

Costless metabolic secretions as drivers of interspecies interactions in microbial ecosystems

Alan R. Pacheco¹, Mauricio Moel², Daniel Segrè^{*1,2,3}

¹ Graduate Program in Bioinformatics and Biological Design Center, Boston University,
Boston, MA 02215, USA

² Department of Biology, Boston University, Boston, MA 02215, USA

³ Department of Biomedical Engineering and Department of Physics, Boston University,
Boston, MA 02215, USA

* To whom correspondence should be addressed: dsegre@bu.edu

1 **ABSTRACT**

2 Metabolic exchange can mediate beneficial interactions among microbes, helping
3 explain diversity in microbial communities. These interactions are often assumed to
4 involve a fitness cost, prompting questions on how cooperative phenotypes can be
5 stable and withstand the emergence of cheaters. Here we use genome-scale models of
6 metabolism to investigate whether a radically different scenario, the pervasive release
7 of “costless” metabolites (i.e. those that cause no fitness cost to the producing
8 organism), can serve as a prominent mechanism for inter-microbial interactions. By
9 carrying out over 1 million pairwise growth simulations for 14 microbial species in a
10 combinatorial assortment of environmental conditions, we find that there is indeed a
11 large space of metabolites that can be secreted at no cost, which can generate ample
12 cross-feeding opportunities. In addition to providing an atlas of putative costless
13 interdependencies, our modeling also demonstrates that oxygen availability significantly
14 enhances mutualistic interactions by providing more opportunities for metabolic
15 exchange through costless metabolites, resulting in an over-representation of specific
16 ecological network motifs. In addition to helping explain natural diversity, we show how
17 the exchange of costless metabolites can facilitate the engineering of stable synthetic
18 microbial consortia.

19

20 **Keywords:** Microbial communities, cross-feeding, cooperation, synthetic ecology,
21 genome-scale modeling, microbiome

22

23 **INTRODUCTION**

24 The astonishing number of microbial species observed in nature ^{1–3} seems to contradict
25 classical ecological theory, which predicts far less biodiversity in many nutrient-poor
26 environments ^{4,5}. A variety of different explanations have been proposed as possible
27 solutions to this inconsistency, including resource partitioning ⁶, differential nutrient use ⁷,
28 spatial segregation ⁸, and metabolic cross-feeding ^{9–11}. In environments poor in
29 resources, cross-feeding has been shown to enhance the capacity of microbes to survive,
30 either through the secretion of valuable compounds ^{12–14}, or by maintaining
31 thermodynamic gradients necessary for continued metabolism ¹⁵. Despite their
32 prevalence, it is not clear how these cooperative phenotypes emerge, as they often
33 involve the exchange of metabolites that are costly for the producer. This apparent
34 altruism introduces the potential for the rise of cheating organisms that do not contribute
35 common goods but still benefit metabolically from others, challenging community stability

36 16. Previous studies have addressed this dilemma in different ways, either by suggesting
37 fundamental boundaries to the feasibility of costly cross-feeding based on theoretical
38 considerations¹⁷, or by establishing balances between production costs and reciprocation
39 benefits in specific communities and environments^{18–20}. However, it is not fully
40 understood whether costly exchanges can account for the degree of biodiversity observed
41 in nature, as the conditions necessary for the rise and maintenance of these costly
42 interdependencies may not frequently manifest themselves.

43
44 We therefore ask whether a radically different interaction mechanism, one that hinges on
45 organisms secreting metabolic products at no cost to their own fitness, may be prevalent
46 enough in the microbial world to help explain the abundance of cross-feeding
47 opportunities. Key to this mechanism, which we will term “costless,” is the emergence of
48 community benefits as a product of otherwise selfish acts by individual microbial species.
49 This phenomenon has been explored in a macroecological context^{21–23} and can be
50 illustrated by the example of a vulture consuming the remains of a lion kill. Here, the lion
51 gains nutritional benefit from its hunt and leaves behind scraps of food that are in turn
52 eaten by the vulture. In this way, though the lion did not expend energy to facilitate access
53 to food explicitly for the vulture, it did unintentionally contribute to the vulture’s success
54 through its own selfish action²⁴. It is known that, in the microbial world, metabolic waste
55 products secreted at no cost to the producing organism (e.g. *E. coli* secreting acetate
56 under limited oxygen) can serve to support other species¹³. However, it is not obvious
57 whether such behavior extends beyond a few fermentation byproducts. Moreover, little
58 information exists on how costless secretions vary across microbial species and growth
59 media composition. Most importantly, even if the metabolites secreted by an organism
60 under a given condition were to be known, it still would be difficult to ascertain whether
61 such byproducts would be likely to enable or enhance growth of other species.

62
63 In this study, we use computational metabolic modeling to quantify the magnitude of
64 environmental modification brought about by costless metabolite secretion, as well as the
65 interspecies interactions that can arise from this type of exchange. In a microbial analog

66 to the lion-vulture interaction, we seek to understand how metabolites released as a
67 product of selfish action by individual species yield unintended benefits to partner
68 organisms, resulting in emergent interspecies cooperation. Based on this framework, we
69 present a computational pipeline based on flux balance analysis (FBA) ²⁵ that predicts
70 the growth phenotypes and cooperative interactions mediated by costless metabolites for
71 14 microbial species under a large combinatorial set of environmental conditions. In this
72 way, we obtain a global view of cross-feeding opportunities that can mediate the
73 emergence of cooperation and the maintenance of biodiversity in natural communities. In
74 addition, we complement our metabolic modeling with a dynamical modeling framework
75 to understand whether costless secretions on their own can promote long-term stability
76 in model synthetic microbial communities. While the present work focuses entirely on
77 putative secretions and interactions predicted computationally, we wish to highlight that
78 we restricted our analysis to microbes associated with high quality, manually curated (and
79 therefore in most cases individually tested) *in silico* models and that in many cases,
80 specific predictions can be shown to be consistent with previously established empirical
81 knowledge. For the most part, however, the current analysis should be viewed as the
82 exploration of a large space of stoichiometrically possible costless interactions
83 (inscrutable to such an extent at the experimental level), whose global patterns could
84 motivate and inform future experimental and theoretical endeavors.

85

86 RESULTS

87 **Metabolite secretions can be costly or costless, depending on environmental**
88 **context.** Understanding whether or not the secretion of a specific metabolite by a given
89 organism is associated with a decrease in fitness (interpreted here as growth rate) is
90 difficult to assess experimentally, but can be readily addressed using genome-scale
91 models of metabolism (see Methods). For example, it is possible to impose the secretion
92 of a given compound at a given rate, and then ask whether this constraint is expected to
93 cause a reduction in growth. A small set of simulations of this kind for a single organism
94 (Fig. S1) exemplifies the broad spectrum of possible outcomes: based on the specific
95 carbon sources, different metabolites can be produced, sometimes at the expense of

96 growth capacity, other times with no apparent effect (neutral), or even to its benefit. Due
97 to the basic assumptions of the genome-scale models we employed (especially the
98 maximization of growth as the objective function) we know that these last two kinds of
99 secretions are compatible (or even necessary) for metabolism to operate at maximal
100 efficiency. We will refer to these beneficial or neutral secretions as ‘costless.’

101

102 **Secretion of costless metabolites leads to substantive environmental enrichment.**

103 Having illustrated in an individual case how metabolite secretion costs can strongly
104 depend on carbon sources, we sought to map the prevalence of costless secretions
105 across a broad set of organisms and environments, as well as the chance that such
106 secreted metabolites could mediate cross-feeding. We carried out a total of 1,051,596
107 unique *in silico* simulations, each with two organisms i and j from a set of 14 genome-
108 scale metabolic models and two carbon sources α and β from a set of 108 compounds
109 (Figure 1, Supplementary Information 1, 2). Each simulation is conducted as an iterative
110 process that simulates a coculture experiment, and is uniquely defined by the organisms
111 involved, the carbon sources provided, and the availability of oxygen. At each iteration,
112 we used FBA to determine the ability of each organism to grow on the current medium
113 (see Methods, and Fig. 1a). As an outcome of this calculation, we also obtained
114 information about the set of metabolites predicted to be spontaneously (i.e. costlessly)
115 secreted by each microbe. If at the first iteration ($c = 1$) at least one *in silico* organism
116 was able to grow on the carbon sources provided, all costless metabolites were added to
117 the medium for the next iteration. This process was repeated until no new metabolites
118 were produced. The final iteration c before attaining this steady state is defined as c_s .
119 Upon running these iteration simulations for all possible combinations of species and
120 environments and selecting only the cases in which both species grew, we obtained
121 distributions for the value of c_s (Figure 2a). A large number of cases reached a steady
122 state after only one iteration (92% of cases with oxygen, and in 82% without oxygen).
123 This could in principle be due to a complete lack of costless secretions at the first iteration.
124 However, as demonstrated by Fig. 2b, the skewness in the distribution of c_s is better

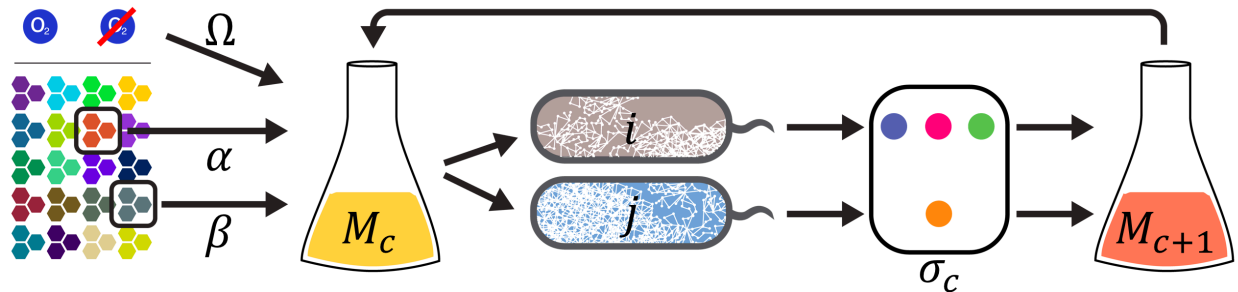


Figure 1. Simplified schematic of example computational pairwise cross-feeding simulation. Simplified schematic of an *in silico* experiment: A growth medium (M_c) containing two carbon sources (α, β) with or without oxygen (Ω) is provided to genome-scale metabolic models of two microbial organisms (i, j). If at least one organism grows, any costlessly-secreted metabolites (σ_c) are added to the medium, which is fed back to the organisms. This process is repeated for a series of iterations c , and terminates at iteration c_s , defined as the last iteration in which any new metabolites were secreted into the medium.

125 explained by the alternative hypothesis that organisms do secrete multiple byproducts in
126 the first iteration, but these byproducts contribute weakly to additional secretions in
127 subsequent iterations.

128

129 In aggregate, our simulations showed a rightward shift in the diversity of metabolites
130 secreted under anoxic conditions when compared to the number secreted when oxygen
131 was present. To understand this effect, we looked at the distribution of the number of
132 metabolites secreted after the first iteration, which is equivalent to growing each organism
133 on its own in the provided medium. This distribution for $c = 1$ was unimodal for both
134 conditions, centered between two and three metabolites with oxygen and around five
135 metabolites without oxygen (Figure 2b). After this first iteration, the maximum number of
136 secreted metabolites was 11 with oxygen and 16 without oxygen. In the anoxic
137 simulations, the central carbon metabolites most commonly secreted after the first
138 iteration were fermentation byproducts such as acetate, formate, succinate, and ethanol.
139 These metabolites were secreted in 87.5%, 74.5%, 25.7%, and 20.2% of growth-yielding
140 simulations respectively. With oxygen, the most commonly secreted central carbon
141 metabolites after the first iteration were formate and acetate, secreted in 46.8% and
142 18.3% of growth-yielding simulations respectively. We may therefore chiefly attribute the
143 shift between the oxic and anoxic secretion curves to the anoxic export of incompletely-
144 reduced core metabolism intermediates.

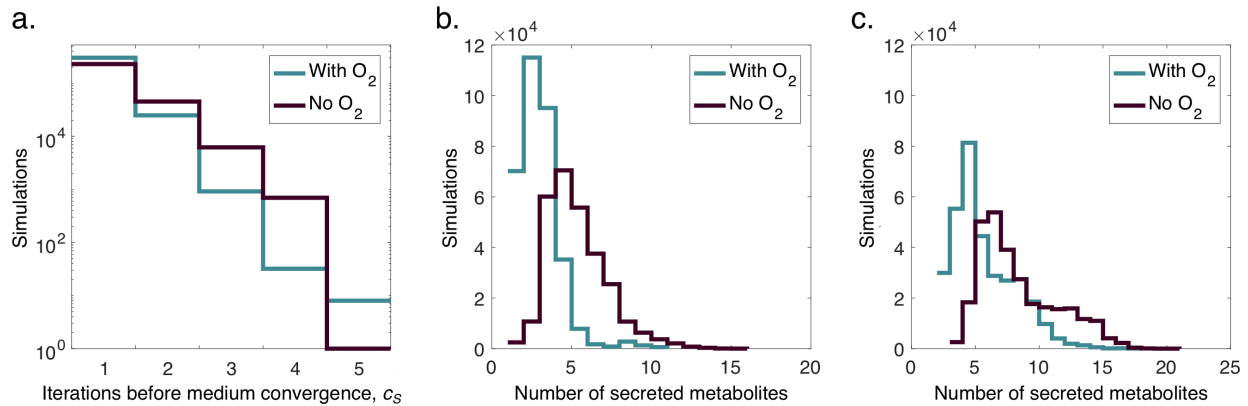


Figure 2. Analysis of costlessly-secreted metabolites in pairwise *in silico* experiments that led to growth of at least one organism. (a) Distribution of number of expansions until final medium expansion iteration. (b) Distribution of the number of metabolites secreted into the medium by one or both organisms in a pair after one iteration of FBA. (c) Distribution of the number of metabolites secreted by one or both organisms after the last iteration of FBA (c_s). The last iteration is defined as the iteration in which no additional metabolites were secreted into the medium. Despite the large variability in number of expansions and number of secreted metabolites, we observe a poor correlation between these distributions, indicating that a simulation resulting in a high number of expansions does not necessarily result in a high number of metabolites being secreted (Figure S3).

