

A Machine Learning Approach Predicts Tissue-Specific Drug Adverse Events

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ABSTRACT

One of the main causes for failure in the drug development pipeline or withdrawal post approval is the unexpected occurrence of severe drug adverse events. Even though such events should be detected by in vitro, in vivo, and human trials, they continue to unexpectedly arise at different stages of drug development causing costly clinical trial failures and market withdrawal. Inspired by the “moneyball” approach used in baseball to integrate diverse features to predict player success, we hypothesized that a similar approach could leverage existing adverse event and tissue-specific toxicity data to learn how to predict adverse events. We introduce MAESTER, a data-driven machine learning approach that integrates information on a compound’s structure, targets, and phenotypic effects with tissue-wide genomic profiling and our toxic target database to predict the probability of a compound presenting with different types of tissue-specific adverse events. When tested on 6 different types of adverse events MAESTER maintains a high accuracy, sensitivity, and specificity across both the training data and new test sets. Additionally, MAESTER scores could flag a number of drugs that were approved, but later withdrawn due to unknown adverse events - highlighting its potential to identify events missed by traditional methods. MAESTER can also be used to identify toxic targets for each tissue type. Overall MAESTER provides a broadly applicable framework to identify toxic targets and predict specific adverse events and can accelerate the drug development pipeline and drive the design of new safer compounds.

INTRODUCTION

Drug adverse events are currently one of the main causes of failure in drug development and are one of the top 10 causes of death in the developed world^{1, 2}. Toxicity issues remain a leading cause for the rising clinical trial attrition rates^{3, 4}. Even after a drug has been approved, adverse drug reactions remain a large burden on the medical system with the costs amounting to as much as \$30 billion dollars annually in the USA⁵. Furthermore the identification of the serious adverse events associated with drugs

47 frequently does not occur until after FDA approval, with as many as 50% of adverse
48 events going undetected during human trials⁶. Due to the prevalence and impact of this
49 problem, the U.S. Food and Drug Administration (FDA) has established the US FDA
50 Adverse Event Reporting System (FAERS).

51
52 Most adverse event detection experiments are carried out in pre-clinical phases based
53 on animal results or during early clinical trials. However not all adverse events are
54 detected, due to several factors including limited relevance of animal models to human
55 physiology, limited sample sizes during trials, and patient populations that may not be
56 representative of the overall population⁵. Further complications may include the low
57 frequency or late onset of some adverse events⁵. As a result, retrospective studies are
58 currently an important method for further characterization of the side effects associated
59 with drugs. However this requires a large number of patients to be treated first and is
60 dependent on voluntary reporting, which is especially problematic as only 10% of all drug
61 adverse events are reported post-approval⁷.

62
63 Ideally possible adverse events would be detected during the pre-clinical phases of drug
64 development, even before animal studies. Cell lines and reporter assays may help detect
65 unwanted side effects early, but are often imprecise. Computational screening methods
66 are also critical components of current drug development pipelines for evaluating pre-
67 clinical toxicity. In particular, drug-likeness measures, which use molecular features to
68 estimate oral bioavailability as a proxy for drug toxicity, have been widely adopted.
69 Examples of drug-likeness methods include Lipinski's Rule of Five⁸ and the Quantitative
70 Estimate for Drug Likeness⁹. More recently machine learning based methods have been
71 proposed for predicting drug toxicity, including previous work from our group (PrOCTOR)
72 which integrates established molecular properties with target-based features to directly
73 predict broad clinical trial toxicity¹⁰. Other groups have developed diverse methods
74 focused on predicting toxicity specific to the liver¹¹. However no method has yet been
75 developed with the granularity to predict multiple specific adverse events across different
76 tissue types, such as heart attacks or neutropenia, for a specific drug. Better methods
77 for predicting such adverse events could improve fast-fail procedures and facilitate better
78 trial design. To address this problem, we introduce MAESTER, a new machine-learning
79 platform for the prediction of tissue-specific drug adverse events. We show that for a set
80 of 6 serious adverse events MAESTER achieves unprecedented accuracy while
81 maintaining high specificity and sensitivity. Additionally we demonstrate how MAESTER
82 could have identified drug adverse events that were missed by traditional screening
83 methodologies but led to costly market withdrawal.

84 85 **RESULTS**

86 87 ***Identifying determinants of tissue-specific toxicities and adverse events***

88 We first sought to identify drugs or compounds that are specifically toxic within individual
89 tissues and compare them with compounds with no reported toxicities in these tissues.
90 We focused on a set of six tissues whose corresponding AEs are correlated with clinical

91 trial failures: liver, kidney, blood, heart, lung, and pancreas (**Fig.S1A**). We used the
92 SIDER database of drug side effects to identify subsets of drugs that are associated with
93 tissue-specific adverse events (TSAEs) (**Table 1**)¹². For example we identified all drugs
94 that have been associated with liver toxicities. For each tissue, we also established a
95 “safe” set of drugs for comparisons identifying any drugs not associated with those TSAEs
96 or other AEs highly correlated with fatalities in openFDA (<https://open.fda.gov/>) defined
97 as having a fatality frequency > 13% (**Fig.1A**). For each drug, we compiled structural
98 representations in the format of SMILES from DrugBank, differential gene expression
99 profiles from the Broad Institute’s Connectivity Map (CMAP)¹³, growth inhibition patterns
100 across the NCI60 cell lines (NCI60) from the NCI’s Developmental Therapeutics
101 Program¹⁴, and bioassay data from PubChem¹⁵.

