A rapid and robust method for single cell chromatin accessibility profiling

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Abstract

The assay for transposase-accessible chromatin using sequencing (ATAC-seq) is widely used to identify regulatory regions throughout the genome. However, very few studies have been performed at the single cell level (scATAC-seq) due to technical challenges. Here we developed a simple and robust plate-based scATAC-seq method, combining upfront bulk Tn5 tagging with single-nuclei sorting. We demonstrated that our method worked robustly across various systems, including fresh and cryopreserved cells from primary tissues. By profiling over 3,000 splenocytes, we identify distinct immune cell types and reveal cell type-specific regulatory regions and related transcription factors.

Due to its simplicity and sensitivity, ATAC-seq[1] has been widely used to map open chromatin regions across different cell types in bulk. Recent technical developments have allowed chromatin accessibility profiling at the single cell level (scATAC-seq) and revealed distinct regulatory modules across different cell types within heterogeneous samples[2–9]. In these approaches, single cells are first captured by either a microfluidic device[3] or a liquid deposition system[7], followed by independent tagmentation of each cell. Alternatively, a combinatorial indexing strategy has been reported to perform the assay without single cell isolation[2,4,9]. However, these approaches require either a specially engineered and expensive device, such as a Fluidigm C1[3] or Takara ICELL8[7], or a large quantity of customly modified Tn5 transposase[2,4,5,9].

Here, we overcome these limitations by performing upfront Tn5 tagging in the bulk cell population, prior to single nuclei isolation. It has been previously demonstrated that Tn5 transposase-mediated tagmentation contains two stages: (1) a tagging stage where the Tn5 transposome binds to DNA, and (2) a fragmentation stage where the Tn5 transposase is released from DNA using heat or denaturing agents, such as sodium dodecyl sulfate (SDS)[10–12]. Since the Tn5 tagging does not fragment DNA, we reasoned that the nuclei would remain intact after incubation with the Tn5 transposome in an ATAC-seq experiment. Based on this idea, we developed a simple, robust and flexible plate-based scATAC-seq protocol, performing a Tn5 tagging reaction[6,13] on a pool of cells (5,000 - 50,000) followed by sorting individual nuclei into plates containing lysis buffer.

Tween-20 is subsequently added to quench the SDS in the lysis buffer[14], which otherwise will interfere the downstream reactions. Library indexing and amplification are done by PCR, followed by sample pooling, purification and sequencing. The whole procedure takes place in one single plate, without any intermediate purification or plate transfer steps (Fig. 1a). With this easy and quick workflow, it only takes a few hours to prepare sequencing-ready libraries, and the method can be implemented by any laboratory using standard equipment.

We first tested the accuracy of our sorting by performing a species mixing experiment, where equal amounts of HEK293T and NIH3T3 cells were mixed, and scATAC-seq was performed with our method. Using a stringent cut-off (Online Methods), we recovered 307 wells, among which 303 wells contain predominantly either mouse fragments (n = 136) or human fragments (n=167). Only 4 wells are categorised as doublets (Fig. 1b).

To compare our plate-based method to the existing Fluidigm C1 scATAC-seq approach, we performed side-by-side experiments, where cultured K562 and mouse embryonic stem cells (mESC) were tested by both approaches. We used three

metrics to evaluate the quality of the data generated by both methods (Fig. 1c and Supplementary Fig. 1a). Our plate-based method has higher library complexity (library size estimated by the Picard tool), comparable or lower amount of mitochondrial DNA, and higher signal-to-noise ratio measured by fraction of reads in peaks (FRiP) (Fig. 1c). In addition, visual inspection of the read pileup from the aggregated single cells suggested both methods were successful, but data generated from our plate-based method exhibited higher signal (Fig. 1d and e).

The main difference between our method and Fluidigm C1 is the Tn5 tagging strategies. The plate-based method performed Tn5 tagging using a population of cells, while it was done in individual microfluidic chambers in the Fluidigm C1. It is possible that the upfront Tn5 tagging is more efficient than tagging in microfluidic chambers.

To evaluate the generality of our method, we tested the plate-based method on cryopreserved cells from four tissues: human and mouse skin fibroblasts (hSF and mSF)[15] and mouse cardiac progenitor cells (mCPC) at embryonic day E8.5 and E9.5[16]. Cells were revived from liquid nitrogen, and our plate-based method was carried out immediately after revival. The library complexities varied among cell types (Fig. 2a). We obtained median library sizes ranging from 52,747 (mSF) to 104,608.5 (mCPC_E8.5) unique fragments (Fig. 2a). The amount of mitochondrial DNA also varied across cell types but was low in all samples (<13%). All four tested samples had very high signal-to-noise ratio, with a median FRiP ranging from 0.50 (mSF) to 0.60 (hSF) (Fig. 2a). The insert size distributions of the aggregated single cells from all four samples exhibited clear nucleosomal banding patterns (Fig. 2b), which is a feature of high quality ATAC-seq libraries[1]. Finally, visual inspection of aggregate of single cell profiles showed clear open chromatin peaks around expected genes (Fig. 2c and d). Details of all tested cells/tissues are summarised in Supplementary Table 1.

After this validation of the technical robustness of our plate-based method, we further tested it by generating the chromatin accessibility profiles of 3,648 splenocytes (after red blood cell removal) from two C57BL/6Jax mice. In total, we performed two 96-well plates and nine 384-well plates. By setting a stringent quality control threshold (>10,000 reads and >90% mapping rate), 3,385 cells passed the technical cut-off (>90% successful rate) (Supplementary Fig. 3b). The aggregated scATAC-seq profiles exhibited good coverage and signal and resembled the bulk data generated from 10,000 cells by the Immunological Genome Project (ImmGen)[17] (Fig. 3a). The library fragment size distribution before and after sequencing both displayed clear nucleosome banding patterns (Fig. 3b and Supplementary Fig. 2a). In addition, sequencing reads showed strong enrichment around transcriptional start sites (TSS) (Fig. 3c), further demonstrating the quality of the data was high.

Importantly, for the majority of the cells, less than 10% (median 2.1%) of the reads were mapped to the mitochondrial genome (Supplementary Fig. 3a). Overall, we obtained a median of 643,734 reads per cell, while negative controls (empty wells) generated only ~ 100-1,000 reads (Supplementary Fig. 3b). In most cells, more than 98% of the reads were mapped to the mouse genome (Supplementary Fig. 3b), indicating low level of contamination. The median of estimated library sizes is 31,808.5 (Supplementary Fig. 3c). At the sequencing depth of this experiment, the duplication rate of each single cell library is ~ 95% (Supplementary Fig. 3d), indicating that the libraries were sequenced to near saturation. Downsampling the raw reads (from the fastq files) and repeating the analysis suggest that at 20 - 30% of our current sequencing depth, the majority of the fragments would have already been captured (Supplementary Fig. 4a and b). Therefore, in a typical scATAC-seq experiment, ~ 120,000 reads per cell are sufficient to capture most of the unique fragments, with higher sequencing depth still increasing the number of detected unique fragments (Supplementary Fig. 3e).

Next, we examined the data to analyse signatures of different cell types in the mouse spleen. Reads from all cells were merged, and a total of 78,048 open chromatin regions were identified by peak calling with q values less than 0.01[18] (Online Methods). We binarised peaks as "open" or "closed" (Online Methods) and applied a Latent Semantic Indexing (LSI) analysis to the cell-peak matrix for dimensionality reduction[2] (Online Methods). Consistent with previous findings[2], the first dimension is primarily influenced by sequencing depth (Supplementary Fig. 3f). Therefore, we only focused on the second dimension and upwards and visualised the data by t-distributed stochastic neighbour embedding (t-SNE)[19]. We did not observe batch effects from the two profiled spleens, and several distinct populations of cells were clearly identified in the t-SNE plot (Fig. 3d). Read counts in peaks near key marker genes (*e.g. Bcl11a* and *Bcl11b*) suggested that the major populations are B and T lymphocytes, as expected in this tissue (Fig. 4a). In addition, we found a small number of antigen-presenting cell populations (Supplementary Fig. 5), consistent with previous analyses of mouse spleen cell composition[20].

To systematically interrogate various cell populations captured in our experiments, we applied a spectral clustering technique[21] which revealed 12 different cell clusters (Fig. 4b). Reads from cells within the same cluster were merged together to form 'pseudo-bulk' samples and compared to the bulk ATAC-seq data sets generated by ImmGen (Supplementary Fig. 6 and 7). Cell clusters were assigned to the most similar ImmGen cell type (Fig. 4b and Supplementary Fig. 7). In this way, we identified most clusters as different subtypes of B, T and Natural Killer (NK) cells, as well as a small population of granulocytes (GN), dendritic cells (DC) and macrophages (MF) (Fig. 4b and Supplementary Table 2). An aggregate of all single

cells within the same predicted cell type agrees well with the ImmGen bulk ATAC-seq profiles (Supplementary Fig. 8). Remarkably, the aggregate of as few as 55 cells (*e.g.* the predicted MF cell cluster) already exhibited typical bulk ATAC-seq profiles (Supplementary Fig. 8). This finding opens the door for a novel ATAC-seq experimental design, where Tn5 tagging can be performed upfront on large populations of cells (*e.g.* 5,000 - 50,000 cells). Subsequently, cells of interest (for example, marked by surface protein antibodies or fluorescent RNA/DNA probes) can be isolated by FACS, and libraries generated for subsets of cells only. This will be a simple and fast way of obtaining scATAC-seq profiles for rare cell populations.

To test the feasibility of this idea, we stained mouse splenocytes with an anti-CD4 antibody conjugated with PE and performed tagmentation afterwards. The PE signal remained after tagmentation (Supplementary Fig. 9), allowing us to specifically sort out CD4 positive T cells from the rest of the splenocytes for analysis (we named these "TagSort" libraries). As a control, we first purified CD4 T cells using an antibody-based depletion method (Online Method), and subsequently performed scATAC-seq on the purified CD4 T cells (we named these "SortTag" libraries). The data of CD4 T cells generated from these two strategies agree very well (Fig. 4c). The library complexity is comparable with median library sizes of 30,953 and 25,830 respectively (Fig. 4c, top left panel). The binding signals around open chromatin peaks are highly correlated (Pearson r=0.96) (Fig. 4c, top right panel). Visual inspection of read pileup profiles around the *Cd4* gene locus from single cell aggregates suggested the data are of good quality (Fig. 4c, bottom panel).

This experiment serves as a proof-of-principle test where staining of a surface marker can be done before Tn5 tagging, and a specific population can be sorted by FACS afterwards for scATAC-seq analysis. It should be noted that we have only tested CD4 - an abundant marker in a subpopulation of splenocytes. Other surface markers in different tissues would need to be investigated individually. In addition, the ability to investigate rare cell populations using this approach is limited by the frequency of the rare cell types and the amount of cells that can be tagged upfront.

The spectral clustering was able to distinguish different cell subtypes, such as naïve and memory CD8 T cells, naïve and regulatory CD4 T cells and CD27+ and CD27- NK cells (Fig. 4b). Previous studies have identified many enhancers that are only accessible in certain cell subtypes, and these are robustly identified in our data. Examples are the *Il2rb* and *Cd44* loci in memory CD8 T cells[22] and *Ikzf2* and *Foxp3* in regulatory T cells[23] (Supplementary Fig. 10a and b). Interestingly, our clustering approach successfully identified two subtle subtypes of NK cells (CD27- and CD27+ NK cells), as determined by their open chromatin profiles (Fig. 4b and d). It has been shown that, upon activation, NK cells can express CD83[24], a well-known marker for mature dendritic cells[25]. In mouse spleen, *Cd83* expression was barely detectable in

the two NK subpopulations profiled by the ImmGen consortium (Supplementary Fig. 10c). However, in our data, the *Cd83* locus exhibited different open chromatin states in the two NK clusters (Fig. 4d). Multiple ATAC-seq peaks were observed around the *Cd83* locus in the CD27+ NK cell cluster but not in the CD27- NK cluster (Fig. 4d). This suggests that *Cd83* is in a transcriptionally permissive state in the Cd27+ NK cells, and the CD27+ NK cells have a greater potential for rapidly producing CD83 upon activation. This may partly explain the functional differences between CD27+ and CD27- NK cell states[26].

Finally, we investigated whether we could identify the regulatory regions that define each cell cluster. To this end, we trained a logistic regression classifier using the spectral clustering labels and the binarised scATAC-seq count data (Online Methods). From the classifier, we extracted the top 500 open chromatin peaks (marker peaks) that can distinguish each cell cluster from the others (Fig. 4e and Online Methods). By looking at genes in the vicinity of the top 50 marker peaks, we recapitulated known markers, such as *Cd4* for the helper T cell cluster (cluster 3), *Cd8a* and *Cd8b1* for the cytotoxic T cell cluster (cluster 6) and *Cd9* for marginal zone B cell cluster (cluster 4) (Supplementary Figure 11 and Supplementary Table 3). These results are consistent with our correlation-based cell cluster annotation (Fig. 4b).

