated, according to niche preferences such as resource requirements

(Valladares *et al.* 2015). Model simulations reveal the potentially important role of

heterogeneity and its complex and delicate interplay with dispersal in mediating long-

term outcomes of species coexistence (Schreiber and Killingback 2013; Valladares *et al.*

2015). For example, an increase in the amount of habitat types, resources and structural

complexity should increase the available niche space and thus allow more species to

coexist (Currie 1991). Equally important for the maintenance of coexistence is the

heterogeneity in time, with an influence on natural communities also variable depending

upon the temporal scales (Valladares *et al.* 2015). Temporal fluctuations can stabilize

coexistence via storage effect (Chesson 2000), when inter-annual variation in climate or

resource availability favors alternatively one group of species over the others (Zavaleta

*et al.* 2003). In particular, wild small mammals as *Peromyscus* mice are an ideal system

to evaluate whether changes in spatiotemporal habitat heterogeneity promote coexistence,

resource partitioning, spatial segregation and competition among syntopic populations

(M'Closkey and Fieldwick 1975; M'Closkey and Lajoie 1975; Monamy and Fox 1999;

Morris 1984; Seagle 1985).

Heterogeneous habitats contain patches of varying size, distribution, resources,

environmental conditions, and species composition (Fahrig and Merriam 1994) that vary

spatiotemporally, depending on observation scale and habitat type under study (Wiens

2000). It is highly likely that small size species such as small mammals perceive habitat

spatial heterogeneity at fine scale (microhabitat); therefore, both movement and foraging

by this kinds of mammals are affected by habitat heterogeneity (Bowne *et al.* 1999),

having different responses among species or demographic groups within them (Dooley and Bowers 1996). Here we focus on whether small mammals are capable to perceive and to response to microhabitat heterogeneity.

Small mammals use resources selectively, based on their requirements for growth, survival, and reproduction, (Johnson and Gaines 1980). Several studies have quantified variation in resource use at heterogeneous environments, among congeneric and co-family mice species within a community (Kaufman and Kaufman 1989 and references therein). Indeed, syntopic related species are especially valuable in studies of spatial and resource partitioning, since they are most likely to be current or past competitors (Kalcounis-Rüppell and Millar 2002). Since habitat use varies spatiotemporally (Haim and Rozenfeld 1993) due to distribution and availability of resources, we expect that such variation also affect use of resources and relationships among non-closely related species. Here we analyze how heterogeneity of microhabitats is structured and change through pluvial seasons in a temperate, mixed forest, and how such changes affect coexistence of two syntopic species of *Peromyscus* (*i. e., P. difficilis* and *P. melanotis*), over the following objectives. 1) To assess microhabitats heterogeneity according to vertical and horizontal structure indicators. 2) To analyze spatiotemporal dispersion and availability of microhabitat resources (*e. g.,* space, food in a broad sense, refuges). 3) To determine what kind of spatial relationships occur between both species, depending on seasonal changes in microhabitats heterogeneity (*i.e.,* attraction or positive: sharing of same microhabitats and resources; repulsion or negative: not sharing). 4) To disclose which specific structural elements of the microhabitats better explain abundance in the two *Peromyscus* between seasons (*i. e.,* partitioning of microhabitat resources).

 **Material and methods**

*Ethics statement*

Animal capture and handling followed guidelines of the American Society of

Mammalogists (Sikes *et al.* 2011). If animals died, corpses were prepared as voucher

specimens to house them at the Mammal Collection of the Universidad Autónoma

Metropolitana-Iztapalapa (UAMI, Ramírez-Pulido *et al.* 1989). Collecting permit,

SEMARNAT-08-049-B, was issued by DGVS, SGPA-09712/13, SEMARNAT, to

Alondra Castro-Campillo (ACC).

*Study area*

Our study area involves a temperate forest of conifers and broad-leaved trees at Parque

Nacional Desierto de los Leones (PNDL, CONANP 2006), Mexico City, within the

Transmexican Neovolcanic Range. Temperature averages $12.6 \pm 6$ °C from April to July,

while it descends to $8.1 \pm 2$ °C from December to February. Rainfalls occur from late

spring throughout summer (May to August), with an average precipitation of $235 \pm 30$

mm, though this period can go until mid-fall (October). From winter to early spring

(December to March), precipitation mean becomes $12 \pm 4$ mm, introducing a dry and cold

climate. Dominating vegetation by strata include canopy, *Abies religiosa*, *Pinus*

*hartwegii*, *P. leiophylla*, *P. montezumae*, *Prunus serotina*, *Garrya laurifolia*, *Salix*

*paradoxa*, and *Buddleia cordata*; understory shrubs, *Baccharis conferta* and *Senecio* spp.,

and herbs, *Acaena elongata*, *Sigesbeckia jorullensis,* and *Alchemilla procumbens*; ground

level, a rich variety of mosses and fungi.