102
103 For each tissue we then investigated how these safe and toxic drugs compare to each
104 other. For each pair of drugs, we calculated a similarity score for each of the considered
105 data types (**Methods**). We found that in all tissues, tissue-specific toxic drugs were most
106 structurally similar to each other (**Fig.1B**). Additionally, toxic drugs tended to also be most
107 similar to other toxic drugs in terms of differential gene expression profiles (**Fig.1C**),
108 growth inhibition screens (**Fig.1D**) and bioassays (**Fig.1E**). Interestingly we found distinct
109 patterns across the different tissue types – for instance, growth inhibition was best able
110 to separate out drugs with blood specific adverse events, whereas gene expression
111 changes had the greatest utility in the liver. These patterns could be incredibly valuable
112 for adverse event prediction as they highlight how we can model the diversity across
113 drugs with a given side effect. For example high structural similarity between a new
114 compound and compounds known to be toxic in the heart could indicate potential cardiac
115 toxicity for that new compound. Additionally high similarity between the compound-
116 induced expression changes of a new compound with expression changes of compounds
117 with known liver toxicity could suggest liver toxicity for the new compound.

118
119 We next examined how expression of a drug’s targets could be used to predict TSAEs.
120 For this analysis we integrated tissue-specific expression data measured by the GTEX
121 database. For each toxic or safe drug in a given tissue set (**Fig.1A**), we quantified the
122 expression of all of that drug’s targets in the specific tissue. Overall drugs with adverse
123 events in a specific tissue tended to also have higher target expression in that tissue than
124 their safe drug counterparts (**Fig.2A-E**). This information helps illustrate how it is
125 important to consider target based features and tissue-specific expression when
126 predicting adverse events. This analysis also confirms that high expression of a drug’s
127 target in a given tissue can help predict toxicity in that tissue.

128 129 ***Distinct Patterns of Tissue-Specific Toxic and Safe Target Sets***

130 Due to the significant relationship between drug target expression and related tissue
131 adverse events, we next sought to define a set of tissue-specific “toxic targets”– proteins
132 that are only targeted by drugs with known toxicity in that tissue – and “safe targets” –
133 proteins only targeted by drugs with no related tissue toxicities. To do this, we begin by
134 taking the safe and toxic drug sets described in **Fig.1A** and identifying any targets

135 exclusive to each drug subset (**Fig.2F**). Interestingly we found that though there was a
136 significant degree of overlap between the toxic and safe gene sets across multiple
137 tissues, there were a number of proteins identified that were specifically associated with
138 toxicity or non-toxicity in a single tissue (**Fig.2G-H**). For instance, ABL1 was flagged as a
139 toxic target in all six tissues, whereas KCNJ3 and KCNJ6 – proteins involved in voltage
140 gated potassium channels and the regulation of heartbeats – were only marked as toxic
141 targets in the heart.

142
143 To further investigate features of tissue-specific toxic targets we expanded the procedure
144 described in **Fig.2F** to generate toxic and safe targets for 30 different tissue types –
145 including the 6 prior tested tissues. For each target, we computed a number of features,
146 including tissue-specific expression, network properties (betweenness and degree), loss
147 of function (LoF) mutation frequency, and essentiality status. We found that toxic gene
148 sets tend to be more connected in an aggregated protein-protein interaction network
149 (**Fig.3A-B**), be more intolerant for LoF mutations (**Fig.3C**), and be enriched for essential
150 genes (**Fig.3D**). Finally, we used the ConsensusPathDB framework¹⁶ to measure for GO
151 term enrichment and observed that for toxic gene sets the most commonly enriched terms
152 had to do with cell death, receptor signaling, and apoptotic processes (**Fig.3E**) –
153 pathways one would expect to be related to toxicity – whereas safe targets did not appear
154 to be related to any toxicity related processes (**Fig.3F**) – likely due to the diverse nature
155 and function of safe targets. Altogether these results suggest that tissue-specific toxic
156 targets have specific recognizable features and that such features may be used to predict
157 whether a new compound whose targets are known is likely to be toxic in a given tissue.

158 159 ***Computational approach predicts likelihood of specific adverse events***

160 To utilize these findings and more directly address the problem of adverse event
161 prediction, we developed MAESTER (a **M**oneyball **A**pproach for **E**stimating **S**pecific
162 **T**issue adverse **E**vents using **R**andom forests) to compute the probability of a compound
163 presenting with a specific adverse event (**Fig.4A**). To do this, we expanded upon the
164 framework of our previously published work on predicting broad clinical trial toxicities,
165 ProCTOR¹⁰, and narrowed down the classification task to a set of specific adverse
166 events that are correlated with clinical toxicity and have high reported frequencies of
167 fatality in openFDA: drug-induced liver injury (DILI), nephrotoxicity, neutropenia, heart
168 attack, pleural effusion, and pancreatitis (**Fig.S1A**). We began by using the framework
169 described in **Fig.1A** to define a training set of safe and toxic drugs for each adverse event
170 and its corresponding tissue. For the toxic drugs, we directly queried the database for
171 drugs that are linked to each adverse event or its synonyms. We then took drugs that are
172 not associated with any adverse event in the related tissue or any other severe adverse
173 events to be the set of safe drugs (**Fig.S1B**). The set of keywords used to construct these
174 training sets are described in **Table 1**.