145 In addition to a positive shift observed between anoxic and oxic conditions, our results
146 also show a shift in the quantity of metabolites secreted between the first and last iteration
147 of each computational experiment (Figure 2c). This effect reflects organisms taking up
148 metabolites secreted by themselves or their partner, and secreting different metabolites
149 as a response. After the last medium expansion iteration for all simulations, the total
150 number of secreted metabolites followed similar distributions with a maximum at 18 and
151 21 metabolites for oxic and anoxic conditions, respectively. This positive shift suggests a
152 response from one or both organisms to a medium iteratively enriched by costless
153 byproducts, which hints at their potential metabolic utility. Principal component analysis
154 (PCA) shows that neither the environment nor the species alone can explain the variability
155 in secretion profiles (Figure S4), suggesting that a combination of both variables accounts
156 for the range in costlessly-secreted products.

157

158 **Useful costlessly-secreted byproducts are abundant.** Our analysis reveals a broad
159 distribution of metabolically useful compounds secreted without cost in a variety of
160 environmental conditions by most organisms (Figure 3, Figure S5a). Though inorganic
161 compounds such as water and carbon dioxide were, as expected, the most commonly

162 secreted compounds across all simulations, nitrogen-containing compounds such as
163 nitrite, ammonium, urea, and trimethylglycine were secreted in 73.5% of the analyzed
164 cases, suggesting maintenance of an appropriate carbon-to-nitrogen ratio in the cell. We
165 note specifically that nitrite is secreted in fewer than 100 simulations with oxygen, but
166 almost universally in anoxic simulations - a phenomenon previously observed in
167 anaerobic enteric bacteria ²⁶. Organic acids make up the second most abundant category
168 of costlessly-secreted byproducts, constituting 23% and 36% of unique metabolites with
169 and without oxygen respectively. Notably, we also observe secretion of nucleotides,
170 peptides, and carbohydrates in a combined 9% and 13% of simulations with and without
171 oxygen respectively. Altogether, this space of secreted metabolites points to a large
172 variety of molecules that can be freely produced, suggesting that costless metabolic
173 secretion may provide substantial degrees of environmental enrichment. This effect
174 becomes magnified considering the relative scarcity of resources provided in our minimal
175 medium, which suggests that costless secretions play a fundamental role in promoting
176 metabolic diversity in natural environments.

177

178 Given the abundance and complexity of secretions from different organisms, as well as
179 the possible ecological connections they may promote, we asked whether specific
180 metabolite secretions were highly correlated. As patterns in environmental modification
181 through secretion have an impact on the species composition of a microbial community
182 ²⁷, it becomes important to understand which metabolites co-occur within our set of
183 simulations. To address this question, we performed a Spearman correlation analysis to
184 determine common secretion patterns (Figure S6). In the presence of oxygen, we observe
185 a strong co-occurrence of glycerol, lactate, succinate, malate, and acetate, which
186 correlates with the high frequency of secretion of these carbon-containing compounds
187 (Figure S5a). We also observe positive, but weaker correlations between these
188 metabolites and other central carbon compounds such as fumarate, citrate, and 2-
189 oxoglutarate. Our analysis also points to the simultaneous release of multiple nitrogen-
190 containing compounds, chiefly urea, ammonium, and nitrate. Without oxygen, we observe

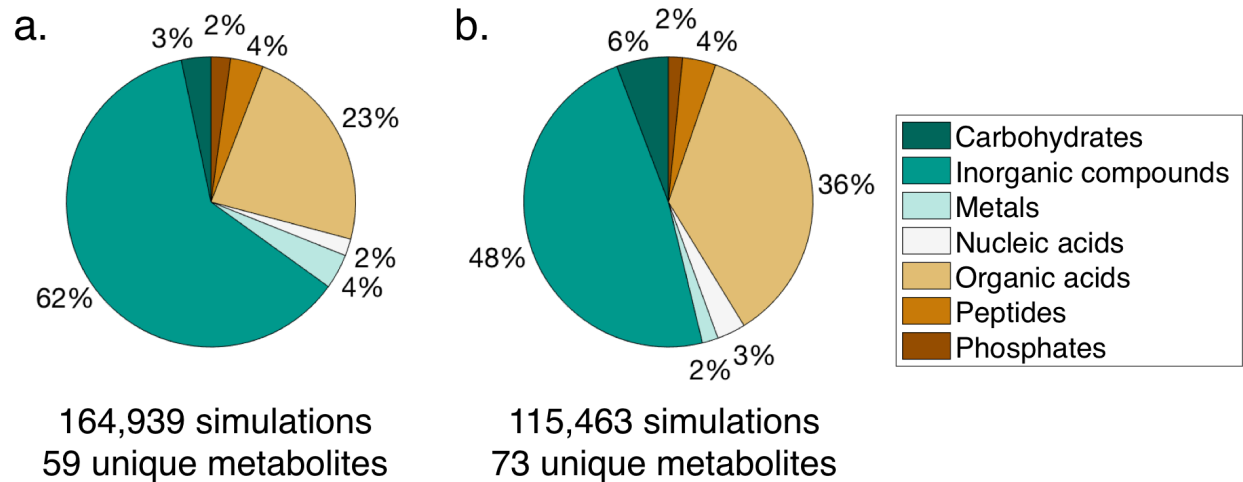


Figure 3. Categorization of metabolites secreted costlessly in oxic (a) and anoxic (b) conditions.

191 stronger correlations between secretion of nitrogen-containing compounds and
192 fermentation byproducts. Amino acids also co-occur with high frequency without oxygen,
193 particularly cysteine, methionine, and alanine – which itself is associated with export of
194 proline and glutamine. These patterns are consistent with specific examples of previously
195 studied exometabolomic profiles, including those showing co-secretion of central carbon
196 intermediates in *E. coli* and of amino acids in yeast²⁸, as well as time-dependent patterns
197 of metabolites released simultaneously in soil communities²⁹. In summary, by promoting
198 secretion of a larger number of metabolites across a wide space of conditions, these co-
199 secretion profiles may result in enhanced metabolic enrichment of the environments in
200 our simulation set.

201

202 We note that while a potentially useful metabolite can be secreted into the environment
203 by one species, it does not necessarily mean that it will be consumed by a second
204 organism. We place particular importance on this distinction, as any interspecies
205 interaction must also take into account the decision to import a novel metabolite found in
206 the environment. To map this distinction, we examine the space of costless metabolites
207 that are exchanged by each organism across all *in silico* experiments (Figure S5b). Here,
208 the most commonly exchanged organic metabolites were central carbon intermediates,
209 secreted mostly in anoxic conditions. These secretion patterns mirror those of anoxic gut
210 bacteria, which divide the task of digesting complex polysaccharides by exchanging

211 intermediate organic acids^{9,30}. Importantly, we observed that amino acids, secreted
212 chiefly by *S. cerevisiae*, but also in a substantial number of simulations by *S. enterica*, *K.*
213 *pneumoniae*, and *E. coli*, were among the most highly-exchanged costless metabolites.
214 This phenomenon has been previously documented in relation to overflow metabolism in
215 *S. cerevisiae*³¹ and *E. coli*^{32,33}, as well as in yeast-bacteria symbioses^{34,35}, and account
216 for exchange in over 10⁴ simulations with and without oxygen in our study. This high
217 prevalence of exchange underscores the metabolic utility of these secreted byproducts,
218 particularly when contrasted with patterns of secretion in which the most commonly
219 released metabolites were of low or no metabolic utility to a partner organism (e.g. water).

220
221 **Costless metabolite exchange enhances growth capabilities.** Having mapped the
222 space of metabolites that can be secreted costlessly across a large variety of contexts,
223 we asked if these secreted byproducts could directly enable the growth of other
224 organisms. We find that with oxygen, 95,519 *in silico* experiments predicted growth of
225 both organisms in the minimal medium, accounting for 18.2% of all 525,798 oxic
226 simulations (Figure 4a). Under anoxic conditions, only 11.9% of simulations resulted in
227 growth of both organisms in the minimal medium alone. After the organism pairs were
228 allowed to exchange costlessly-secreted metabolites, our algorithm predicted that 31.4%
229 and 22.0% of simulations would result in both species growing with and without oxygen,
230 respectively. This stage of growth, at $c = c_s$, is analogous to both species growing in the
231 presence of each other's secreted metabolites *in vivo*. This enhanced growth potential in
232 coculture represents a 72.7% increase in growth-supporting environments with oxygen
233 and an 82.5% increase in environments without oxygen, suggesting that exchange of
234 costlessly-secreted metabolites can enable growth of additional organisms in resource-
235 poor environments.

236
237 Though application of our algorithm resulted in a global increase in growth capabilities
238 due to costless metabolite secretion, species-specific growth patterns varied widely
239 across our dataset (Figure 4b). We look specifically at *L. lactis* and *P. gingivalis*, host-
240 associated microbes present in the human gut and oral microbiomes respectively. Both

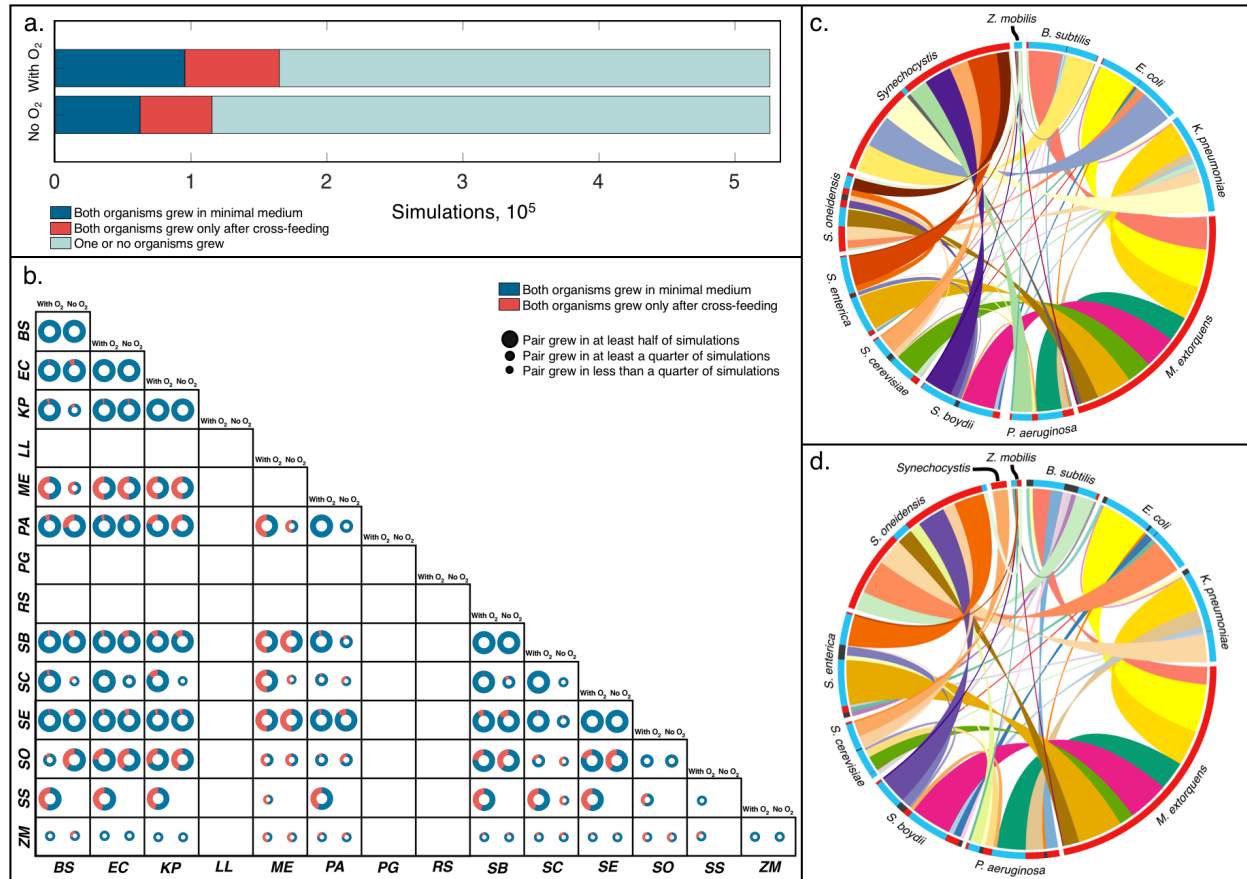


Figure 4. Growth outcomes of pairwise cross-feeding simulations based on organisms and carbon sources. (a) Growth outcomes of all *in silico* experiments with and without oxygen, grouped by pairwise growth phenotype. **(b)** Organism-specific growth outcomes. Size of circles represent the relative number of environments in which an organism was able to grow out of 5,774 *in silico* experiments with each partner. Organisms are abbreviated as follows: BS: *B. subtilis*; EC: *E. coli*; KP: *K. pneumoniae*; LL: *L. lactis*; ME: *M. extorquens*; PA: *P. aeruginosa*; PG: *P. gingivalis*; RS: *R. sphaeroides*; SB: *S. boydii*; SC: *S. cerevisiae*; SE: *S. enterica*; SO: *S. oneidensis*; SS: *Synechocystis*; ZM: *Z. mobilis*. **(c, d)** Frequency of obligate pairwise growth by species in single carbon source simulations for oxic (N = 69,420, c) and anoxic (N = 52,897, d) conditions. Each color ribbon is unique to an individual species pair. Width of ribbons is proportional to the number of experiments in which obligate syntrophy was predicted for each species pair. Radial axis colors represent directionality of exchange: Blue: Organism provided essential metabolites to partner organism in over 75% of simulations; Red: Organism received essential metabolites in over 75% of simulations; Gray: Both organisms gave and received essential nutrients in most simulations.

241 organisms are auxotrophic for a wide range of amino acids and other central metabolites,
 242 necessitating dependence on a rich set of metabolic products produced by the host or
 243 other commensal microbes. In our simulations, however, these organisms failed to grow
 244 in all environments and with all species pairs even after any costless metabolites were
 245 secreted. This failure to sustain growth of highly dependent organisms suggests that there
 246 is an upper limit to the degree to which costless metabolite production can enable species

247 growth, especially in the minimal environments that were tested. Aside from these
248 extreme cases, our analysis sheds light on the performance of generalist organisms, such
249 as *E. coli*, *K. pneumoniae*, *S. cerevisiae*, and *S. enterica*. These organisms grew in at
250 least half of all tested environmental conditions, in contrast with organisms such as *M.*
251 *extorquens* or *Z. mobilis*, which exhibited much more limited pairwise growth capabilities.
252 These patterns suggest a greater dependence of these organisms on the metabolic
253 byproducts of their partners, particularly in anoxic conditions. These patterns underscore
254 the importance of not only the number of metabolites secreted, but also of the specific
255 metabolic needs of the receiving organism in determining the contribution of costless
256 metabolites to the growth of a partner.

