

1 **Title:** Natural variation in *couch potato* mediates rapid evolution of learning and reproduction in
2 natural populations of *Drosophila melanogaster*

3
4 **Authors:** Emily L. Behrman^{1,2,3}, Tadeusz J. Kawecki^{2*} and Paul Schmidt^{1*}

5
6 **Author Affiliations:**

7 1. Department of Biology, University of Pennsylvania, 433 S. University Ave. Philadelphia, PA
8 19104

9 2. Department of Ecology and Evolution, University of Lausanne, Biophore, CH-1015
10 Lausanne, Switzerland

11 3. Current address: Janelia Research Campus, Howard Hughes Medical Institute, 19700 Helix
12 Drive Ashburn, VA 20147

13 * Authors contributed equally

14
15 **Orcid IDs:**

16 Emily Behrman: 0000-0002-2472-9635

17 Tadeusz Kawecki: 0000-0002-9244-1991

18 Paul Schmidt: 0000-0002-8076-6705

19
20
21 **Running head:** Rapid learning evolution

22
23 **Corresponding Author:**

24 Emily L. Behrman

25 Email Address: Bemily@sas.upenn.edu

26 Mailing address: 19700 Helix Drive

27 Ashburn, VA 20147

28 Phone: 571.209.4000

29
30 **Keywords**

31 Rapid adaptation, learning, fecundity, trade-offs, *Drosophila melanogaster*, *couch potato*

32

33

34

35

36 **Abstract**

37 Seasonal oscillations in temperate environments between the different selection regimes
38 of winter and summer produce cyclical selection pressures that may drive rapid evolution of
39 diverse traits. We investigated the evolutionary dynamics of learning ability in natural
40 populations over this rapid seasonal timescale. Associative learning was tested in common
41 garden-raised *Drosophila melanogaster* collected from a natural population in the spring and fall
42 over three consecutive years. The spring flies learned consistently better than fall flies, revealing
43 seasonal evolution of improved learning performance in nature. Fecundity showed the opposite
44 seasonal pattern, suggesting a trade-off between learning and reproduction, which we confirmed
45 at the level of individual females. This trade-off is mediated at least in part by natural
46 polymorphism in the RNA binding protein *couch potato* (*cpo*), with a haplotype favored during
47 summer showing poorer learning performance and higher fecundity than a haplotype favored
48 over winter. Thus, seasonal environments can drive rapid cyclical evolution of learning
49 performance, but the evolutionary dynamics may be driven by pleiotropic effects of alleles
50 selected for other reasons.

51

52

53 **Impact statement**

54 Evolution is traditionally considered to be a very slow, gradual process, but recent studies show
55 that some organisms evolve rapidly for a variety of traits. However, there is still little known
56 about the rate at which behaviors evolve in the wild. Complex behaviors may evolve due to their
57 fitness benefits or because natural selection acts on a trait that is genetically correlated with that
58 behavior. Learning ability is an important behavior for many aspects of an organism's biology,

Rapid learning evolution

59 but may be costly and use resources that could be otherwise allocated to other important
60 functions, such as reproduction. We measure learning ability in fruit flies collected from a
61 natural population as it evolves across seasons. We find that the complex behavior of learning
62 ability evolves very fast the wild at the cost of reproduction: the spring populations have higher
63 learning ability but lay fewer eggs compared to the fall. We found that natural variants of the
64 *couch potato* help mediate the trade-off between reproduction and learning in natural
65 populations. This shows how evolution of complex traits can occur due to selection on genes that
66 affect multiple traits.

67

68

69 **Introduction**

70 It is becoming clear that many life history traits can evolve very rapidly over time scales
71 that were previously assumed to be constant (Kingsolver et al. 2001; Grant and Grant 2002;
72 Schmidt and Conde 2006; Carroll et al. 2007; Thompson 2013; Behrman et al. 2015, 2018), but
73 little is known about the rate at which behaviors evolve. Behaviors may be less prone to evolve
74 rapidly because behavior can be highly plastic and shaped by learning. Learning is important for
75 many aspects of animal biology including foraging, spatial orientation, predator avoidance,
76 aggression, social interactions, and sexual behavior (Yurkovic et al. 2006; Dukas 2008; Stensmyr
77 et al. 2012; Mansourian et al. 2016). Learning decouples behavioral phenotypes from genotypes
78 by allowing individuals to develop adaptive behavioral responses to changing environments.
79 Under a narrow set of circumstances learning may accelerate genetically-based evolutionary
80 change of behavior (Mayr 1974; Mery and Kawecki 2004b; Paenke et al. 2007), but under most
81 circumstances it is predicted to slow the rate at which behaviors evolve (Dukas 2004; Paenke et
82 al. 2007).

83 The ability to learn is itself a product of evolution, but we know next to nothing about
84 how rapidly its characteristics evolve in nature. Examples of genetic differences in learning
85 performance have been reported between closely related species (Odling-Smee et al. 2008;
86 Hoedjes and Smid 2014) and between conspecific populations (Croston et al. 2015; Froissart et
87 al. 2017), but the timescale over which these differences have evolved is unknown. Rapid
88 evolution of improved learning performance in laboratory selection experiments in rats, bees,
89 blowflies and *Drosophila* indicates that natural populations have copious standing genetic
90 variation for learning ability (Tryon 1940; McGuire and Hirschth 1977; Brandes et al. 1988;
91 Mery and Kawecki 2002; Zwoinska et al. 2017). However, in those experiments learning was

Rapid learning evolution

92 directly under selection, whereas selection on learning in nature is indirect and mediated by the
93 fitness consequences of behaviors it modifies. An environmental change that results in an
94 increased need for learning in one behavior (e.g., foraging) might have no effect, or even
95 decrease, its benefits in a different context (e.g., predator avoidance), dampening changes in net
96 selection on learning.

97 The evolutionary dynamics of learning ability may be driven by fitness consequences of
98 the behavioral modifications it causes or by selection on genetically correlated traits. Correlated
99 selection due to trade-offs may arise because resources that might be invested in growth,
100 reproduction, maintenance or defense are diverted to develop and maintain the energetically
101 costly neural tissue required for learning and memory (Johnston 1982; Mery and Kawecki 2005;
102 Dukas 2008). The hypothesized reproductive cost of learning (Johnston 1982) is supported by
103 genetic trade-offs between learning and reproduction in the cabbage white butterfly, *Pieris*
104 *rapae*, (Snell-Rood et al. 2011) and an operant reproductive cost of learning in *Drosophila* (Mery
105 and Kawecki 2004). Thus, rapid evolutionary change of learning ability could be triggered by
106 environmental changes that alter selection on either learning ability or on correlated traits such as
107 fecundity.

108 Multivoltine species are a promising system to study rapid evolution because each
109 generation may experience significantly different conditions across seasonal time. The
110 alternating conditions between different selection regimes of winter and summer drive rapid
111 cyclical evolution of *Drosophila melanogaster* life history, stress resistance and immunity traits
112 with a hypothesized reproductive trade-off (Schmidt and Conde 2006; Behrman et al. 2015,
113 2018). Here we investigate if learning performance evolves rapidly over an annual timescale in a
114 natural population of *D. melanogaster* and if reproductive costs of learning affect the dynamics

115 in the wild. We predicted learning ability of wild *Drosophila* to vary with season as the
116 behaviors that involve learning change over seasonal time. Increased ability to learn may be
117 favored in the summer as many behavioral tasks associated with increased summer fitness are
118 modulated by learning in *Drosophila* laboratory studies: foraging, pathogen avoidance, and
119 sexual and social interactions (Dukas 1998, 2005; Sarin and Dukas 2009; Battesti et al. 2012;
120 Zrelec et al. 2013; Hollis and Kawecki 2014). However, it is possible that learning may be
121 involved behaviors that are important for overwintering survival (e.g., shelter identification,
122 pathogen avoidance). Alternatively, if selection acts on correlated traits, then seasonal variation
123 in learning ability could be driven by these indirect effects – particularly on fecundity in the
124 summer (Mery and Kawecki 2004) and stress tolerance in the winter (Mery and Kawecki 2005).