While the peaks at TSS are useful for cell type annotation, the majority of the cluster-specific marker peaks are in intronic and distal intergenic regions, in line with the global peak distribution (Supplementary Fig. 12). To identify transcription factors that are important for the establishment of these marker peaks, we investigated them in more detail by motif enrichment analysis using HOMER[27]. The full results of these motif enrichment analyses are included in Supplementary Table 4. As expected, different ETS motifs and ETS-IRF composite motifs were significantly enriched in marker peaks of many clusters (Fig. 4f), consistent with the notion that ETS and IRF transcription factors are important for regulating immune activities[28]. Furthermore, we found motifs that were specifically enriched in certain cell clusters (Fig. 4f). Our motif discovery is consistent with previous findings, such as the importance of T-box (*e.g.* Tbx21) motifs in NK[29] and CD8T memory cells[30] and POU domain (*e.g.* Pou2f2) motifs in marginal zone B cell[31]. This suggests that our scATAC-seq data is able to identify known gene regulation principles in different cell types within a tissue.

In recent years, other methods, such as DNase-seq[32], MNase-seq[33] and NOMe-seq[34,35], have investigated chromatin status at the single cell level. However, due to its simplicity and reliability, ATAC-seq currently remains the most popular technique for chromatin profiling. Several recent studies have demonstrated the power of using scATAC-seq for investigating regulatory principles, *e.g.* brain

development[4,9], Mouse sci-ATAC-seq Atlas[36] and pseudotime inference[37]. The combined multi-omics approaches also began to emerge, such as sci-CAR-seq[38], scCAT-seq[39] and piATAC-seq[8]. Our study added on top of those methods to provide a simple and easy-to-implement scATAC-seq approach that can successfully detect different cell populations, including subtle and rare cell subtypes, from a complex tissue. More importantly, it is able to reveal key gene regulatory features, such as cell-type specific open chromatin regions and transcription factor motifs, in an unbiased manner. Future studies can utilise this method to unveil the regulatory characteristics of novel and rare cell populations and the mechanisms behind their transcriptional regulation.

Figure Legends

Figure 1. Simple and robust analysis of chromatin status at the single cell level. (a) Schematic view of the workflow of the scATAC-seq method. Tagmentation is performed upfront on bulk cell populations, followed by sorting single-nuclei into 96/384 well plates containing lysis buffer. The lysis buffer contains a low concentration of proteinase K and SDS to denature the Tn5 transposase and fragment the genome. Tween-20 is added to quench SDS[14]. Subsequently, library preparation by indexing PCR is performed, and the number of PCR cycles needed to amplify the library is determined by quantitative PCR (qPCR) (**Supplementary Fig. 2b**). (b) Species mixing experiments to show the accuracy of FACS. Equal amounts of HEK293T (Human) and NIH3T3 (Mouse) cells were mixed, and scATAC-seq was performed as described in (a). Successful wells with more than 90% of reads uniquely mapped to either human or mouse were categorised as singlets (n=303). Otherwise, they will be categorised as doublets (n=4) (see Online Methods). (c) Comparison of the median library size (estimated by the Picard tool), fraction of mitochondrial DNA (MT content) and fraction of reads in peaks (FRiP) in single cells from either C1 (blue) or plate-based (red) scATAC-seq approach. (d) UCSC genome browser tracks displaying the signal around the *Nanog* gene locus from the aggregate of mESCs obtained from Fluidigm C1 (top) and plate (bottom). (e) The same type of tracks as (d) around the *ZBTB32* gene locus in K562 cells.

Figure 2. Plate-based scATAC-seq worked robustly on cryopreserved cells from primary tissues. (a) Comparison of the median library size (estimated by the Picard tool), fraction of mitochondrial DNA (MT content) and fraction of reads in peaks (FRiP) in cryopreserved single cells from four different tissues: human skin fibroblasts (hSF), mouse skin fibroblasts (mSF), mouse cardiac progenitor cells (mCPC) at embryonic day E8.5 and E9.5. (b) Insert size frequencies from the aggregated data of the cells from the four tissues. (c and d) UCSC genome browser tracks displaying the signal around the *RPS18* gene locus from the aggregate of hSFs (c) and around the *Gapdh* gene locus from the aggregate of mSFs, mCPC_E8.5 and mECP_E9.5 (d).

Figure 3. Plate-based scATAC-seq applied to over 3000 mouse splenocytes. (a) UCSC genome browser tracks displaying the signal around the *Cxcr5* gene locus from the aggregate of all single cells in this study. Bulk ATAC-seq profiles from the ImmGen consortium are also shown. Randomly selected 100 single cell profiles are show below the aggregated profile. (b and c) Insert size frequencies (b) and sequencing read distributions across transcriptional start sites (c) of libraries from the aggregated data (the red line) and individual single cells (grey lines, 24 examples are shown). (d) A two-dimensional projection of the scATAC-seq data using t-SNE.

Colours represent two different batches, showing excellent agreement between batches. Sp: spleen.

Figure 4. **Identification of different cell types and cell-type specific open chromatin regions and transcription factor motifs.** (a) The same t-SNE plot as in **Figure 3d**, coloured by the number of counts in the peaks near indicated gene locus. (b) The same t-SNE plot as in **Figure 3d** coloured by spectral clustering and cell type annotation. (c) Comparisons of spleen CD4 T cells scATAC-seq obtained by two strategies. TagSort: cells were stained with anti-CD4-PE, tagged with Tn5 and CD4-PE positive cells were sorted for scATAC-seq; SortTag: CD4 T cells were purified first and scATAC-seq was performed on the purified cells. Top: comparison of library size and binding signal correlation (pearson r=0.96) around called peaks; bottom: UCSC genome browser tracks of the indicated single cell aggregates around the *Cd4* gene locus. (d) UCSC genome browser tracks around *Cd27* and *Cd83* gene loci, displaying the aggregate (top panel) and single cell (bottom panel) signals of the two NK clusters. ATAC-seq peaks specific to the CD27+ NK cells are highlighted. For visual comparison reason, we randomly choose 65 out of 75 CD27- NK cells. (e) Z-score of normalised read counts in the top 500 peaks that distinguish each cell cluster based on the logistic regression classifier, across each peak (row) in each cell (column). Top 500 marker peaks were picked per cell cluster, so there are 500 x 12 = 6,000 peaks in the heatmap. Cells are ordered by cluster labels. (f) Heatmap representation of transcription factor motif (rows) enrichments (binomial test *p*-values) in the top 500 marker peaks in different cell clusters (columns). Some key motifs are enclosed by black rectangles and motif logos are shown to the right. Motif names are taken from the HOMER software suite.

Supplementary Figure 1. (a) Violin plots showing the comparisons of distributions of the median library size (estimated by the Picard tool), fraction of mitochondrial DNA (MT content) and fraction of reads in peaks (FRiP) in single cells from either plate or C1 scATAC-seq approach. (b) the same metrics as in (a) showing data obtained from cryopreserved cells of four different primary tissues.

Supplementary Figure 2. (a) Bioanalyzer results of pools of 11 different plates (two spleens) of scATAC-seq in this study. (b) qPCR amplification plot of 64 different single cell libraries. The qPCR was performed after 8 cycles of pre-amplification. Dotted line indicates the number of cycles used for final amplification. A total of 8 + 10 = 18 cycles were performed in this study.

Supplementary Figure 3. Scatter plots of different quality control metrics. Single cells from different batches are indicated by different colours, and empty well controls are also indicated. We removed cells that have less than 10,000 reads or less than 90% mapping rate, as indicated by dotted lines in (b). Sp: spleen; MT

content: fraction of mitochondrial reads; LSI: latent semantic indexing; Library size is estimated by the Picard tool.

Supplementary Figure 4. (a) The median library size after downsampling (at the fastq stage) to different fractions relative to the full data sets. (b) Violin Plot of the library size at the different level of downsampling.

Supplementary Figure 5. Number of counts from all peaks that assigned to the indicated genes by HOMER.

Supplementary Figure 6. Hierarchical clustering of the Pearson's correlation between aggregated single cell clusters and the bulk ATAC-seq data sets from ImmGen. The full matrix is shown here, and the ImmGen sample labels were taken directly from the ImmGen ATAC-seq data deposited at the European Nucleotide Archive (ENA) (https://www.ebi.ac.uk/ena/data/view/PRJNA392905).

Supplementary Figure 7. The top correlated ImmGen bulk samples to each aggregated single cell clusters. Top 5 pearson r scores for each clusters are shown.

Supplementary Figure 8. UCSC genome browser tracks showing ATAC-seq profiles of indicated ImmGen bulk samples and aggregated single cell clusters around the *Ptprc* (*Cd45*) promoter region.

Supplementary Figure 9. FACS results showing the anti-CD4-PE and DAPI stain on mouse splenocytes before (top) and after (bottom) Tn5 tagging. Note, all cells are DAPI negative before Tn5 tagging but become DAPI positive afterwards. CD4-PE signal remains after Tn5 tagging.

Supplementary Figure 10. (a and b) UCSC genome browser tracks showing ATAC-seq profiles of aggregate (top panel) and individual single cells (bottom panels). Known enhancers are highlighted. (c) *Cd83* expression from the ImmGen bulk RNA-seq of the indicated sample.

Supplementary Figure 11. Nearest genes assigned to the top 50 marker peaks in each single cell cluster.

Supplementary Figure 12. Genomic distribution (by HOMER) of all peaks and the marker peaks in each single cell cluster.

Author contributions

X.C, K.N.N and S.A.T conceived the project. X.C designed the protocol. X.C, R.J.M and K.N.N performed the experiments. X.C carried out the computational analysis. S.A.T supervised the entire project. All authors contributed to the writing.

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Data and materials availability

The sequencing data has been deposited at ArrayExpress (accession E-MTAB-6714). The code used for the analysis is available on the Github repository https://github.com/dbrg77/plate_scATAC-seq.

The UCSC genome browser tracks containing both the ImmGen bulk ATAC-seq and scATAC-seq from this study can be viewed via this link: http://genome-euro.ucsc.edu/cgi-bin/hgTracks?hgS_doOtherUser=submit&hgS_otherUserName=dbrg77&hgS_otherUserSessionName=mSpleen_scATAC_cluster.

Competing financial interests

None declared.

Methods

Cell isolation

For splenocytes, the spleen from a C57BL/6Jax mouse was mashed by a 2-ml syringe plunger through a 70 μm cell strainer (Fisher Scientific 10788201) into 30 ml 1X DPBS (ThermoFisher 14190169) supplied with 2 mM EDTA and 0.5% (w/v) BSA (Sigma A9418). Cells were centrifuged down, supernatant was removed, and the cell pellet was briefly vortexed. 5 ml 1X RBC lysis buffer (ThermoFisher 00-4300-54) was used to resuspend the cell pellet, and the cell suspension was vortexed again, and left on bench for 5 minutes to lyse red blood cells. Then 45 ml 1X DPBS was added, and cells were centrifuged down. 30 ml 1X DPBS were used to resuspend the cell pellet. The cell suspension was passed through a Miltenyi 30 μm Pre-Separation Filter (Miltenyi 130-041-407), and the cell number was determined using C-chip counting chamber (VWR DHC-N01). All centrifugations were done at 500 g, 4 °C, 5 minutes. For human and mouse skin fibroblasts, cells were extracted as previously described[15]. For mouse cardiac progenitor cells, cells were extracted as previously described[16]. Cells were cryopreserved in 90% FBS and 10% DMSO and stored in liquid nitrogen until experiments.

Plate-based single-cell ATAC-seq (scATAC-seq).

A detailed step-by-step protocol can be found in the Supplementary Protocol. Briefly, 50,000 cells were centrifuged down at 500 g, 4 °C, 5 minutes. Cell pellets were resuspended in 50 μl tagmentation mix (33 mM Tris-acetate, pH 7.8, 66 mM potassium acetate, 10 mM magnesium acetate, 16% dimethylformamide (DMF), 0.01% digitonin and 5 μl of Tn5 from the Nextera kit from Illumina, Cat. No. FC-121-1030). The tagmentation reaction was done on a thermomixer (Eppendorf 5384000039) at 800 rpm, 37 °C, 30 minutes. The reaction was then stopped by adding equal volume (50 μl) of tagmentation stop buffer (10 mM Tris-HCl, pH 8.0, 20 mM EDTA, pH 8.0) and left on ice for 10 minutes. 200 μl 1X DPBS with 0.5% BSA was added and the nuclei suspension was transferred to a FACS tube. DAPI (ThermoFisher 62248) was added at a final concentration of 1 μg/μl to stain the nuclei.

Species mixing experiments

25,000 HEK293T (Human) and 25,000 NIH3T3 (Mouse) cells were mixed together, and scATAC-seq was performed as described in the Supplementary Protocol. The obtained sequencing reads were mapped to a concatenated genome of mouse and human by hisat2[40]. One 384-well plate was performed. We first set a technical cutoff where a successful well must contain more than 10,000 total reads and more than 90% of reads are mapped to the concatenated genome. 307 wells were marked as

successful. Among the successful wells, we calculated the ratio of reads that mapped to the human genome and the mouse genome. If the ratio is larger than 10, the well is categorised as containing human single cells; if the ratio is less than 0.1, the well is categorised as containing mouse single cells; otherwise, the well is categorised as containing human-mouse doublets.