257

258 **Exchange mediated by costless metabolites yields species-specific obligate**
259 **partnerships.** After analyzing general growth outcomes across our entire simulation set,
260 we sought to determine which specific organisms could not grow in our environments
261 without the costless secretions of a partner. This question is of particular interest as co-
262 occurrence in natural communities is widespread^{36–38} and suggests patterns of species
263 codependence³⁹, potentially providing a mechanistic view into the assembly of complex
264 microbial ecosystems. Our simulations identified a diverse space of codependent
265 organisms, with most species exhibiting at least one case of obligate syntrophy with all
266 others (Figure 4 c, d). Many organisms had balanced distributions of codependence
267 (organism A enabled the growth of organism B in some cases, and organism B enabled
268 the growth of A in others), but the majority of co-dependent relationships were
269 unidirectional. One striking example of this phenomenon is that of cyanobacteria and
270 heterotrophic organisms, with *Synechocystis* (grown here in the absence of light)
271 indicating high degrees of dependence on other organisms. With oxygen, *Synechocystis*
272 was dependent on 9 different organisms across the vast majority of simulations in which
273 it grew with a partner. As all organisms were grown heterotrophically, carbon dioxide and
274 ammonium were the main byproducts that enabled growth of *Synechocystis* in these
275 simulations. Previous studies have confirmed ammonium as the preferred nitrogen
276 source of cyanobacteria^{40–42}, indicating that the ability to fix carbon and consume nitrogen

277 are accurately reflected in the *in silico* metabolic requirements of *Synechocystis*. We also
278 observed that *E. coli*, *B. subtilis*, and *S. cerevisiae*, three species commonly used as
279 model microbial organisms, were more frequently the giving organisms in cases of
280 obligate syntrophy. These pairings not only shed light on the mechanisms behind
281 interspecies codependencies, but may also serve as a map for assembling co-dependent
282 synthetic communities stabilized by costless metabolic exchange.

283
284 **Carbon sources exhibit cooperativity in determining growth potential.** In addition to
285 characterizing the global space of *in silico* growth phenotypes, we examined how
286 cooperativity of primary carbon sources could enhance growth capabilities in organism
287 pairs. Drawing from techniques used to quantify epistatic interactions⁴³, we defined the
288 cooperativity index C of two carbon sources α and β as the difference between the number
289 of simulations that result in growth from both carbon sources and the product of the
290 number of simulations that result from single carbon sources. These counts were
291 normalized by the total number of simulations involving the specific pairing of carbon
292 sources being analyzed (represented here by the combinatorial formula $\binom{N}{2}$), as follows:

293

$$C^{\alpha,\beta} = \frac{g_{\alpha,\beta}}{\binom{N_{\alpha,\beta}}{2}} - \left(\frac{g_{\alpha}}{\binom{N_{\alpha}}{2}} * \frac{g_{\beta}}{\binom{N_{\beta}}{2}} \right) \quad (\text{E1})$$

294
295 This metric therefore aims to reflect the cooperative potential of each carbon source pair
296 relative to that of each carbon source in isolation. In this way, when averaging a single
297 carbon source over its cooperativity index, we obtain a relative degree to which a carbon
298 source “depends” on another to sustain growth. By framing cooperativity in this context,
299 we observed that simple sugars such as glucose and sucrose had relatively low
300 cooperativity indices, that is, they were able to sustain growth efficiently on their own. In
301 contrast, more complex molecules and dipeptides had higher average cooperativity
302 indices, indicating they performed better in the presence of another carbon source. We
303 grouped these average cooperativity indices through hierarchical clustering (Figure S7)

304 and observed general clustering by carbon source type – especially with sugars and
305 amino acids appearing in distinct groups. This analysis illustrates the nonlinear effects of
306 adding additional nutrients to a minimal medium, underscoring the observed complex
307 metabolite usage patterns in organism pairs.

308

309 **Organisms competing for the same carbon source can simultaneously benefit each**
310 **other through costless secretions.** Our analysis so far has examined the contexts in
311 which a metabolite can be secreted costlessly, as well as the potential for these
312 metabolites to promote growth. Based on these insights, we wished to more
313 fundamentally understand these interspecies interactions and how they compare to
314 ecological expectations of cooperation and competition. To do this, we defined six types
315 of possible interactions: non-interaction, commensalism (unidirectional exchange), and
316 mutualism (bidirectional exchange), each with or without competition for a primary carbon
317 source. We chose to decouple competition for nutrients from cooperation via secreted
318 metabolites in order to more fully understand the degree to which the latter can promote
319 organism coexistence despite resource scarcity (Figure 5a). When analyzing our dataset
320 under this framework, we found that competition for one or both carbon sources
321 constituted the majority of the space of all interactions across all simulations (Figure 5b),
322 as previously observed experimentally ⁴⁴. However, these predicted competitive
323 phenotypes were observed to occur simultaneously with potentially beneficial interactions
324 mediated by metabolic byproducts. Here, we found that uni- and bidirectional exchange
325 accounted for a majority of all interactions predicted with and without the presence of
326 oxygen.

327

328 Our modeling predicted bidirectional interactions to be far more common without oxygen
329 than with oxygen (Figure 5c). We obtained a more fine-grained perspective on costless
330 metabolic interactions by considering the distributions of interaction types by species
331 pairs (Figure 5d). For example, the majority of pairings of *M. extorquens* with *B. subtilis*,
332 *E. coli*, and *K. pneumoniae* exhibited commensal interactions (chiefly with *M. extorquens*
333 receiving). In contrast, the distribution of interactions shifted toward mutualism when

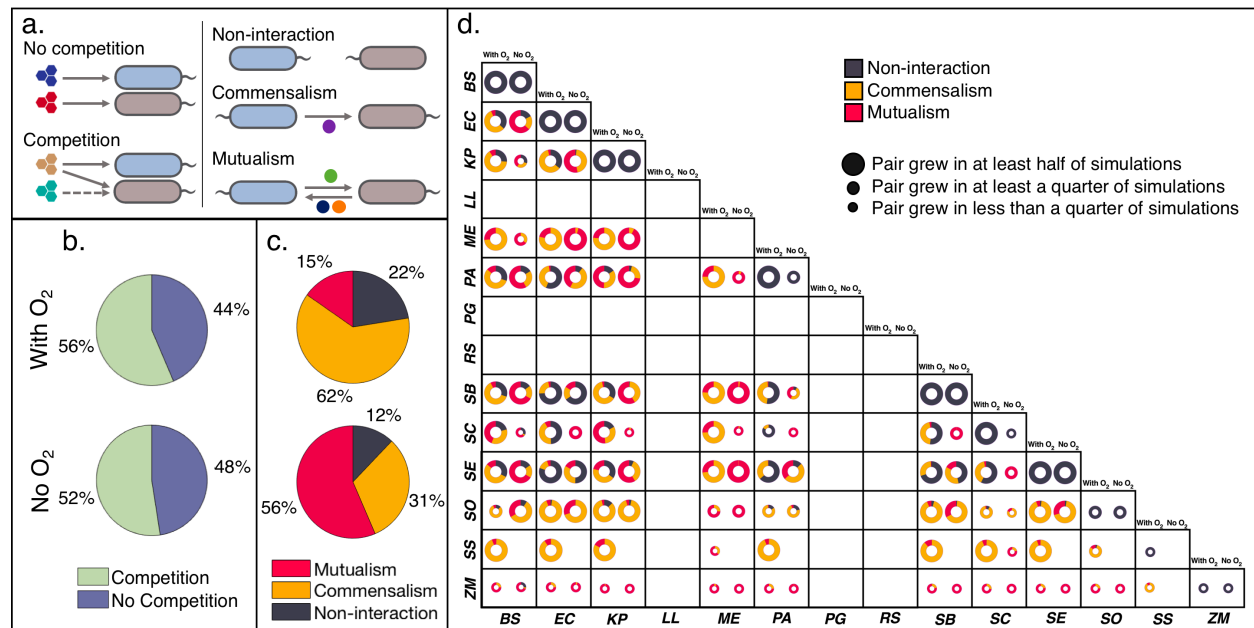


Figure 5. Distribution of metabolic interaction types. (a) Schematic representation of interaction types arising from costlessly-secreted metabolites. Competition is defined as both organisms consuming the same carbon source. Commensalism is defined as a unidirectional exchange of one or more costlessly-secreted metabolites, and mutualism is defined as a bidirectional exchange of one or more costlessly-secrete metabolites. (b) Overall distributions of competitive/noncompetitive interactions for oxic (out of 164,939 simulations that yielded pairwise growth) and anoxic conditions (out of 115,463 simulations that yielded pairwise growth). (c) Overall distributions of general interactions mediated by costless metabolites for oxic and anoxic conditions. These interactions at the level of secreted metabolites exist simultaneously with competition or no competition for a primary carbon source. (d) Organism-specific growth outcomes and interaction type distributions. Size of circles represent the relative number of environments in which an organism was able to grow out of 5,774 *in silico* experiments with each partner. Organisms are abbreviated as follows: BS: *B. subtilis*; EC: *E. coli*; KP: *K. pneumoniae*; LL: *L. lactis*; ME: *M. extorquens*; PA: *P. aeruginosa*; PG: *P. gingivalis*; RS: *R. sphaeroides*; SB: *S. boydii*; SC: *S. cerevisiae*; SE: *S. enterica*; SO: *S. oneidensis*; SS: *Synechocystis*; ZM: *Z. mobilis*.

334 oxygen was made unavailable. These patterns were also mirrored in a majority of
 335 individual species pairings. As with the positive shift observed in the distributions of
 336 secreted metabolites (Figure 2b, c), we may attribute the increased prevalence of
 337 mutualistic interactions without oxygen to a greater availability of metabolic byproducts
 338 that can contribute to reciprocity. To test this hypothesis, we performed a small subset of
 339 “hybrid” *in silico* experiments, where we analyzed the interactions that arose from one
 340 species being grown in the presence of oxygen and the other anaerobically. We looked at
 341 the examples of *E. coli* with *B. subtilis* and *S. enterica*, whose pairwise simulations
 342 showed greater amounts of mutualistic interactions without oxygen. When *E. coli* was
 343 grown anaerobically but its partner was grown with oxygen, the vast majority of

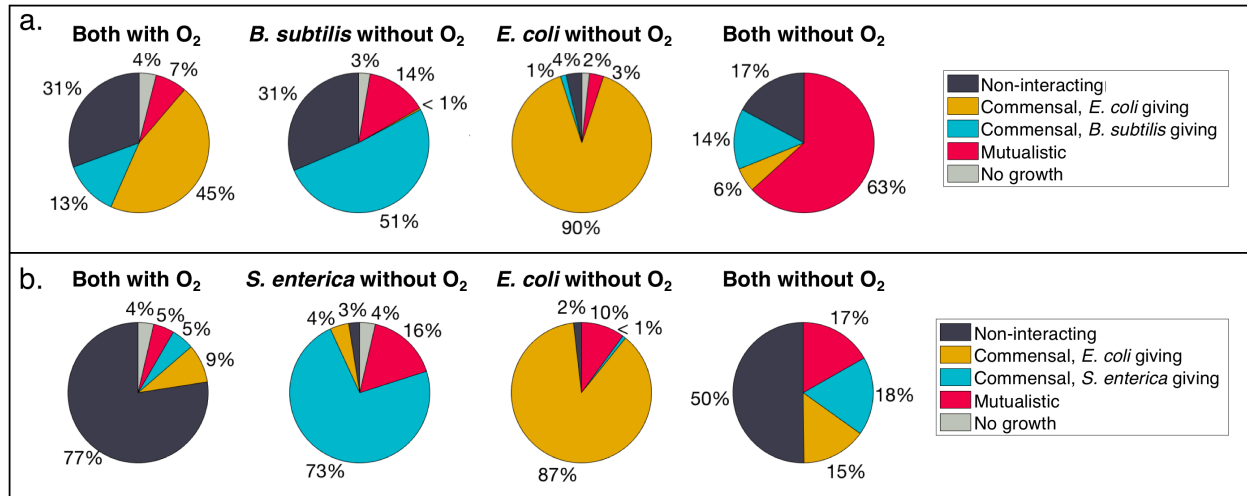


Figure 6. Interaction type distributions from hybrid oxico-anoxic *in silico* experiments for two organism pairs: *E. coli* with *B. subtilis* (a), and *E. coli* with *S. enterica* (b).

344 interactions observed were unidirectional, with *E. coli* providing costless metabolites to
 345 its partner (Figure 6). When *E. coli* was grown with oxygen, its anoxic partner then
 346 provided the majority of metabolites that were exchanged. These intermediate hybrid
 347 simulations thus serve as a type of stepping stone, in which an organism grown anoxically
 348 can provide a higher number of useful byproducts to its aerobic partner, leading to
 349 bidirectional interactions when both are grown without oxygen.

350

351 **Interaction motifs form a basis for synthetic community assembly.** Lastly, we wished
 352 to use data generated by our algorithm to understand how multiple simultaneous
 353 interactions between two organisms could combine into network patterns (motifs) with
 354 different chance of appearance in a community and different dynamical stability
 355 properties. In particular, we sought to understand how the competition for common
 356 nutrients and the rise of costless exchange could jointly affect the stability of microbial
 357 consortia in resource-poor environments. These criteria could also serve as an atlas for
 358 guiding the engineering of stable synthetic consortia built off of costless metabolic
 359 relationships. As a first step in this analysis, we enumerated possible interaction network
 360 motifs based on our three interaction types and competition statuses (Figure 7a). These
 361 motifs encompassed all the possible permutations of interactions we identified in our
 362 dataset, accounting for non-interaction, commensalism, and mutualism with or without

363 competition. For non-interacting motifs, our simulations predicted an almost exclusive
364 representation of relationships involving competition for a primary carbon source (Figure
365 7b). The distribution between competitive and non-competitive types motifs was more
366 balanced for commensal and mutualistic interactions, showing a slight preference for
367 interactions involving competition.

