Lipoprotein Signatures of Cholesteryl Ester Transfer Protein and HMG-CoA Reductase Inhibition

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Disclosures

None

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Abstract

Background: CETP inhibition reduces vascular event rates but confusion surrounds its low-density lipoprotein (LDL)-cholesterol effects. We sought to clarify associations of genetic inhibition of CETP on detailed lipoproteins.

Methods and Results: We used variants associated with *CETP* (rs247617) and *HMGCR* (rs12916) expression in 62,400 Europeans with detailed lipoprotein profiling from nuclear magnetic resonance spectroscopy. Genetic associations were scaled to 10% lower risk of coronary heart disease (CHD). Associations of lipoprotein measures with risk of incident CHD in three population-based cohorts (770 cases) were examined.

CETP and HMGCR had near-identical associations with LDL-cholesterol concentration estimated by Friedewald-equation. HMGCR had a relatively consistent effect on cholesterol concentrations across all apolipoprotein B-containing lipoproteins. CETP had stronger effects on remnant and very-low-density lipoprotein cholesterol but no effect on cholesterol concentrations in LDL defined by particle size (diameter 18–26 nm) (-0.02SD 95%CI: -0.10, 0.05 for CETP versus -0.24SD, 95%CI -0.30, -0.18 for HMGCR). CETP had profound effects on lipid compositions of lipoproteins, with strong reductions in the triglyceride content of all high-density lipoprotein (HDL) particles. These alterations in triglyceride composition within HDL subclasses were observationally associated with risk of CHD, independently of total cholesterol and triglycerides (strongest HR per 1-SD higher triglyceride composition in very-large HDL 1.35; 95%CI: 1.18, 1.54).

Conclusion: CETP inhibition does not affect size-specific LDL cholesterol but may lower CHD risk by lowering cholesterol in other apolipoprotein-B containing lipoproteins and lowering triglyceride content of HDL particles. Conventional composite lipid assays may mask heterogeneous effects of lipid-altering therapies.

Keywords: CETP, HMGCR, lipoproteins, drug target Mendelian randomization

Introduction

Definitive evidence on the causal role of low-density lipoproteins (LDL) in cardiovascular disease comes from trials of LDL cholesterol lowering compounds, which have shown beneficial effects on risk of coronary heart disease (CHD) and stroke. Consistent effects have been seen for drugs acting on related pathways, such as 3-hydroxy-3-methylglutaryl-coenzyme A reductase (HMGCR) inhibitors, i.e., statins, and proprotein convertase subtilisin-kexin type 9 (PCSK9) inhibitors, both of which upregulate hepatic LDL receptor expression, and for drugs acting on other pathways, such as ezetimibe, which inhibits intestinal absorption of cholesterol. 3

However, trials of drugs primarily designed to alter concentrations of lipids other than LDL cholesterol have had mixed results. One such example is the class of drugs designed to inhibit cholesteryl ester transfer protein (CETP), a lipid transport protein responsible for the exchange of triglycerides and cholesteryl esters between apolipoprotein B-containing particles and high-density lipoprotein (HDL) particles. CETP inhibitors were developed initially on the basis of their HDL cholesterol raising effects. While accumulating genetic evidence suggests that HDL cholesterol concentration is unlikely to be causally related to CHD, there were two strong reasons to believe that CETP inhibition may still reduce vascular risk: (i) genetic studies of *CETP* variants have shown associations with CHD and (ii) some CETP inhibitors not only increase HDL cholesterol but also appear to lower LDL cholesterol as measured by conventional assays.

The recent findings from the phase III REVEAL trial showed that treatment with the CETP inhibitor anacetrapib led to a reduction in risk of coronary events that was proportional to the reduction in non-HDL cholesterol. Interestingly, anacetrapib appeared to have discrepant effects based on the assay used to quantify LDL cholesterol (using beta-quant, direct or Friedewald estimation). This discrepant effect was also identified in a genetic study that approximated a factorial clinical trial of CETP inhibition and statin therapy. Thus, while both CETP inhibitors and statins lower Friedewald-estimated LDL cholesterol, which also includes cholesterol carried by other lipoprotein particles, it is possible that the drugs have differential effects on the concentration and content of lipids in different apolipoprotein-B containing lipoproteins.

In this study, we used the established approach of exploiting genetic variants near the protein-coding genes of drug targets to investigate detailed lipid and lipoprotein subclass signatures of CETP inhibition. We compared the association of variants in *CETP* with *HMGCR*¹⁰ (to proxy statin treatment) to gauge insight into how these two therapies alter the lipoprotein milieu. We also present findings that the triglyceride composition, in contrast to circulating concentrations, of HDL particles is associated with CHD and may relate to a new mechanism by which CETP inhibition reduces risk of CHD.

