1 bmotif: a package for counting motifs in bipartite networks

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22 Abstract

23 1. Bipartite networks are widely-used to represent a diverse range of species interactions, such as 24 pollination, herbivory, parasitism and seed dispersal. The structure of these networks is usually 25 characterised by calculating one or more metrics that capture different aspects of network architecture. While these metrics capture useful properties of networks, they only consider structure at the scale of 26 27 the whole network (the macro-scale) or individual species (the micro-scale). 'Meso-scale' structure between these scales is usually ignored, despite representing ecologically-important interactions. 28 29 Network motifs are a framework for capturing this meso-scale structure and are gaining in popularity. However, there is no software available in R, the most popular programming language among 30 31 ecologists, for conducting motif analyses in bipartite networks. Similarly, no mathematical 32 formalisation of bipartite motifs has been developed.

2. Here we introduce bmotif: a package for counting motifs, and species positions within motifs, in
bipartite networks. Our code is primarily an R package, but we also provide MATLAB and Python code
of the core functionality. The software is based on a mathematical framework where, for the first time,
we derive formal expressions for motif frequencies and the frequencies with which species occur in
different positions within motifs. This framework means that analyses with bmotif are fast, making
motif methods compatible with the permutational approaches often used in network studies, such as
null model analyses.

3. We describe the package and demonstrate how it can be used to conduct ecological analyses, using
two examples of plant-pollinator networks. We first use motifs to examine the assembly and
disassembly of an Arctic plant-pollinator community, and then use them to compare the roles of native
and introduced plant species in an unrestored site in Mauritius.

44 4. bmotif will enable motif analyses of a wide range of bipartite ecological networks, allowing future45 research to characterise these complex networks without discarding important meso-scale structural

- 46 detail.
- 47

48 Key-words: motifs, bipartite networks, R, MATLAB, Python, pollination, food web, seed dispersal

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50 Introduction

51 Bipartite networks are widely used to study the structure of interactions between two groups of species, such as plants and pollinators, hosts and parasitoids, and plants and seed dispersers (Borrett, Moody, & 52 53 Edelmann, 2014). Studies of bipartite networks have vielded many new insights (Bascompte & Jordano, 54 2007), such as uncovering widespread nestedness and modularity in mutualistic communities 55 (Bascompte, Jordano, Melián, & Olesen, 2003; Olesen, Bascompte, Dupont, & Jordano, 2007), and showing that community structure is stable despite substantial turnover in species and interactions over 56 space and time (Petanidou, Kallimanis, Tzanopoulos, Sgardelis, & Pantis, 2008; Dáttilo, Guimarães, & 57 58 Izzo, 2013). Such studies typically describe networks with one or more metrics, such as connectance 59 (the proportion of possible interactions which are realised), nestedness (the extent to which specialist 60 species interact with subsets of the species generalist species interact with), degree (number of partners a species has) and d' (the extent to which a species' interactions deviate from a random sampling of its 61 62 partners).

While these metrics describe useful properties of networks, macro-scale measures, such as connectanceand nestedness, can be too broad to capture fine-scale details, while micro-scale metrics, such as degree

as d', can be too narrow to capture a species' indirect interactions (Cirtwill, Roslin, Rasmussen, Olesen,

66 & Stouffer, 2018). Capturing network structure at the meso-scale between these macro and micro scales 67 is necessary to overcome these issues (Cirtwill et al., 2018). For example, a micro-scale metric such as 68 degree might show that a plant is visited by two pollinators, while meso-scale structure could reveal 69 that one of these pollinators is a generalist visiting three other generalist plants, while the other is a 70 specialist visiting only the focal plant. Such distinctions can have important consequences for 71 understanding the ecology and evolution of communities and so are essential to incorporate in network 72 analyses.