*Mice sampling*

We captured-recaptured mice along ten months, including the dry and wet seasons (October 2013- August 2014), within a 2,475 m$^2$ surface plot (Fig. A.1) at 2289 m. The plot was gridded (12 vertical lines, A-L; 10 horizontal lines, 1-10) marking intersections with buried wood stakes (150x2.5x2.5 cm) to construct a coordinate system for independent sampling stations, where we set 120 Sherman live traps (Tallahassee, FL 32303, USA), baited with oat flakes and vanilla scent (Fig. A.1). Traps were set for two nights (2400 trap-nights), shortly before dusk, left open overnight, and checked at next day dawn. To avoid bias in abundance quantification, we code-marked temporally each mouse with gentian violet (De la Cruz-Arguello 2015). We recorded coordinates of any station with a captured mouse, its species (*Peromyscus melanotis*, *P. difficilis*) and its sex by inspection of its external genitalia.

*Microhabitat features*

To assess environmental spatiotemporal heterogeneity of microhabitat within the entire plot, we delimited an influence zone (sub-plot, Fig. A.1; close-up, Fig. A.1), drawing a rectangle of 2.5 m$^2$ around each sampling station. We sampled 23 variables (Table 1) in each influence zone during the rainy (June-October) and dry seasons (January-April) to assess components of vertical and horizontal structure of microhabitat, as well as factors affecting distribution of small mammals at fine scales (Morris 1984, 1987; Stapp 1997; Jorgensen 2004, Coppeto *et al.* 2006; Villanueva-Hernández *et al.* 2017). Table 1 shows assigned ecological meanings for each variable.

We adapted Canfield's (1941) method of Line Intercept (LI) to measure all

variables in each pluvial station, for it allows sampling within-plot variation and

quantifies changes, both in plant species cover and height over time, by using transects

located within a plot. We drew eight graduated transects (cm) from the respective

influence zone of each station to cover the 2.5 m$^2$ sub-plots (Fig. A.1); four 2.5 m transects

were cardinally oriented (N, S, E, W), while other four 3.30 m ones were diagonally

oriented (NE, NW, SE, SW). Only plants intercepted by diagonal transects were counted

for vegetation species richness, number of life forms at different heights (herbaceous and

woody plants), and total herbaceous and woody plants; while all transects were used to

measure plant coverage at different heights, as well as coverage of fixed elements (*i. e.,*

rocks, logs, bare soil, dead organic matter). We calculated coverage percentage of each

variable within the sub-plots with the formula $\sum(Xi)\frac{100}{2400}$, where *Xi* is centimeters

occupied by every plant in each transect, and 2400 equals total longitude (sum of the eight

transects). We counted directly the number of woody and herbaceous plant life forms at

different heights and burrows within each influence zone of every sampling station. We

took each hole in the ground ca. 8-12 cm of diameter as a potential burrow for both

*Peromyscus* (Álvarez-Castañeda 2005; Fernandez *et al.* 2010).

*Statistical data analysis*

We standardized all variables and run statistical tests at $p \leq 0.05$ (Zar 1999). To assess

microhabitat heterogeneity within the grid, we performed a Hierarchical Cluster Analysis

(HCA) in JMP® (ver. 9, SAS Institute Inc., Cary, NC, 1989-2007), using Ward´s method

(1963), where distance between two clusters is the ANOVA' sum of squares between

them, added up over all variables. Visual inspection of the dendrogram and of no drastic

changes in the variance screeplot, gave us three general groups of stations, which we

interpreted as distinct microhabitat types (M1-3). We further statistically validated such

microhabitats, as *a priori* groups in Discriminant Analyses (DA, Addinsoft SARL's

XLSTAT 2013; $F = 9.99$ dry; $F = 9.64$, rains; Wilk's Lambda = 0.0001 in both).

