bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Cellular Adaptive Response and Regulation of HIF after Low Dose Gamma-radiation Exposure

Nitin Motilal Gandhi *

Radiation Biology & Health Sciences Division,

Bhabha Atomic Research Centre, Trombay Mumbai 400 085

India

Email: nmgandhi@barc.gov.in

Key words: LNT; HIF; Low Dose; Adaptive response; comet assay; MTT assay;

*Corresponding author

Nitin Motilal Gandhi

Radiation Biology and Health Sciences Division

Bhabha Atomic Research Centre, Trombay, Mumbai 400 085

India Tel: +91-022-25590409

E mail: nmgandhi@barc.gov.in

orcid.org/0000-0003-0581-5869

Conflict of Interest: None

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Abstract

Purpose: Cellular damage due to low dose of γ-radiation (≤0.1 Gy) is generally extrapolated from observing the effects at higher doses. These estimations are not accurate. This has led to uncertainties while assessing the radiation risk factors at low doses. Although there are reports on the radiation induced adaptive response, the mechanism of action is not fully elucidated, leading to the uncertainties. One of the outcomes of low dose radiation exposure is believed to be adaptive response. The mechanism of adaptive response is not fully understood. Therefore, the study was undertaken to understand the role of hypoxia inducible factor (HIF) on radiation induced adaptive response.

Materials and methods: MCF-7 cells pre-exposed to low dose γ-radiation (0.1 Gy; Priming dose) were exposed to 2 Gy (challenging dose) 8 hrs after the priming dose and studied for the adaptive response. Cell death was measured by MTT assay, and apoptosis was measured by FACS analysis. DNA damage was measures by alkaline comet assay. HIF transcription activity was assayed using transiently transfected plasmid having HIF consensus sequence and luciferase as the reporter gene.

Results: Cells when exposed to 0.1 Gy priming dose 8 hrs prior to the higher dose (2 Gy; Challenging dose) results in lower amount of radiation induced damages compared to the cells exposed to 2 Gy alone. Cobalt chloride treatment in place of priming dose also results in the protection to cells when exposed to challenging dose. There was up-regulation of HIF activity when cells were exposed to priming dose, indicating the role of HIF in radiation induced response.

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Conclusion: Results indicate the γ-radiation induced adaptive response. One of the mechanism proposed is up-regulation of HIF after low dose exposure, which protects the cells from damages when they are exposed to challenging dose of 2 Gy radiation dose.

.

Introduction

The widespread use of medical diagnostic imaging and the fear it has generated has highlighted the need to determine the effects of radiation at low doses (≤ 0.1 Gy). There are uncertainties while measuring the biological effects of radiation in this dose range. (Shore et al. 2017) It is widely accepted that there is no threshold for damaging effect of radiation on living organisms. This is termed as "Linear-no-threshold" (LNT) model. Most of the radiological guidelines are based on this model. There are alternate models put forward like threshold model and adaptive response (Feinendegen et al. 2013; Mullenders et al. 2009). The conundrum is not resolved as all the three models are supported in various studies (Morgan and Bair 2013). These uncertainties in studying the radiobiological effects at low doses are due to the difficulties in assaying radiological damages at low doses accurately. Most of the current knowledge on effects of low dose is based on the atomic bomb survivors by measuring the increase in cancer incidence compared to control population. However, there are several factors which can lead to skewed estimates in population exposed to low dose radiation (Little et al. 2014; Sasaki et al. 2014), as the carcinogenesis being a multi-model disease, it is difficult to estimate the increase in cancer incidence assigned to any one parameter. The use of LNT model results in the stringent conditions to determine the permissible levels of radiation exposure.

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

It is well known that ionizing radiation damages cellular system through generation of free radicals by breakdown of the water molecule, which is abundant in the cell. These free radicals when scavenged in the cells generate reactive oxygen species (ROS). ROS under these conditions functions as an important physiological regulator of HIF (Finkle T 2012) HIF is a transcription factor which is a heterodimeric protein composed of HIF-1(α) and HIF-1(β) subunit. HIF-1(α) is constantly under degradation pathway in response to the oxygen levels; HIF-1(β) is constitutively expressed. HIF binds to HRE (hypoxia response elements) on the consensus sequence of DNA and regulates the corresponding gene. In the presence of oxygen, the proline residue at 402 and 564 of HIF-1(α) is hydroxylated by prolyl-hydroylase domain (PHD), the hydroxylated protein gets attached to von Hippel-Lindau protein, and is broken down through ubquitination and ptoteosomal degradation. HIF activation is known to provide the increased survival in the cells exposed to ionizing radiation. (Semenza 2012a)