Methods

Prospective and cross-sectional studies and lipoprotein quantification

We used genetic and lipoprotein data from four population-based Finnish cohorts and one cross-sectional study in the UK (cohort characteristics are presented in Online Table 1 and study descriptions are given in the Supplementary Note in the Supplementary Appendix). For prospective analyses we used two of the abovementioned Finnish cohorts and additionally a UK-based multiethnic SABRE (Southall And Brent REvisited) cohort. Briefly, the cohorts used were the Northern Finland Birth Cohort 1966 (NFBC66) (n = 4,702 individuals aged 31 y at blood draw), ¹¹ the Cardiovascular Risk in Young Finns Study (YFS, n = 1,948 individuals aged 24–39 y in 2007), ¹² two population based Finnish cohorts FINRISK 1997 (n = 6,942 individuals aged 24–74 y) and DILGOM subsample of FINRISK 2007 (n = 4,124 individuals aged 24–74 y), ¹³ a study of healthy blood donors from the UK (INTERVAL: n = 40,958 individuals aged 18–80 y). ¹⁴ and a tri-ethnic UK community-based cohort SABRE (n = 4,857 individuals aged 40–69 y). ¹⁵ A nuclear magnetic resonance (NMR)-based methodology was used to quantify lipoprotein lipids and subclasses. Details of this platform have been published previously, ^{16,17} and it has been widely applied in genetic and epidemiological studies. ^{10,18}

Where possible, we excluded individuals receiving lipid lowering medication, pregnant women and those who had a high proportion (>30%) of values missing across the lipid traits. All measures were first adjusted for sex, age (if applicable), genotyping batch (if applicable) and ten first principal components from genomic data and the resulting residuals were transformed to

normal distribution by inverse rank-based normal transformation. Details of study-specific genotyping are provided in Online Table 2 in the Supplementary Appendix.

SNP analysis

We selected variants as genetic proxies of CETP and HMGCR inhibition on the basis of robust associations with circulating lipids in GWAS consortia^{19, 20} and target gene expression. The HMGCR variant (rs12916) LDL cholesterol lowering T allele (-0.24 SD LDL cholesterol per T allele; P=1.3x10⁻¹⁴) has been shown to lower *HMGCR* expression²¹ and the *CETP* variant (rs247617) HDL cholesterol increasing A allele (0.84 SD HDL cholesterol per A allele: P=5.4x10⁻⁹⁴) associates with lower gene expression across several tissues, verified through Genotype To Expression (https://gtexportal.org) project data.²² We used an additive model for each cohort separately (see Online Table 1 for details of analysis software). In order to make the lipoprotein and lipid estimates comparable, the estimates for CETP rs247617 and HMGCR rs12916 were scaled to the same CHD association as reported by the CARDIoGRAMplusC4D GWAS Consortium.²³ The per-allele log odds (logOR) for CHD was 0.0358 for *HMGCR* rs12916 and 0.0309 for CETP rs247617; subsequently the summary statistics of each individual cohort and each metabolite were scaled to -0.105 logOR of CHD (equivalent to an odds ratio [OR] of CHD of 0.90) to align the estimates to a 10% lower risk of CHD. The cohort specific association results of lipoprotein and lipid measures with both variants were then combined using an inverse variance weighted fixed effect meta-analysis.

Our focus for this study was to evaluate the impact of variants in *CETP* and *HMGCR* on the entire cascade of apolipoprotein B-containing lipoproteins and HDL subclasses. Therefore, we decided *a priori* to examine 191 lipoprotein and lipid traits available from the NMR platform.^{18, 24} Focusing on these 191 traits, we estimated that 28 principal components explain 99% of their variation in the Finnish cohorts and therefore we used a P-value threshold of 0.05/28=0.002 to denote evidence in favor of an association. Abbreviations and full descriptions of the lipoprotein measures studied are listed in Online Table 3.

Association of lipoprotein measures with risk of incident CHD

Cohorts contributing to the associations of lipoprotein lipid concentration and composition measures and the hazard of incident CHD were FINRISK 1997, DILGOM and SABRE. Participants with prevalent CHD were excluded from the analysis. Following exclusion, data were available from FINRISK 1997 for 7,076 individuals (291 cases / 6,785 controls) and 4,736 individuals from DILGOM (192 cases / 4,544 controls) and for SABRE 4,689 individuals with non-missing data (287 cases / 4,402 controls). The follow up time of FINRISK 1997 and SABRE were censored to 8 years to match the follow up time in DILGOM.

Prior to statistical analyses, metabolic measures were log-transformed and scaled to standard deviations (SD) in each cohort. The relationships of lipid measures with the risk of CHD were analysed using Cox proportional hazards regression models with age, sex, mean arterial pressure, smoking, diabetes mellitus, lipid medication and geographical region (Finnish cohorts), ethnicity (SABRE), total cholesterol and total triglyceride concentrations as covariates. The cohort-specific association results of 191 lipid measures were then combined using an inverse variance weighted fixed effect meta-analyses. Analyses were conducted in R studio (version 1.0.153, R version 3.3.3). As above, we used a P-value threshold of ≤0.002 to denote evidence in favor of an association.

Results

Data from 62,400 individuals with extensive lipoprotein subclass profiling and genotypes were available. We combined data from five adult cohorts (mean age range from 31 to 52 years) and one cohort of adolescents (mean age 16 years) for the genetic analyses where 51% of participants of all six studies were female. Study specific and pooled estimates from meta-analyses of genetic and observational analyses for all 191 traits are presented in Online Figures 1-15.