For distribution changes, related to the rainy and dry seasons, on each

microhabitat variable, we used Student's *t*-parametric tests in normally distributed

microhabitat variables and non-parametric Wilcoxon test for non-normal variables (Table

2), in either one or both seasons. We checked distribution normality of every variable

with Kolmogorov-Smirnov tests; there was no multicollinearity among the 23 variables

in correlation tests (Zar 1999). In order to know whether the number of stations

structuring each type of microhabitat changed between seasons (seasonal microhabitat

heterogeneity), we used a contingency table and $X^2$ test. All these analyses were

performed with XLSTAT 13 (Addinsoft SARL).

To examine association between changes in spatial heterogeneity of microhabitats

and dispersion patterns of *Peromyscus*, along the entire grid, we conducted a Contingency

Tables Analysis. Then, for visualizing associations between the *Peromyscus* and

microhabitats, we performed a Correspondence Analysis with the constructed

Contingency Tables for both seasons, including abundance of each species in the three

microhabitats (Table A.2).

*Second order spatial analysis*

To map variations in point density captures of small mammals, and to find density

gradients across the plot area, we used the Kernel function (PAST, ver. 3.14, Hammer *et*

*al.* 2001). To assess whether both species were associated or disassociated, we compared

dispersion patterns between the two mice, using Ripley's K bivariate function (Ripley

1977), since the method considers all distances among individuals located under a

Cartesian scheme (X,Y) in a quadrat plot (Ripley 1976, Dale 1999, Zavala-Hurtado *et al*.

2000). A bivariate pattern occurs when there are only two point categories (the two

*Peromyscus*); thence, each capture point was assigned to either *P. difficilis* or to *P.*

*melanotis*, beyond its simple location within the grid in Ripley's K bivariate function,

$K_{12}(d)$. To evaluate statistical significance of $K_{12}(d)$, we estimated 95 % confidence

intervals (95%CI), using a Monte Carlo procedure, based on 1000 random relocation

simulations of the sampling stations in the plot (Zavala-Hurtado *et al.* 2000). When L(*d*)

was positive and took values over the upper limit of the confidence region, we inferred

dissociation or repulsion between the *Peromyscus* at the corresponding (*d*) scale; whereas

a significant negative deviation indicated us a pattern of association or attraction between

the two mice (Dale 1999). If L(*d*) remains between the limits of the 95%CI for a given

value of *d*, the null hypothesis of independence between the two contrasts cannot be

rejected (Dale 1999). We used PASSaGE (ver. 2, Rosenberg and Anderson 2011) to carry

out Ripley's bivariate K analysis.

*Prediction of microhabitat elements affecting use of space by each species*

To elucidate what elements of microhabitat structure, directly affected use of space by

*Peromyscus difficilis* and *P. melanotis*, we performed General Linear Models (GLIMs),

using JMP® (ver. 9, SAS Institute Inc., 1989-2007, Cary, NC,). First, we conducted a

Principal Component Analysis (PCA) onto the means of the 23 variables to obtain

functions summarizing the greatest variance at each season. Thenceforth, we only used

functions explaining > 10 % of variance, as independent variables to predict likelihood

that a mouse of *P. difficilis* or *P. melanotis* could be present in each of the sampling

stations, either in dry or rainy seasons, within the GLIMs. In each model, response

variable was capture frequency of both species in each station of the plot, and we assumed

a Poisson distribution with a log link function. Criteria for selecting models that most

explained number of mice for each species at each season in the sampling stations,

included: model significance of $p > 0.05$; lowest Akaike Information Criterion (AIC), *i.*

*e.,* measure of variance explained by the model (Posada and Buckley 2004); and its

deviation (Deviance); as well as biological and ecological meaning of results.

235                                                    **Results**

*Behavior of 23 variables of microhabitat between seasons*

Sixteen variables from the microhabitat structure showed statistical differences in their

mean values (Table 2) between pluvial seasons, main changes occurring in variables that

represented herbaceous vegetation, according to plant coverage at different heights, and

vegetation species richness (VSR). As expected in overall, woody life forms were more

stable between seasons (Table 2). Components of microhabitat showing no statistical

change between seasons were frequency of 10 cm height herbaceous plants (FH11),

frequency of both 35 cm (FW23) and 50 cm (FW24) woody plants, as well as coverage

of logs (CLO), rocks (RC), and bare soil (BS). Therefore, space configuration given by

these structural features remained stable in both seasons.