368
369 In order to simulate how these interactions could contribute to stable symbioses, we
370 created a dynamical chemostat model of two arbitrary species consuming carbon sources
371 and exchanging costless metabolites according to each motif type (see Methods). By
372 varying the specific growth rates of each species from 0 to 1 hr⁻¹, we simulated the growth
373 of the pair under each motif type for 500 hours. If both species were still present at the
374 end of the simulation, we determined the motif type to enable stability at that combination
375 of specific growth rates. We mapped the space of stable species pairs under each motif
376 type, observing that competitive interactions generally have a reduced parameter space
377 for enabling stability (Figure 7c). Notably, though motif N1b was highly prevalent in the
378 costless FBA simulation set, this motif represents classic competitive exclusion and
379 cannot result in long-term stability. In contrast, though complete nutrient-organism
380 orthogonality can yield stability over the whole space of parameters (N2a), this motif was
381 not predicted to occur in the mechanistic simulations. An intermediate case between
382 these two extremes (N2b) is the one in which there is a balance between competition and
383 independence with respect to external carbon source utilization: in this case, which
384 frequently occurs in our dataset, stability is achievable only for a narrow set of parameters.

385
386 A marked increase in stability is predicted when costless metabolite exchange is enabled
387 (commensalism and mutualism). For motif C2b, for example, both organisms are
388 competing for a carbon source and organism 1 is providing one or more costless
389 metabolites to organism 2. Our dynamical modeling showed that the growth rate of
390 organism 1 must be greater than that of organism 2 in order for both species to be stable.
391 When feedback was allowed to occur (mutualism), the potential for stability vastly
392 increases across our parameter space. M2a and M2b even allowed for very low specific

393 growth rates for both organisms, indicating a strong dependence on costless metabolites
 394 for long-term coexistence.

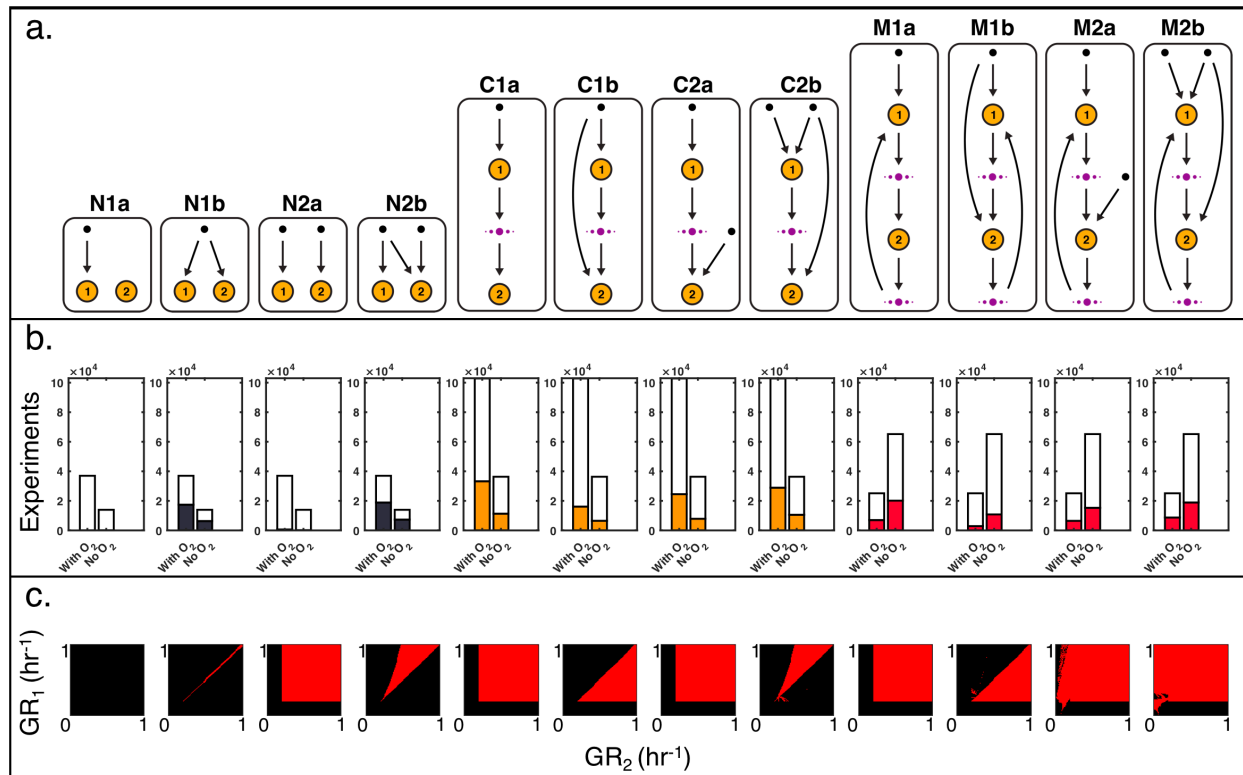


Figure 7. Interaction motif analysis and dynamical modeling of motif stability. (a) Schematic representation of specific motif types. Motifs are named according to three features: the interaction type (non-interacting, N; commensal, C; mutualistic, M), the number of carbon sources consumed by the pair (1-2), and competition for a primary carbon source (no competition, a; competition, b). Orange circles denote organisms, black dots denote primary carbon sources, and violet dots indicate any arbitrary number of costlessly-shared metabolites. Arrows indicate direction of metabolite flow. (b) Frequency of specific motif types. Height of empty white bars indicate the total number of simulations that exhibited the general motif type (Non-interacting, commensal, mutualistic). Colored bars within indicate the number of the specific motif type (N1a, N1b, etc.). (c) Stability space of motifs from dynamical chemostat modeling, as a function of the specific growth rates of the two organisms involved (GR_1 , GR_2). Red indicates area of stable coculture.

395

396 DISCUSSION

397 We have investigated the pairwise growth phenotypes and interactions of 14 diverse
 398 microbial species in over 10^6 computational experiments. We found that resource-poor
 399 environments provide the basis for release of a wide variety of useful metabolic products
 400 secreted without cost by their producing organism; these costless metabolic products
 401 provide, in an oxygen-dependent manner, valuable environmental enrichment, nearly

402 doubling the potential of minimal environments to sustain growth. We further found that
403 exchange of costless metabolites establishes beneficial uni- and bidirectional
404 interspecies interactions, associated with different chance of stability of the ensuing
405 consortia. Overall, both the metabolic capabilities of the organisms and the environmental
406 contexts in which they are grown (particularly oxygen availability) determine which
407 metabolites will be secreted without cost and how these secretions will contribute to
408 interspecies interactions.

409
410 Our modeling pipeline represents a novel *in silico* representation of distinct organisms
411 growing in environments progressively enriched by their partner's secreted byproducts.
412 This iterative medium expansion method provides a useful lens into the emergence of
413 higher-order interactions in microbial communities, allowing us to observe which
414 metabolites are secreted in response to others in a mechanistic fashion. We highlight the
415 utility of applying metabolic modeling to this area, particularly considering the
416 experimental inaccessibility of measuring metabolic secretions, interactions, and stability
417 across all the species and environmental conditions we tested. We observe that, despite
418 allowing organisms to secrete new metabolic products in response to changing medium
419 conditions, the amount and types of metabolites secreted is not enough to sustain
420 prolonged expansion iterations in most cases. This medium expansion distribution hints
421 at an upper limit to higher-order interactions mediated by costless metabolites in microbial
422 ecology.

423
424 We nonetheless emphasize that even in the simple, minimal environments we studied,
425 our modeling framework, based on fundamental stoichiometric constraints and metabolic
426 efficiency assumptions, predicts the widespread prevalence of molecular products that
427 are secreted without a metabolic burden and that can benefit other organisms. An
428 important implication of this prediction is that costless metabolites may significantly
429 contribute to enriching environments and sustaining biodiversity, even when organisms
430 are competing for the same primary nutrients. By using costless secretions to cooperate
431 while simultaneously competing for primary nutrients, organisms may escape some of the

432 limitations of pure competition, which has been predicted to limit biodiversity ⁴⁵. This
433 inference could help understanding microbial metabolic dynamics in many different
434 environments, ranging from structured soil communities to large oligotrophic microbial
435 communities, such as those found in the open ocean. This type of exchange, similar to
436 metabolic leakage behind the Black Queen Hypothesis ^{17,20}, may contribute to the
437 maintenance of small genomes in resource-poor environments, as the metabolic needs
438 of some organisms can be fulfilled by others. We look specifically at the obligate
439 partnerships predicted by our analysis, which mirror previously-studied codependencies
440 ^{9,40,41}. While our algorithm explored only pairs of organisms in coculture, one may wonder
441 whether more complex communities would display qualitatively different features. Our
442 analysis indeed suggests that higher order communities can support growth of highly
443 auxotrophic organisms such as *L. lactis*, *P. gingivalis*, and *R. sphaeroides*: in our pairwise
444 combinations, these organisms did not obtain enough byproducts from any single partner;
445 however, most of the metabolites that these organisms require to grow on a minimal
446 medium were producible separately by multiple species.

447
448 Our interaction analysis also provides deeper mechanistic insight into the increased
449 prevalence of mutualistic interactions without oxygen, a phenomenon that has been
450 previously predicted computationally ⁴⁶ and that provides a window into metabolic
451 relationships in environments harboring steep oxygen gradients, such as the human gut
452 ⁴⁷. By carrying out a set of hybrid oxic-anoxic *in silico* experiments, we observed that the
453 additional metabolites secreted anoxically by a facultative anaerobe (e.g. fermentation
454 byproducts) could provide extensive food supply for aerobically growing organisms. This
455 phenomenon has been suggested to play an important role in maintaining equilibrium in
456 communities at oxic-anoxic interfaces in the mammalian gut ^{48,49} and could be the subject
457 of further mechanistic studies.

458
459 Although our modeling method considers a wide space of mechanistic constraints in
460 predicting costless metabolic exchange, we acknowledge that secretion patterns and
461 exchange potential are also defined by a variety of other biological factors that fall outside

462 the scope of constraint-based modeling⁵⁰, such as signaling-based decisions, regulatory
463 states, and thermodynamic gradients induced by metabolite concentrations. Thus, our
464 analysis, in addition to demonstrating the plausibility of widespread costless cross-
465 feeding, could serve as the basis for prioritization of future specific experiments, for which
466 model predictions could be thought of as a null hypothesis against which to compare
467 empirical measurements. Moreover, though our analysis may accurately predict some
468 instances of metabolism-driven synergistic interactions, there may exist experimental
469 barriers (e.g. temperature or pH incompatibilities) to co-culturing some of the organisms
470 in our list, which are not captured in our modeling method. Nonetheless, our mechanistic
471 modeling framework may be applied to finding candidate species-environment pairs that
472 yield mutualistic relationships. Dynamical modeling coupled with these metabolic
473 analyses could then be used to obtain the parameter space most likely to yield desired
474 stable partnerships *in vivo*. Because this approach relies on screening environments that
475 can yield synergy as opposed to engineering individual strains, this approach has the
476 potential to simplify the process of assembling novel synthetic communities⁵¹. Our
477 analysis is also easily scalable to a large number of organisms and environments, and
478 could help produce a global atlas of expected, environment-dependent costless
479 secretions and their potential roles in mediating ecological interactions, with applications
480 in understanding and engineering microbiomes.

481

482 **METHODS**

483 **Selection and modification of genome-scale metabolic models.** A genome-scale
484 metabolic reconstruction was obtained for each of the 14 facultative anaerobic organisms
485 used in the analysis^{52–65}. Genome-scale metabolic models are mathematical
486 representations of an organism's known metabolic network, which are used to generate
487 mechanistic predictions of growth and resource allocation in a variety of environmental
488 conditions. The process of generating a genome-scale metabolic model has been
489 outlined conceptually^{66–69} and described procedurally⁷⁰ by various groups, and generally
490 comprises an automatic generation of a model based on pathway and genome data
491 followed by manual curation by integrating phenotyping, metabolomic, or transcriptomic

492 data ⁷¹. We note that although an automatically-generated draft metabolic model can be
493 constructed for virtually any organism for which a genome annotation exists, the space of
494 high-quality, experimentally-verified metabolic models that have undergone the manual
495 curation process summarized above is comparatively very small ⁷². This is due to the time
496 and resources needed to complete the curation process, which can span from six months
497 ⁷⁰ to more than ten years for the iteratively-refined model of *E. coli* K-12 ⁶⁴. We
498 nonetheless consider this process to be essential in producing models that can generate
499 the mechanistic cross-feeding predictions detailed here, which rely on verified metabolic
500 capabilities in monoculture.

501

502 The models used in this analysis span four taxonomic kingdoms, including
503 representatives from eight bacterial taxa, as well as a variety of primary metabolic
504 strategies (Supplementary Information 1). In addition, these models describe several
505 organisms that are commonly used for *in vivo* studies (*E. coli* K-12, *S. enterica* LT2, etc.),
506 making the resulting costless cross-feeding predictions particularly useful for synthetic
507 ecology experiments and microbial community assembly.

508

509 Each model was imported into MATLAB (The MathWorks, Inc., Natick, Massachusetts)
510 using the COntstraint-Based Reconstruction and Analysis (COBRA) Toolbox ⁷³, a
511 software platform for constraint-based modeling of metabolic networks. In order to enable
512 *in silico* cross-feeding to be correctly classified, the namespace of all of the metabolic
513 compounds in each of the models was standardized to be internally consistent. This was
514 performed via a computational pipeline with additional manual curation for irregularly-
515 annotated metabolites.

516

517 **Computational methodology description and inputs.** Our computational method
518 comprises a set of programs written in MATLAB that use Flux Balance Analysis (FBA) to
519 mechanistically define the growth status and metabolic exchange of microbes through
520 costlessly-secreted byproducts. Briefly, FBA is a mathematical method that determines
521 an optimal distribution of metabolic flux through a biochemical network that will maximize

522 a given objective, usually biomass⁷⁴. An FBA problem is framed in the context of several
523 constraints, namely: (i) S , the stoichiometric matrix of dimensions $m \times n$ where m is the
524 number of metabolites and n is the number of reactions in the model; (ii) v , the vector of
525 all reaction fluxes; and (iii) v_{min} and v_{max} , flux constraints placed on v , defined by
526 enzymatic capacity and experimentally measured uptake rates.