73 To capture meso-scale structure, ecologists are increasingly using bipartite motifs: subnetworks 74 representing interactions between a given number of species (Fig. 1). These subnetworks can be thought 75 of as the basic 'building blocks' of networks (Milo et al., 2002). Bipartite motifs are used in two main 76 ways. First, to calculate how frequently different motifs occur in a network. For example, Rodríguez-77 Rodríguez et al. (2017) used this approach to show that plant species involved in both mutualistic and 78 antagonistic interactions with animals were the most important for pollination. Second, to quantify 79 species roles in a community by counting the frequency with which species occur in different positions 80 within motifs. For example, Baker et al. (2015) used this method to demonstrate that species' roles in host-parasitoid networks are an intrinsic property of species. However, while the motif framework is 81 gaining in popularity, no software currently exists to conduct motif analyses of bipartite networks in R, 82 83 the most popular programming language among ecologists.

84 To fill this gap, we introduce bmotif: an R package, based on a formal mathematical framework, for counting motifs, and species positions within motifs, in bipartite networks. While bmotif is primarily 85 86 an R package, we additionally provide MATLAB and Python code that replicates the core package 87 functionality. Here, we introduce the motifs and motif positions counted by bmotif and describe the package's main functions and performance. We then provide two examples showing how bmotif can 88 89 be used to answer questions about ecological communities. While here we focus on mutualistic bipartite 90 networks, our methods are general and can also be applied to other types of interaction, such as parasitism and herbivory, and even non-biological systems, such as trade networks (Saracco, Di 91 92 Clemente, Gabrielli, & Squartini, 2015) and finance networks (Gualdi, Cimini, Primicerio, Di 93 Clemente, & Challet, 2016).

94 Description

95 **Defining bipartite motifs**

In a bipartite network containing N species, a motif is a subnetwork comprising n species and their interactions (where n < N and all species have at least one interaction). Fig. 1 shows the motifs included in bmotif: all 44 possible motifs containing up to six nodes. Within motifs, species can appear in different positions (Fig. 1). For reasons of symmetry, not all these positions are topologically unique (Stouffer, Sales-Pardo, Sirer, & Bascompte, 2012). For example, motif six contains four species, but only two positions (Fig. 1). The 148 unique positions a species can occupy across all motifs up to six nodes are shown in Fig. 1. Motifs and positions are ordered as in Baker et al. (2015) Appendix 1.

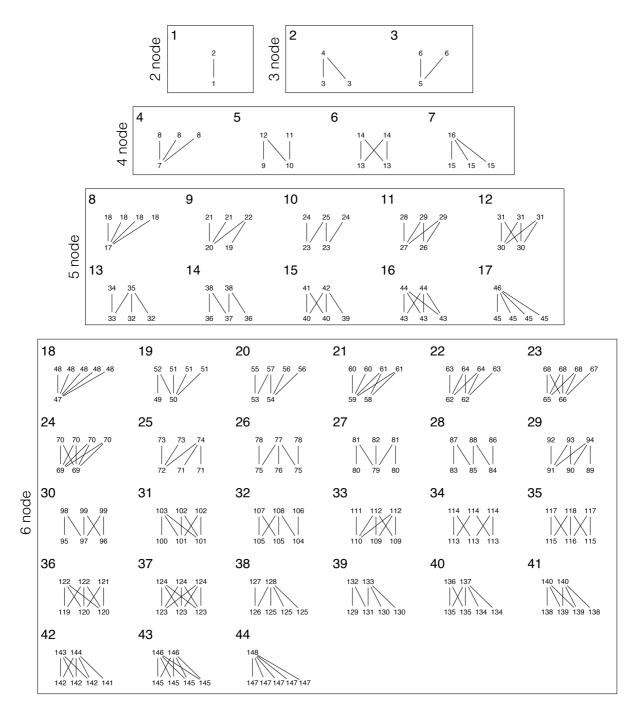




Figure 1: All bipartite motifs containing up to 6 nodes (species). Large numbers identify each motif. Small numbers represent the unique positions species can occupy within motifs, following Baker et al. (2015)
 Appendix 1. Lines between small numbers indicate undirected species interactions. There are 44 motifs containing 148 unique positions.