*Microhabitat heterogeneity*

The dendrogram from cluster analysis revealed three different kinds of microhabitats

(M1, M2, and M3) within the sampling plot in each season. M1 included more sampling

stations during the dry season, while M2 covered the majority of sampling stations during

the rainy season (Table A.3). Indeed, Fisher's exact test of Contingency Tables Analysis

revealed significant changes in number of sampling stations ($p > 0.004$) from dry to rainy

seasons with main changes occurring between M1 ($X^2 = 2.123$, $0.05 > p$) and M2 ($X^2 =$

$3.348$, $0.05 > p$), especially in the latter, since it augmented 24 sampling stations from the

dry season to the rainy season. Conversely, number of sampling stations remained quite

stable in M3 ($X^2 = 0.011$) throughout the study (Table A.3).

Discriminant analyses validated heterogeneity of the three microhabitats (Fig. 1)

within and between pluvial seasons. Differences in apportion of variance percentages in

the discriminant functions was more clear during the rainy season (dry season: F1 = 58.67

%, F2 = 42.32 %; rainy season: F1 = 72.581 %, F2 = 27.419 %) (Fig. 1). Discriminant

functions explained variation due to woody plants in the understory in the dry season,

while herbaceous vegetation together with decayed matter and logs became more relevant

at rains. Wilk's Lambda showed that at least one microhabitat mean differed statistically

from the others in both dry ($p > 0.0001$) and wet ($p > 0.0001$) seasons (Fig. 1). These

results corroborated spatial heterogeneity drawn from cluster analysis of individual

station features in the grid plot. During the dry season, 93.33 % of individual sampling

stations remained correctly classified, and 94.17 % of them during the rainy season (Table

A.3, Fig. 1).

*Description of microhabitats within the plot*

*Microhabitat 1.* M1 (Fig. 1) was characterized by low amounts, or even absence, of

vegetation cover at different heights (VC) and low frequency of herbaceous and woody

plants (FH and FW, respectively) in the understory. In contrast, organic matter on the

ground (OM) was the most frequent component. This microhabitat was also distinctive

for having a large coverage of logs (CLO) on the ground and for being the microhabitat

with the major presence of burrows (BW).

*Microhabitat 2.* M2 (Fig. 1) was the largest area covered by herbaceous life forms at

different heights; vegetation species richness (VSR) and total number of herbaceous

plants (TH) remained also very high. Woody life forms at 25 cm (FW22) were present

but at low frequencies, while FW24 and FW25 remained at higher frequencies. Coverage

on the ground of small logs or dead wood (CLO), of rocks (RC), and organic matter (OM)

were very low during the study. Bare soil surface (BS) increased to higher amounts during

the dry season, while it decreased at the rainy season. There were no burrows (BW) in

this microhabitat.

*Microhabitat 3.* In M3 (Fig. 1), VC25, VC35, VC50, and VC100 showed high values,

while vegetation species richness (VSR) averaged the highest compared to other

microhabitats. Total number of woody plants (TW) and frequency of woody plants at

different heights (FW21-25) remained very high, while total number of herbaceous plants

(TH) was very low. Vegetation coverage at 10 cm (VC10), rocks (RC), logs (CLO), and

bare soil surface (BS) were very low during the dry season; however, VC10 increased in

the rainy season. Conversely, OM was present in the dry season, but decreased in the

rainy season. Number of burrows (BW) remained quite scarce.

*Association between the microhabitats and species*

Kernel Graphs (Fig. 2) indicated that the two species used space distinctively, since they

were differentially scattered along the plot. *Peromyscus difficilis* was more abundant

during the dry season, occupying a large portion of the plot. Instead, *P. melanotis*

increased its abundance during the rains when it seemed to displace *P. difficilis* into other

sampling stations (Fig. 2). Distribution rearrangements between seasons implied that one

species occupied some stations more frequently than the other, and vice versa (Fig. 2).

Indeed, the two $X^2$ tests yielded statistical abundance differences in each species at both

microhabitats and seasons (Table A.2: dry season; $R^2$ (U) = 0.09, $n$ = 111, df = 2,

Likelihood Ratio $X^2$ = 15.07, $p$ = 0.00005; rainy season; $R^2$ (U) = 0.02, $n$ = 168, df = 2,

Likelihood Ratio $X^2$ = 6.40, $p$ = 0.0406).