527

528 We employ FBA to determine if an organism is able to grow on the *in silico* growth media
529 conditions we define, in addition to which metabolites are taken up and costlessly
530 secreted. We first apply FBA by maximizing for growth and obtaining an optimal growth
531 rate for an organism, $v_{growth}^{(max)}$. To determine which metabolites are secreted costlessly,
532 we set this growth rate as a minimum for the biomass flux and apply FBA again, recording
533 any metabolites that were secreted. We also apply the additional constraint of minimizing
534 all reaction fluxes across the network to more closely simulate efficient use of the
535 proteome and minimize cycling of metabolites through the network⁷⁵. Our linear program
536 therefore becomes:

537

538

$$\min |v|,$$

539

$$\text{s.t.:$$

540

$$S \cdot v = 0,$$

541

$$v_{min} \leq v \leq v_{max},$$

542

$$v_{growth} \geq v_{growth}^{(max)}.$$

543

544 This optimization aims to encompass any enzymatic cost incurred by the organism in
545 synthesizing and exporting any metabolite we deem to be ‘costless.’ During each step in
546 which growth or metabolite absorption and secretion are computed, FBA optimizations
547 are performed separately for each *in silico* organism i and j , with biomass production set
548 as the objective function while minimizing the sum of the absolute value of v . Because
549 we focus on the emergence of potential metabolic exchange through the availability of
550 costlessly-secreted metabolites, our modeling framework purposefully keeps FBA

551 optimizations separate for each model without accounting for spatial or temporal
552 community structure. It is also for this reason that we establish the biomass fluxes of each
553 *in silico* organism as the objective functions to be optimized, as we are concerned with
554 secretion of potentially useful metabolic byproducts that arise out of “selfish” optimal
555 growth. This assumption of maximum growth with proteome optimality is also key for
556 translating these organisms and predictions to *in vivo* synthetic ecologies, where biomass
557 optimization more closely describes the behavior of organisms in batch or continuous
558 culture ⁷⁶.

559
560 Our algorithm requires six inputs: 1: a data structure containing the genome-scale
561 metabolic models to be used, 2: a list of carbon sources, 3: the number N_M of *in silico*
562 organisms to be simulated together (for pairwise simulations $N_M = 2$), 4: the number N_{CS}
563 of carbon sources to be provided to each simulation, 5: a Boolean variable $\Omega = \{1,0\}$ that
564 specifies if oxygen will be made available to the *in silico* organisms, and 6: a list of
565 metabolites that makes up a simulated base growth medium, M_{min} . This base medium
566 contains various nitrogen, sulfur, and phosphorus sources, as well as vitamins, ions, and
567 metals needed for growth of the organisms (Supplementary Information 3).

568
569 We focused on pairwise species growth with two carbon sources ($N_M, N_{CS} = 2$). Although
570 each genome-scale metabolic model we used has been manually curated to reflect *in*
571 *vivo* metabolic capabilities, very few experiments have been performed to verify FBA-
572 generated predictions for more than a single species ^{77,78}. We therefore limit the number
573 of *in silico* species to two, in order to interpret the growth and cross-feeding predictions
574 with greater confidence. This limit also constrains the combinatorial space of the
575 simulations, which grows exponentially and becomes numerically intractable with more
576 models and carbon sources. In addition, limiting simulations to $N_M = 2$ allows for greater
577 experimental accessibility for assembling synthetic ecologies based on costless
578 metabolite exchange. Our algorithm can nonetheless be applied to any $\{N_M, N_{CS} > 0\}$.

579

580 The list of all possible carbon sources was defined primarily from the carbon sources
581 contained in the BIOLOG Phenotyping MicroArray 1 (PM1) plate, which is used for
582 phenotyping and curation of genome-scale metabolic models^{79–81}. The carbon sources
583 we selected are common mono- di- and polysaccharides, all 20 amino acids, dipeptides,
584 and organic acids contained in the PM1 plate. We also supplemented the list with
585 additional carbon sources known to be consumed by the *in silico* organisms, for a total of
586 108 (Supplementary Information 2).

587

588 To permit uptake of the metabolites in the medium, the constraint on the uptake flux bound
589 v_{max} for each exchange reaction pertaining to a medium metabolite was removed in each
590 of the models i and j . This bound was fully removed ($v_{max} = 1000 \text{ mmol/gDW} * \text{hr}$) for
591 non-limiting medium components, and was set to $v_{max} = 10 \text{ mmol/gDW} * \text{hr}$ for the
592 growth-limiting carbon sources α and β . This latter value is drawn from experimentally-
593 estimated uptake rates of sugars by *E. coli* in exponential growth conditions⁶⁴, and is
594 applied equally to all other species to simulate general availability of the carbon sources
595 in the environment. All other exchange reaction v_{max} values are set to zero to block
596 uptake of metabolites not in the medium.

597

598 **Computing growth, secretion, and cross-feeding.** We describe the FBA operations at
599 the core of our algorithm as a function F that, given a medium condition M and organisms
600 i and j , outputs the binary growth status g of the organisms, as well as the set of
601 metabolites σ secreted costlessly by the organisms:

602

$$603 \quad F(\{M, i, j\}) = \{g, \sigma\}$$

604

605 Each *in silico* experiment E for a given organism pair with a pair of carbon sources is
606 made up of an initialization step, an expansion step consisting of series of applications of
607 F , and a completion step (Figure S2). In the initialization step, two organisms i and j are
608 selected, and a medium M_0 is defined. M_0 contains the minimal medium M_{min} , two carbon
609 sources α and β , and the variable Ω , which denotes the presence or absence of oxygen.

610

611 In the expansion step, the function F is applied for a series of iterations c . In each
612 iteration, F simulates the growth of both organisms in the current medium condition and
613 returns the Boolean growth statuses $g_c = \{g_i, g_j\}$ (where $g_i, g_j = \{0,1\}$) of both organisms
614 and the set of any costlessly-secreted metabolites, σ_c . To avoid recording metabolites
615 reported to be secreted only as a result of numerical uncertainty in FBA, a minimal lower
616 flux bound of $0.01 \text{ mmol/gDW} * \text{hr}$ was applied as a cutoff for determining secretion. If
617 at least one organism in the pair grows, the medium is supplemented with σ_c :

618

$$619 \quad M_{c+1} = M_c + \sigma_c.$$

620

621 As long as new metabolites continue to be secreted into the medium, that is,

622

$$623 \quad M_c > M_{c-1},$$

624

625 F continues to be applied. This stepwise expansion simulates the organisms responding
626 to the costlessly-secreted metabolites being secreted and generating a richer medium.
627 The completion step occurs when no new metabolites are secreted,

628

$$629 \quad M_c == M_{c-1}$$

630

631 and the final iteration before this stabilization occurs is defined as c_s . Our algorithm
632 therefore carries out individual *in silico* experiments $E_{i,j}^{\alpha,\beta,\Omega}$, defined as the output resulting
633 from c_s applications of F given organisms i and j , carbon sources α and β , and the
634 presence or absence Ω of oxygen:

635

$$636 \quad E_{i,j}^{\alpha,\beta,\Omega} \equiv \{g_c, M_c\}_{c=1}^{c_s} = F(\{M_0, i, j\})_{c=1}^{c_s}.$$

637

638 **Dynamical modeling of interaction motifs.** We designed a dynamical modeling method

639 to simulate the long-term stability of each pairwise interaction type observed in our *in*
640 *silico* experiments. We first established a graph theory framework to map each simulation
641 to a specific interaction motif, each of which accounted for the general interaction type
642 (non-interacting, commensal, or mutualistic), the number of carbon sources consumed by
643 the pair, and the competition status for the carbon sources (“a” denotes no competition,
644 “b” denotes competition) (Figure 5a). We next applied a differential equation-based
645 growth model to each specific motif. Since motifs with two carbon sources can be
646 represented by more than one motif topology, we selected one representative topology
647 from these motifs to simplify the space of dynamical modeling simulations. These
648 equations were modeled off Monod dynamics⁸² and are intended to simulate growth of
649 species in a chemostat, with constant replenishment of medium components. The
650 abundance of each organism s_i , in g/L, is modeled as follows:

651

$$\frac{ds_i}{dt} = s_i \mu_{max,i} \left(\frac{m_\alpha}{k_{s_i,m_\alpha} + m_\alpha} \right) - D s_i \quad (\text{E2})$$

652

653 where $\mu_{max,i}$ is the specific growth rate of organism i in h^{-1} , m_α is the concentration of
654 carbon source α in g/L, k_{s_i,m_α} is the concentration of α at which organism i reaches half
655 its maximal growth rate in g/L, and D is the chemostat dilution rate in h^{-1} . If two carbon
656 sources are present and the organism is determined to take up both by the motif definition,
657 the equation is modified to include a carbon source β as follows:

658

$$\frac{ds_i}{dt} = s_i \mu_{max,i} \left(\frac{m_\alpha}{k_{s_i,m_\alpha} + m_\alpha} \right) \left(\frac{m_\beta}{k_{s_i,m_\beta} + m_\beta} \right) - D s_i$$

660

661 The concentrations of each carbon source are defined as follows:

662

$$\frac{dm_\alpha}{dt} = I_{m_\alpha} - \frac{s_i}{K_{m_\alpha}} \mu_{max,i} \left(\frac{m_\alpha}{k_{s_i,m_\alpha} + m_\alpha} \right) - D m_\alpha \quad (\text{E3})$$

663

664 where I_{m_α} is the nutrient stock concentration for m_α in g/L, and K_{m_α} is the ratio of
665 nutrient consumed by the organism i in g_{nutrient}/g_{cells}. This equation is modified with an
666 additional term (organism j) to simulate competition for m_α .

667

668 To simulate metabolic exchange, equations for the abundances of costlessly-produced
669 metabolites (\tilde{m}) in g/L were defined as follows:

670

$$\frac{d\tilde{m}_i}{dt} = k_{\tilde{m}_i} * s_i - \frac{s_j}{K_{\tilde{m}_i, s_j}} \mu_{max, j} \left(\frac{\tilde{m}_i}{k_{s_j, \tilde{m}_i} + \tilde{m}_i} \right) - D\tilde{m}_i \quad (\text{E4})$$

671

672 Here, metabolite \tilde{m}_i is produced by organism i and consumed by organism j . $k_{\tilde{m}_i}$ is the
673 synthesis rate of the metabolite in hr⁻¹, $K_{\tilde{m}_i, s_j}$ is the ratio of metabolite consumed by the
674 population s_j in g_{metabolite}/g_{cells}, and k_{s_j, \tilde{m}_i} is the concentration of metabolite needed for
675 the population s_j to reach half of its maximum growth rate in g/L.

676

677 We then combine equations E2-4 to fit the particular motif being modeled (Figure S8).
678 The values of the parameter values are described in Supplementary Information 4 and
679 are based on values reported by Smith⁸³, Balagaddé *et al.*⁸⁴, and those based on
680 reasonable estimates for resource consumption. For each motif, we vary the specific
681 growth rate of both organisms from 0 to 1 hr⁻¹ and run the simulation for 500 hours. If
682 both organism abundances are above 0.05 g/L at the end of the simulation, we
683 determine the motif to be stable at the prescribed growth rates.

684

685 **ACKNOWLEDGEMENTS**

686 We thank Dr. Niels Klitgord for pioneering ideas that inspired launch of this work. We are
687 also grateful to David Bernstein, Joshua E. Goldford, Meghan Thommes, Demetrius
688 DiMucci, and all members of the Segrè Lab for helpful discussions. This work was
689 supported by funding from the Defense Advanced Research Projects Agency (Purchase
690 Request No. HR0011515303, Contract No. HR0011-15-C-0091), the U.S. Department of

691 Energy (Grants DE-SC0004962 and DE-SC0012627), the NIH (Grants 5R01DE024468,
692 R01GM121950 and Sub_P30DK036836_P&F), the National Science Foundation (Grants
693 1457695 and NSFOCE-BSF 1635070), MURI Grant W911NF-12-1-0390, the Human
694 Frontiers Science Program (grant RGP0020/2016), and the Boston University Inter-
695 disciplinary Biomedical Research Office. A.R.P. is supported by a National Academies of
696 Sciences, Engineering, and Medicine Ford Foundation Predoctoral Fellowship and a
697 Howard Hughes Medical Institute Gilliam Fellowship.

698

699 **CONTRIBUTIONS**

700 A.R.P. and D.S. designed the research. A.R.P. designed the computational framework,
701 carried out all simulations, and conducted data analysis. M.M. contributed to the
702 generation of standardized genome-scale models. A.R.P. and D.S. wrote the manuscript.
703 All authors read and approved the final manuscript.