125 We assessed aversive and appetitive learning in common-garden-raised flies derived from
126 wild spring and fall field collections over three consecutive years. We found that learning
127 performance was consistently higher in the post-winter collections in the spring compared to the
128 post-summer collections in the fall. An inverse relationship between learning ability and
129 fecundity between seasonal collections and at an individual level indicated that a reproductive
130 cost of learning was involved in the seasonal dynamics. This pointed to a potential role of *couch*
131 *potato* (*cpo*), a pleiotropic RNA-binding protein highly expressed in the nervous system
132 including the mushroom body (Bellen et al. 1992a,b). Fecundity and reproductive diapause
133 differences in natural *cpo* variants correlate with differential *cpo* expression; the frequency of the
134 variants in wild *Drosophila* changes with latitude and season (Behrman et al. n.d.; Schmidt et al.
135 2008; Kolaczowski et al. 2011; Fabian et al. 2012; Cogni et al. 2013; Bergland et al. 2014). We
136 found that learning performance and fecundity in flies carrying natural *cpo* haplotypes that are
137 more common in the spring versus the fall paralleled the seasonal pattern in the natural

138 population. These results suggest that rapid fluctuating evolution of learning ability in wild
139 *Drosophila* is at least in part driven by pleiotropic effects of *cpo* polymorphism, which mediates
140 a trade-off between reproduction and traits that promote overwinter survival.

141

142 **Methods**

143 ***Drosophila* samples**

144 *Seasonal populations derived from nature*

145 Seasonal differences in learning were assessed by comparing outbred seasonal
146 populations reconstructed from isofemale lines, hereafter referred to as spring and fall
147 populations. Gravid females aspirated off of decaying fruit in the spring (June) and fall
148 (November) at Linvilla Orchards in Media, PA (39.9°N, -75.4°E), across three consecutive years
149 (2012-2014) were used to establish isofemale lines that were maintained in standard laboratory
150 conditions (25°C, 12L:12D) on a four-week transfer cycle. After all collections were complete,
151 representative populations from each collection were re-constructed using 40 isofemale lines per
152 collection and were maintained in common garden culture for more than 10 non-overlapping
153 generations. We infer that differences in learning ability among the populations tested in the
154 standard laboratory environment are due to genetic differences among the populations.

155

156 *cpo* recombinant outbred populations

157 Three SNPs in *cpo* were used as markers for the temperate haplotype (*cpo*^{TTA}) that is
158 more common at high latitudes and in the spring versus the tropical haplotype (*cpo*^{CGT}) that is
159 more common at low latitudes and in the fall. (Behrman et al. n.d.): two intronic SNPs
160 (3R:13790130 and 3R:13791280) and one putative non-synonymous coding change

161 (3R:13793588, *D. melanogaster* reference genome v.5.39, (Cogni et al. 2013; Bergland et al.
162 2014). The non-synonymous coding change is validated using in situ hybridization (Bellen et al.
163 1992a) but the exon is not included in MODENCODE (Celniker et al. 2009). Learning
164 performance of the haplotypes was assessed using recombinant outbred populations (ROPs;
165 (Paaby et al. 2014; Behrman et al. 2018, n.d.)) that were each fixed for one haplotype with a
166 randomized genetic background. ROPs for *cpo*^{TTA} and *cpo*^{CGT} were constructed using nine
167 independent, homozygous *Drosophila* Genetic References Panel (MacKay et al. 2012) lines. Ten
168 gravid females from each line were permitted to lay eggs for 48h; after at least 10 non-
169 overlapping generations of recombination with the other lines containing the same haplotype
170 among the offspring, each ROP was fixed for either *cpo*^{TTA} or *cpo*^{CGT} in a heterogeneous, outbred
171 background.

172

173 *cpo* knockdown

174 We tested if *cpo* regulation changes learning ability because the *cpo* ROPs have different
175 levels of whole-body *cpo* expression (Behrman et al. n.d.) and other traits are correlated with
176 differential *cpo* expression (Schmidt et al. 2008). The UAS/GAL4 system was used to express
177 dsRNA for RNA interference (RNAi) to knockdown *cpo* expression. Two biological replicate
178 UAS constructs were created using different insertion sites paired with their respective insertion
179 site controls located on the second and third chromosomes, respectively, from the Transgenic
180 RNAi Project (TRiP): (BDSC-60388 with attP40 control BDSC-36304) and (BDSC-28360 with
181 attP2 control BDSC-36303). The hypothesis that lower *cpo* expression increases learning was
182 tested using lines that drive GAL4 expression in the mushroom, ellipsoid and fan shaped bodies,
183 subesophageal ganglion, antennal & optic lobes, protocerebrum & median bundle (BDSC-

184 30818). Three *cpo*-specific steroid-activated Gal4 geneswitch drivers were used knock down *cpo*
185 expression in different neuron combinations of adult flies: trachea-associated cells and subsets of
186 ventral nerve cord and sensory neurons (BDSC-40315), a subset of sensory neurons (BDSC-
187 40319) and sense organ support cells and subsets of ventral nerve cord, brain and sensory
188 neurons (BDSC-40334). BDSC-38461 was used as a negative control to express GAL4 in flight
189 muscles under control of the Actin 88F promotor. All crosses were made with the female
190 containing the GAL4 driver and male containing the UAS construct.

191

192 **Learning**

193 *Aversive shock learning in the laboratory*

194 Flies were conditioned to associate one of two odorants with an aversive mechanical
195 shock in an aversive olfactory learning assay (Mery and Kawecki 2005). Flies were reared in
196 common laboratory conditions (25°C, 12L:12D) at standardized density of 100 eggs per vial,
197 sorted into groups by sex under light CO₂ anesthesia 24h prior to the learning assessment, and
198 then assayed at 3-5 days of age. Single-sex groups of 30 flies were conditioned to associate
199 either methylcyclohexanol (MCH, 800 uL/L) or 3-octanol (OCT, 600 uL/L) with a mechanical
200 shock (CS⁺). Three cycles of conditioning were conducted, each consisting of the sequence: 30 s
201 exposure to one odor paired with mechanical shock (pulsating 1s every 5s), 60 s break of humid
202 air, 30 s exposure to other odor with no shock (CS⁻), 60 s humid air. We tested odorant
203 preference of the conditioned flies after a 1 h retention interval by giving them 60 s to choose
204 between odorants in an elevator T-maze. The *cpo* haplotypes were also assessed for immediate
205 response (5 min retention) and long-term memory (24 h retention). Half of the fly groups were
206 conditioned to avoid each odorant to account for innate preference for either odor.

207 To exclude impairment in odor perception or innate preference we performed two
208 additional controls. Controls for absolute preference subjected flies to mechanical shock cycle
209 without odorant before presenting a choice between a single odorant and air in the T-maze.
210 Controls for relative preference tested innate odor preference in naïve flies by giving a choice
211 between the two odors in the T-maze.