Networks in bmotif are represented as incidence matrices (**M**), with one row for each species in the first set (such as pollinators) and one column for each species in the second set (such as plants). When species *i* and *j* interact, $m_{ij} = 1$; if they do not interact $m_{ij} = 0$. This widely-used representation was chosen for compatibility with other packages (Dormann, Frund, Bluthgen, & Gruber, 2009) and openaccess network repositories, such as the Web of Life (www.web-of-life.es). Species in rows correspond to nodes in the top level of the motifs in Fig. 1; species in columns correspond to nodes in the bottom level.

117 Main functions

bmotif has two functions: (i) *mcount*, for calculating how frequently different motifs occur in a network,
 and (ii) *positions*, for calculating the frequency with which species (nodes) occur in different positions

120 within motifs to quantify a species' structural role. To enumerate motif frequencies and species position

121 counts, bmotif uses mathematical operations directly on the incidence matrix: for the first time, we

- derive 44 expressions for each of the 44 motifs and 148 expressions for each of the 148 positions within
- 123 motifs (Appendix S2).

124 mcount takes a network as input and returns a data frame with one row for each motif (17 or 44 rows depending on whether motifs up to five or six nodes are requested, respectively) and three columns. 125 126 The first column is the motif identity as in Fig. 1; the second column is the motif size class (number of 127 nodes each motif contains); and the third column is the frequency with which each motif occurs in the 128 network (a network's motif profile). For comparing multiple networks it is important to normalise motif frequencies. Therefore, if the 'normalise' argument is TRUE, three columns are added to the data frame, 129 each corresponding to a different method for normalising motif frequencies. The first column 130 ('normalise sum') expresses the frequency of each motif as a proportion of the total number of motifs 131 132 in the network. The second column ('normalise sizeclass') expresses the frequency of each motif as a proportion of the total number of motifs within its size class. The final column ('normalise nodesets') 133 expresses the frequency of each motif as the number of species combinations that occur in a motif as a 134 proportion of the number of species combinations that could occur in that motif. For example, in motifs 135 136 9, 10, 11 and 12, there are three species in the top set (A) and two species in the lower set (B) (Fig. 1). 137 Therefore, the maximum number of species combinations that could occur in these motifs is given by the product of binomial coefficients, choosing three species from A and two from P: $\binom{A}{3}\binom{B}{2}$ (Poisot & 138 Stouffer, 2016). The most appropriate normalisation depends on the question being asked. For example, 139 'normalise sum' allows for consideration of whether species are more involved in smaller or larger 140 141 motifs. Conversely, 'normalise sizeclass' focuses the analysis on how species form their interactions among different arrangements of *n* nodes. 142

143 positions takes a network as input and returns a data frame, W, with one row for each species and one 144 column for each position (46 or 148 columns, depending on whether motifs up to five or six nodes are requested, respectively; Fig. 1). w_{rc} gives the number of times species r occurs in position c. Each row 145 146 thus represents the structural role or 'interaction niche' of a species. The 'level' argument allows positions to be requested for all species, species in set A only or species in set B only, returning a data 147 frame with A + B rows, A rows or B rows, respectively. Two types of normalisation are provided: 'sum' 148 149 normalisation expresses a species' position frequencies as a proportion of the total number of times that species appears in any position; 'size class' normalisation uses the same approach, but normalises 150 frequencies within each motif size class. Again, the most appropriate normalisation depends on the 151 152 question being asked: if movements between motif size classes are of interest, 'sum' normalisation is most appropriate; if the focus is on how species form interactions among a given number of nodes, then 153 154 'size class' normalisation should be chosen.