175
176 Building upon the framework of ProCTOR, MAESTER integrates 13 structural features,
177 35 target and tissue features, and 8 drug similarity properties to produce a suite of
178 classifiers that are able to predict the likelihood of each adverse event (**Fig.4A**). Given

179 the established validity of drug-likeness measures in capturing toxicity, we also included
180 properties considered by the Lipinski⁸, Veber¹⁷, and Ghose¹⁸ rules, and the Quantitative
181 estimate for Drug-Likeness (Q.E.D.)⁹ as well as the measures themselves. For tissue-
182 based features, we considered the number of known drug targets that fall in the
183 associated tissue-specific safe and toxic gene sets we created earlier. We also included
184 the above described tissue expression features from GTEx¹⁹, network properties
185 (connectivity and degree), and loss of function mutation frequency²⁰. Finally we integrated
186 the different similarity scores (structural, CMAP, NCI60, and bioassay) through two
187 different measures. The first similarity metric represents whether the drug is more similar
188 to known safe or toxic molecules by using a signed Kolmogorov-Smirnov D-statistic. The
189 second similarity metric is a count of the number of highly similar drugs with known
190 TSAEs.

191
192 The classifiers were evaluated using 10-fold cross validation. All adverse events achieved
193 significant predictive performances with an average accuracy of 72% and area-under-
194 the-receiver-operator curve (AUC) of .81 (**Fig.4B, Table 2**). Focusing specifically on
195 neutropenia – a major cause of clinical trial failure and mortality in cancer and
196 immunocompromised patients²¹– MAESTER achieved an AUC, accuracy, specificity, and
197 sensitivity of 0.8843, 0.7839, 0.7778 and 0.7891 respectively (**Fig.4C, Table 2**) – to our
198 knowledge the highest reported results for the computational prediction of neutropenia.

199
200 ***MAESTER identifies adverse events in independent test sets***
201 We further assessed MAESTER's performance using an independent validation test set.
202 For liver toxicity, the FDA has curated the Liver Toxicity Knowledge Base (LTKB) that
203 classifies a number of compounds based on their risk of causing liver toxicity. We found
204 that MAESTER can significantly distinguish drugs that are of DILI-concern from those
205 classified as no concern using this independent database (**Fig.4D**) ($p < 2.2e-16$, Mann-
206 Whitney U test). For heart attacks, pleural effusion, and neutropenia we turned to FDA
207 drug label warnings as reported in openFDA. We found that MAESTER correctly identified
208 76.3% of drugs with heart attack risk ($p=0.04589$, Binomial test), 75.0% with pleural
209 effusion risk ($p=0.01474$, Binomial test), and 87.5% with neutropenia risk ($p=0.0782$,
210 Binomial test) (**Fig.4E**). These tested compounds did not have their specific adverse
211 events listed in SIDER and thus were not in our original training set, further highlighting
212 MAESTER's potential to predict adverse events for new compounds.

213
214 A feature importance analysis revealed that there is a subset of features that were
215 consistently predictive across all of MAESTER's adverse event models (**Fig.S2A**). The
216 toxic and safe gene sets, structural and bioassay similarity features, polar surface area,
217 and expression of the drug target in mature B cells are important in a majority of models.
218 We also identified a subset of features that are uniquely predictive in specific models. For
219 example, target expression in digestive organs (e.g., colon, small intestine, stomach)
220 were highly important in the prediction of DILI (**Fig.S2B**), expression in immune-related
221 cells (centroblasts, T cells, spleen) were important for neutropenia prediction (**Fig.S2C**),

222 and the network degree of the drug target was the most important feature in prediction of
223 pleural effusion (**Fig.S2D**).

224
225 We then compared the predictions for drugs across all models (**Fig.S3A**). We found that
226 there were subsets of drugs that are predicted to be safe or toxic by most or all models.
227 We found that drugs predicted to have many TSAEs tended to have higher predicted
228 toxicity levels (measured by the ProCTOR score) (**Fig.S3B**) than drugs that were
229 predicted to have one or less TSAEs (**Fig.S3C**, $p=1.178e-06$, Mann-Whitney U test).

230

231 ***MAESTER predicts specific adverse events for withdrawn drugs***

232 To test MAESTER's ability to detect adverse events that may have been missed by
233 traditional approaches, we next focused on drugs that been approved but were later
234 withdrawn due to toxicity concerns. This is especially relevant because cardiotoxicity and
235 hepatotoxicity – two of MAESTER's adverse event models – are the largest causes of
236 toxicity related withdrawal²². We began by focusing on two well-known cases of drug
237 withdrawal – Vioxx and Avandia, both withdrawn for cardiac toxicity– and found that
238 MAESTER scored each as highly likely to cause cardiac toxicity (**Fig.5A-B**). In fact,
239 comparing Avandia (Rosiglitazone) to a less toxic analog (Pioglitazone) we observed that
240 the difference in reported toxicities corresponded to a difference in their MAESTER
241 scores. We found that these predictions did not change substantially when we removed
242 both drugs (and their analogs) from the original training set, retrained MAESTER's
243 underlying model, and rescored each compound. To further expand this analysis we
244 curated a list of withdrawn drugs (that were not part of MAESTER's original training set)
245 and their reason for withdrawal (**Methods**). For each drug we computed a MAESTER
246 probability corresponding to the specific reason for withdrawal (**Table 3**). We found that
247 for 87.5% of the withdrawn drugs MAESTER predicted that specific adverse event with a
248 probability greater than 0.5 – significantly more than would have been expected by
249 random chance ($p=0.0003$, Fisher's exact test). To further evaluate MAESTER's ability
250 to flag withdrawn drugs, we compared MAESTER probabilities of withdrawn drugs against
251 probabilities for drugs of similar indications that were never withdrawn and were not
252 known to have the reported adverse event (**Fig.5C-F**). We found that withdrawn drugs
253 had significantly higher MAESTER adverse event probabilities than approved drugs of
254 the same indication ($p=0.0027$ and 0.0424 , Fisher's exact test). Overall these results
255 highlight MAESTER's ability to specifically identify compounds with adverse events that
256 were missed by traditional approaches.