704

705 **COMPETING FINANCIAL INTERESTS**

706 The authors declare no competing financial interests.

707

708 **CORRESPONDING AUTHORS**

709 Correspondence to: dsegre@bu.edu.

710

711 **REFERENCES**

- 712 1. Welch, D. B. M. & Huse, S. M. Microbial Diversity in the Deep Sea and the
713 Underexplored 'Rare Biosphere'. *Handb. Mol. Microb. Ecol. II Metagenomics*
714 *Differ. Habitats* 243–252 (2011). doi:10.1002/9781118010549.ch24
- 715 2. Tecon, R. & Or, D. Biophysical processes supporting the diversity of microbial life
716 in soil. *FEMS Microbiol. Rev.* **41**, 599–623 (2017).
- 717 3. Qin, J. *et al.* A human gut microbial gene catalogue established by metagenomic
718 sequencing. *Nature* **464**, 59–65 (2010).
- 719 4. Hardin, G. The competitive exclusion principle. *Science* **131**, 1292–7 (1960).
- 720 5. Hutchinson, G. E. The paradox of the plankton. *Am. Nat.* **91**, (1961).
- 721 6. Wilson, M. & Lindow, S. E. Coexistence among Epiphytic Bacterial Populations
722 Mediated through Nutritional Resource Partitioning. *Appl. Environ. Microbiol.* **60**,

- 723 4468–77 (1994).
- 724 7. Inouye, R. S. & Tilman, D. Convergence and Divergence of Old-Field Plant
725 Communities Along Experimental Nitrogen Gradients. *Ecology* **69**, 995–1004
726 (1988).
- 727 8. Kim, H. J., Boedicker, J. Q., Choi, J. W. & Ismagilov, R. F. Defined spatial
728 structure stabilizes a synthetic multispecies bacterial community. *Proc. Natl.
729 Acad. Sci. U. S. A.* **105**, 18188–93 (2008).
- 730 9. Morris, B. E. L., Henneberger, R., Huber, H. & Moissl-Eichinger, C. Microbial
731 syntrophy: Interaction for the common good. *FEMS Microbiol. Rev.* **37**, 384–406
732 (2013).
- 733 10. Fildes, P. Production of Tryptophan by *Salmonella typhi* and *Escherichia coli*. *J.
734 Gen. Microbiol.* **15**, 636–642 (1956).
- 735 11. Goldford, J. E. *et al.* Emergent Simplicity in Microbial Community Assembly.
736 *bioRxiv* 205831 (2017). doi:10.1101/205831
- 737 12. Ponomarova, O. & Patil, K. R. Metabolic interactions in microbial communities:
738 untangling the Gordian knot. *Curr. Opin. Microbiol.* **27**, 37–44 (2015).
- 739 13. Harcombe, W. Novel cooperation experimentally evolved between species.
740 *Evolution (N. Y.)*. **64**, 2166–72 (2010).
- 741 14. Wintermute, E. H. & Silver, P. A. Emergent cooperation in microbial metabolism.
742 *Mol. Syst. Biol.* **6**, 1–7 (2010).
- 743 15. Stolyar, S. *et al.* Metabolic modeling of a mutualistic microbial community. *Mol.
744 Syst. Biol.* **3**, 92 (2007).
- 745 16. Vacca, I. Bacterial ecology: Cheaters take advantage. *Nat. Rev. Microbiol.* **15**,
746 575–575 (2017).
- 747 17. Morris, J. J., Lenski, R. E. & Zinser, E. R. The Black Queen Hypothesis : Evolution
748 of Dependencies through Adaptative Gene Loss. *MBio* **3**, 1–7 (2012).
- 749 18. Germerodt, S. *et al.* Pervasive Selection for Cooperative Cross-Feeding in
750 Bacterial Communities. *PLoS Comput. Biol.* **12**, 1–21 (2016).
- 751 19. Hoek, M. J. A. va. & Merks, R. M. H. Emergence of microbial diversity due to
752 cross-feeding interactions in a spatial model of gut microbial metabolism. *BMC
753 Syst. Biol.* **11**, 1–18 (2017).
- 754 20. Zomorodi, A. R. & Segrè, D. Genome-driven evolutionary game theory helps
755 understand the rise of metabolic interdependencies in microbial communities. *Nat.
756 Commun.* **8**, 1563 (2017).
- 757 21. Sachs, J. L., Mueller, U. G., Wilcox, T. P. & Bull, J. J. The evolution of
758 cooperation. *Q. Rev. Biol.* **79**, 135–60 (2004).
- 759 22. West-Eberhard, M. J. The evolution of social behavior by kin selection. *Q. Rev.
760 Biol.* **50**, 1–33 (1975).
- 761 23. Connor, R. C. The Benefits of Mutualism: A Conceptual Framework. *Biol. Rev.* **70**,

- 762 427–457 (1995).
- 763 24. Brown, J. L. Cooperation—A Biologist’s Dilemma. *Adv. Study Behav.* **13**, 1–37
764 (1983).
- 765 25. Orth, J. D., Thiele, I. & Palsson, B. O. What is flux balance analysis? *Nat Biotech*
766 **28**, 245–248 (2010).
- 767 26. Tiso, M. & Schechter, A. N. Nitrate reduction to nitrite, nitric oxide and ammonia
768 by gut bacteria under physiological conditions. *PLoS One* **10**, e0119712 (2015).
- 769 27. Zelezniak, A. *et al.* Metabolic dependencies drive species co-occurrence in
770 diverse microbial communities. *Proc. Natl. Acad. Sci.* **112**, 6449–6454 (2015).
- 771 28. Paczia, N. *et al.* Extensive exometabolome analysis reveals extended overflow
772 metabolism in various microorganisms. *Microb. Cell Fact.* **11**, 1–14 (2012).
- 773 29. Swenson, T. L., Karaoz, U., Swenson, J. M., Bowen, B. P. & Northen, T. R.
774 Linking soil biology and chemistry in biological soil crust using isolate
775 exometabolomics. *Nat. Commun.* **9**, (2018).
- 776 30. Embree, M., Liu, J. K., Al-Bassam, M. M. & Zengler, K. Networks of energetic and
777 metabolic interactions define dynamics in microbial communities. *Proc. Natl.*
778 *Acad. Sci.* **112**, 15450–15455 (2015).
- 779 31. Velasco, I., Tenreiro, S., Calderon, I. L. & André, B. *Saccharomyces cerevisiae*
780 *Aqr1* is an internal-membrane transporter involved in excretion of amino acids.
781 *Eukaryot. Cell* **3**, 1492–503 (2004).
- 782 32. Dassler, T., Maier, T., Winterhalter, C. & Bock, A. Identification of a major
783 facilitator protein from *Escherichia coli* involved in efflux of metabolites of the
784 cysteine pathway. *Mol. Microbiol.* **36**, 1101–1112 (2000).
- 785 33. Airich, L. G. *et al.* Membrane topology analysis of the *Escherichia coli* aromatic
786 amino acid efflux protein YddG. *J. Mol. Microbiol. Biotechnol.* **19**, 189–97 (2010).
- 787 34. Ponomarova, O. *et al.* Yeast Creates a Niche for Symbiotic Lactic Acid Bacteria
788 through Nitrogen Overflow. *Cell Syst.* **5**, 345–357.e6 (2017).
- 789 35. Stadie, J., Gulitz, A., Ehrmann, M. A. & Vogel, R. F. Metabolic activity and
790 symbiotic interactions of lactic acid bacteria and yeasts isolated from water kefir.
791 *Food Microbiol.* **35**, 92–98 (2013).
- 792 36. Williams, R. J., Howe, A. & Hofmockel, K. S. Demonstrating microbial co-
793 occurrence pattern analyses within and between ecosystems. *Front. Microbiol.* **5**,
794 1–10 (2014).
- 795 37. HilleRisLambers, J., Adler, P. B., Harpole, W. S., Levine, J. M. & Mayfield, M. M.
796 Rethinking Community Assembly through the Lens of Coexistence Theory. *Annu.*
797 *Rev. Ecol. Evol. Syst.* **43**, 227–248 (2012).
- 798 38. Tilman, D. *Resource competition and community structure*. (Princeton University
799 Press, 1982).
- 800 39. Faust, K. *et al.* Microbial co-occurrence relationships in the Human Microbiome.

- 801 *PLoS Comput. Biol.* **8**, (2012).
- 802 40. Lindell, D. & Post, A. F. Ecological Aspects of *ntcA* Gene Expression and Its Use
803 as an Indicator of the Nitrogen Status of Marine *Synechococcus* spp. *Appl.*
804 *Environ. Microbiol.* **67**, 3340–3349 (2001).
- 805 41. Glibert, P. M. & Ray, R. T. Different patterns of growth and nitrogen uptake in two
806 clones of marine *Synechococcus* spp. *Mar. Biol.* **107**, 273–280 (1990).
- 807 42. Flores, E. & Herrero, A. in *The Molecular Biology of Cyanobacteria* 487–517
808 (Springer Netherlands, 1994). doi:10.1007/978-94-011-0227-8_16
- 809 43. Segrè, D., DeLuna, A., Church, G. M. & Kishony, R. Modular epistasis in yeast
810 metabolism. *Nat. Genet.* **37**, 77–83 (2005).
- 811 44. Foster, K. R. & Bell, T. Competition, not cooperation, dominates interactions
812 among culturable microbial species. *Curr. Biol.* **22**, 1845–1850 (2012).
- 813 45. Ashby, B., Watkins, E., Lourenço, J., Gupta, S. & Foster, K. R. Competing
814 species leave many potential niches unfilled. *Nat. Ecol. Evol.* **1**, 1495–1501
815 (2017).
- 816 46. Heinken, A. & Thiele, I. Anoxic Conditions Promote Species-Specific Mutualism
817 between Gut Microbes In Silico. *Appl. Environ. Microbiol.* **81**, 4049–61 (2015).
- 818 47. Espey, M. G. Role of oxygen gradients in shaping redox relationships between
819 the human intestine and its microbiota. *Free Radic. Biol. Med.* **55**, 130–140
820 (2013).
- 821 48. Donaldson, G. P., Lee, S. M. & Mazmanian, S. K. Gut biogeography of the
822 bacterial microbiota. *Nat. Rev. Microbiol.* **14**, 20–32 (2015).
- 823 49. He, G. *et al.* Noninvasive measurement of anatomic structure and intraluminal
824 oxygenation in the gastrointestinal tract of living mice with spatial and spectral
825 EPR imaging. *Proc. Natl. Acad. Sci. U. S. A.* **96**, 4586–91 (1999).
- 826 50. Bordbar, A., Monk, J. M., King, Z. A. & Palsson, B. O. Constraint-based models
827 predict metabolic and associated cellular functions. *Nat. Rev. Genet.* **15**, 107–120
828 (2014).
- 829 51. Lindemann, S. R. *et al.* Engineering microbial consortia for controllable outputs.
830 *ISME J.* **10**, 2077–2084 (2016).
- 831 52. Mazumdar, V., Snitkin, E. S., Amar, S. & Segrè, D. Metabolic network model of a
832 human oral pathogen. *J. Bacteriol.* **91**, 74–90 (2009).
- 833 53. Motamedian, E., Saeidi, M. & Shojaosadati, S. A. Reconstruction of a charge
834 balanced genome-scale metabolic model to study the energy-uncoupled growth of
835 *Zymomonas mobilis* ZM1. *Mol. BioSyst.* **12**, 1241–1249 (2016).
- 836 54. Peyraud, R. *et al.* Genome-scale reconstruction and system level investigation of
837 the metabolic network of *Methylobacterium extorquens* AM1. *BMC Syst. Biol.* **5**,
838 (2011).
- 839 55. Imam, S. *et al.* IRsp1095: A genome-scale reconstruction of the Rhodobacter

- 840 sphaeroides metabolic network. *BMC Syst. Biol.* **5**, 116 (2011).
- 841 56. Flahaut, N. A. L. *et al.* Genome-scale metabolic model for *Lactococcus lactis*
842 MG1363 and its application to the analysis of flavor formation. *Appl. Microbiol.*
843 *Biotechnol.* **97**, 8729–8739 (2013).
- 844 57. Pinchuk, G. E. *et al.* Constraint-Based Model of *Shewanella oneidensis* MR-1
845 Metabolism: A Tool for Data Analysis and Hypothesis Generation. *PLoS Comput.*
846 *Biol.* **6**, e1000822 (2010).
- 847 58. Liao, Y. C. *et al.* An experimentally validated genome-scale metabolic
848 reconstruction of *Klebsiella pneumoniae* MGH 78578, iYL1228. *J. Bacteriol.* **193**,
849 1710–1717 (2011).
- 850 59. Nogales, J., Gudmundsson, S., Knight, E. M., Palsson, B. O. & Thiele, I. Detailing
851 the optimality of photosynthesis in cyanobacteria through systems biology
852 analysis. *Proc. Natl. Acad. Sci.* **109**, 2678–2683 (2012).
- 853 60. Monk, J. M. *et al.* Genome-scale metabolic reconstructions of multiple *Escherichia*
854 *coli* strains highlight strain-specific adaptations to nutritional environments. *Proc.*
855 *Natl. Acad. Sci.* **110**, 20338–20343 (2013).
- 856 61. Zomorodi, A. R. & Maranas, C. D. Improving the iMM904 *S. cerevisiae* metabolic
857 model using essentiality and synthetic lethality data. *BMC Syst. Biol.* **4**, 178
858 (2010).
- 859 62. Thiele, I. *et al.* A community effort towards a knowledge-base and mathematical
860 model of the human pathogen *Salmonella Typhimurium* LT2. *BMC Syst. Biol.* **5**, 8
861 (2011).
- 862 63. Oberhardt, M. A., Puchałka, J., Fryer, K. E., Martins dos Santos, V. A. P. & Papin,
863 J. A. Genome-scale metabolic network analysis of the opportunistic pathogen
864 *Pseudomonas aeruginosa* PAO1. *J. Bacteriol.* **190**, 2790–803 (2008).
- 865 64. Orth, J. D. *et al.* A comprehensive genome-scale reconstruction of *Escherichia*
866 *coli* metabolism--2011. *Mol. Syst. Biol.* **7**, 535 (2011).
- 867 65. Henry, C. S., Zinner, J. F., Cohoon, M. P. & Stevens, R. L. iBsu1103: a new
868 genome-scale metabolic model of *Bacillus subtilis* based on SEED annotations.
869 *Genome Biol.* **10**, R69 (2009).
- 870 66. Reed, J. L., Famili, I., Thiele, I. & Palsson, B. O. Towards multidimensional
871 genome annotation. *Nat. Rev. Genet.* **7**, 130–141 (2006).
- 872 67. Feist, A. M., Herrgård, M. J., Thiele, I., Reed, J. L. & Palsson, B. Ø.
873 Reconstruction of biochemical networks in microorganisms. *Nat. Rev. Microbiol.*
874 **7**, 129–143 (2008).
- 875 68. Price, N. D., Papin, J. A., Schilling, C. H. & Palsson, B. O. Genome-scale
876 microbial in silico models: the constraints-based approach. *Trends Biotechnol.* **21**,
877 162–169 (2003).
- 878 69. Durot, M., Bourguignon, P.-Y. & Schachter, V. Genome-scale models of bacterial