Scaled to a 10% lower risk of CHD, *CETP* rs247617 and *HMGCR* rs12916 had near-identical associations with Friedewald estimated LDL cholesterol (Fig. 1) and similar associations for apolipoprotein B. In contrast, when LDL cholesterol was defined on the basis of cholesterol transported in LDL based on particle size (diameter 18–26 nm), and measured via NMR spectroscopy, *CETP* had no association with this size-specific LDL cholesterol (0.02 SDs; 95%CI: -0.10, 0.05). While *HMGCR* had a relatively consistent association with individual apolipoprotein B-containing lipoproteins (effect estimates ranging from -0.25 for IDL cholesterol to -0.18 for VLDL cholesterol), *CETP* had the most pronounced associations with VLDL cholesterol, a weaker association with IDL cholesterol but no association with LDL cholesterol defined by particle size or cholesterol transported by any of the large, medium or small LDL subclasses (Fig. 1).

When examining triglycerides in apolipoprotein B-containing particles, *CETP* associated with lower circulating triglyceride concentrations in VLDL and IDL subclasses, while *HMGCR* had weaker effects on these measures, except in LDL subclasses (Fig. 2). *CETP* had a very strong association with higher HDL cholesterol (0.84; 95%CI: 0.76, 0.92) but *HMGCR* did not (0.04; 95%CI: -0.02, 0.10) (Fig. 3). Similarly, *CETP* was inversely associated with the total quantity of triglycerides in HDL particles (-0.23; 95%CI: -0.31,-0.15) but *HMGCR* was not (-0.03; 95%CI: -0.09, 0.02).

The lipoprotein particle structure is biophysically constrained, generating strong correlations between lipid measures within individual lipoprotein subclasses.²⁵⁻²⁸ Notable differences in lipid concentrations in subclass particles would therefore suggest changes in the compositional proportions of these lipids. For genetic inhibition of CETP, the effects on circulating triglyceride

concentrations in all HDL subclasses were weaker (XL-HDL and L-HDL) or even in the opposite direction (M-HDL and S-HDL) than the effects on cholesterol concentration in these subclasses (Fig. 3). Examining the genetic associations with the particle lipid compositions, the relative amount of triglycerides (in relation to all lipid molecules in the particles) was remarkably diminished in all HDL subclass particles by genetic inhibition of CETP (Fig. 4). Genetic inhibition of HMGCR did not associate with the triglyceride concentration or composition of any HDL subclass. These associations are in line with the known physiological roles of CETP and HMGCR and their inhibition.^{29, 30} In addition, as expected, *CETP* associated with higher compositions of triglycerides in most VLDL subclass particles and *HMGCR* showed directionally similar, albeit weaker associations.

To understand the clinical relevance of these HDL-related compositional changes arising from CETP inhibition, beyond reductions in cholesterol concentrations of apolipoprotein B-containing lipoprotein particles, we studied the observational associations of lipoprotein subclass lipid concentrations and compositions with CHD in three prospective population cohorts. The triglyceride concentration of HDL was associated with incident CHD when adjusted for non-lipid cardiovascular risk factors (Fig. 5)., However, when serum cholesterol and serum triglycerides were added to the model, as expected, the associations attenuated. In contrast, the triglyceride compositions of all the HDL subclass particles were positively associated with CHD, independent of circulating concentrations of cholesterol and triglycerides, with hazard ratios around 1.3 for all HDL subclasses (Fig. 5). In addition, compositional enrichment of cholesteryl esters in the largest VLDL particles (XXL-VLDL and XL-VLDL) was associated with risk of CHD (Online Fig. 11); genetic inhibition of CETP also impacted on these traits (Online Fig. 2).

Discussion

We used genetic variants in *CETP* and *HMGCR* to gain insight into the expected effects of therapeutic inhibition of CETP and HMG-coA reductase on circulating lipoproteins and lipids. Our data show that while *CETP* and *HMGCR* have near identical effects on Friedewald-estimated LDL cholesterol, this result masks a very different association of *CETP* and *HMGCR* with size-specific LDL cholesterol. Genetic inhibition of HMGCR showed similar effects with cholesterol across the apolipoprotein B-containing lipoproteins but genetic inhibition of CETP showed stronger associations with larger apolipoprotein B particles, namely VLDL and remnant cholesterol,³¹ but no association with cholesterol carried specifically in LDL particles defined by size.

Friedewald-estimated LDL cholesterol (and other assays such as 'direct' and betaquant) are nonspecific measures of cholesterol. 32-34 For example, in addition to the cholesterol in size-specific LDL particles, Friedewald LDL cholesterol also includes, to varying degrees, cholesterol in IDL, VLDL and lipoprotein(a).³⁵ This non-specificity of commonly-used "LDL" cholesterol assays is under-recognized and underlies the prevailing opinion that inhibitors of HMGCR and CETP both alter LDL cholesterol. However, our data show this not to be the case: using NMR spectroscopybased lipoprotein particle quantification, which defines individual lipoprotein subclasses based on particle size, ^{18, 25, 27} our findings demonstrate that CETP has negligible effect on cholesterol in size-specific LDL particles. In this way, the use of a composite lipid measure can obscure differential associations of a therapy or gene²⁶ with individual constituents of the composite, and can have clinical ramifications. For example, if a trial is powered to a given reduction in Friedewald LDL cholesterol, under the naïve assumption that the drug uniformly alters all the subcomponents, then the trial may not have the expected result if the drug has differential effects on these subcomponents. This is exemplified in the recent phase III ACCELERATE trial of evacetrapib, which was terminated for futility, and was powered to a difference in LDL cholesterol based on a composite assay. 36 The differential effects of CETP inhibition on composite markers such as Friedewald and directly-quantified LDL cholesterol compared to apolipoprotein B concentrations identified in the subsequent phase III REVEAL trial of anacetrapib⁷ suggest that had ACCELERATE used an alternative measure of pro-atherogenic

lipoproteins (e.g. apolipoprotein B or non-HDL-C⁸) to gauge the expected vascular effect, the trial may have been more appropriately powered.