257 **DISCUSSION**

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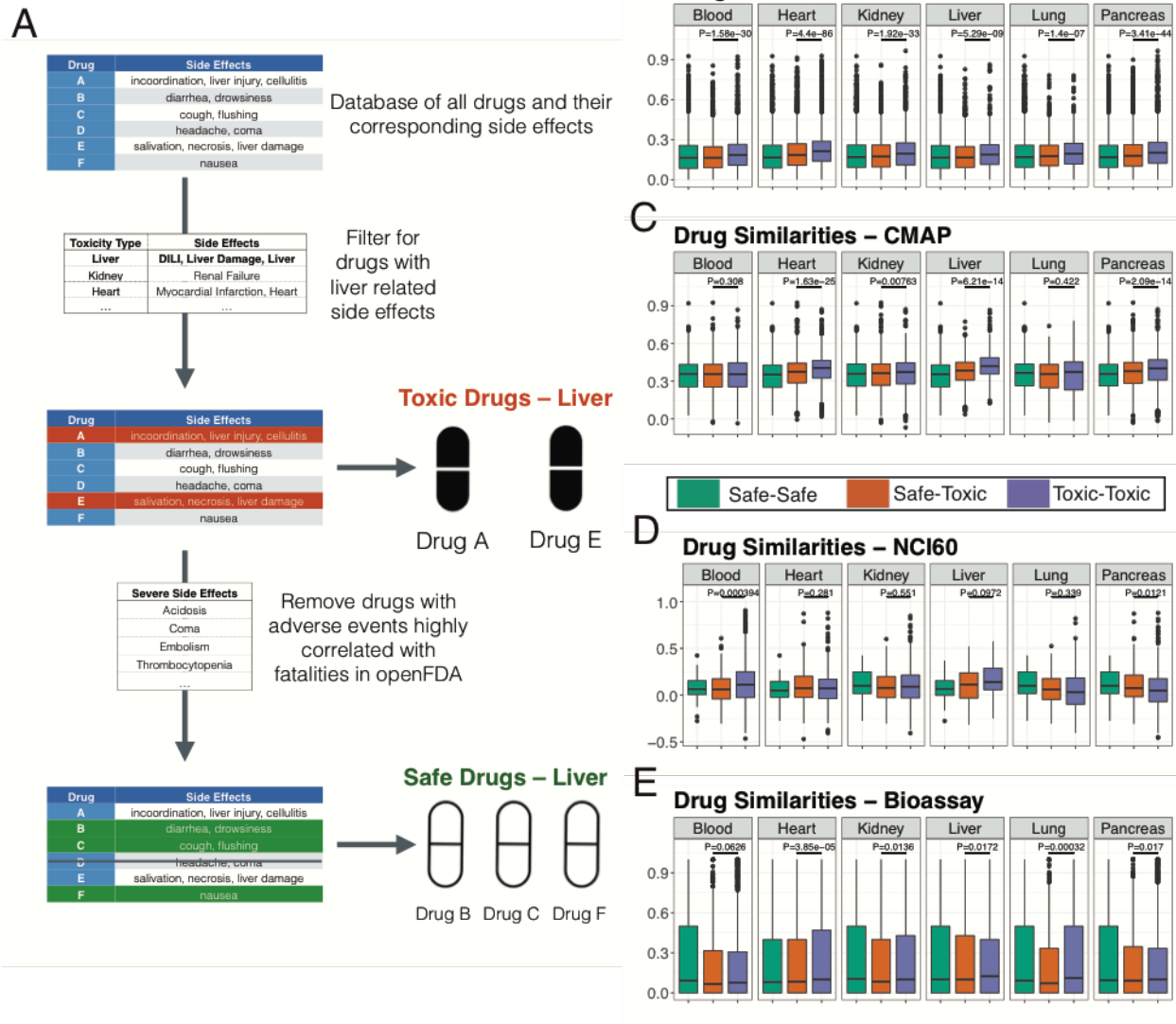
259 Pre-clinical toxicity screening is one of the most important parts of drug development.
260 Existing experimental methods are cumbersome and often do not translate to clinical
261 results. Computational methods for predicting toxicity can complement and perhaps guide
262 experimentation to evaluate toxicities. However prior methods have for the most part
263 focused only on molecular properties and predicting broad clinical toxicities rather than
264 specific adverse events. We have proposed MAESTER, a data-driven machine learning

265 approach that integrates information on a compound's structure, targets, and downstream
266 effects to predict the probability of a compound presenting with different adverse events.
267 When trained on drugs with known adverse events, MAESTER performs at high
268 accuracy, sensitivity, and specificity across six different prediction tasks. Additionally
269 MAESTER performs with high accuracy on external FDA test sets and drug warning
270 labels, and could accurately identify adverse events for withdrawn drugs that may have
271 been missed during traditional analyses.