212

213 *Appetitive learning in a natural environment*

214 Appetitive conditioning learning assays were adapted from previously developed food
215 substrate-based protocols (Mery and Kawecki 2002; Zrelec et al. 2013), but were implemented in
216 a natural setting in an experimental orchard over three replicate days. Flies were reared in
217 standard laboratory conditions at controlled density and 3d cohorts were sorted into groups of 50
218 by sex. Flies were marked by treatment using fluorescent powder according to learning treatment
219 and source population and kept for 12h on an agar substrate. The flies were then exposed to
220 either strawberry or apple food for 8h and had a 4h rest period on a fresh agar substrate before
221 being released into the outdoor testing phase. Four hundred flies from each treatment, season and
222 sex combination were released together into the same outdoor mesocosm, a 0.6x0.6x1.8m mesh
223 cage with plant bedding covering the ground. Each mesocosm was placed underneath a peach
224 tree inside a larger 8 m³ cage. Eight pairs of strawberry and apple unidirectional traps were
225 dispersed around the mesocosm and flies were scored as learning if they selected the same food
226 type that they had been previously exposed to during the conditioning period. The powder
227 marking was an effective method of labeling the flies and all of the trapped flies showed traces
228 of the powder; this also indicates that no additional flies infiltrated the experiment from outside.

229

230 **Fecundity**

231 Fecundity of the reconstructed seasonal populations was measured by placing 25 virgin
232 females from each reconstructed seasonal population into culture bottles (Paaby et al. 2014) with
233 25 males from a standardized stock (BDSC-3605). Food plates were changed daily for ten days
234 to count the number of eggs. Average fecundity was calculated daily across 10 replicate bottles
235 per population.

236 Individual trade-off between learning and reproduction was assessed in the spring
237 reconstructed seasonal populations from 2013 and 2014. Virgins were collected, aged in vials of
238 standard food for 3 days, and subject to the aversive olfactory conditioning and test described
239 above. The flies were thus divided in "learners" (those that chose CS-) and "non-learners" (those
240 that chose CS+). From each replicate test, five "learner" and five "non-learner" females were
241 placed onto food with 5 males; all available flies were used in the few replicates with less than
242 five flies. The flies were housed on food supplemented with topical yeast to promote egg
243 production before the assay and on grape-agar substrate during the egg collection. Daily
244 fecundity was counted for 3d post learning assessment and the mean fecundity was calculated
245 based on the number of females in the vial.

246

247 **Statistical analysis**

248 All statistical analyses were performed using the R software (R Core Team, version R
249 3.2.2). To analyze learning performance, we modeled proportion of flies selecting the odor
250 against which they had been conditioned (CS⁺) using generalized linear models with mixed
251 effects fit by maximum likelihood (package *lme4*, Bates et al. 2014). To test for the significance
252 of main effects and interactions, we used type II Wald chi-square test from the package *car* and

Rapid learning evolution

253 functional analysis of variance (ANOVA, Fox and Weisberg 2011). Flies that chose no odor and
254 remained in the center of the T-maze were excluded from analysis. For illustration purposes, the
255 proportion of flies learning was rescaled as $2 \times \text{proportion} - 1$ to make it range from -1 to 1 with
256 0 indicating no learning.

257 For the seasonal populations, we used the following model:

258

259 $\text{Response} = \text{Season} + \text{Year} + \text{Direction of conditioning} + \text{Replicate}$

260

261 where Season, Year and Direction of conditioning are fixed effects and Replicate is a random
262 effect.

263

264 The effect of the natural *cpo* haplotypes was determined using the following model:

265

266 $\text{Response} = \text{Genotype} + \text{Sex} + \text{Direction of conditioning} + \text{Retention Interval} + \text{Replicate}$

267

268 where Genotype, Sex, Direction of conditioning and Retention Interval are fixed effects and
269 Replicate is a random effect.

270

271 To assess the RNAi knockouts, we used the following model:

272

273 $\text{Response} = \text{cpo} * \text{Gal4} * \text{Direction of conditioning} + \text{UAS} + \text{Replicate}$

274

275 where *cpo* (presence or absence), Gal4 (location), Sex and Direction of conditioning are fixed
276 effects and UAS driver and Replicate are random effects. Tukey's Honestly Significant
277 Difference test was used as a post-hoc evaluation of the effect of *cpo* in specific tissues using the
278 lsmeans package (Lenth 2016) in R.

279 Population measurement of fecundity was assessed using mixed model ANOVAs with
280 the fixed effects of season and year (seasonal populations) and genotype (*cpo* ROP) and the
281 random effect of replicate bottle. Individual fecundity of flies that learned compared to those that
282 did not learn was calculated using a paired T-test.

283

284 **Results**

285 *Rapid evolution of learning and fecundity in a natural population*

286 Learning ability in the wild *D. melanogaster* population evolved rapidly and repeatedly
287 across seasonal time. The spring collections learned better than the fall from the same year when
288 assessed using laboratory aversive conditioning ($\chi^2_1=4.37$, $p=0.036$; Figure 1a). These behavioral
289 differences were not due to differences in ability to perceive odor, as there was no effect of
290 season in absolute preference between odorant and solvent in unconditioned flies ($\chi^2_1=0.24$,
291 $p=0.62$ Figure 1b). The spring collections also tended to show higher appetitive learning in an
292 assay performed in outdoor mesocosms: spring flies were better at learning the association
293 between food and odor as they returned to the known high-quality food source at higher
294 frequency than the fall flies ($\chi^2_1= 2.56$, $p=0.11$, Figure 1c). Taken together, the combined
295 aversive and appetitive conditioning results strengthen the evidence for a higher learning ability
296 of the spring flies ($\chi^2_4 = 11.0$, $p = 0.026$, Fisher's method for combining p -values).

297 The seasonal pattern of fecundity was opposite to that for learning ability at the

298 population level: the fall populations laid on average 30% more eggs per female per day than the
299 spring populations (Figure 1d; $F_{1,57} = 44.2$, $p < 0.0001$). A negative association between learning
300 and reproduction was also apparent at the individual level. Females that did not learn in the
301 aversive learning assay laid an average of six more eggs (25% more) per day compared to the
302 females that avoided the shock-associated odor ($t_{14} = -2.84$, $p = 0.013$, Figure 1e).

303

304 *Natural variation in cpo sequence affects learning and fecundity*

305 Recombinant outbred populations (ROPs) homozygous for the spring *cpo*^{TTA} haplotype
306 had higher aversive learning than the fall ROP; this pattern persisted across all retention intervals
307 between conditioning and testing ($\chi^2_1 = 34.88$, $p = 3.5 \times 10^{-9}$; Figure 2a). In the absence of
308 conditioning, the spring ROPs showed a weaker avoidance of odors paired with air than the fall
309 with 1h between training and conditioning ($\chi^2_1 = 8.46$, $p = 3.6 \times 10^{-3}$, Figure 2b), but not at 5m or
310 24h retention intervals. The *cpo* ROPs did not differ in the relative preference of naïve flies
311 choosing between the odors ($\chi^2_1 = 0.016$, $p = 0.90$, Figure 2c). The spring ROPs also had higher
312 appetitive associative learning when assessed in the natural mesocosms ($\chi^2_1 = 3.62$, $p = 0.057$,
313 Figure 2a). The spring *cpo*^{TTA} ROP was characterized by lower fecundity, with females laying
314 36% fewer eggs than females from the fall *cpo*^{CGT} ROP (Figure 2d, $F_2 = 8.23$, $p = 0.01$).