This highlights the need to understand, in detail, the consequences of lipid modifying therapies on lipoproteins and lipids in order to be able to gauge whether a composite measure (such as Friedewald LDL cholesterol) can be reliably used as an indicator of the likely beneficial effect of a therapy. This is unlikely to be limited to assays for LDL cholesterol. For example, assays that quantify triglycerides, measure the summation of triglycerides across multiple lipoprotein particle categories. Drugs currently under development that target triglycerides (such as apolipoprotein C-III inhibitors³⁷) have differential effects on triglycerides in lipoprotein subclass particles as demonstrated in a recent genetic study. ³⁸ If triglycerides within different lipoprotein subclasses have heterogeneous effects on vascular disease, a clinical trial powered to the overall concentration of circulating triglycerides may give an inaccurate portrayal of the cardiovascular consequences arising from apolipoprotein C-III inhibition.

Another key finding is that the lipid compositions of lipoprotein particles can associate with disease risk independently of total lipid concentrations. While genetic inhibition of CETP increased circulating concentrations of cholesterol in all HDL subclasses, the triglyceride composition, i.e. the percentage of triglyceride molecules of all the lipid molecules in the particle, was markedly lower in all HDL particles. Intriguingly, our observational analyses, the first to explore lipoprotein particle lipid composition with CHD outcomes, revealed that triglyceride enrichment of HDL particles associates with higher risk for future CHD, independently of total circulating cholesterol and triglycerides. The largest hazard ratio for the triglyceride enrichment in medium HDL subclass particles was of a similar magnitude (~1.3) as that for LDL cholesterol and apolipoprotein B. These findings suggest that lipoprotein particle compositions, independent of circulating lipid concentrations, could have a role in the development of CHD. While the causal role of these lipoprotein compositions remains unclear, these findings advocate the importance of moving from simple composite lipid measures towards more detailed molecular phenotyping of lipoprotein metabolism.

Key strengths of our analyses include the availability of detailed measurements of blood lipoprotein subclass concentrations and compositions from general population studies with incident CHD events, together with the availability of genome-wide genotyping. While we used *CETP* and *HMGCR* variants as genetic proxies for therapeutic inhibition, we note that the *CETP* genetic variant recapitulated the effects of CETP enzyme activity in relation to the role the enzyme has in shuttling esterified cholesterol from HDL to apolipoprotein B-containing particles in exchange for triglycerides.²⁹ Furthermore, prospective population-based data of patients taking statins with blood sampling before and after the commencement of therapy showed that genetic variants in *HMGCR* robustly recapitulated the effects of statin therapy on lipoprotein subclasses and lipids.¹⁰

In conclusion, we have shown that, in contrast to genetic inhibition of HMG-CoA (proxying statin therapy), genetic inhibition of CETP does not alter circulating size-specific LDL cholesterol concentrations. This is masked by using conventional, non-specific assays for LDL cholesterol and may be problematic for ongoing and future clinical trials of lipid lowering therapies, especially when a non-specific marker of lipids is used to derive an expected effect of a drug with risk of disease. Our findings suggest potential additional mechanisms by which CETP inhibition could prevent CHD through reductions in the triglyceride composition of HDL particles. Our findings also call attention to the need for metabolic precision in measurements of lipoprotein lipids and in assessing the role of lipoprotein metabolism in cardiovascular disease in relation to ongoing treatment trials of novel lipid-altering therapies.