272
273 We have identified sets of toxic and safe drugs and genes that are associated with
274 adverse events in specific tissues. We found that tissue-specific toxic drugs tend to be
275 more similar to each other than known safe drugs and that their associated targets are
276 more highly expressed in corresponding tissues. We found tissue-specific toxic targets
277 tend to be enriched for apoptosis and cell death related biological processes, more
278 connected in protein-protein interaction networks, and are classified as more essential.
279 Leveraging this data, we developed MAESTER to combine compound and target
280 properties to predict the likelihood of specific adverse events. Because it is trained on
281 drugs with known adverse events, MAESTER can directly predict clinical effects
282 compared to cell or animal screening methods whose toxicity predictions may not
283 translate to the clinic.

284
285 One of the strengths of our big data approach is that it can consider a large number of
286 features without prior bias. This will become especially powerful in the coming years as
287 more large pharmacogenomics datasets become available to integrate. Analysis of these
288 features can aid in future drug design by providing insight into what types of drugs are
289 likely to be toxic and feeding this information back to the chemists. Additionally, while
290 toxicity is often modeled as a broad feature, often times it is a patient specific effect. As
291 more patient specific data becomes available MAESTER can be improved to predict
292 patient specific adverse events. This could be used to guide clinical trial design by
293 specifically selecting patients unlikely to present with toxic effects and radically change
294 how people approach precision medicine.

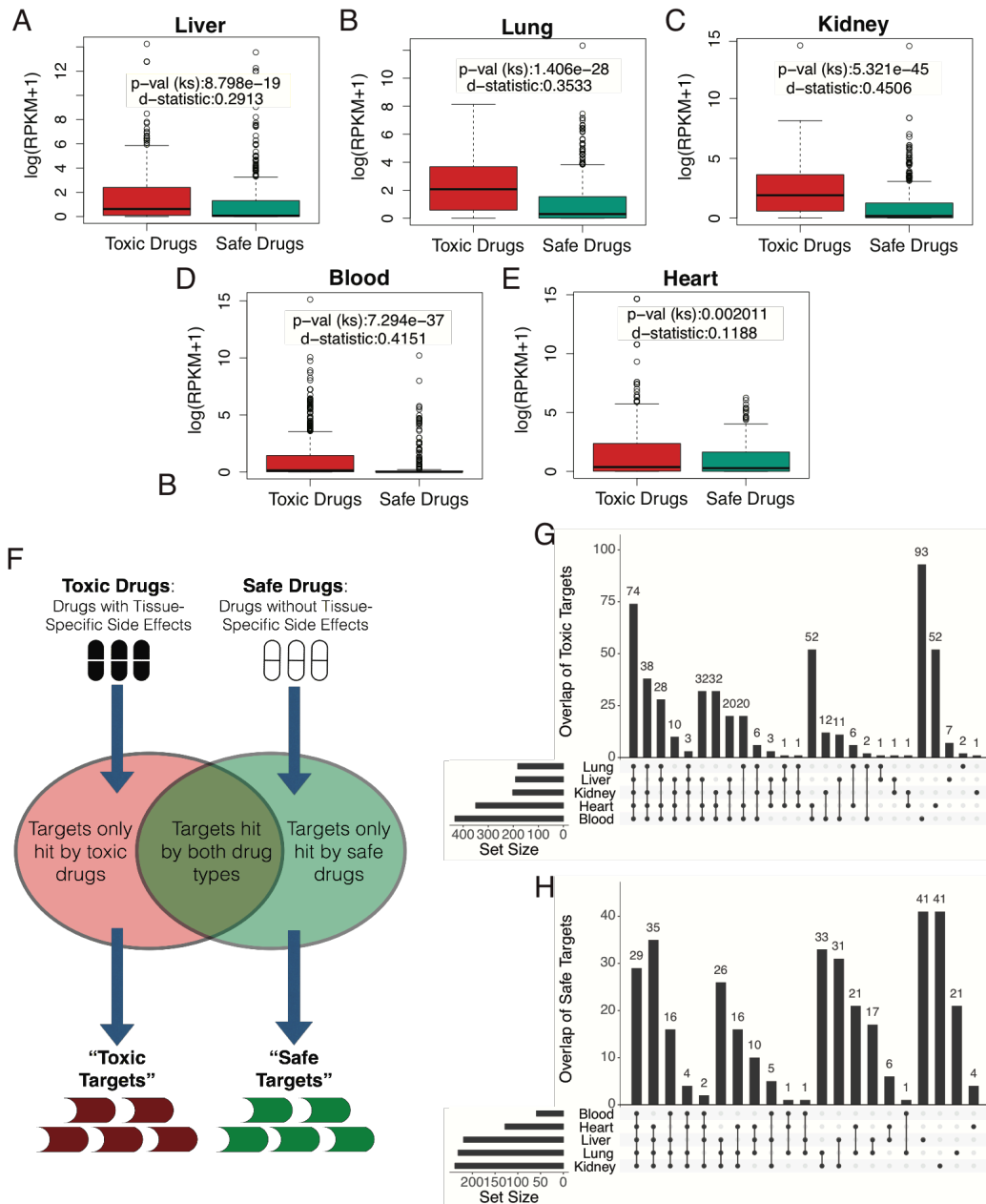
Figure 1



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Figure 1 – A) Schematic describing the process by which we selected our toxic and safe drugs for each specific tissue. B) Similarities of across all toxic drugs pairs, safe drug pairs, and all combinations of toxic and safe drugs for drug structures, C) gene expression changes, D) growth efficacies, and E) bioassays. P values were calculated using a Wilcoxon Rank Sum test.