Plate scATAC-seq on CD4+ T cells (TagSort vs SortTag)

For the "TagSort" strategy, 50,000 splenocytes were stained with anti-Mouse CD4 PE (eBioscience cat no. 12-0043-82) at room temperature for 30 minutes according to the manufacturer's instructions. The stained cells were washed with ice-cold 1X PBS twice and pelleted down at 500 g, 4 °C, 5 minutes. Experimente were carried out following the procedures described in the Supplementary Protocol. DAPI and PE double positive cells were sorted into a 384-well plate for library construction. For the "SortTag" strategy, CD4+ T cells were purified first from mouse splenocytes using the Naive CD4 T Cell Isolation Kit, Mouse (Miltenyi, cat. no. 130-104-453) following the manufacturer's instruction without the anti-CD44 depletion step. The purified CD4 T cells were processed according to the procedures described in the Supplementary Protocol.

scATAC-seq using Fluidigm C1

Experiments were performed as previously described[3] using the medium-sized (1862x) Open App chip. We followed the manufacturer's instructions described in the "ATAC Seq No Stain (Rev C)" from the Fluidigm ScriptHub (https://www.fluidigm.com/c1openapp/scripthub), except that we replace the detergent NP-40 in the original protocol with digitonin so that the final concentration of digitonin in the reaction chamber is 0.005%. After collecting the pre-amplified material from the Fluidigm chip, the libraries were indexed by library PCR for 14 cycles as previously described[3].

Costs involved in plate-based and Fluidigm C1 scATAC-seq

For our plate-based scATAC-seq method, most reagents and buffers are available in a standard molecular biology lab. Exceptions are the Tn5 transposase, which can be purchased from Illumina (Cat No. FC-121-1030), and the PCR master mix, which can be purchased from various vendors (we used the 2X NEBNext® High-Fidelity 2X PCR Master Mix from NEB). Since the Tn5 tagging reaction was performed upfront at the bulk level, the Tn5 cost per cell depends on how many cells are sorted during the sorting. Based on our experience, when 50,000 cells are used at the beginning, two to eight 384-well plates can be sorted. Therefore, the cost of Tn5 is negligible. The major cost per unit for the plate-based scATAC-seq is the PCR master mix used during library amplification. Currently, 10 µl of PCR master mix are needed per cell in a 20 µl library amplification reaction, but we have been successfully and consistently generated libraries from half of the volume described in the protocol.

For scATAC-seq using the Fluidigm C1, all the aforementioned reagents are needed, and a microfluidic chip is required per 96 cells.

Hands-on time for plate-based versus C1 scATAC-seq approaches

For our plate-based scATAC-seq method, the most time-consuming part is the lysis plate preparation (mixing lysis buffer and indexing primers). For maximum efficiency, this can be done upfront in bulk, and the lysis plate is stable in -80 °C for a long time. Another time/labour-consuming step is the pooling of single cell libraries after PCR using a multi-channel pipette. We provide online advice to perform the whole procedure in minutes. This information is included in the accompanying GitHub page: https://github.com/dbrg77/plate_scATAC-seq. For scATAC-seq using the Fluidigm C1, an extra ~4 hours of C1 runtime are needed.

qPCR for library amplification

After assembly of the 20 µl PCR reaction (see **Supplementary Protocol**), a pre-amplification step was performed on a PCR machine (Alpha Cycler 4, PCRmax) with 72 °C 5 minutes, 98 °C 5 minutes, 8 cycles of [98 °C 10 seconds, 63 °C 30 seconds, 72 °C 20 seconds]. Of the product, 19 µl of pre-amplified library were transferred to a 96 well qPCR plate, 1 µl 20X EvaGreen (Biotium #31000) was added, and qPCR was performed on an ABI StepOnePlus system with the following cycle conditions: 98 °C 1 minutes, 20 cycles of [98 °C 10 seconds, 63 °C 30 seconds, 72 °C 20 seconds]. Data was acquired at 72 °C. We qualitatively chose the cycle number to where the fluorescence signals just about to start going up (**Supplementary Fig. 1b**). In this study, a total of 18 cycles were used to amplify the libraries.

Sequencing data processing

All sequencing data were processed using a pipeline written in snakemake[41]. The software/packages and the exact flags used in this study can be found in the 'Snakefile' provided in the GitHub repository https://github.com/dbrg77/plate_scATAC-seq. Briefly, reads were trimmed with cutadapt[42] to remove the Nextera sequence at the 3' end of short inserts. The trimmed reads were mapped to the reference mouse genome (UCSC mm10) using hisat2[40]. Reads with mapping quality less than 30 were removed by samtools[43] (-q 30 flag) and deduplicated using the MarkDuplicates function of the Picard tool (http://broadinstitute.github.io/picard). All reads from single cells were merged together using samtools, and the merged BAM file was deduplicated again. Peak calling was performed on the merged and deduplicated BAM file by MACS2[18]. For bulk ATAC-seq and single cell aggregate coverage visualisation, bedGraph files generated from MACS2 callpeak were converted to bigWig files and visualised via UCSC genome browser. For individual single cell ATAC-seq visualisation, aligned

reads from individual cells were converted to bigBed files. A count matrix over the union of peaks was generated by counting the number of reads from individual cells that overlap the union peaks using coverageBed from the bedTools suite[44].

Public ATAC-seq data processing

FASTQ files were all downloaded from the European Nucleotide Archive (ENA). The ImmGen bulk ATAC-seq data (study accession PRJNA392905) and the scATAC-seq data using Fluidigm C1 (study accessions PRJNA274006 and PRJNA299657) were processed in the same way as described in this study. The 'Snakefile' used to process the data can be found at the the same GitHub repository.

Bioinformatics analysis

Codes used to carry out all the analyses were provided as Jupyter Notebook files, which can be found in the same GitHub repository. Briefly, downsampling was performed by randomly selecting a fraction of reads from the original FASTQ files using seqtk (https://github.com/lh3/seqtk), and the same pipeline was run on the sub-sampled FASTQ files. For binarising the scATAC-seq data, peak calling was performed on reads merged from all cells, and we labelled the peak '1' (open) if there was at least one read overlapping the peak, and '0' (closed) otherwise. Latent semantic indexing analysis was performed by first normalising the binarized count matrix by term frequency inverse document frequency (TF-IDF) and then performing a Singular-Value Decomposition (SVD) on the normalised count matrix. Only the 2nd - 50th dimensions after the SVD were passed to t-SNE for visualisation. To compare with ImmGen bulk ATAC-seq data, a reference peak set was created by taking the union of peaks from the peak calling results of aggregated scATAC-seq (this study) and different samples of ImmGen bulk ATAC-seq using mergeBed from the bedTools suite[44]. All comparisons were done based on this reference peak set. The annotatePeaks.pl from HOMER[27] was used to assign genes to peaks. Latent semantic indexing, spectral clustering and logistic regression were carried out using Scikit-learn[45].

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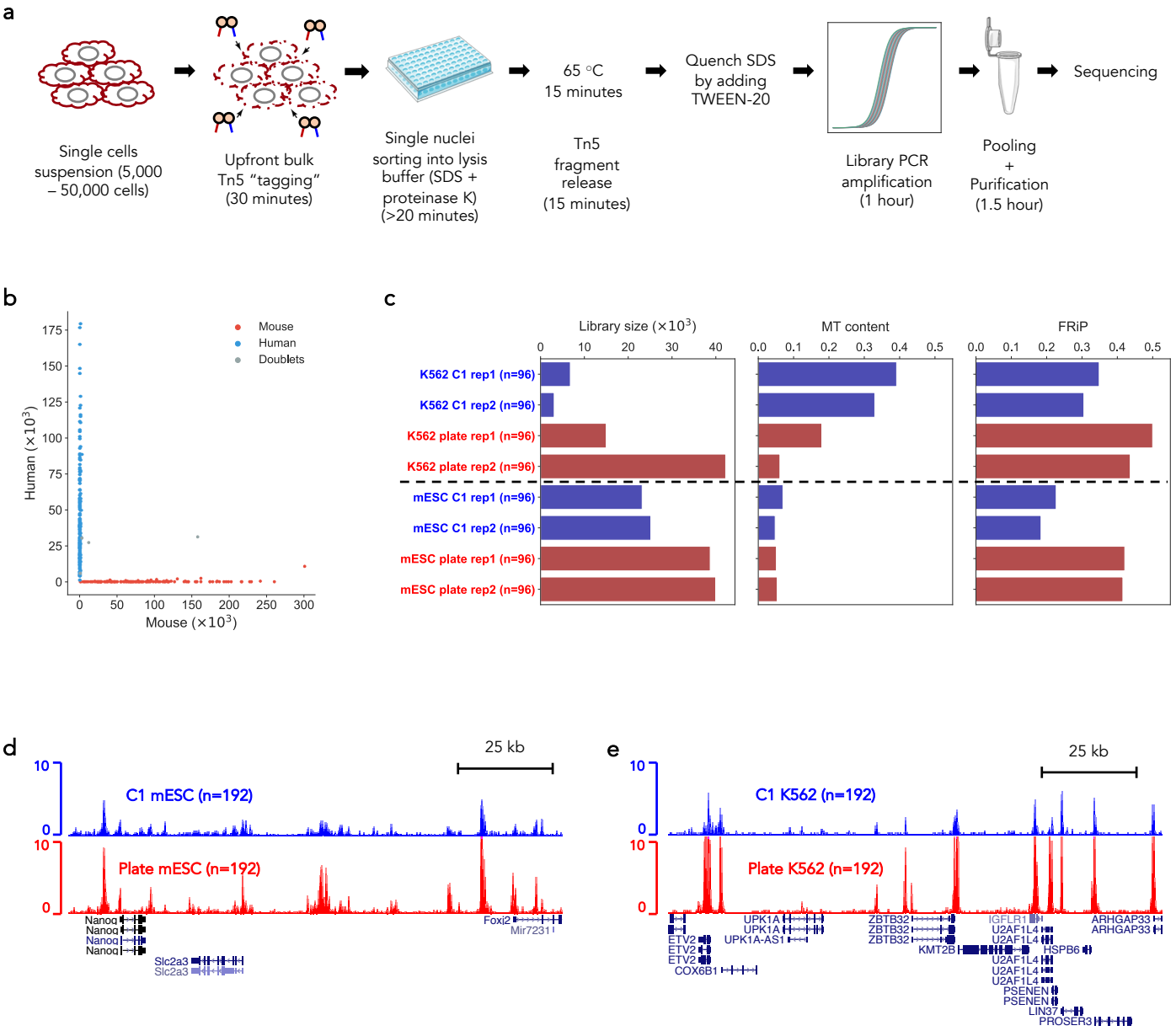

a
Single cells suspension (5,000 – 50,000 cells)
Upfront bulk Tn5 "tagging" (30 minutes)
Single nuclei sorting into lysis buffer (SDS + proteinase K) (>20 minutes)
65 °C 15 minutes
Tn5 fragment release (15 minutes)
Quench SDS by adding TWEEN-20
Library PCR amplification (1 hour)
Pooling + Purification (1.5 hour)
Sequencing
b
Mouse
Human
Doublets
Human (×10³)
Mouse (×10³)
c
Library size (×10³)
MT content
FRiP
K562 C1 rep1 (n=96)
K562 C1 rep2 (n=96)
K562 plate rep1 (n=96)
K562 plate rep2 (n=96)
mESC C1 rep1 (n=96)
mESC C1 rep2 (n=96)
mESC plate rep1 (n=96)
mESC plate rep2 (n=96)
d
25 kb
C1 mESC (n=192)
Plate mESC (n=192)
Nanog
Foxi2
Mir7231
Slc2a3
e
25 kb
C1 K562 (n=192)
Plate K562 (n=192)
UPK1A
UPK1A-AS1
ETV2
COX6B1
ZBTB32
KMT2B
IGFLR1
U2AF1L4
HSPB6
ARHGAP33
PSENEN
LIN37
PROSER3