During dry season (Fig. 3a and Table A.2), *Peromyscus difficilis* was highly

associated to M2 (80 % captured mice) and frequently associated to M1 (64 % captured

mice); conversely, *P. melanotis* showed frequent association to M3 (66 % captured mice).

Spatial use of microhabitats changed for both species in the rains (Fig. 3b, and Table A.2),

showing again a microhabitat partition, though less obvious: in the Correspondence

Analysis, 61 % of captured *P. difficilis* occurred in M1 and this species was also

associated to M3, while 60% captured mice of *P. melanotis* was associated to M2.

*Spatial Patterns of attraction or repulsion between species*

During the dry season, Ripley's bivariate K showed a statistical significant pattern of

repulsion for both *Peromyscus* in all analyzed distances of the entire plot (Fig. 4a). Such

repulsion pattern turned over in almost all distances during the rainy season (Fig. 4b),

since the two species became more associated, sharing microhabitats in almost all capture

stations. However, in the rainy season statistical significant peaks of repulsion reappeared

between the species at distances of five, ten, and 14 m (Fig. 4b). Analyses also revealed

intensity of these patterns; e.g., likelihood for finding individuals of *P. difficilis* and *P.*

*melanotis* together at the same capture station was very low during the dry season (Fig.

4a), while this probability increased in the rainy season (Fig. 4b).

*Structural components of microhabitats affecting distribution of small mammals*

In the dry season, Generalized Linear Model (GLIM) of the effect of PC1 and PC3 from

the PCA on abundance of *P. difficilis* (Fig. A.2), resulted statistically significant; indeed

with a positive relationship (AICc 212.55). Environmental variables with higher scores

in the PC1 ($p = 0.0186$) were total number of herbaceous plants (TH), vegetation species

richness (VR), and vegetation cover at 10 cm height (VC10). In PC3 ($p = 0.0001$),

characterizing variables were number of logs on the ground (CLO), as well as presence

of burrows (BW). These five variables were also the main elements characterizing M2

(PC1) and M3 (PC3), the two microhabitats to which *P. difficilis* was associated in this

season. As for *P. melanotis* (Fig. A.3), the GLIM model showed it was only positively

associated to PC2 (AICc = 204.47, $p = 0.0001$). Here, among the four variables with

higher scores, were total number of woody vegetation (TW) and frequency of woody life

forms at 10 cm height (FW21), the two main elements at M3 for the dry season

During the rainy season, a similar pattern of microhabitat partition occurred

between the two species. *P. difficilis* (Fig. A.2) was now mainly associated (AICc =

281.77) to total number of woody plants (TW), and vegetation cover at 10 cm of height

(VC10), in PC2 ($p = 0.0065$); to number of logs on the ground (CLO), in PC3 ($p =

0.0210$); and to vegetation cover at 50 cm and 100 cm (VC50, VC100), in PC4 ($p =$

0.0076). The latter involved variables with high presence at M1 and M3 during the wet

season, where more *P. difficilis* occurred. On the other hand, *P. melanotis* (Fig. A.2) was

only associated to PC1 (AICc = 246.46, *p* = 0.0262), increasing as the number of total

herbaceous plants (TH), vegetation richness (VR), and the plant cover at all analyzed

heights (VC10-100 cm) also increased. Thence, PC1 represented variables commonly

found in greater amounts at M2, the microhabitat with more abundance of *P. melanotis.*

Therefore*,* both results of Contingency Tables Analysis, Correspondence Analysis, and

GLIMs models showed that the two syntopic species of *Peromyscus* switched and split

resources in space, as environmental conditions changed (Fig. A.2).

361                                                         **Discussion**

Our findings on microhabitat associations of rodents in a patch of a mid-latitude

temperate forest, concur with the habitat heterogeneity coexistence hypothesis (Cramer

and Willig 2002; Valladares *et al.* 2015). We found that both *Peromyscus* were sensitive

to slight modifications of habitat structure and that the three microhabitats provided

different resources for each one. Our results also suggest that coexistence between these

syntopic *Peromyscus* is mainly facilitated by temporal differences in space structure, and

probably also by changes on availability of both food and shelter that go with it (Pianka

1973; Schoener 1974). For instance, *P. difficilis* was more associated to M1 and M2 by

the dry season, while *P. melanotis* was highly associated to M3. Conversely, *P. difficilis*

was more associated to M1 and slightly associated to M3 by the rainy season, while *P.*

*melanotis* was highly attracted to M2. Partition in space use has already been documented

for coexisting species of *Peromyscus* within a community; *i.e.,* extensive studies of niche

partitioning by *Peromyscus leucopus* and *P. maniculatus* (Kaufman and Kaufman 1989).