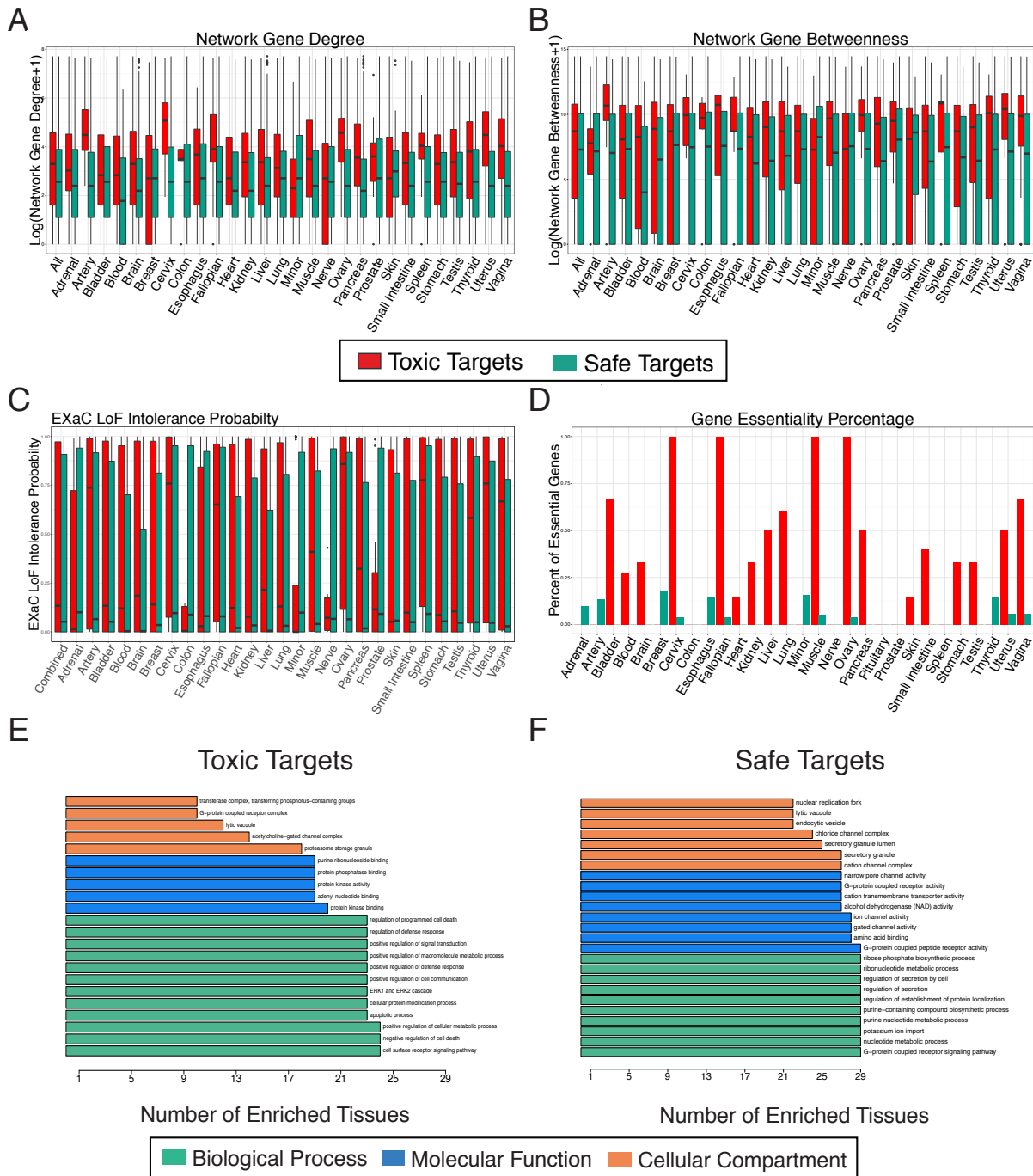
Figure 2



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302 **Figure 2** – A–E) Distribution of target expression in a specific tissue for drugs
 303 with and without any tissue-specific adverse events (in that given tissue). P values and
 304 D statistic calculated using a KS test. F) Schematic for the selection of toxic and safe
 305 targets. G) UpSetR plot highlighting the overlap across tissue types for their respective
 306 toxic and H) safe targets.