315

316 *cpo expression in the peripheral nervous system affects learning*

317 The two intronic SNPs in the *cpo* haplotype may regulate *cpo* expression as the flies with
318 the spring *cpo* haplotype have lower full body *cpo* expression than flies that carry the fall *cpo*
319 haplotype (Behrman et al. n.d.). Therefore, we tested if *cpo* expression mediates the differences
320 in learning and found a tissue-specific effect of *cpo* on learning with a significant interaction

321 between *cpo* expression and Gal4 driver tissue ($\chi^2_5=14.26$ p=0.014, Figure 3). There was no
322 difference in learning when *cpo* was knocked down in the Actin 88F promotor control
323 (BSC38461), in the broad brain knockout (BDSC30818), in a subset of ventral nerve cord and
324 sensory neurons (BDSC-40315) or in the sense organ support cells and subsets of ventral nerve
325 cord, brain and sensory neurons (BDSC-40334). Knocking down *cpo* expression in a subset of
326 the sensory neurons (BDSC-40319) decreased learning (Tukey HSD: $z=3.35$ p=0.0008).

327

328 **Discussion**

329

330 *Rapid evolution in natural populations*

331 Learning and fecundity both evolve rapidly in a natural population of *D. melanogaster*
332 over the scale of approximately 10-15 generations from spring to fall and approximately 1-2
333 generations between fall and spring. The differences can be attributed to annual cyclical genetic
334 changes in natural populations because environmental effects are removed by rearing and testing
335 these populations in common laboratory conditions. The repeatability across years indicates that
336 this is not a result of genetic drift but instead a deterministic evolutionary process and genomic
337 data excludes gene flow through migration generating the seasonal cycles (Bergland et al. 2014).
338 Therefore, the rapid and repeatable seasonal changes in learning and reproduction are consistent
339 with seasonally fluctuating selection. Learning is thus not only a mechanism of plasticity that
340 allows organisms to respond rapidly to environmental change (Crombach and Hogeweg 2008);
341 we demonstrate that learning ability itself can evolve rapidly in nature.

342 Learning evolved rapidly across seasonal time with higher learning in the spring
343 compared to the fall. The results are counter to the prediction that learning is favored during the

Rapid learning evolution

344 summer when flies are active and use behaviors that involve learning when assessed in
345 laboratory. However, cognitive abilities may evolve to match the demands posed by an
346 organism's biological and physical environment (Healy and Jones 2002; Smid and Vet 2016).
347 Energetically challenging environments (e.g., cold winters or severe droughts) are hypothesized
348 to favor cognitive performance at the cost of other physiological systems receiving less resources
349 (Maille and Schradin 2016). For example, cache seed recovery success in several bird species
350 suggests that learning may be important for overwintering survival in harsh environments
351 (Bednekoff et al. 1997; Pravosudov and Clayton 2002; Olson et al. 2004). Although flies do not
352 cache food, it is possible that learning is also important for other aspects of *D. melanogaster*
353 overwintering survival in harsh climates, such as the ability to find a suitable overwintering site.
354 Alternatively, rapid evolution of learning may be driven by pleiotropic effects of alleles that
355 fluctuate in frequency for other reasons such as a correlated trade-off with fecundity.

356

357 *Trade-off between learning and fecundity*

358 The rapid evolution of learning and fecundity are consistent with patterns of seasonal
359 evolution of other life history traits in natural populations of *D. melanogaster*. Previous studies
360 have demonstrated that spring populations collected after the winter bottleneck are more
361 vigorous with higher propensity for reproductive diapause (Schmidt and Conde 2006), greater
362 stress resistance (Behrman et al. 2015), higher post-infection survival (Behrman et al. 2018).
363 Here, we show that spring flies are also better at learning compared to fall flies collected from
364 the same location. Our finding of higher reproductive output in fall supports the hypothesis that
365 seasonal oscillations in traits and allele frequencies are caused by alternating selection for
366 robustness required for survival in the harsh winter conditions and for reproduction and

367 population growth during the summer expansion (Schmidt et al. 2005; Schmidt and Paaby 2008).

368 Our data demonstrate a pattern of cyclical seasonal selection with general robustness
369 favored in the winter and fecundity selected for during the summer. However, it remains unclear
370 which traits are directly being selected for, particularly given genetic correlations among traits
371 and complexity of genetic architecture. The negative correlation between learning and
372 reproduction is consistent with previous studies across a range of taxa (Galea et al. 1994; Mery
373 and Kawecki 2004; Snell-Rood et al. 2011). However, artificial selection in *D. melanogaster*
374 indicates independent trajectories of cognitive and reproductive aging, indicating some
375 differences in the genetic architecture of these traits with age (Zwoinska et al. 2017). It is
376 possible that rapid evolution of learning may be a product of the negative correlation between
377 learning and fecundity. The exponential population growth throughout the summer may select
378 for increased fecundity and therefore result in lower learning ability as a correlated response.

379

380

381 *Natural variants in cpo affect learning*

382 Selection on pleiotropic genes could result in the correlated trait changes in the
383 population across seasonal time. Variants of *cpo* show latitudinal clines and seasonal fluctuations
384 in frequency (Schmidt et al. 2008; Kolaczowski et al. 2011; Fabian et al. 2012; Cogni et al.
385 2013; Bergland et al. 2014). *cpo* is known to be involved in many traits including activity,
386 dormancy, fecundity and lifespan (Behrman et al. n.d.; Bellen et al. 1992b; Schmidt et al. 2008),
387 but learning is a new phenotype for this pleiotropic gene. Learning performance and fecundity of
388 the recombinant outbred populations homozygous for the *cpo* haplotype variants matched the
389 seasonal pattern of those traits in the wild. The haplotype with SNP variants that are more

390 frequent in the spring showed higher learning and lower reproduction than the fall haplotype.
391 The flies containing the spring *cpo* haplotype showed a weaker avoidance of odors when paired
392 with air during the control. Previous work has shown that a knockdown of *cpo* through p-element
393 insertion also reduced odor avoidance (Sambandan et al. 2006). This leaves the possibility that
394 flies containing the spring *cpo* haplotypes have a reduced perception of odors; however, this
395 should have led to impaired rather than improved learning. Thus, the difference in learning
396 between the haplotypes is thus unlikely to be due to differences in ability to perceive odor.
397 Rather, it appears that natural variation in *cpo* haplotypes, or genetic variants linked to them,
398 serve as an integrator for sensing and responding to environmental changes. Lower full-body *cpo*
399 expression in the spring ROPs (Behrman et al. n.d.) may contribute to the higher learning ability
400 in these flies. We tested if *cpo* regulation affects learning ability using GAL4-UAS and found the
401 opposite pattern: decreasing *cpo* expression in the sensory neurons and ventral nerve cord
402 decreased learning ability. Our results indicate that *cpo* expression in the peripheral nervous
403 system is involved in learning. The contrary pattern in the ROPs may due to masking by other
404 tissues (e.g, ovaries) that express high levels of *cpo*. Altogether, these results imply that the
405 seasonally fluctuating evolution of learning ability is at least in part mediated by polymorphisms
406 in *cpo*.

407

408 *Pleiotropy as a driving force in rapid evolution of learning*

409 The effect of *cpo* on numerous fitness traits beside learning is consistent with evidence
410 that many learning genes have broad pleiotropic effects (Dubnau et al. 2002; Butcher et al.
411 2006). For example, a natural polymorphism in *D. melanogaster foraging (for)* gene is highly
412 pleiotropic: in addition to learning (Kaun et al. 2007; Mery et al. 2007) it affects many traits,

413 including foraging behavior (Fitzpatrick and Sokolowski 2004) and aggregation (Wang and
414 Sokolowski 2017).