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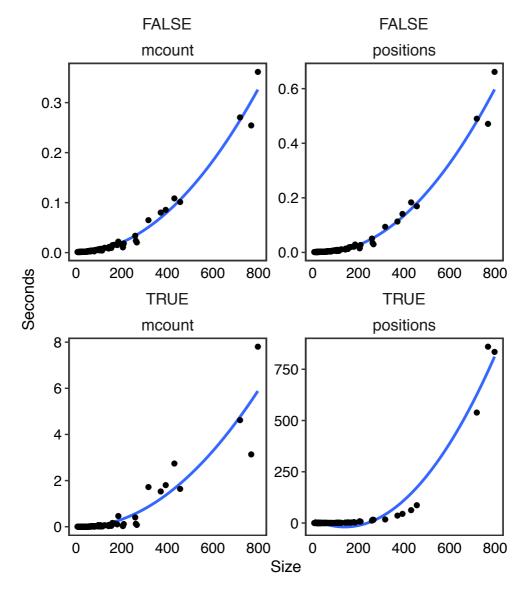
156 Computational performance

To assess the speed of bmotif functions, we used *mcount* and *positions* to calculate the complete motif profiles of 175 empirical pollination and seed dispersal networks and the positions of all their constituent species. While most of these networks use the frequency of animal visits to plants as a surrogate for true pollination or seed dispersal, this has been shown to be a reasonable proxy (Vázquez, Morris, & Jordano, 2005; Simmons et al., 2018). The networks varied in size from 6 to 797 species (mean: 77.1; standard deviation: 117.8). Further details of the networks used for this analysis are given

in Supplementary Data 1. Analyses were carried out on a computer with a 4.0 GHz processor and 32

164 GB of memory. Functions were timed using the R package 'microbenchmark' (Mersmann, 2015).

165 Results are shown in Fig. 2.



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Figure 2: Relationship between network size and computational performance for *mcount* and *positions*.
 Functions were timed on 175 empirical networks, for motifs containing up to five and six nodes. Lines are best fit polynomial curves of degree 2.

As expected, the time taken for a function to run increases monotonically with the size of the network 170 171 (number of species). When six-node motifs were excluded, *mcount* and *positions* took 0.36 and 0.66 172 seconds, respectively, to complete for the largest network in our dataset (797 species). For smaller networks which are more typical of the communities analysed by ecologists, both functions completed 173 in substantially less than one second. This speed is possible as all formulae involved in calculations of 174 175 motifs up to five-nodes use relatively simple operations, such as matrix multiplication or the binomial coefficient. When six-node motifs were included, for a network with 78 species (close to the mean 176 network size of 77.1 species), mcount completed in 0.01 seconds, while positions completed in 0.32 177 seconds. For the largest network, mcount completed in 7.8 seconds, while positions took 13.9 minutes. 178 Six-node motifs slow down calculations as, unlike five-node motifs, their algorithms require the use of 179 180 the tensor product.

We carried out additional analyses using randomly-generated networks to disentangle the effects of both network dimensions and connectance on computational performance (Appendix S1). We found that connectance had little effect on the performance of individual motif and position calculations, while a polynomial of degree two explained the increase in time with network size ($R^2 > 0.99$) (Appendix S1).

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187 Example analyses

188 Comparing community structures

Here we use bmotif to examine the assembly and disassembly of an Arctic plant-pollinator community. 189 190 Networks were sampled daily, when weather conditions allowed, at the Zackenberg Research Station in northeastern Greenland, across two full seasons in 1996 (24 days) and 1997 (26 days) (Olesen, 191 192 Bascompte, Elberling, & Jordano, 2008). Basic network properties are given in Supplementary Data 2. We used *mcount* to calculate motif frequencies in each daily network in both years, normalised using 193 'normalise nodesets'. Days 1 and 24 in 1996, and days 1 and 26 in 1997, were excluded from the 194 195 analysis as they were too small for some motifs to occur. Using nonmetric multidimensional scaling 196 (NMDS), we visualised how the community structure changed from assembly after the last snow melt 197 to disassembly at the first snow fall, in two consecutive years (Fig. 3). More positive values of the first 198 NMDS axis are associated with motifs where generalist pollinators compete for generalist plants, while 199 negative values are associated with motifs where more specialist pollinators have greater 200 complementarity in the specialist plants they visit. More positive values of the second NMDS axis are associated with loosely connected motifs containing specialist plants interacting with both specialist 201 and generalist pollinators, while negative values are associated with highly connected motifs containing 202 pollinators competing for generalist plants. While the community was relatively stable over time in the 203 1996 season, there were larger structural changes in 1997, with a largely monotonic shift from high 204 competition between generalist pollinators at the start of the season, to lower competition between more 205 specialist pollinators at the end of the season, with a more complementary division of plant resources 206 (Fig. 3). Thus while network structure may appear stable when analysed with traditional indices such 207 208 as connectance (Olesen et al., 2008), motifs reveal the presence of complex, ecologically-important 209 structural dynamics. Additionally, it is clear that, even in consecutive years, the community followed different structural trajectories, emphasising the danger of treating networks as static entities 210 (Rasmussen, Dupont, Mosbacher, Trøjelsgaard, & Olesen, 2013). 211