Moreover, several studies have demonstrated variation in spatial resource use by

congeneric species (Barry *et al.* 1990; Dooley and Dueser 1996; Kalcounis-Rüppell and

Millar 2002).

In our study, *P. difficilis* remained associated to M1 and M3 in both seasons,

especially to the former during the wet season. Number of fallen logs was very high at

M1, while understory complexity, together with woody vegetation, were more relevant

in M3. Such microhabitats represent small patches of food for small mammals, offering

invertebrates (Bellows *et al.* 2001), flowers, grains, and fungi items (Bowman *et al.*

2001). In these microhabitats, individuals of *P. difficilis* can also use fallen logs as shelter,

as well as pathways for quick and straight locomotion inside the forest (Bellows *et al.*

2001; Dewalt *et al.* 2003). Indeed, fallen logs promote structural complexity of forests

and may enhance positive interaction among species of small mammals (Bowman and

Facelli 2013). GLIM's analyses showed that abundance of *P. difficilis* was explained by

PC1 and PC3 during the dry season and by PC2-4 in the rainy season. In the dry season,

variables contributing most to PCA ordination in PC1 and PC3 were total number of

herbaceous vegetation (TH) and number of logs on the ground (CLO), as well as burrows

(BW). While in the dry season, PC2-4 represented woody vegetation (TW), number of

logs, burrows, and understory complexity (VR). Bellows *et al.* (2001) found a similar

result in a high latitude temperate forest (Virginia, USA), where distribution of a small

mammal, generalist species was associated with the diameter of fallen logs, frequency of

shrubs, and degree of canopy closure. Association between rodents and fallen logs was

also documented for *Nectomys squamipes* that builds up its nests inside decomposed,

fallen logs (Briani *et al.* 2004); this has also been recorded for other rodent species in

several biomes with different vegetation types (*e. g.*, *Rattus rattus*, *Nesomys audeberti* in

Lehtonen *et al.* 2001; *Oligoryzomys nigripes* in Dalmagro and Vieira 2005).

In contrast, *P. melanotis* showed a close association with only one PC over each

pluvial season. During the dry season, abundance of this species was only associated to

PC2, which represented total number of woody vegetation (TW). Then by the rainy

season, the species became only related to PC1, which accounted for understory

complexity dominated by herbaceous life forms (TH). Association between abundance of

rodents and habitats with increased understory density, has been interpreted as protection

against aerial predators (Dalmagro and Vieira 2005), or as preference for more complex

habitats that enable for vertical stratifications and, thus for extended coexistence inside

fragments (Grelle 2003; Johnson 2007).

Coexistence of both *Peromyscus* at the study plot, must be also related to its

intrinsic factors. For instance, partition of space by these syntopic deermice must be

facilitated by their respective locomotive habits as semiarboreal (*P. difficilis*) and

cursorial (*P. melanotis*). Both the long tail and wide sole surface of *P. difficilis*, must

enable it to rush and climb along shrubs or trees (Álvarez-Castañeda 2005), thus easing

its preference for habitats where it can escape from predators. Indeed, adult color coat

changed in *P. difficili*s between pluvial seasons, becoming more similar to ground litter;

*e. g.,* in this resident deermouse, color phenotype plasticity must be a cryptic response

elicited against predators. On the other hand, a smaller sized body, together with a shorter

tail and narrower soles (Fernandez *et al.* 2010), should enable the cursorial *P. melanotis*

to occupy such zones as M2 and M3, where it cannot be easily spotted by predators

through a dense vegetation cover, so it can scape very quickly. Indeed, high shrubs cover

provide both protection from predators and food sources, since seeds may be concentrated

under shrub canopies (Thompson 1982; Mohammadi 2010). In fact, rodents usually avoid

foraging in unsheltered microhabitats and forest edges where they are more likely to be

spotted by avian (Kotler *et al.* 1991) and other vertebrate predators (Morris and Davidson

2000; Mohammadi 2010). *P. melanotis* generally showed a more opportunistic pattern of

habitat occupation as compared to the larger sized *P. diffiilis*. Chupp (2002) also showed

an opportunistic habitat by small sized *Peromyscus leucopus*, which he related to

predation risk. Further studies about influence of predation on opportunistic behavior of

*P. melanotis* are needed.