415 The overall force of selection acting on pleiotropic polymorphisms in natural populations
416 reflects their aggregate impact on survival and reproduction mediated by the diverse ecologically
417 relevant traits they influence. It is unclear which traits are directly under selection for their
418 fitness benefits and which evolve as a byproduct of natural selection on correlated traits. The
419 learning differences between *cpo* haplotypes are likely to generate fitness differences that
420 contribute to seasonal adaptation in natural populations, either through alternative coadapted
421 strategies or are a non-adaptive mechanistic consequence of gene action.

422

423

424 **Acknowledgements:**

425 This work was supported by NSF GRF DGE-0822 (ELB), the Rosemary Grant Award from the
426 Society for the Study of Evolution (ELB), the Teece Dissertation Research Fellowship from the
427 University of Pennsylvania (ELB), the Peachey Environmental Fund from the Department of
428 Biology, University of Pennsylvania (ELB), and NIH R01GM100366 (PS), as well as a research
429 funding from the University of Lausanne to TJK.

430 **Author contributions:**

431 ELB, TJK and PS designed the experiment. ELB performed the experiment and did the analyses.

432 ELB, TJK and PS wrote the manuscript.

433 **References:**

434 Bates, D., M. Mächler, B. Bolker, and S. Walker. 2014. Fitting Linear Mixed-Effects Models
435 using lme4. *J. Stat. Softw.* 67:1–48.

436 Battesti, M., C. Moreno, D. Joly, and F. Mery. 2012. Report Spread of Social Information and
437 Dynamics of Social Transmission within *Drosophila* Groups. *Curr. Biol.* 22:309–313.

438 Bednekoff, P. A., R. P. Balda, A. C. Kamil, and A. G. Hile. 1997. Long-term spatial memory in
439 four seed-caching corvid species. *Anim. Behav* 53:335–341.

440 Behrman, E. L., A. O. Bergland, D. A. Petrov, and P. S. Schmidt. n.d. Intragenic epistasis in
441 couch potato and its effect on climatic adaptation in natural populations in *Drosophila*
442 *melanogaster*.

443 Behrman, E. L., V. M. Howick, M. Kapun, F. Staubach, A. O. Bergland, D. A. Petrov, B. P.
444 Lazzaro, and P. S. Schmidt. 2018. Rapid seasonal evolution in innate immunity of wild
445 *Drosophila melanogaster*. *Proc. R. Soc. B* 285:20172599.

446 Behrman, E. L., S. S. Watson, K. R. O’Brien, M. S. Heschel, and P. S. Schmidt. 2015. Seasonal
447 variation in life history traits in two *Drosophila* species. *J. Evol. Biol.* 28:1691–1704.

448 Bellen, H. J., S. Kooyer, D. D’Evelyn, and J. Pearlman. 1992a. The *Drosophila* couch potato
449 protein is expressed in nuclei of peripheral neuronal precursors and shows homology to
450 RNA-binding proteins. *Genes Dev.* 6:2125–36.

451 Bellen, H. J., H. Vaessin, E. Bier, A. Kolodkin, D. D’Evelyn, S. Kooyer, and Yuh Nung Jan.
452 1992b. The *Drosophila* couch potato gene: An essential gene required for normal adult

- 453 behavior. *Genetics* 131:365–375.
- 454 Bergland, A. O., E. L. Behrman, K. R. O’Brien, P. S. Schmidt, D. A. Petrov, K. R.
455 O’Brien, P. S. Schmidt, and D. A. Petrov. 2014. Genomic Evidence of Rapid and Stable
456 Adaptive Oscillations over Seasonal Time Scales in *Drosophila*. *PLoS Genet.* 10:e1004775.
- 457 Brandes, C., B. Frish, and R. Menzel. 1988. Time course of memory formation differs in
458 honeybee lines selected for good and poor learning. *Anim. Behav.* 36:981–985.
- 459 Butcher, L. M., J. K. Kennedy, and R. Plomin. 2006. Generalist genes and cognitive
460 neuroscience. *Curr. Opin. Neurobiol.* 16:145–151.
- 461 Carroll, S. P., A. P. Hendry, D. N. Reznick, and C. W. Fox. 2007. Evolution on ecological time-
462 scales. *Funct. Ecol.* 21:387–393.
- 463 Celniker, S. E., L. A. L. Dillon, M. B. Gerstein, K. C. Gunsalus, S. Henikoff, G. H. Karpen, M.
464 Kellis, E. C. Lai, J. D. Lieb, D. M. MacAlpine, G. Micklem, F. Piano, M. Snyder, L. Stein,
465 K. P. White, and R. H. Waterston. 2009. Unlocking the secrets of the genome. *Nature*
466 459:927–930.
- 467 Cogni, R., C. Kuczynski, S. Koury, E. Lavington, E. L. Behrman, K. R. O’Brien, P. S.
468 Schmidt, W. F. Eanes, K. R. O’Brien, P. S. Schmidt, and W. F. Eanes. 2013. The Intensity
469 of Selection Acting on the Couch Potato Gene-Spatial-Temporal Variation in a Diapause
470 Cline. *Evolution (N. Y.)*. 68:538–548.
- 471 Crombach, A., and P. Hogeweg. 2008. Evolution of evolvability in gene regulatory networks.
472 *PLoS Comput. Biol.* 4:e1000112.

- 473 Croston, R., C. L. Branch, D. Y. Kozlovsky, T. C. Roth, L. D. LaDage, C. A. Freas, and V. V.
474 Pravosudov. 2015. Potential mechanisms driving population variation in spatial memory
475 and the hippocampus in food-caching chickadees. Pp. 354–371 *in* Integrative and
476 Comparative Biology.
- 477 Dubnau, J., A.-S. Chiang, and T. Tully. 2002. Neural substrates of memory: From synapse to
478 system. *J. Neurobiol.* 54:238–253.
- 479 Dukas, R. 2004. Evolutionary Biology of Animal Cognition. *Annu. Rev. Ecol. Evol. Syst.*
480 35:347–374.
- 481 Dukas, R. 2008. Evolutionary Biology of Insect Learning Learning: the acquisition of neuronal
482 representations of new information. *Annu. Rev. Entomol* 53:145–60.
- 483 Dukas, R. 1998. Evolutionary Ecology of Learning. Pp. 129–174 *in* Cognitive Ecology: The
484 Evolutionary Ecology of Information Processing and Decision Making.
- 485 Dukas, R. 2005. Learning affects mate choice in female fruit flies. *Behav. Ecol.* 16:800–804.
- 486 Fabian, D. K., M. Kapun, V. Nolte, R. Kofler, P. S. Schmidt, C. Schlötterer, and T. Flatt. 2012.
487 Genome-wide patterns of latitudinal differentiation among populations of *Drosophila*
488 *melanogaster* from North America. *Mol. Ecol.* 21:4748–4769.
- 489 Fitzpatrick, M. J., and M. B. Sokolowski. 2004. In Search of Food: Exploring the Evolutionary
490 Link Between cGMP-Dependent Protein Kinase (PKG) and Behaviour. *Integr. Comp. Biol.*
491 44:28–36.
- 492 Fox, J., and S. Weisberg. 2011. An {R} Companion to Applied Regression.