HIF can also be experimentally stabilized through the pharmacological intervention. The function of PHD requires α-ketoglutarate as co-substrate and ferrous ions in their catalytic site. Therefore, iron chelator like desferrioxamine (Woo et al. 2003), and $CoCl_2$ can be used to stabilize HIF under normoxic conditions (Yuan et al. 2003)

The work reported here describes the adaptive response in the cells which pre-exposed to the low dose radiation prior to the challenging dose compared to the radiation damage in the cells exposed to the challenging dose alone. One of the mechanism of adaptive response is proposed is due to the activation of HIF on γ-radiation exposure. This stems due to the observation that the $CoCl_2$ treatment in place of priming dose can also results in reducing the cellular toxicity by challenging dose.

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Materials and Methods

Chemicals

Dulbecco's Modified Eagle Medium (DMEM), Fetal Calf Serum, antibiotic solution, trypsin-EDTA was from Hi media, Mumbai India. Anti-HIF antibody, Annexin V/Pi kit was from BD bioscience. Propidium iodide, Radioimmunoprecipitation (RIPA) buffer, was obtained from Sigma-Aldrich Corp. (St. Louis MO). Lipofectamine3000 was from Invitrogen. Other chemicals used were of analytical grade obtained from local suppliers.

Cell culture

Human breast cancer cell line MCF-7 was grown in vitro in DMEM with 10% v/v fetal bovine serum. 50 units/ml penicillin and 50 µg/ml streptomycin were added to the media. The cells were grown in CO_2 incubator set at 5% CO_2 and 95% air, at 37 ºC with relative humidity greater than 85%.

Radiation exposure

The cells were subjected to γ-irradiation at room temperature (dose rate: 1 Gy/min) using ^{60}Co gamma teletherapy machine (Bhabhatron-II, Panacea Medical Technologies, Bangalore, India).

Cell viability assays

To determine the toxicity of γ-radiation exposure on MCF-7 cells viability assay using MTT assay were conducted. CCK-8 reagent (Sigma) was used to assay the viability of cells treated with $CoCl_2$.

Single cell gel electrophoresis (Comet assay)

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

The basic alkaline technique described by Singh et al. (2000) was followed with some modifications (Gade and Gandhi 2015) Cell suspension containing 10,000 cells were mixed with 200 µl of 0.7% low melting agarose at 37 ºC in a microfuge tube and spread on a fully frosted microscopic slide. The slide was covered with a cover slip and left on ice-cold surface for 2 min. After gelling, cover slip was gently removed. The cells were lysed by dipping the slides into lysing solution (100 mM Na-EDTA, 10 mM Tris, 2.5 M NaCl, 1% Triton X-100 and 10 % DMSO, pH 10) for 1 hr at 4 ºC. The slides were rinsed free of salt and detergent in a buffer (1 mM Na-EDTA, 300 mM NaOH, pH>13) and subsequently submerged in a horizontal gel-electrophoresis apparatus by adding fresh buffer, and left in the buffer for 20 min to allow the unwinding of the DNA and the expression of alkali labile damage. Then, an electric field was applied (300 mA; 25 mV) for 20 min to draw negatively charged DNA towards the anode. After electrophoresis, the slides were washed twice for 5 min in a neutralizing buffer (0.4 M Tris, pH 7.4) and stained with 75 µl of propidium iodide (20 µg/ml), and visualized using a fluorescent microscope (Carl Ziess Axioskop) attached with high-performance Carl-Zeiss camera. Fifty cells/slide were captured. The quantification of the DNA strand breaks of images was done using the CASP software (Konca et al. 2000) by which Olive tail moment (OTM) was obtained. OTM is measure of DNA damage which incorporates the percent DNA in tail and tail length.

Apoptosis assay

Annexin V-FITC/PI dual staining was used for estimating apoptotic cells. Cells were labelled with 5 µl of Annexin V-FITC and 2 µl of propidium iodide (PI, 100 µg/ml) using apoptosis detection kit (BD pharmingen) as per the manufacturer's protocol. Within one hour the stained

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

cells were acquired in a Partec CyFlow Space™ flow cytometer. Percentage apoptotic cells are determined from the scatter plots using FloJo software.