One possible consequence of such extrinsic and intrinsic operating factors is that

*P. difficilis* remained more abundant and always present at the microhabitats with more

stable elements in the study area (our unpublished data), while *P. melanotis* only

increased its abundance during the rainy season, when environmental conditions became

more benign and plant cover became denser. Since rains promote increase of primary

productivity, allowing more resource availability (mainly food) and enhancing

microhabitat carrying capacity, such habitat changes facilitate coexistence between both

*Peromyscus*, and with other small land mammals in the area (Castro-Campillo *et al*. 2008,

2012). Ripley's bivariate K supported and shed light into such outcome, since the species

showed repulsion between them in all analyzed distances during the dry season; *i.e.,* it

was almost null that a mixed pair used the same space or microhabitat during this season.

Conversely, intensity of such repulsion decreased substantially during the rainy season;

moreover, at some analyzed distances, the two deermice showed an association pattern.

Our results also indicate that in these two syntopic *Peromyscus*, both microhabitat use

and spatial organization are seasonally variable (Brown and Zeng 1989; Cramer and

Willig 2002).

Finally, we want to highlight and recommend our sampling of environmental

features and quantitative analyses, especially for Ripley's bivariate K, as neither one has

been extensively used in studies focusing on the spatial dynamics of dispersion in small

rodents, despite its helpful advantages. The methodological approach used here, together

with the gathered evidences became relevant for providing us with basic information

about the ability of this two *Peromyscus* to split resources in a mid-latitude temperate

forest at the edge of a megalopolis, which continuous enlargement produces

fragmentation and patching of natural microhabitats. Therefore, our local scope was

explicitly designed at this initial point, to focus on small sized rodents with low vagility.

Besides, we successfully aimed to reconstruct vegetation structure, in order to provide us

with clues as how to eventually manage such perturbed ecosystems for conservation

purposes. We also think that our methodology can be helpful in other scenarios for

understanding small rodents, such as ethological dynamics, activity patterns, and

reproductive systems at either or both intra or interpopulation levels.

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**Table 1.** Environmental variables used to measure horizontal and vertical structure of microhabitat at each sampling station, showing its ecological interpretation (kind or resource type) for both species of *Peromyscus*.

| Variable code | Structure | Predation | | Resources | | | | |
|---|---|---|---|---|---|---|---|---|
| | Spatial heterogeneity | Diminished risk | Refuges | Breeding Space | Food | Temporal food and refuges | Permanent | Resting places |
| **Vegetation Cover (cm)** | | | | | | | | |
| **VC10** | X | X | | | X | X | | X |
| **VC25** | X | X | | X | X | X | | X |
| **VC35** | X | X | | X | X | X | | X |
| **VC50** | X | X | | X | X | X | | X |
| **VC100** | X | X | | X | X | X | | X |
| **Frequency of Herbaceous Plants (cm)*** | | | | | | | | |
| **FH11** | X | X | | X | X | X | | X |
| **FH12** | X | X | | X | X | X | | X |
| **FH13** | X | X | | X | X | X | | X |
| **FH14** | X | X | | X | X | X | | X |
| **FH15** | X | X | | X | X | X | | X |
| **Frequency of Woody Plants (cm)*** | | | | | | | | |
| **FW21** | X | X | X | X | X | X | X | X |
| **FW22** | X | X | X | X | X | X | X | X |
| **FW23** | X | X | X | X | X | X | X | X |
| **FW24** | X | X | X | X | X | X | X | X |
| **FW25** | X | X | X | X | X | X | X | X |
| **Vegetation Species Richness** | | | | | | | | |
| **VR** | X | X | X | X | X | X | X | X |
| **Total Herbaceous Plants** | | | | | | | | |
| **TH** | X | X | | X | X | X | | X |
| **Total Woody Plants** | | | | | | | | |
| **TW** | X | X | X | X | X | X | X | X |
| **Rock Coverage** | | | | | | | | |
| **RC** | X | X | X | X | | | X | X |
| **Organic Matter (litter, twigs, small trunks)** | | | | | | | | |
| **OM** | X | | | | X | X | | |
| **Bare Soil** | | | | | | | | |
| **BS** | X | | | | | X | | |
| **Coverage of Logs (stumps, fallen trunks)** | | | | | | | | |
| **CLO** | X | X | X | X | | | X | X |
| **Number of Burrows** | | | | | | | | |
| **BW** | X | X | X | X | | | X | X |

* = Frequencies of plant types (FH, FW) involve the same heights as in VC.