- 493 Froissart, L., M. Giurfa, S. Sauzet, and E. Desouhant. 2017. Cognitive adaptation in asexual and
494 sexual wasps living in contrasted environments. *PLoS One* 12.
- 495 Galea, L. A. M., M. Kavaliers, K. P. Ossenkopp, D. Innes, and E. L. Hargreaves. 1994. Sexually
496 dimorphic spatial learning varies seasonally in two populations of deer mice. *Brain Res.*
497 635:18–26.
- 498 Grant, P. R., and B. R. Grant. 2002. Unpredictable evolution in a 30-year study of Darwin’s
499 finches. *Science* (80-.). 296:707–711.
- 500 Healy, S. D., and C. M. Jones. 2002. Animal learning and memory: an integration of cognition
501 and ecology. *Zoology* 105:321–327.
- 502 Hoedjes, K. M., and H. M. Smid. 2014. Natural variation in long-term memory formation among
503 *Nasonia* parasitic wasp species. *Behav. Processes* 105:40–45.
- 504 Hollis, B., and T. J. Kawecki. 2014. Male cognitive performance declines in the absence of
505 sexual selection. *Proc. R. Soc. B Biol. Sci.* 281:20132873.
- 506 Johnston, T. D. 1982. Selective costs and benefits in the evolution of learning. *Adv. Study*
507 *Behav.* 12:65–106.
- 508 Kaun, K. R., T. Hendel, B. Gerber, and M. B. Sokolowski. 2007. Natural variation in *Drosophila*
509 larval reward learning and memory due to a cGMP-dependent protein kinase. *Learn. Mem.*
510 14:342–349.
- 511 Kingsolver, J. G., H. E. Hoekstra, J. M. Hoekstra, D. Berrigan, S. N. Vignieri, C. E. Hill, A.
512 Hoang, P. Gibert, and P. Beerli. 2001. The strength of phenotypic selection in natural

- 513 populations. *Am. Nat.* 157:245–61.
- 514 Kolaczowski, B., A. D. Kern, A. K. Holloway, and D. J. Begun. 2011. Genomic Differentiation
515 Between Temperate and Tropical Australian Populations of *Drosophila melanogaster*.
516 *Genetics* 187:245–260.
- 517 Lenth, R. V. 2016. Least-Squares Means: The R Package lsmeans. *J. Stat. Softw.* 69.
- 518 MacKay, T. F. C., S. Richards, E. A. Stone, A. Barbadilla, J. F. Ayroles, D. Zhu, S. Casillas, Y.
519 Han, M. M. Magwire, J. M. Cridland, M. F. Richardson, R. R. H. Anholt, M. Barrón, C.
520 Bess, K. P. Blankenburg, M. A. Carbone, D. Castellano, L. Chaboub, L. Duncan, Z. Harris,
521 M. Javaid, J. C. Jayaseelan, S. N. Jhangiani, K. W. Jordan, F. Lara, F. Lawrence, S. L. Lee,
522 P. Librado, R. S. Linheiro, R. F. Lyman, A. J. MacKey, M. Munidasa, D. M. Muzny, L.
523 Nazareth, I. Newsham, L. Perales, L. L. Pu, C. Qu, M. Ràmia, J. G. Reid, S. M. Rollmann,
524 J. Rozas, N. Saada, L. Turlapati, K. C. Worley, Y. Q. Wu, A. Yamamoto, Y. Zhu, C. M.
525 Bergman, K. R. Thornton, D. Mittelman, and R. A. Gibbs. 2012. The *Drosophila*
526 *melanogaster* Genetic Reference Panel. *Nature* 482:173–178.
- 527 Maille, A., and C. Schradin. 2016. Ecophysiology of cognition: How do environmentally
528 induced changes in physiology affect cognitive performance? *Biol. Rev.* 92:1101–1112.
- 529 Mansourian, S., J. Corcoran, A. Enjin, C. Löfstedt, M. Dacke, and M. C. Stensmyr. 2016. Fecal-
530 Derived Phenol Induces Egg-Laying Aversion in *Drosophila*. *Curr. Biol.* 26:2762–2769.
- 531 Mayr, E. 1974. Behavior Programs and Evolutionary Strategies: Natural selection sometimes
532 favors a genetically “closed” behavior program, sometimes an “open” one. *Am. Sci.*
533 62:650–659.

Rapid learning evolution

- 534 Mcguire, T. R., and J. Hirschth. 1977. Behavior-genetic analysis of *Phormia regina*:
535 Conditioning, reliable individual differences, and selection. 74:5193–5197.
- 536 Mery, F., A. T. Belay, A. K-C So, M. B. Sokolowski, T. J. Kawecki, and G. E. Robinson. 2007.
537 Natural polymorphism affecting learning and memory in *Drosophila*. *Proc. Natl. Acad. Sci.*
538 104:13051–13055.
- 539 Mery, F., and T. J. Kawecki. 2004. An operating cost of learning in *Drosophila melanogaster*.
540 *Anim. Behav.* 68:589–598.
- 541 Mery, F., and T. J. Kawecki. 2005. Cost of Long-Term Memory in *Drosophila*. *Science* (80-.).
542 308:1148.
- 543 Mery, F., and T. J. Kawecki. 2002. Experimental evolution of learning ability in fruit flies. *Pnas*
544 99:14274–14279.
- 545 Mery, F., and T. J. Kawecki. 2004b. The effect of learning on experimental evolution of
546 resource preference in *Drosophila melanogaster*. *Evolution* (N. Y). 58:757–767.
- 547 Odling-Smee, L. C., J. W. Boughman, and V. A. Braithwaite. 2008. Sympatric species of
548 threespine stickleback differ in their performance in a spatial learning task. *Behav. Ecol.*
549 *Sociobiol.* 62:1935–1945.
- 550 Olson, D. J., A. C. Kamil, R. P. Balda, and P. J. Nims. 2004. Performance of Four Seed-Caching
551 Corvid Species in Operant Tests of Nonspatial and Spatial Memory. *J. Comp. Psychol.*
552 109:1–9.
- 553 Paaby, A. B., A. O. Bergland, E. L. Behrman, and P. S. Schmidt. 2014. A highly pleiotropic