References

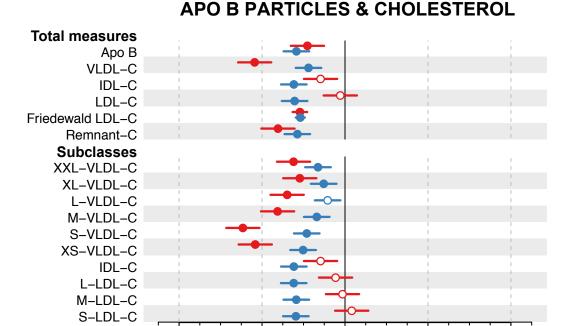
- 1. Collins R, Reith C, Emberson J, Armitage J, Baigent C, Blackwell L, Blumenthal R, Danesh J, Smith GD, DeMets D, Evans S, Law M, MacMahon S, Martin S, Neal B, Poulter N, Preiss D, Ridker P, Roberts I, Rodgers A, Sandercock P, Schulz K, Sever P, Simes J, Smeeth L, Wald N, Yusuf S, Peto R. Interpretation of the evidence for the efficacy and safety of statin therapy. Lancet 2016;**388**(10059):2532-2561.
- 2. Sabatine MS, Giugliano RP, Keech AC, Honarpour N, Wiviott SD, Murphy SA, Kuder JF, Wang H, Liu T, Wasserman SM, Sever PS, Pedersen TR, Committee FS, Investigators. Evolocumab and Clinical Outcomes in Patients with Cardiovascular Disease. N Engl J Med 2017.
- 3. Silverman MG, Ference BA, Im K, Wiviott SD, Giugliano RP, Grundy SM, Braunwald E, Sabatine MS. Association Between Lowering LDL-C and Cardiovascular Risk Reduction Among Different Therapeutic Interventions: A Systematic Review and Meta-analysis. JAMA 2016;316(12):1289-97.
- 4. Group HTC, Landray MJ, Haynes R, Hopewell JC, Parish S, Aung T, Tomson J, Wallendszus K, Craig M, Jiang L, Collins R, Armitage J. Effects of extended-release niacin with laropiprant in high-risk patients. N Engl J Med 2014;**371**(3):203-12.
- 5. White J, Swerdlow DI, Preiss D, Fairhurst-Hunter Z, Keating BJ, Asselbergs FW, Sattar N, Humphries SE, Hingorani AD, Holmes MV. Association of Lipid Fractions With Risks for Coronary Artery Disease and Diabetes. JAMA Cardiol 2016;1(6):692-9.
- 6. Thompson A, Di Angelantonio E, Sarwar N, Erqou S, Saleheen D, Dullaart RP, Keavney B, Ye Z, Danesh J. Association of cholesteryl ester transfer protein genotypes with CETP mass and activity, lipid levels, and coronary risk. JAMA 2008;**299**(23):2777-88.
- 7. HPS3/TIMI55-REVEAL Collaborative Group, Bowman L, Hopewell JC, Chen F, Wallendszus K, Stevens W, Collins R, Wiviott SD, Cannon CP, Braunwald E, Sammons E, Landray MJ. Effects of Anacetrapib in Patients with Atherosclerotic Vascular Disease. N Engl J Med 2017;**377**(13):1217-1227.
- 8. Holmes MV, Davey Smith G. Dyslipidaemia: REVEALing the effect of CETP inhibition in cardiovascular disease. Nat Rev Cardiol 2017;**14**(11):635-636.

- 9. Ference BA, Kastelein JJP, Ginsberg HN, Chapman MJ, Nicholls SJ, Ray KK, Packard CJ, Laufs U, Brook RD, Oliver-Williams C, Butterworth AS, Danesh J, Smith GD, Catapano AL, Sabatine MS. Association of Genetic Variants Related to CETP Inhibitors and Statins With Lipoprotein Levels and Cardiovascular Risk. JAMA 2017;**318**(10):947-956.
- 10. Wurtz P, Wang Q, Soininen P, Kangas AJ, Fatemifar G, Tynkkynen T, Tiainen M, Perola M, Tillin T, Hughes AD, Mantyselka P, Kahonen M, Lehtimaki T, Sattar N, Hingorani AD, Casas JP, Salomaa V, Kivimaki M, Jarvelin MR, Davey Smith G, Vanhala M, Lawlor DA, Raitakari OT, Chaturvedi N, Kettunen J, Ala-Korpela M. Metabolomic Profiling of Statin Use and Genetic Inhibition of HMG-CoA Reductase. J Am Coll Cardiol 2016;67(10):1200-10.
- 11. Sabatti C, Service SK, Hartikainen AL, Pouta A, Ripatti S, Brodsky J, Jones CG, Zaitlen NA, Varilo T, Kaakinen M, Sovio U, Ruokonen A, Laitinen J, Jakkula E, Coin L, Hoggart C, Collins A, Turunen H, Gabriel S, Elliot P, McCarthy MI, Daly MJ, Jarvelin MR, Freimer NB, Peltonen L. Genome-wide association analysis of metabolic traits in a birth cohort from a founder population. Nat Genet 2009;41(1):35-46.
- 12. Raitakari OT, Juonala M, Ronnemaa T, Keltikangas-Jarvinen L, Rasanen L, Pietikainen M, Hutri-Kahonen N, Taittonen L, Jokinen E, Marniemi J, Jula A, Telama R, Kahonen M, Lehtimaki T, Akerblom HK, Viikari JS. Cohort profile: the cardiovascular risk in Young Finns Study. Int J Epidemiol 2008;37(6):1220-6.
- 13. Konttinen H, Silventoinen K, Sarlio-Lahteenkorva S, Mannisto S, Haukkala A. Emotional eating and physical activity self-efficacy as pathways in the association between depressive symptoms and adiposity indicators. Am J Clin Nutr 2010;**92**(5):1031-9.
- 14. Moore C, Sambrook J, Walker M, Tolkien Z, Kaptoge S, Allen D, Mehenny S, Mant J, Di Angelantonio E, Thompson SG, Ouwehand W, Roberts DJ, Danesh J. The INTERVAL trial to determine whether intervals between blood donations can be safely and acceptably decreased to optimise blood supply: study protocol for a randomised controlled trial. Trials 2014;15:363.
- 15. Tillin T, Forouhi NG, McKeigue PM, Chaturvedi N, Group SS. Southall And Brent REvisited: Cohort profile of SABRE, a UK population-based comparison of cardiovascular disease and diabetes in people of European, Indian Asian and African Caribbean origins. Int J Epidemiol 2012;**41**(1):33-42.
- 16. Inouye M, Kettunen J, Soininen P, Silander K, Ripatti S, Kumpula LS, Hamalainen E, Jousilahti P, Kangas AJ, Mannisto S, Savolainen MJ, Jula A, Leiviska J, Palotie A, Salomaa V,