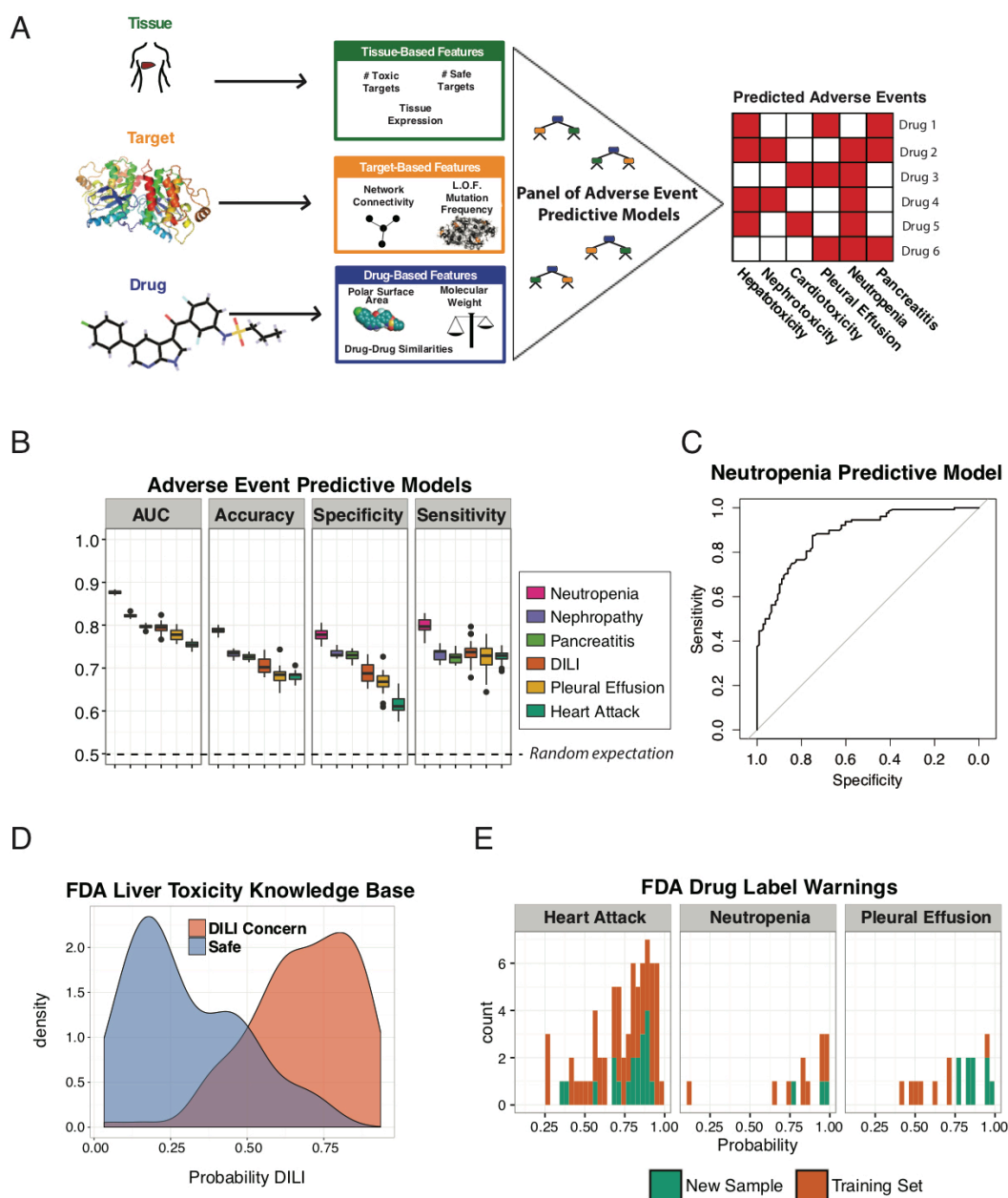
Figure 3



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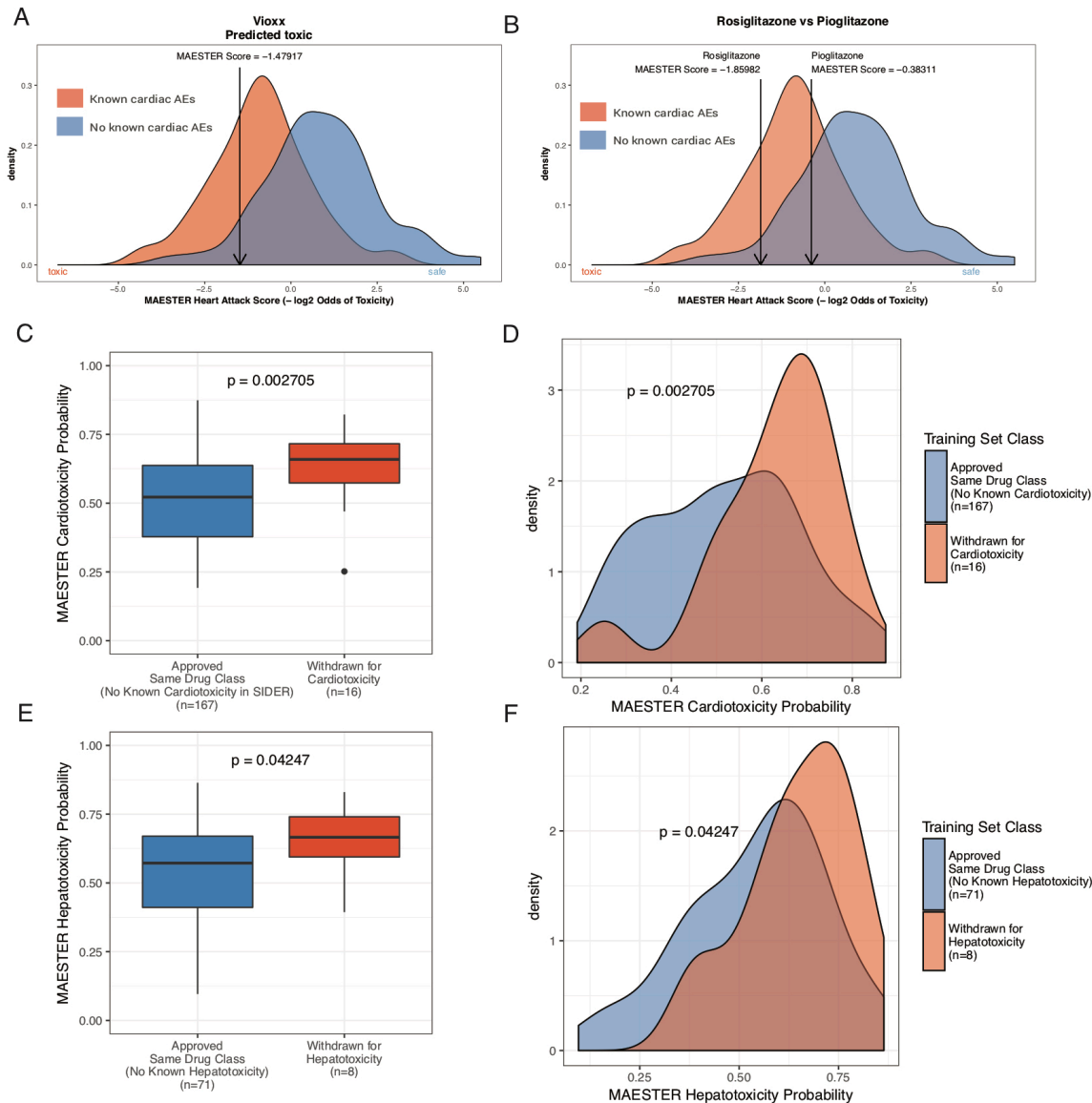
Figure 3 – A-D) Distribution of features across multiple tissues for their individual toxic and safe targets. E) Number of tissues whose respective toxic or F) safe targets are enriched for a specific Gene Ontology category.