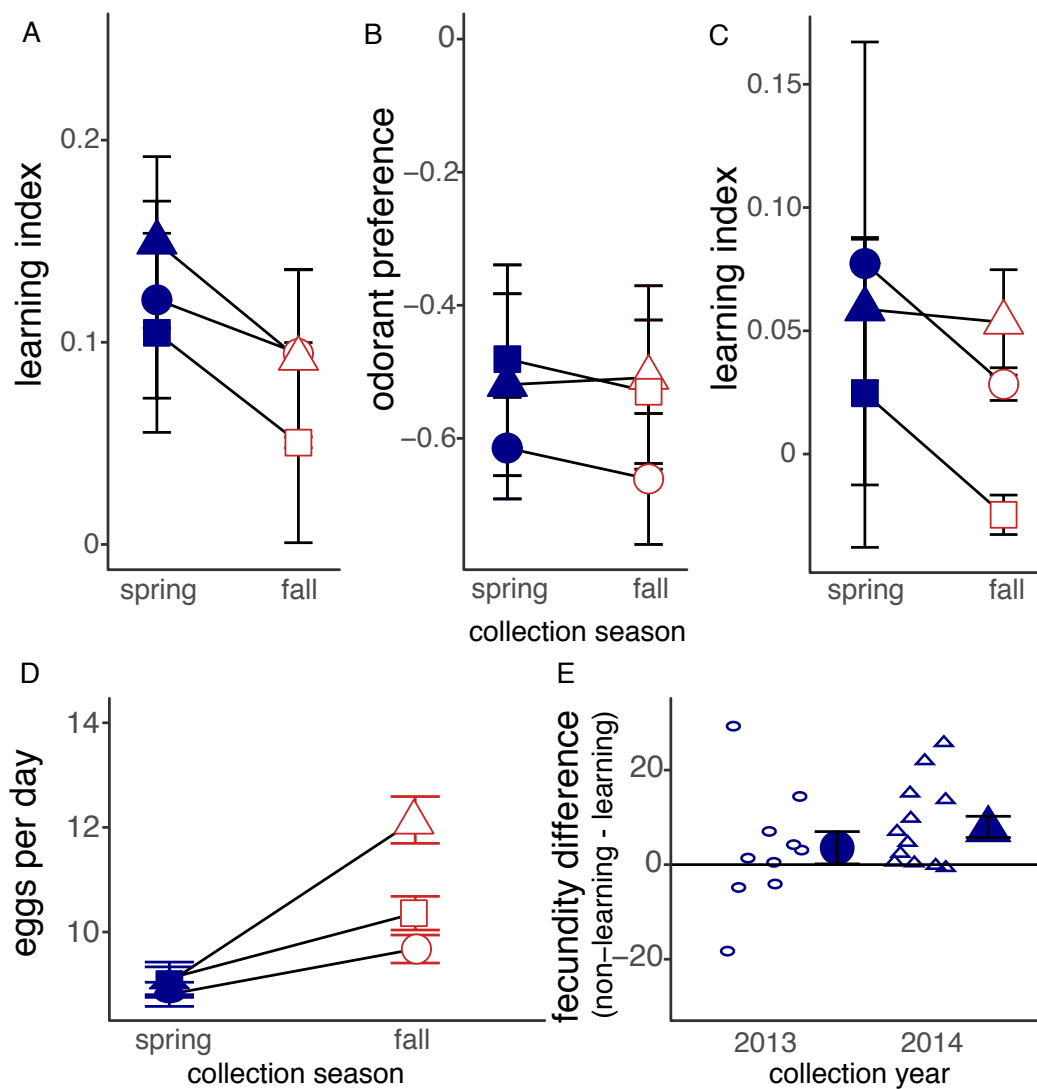
- 554 amino acid polymorphism in the *Drosophila* insulin receptor contributes to life-history
555 adaptation. *Evolution* (N. Y). 68:3395–3409.
- 556 Paenke, I., B. Sendhoff, and T. J. Kawecki. 2007. Influence of Plasticity and Learning on
557 Evolution under Directional Selection. *Am. Nat.* 170:E47–E58.
- 558 Pravosudov, V. V., and N. S. Clayton. 2002. A test of the adaptive specialization hypothesis:
559 Population differences in caching, memory, and the hippocampus in black-capped
560 chickadees (*Poecile atricapilla*). *Behav. Neurosci.* 116:515–522.
- 561 Sambandan, D., A. Yamamoto, J.-J. Fanara, T. F. C. Mackay, and R. R. H. Anholt. 2006.
562 Dynamic genetic interactions determine odor-guided behavior in *Drosophila melanogaster*.
563 *Genetics* 174:1349–63.
- 564 Sarin, S., and R. Dukas. 2009. Social learning about egg-laying substrates in fruitflies. *Proc. R.*
565 *Soc. B Biol. Sci.* 276:4323–4328.
- 566 Schmidt, P. S., and D. R. Conde. 2006. Environmental heterogeneity and the maintenance of
567 genetic variation for reproductive diapause in *Drosophila melanogaster*. *Evolution* 60:1602–
568 11.
- 569 Schmidt, P. S., and A. B. Paaby. 2008. Reproductive diapause and life-history clines in North
570 American populations of *Drosophila melanogaster*. *Evolution* (N. Y). 62:1204–1215.
- 571 Schmidt, P. S., A. B. Paaby, and M. S. Heschel. 2005. Genetic variance for diapause expression
572 and associated life histories in *Drosophila melanogaster*. *Evolution* (N. Y). 59:2616–2625.
- 573 Schmidt, P. S., C.-T. C.-T. Zhu, J. Das, M. Batavia, L. Yang, and W. F. Eanes. 2008. An amino

- 574 acid polymorphism in the couch potato gene forms the basis for climatic adaptation in
575 *Drosophila melanogaster*. Proc. Natl. Acad. Sci. U. S. A. 105:16207–16211.
- 576 Smid, H. M., and L. E. Vet. 2016. The complexity of learning, memory and neural processes in
577 an evolutionary ecological context. Curr. Opin. Insect Sci. 15:61–69.
- 578 Snell-Rood, E. C., G. Davidowitz, and D. R. Papaj. 2011. Reproductive tradeoffs of learning in a
579 butterfly. Behav. Ecol. 22:291–302.
- 580 Stensmyr, M. C., H. K. M. Dweck, A. Farhan, I. Ibba, A. Strutz, L. Mukunda, J. Linz, V. Grabe,
581 K. Steck, S. Lavista-Llanos, D. Wicher, S. Sachse, M. Knaden, P. G. Becher, Y. Seki, and
582 B. S. Hansson. 2012. A Conserved Dedicated Olfactory Circuit for Detecting Harmful
583 Microbes in *Drosophila*. Cell 151:1345–1357.
- 584 Thompson, J. N. 2013. Relentless evolution. University of Chicago.
- 585 Tryon, R. C. 1940. Studies in individual differences in maze ability. VII. The specific
586 components of maze ability, and a general theory of psychological components. J. Comp.
587 Psychol. 30:283–335.
- 588 Wang, S., and M. B. Sokolowski. 2017. Aggressive behaviours, food deprivation and the
589 foraging gene. R. Soc. Open Sci. 4:170042.
- 590 Yurkovic, A., O. Wang, A. C. Basu, and E. A. Kravitz. 2006. Learning and memory associated
591 with aggression in *Drosophila melanogaster*. Proc. Natl. Acad. Sci. U. S. A. 103:17519–
592 17524.
- 593 Zrelec, V., M. Zini, S. Guarino, J. Mermoud, J. Oppliger, A. Valtat, V. Zeender, and T. J.

Rapid learning evolution

- 594 Kawecki. 2013. *Drosophila* rely on learning while foraging under semi-natural conditions.
595 Ecol. Evol. 3:4139–4148.
- 596 Zwoinska, M. K., A. A. Maklakov, T. J. Kawecki, and B. Hollis. 2017. Experimental evolution
597 of slowed cognitive aging in *Drosophila melanogaster*. *Evolution* (N. Y). 71:662–670.
- 598

599 Figures:



600

601 Figure 1. Rapid evolution of learning and reproduction in rapid evolution in natural populations

602 of *Drosophila melanogaster*. (a) Higher learning (mean +/-SE) in the spring populations

603 compared to the fall replicated across three years: 2012 (square), 2013 (circle), 2014 (triangle).