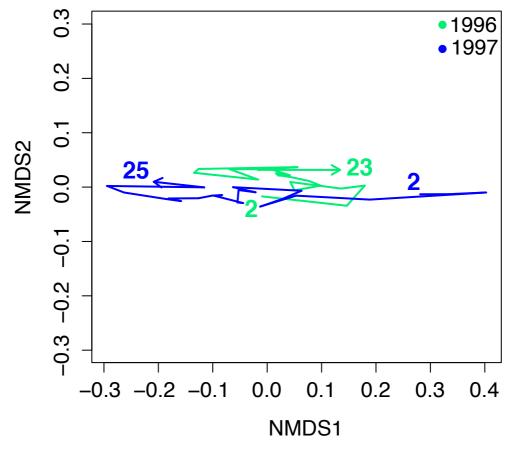


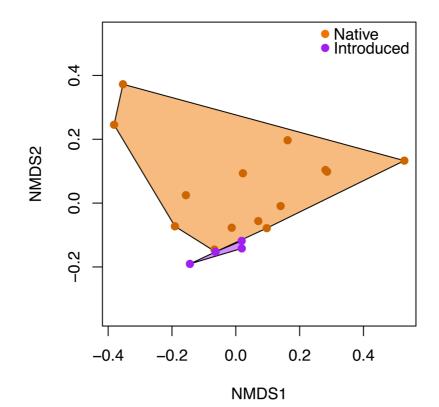
Figure 3: Nonmetric multidimensional scaling plot (NMDS) showing change in Arctic plant-pollinator network
 structure over the 1996 and 1997 seasons, quantified using motifs. Numbers represent the days of sampling.

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217 Comparing species' structural roles

We use *positions* to compare the roles of native and introduced plant species in a plant-pollinator 218 219 community sampled in Mauritius in November 2003 (Kaiser-Bunbury, Memmott & Müller 2009; 48 species, 75 interactions, connectance = 0.134). We calculated the sum-normalised roles of all plant 220 species (16 native and 4 introduced) and plotted them on two NMDS axes (Fig. 4). This figure shows 221 222 three striking features. First, there is almost no overlap between native and introduced species' 223 interaction niches. Similar to research showing that non-native species can occupy different functional niches to native species (Ordonez, Wright, & Olff, 2010), these results suggest they may also occupy 224 225 unexploited interaction niches. Further research could use motifs to investigate whether introduced species 'pushed' native species out of previously occupied interaction niche space, or whether 226 introduced species colonised previously-unused space. Second, the interaction niche of introduced 227 species is much smaller than that of native species: the four introduced species all occupy similar areas 228 229 of motif space, possibly suggesting a single 'invader role'. Third, introduced species occupy lower 230 values on the second NMDS axis, corresponding to motif positions where they are visited by generalist pollinator species, possibly due to the absence of co-evolutionary associations with specialists. NMDS 231 analyses were conducted with the metaMDS function in the R package vegan (Oksanen et al., 2016). 232



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Figure 4: The roles of native and introduced species in a plant-pollinator network. Each point represents the role
 of a species in the network. Shaded polygons are convex hulls either containing all introduced species or all
 alien species.