- 879 metabolism: reconstruction and applications. *FEMS Microbiol. Rev.* **33**, 164–190
880 (2009).
- 881 70. Thiele, I. & Palsson, B. O. A protocol for generating a high-quality genome-scale
882 metabolic reconstruction. *Nat. Protoc.* **5**, (2010).
- 883 71. Oh, Y.-K., Palsson, B. O., Park, S. M., Schilling, C. H. & Mahadevan, R. Genome-
884 scale reconstruction of metabolic network in *Bacillus subtilis* based on high-
885 throughput phenotyping and gene essentiality data. *J. Biol. Chem.* **282**, 28791–9
886 (2007).
- 887 72. King, Z. A. *et al.* BiGG Models: A platform for integrating, standardizing and
888 sharing genome-scale models. *Nucleic Acids Res.* **44**, D515–D522 (2016).
- 889 73. Becker, S. A. *et al.* Quantitative prediction of cellular metabolism with constraint-
890 based models: the COBRA Toolbox. *Nat Protoc* **2**, (2007).
- 891 74. Orth, J. D., Thiele, I. & Palsson, B. Ø. What is flux balance analysis? *Nat.*
892 *Biotechnol.* **28**, 245–248 (2010).
- 893 75. Holzhütter, H.-G. The principle of flux minimization and its application to estimate
894 stationary fluxes in metabolic networks. *Eur. J. Biochem.* **271**, 2905–2922 (2004).
- 895 76. Lewis, N. E. *et al.* Omic data from evolved *E. coli* are consistent with computed
896 optimal growth from genome-scale models. *Mol. Syst. Biol.* **6**, 390 (2010).
- 897 77. Harcombe, W. R. *et al.* Metabolic resource allocation in individual microbes
898 determines ecosystem interactions and spatial dynamics. *Cell Rep.* **7**, 1104–1115
899 (2014).
- 900 78. Zomorodi, A. R. & Segrè, D. Synthetic Ecology of Microbes: Mathematical
901 Models and Applications. *J. Mol. Biol.* (2015). doi:10.1016/j.jmb.2015.10.019
- 902 79. Reed, J. L. *et al.* Systems approach to refining genome annotation. *Proc. Natl.*
903 *Acad. Sci.* **103**, 17480–17484 (2006).
- 904 80. Oberhardt, M. A., Puchałka, J., Martins dos Santos, V. A. P. & Papin, J. A.
905 Reconciliation of Genome-Scale Metabolic Reconstructions for Comparative
906 Systems Analysis. *PLoS Comput. Biol.* **7**, e1001116 (2011).
- 907 81. Henry, C. S. *et al.* High-throughput generation, optimization and analysis of
908 genome-scale metabolic models. *Nat. Biotechnol.* **28**, 977–982 (2010).
- 909 82. Monod, J. The Growth of Bacterial Cultures. *Annu. Rev. Microbiol.* **3**, 371–394
910 (1949).
- 911 83. Smith, H. L. Bacterial growth. (2006).
- 912 84. Balagaddé, F. K. *et al.* A synthetic *Escherichia coli* predator-prey ecosystem. *Mol.*
913 *Syst. Biol.* **4**, 187 (2008).

SUPPLEMENTARY FIGURES

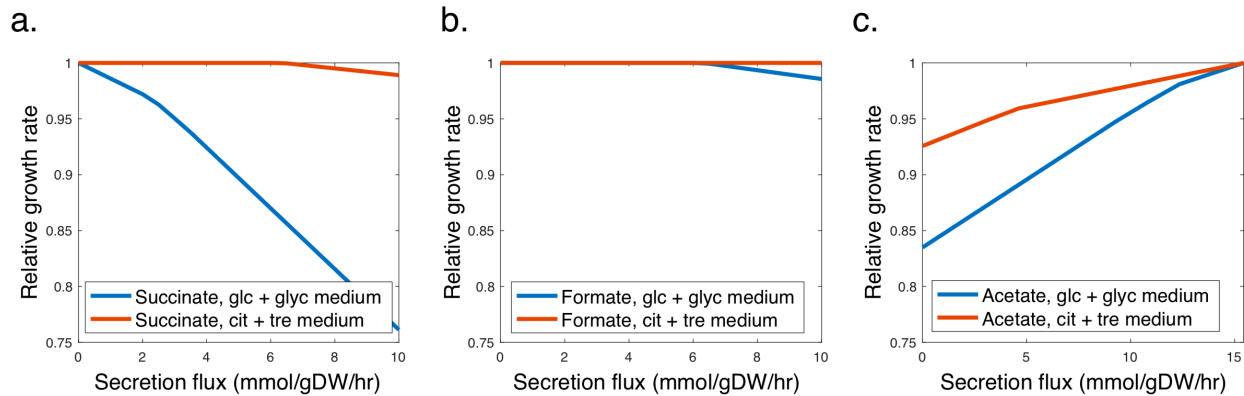


Figure S1. Three modes of *in silico* metabolite secretion by *E. coli* (iJO1366) in anoxic conditions as defined by FBA. What makes a metabolite costless is dependent on the environment. (a) Increasing the secretion flux of a 'costly' product, such as succinate, imposes a reduction in growth rate when glucose and glycerol are supplied as carbon sources. When the carbon sources are replaced with citrate and trehalose, succinate is secreted without a cost to growth rate. **(b)** With glucose and glycerol as carbon sources, *E. coli* is predicted to have a wide range of fluxes at which formate can be secreted without a cost to its growth rate. Formate would, according to our definition, be secreted 'costlessly' by *E. coli* under the applied environmental conditions. **(c)** Some costlessly-secreted metabolites must be secreted at a given rate in order to maximize growth. If an upper bound is placed on acetate secretion, *E. coli* must allocate resources away from biomass in order to cope with its limited ability to secrete fermentation byproducts. Acetate would therefore also be considered a costlessly-secreted metabolite by our definition.

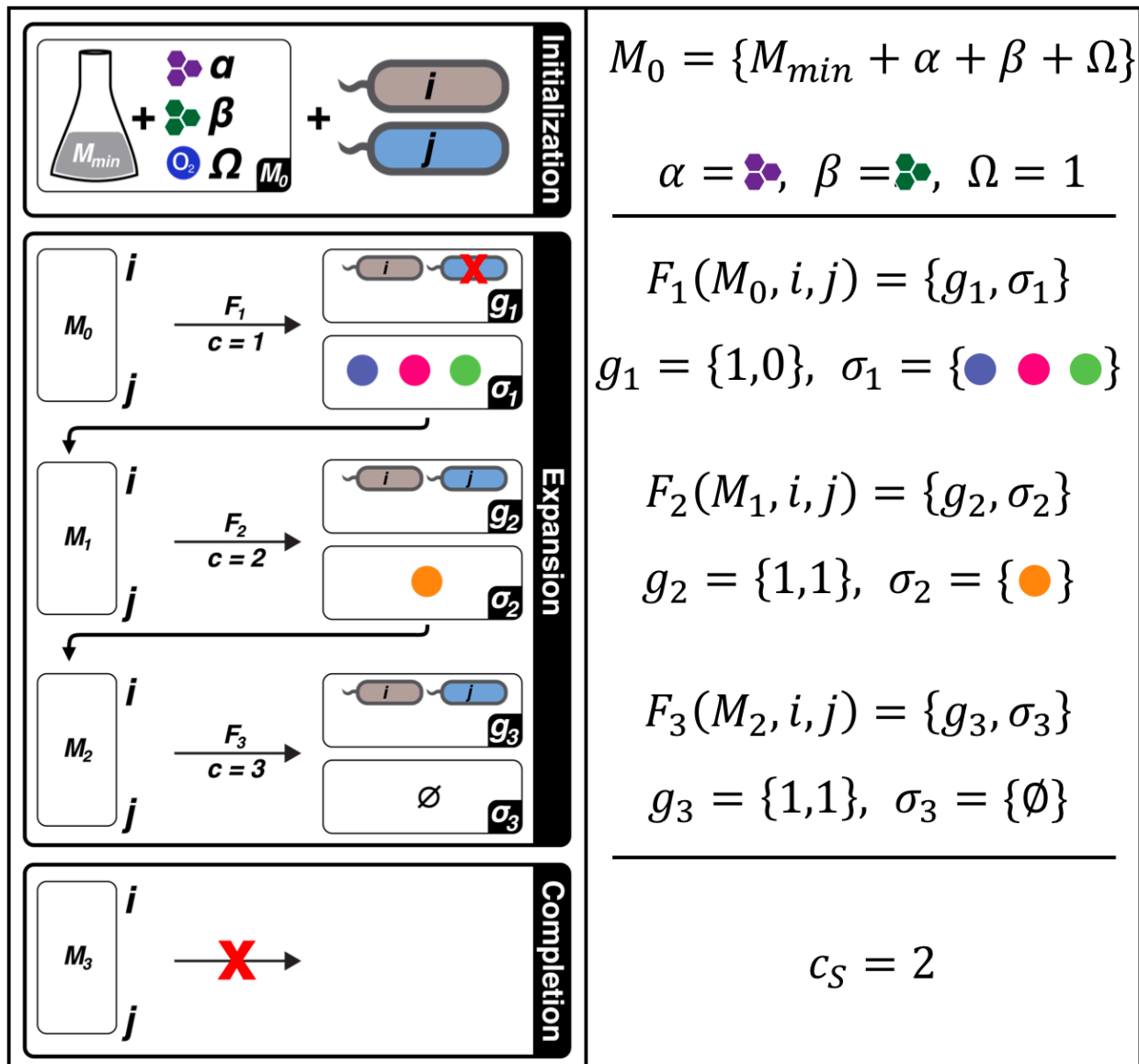


Figure S2. Detailed example of single *in silico* experiment, illustrating three phases. Initialization: A minimal medium M_{min} common to all simulated conditions (composed of salts, metals, vitamins, as well as nitrogen, phosphorous, and sulphur sources) is defined prior to execution of the pipeline. This medium is supplemented with two carbon sources, α and β . The Boolean variable $\Omega = \{0,1\}$ defines whether or not oxygen is present in the environment. Here, $\Omega = 1$. These together define the initial medium set, M_0 . **Expansion:** The function F is applied to genome-scale metabolic models of two organisms (i, j) in a series of iterations, c . In each iteration, F simulates the growth of both organisms in the current medium condition and returns the Boolean growth statuses $g_c = \{g_i, g_j\}$ of both organisms and the set of any costlessly-secreted metabolites, σ_c . Here, in the first iteration, $g_1 = \{1, 0\}$ since organism i grew but organism j did not. Since at least one organism in the pair grew, the medium is updated ($M_{c+1} = M_c + \sigma_c$) and F is applied again until no new metabolites are secreted. **Completion:** When no new metabolites are added to the medium, the experiment is complete. The last iteration with any new secreted metabolites is defined as c_S .

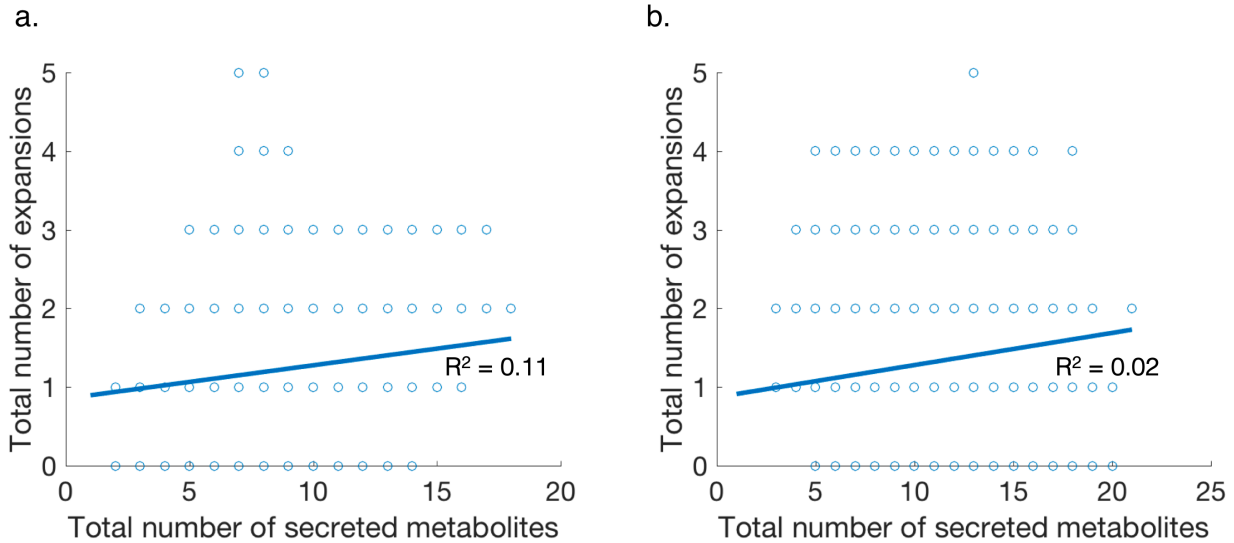


Figure S3. Correlation between total number of metabolites secreted costlessly and the number of expansions in each *in silico* experiment for (a) oxic and (b) anoxic conditions. We observe a poor correlation between number of secreted metabolites and number of expansions in both oxic and anoxic simulations. This lack of correlation suggests a lower rate of metabolite exchange with increasing iterations, with most organisms quickly stabilizing their environment within one or two expansions. With oxygen, for example, only the *K. pneumoniae* and *Synechocystis* pair exhibited more than three medium expansions, with acetate, formate, citrate, and L-malate being the only metabolites secreted at these iterations. These scenarios accounted for only 40 simulations. Without oxygen, there were 697 experiments that reached more than three medium expansions, with 10 organisms being represented. However, this anaerobic set was dominated by the *S. cerevisiae*-*P. aeruginosa* pair, with fermentation byproducts being secreted at late iterations.