- Perola M, Ala-Korpela M, Peltonen L. Metabonomic, transcriptomic, and genomic variation of a population cohort. Mol Syst Biol 2010;6:441.
- 17. Soininen P, Kangas AJ, Wurtz P, Tukiainen T, Tynkkynen T, Laatikainen R, Jarvelin MR, Kahonen M, Lehtimaki T, Viikari J, Raitakari OT, Savolainen MJ, Ala-Korpela M. Highthroughput serum NMR metabonomics for cost-effective holistic studies on systemic metabolism. Analyst 2009;**134**(9):1781-5.
- 18. Soininen P, Kangas AJ, Wurtz P, Suna T, Ala-Korpela M. Quantitative serum nuclear magnetic resonance metabolomics in cardiovascular epidemiology and genetics. Circ Cardiovasc Genet 2015;8(1):192-206.
- 19. Global Lipids Genetics Consortium. Discovery and refinement of loci associated with lipid levels. Nat Genet 2013;**45**(11):1274-83.
- 20. Kettunen J, Demirkan A, Wurtz P, Draisma HH, Haller T, Rawal R, Vaarhorst A, Kangas AJ, Lyytikainen LP, Pirinen M, Pool R, Sarin AP, Soininen P, Tukiainen T, Wang Q, Tiainen M, Tynkkynen T, Amin N, Zeller T, Beekman M, Deelen J, van Dijk KW, Esko T, Hottenga JJ, van Leeuwen EM, Lehtimaki T, Mihailov E, Rose RJ, de Craen AJ, Gieger C, Kahonen M, Perola M, Blankenberg S, Savolainen MJ, Verhoeven A, Viikari J, Willemsen G, Boomsma DI, van Duijn CM, Eriksson J, Jula A, Jarvelin MR, Kaprio J, Metspalu A, Raitakari O, Salomaa V, Slagboom PE, Waldenberger M, Ripatti S, Ala-Korpela M. Genome-wide study for circulating metabolites identifies 62 loci and reveals novel systemic effects of LPA. Nat Commun 2016;7:11122.
- 21. Do R, Willer CJ, Schmidt EM, Sengupta S, Gao C, Peloso GM, Gustafsson S, Kanoni S, Ganna A, Chen J, Buchkovich ML, Mora S, Beckmann JS, Bragg-Gresham JL, Chang HY, Demirkan A, Den Hertog HM, Donnelly LA, Ehret GB, Esko T, Feitosa MF, Ferreira T, Fischer K, Fontanillas P, Fraser RM, Freitag DF, Gurdasani D, Heikkila K, Hypponen E, Isaacs A, Jackson AU, Johansson A, Johnson T, Kaakinen M, Kettunen J, Kleber ME, Li X, Luan J, Lyytikainen LP, Magnusson PK, Mangino M, Mihailov E, [multiple authors], Wallentin L, Wareham NJ, Whitfield JB, Wolffenbuttel BH, Altshuler D, Ordovas JM, Boerwinkle E, Palmer CN, Thorsteinsdottir U, Chasman DI, Rotter JI, Franks PW, Ripatti S, Cupples LA, Sandhu MS, Rich SS, Boehnke M, Deloukas P, Mohlke KL, Ingelsson E, Abecasis GR, Daly MJ, Neale BM, Kathiresan S. Common variants associated with plasma triglycerides and risk for coronary artery disease. Nat Genet 2013;45(11):1345-52.

- 22. GTEx Consortium. The Genotype-Tissue Expression (GTEx) project. Nat Genet 2013;**45**(6):580-5.
- 23. Consortium. tCD. A comprehensive 1,000 Genomes-based genome-wide association meta-analysis of coronary artery disease. Nat Genet 2015;47(10):1121-1130.
- 24. Wurtz P, Kangas AJ, Soininen P, Lawlor DA, Davey Smith G, Ala-Korpela M. Quantitative Serum NMR Metabolomics in Large-Scale Epidemiology: A Primer on -Omic Technology. Am J Epidemiol 2017;**186**:1084–1096.
- 25. Wang J, Stancakova A, Soininen P, Kangas AJ, Paananen J, Kuusisto J, Ala-Korpela M, Laakso M. Lipoprotein subclass profiles in individuals with varying degrees of glucose tolerance: a population-based study of 9399 Finnish men. J Intern Med 2012;**272**(6):562-72.
- 26. Tukiainen T, Kettunen J, Kangas AJ, Lyytikainen LP, Soininen P, Sarin AP, Tikkanen E, O'Reilly PF, Savolainen MJ, Kaski K, Pouta A, Jula A, Lehtimaki T, Kahonen M, Viikari J, Taskinen MR, Jauhiainen M, Eriksson JG, Raitakari O, Salomaa V, Jarvelin MR, Perola M, Palotie A, Ala-Korpela M, Ripatti S. Detailed metabolic and genetic characterization reveals new associations for 30 known lipid loci. Hum Mol Genet 2012;**21**(6):1444-55.
- 27. Lounila J, Ala-Korpela M, Jokisaari J, Savolainen MJ, Kesaniemi YA. Effects of orientational order and particle size on the NMR line positions of lipoproteins. Phys Rev Lett 1994;**72**(25):4049-4052.
- 28. Kumpula LS, Kumpula JM, Taskinen MR, Jauhiainen M, Kaski K, Ala-Korpela M. Reconsideration of hydrophobic lipid distributions in lipoprotein particles. Chem Phys Lipids 2008;**155**(1):57-62.
- 29. Barter PJ, Kastelein JJ. Targeting cholesteryl ester transfer protein for the prevention and management of cardiovascular disease. J Am Coll Cardiol 2006;47(3):492-9.
- 30. Istvan ES, Deisenhofer J. Structural mechanism for statin inhibition of HMG-CoA reductase. Science 2001;**292**(5519):1160-4.
- 31. Varbo A, Nordestgaard BG. Remnant lipoproteins. Curr Opin Lipidol 2017;**28**(4):300-307.
- 32. Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clin Chem 1972;**18**(6):499-502.