604 (b) No difference absolute preference in either population when unconditioned flies are given the

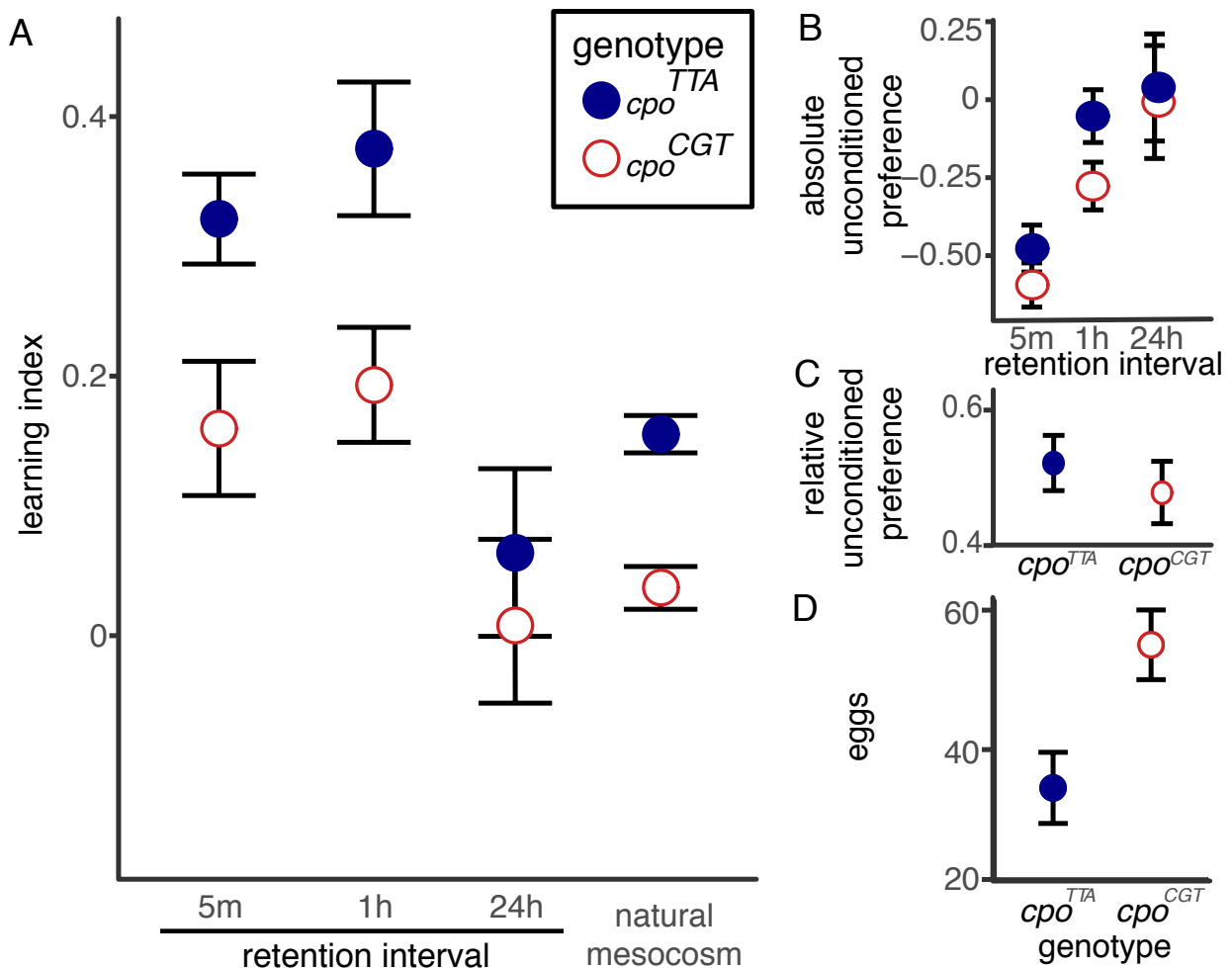
605 choice between one odor and solvent. (c) Spring populations return to positive conditioning food

606 at a higher rate than fall populations. (d) Lower reproduction in the spring populations compared

Rapid learning evolution

607 to the fall over ten days. (e) Individuals that do not learn in the aversive conditioning assay have
608 higher daily reproductive output than those that learn over three days. Difference between
609 individuals is shown in small outlined shapes and the mean difference +/- SE in the large, filled
610 shapes.

611



612

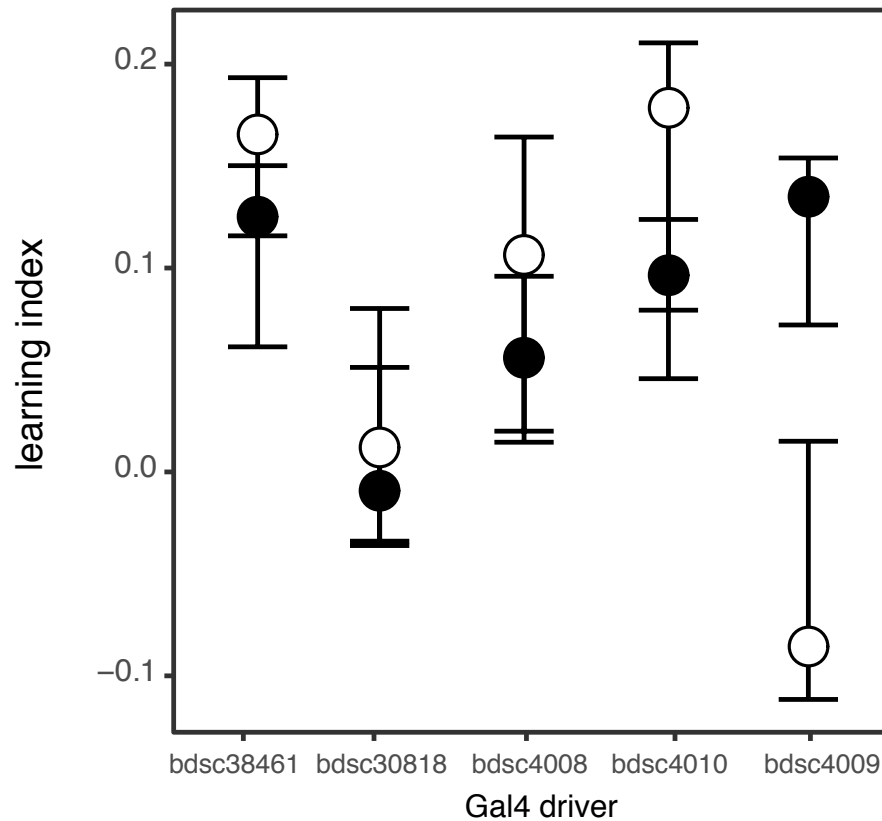
613 Figure 2. Natural variants in *couch potato* (*cpo*) involved in trade-off between learning and

614 reproduction. (a) Flies containing the spring (*cpo*^{TTA}) haplotype have higher learning (mean +/-

Rapid learning evolution

615 SE) than flies that contain the fall (*cpo*^{CGT}) haplotype across all retention intervals between
616 conditioning and testing using aversive shock, as well as in the appetitive conditioning of the
617 natural mesocosm. (b) Unconditioned flies containing the spring haplotype have a slightly higher
618 absolute preference for odor instead of solvent compared to the flies containing the fall
619 haplotype. (c) However, there is no relative preference for either of the experimental odors in
620 unconditioned flies. (d) Flies containing the fall haplotype had higher daily reproductive output
621 than flies containing the spring haplotype.

622



623

624 Figure 3. Effect of *couch potato* (*cpo*) expression on aversive shock learning. Learning scores
625 (mean +/- SE) for flies with normal *cpo* expression (filled) and *cpo* knockdown (outline). No
626 difference in learning when *cpo* was knocked down in the Actin 88F promotor control
627 (BSC38461), in the broad brain knockout (BDSC30818), in a subset of ventral nerve cord and
628 sensory neurons (BDSC-40315) or in the sense organ support cells and subsets of ventral nerve
629 cord, brain and sensory neurons (BDSC-40334). Knocking down *cpo* expression in a subset of
630 the sensory neurons (BDSC-40319) decreased learning.

631