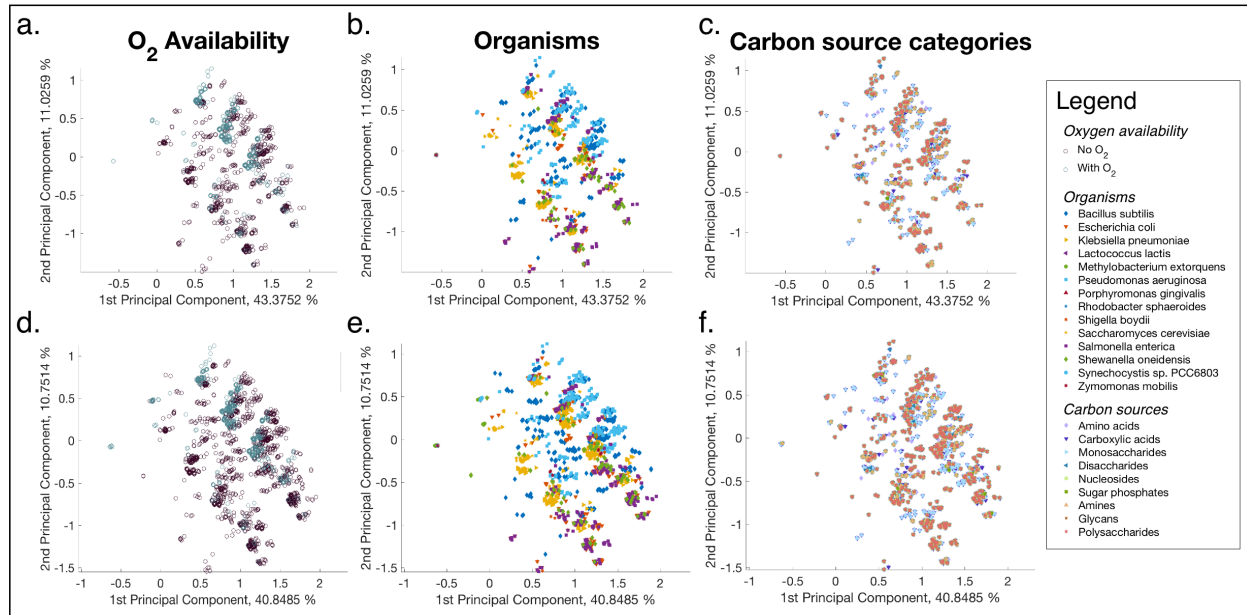


Figure S4. Principal component analysis (PCA) plots for metabolite secretion profiles. To address whether there is one chief contributing factor to patterns of costless metabolite secretion, we carried out principal component analysis (PCA), a dimensionality reduction technique. Each point represents the secreted metabolites of a single organism in one *in silico* experiment. **(a-c)** PCA plots for metabolites secreted before medium expansions ($c = 1$). **(d-f)** PCA plots for secreted metabolites after all medium expansions. Points are clustered by oxygen availability (a, d), organisms (b, e), and carbon source category (c, f).

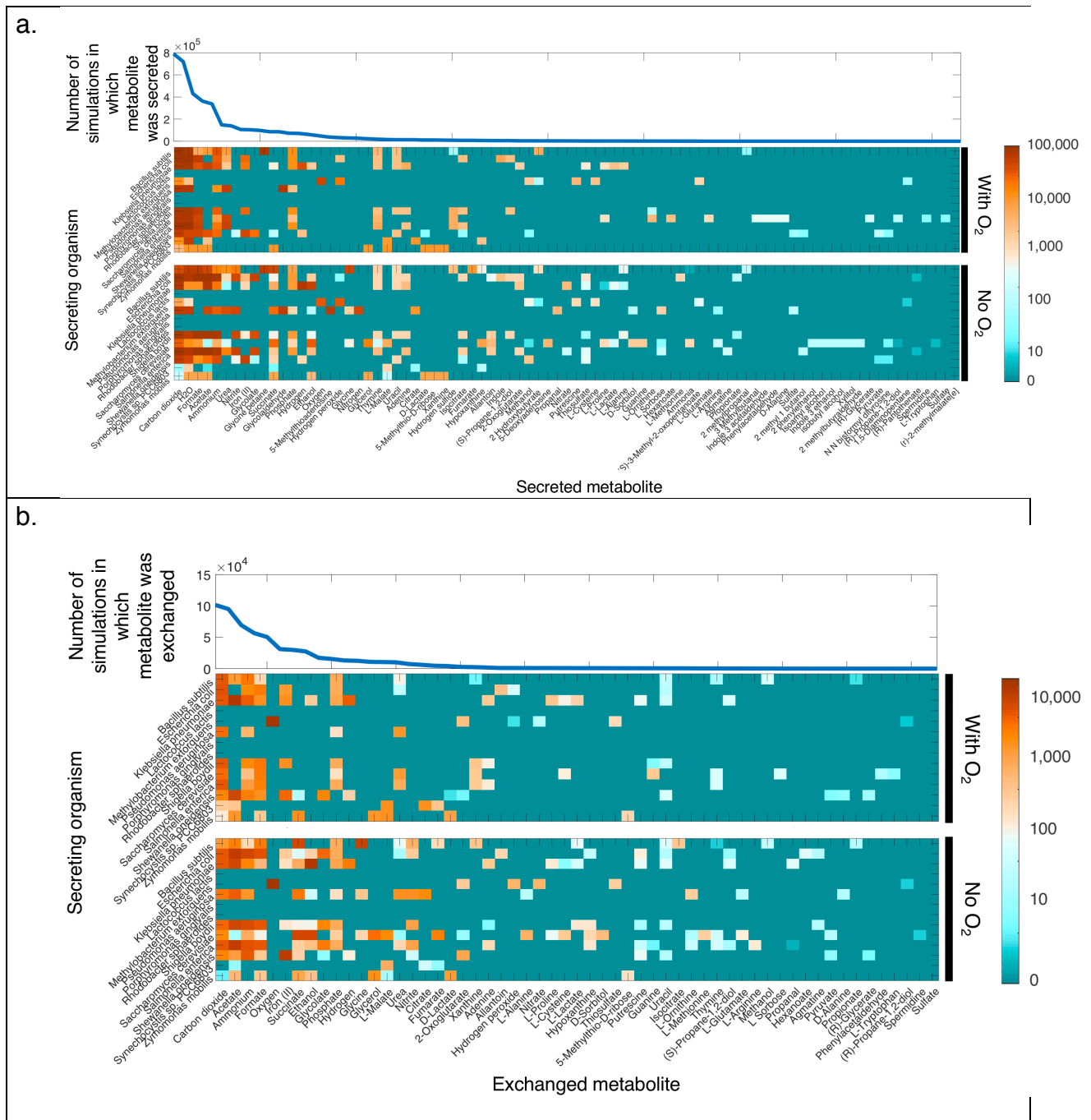


Figure S5. Range of costlessly-secreted and exchanged metabolites. (a) Cumulative sum of *in silico* experiments in which metabolite was secreted (top), and sorted heatmap of metabolites secreted in at least one simulation, arranged by secreting organism (bottom). **(b)** Cumulative sum of *in silico* experiments in which each secreted metabolite was taken up by another organism (top), and sorted heatmap of metabolites secreted and taken up in at least one simulation, arranged by secreting organism (bottom).

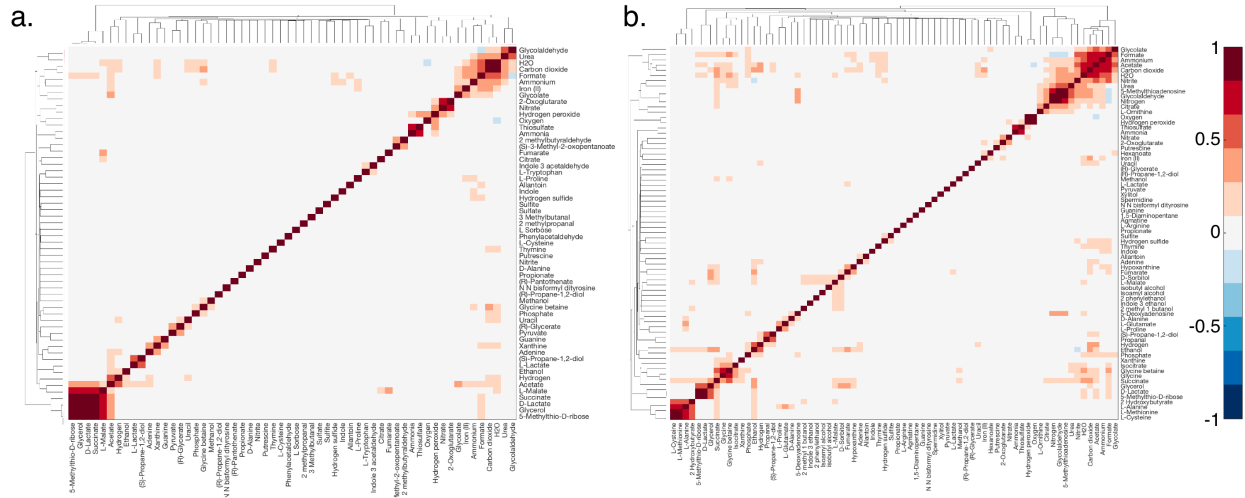


Figure S6. Clustered Spearman correlation of secreted metabolites for (a) oxic and (b) anoxic *in silico* experiments.

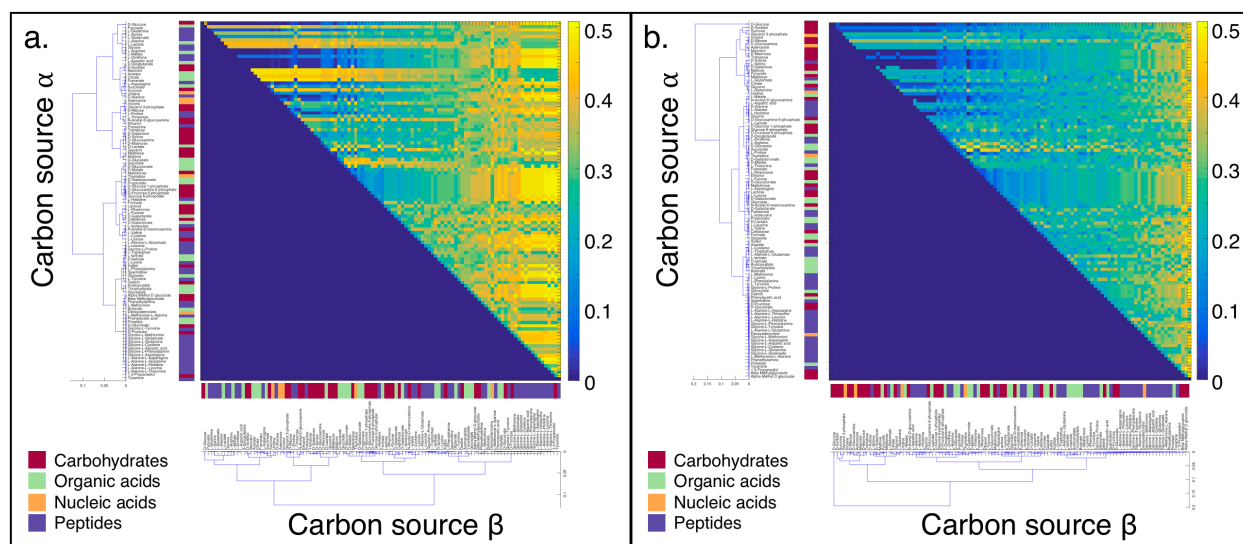


Figure S7. Cooperativity indices of all carbon source pairs in oxic (a) and anoxic (b) conditions, clustered by average carbon source cooperativity index.

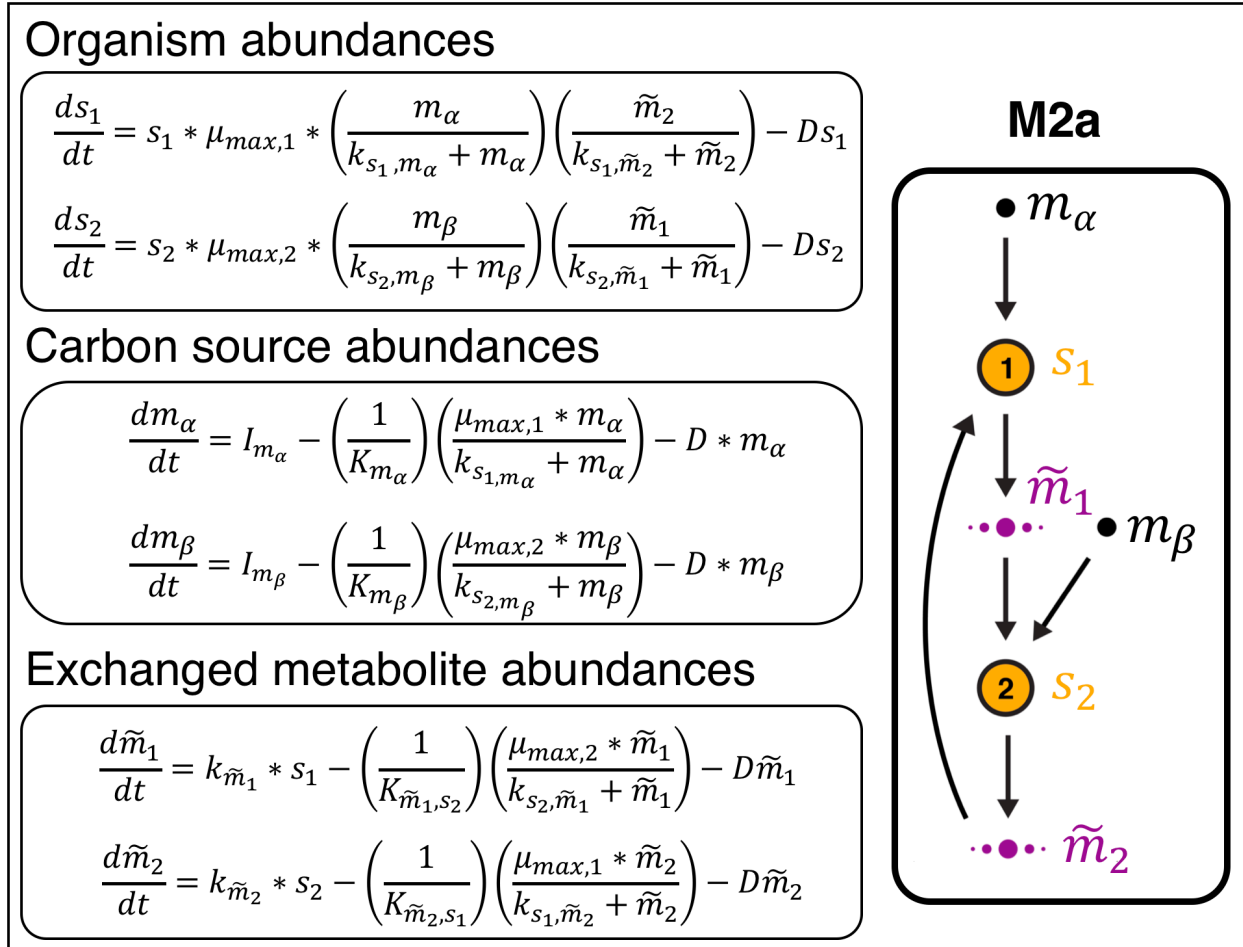


Figure S8. Example of dynamical modeling equations for motif M2a (two carbon sources consumed, no competition, mutualism).