Figure 1

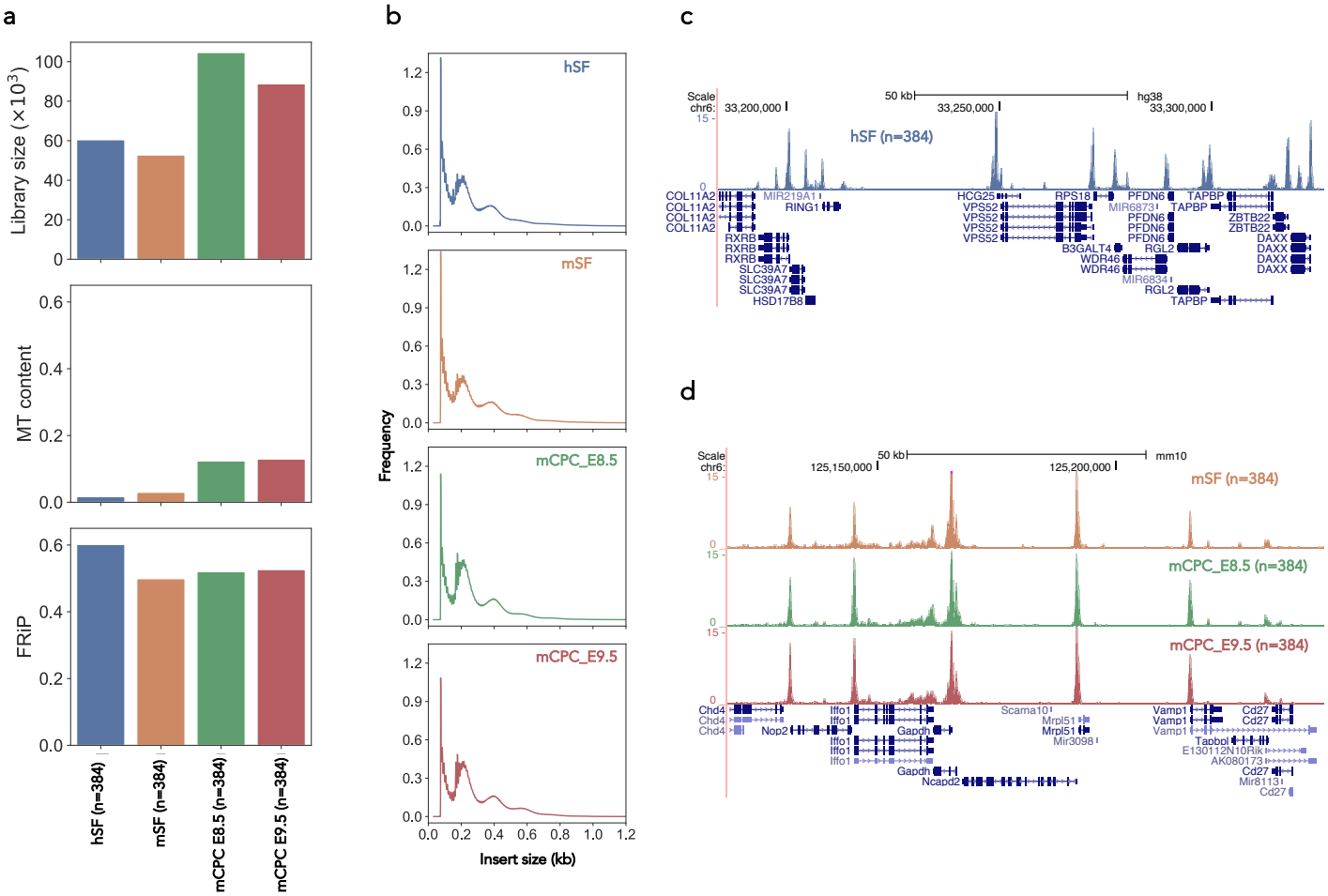

a
Library size (×10³)
MT content
FRiP
hSF (n=384)
mSF (n=384)
mCPC E8.5 (n=384)
mCPC E9.5 (n=384)
b
hSF
mSF
mCPC_E8.5
mCPC_E9.5
Frequency
Insert size (kb)
c
hg38
hSF (n=384)
d
mm10
mSF (n=384)
mCPC_E8.5 (n=384)
mCPC_E9.5 (n=384)

Figure 2

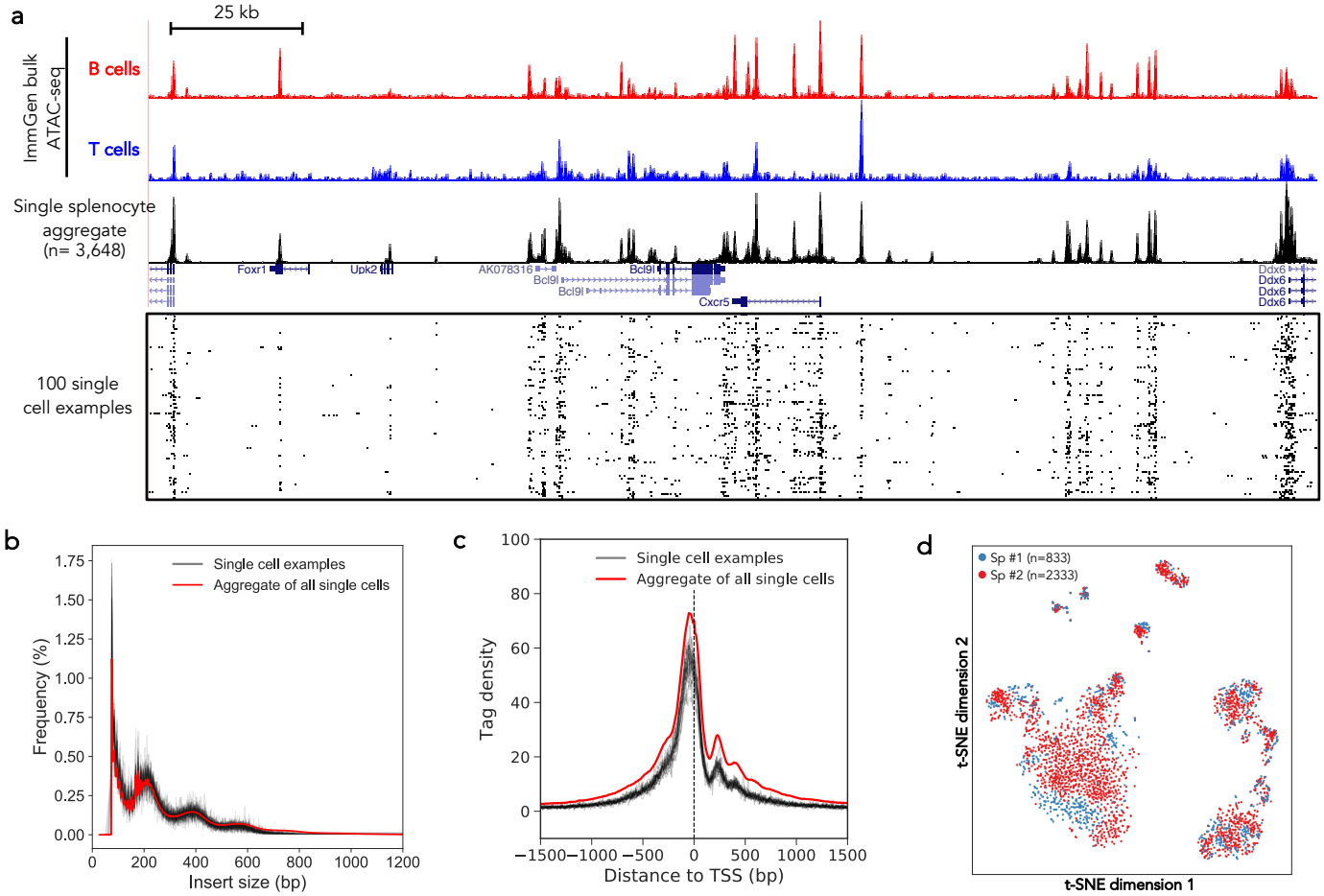

a
25 kb
ImmGen bulk ATAC-seq
B cells
T cells
Single splenocyte aggregate (n= 3,648)
Foxr1
Upk2
AK078316
Bcl9l
Cxcr5
Ddx6
100 single cell examples
b
Single cell examples
Aggregate of all single cells
Frequency (%)
Insert size (bp)
c
Tag density
Distance to TSS (bp)
d
Sp #1 (n=833)
Sp #2 (n=2333)
t-SNE dimension 1
t-SNE dimension 2

Figure 3

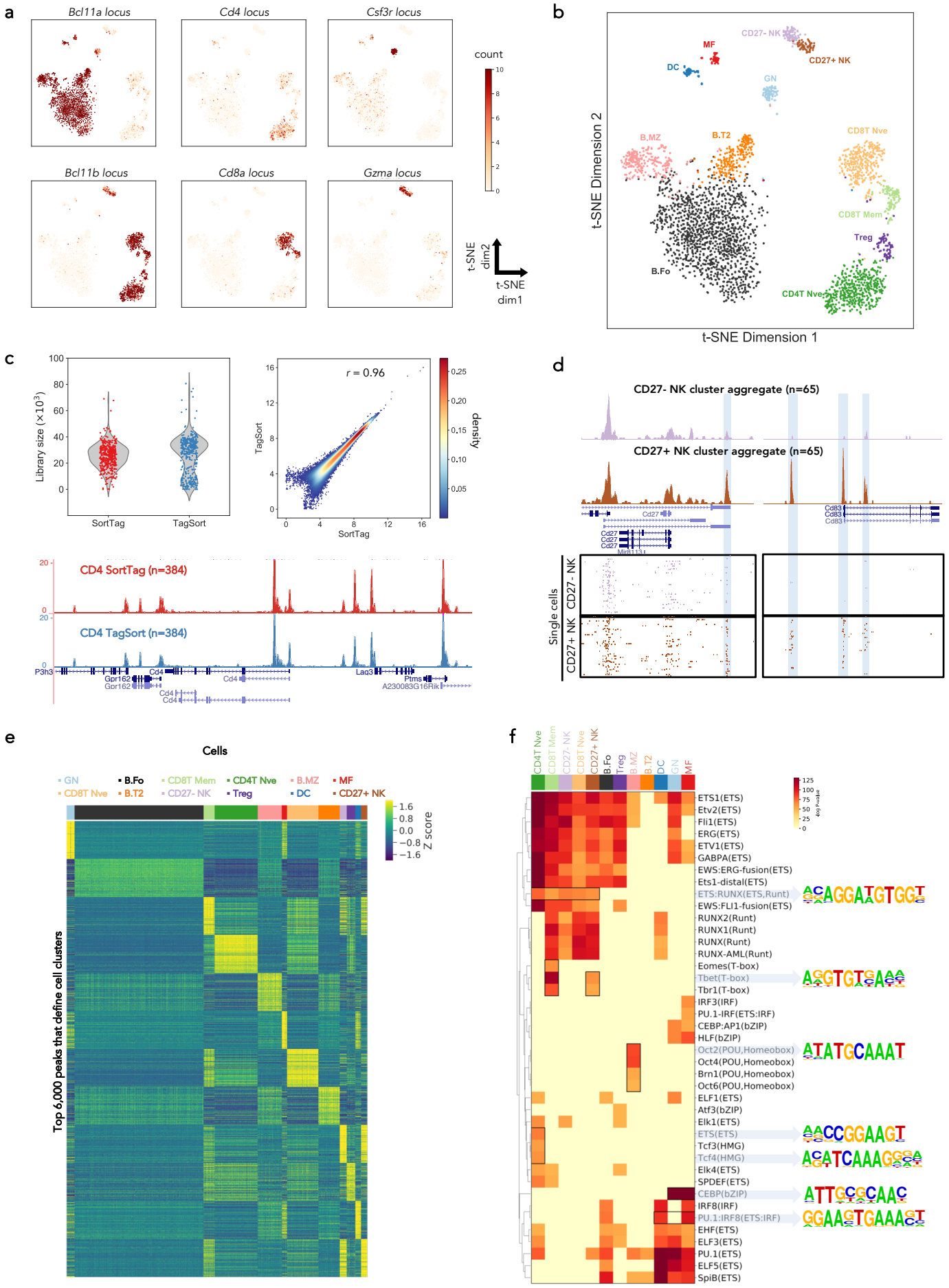
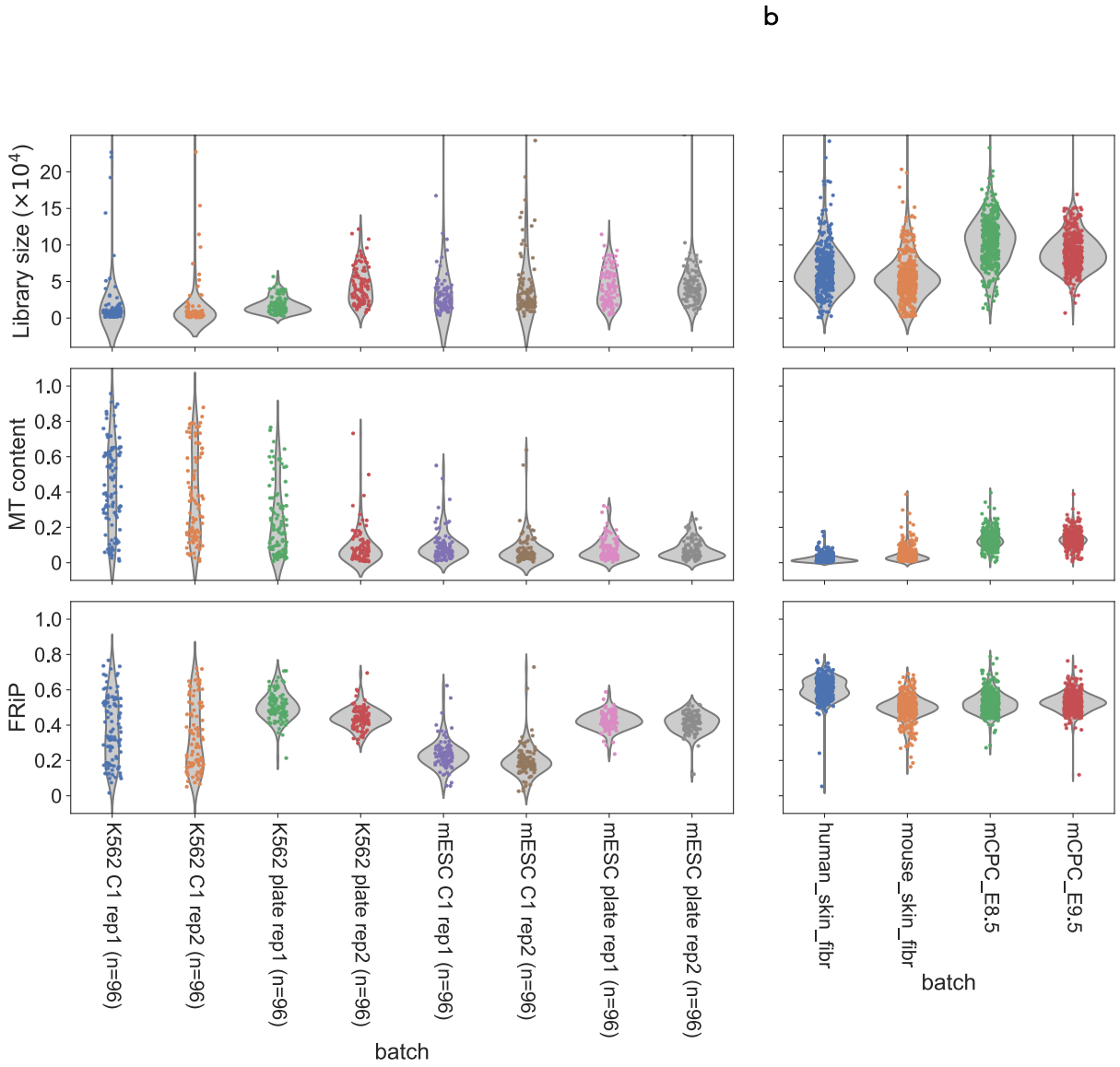
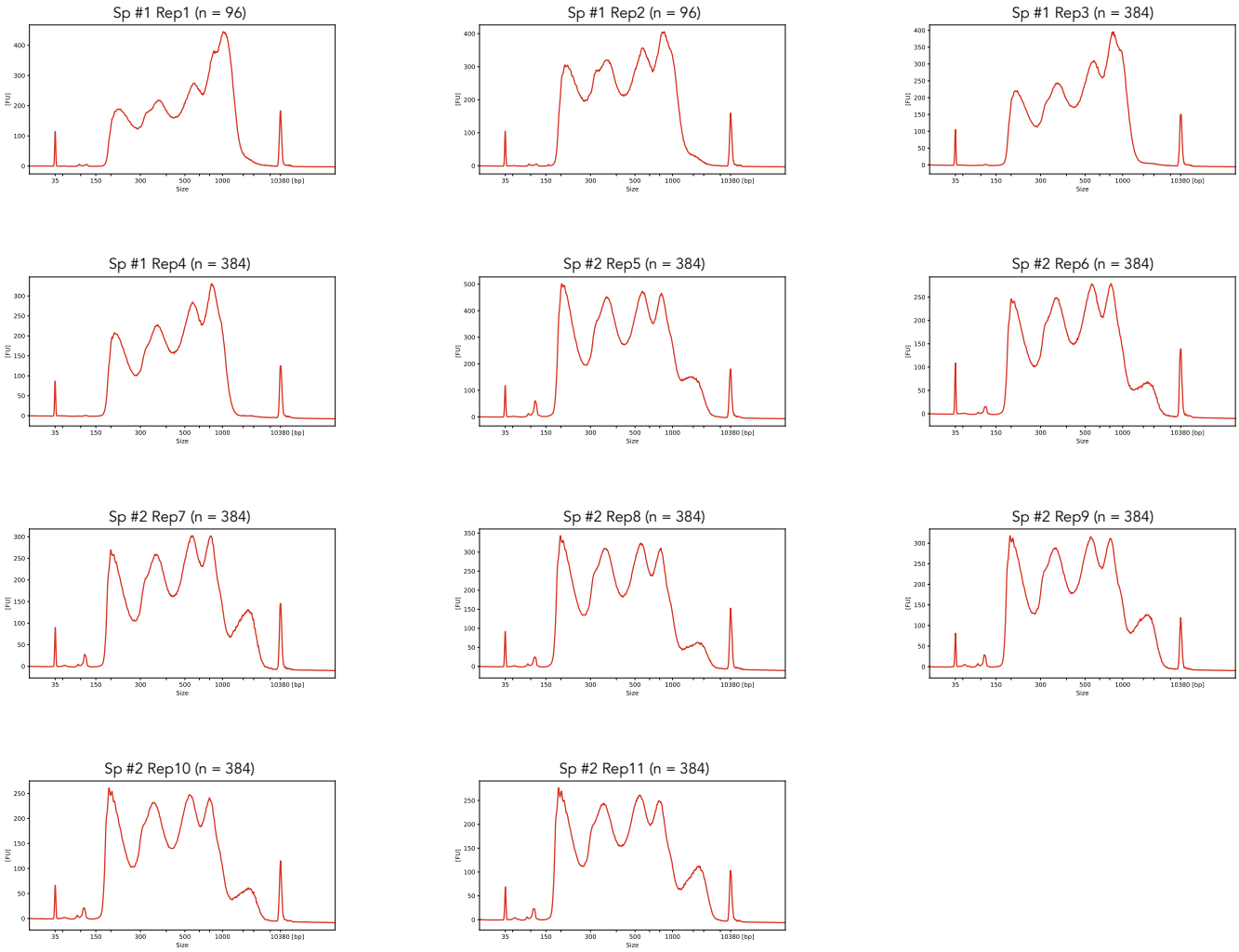