Figure 4



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 312 **Figure 4** – A) Schematic of MAESTER’s method of integrating multiple feature types to
 313 predict tissue-specific adverse events. B) Performance metrics for multiple MAESTER
 314 prediction models. C) Area under the receiver-operating curve for MAESTER’s
 315 Neutropenia model. D) Distribution of MAESTER DILI probabilities for drugs marked as
 316 “DILI Concern” or “Safe” by the FDA Liver Toxicity Knowledge base. E) MAESTER
 317 Predictions for drugs with FDA warning labels for heart attacks, neutropenia, or pleural
 318 effusion.

Figure 5



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Figure 5 – A) Distributions of MAESTER scores for all drugs known to cause heart attacks and those considered safe. MAESTER scores for Vioxx, B) Rosiglitazone, and Pioglitazone are indicated with arrows. C-D) MAESTER scores for drugs withdrawn for cardiac toxicity compared to approved drugs of the same class with no known cardiac toxicities. E-F) MAESTER scores for drugs withdrawn for liver toxicity compared to approved drugs of the same class with no known liver toxicities. All p values were calculated using a Wilcoxon rank sum test.

327 **Table 1 – MAESTER Training Set Definitions.** Table of the 6 major adverse event
328 categories. In addition to the given adverse event, certain synonymous adverse events
329 were also included and any drugs with containing an adverse event in the “other
330 removed terms” category were removed excluded from the safe set.

Adverse Event	Synonyms	Tissue	Other Removed Terms
DILI	Liver Disease, Liver Injury, Liver Damage	Liver	"Nephro*"
Heart Attack	Myocardial Infarction	Heart	"Immun"
Renal Failure	Kidney Failure	Kidney	
Neutropenia	-	Blood	
Pleural Effusion	-	Lung	-
Pancreatitis	-	Pancreas	-

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332 **Table 2 – MAESTER Model Performances.** AUROC, Accuracy, Specificity, and
333 Sensitivity values for each of MAESTER’s underlying models.

Adverse Event	# safe drugs	# toxic drugs	AUROC	Accuracy	Specificity	Sensitivity
DILI	105	268	0.8079	0.7373	0.7333	0.7388
Heart Attack	113	166	0.7552	0.6882	0.6283	0.7289
Renal Failure	127	165	0.8198	0.7329	0.7087	0.7515
Neutropenia	108	128	0.8843	0.7839	0.7778	0.7891
Pleural Effusion	128	59	0.7761	0.6631	0.6562	0.678
Pancreatitis	126	153	0.7967	0.7348	0.7302	0.7386

334

335 **Table 3 – MAESTER Performance on Withdrawn Drugs.** List of withdrawn drugs,
336 their reason for withdrawal, and the corresponding MAESTER score.

Drug	Specific MAESTER Probability*	Reason for Withdrawal
Sitaxentan	0.83	Hepatotoxicity
Sparfloxacin	0.822	Cardiotoxicity
Flecainide	0.772	Cardiotoxicity
Nialamide	0.76	Hepatotoxicity
Dexfenfluramine	0.738	Cardiotoxicity
Acetohexamide	0.734	Hepatotoxicity
Cisapride	0.716	Cardiotoxicity
Tegaserod	0.716	Cardiotoxicity
Bepidil	0.714	Cardiotoxicity
Tolrestat	0.708	Hepatotoxicity
Alprenolol	0.682	Cardiotoxicity
Fenfluramine	0.664	Cardiotoxicity
Encainide	0.654	Cardiotoxicity
Nimesulide	0.624	Hepatotoxicity
Sertindole	0.62	Cardiotoxicity
Nomifensine	0.616	Hepatotoxicity
Hexylcaine	0.614	Cardiotoxicity
Mibefradil	0.588	Cardiotoxicity
Astemizole	0.53	Cardiotoxicity
Zimelidine	0.53	Hepatotoxicity
Prenylamine	0.512	Cardiotoxicity
Terfenadine	0.47	Cardiotoxicity
Ximelagatran	0.394	Hepatotoxicity
Dextropropoxyphen e	0.252	Cardiotoxicity

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338 * = Probability corresponds to probability of presenting with either cardiac or
339 hepatotoxicity depending on the reason for withdrawal

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