671 **Table 2.** Normality (**NOR**) tests for 22 variables (**VAR**) of microhabitat, and

672 comparisons[abc] (**COM**) for average values between the dry (**DRY**) and rainy (**RAINS**)

673 seasons, respectively. Variable names in Table 1.

| VAR | NOR[a] | | COM | VAR | NOR[a] | | COM | VAR | NOR[a] | | COM |
|---|---|---|---|---|---|---|---|---|---|---|---|
| | DRY | RAINS | | | DRY | RAINS | | | DRY | RAINS | |
| BS | 0.0001 | 0.0001 | *1.0[c]* | VC100 | 0.002 | 0.003 | 0.0001[c] | TW | *0.143* | *0.08* | 0.0004[b] |
| CLO | 0.0001 | 0.0001 | *0.972[c]* | VR | 0.006 | *0.082* | 0.0001[b] | FW21 | 0.0001 | 0.0001 | 0.004[c] |
| OM | 0.010 | 0.034 | 0.0001[c] | TH | 0.0001 | *0.072* | 0.0001[b] | FW22 | 0.0001 | 0.0001 | 0.026[c] |
| RC | 0.0001 | 0.0001 | *0.432[c]* | FV11 | 0.015 | 0.028 | *0.658[c]* | FW23 | 0.0001 | 0.0001 | *0.875[a]* |
| VC10 | 0.002 | 0.041 | 0.0001[c] | FV12 | 0.0001 | 0.0001 | 0.001[c] | FW24 | 0.0001 | 0.0001 | *0.378[c]* |
| VC25 | 0.009 | 0.003 | 0.0001[c] | FV13 | 0.0001 | 0.0001 | 0.0001[c] | FW25 | 0.0001 | 0.001 | 0.0001[c] |
| VC35 | 0.0001 | 0.002 | 0.0001[c] | FV14 | 0.0001 | 0.0001 | 0.0001[c] | | | | |
| VC50 | 0.0001 | 0.019 | 0.0001[c] | FV15 | 0.0001 | 0.0001 | 0.0001[a] | | | | |

677 Parametric: [a]Kolmogorov-Smirnov; [b]Student for two samples; non-parametric: [c]Wilcoxon). The *p*-values in italics do

678 not reject normality ($p \geq 0.05$), or do not indicate significant differences between pluvial seasons.

 **Figure legends**

**Fig. 1.** Classification of the three types of microhabitats (M1 = red, M2 = green, M3 =

blue), in the dry (a) and rainy (b) seasons, respectively, resulting from the behavior and

the distributions of the 23 structural variables. Below each figure is observed the

classification by Discriminant analysis made with the *a priori* classification of the cluster

analysis.

**Fig. 2.** Spatial analysis with Kernel density points showing capture frequencies for *P.*

*difficilis* at dry (a) and rainy (b) seasons, and for *P. melanotis* at dry (c) dry and rainy (d).

Dots depict actual capture points, while darker shadowing indicates higher density.

**Fig. 3.** Correspondence analysis from contingency tables of frequency capture data for

each deermouse (*P. difficilis* and *P. melanotis*) in each microhabitat (M1-3), during the

dry (a) and rainy (b) seasons, respectively. Axis c1 and c2 indicate the coordinates

resulted from the ordination of each variable from the analysis.

**Fig. 4.** Ripley's bivariate K Analysis for interactions between the two *Peromyscus*. (a)

Repulsion between both species in all analyzed distances during the dry season. (b)

Attraction from 6 - 12 m, and repulsion at 5, 11, and 14 m during the rainy season. Black

solid line represent the Ripleys K bivariate pattern; red dashed lines represent the 95%

CI; blue solid line shows the null hypothesis.

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745   **Fig. 2**

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830     **Fig. 4**