a
Bcl11a locus
Cd4 locus
Csf3r locus
Bcl11b locus
Cd8a locus
Gzma locus
count
t-SNE dim2
t-SNE dim1
b
CD27- NK
MF
CD27+ NK
DC
GN
B.MZ
B.T2
CD8T Nve
CD8T Mem
Treg
B.Fo
CD4T Nve
t-SNE Dimension 2
t-SNE Dimension 1
c
Library size (×10³)
SortTag
TagSort
r = 0.96
density
CD4 SortTag (n=384)
CD4 TagSort (n=384)
d
CD27- NK cluster aggregate (n=65)
CD27+ NK cluster aggregate (n=65)
Single cells
CD27- NK
CD27+ NK
e
Cells
GN
B.Fo
CD8T Mem
CD4T Nve
B.MZ
MF
CD8T Nve
B.T2
CD27- NK
Treg
DC
CD27+ NK
Z score
Top 6,000 peaks that define cell clusters
f
ETS1(ETS)
Etv2(ETS)
Fli1(ETS)
ERG(ETS)
ETV1(ETS)
GABPA(ETS)
EWS:ERG-fusion(ETS)
Ets1-distal(ETS)
ETS:RUNX(ETS,Runt)
EWS:FLI1-fusion(ETS)
RUNX2(Runt)
RUNX1(Runt)
RUNX(Runt)
RUNX-AML(Runt)
Eomes(T-box)
Tbet(T-box)
Tbr1(T-box)
IRF3(IRF)
PU.1-IRF(ETS:IRF)
CEBP:AP1(bZIP)
HLF(bZIP)
Oct2(POU,Homeobox)
Oct4(POU,Homeobox)
Brn1(POU,Homeobox)
Oct6(POU,Homeobox)
ELF1(ETS)
Atf3(bZIP)
Elk1(ETS)
ETS(ETS)
Tcf3(HMG)
Tcf4(HMG)
Elk4(ETS)
SPDEF(ETS)
CEBP(bZIP)
IRF8(IRF)
PU.1:IRF8(ETS:IRF)
EHF(ETS)
ELF3(ETS)
PU.1(ETS)
ELF5(ETS)
SpiB(ETS)