- 33. Martin SS, Blaha MJ, Elshazly MB, Toth PP, Kwiterovich PO, Blumenthal RS, Jones SR. Comparison of a novel method vs the Friedewald equation for estimating low-density lipoprotein cholesterol levels from the standard lipid profile. JAMA 2013;**310**(19):2061-8.
- 34. Niemi J, Makinen VP, Heikkonen J, Tenkanen L, Hiltunen Y, Hannuksela ML, Jauhiainen M, Forsblom C, Taskinen MR, Kesaniemi YA, Savolainen MJ, Kaski K, Groop PH, Kovanen PT, Ala-Korpela M. Estimation of VLDL, IDL, LDL, HDL2, apoA-I, and apoB from the Friedewald inputs--apoB and IDL, but not LDL, are associated with mortality in type 1 diabetes. Ann Med 2009;41(6):451-61.
- 35. Nauck M, Warnick GR, Rifai N. Methods for measurement of LDL-cholesterol: a critical assessment of direct measurement by homogeneous assays versus calculation. Clin Chem 2002;48(2):236-54.
- 36. Lincoff AM, Nicholls SJ, Riesmeyer JS, Barter PJ, Brewer HB, Fox KAA, Gibson CM, Granger C, Menon V, Montalescot G, Rader D, Tall AR, McErlean E, Wolski K, Ruotolo G, Vangerow B, Weerakkody G, Goodman SG, Conde D, McGuire DK, Nicolau JC, Leiva-Pons JL, Pesant Y, Li W, Kandath D, Kouz S, Tahirkheli N, Mason D, Nissen SE, Investigators A. Evacetrapib and Cardiovascular Outcomes in High-Risk Vascular Disease. N Engl J Med 2017;376(20):1933-1942.
- 37. Gaudet D, Alexander VJ, Baker BF, Brisson D, Tremblay K, Singleton W, Geary RS, Hughes SG, Viney NJ, Graham MJ, Crooke RM, Witztum JL, Brunzell JD, Kastelein JJ. Antisense Inhibition of Apolipoprotein C-III in Patients with Hypertriglyceridemia. N Engl J Med 2015;**373**(5):438-47.
- 38. Drenos F, Davey Smith G, Ala-Korpela M, Kettunen J, Wurtz P, Soininen P, Kangas AJ, Dale C, Lawlor DA, Gaunt TR, Casas JP, Timpson NJ. Metabolic Characterization of a Rare Genetic Variation Within APOC3 and Its Lipoprotein Lipase-Independent Effects. Circ Cardiovasc Genet 2016;**9**(3):231-9.
- 39. Wurtz P, Havulinna AS, Soininen P, Tynkkynen T, Prieto-Merino D, Tillin T, Ghorbani A, Artati A, Wang Q, Tiainen M, Kangas AJ, Kettunen J, Kaikkonen J, Mikkila V, Jula A, Kahonen M, Lehtimaki T, Lawlor DA, Gaunt TR, Hughes AD, Sattar N, Illig T, Adamski J, Wang TJ, Perola M, Ripatti S, Vasan RS, Raitakari OT, Gerszten RE, Casas JP, Chaturvedi N, Ala-Korpela M, Salomaa V. Metabolite profiling and cardiovascular event risk: a prospective study of 3 population-based cohorts. Circulation 2015;**131**(9):774-85.



-0.4

CETP

-0.8

Figure 1. Associations of genetic variants in CETP rs247617 (red) and HMGCR rs12916 (blue) with circulating apolipoprotein B and cholesterol concentrations in size-specific apolipoprotein B particles. Estimates represent the standardized difference in lipoprotein trait, with per-allele associations scaled to a 10% lower risk of CHD. Analyses were adjusted for age, sex, genotyping batch and ten genetic principal components. Closed circles represent statistical significance of associations at P<0.002 and open circles associations that are non-significant at this threshold. The lipoprotein subclasses are defined by particle size: 17, 18, 25 potential chylomicrons and the largest very-low-density lipoprotein particles (XXL-VLDL; average particle diameter ≥75 nm); five different VLDL subclasses, i.e. very large (average particle diameter 64.0 nm), large (53.6 nm), medium (44.5 nm), small (36.8 nm) and very small VLDL

SD change per 10% decrease in CHD odds ratio

0.4

- HMGCR

(31.3 nm); intermediate-density lipoprotein (IDL; 28.6 nm); and three LDL subclasses, i.e. large (25.5 nm), medium (23.0 nm) and small LDL (18.7 nm).