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²³⁸ Implementation and availability

The bmotif package is available for the R programming language. The package can be installed in R 239 using install.packages("bmotif"). This paper describes version 1.0.0 of the software. The source code 240 241 of the package is available at https://github.com/SimmonsBI/bmotif. Any problems can be reported using the Issues system. The code is version controlled with continuous integration and has code 242 243 coverage of approximately 98%. MATLAB and Python code replicating the core package functionality https://github.com/SimmonsBI/bmotif-matlab 244 available is at and https://github.com/SimmonsBI/bmotif-python respectively. All code is released under the MIT license. 245

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247 Conclusions

bmotif is an R package and set of mathematical formulae enabling motif analyses of bipartite networks.
Specifically, bmotif provides functions for two key analyses: (i) enumerating the frequency of different
motifs in a network, and (ii) calculating how often species occur in each position within motifs. These
two techniques capture important meso-scale variation in network structure that may be missed by
traditional methods. Motif approaches represent a new addition to the network ecologists 'toolbox' for
use alongside other techniques to analyse bipartite networks. We hope bmotif encourages further uptake
of the motif approach to shed light on the ecology and evolution of species and communities.

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264 Author contributions

BIS conceived the project, conducted analyses and wrote the first draft of the manuscript. BIS, MJM
and RDC developed the bmotif package. BIS, WJS, LVD and RDC planned the study. BIS and RDC
coordinated and designed the work. All authors contributed to writing the manuscript.

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269 Data accessibility

All networks used in this study are available from the Web of Life repository (<u>www.web-of-life.es</u>),

with the exception of the Greenland plant-pollinator networks which are available from Data Dryad(Saavedra et al., 2016)

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274 References

- Baker, N. J., Kaartinen, R., Roslin, T., & Stouffer, D. B. (2015). Species' roles in food webs show
 fidelity across a highly variable oak forest. *Ecography*, *38*(2), 130–139. doi:10.1111/ecog.00913
- Bascompte, J., & Jordano, P. (2007). Plant-Animal Mutualistic Networks: The Architecture of
 Biodiversity. *Annual Review of Ecology, Evolution, and Systematics*, *38*(1), 567–593.
 doi:10.1146/annurev.ecolsys.38.091206.095818
- Bascompte, J., Jordano, P., Melián, C. J., & Olesen, J. M. (2003). The nested assembly of plant- animal mutualistic networks. *Proceedings of the National Academy of Sciences*, 100(16), 9383–
 9387.
- Borrett, S. R., Moody, J., & Edelmann, A. (2014). The rise of Network Ecology: Maps of the topic
 diversity and scientific collaboration. *Ecological Modelling*, 293, 111–127.
 doi:10.1016/j.ecolmodel.2014.02.019
- Cirtwill, A. R., Roslin, T., Rasmussen, C., Olesen, J. M., & Stouffer, D. B. (2018). Between-year
 changes in community composition shape species' roles in an Arctic plant-pollinator network.
 Oikos.
- 289 Dáttilo, W., Guimarães, P. R., & Izzo, T. J. (2013). Spatial structure of ant-plant mutualistic networks.
 290 *Oikos*, *122*(11), 1643–1648. doi:10.1111/j.1600-0706.2013.00562.x
- Dormann, C. F., Frund, J., Bluthgen, N., & Gruber, B. (2009). Indices, Graphs and Null Models:
 Analyzing Bipartite Ecological Networks. *The Open Ecology Journal*, 2(1), 7–24.
 doi:10.2174/1874213000902010007
- Gualdi, S., Cimini, G., Primicerio, K., Di Clemente, R., & Challet, D. (2016). Statistically validated
 network of portfolio overlaps and systemic risk. *Scientific Reports*, 6. doi:10.1038/srep39467
- Kaiser-Bunbury, C. N., Memmott, J., & Müller, C. B. (2009). Community structure of pollination
 webs of Mauritian heathland habitats. *Perspectives in Plant Ecology, Evolution and Systematics*,