Figure 4

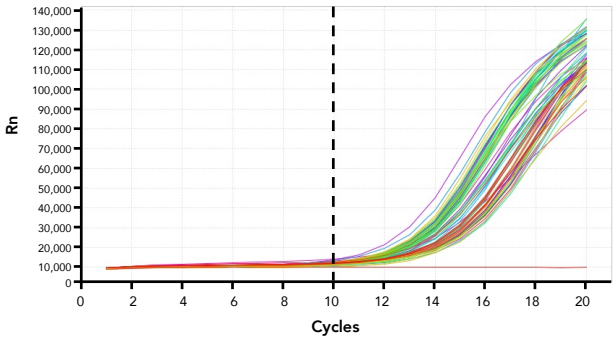


Supplementary Figure 1

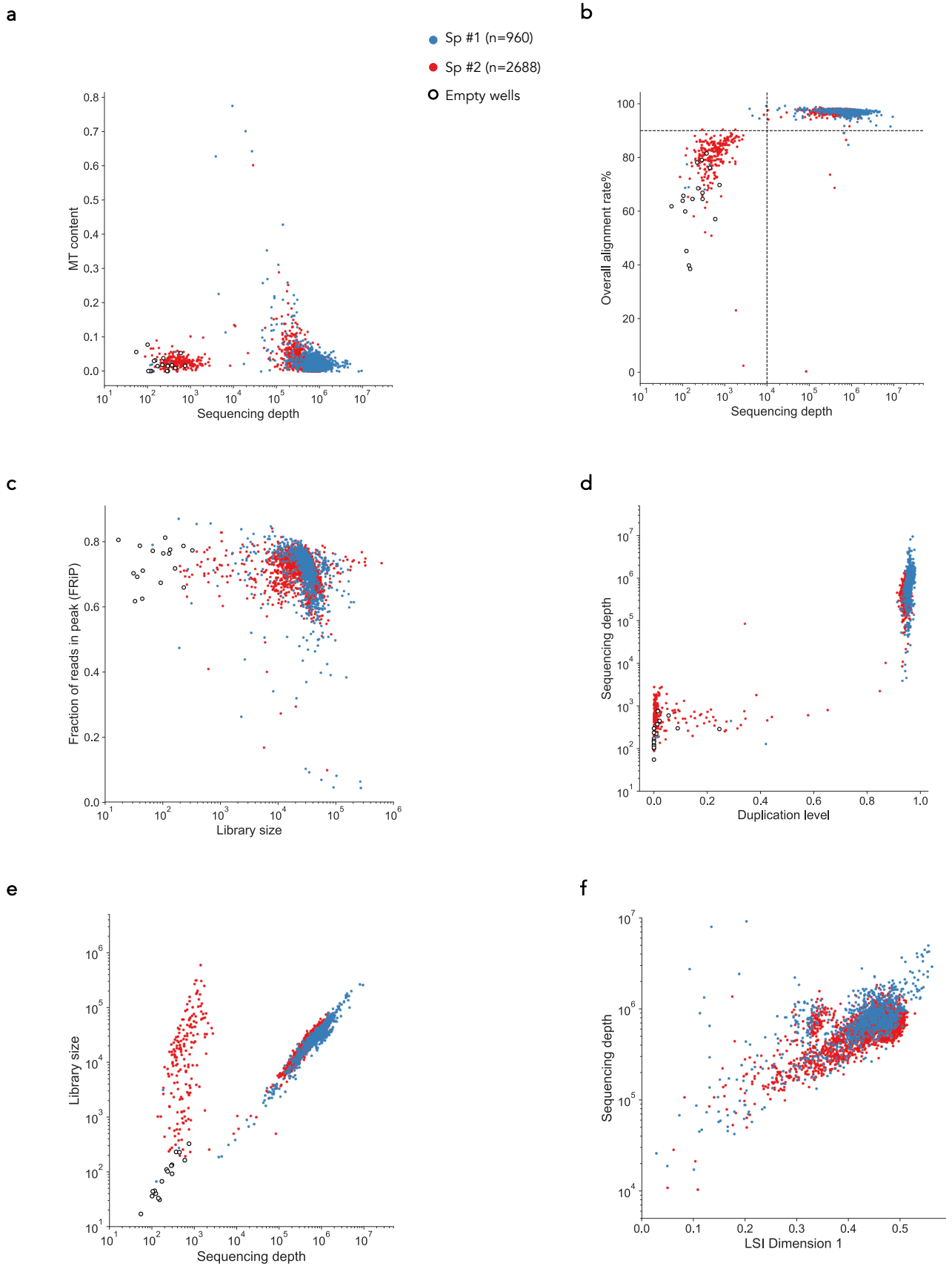
a



b

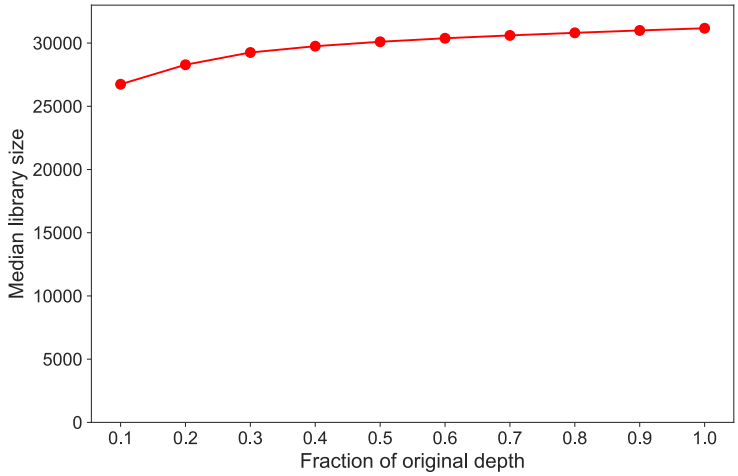


Supplementary Figure 2

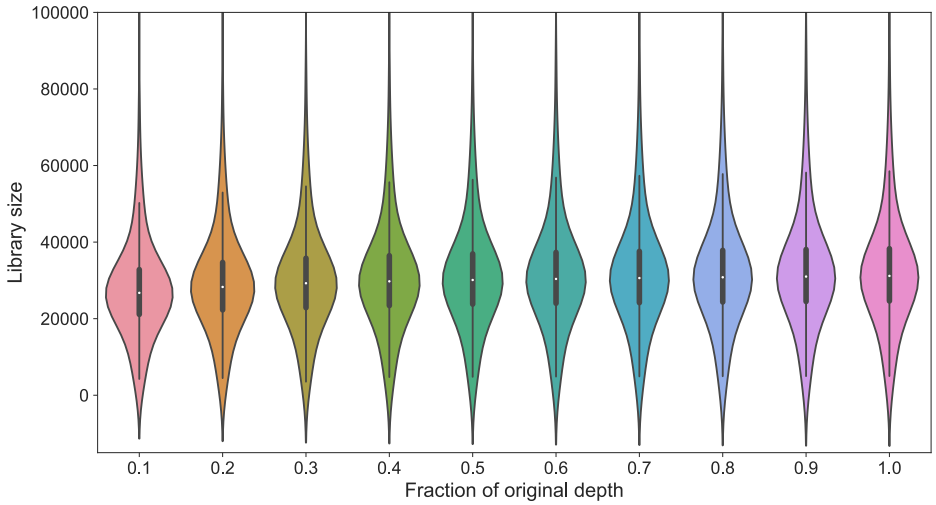


Supplementary Figure 3

a

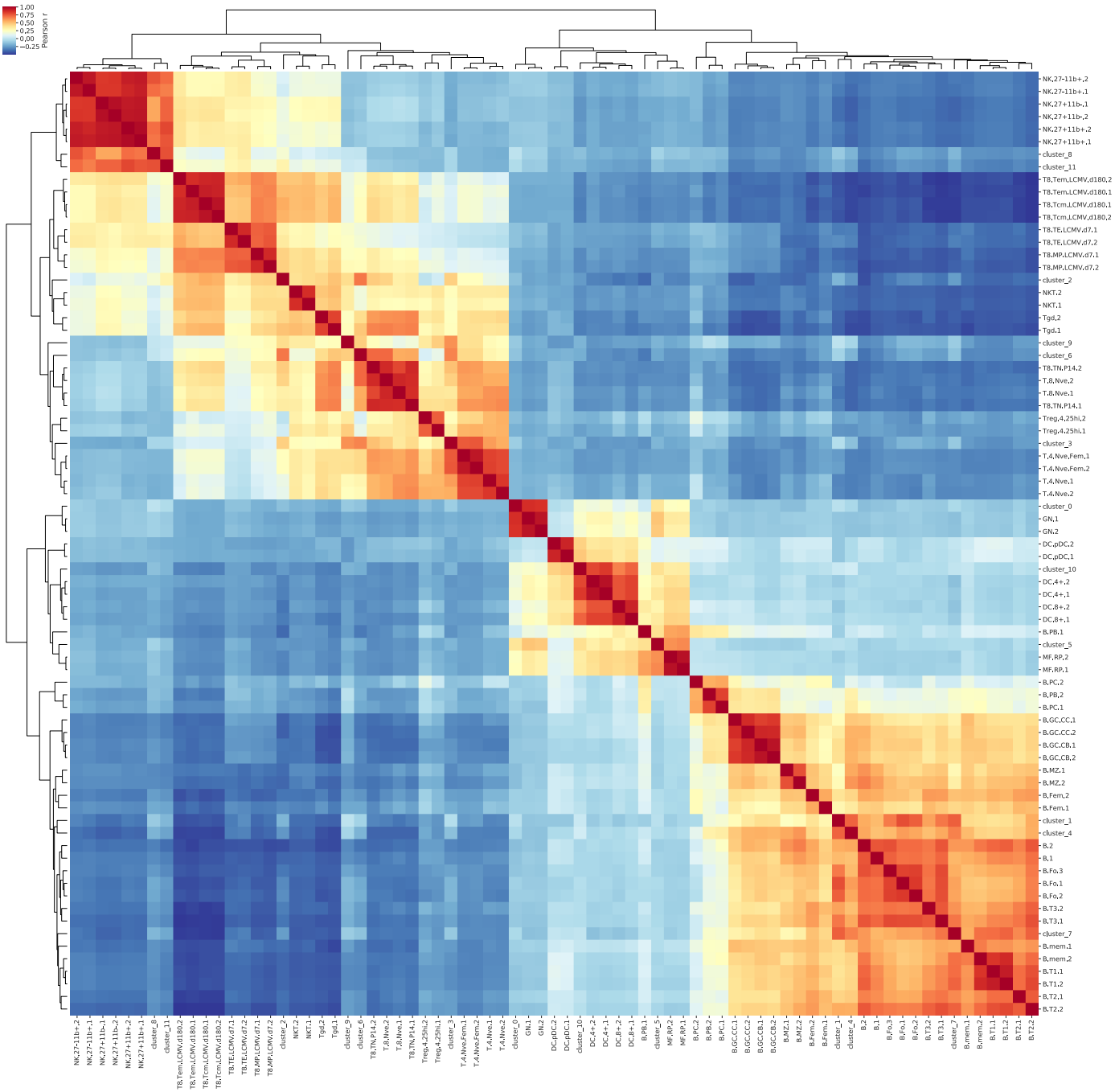


b



Supplementary Figure 4

Supplementary Figure 5



Supplementary Figure 6

cluster_0

```
GN.1       0.858115
GN.2       0.845666
MF.RP.1    0.255730
MF.RP.2    0.244122
DC.4+.1    0.204005
Name: cluster_0, dtype: float64
```

cluster_1

```
B.Fo.1    0.774211
B.Fo.2    0.735329
B.T3.1    0.678556
B.Fo.3    0.672733
B.T3.2    0.575539
Name: cluster_1, dtype: float64
```

cluster_2

```
T8.Tcm.LCMV.d180.1    0.554817
T8.Tcm.LCMV.d180.2    0.538327
T8.Tem.LCMV.d180.1    0.516215
Tgd.1                 0.491312
T8.Tem.LCMV.d180.2    0.478378
Name: cluster_2, dtype: float64
```

cluster_3

```
T.4.Nve.Fem.2    0.750628
T.4.Nve.Fem.1    0.748461
T.4.Nve.1        0.592180
T.4.Nve.2        0.547945
T8.TN.P14.2      0.494832
Name: cluster_3, dtype: float64
```

cluster_4

```
B.MZ.2    0.606645
B.Fo.1    0.592016
B.Fo.2    0.567450
B.2       0.565281
B.MZ.1    0.560363
Name: cluster_4, dtype: float64
```

cluster_5

```
MF.RP.1    0.635129
MF.RP.2    0.616450
GN.1       0.485931
GN.2       0.466141
B.PB.1     0.397456
Name: cluster_5, dtype: float64
```

cluster_6

```
T.8.Nve.2      0.711286
T8.TN.P14.2    0.710398
T8.TN.P14.1    0.563161
T.8.Nve.1      0.552890
Tgd.1          0.477801
Name: cluster_6, dtype: float64
```

cluster_7

```
B.T2.2     0.730364
B.T1.2     0.723927
B.T1.1     0.697820
B.mem.2    0.697162
B.T3.1     0.681272
Name: cluster_7, dtype: float64
```

cluster_8

```
NK.27-11b+.2    0.764921
NK.27-11b+.1    0.702963
NK.27+11b+.2    0.698239
NK.27+11b+.1    0.684591
NK.27+11b-.2    0.568392
Name: cluster_8, dtype: float64
```

cluster_9

```
Treg.4.25hi.1    0.455224
T.4.Nve.Fem.2    0.399113
T.4.Nve.Fem.1    0.390887
Treg.4.25hi.2    0.357982
NKT.2            0.347487
Name: cluster_9, dtype: float64
```

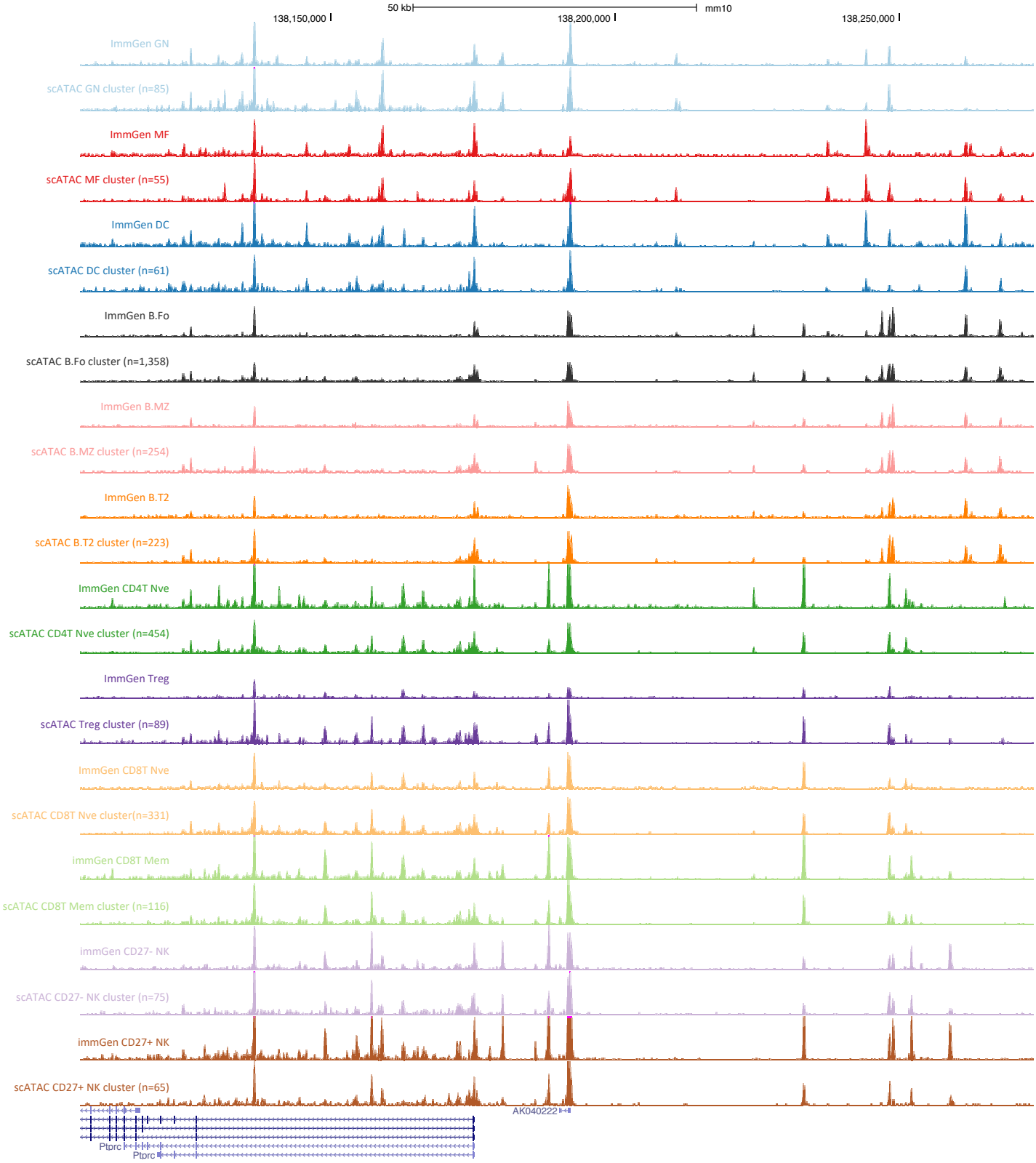
cluster_10

```
DC.4+.1     0.829736
DC.4+.2     0.796127
DC.8+.1     0.725133
DC.8+.2     0.654761
DC.pDC.1    0.470827
Name: cluster_10, dtype: float64
```