APO B PARTICLES & TRIGLYCERIDES

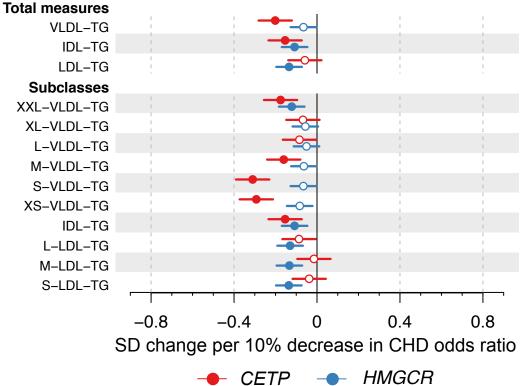


Figure 2. Associations of genetic variants in *CETP* rs247617 (red) and *HMGCR* rs12916 (blue) with circulating triglyceride concentrations in size-specific apolipoprotein B particles. The estimates and lipoprotein subclasses are as defined in the caption for Fig. 1.

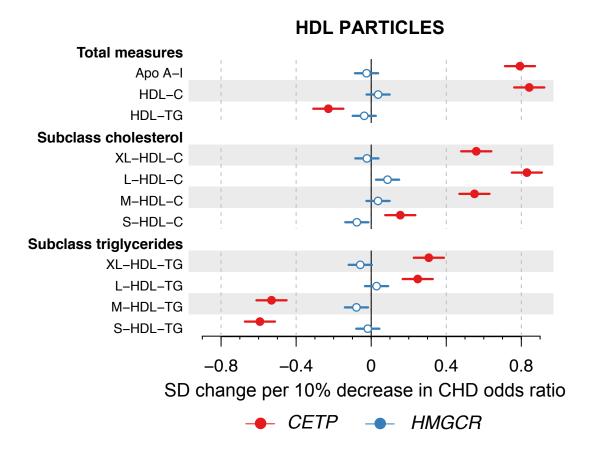


Figure 3. Associations of genetic variants in *CETP* rs247617 (red) and *HMGCR* rs12916 (blue) with circulating apolipoprotein A-I as well as cholesterol and triglyceride concentrations in size-specific HDL particles. The estimates are as defined in the caption for Fig. 1. The four size-specific HDL subclasses are very large (average particle diameter 14.3 nm), large (12.1 nm), medium (10.9 nm) and small HDL (8.7 nm).

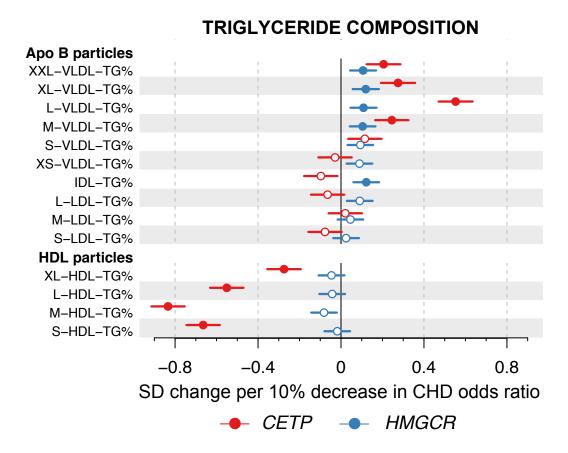


Figure 4. Associations of genetic variants in *CETP* rs247617 (red) and *HMGCR* rs12916 (blue) with the triglyceride composition of size-specific lipoprotein particles. The estimates are as defined in the caption for Fig. 1 and the lipoprotein subclasses are as defined in the captions for Fig. 1 and Fig. 3.

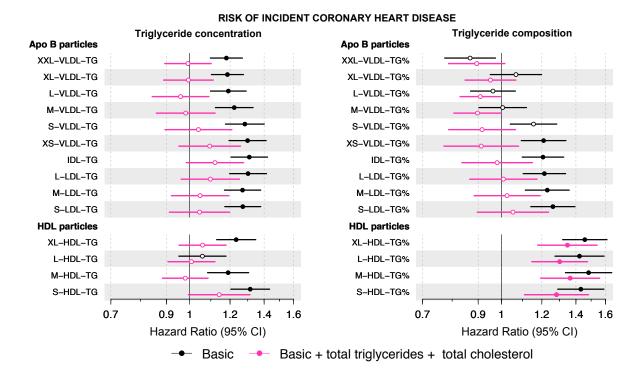


Figure 5. Observational associations of circulating triglyceride concentrations and triglyceride composition in lipoprotein subclass particles and risk of incident coronary heart disease.

Left pane: Black: Hazard ratios for incident CHD per-SD higher triglyceride concentration within each size-specific lipoprotein subclass adjusted for traditional risk factors. Pink: adjusted for traditional risk factors, serum cholesterol and serum triglycerides. Right pane: Black: Hazard ratios for incident CHD per-SD higher percentage of triglycerides (of all lipid molecules) within each size-specific lipoprotein subclass adjusted for traditional risk factors. Pink: adjusted for traditional risk factors, serum cholesterol and serum triglycerides. Basic risk factors include age, sex, mean arterial pressure, smoking, type 2 diabetes mellitus, lipid medication, geographical region (FINRISK) and ethnicity (SABRE). Closed circles represent statistical significance of associations at P<0.002 and open circles associations that are non-significant at this threshold.