- 298 *11*(4), 241–254. doi:10.1016/j.ppees.2009.04.001
- 299 Mersmann, O. (2015). microbenchmark: Accurate Timing Functions. R package version 1.4-2.1.
- Milo, R., Shen-Orr, S., Itzkovitz, S., Kashtan, N., Chklovskii, D., & Alon, U. (2002). Network motifs:
 simple building blocks of complex networks. *Science*, *298*(5594), 824–827.
- 302 Oksanen, J., Blanchet, F. G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., ... Wagner, H.
 303 (2016). vegan: Community Ecology Package. R package version 2.4-0.
- Olesen, J. M., Bascompte, J., Dupont, Y. L., & Jordano, P. (2007). The modularity of pollination
 networks. *Proceedings of the National Academy of Sciences*, 104(50), 19891–19896.
- Olesen, J. M., Bascompte, J., Elberling, H., & Jordano, P. (2008). Temporal dynamics in a pollination
 network. *Ecology*, *89*(6), 1573–1582. doi:10.1890/07-0451.1
- Ordonez, A., Wright, I. J., & Olff, H. (2010). Functional differences between native and alien species:
 A global-scale comparison. *Functional Ecology*, 24(6), 1353–1361. doi:10.1111/j.1365 2435.2010.01739.x
- Petanidou, T., Kallimanis, A. S., Tzanopoulos, J., Sgardelis, S. P., & Pantis, J. D. (2008). Long-term
 observation of a pollination network: Fluctuation in species and interactions, relative invariance
 of network structure and implications for estimates of specialization. *Ecology Letters*, 11(6),
 564–575. doi:10.1111/j.1461-0248.2008.01170.x
- Poisot, T., & Stouffer, D. (2016). How ecological networks evolve. *bioRxiv*. Retrieved from
 http://biorxiv.org/content/early/2016/08/29/071993.abstract
- Rasmussen, C., Dupont, Y. L., Mosbacher, J. B., Trøjelsgaard, K., & Olesen, J. M. (2013). Strong
 Impact of Temporal Resolution on the Structure of an Ecological Network. *PLOS ONE*, 8(12),
 e81694. Retrieved from https://doi.org/10.1371/journal.pone.0081694
- Rodríguez-Rodríguez, M. C., Jordano, P., & Valido, A. (2017). Functional consequences of plant animal interactions along the mutualism-antagonism gradient. *Ecology*, *98*(5), 1266–1276.
 doi:10.1002/ecy.1756
- Saavedra, S., Rohr, R. P., Olesen, J. M., & Bascompte, J. (2016). Data from: Nested species
 interactions promote feasibility over stability during the assembly of a pollinator community.
 Ecology and Evolution. Dryad Digital Repository. doi:doi:10.5061/dryad.3pk73
- Saracco, F., Di Clemente, R., Gabrielli, A., & Squartini, T. (2015). Randomizing bipartite networks:
 the case of the World Trade Web. *Scientific Reports*, *5*, 10595. Retrieved from
 http://dx.doi.org/10.1038/srep10595
- Simmons, B. I., Sutherland, W. J., Dicks, L. V, Albrecht, J., Farwig, N., García, D., ... González Varo, J. P. (2018). Moving from frugivory to seed dispersal: incorporating the functional
 outcomes of interactions in plant-frugivore networks. *Journal of Animal Ecology*.
- Stouffer, D. B., Sales-Pardo, M., Sirer, M. I., & Bascompte, J. (2012). Evolutionary Conservation of
 Species' Roles in Food Webs. *Science*, *335*(6075), 1489–1492. doi:10.1126/science.1216556
- Vázquez, D. P., Morris, W. F., & Jordano, P. (2005). Interaction frequency as a surrogate for the total
 effect of animal mutualists on plants. *Ecology Letters*, 8(10), 1088–1094. doi:10.1111/j.1461 0248.2005.00810.x