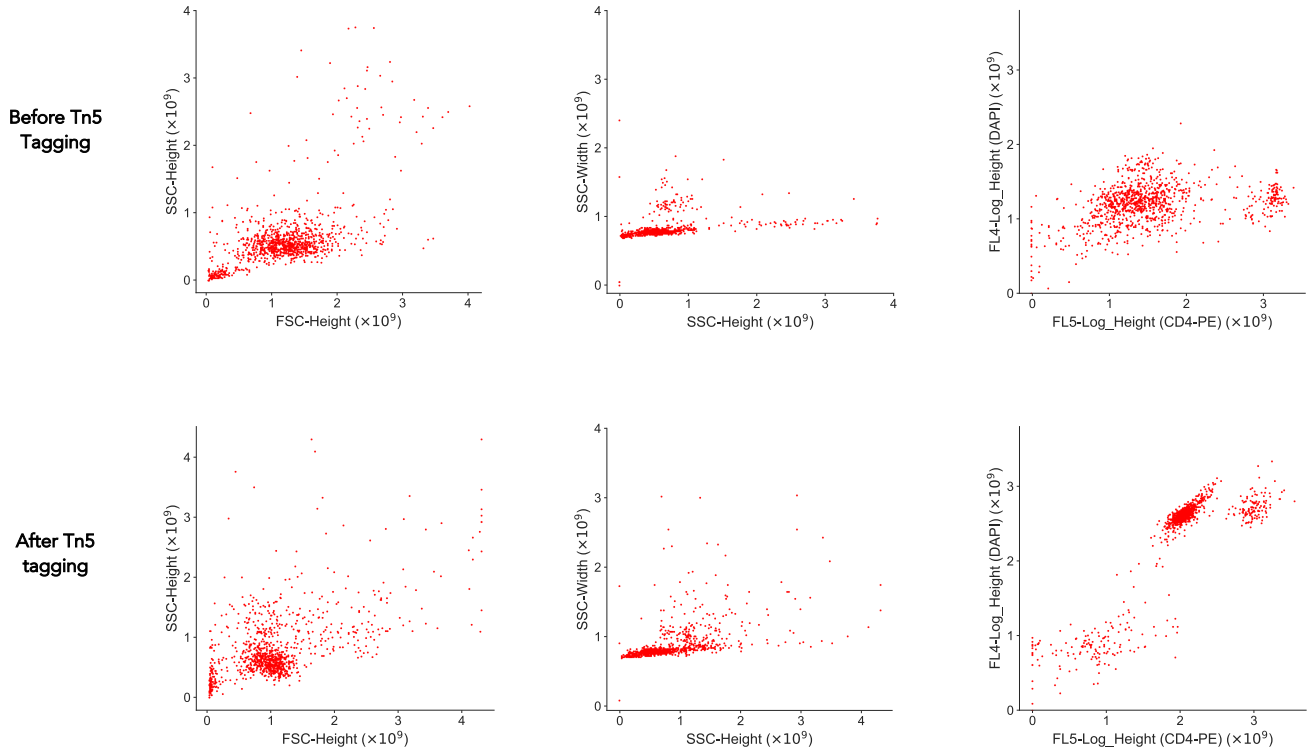
cluster_11

```
NK.27+11b+.2    0.793500
NK.27+11b+.1    0.755976
NK.27+11b-.1    0.721280
NK.27+11b-.2    0.720818
NK.27-11b+.2    0.706145
Name: cluster_11, dtype: float64
```

Supplementary Figure 7

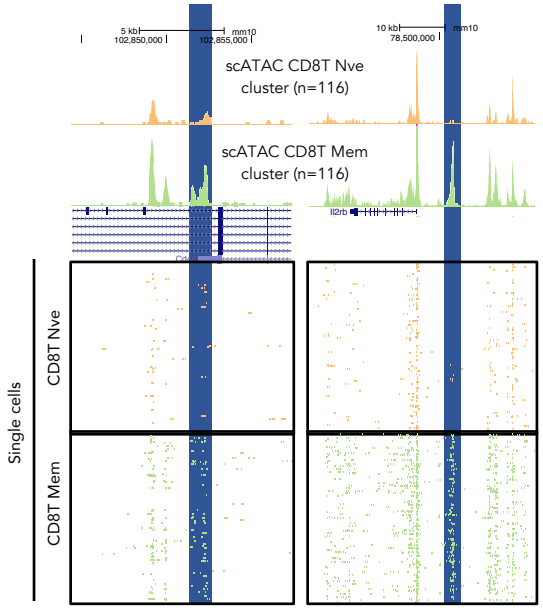


Supplementary Figure 8

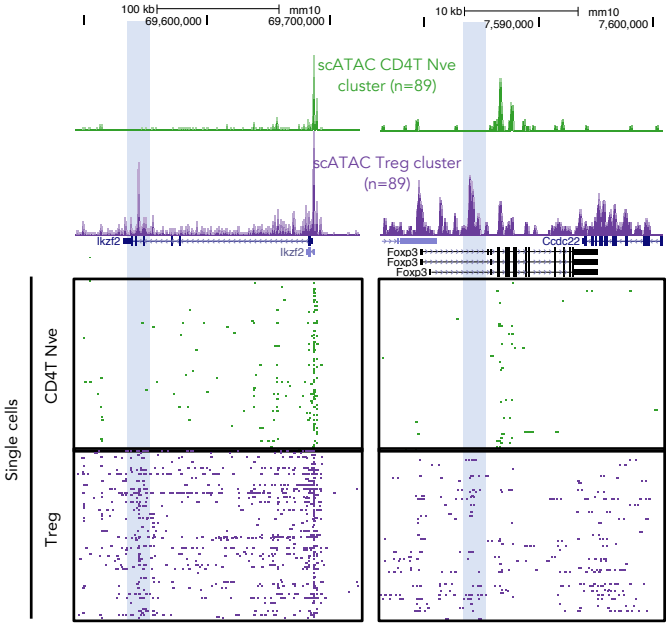


Supplementary Figure 9

a



b



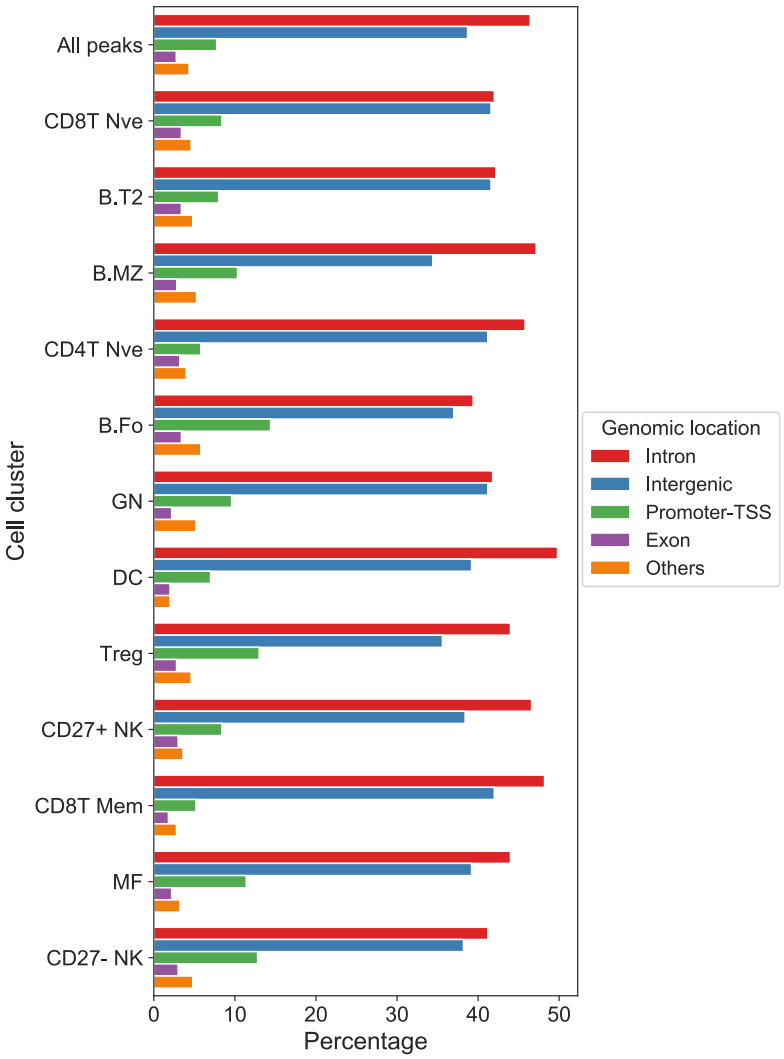
c

ImmGen NK cell RNA-seq sample	*Cd83* expression (tpm)
DC.4+.Sp#1	498.9
DC.4+.Sp#2	712.3
DC.4+.Sp#3	1007.9
NK.27-11b+.Sp#2	0
NK.27+11b+.Sp#1	0
NK.27+11b+.Sp#2	3.5

Supplementary Figure 10

	cluster0	cluster1	cluster2	cluster3	cluster4	cluster5	cluster6	cluster7	cluster8	cluster9	cluster10	cluster11
0	Cd9	Fcer2a	Ptpn12	Myo10	Cdh26	Mafb	Cd8b1	Plxnd1	Lim2	Zdhhc2	Slc46a2	Ermard
1	Lyst	Chil4	Katnb1	Pdlim4	Dtx1	Mgat5b	A630075F10Rik	Psapl1	Kcng3	Nf2	Myo1g	Pofut2
2	Cxcr4	B3gnt7	Ypel1	Trub1	Ackr3	Smad3	5730508B09Rik	Mtx3	Vmn1r234	Spry1	Ctnna1	Magi2
3	Cd300a	Mir7211	S100z	Klhl5	Cd9	Eml6	Cd8a	Akap12	Mir3075	Mir21a	Dusp22	Rftn1
4	Cxcr2	Zfp407	Pigv	Itsn2	AW011738	Mad1l1	Kcnc1	Wnt8b	Gzma	Mpp2	Abcg3	Ccl3
5	Taldo1	Bmp2k	Mir7683	Polr1a	Taok3	Zfyve9	Cd8a	Myl10	9330199G10Rik	Ptpn13	Kit	Iqgap1
6	Map2k3	Xylt1	Gm6981	1700042G15Rik	Smyd2	Parp2	Nkg7	Olfr521	Smarcd3	Tank	Dpp4	Nol4l
7	Cxcl5	Wwox	Il2rb	Siah3	Mirlet7a-1	Hsf2	Cd8b1	Cplx2	Ccl3	Maf	Arhgap26	Gm7550
8	Nfe2	Stpg2	Mylip	Gm11186	Cdh26	Pdgfrb	Itgae	Thoc3	A730036I17Rik	Nr2e3	Macc1	Mir697
9	Tacstd2	1110059E24Rik	Gm13547	1700056E22Rik	Ackr3	Mtl5	Smad7	Thbs4	Lingo1	Tnfsf11	Abi1	Tpp2
10	Fam63a	Tmem252	Runx2	Cd4	Serpine2	Spred1	Haao	Kcnj1	1700063O14Rik	A630076J17Rik	Lonrf2	Frmd4b
11	Cxcr2	Rps15a	Tmevpg1	Gcm2	Thada	Gsdmd	Mir3108	Tnrc18	Snx20	Kbtbd11	Defb25	Tmem171
12	Rplp0	Tmprss13	Cdc42ep1	Gm13986	Dennd2d	4833417C18Rik	Tmed1	Nol4l	Cacna1c	Ctla4	A530013C23Rik	Klhl18
13	Tgif2	Zfp318	Ppp4r2	Epas1	Rgmb	Gm7168	Gosr1	Slc22a12	Bbc3	Acot11	H2-Aa	Bcl11b
14	Taldo1	AI427809	Gpr183	Itgb3	Dtx1	Atg4c	Ric8b	Cep89	Sept5	Axin2	Bloc1s2	Map3k8
15	Pygl	1700016K19Rik	Igsf23	Gm15417	LOC215458	Atp1b3	Orai2	Igf2bp3	Gpx8	B930018H19Rik	Npc1	Lrrc1
16	E2f2	Ciita	Dnaja2	Gcc1	Echdc3	F10	Efcc1	Nt5dc3	Zeb2	Art2b	Tomm20	Mir199a-1
17	Mir7021	Sorl1	Rgs3	Cdhr3	Mzb1	Ak2	2210416O15Rik	Wnt10a	Chpt1	Art2a-ps	Sh3bp4	4921513I03Rik
18	Slc2a3	Shank1	Sycp3	Olfr1510	Cdc42bpb	5430437J10Rik	Hmga2	Cpm	Gm29811	Asxl2	Jak2	Tnfrsf8
19	Ptma	Sacm1l	Gm38403	Gm13582	Gns	4930481A15Rik	Agpat4	Rrm2b	Pisd-ps1	Rgs16	Tbc1d8	Pold2
20	Tacstd2	H2-Aa	Abhd2	St8sia6	Myof	Pld1	Ccdc102a	Zfp608	Fcgr2b	Grb7	Gm13498	Fam131a
21	Fbxo31	Icosl	Cdk5rap3	Adar	Gpr55	AI463170	Cd8a	Cplx2	Chsy1	Rgs1	Jag1	Rasgrp1
22	Cxcr2	Osbpl10	Tmevpg1	Pdlim4	NaN	1700012I11Rik	Slc6a19	1700065J11Rik	Ptpre	Plxnc1	B230217C12Rik	Wisp2
23	Ifitm5	Sergef	Samd3	Mir151	4930581F22Rik	4930552P12Rik	Cd8a	Fads1	1700094M24Rik	Tjp2	Rnf216	Il20rb
24	Ckap4	Nup133	Smndc1	Sugct	Ptpn14	Plcb1	Mta3	Nfatc2	Pla1a	Ctla4	Ffar2	Emb
25	Zbtb16	Stap1	Abhd2	Zfp800	Cxcr4	2900026A02Rik	Hdac7	Igf2bp3	Scrg1	Eea1	Gm4814	Fbxo28
26	Tigd4	Pakap	Galr1	Cd200r3	Ubl3	Atp2b1	Runx3	Adcy9	Pik3r1	Pabpc2	Depdc1b	Gpr21
27	C130050O18Rik	H2-Aa	Drc1	Calcrl	Zfp36l1	Sowahc	Trpm1	Cecr2	Cmklr1	Fam76b	Flt3	Gpr25
28	Csf3r	Gpr137b-ps	Gabrp	Fam105a	Coro2b	Dagla	Nkg7	Man1c1	Gm2176	Lrig1	4833427F10Rik	Chd1
29	Rabac1	H2-Ea-ps	Dynlrb2	Dapk1	Setbp1	A230028O05Rik	Cd226	Gm38404	4930556N09Rik	Khdc1a	Zbtb46	Sacm1l
30	Nlrp12	Ppdpf	Kdm4d	4933406K04Rik	Blk	Pparg	Cd8b1	Bhlhe41	B230217C12Rik	Msl3l2	Acvr2a	Gsap
31	Emb	Cenpe	Txk	Tgfbr3	Mir1941	Nfkbiz	Ldlrad1	4930515G16Rik	Cox4i2	Mamstr	Mir1231	Fxyd3
32	Myh9	Icosl	Agtr1b	Slc19a3	Dtx1	Vcan	Gm29687	4930515G16Rik	Hgsnat	Nek7	Ddr1	Gpr25
33	Pgd	Enpp6	Cmc1	Myo10	Akap5	Gm4262	Col6a1	Cplx2	Pisd-ps3	Zfp41	Myo1h	Gdf1
34	Zeb2	Pxdc1	Mir7057	B4galt5	Sirpa	Xirp1	Wipf2	9430020K01Rik	Fastkd1	Tbc1d4	Zfp800	A630001G21Rik
35	Gcnt2	Capzb	Fosl2	Pno1	Prkcz	Ccr3	Cd8b1	Acaca	1700018C11Rik	Myo3b	Itpr1	Hemgn
36	Itgb2	H2-Eb1	Parp8	Epas1	Gm15713	Pola2	Cxcr4	Ldoc1l	Jam3	Gm10560	Serpina4-ps1	Gm5547
37	Susd1	Stoml1	Serpina12	Msra	Cmah	Galnt9	Irgc1	Tbc1d7	Kcnk13	Comt	Kif16b	Immp2l
38	Grina	Rgs9	Olfr525	Unc80	Kmt2a	Cmklr1	Slc6a19os	Bcl2l1	Efhd2	Ndfip1	Ccnd1	Gm7443
39	Mrpl33	Scd1	Gimap4	Ibtk	Hmgn3	Pld4	Trnp1	Gpr25	Adamts14	Gpr15	Gm9733	Nt5dc3
40	Dmxl2	Ptpn11	Fyn	Chl1	Atxn1	Msr1	Egfl7	Pik3c2b	Irf8	Suco	Grk3	Bcl2l10
41	Reep3	B3gat2	Btbd11	Chdh	Tsga13	Gsap	Cd8a	Gm12159	Id2	Lrrc32	Mefv	Rftn1
42	Cxcr1	Ezr	Runx2	Mtx2	Plac8	Commd9	Mir467h	2310061N02Rik	Nup50	Gm11985	Reps1	P2rx3
43	Slc11a1	Rasgef1b	Ccl5	Tnni1	Atxn1	4933433H22Rik	Ss18	Paqr6	Rps4l	Fxr1	Rrad	Rpl38
44	Slmap	9030404E10Rik	Gm38403	Cck	Syk	Arhgef10l	Sos1	Guca2b	Sytl2	Aven	Rgs2	Mir7235
45	Csf3r	Helz2	Nosip	Zscan10	Cbx4	Pitpna	Ccdc102a	Dcun1d1	St5	1700016G22Rik	Frmd5	Phf3
46	Arntl2	Chrna9	Runx2	Fam65b	Fasl	Tm9sf4	Il21r	Prkcg	Znrf2	Ptger2	Cdyl2	Gm7008
47	Ndel1	Parp8	Kcnj8	Cers6	Rps24	Selenbp1	Bfsp2	Gm17455	Nav2	Slc25a19	Tbc1d4	Serpina3f
48	Ptafr	March1	Mapre2	Kif23	Slc31a2	Zfp397	Cst7	Gopc	Clnk	Ets1	Mreg	Manba
49	Map7	Akap13	Fam169b	Rab11fip4	Dusp16	Agap1	Ap1ar	Myb	Gm20750	Sdcbp2	BC039771	Erc2

Supplementary Figure 11



Supplementary Figure 12

Supplementary Protocol

Protocol for plate-based scATAC-seq using FACS
Timestamp: 15-Feb-2018

1. One day before the experiment, prepare the plates by aliquoting 2 µl 2X Lysis Buffer to each well of the plates (either 96-well or 384-well plate). Then add 2 µl of 10 µM S5xx/N7xx Nextera Index Primer Mix (5 µM each) to each well. Seal the plate and store in -80 °C.

   Recipe for 2X Lysis Buffer:

100 mM	Tris.HCl, pH 8.0
100 mM	NaCl
40 µg/ml	Proteinase K (Ambion, AM2546, 20 mg/ml stock)
0.4%	SDS

2. On the day of the experiment, thaw plates at room temperature.
3. Pre-coat all tubes with 500 µl 0.5% BSA (prepared in 1X PBS) for a few minutes to reduce sample loss. Count or sort 5k - 50k cells into 1.5-ml eppendorf tubes. DO NOT use DNA LoBind tubes for pelleting cells, which does not work well especially when cell numbers are limited.
4. Pellet cells at 500 g, 4 °C, 5 minutes.
5. Wash the cell pellet with 100 µl ice-cold PBS, twice, 500 g, 4 °C, 5 minutes, and carefully remove the supernatant.
6. Resuspend the cell pellet in 50 µl tagmentation mix. The recipe for the tagmentation mix is as follows (THS-seq recipe):

12.5 µl	4X THS-seq TD buffer
5 µl	10X Digitonin
27. 5 µl	H_2O
5 µl	Illumina Tn5 (Nextera kit, Illumina Cat No. FC-121-1030)

Recipe for 4X THS-seq TD buffer:

132 mM	Tris-acetate, pH 7.8
264 mM	Potassium acetate
40 mM	Magnesium acetate
64%	Dimethylformamide (DMF)

Recipe for 10X Digitonin:

1 µl	Digitonin (Promega, G9441, 2% stock)
19 µl	H_2O

7. Put the tagmentation reaction (50 µl) on a thermomixer, 37 °C, 800 rpm, 30 minutes.
8. Stop the reaction by adding 50 µl tagmentation stop buffer (TSB). Recipe for TSB:

10 mM	Tris-HCl, pH 8.0
20 mM	EDTA, pH 8.0

9. Leave on ice for 10 minutes.
10. Add 100 - 300 µl PBS/0.5% BSA to the 100 µl stopped tagmentation mix, and transfer to a FACS tube.
11. Optional: add DAPI to stain nuclei based on manufacturer's instruction.
12. Sort DAPI positive single nuclei into the plates prepared the day before.
13. Quickly spin down and seal the plate well (can be stored in -80 °C for a few weeks from here), and put the plate on a PCR machine, with lid temperature set to 100 °C.
14. Incubate the plate at 65 °C for 15 minutes to perform Tn5 release.
15. Add equal volume (4 µl) of 10% TWEEN-20 to each well to quench SDS. Briefly vortex to mix.
16. Add 2 µl H_2O to each well.
17. Add 10 µl 2X NEBNext® High-Fidelity 2X PCR Master Mix (NEB M0541L) to each well

18. At this stage, each well contains 20 µl PCR reaction.
19. Perform library amplification PCR:

    72 °C 10 minutes

    98 °C 5 minutes

    [98 °C 10 seconds, 63 °C 30 seconds, 72 °C 20 seconds] x 18

    10 °C hold
20. Combine all reactions into a 50-ml falcon, which yields about 20 µl x 384 = 7.68 ml. Normally, the yield will be ~ 7.2 ml.
21. Add 5 volumes (~ 36 ml) Buffer PB (Qiagen), mix well, and pass reaction volume through a single column from a Qiagen MinElute PCR Purification Kit by connecting the column to a vacuum.
22. To wash the column, pass through 40 ml Column Wash Buffer (10 mM Tris-HCl, pH 7.5, 80% ethanol).
23. Spin down the column at top speed on a table top centrifuge to remove all traces of ethanol, and remember to use a pipette to remove the ethanol leftover on the rim of the Qiagen column.
24. Elute the library in 12.5 µl Buffer EB. Perform the elution three times and combine the three elutes to a final volume of ~ 36 µl.
25. Do a final fragment size selection using 0.5X SPRI upper cutoff, followed by 1.2X SPRI lower cutoff, and elute in 30 µl 10 mM Tris-HCl, pH 8.0.
26. Run Nanodrop to obtain a rough estimate of the concentration, and then dilute the library to a range suitable for Bioanalyzer/TapeStation etc.
27. Check for expected results (see **Supplementary Fig. 2a**).
28. Sequencing: we sequenced each 384 pool on one lane of Hiseq 2000 or one rapid run of Hiseq 2500, which nearly saturated the library. From the data obtained, each cell was sequenced to about 1 million reads, but only ~30,000 unique reads were obtained per cell. Further reads were redundant, which is comparable (if not better) to published scATAC-seq by other methods. Theoretically, 30,000 reads per cell should be sufficient to profile the unique reads. However, considering the presence of mitochondrial DNA, non-mapped and non-uniquely mapped reads, it is safer to aim for at least 100,000 reads per cell.

Oligonucleotides sequence

N701 CAAGCAGAAGACGGCATACGAGATTCGCCTTAGTCTCGTGGGCTCGG

N702 CAAGCAGAAGACGGCATACGAGATCTAGTACGGTCTCGTGGGCTCGG

N703 CAAGCAGAAGACGGCATACGAGATTTCTGCCTGTCTCGTGGGCTCGG

N704 CAAGCAGAAGACGGCATACGAGATGCTCAGGAGTCTCGTGGGCTCGG

N705 CAAGCAGAAGACGGCATACGAGATAGGAGTCCGTCTCGTGGGCTCGG

N706 CAAGCAGAAGACGGCATACGAGATCATGCCTAGTCTCGTGGGCTCGG

N707 CAAGCAGAAGACGGCATACGAGATGTAGAGAGGTCTCGTGGGCTCGG

N710 CAAGCAGAAGACGGCATACGAGATCAGCCTCGGTCTCGTGGGCTCGG

N711 CAAGCAGAAGACGGCATACGAGATTGCCTCTTGTCTCGTGGGCTCGG

N712 CAAGCAGAAGACGGCATACGAGATTCCTCTACGTCTCGTGGGCTCGG

N714 CAAGCAGAAGACGGCATACGAGATTCATGAGCGTCTCGTGGGCTCGG

N715 CAAGCAGAAGACGGCATACGAGATCCTGAGATGTCTCGTGGGCTCGG

N716 CAAGCAGAAGACGGCATACGAGATTAGCGAGTGTCTCGTGGGCTCGG

N718 CAAGCAGAAGACGGCATACGAGATGTAGCTCCGTCTCGTGGGCTCGG

N719 CAAGCAGAAGACGGCATACGAGATTACTACGCGTCTCGTGGGCTCGG

N720 CAAGCAGAAGACGGCATACGAGATAGGCTCCGGTCTCGTGGGCTCGG

N721 CAAGCAGAAGACGGCATACGAGATGCAGCGTAGTCTCGTGGGCTCGG

N722 CAAGCAGAAGACGGCATACGAGATCTGCGCATGTCTCGTGGGCTCGG

N723 CAAGCAGAAGACGGCATACGAGATGAGCGCTAGTCTCGTGGGCTCGG

N724 CAAGCAGAAGACGGCATACGAGATCGCTCAGTGTCTCGTGGGCTCGG

N726 CAAGCAGAAGACGGCATACGAGATGTCTTAGGGTCTCGTGGGCTCGG

N727 CAAGCAGAAGACGGCATACGAGATACTGATCGGTCTCGTGGGCTCGG

N728 CAAGCAGAAGACGGCATACGAGATTAGCTGCAGTCTCGTGGGCTCGG

N729 CAAGCAGAAGACGGCATACGAGATGACGTCGAGTCTCGTGGGCTCGG

S502 AATGATACGGCGACCACCGAGATCTACACCTCTCTATTCGTCGGCAGCGTC

S503 AATGATACGGCGACCACCGAGATCTACACTATCCTCTTCGTCGGCAGCGTC

S505 AATGATACGGCGACCACCGAGATCTACACGTAAGGAGTCGTCGGCAGCGTC

S506 AATGATACGGCGACCACCGAGATCTACACACTGCATATCGTCGGCAGCGTC

S507 AATGATACGGCGACCACCGAGATCTACACAAGGAGTATCGTCGGCAGCGTC

S508 AATGATACGGCGACCACCGAGATCTACACCTAAGCCTTCGTCGGCAGCGTC

S510 AATGATACGGCGACCACCGAGATCTACACCGTCTAATTCGTCGGCAGCGTC

S511 AATGATACGGCGACCACCGAGATCTACACTCTCTCCGTCGTCGGCAGCGTC

S513 AATGATACGGCGACCACCGAGATCTACACTCGACTAGTCGTCGGCAGCGTC

S515 AATGATACGGCGACCACCGAGATCTACACTTCTAGCTTCGTCGGCAGCGTC

S516 AATGATACGGCGACCACCGAGATCTACACCCTAGAGTTCGTCGGCAGCGTC

S517 AATGATACGGCGACCACCGAGATCTACACGCGTAAGATCGTCGGCAGCGTC

S518 AATGATACGGCGACCACCGAGATCTACACCTATTAAGTCGTCGGCAGCGTC

S520 AATGATACGGCGACCACCGAGATCTACACAAGGCTATTCGTCGGCAGCGTC

S521 AATGATACGGCGACCACCGAGATCTACACGAGCCTTATCGTCGGCAGCGTC

S522 AATGATACGGCGACCACCGAGATCTACACTTATGCGATCGTCGGCAGCGTC