Cell-based reporter assay

Cell-based luciferase assay was used to study the transcription activity of HIF. A luciferase reporter gene under the control of hypoxia response elements from the erythropoietin gene (pTK-HRE3-luc; obtained from Prof. S. McKnight, University of Texas, USA) was employed to monitor the HIF activity (Tian et al. 1997) MCF-7 cells were transiently transfected using lipofectamine3000 (Invitrogen) following manufacturer's protocol by the plasmid and then were exposed to the γ-radiation in different conditions as shown in figure legend. The transfection was continued for 32 hrs, at that time almost 30-50% transfection efficiency. The cells where then exposed to the radiation at various doses. Cells were then harvested, counted in haemocytometer and then lysed with RIPA buffer containing protease inhibitors; snap-frozen under liquid nitrogen and stored at -80 ºC. Before analysis the cells were thawed on ice centrifuged at 10,000 g for 5 minutes and the supernatant was analyzed for luciferase enzyme using Sigma-Aldrich (Luc-1) kit following manufacturer's protocol. The luciferase assay was normalized by equal number of cells taken for assay. The detailed experimental protocol followed is given in individual figure legend.

Statistical analysis

All data are expressed as the mean ± SEM from three independent experiments. Statistical analyses were performed by the Student's *t* test, and significance was assumed if $P<0.05$. Statistical significance was measured using Mann-Whitney test while assuming the significance in comet assay results.

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Results

Optimizing the concentration of Cobalt chloride treatment

Cobalt chloride treatment is known to stabilize HIF-1(α) under normoxic conditions. In order to select the nontoxic but an effective concentration of $CoCl_2$; toxicity of $CoCl_2$ was determined by exposing the MCF-7 cells under various concentrations of $CoCl_2$ for 48 h (Figure 1). CCK-8 assay results indicate the dose dependent toxicity of $CoCl_2$ on the viability of MCF-7 cells. At 100 µM and 200 µM there is 8.72% and 17.4% of the toxicity to the cells respectively. Stabilization of HIF after various doses of $CoCl_2$ was observed, by western blot analysis (results not shown) In subsequent studies, the concentration 100 µM and 200 µM of $CoCl_2$ was used as indicated in the individual figure legend.

Adaptive response using Cell viability (MTT) assay

MCF-7 cells grown in 96 well culture plates until a confluence of approximately 70%. The different groups were treated as indicated in the Figure 1 in triplicate. After the incubation for 48 h MTT assay was performed. OD570 of control cells were taken as 100 % and then the absorbance of the other groups was factored. Priming dose alone (0.1 Gy) resulted in slight but statistically significant increase in proliferation. Results depicted in Figure 2 indicate that cells when exposed to 2 Gy γ-radiation alone resulted in significantly higher decrease in viability. Priming dose of 0.1 Gy 8 h prior to the challenging dose of 2 Gy rescued the cells from challenging dose induced decrease in viability. $CoCl_2$ treatment in place of priming dose also resulted in the decrease in damage when challenged with 2 Gy γ-radiation.

Measurement of DNA damage using alkaline comet assay

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

MCF-7 cells grown in 6 well plates were treated with various conditions in triplicate as described in the legend of Figure 3. Immediately after the challenging dose of 2 Gy of γ-radiation, cells were harvested; aliquot of cells (approx.10000 cells) were taken into low melting agarose and subjected to alkaline comet assay. Results indicate that cells when exposed to 2 Gy γ-radiation alone resulted in a significantly more DNA damage as measured as OTM (Fig. 3) Priming dose of 0.1 Gy 8 h prior to the challenging dose of 2 Gy rescued the cells from undergoing the challenging dose induced DNA damages. $CoCl_2$ treatment in place of priming dose also resulted in the decrease in 2 Gy γ-radiation induced DNA damages.

Measurement of γ-radiation induced Apoptosis

MCF-7 cells were grown in six well culture plates overnight till a confluence of approximately 70% was reached. Different groups were treated in triplicate; after incubation for 48 h cells were harvested, stained by Annexin V/PI and analyzed by FACS. Results indicate that cells when exposed to 2 Gy γ-radiation alone resulted in a significantly higher percentage of apoptosis after 48 h. compared to sham irradiated group. Priming dose of 0.1 Gy 8 h prior to the challenging dose of 2 Gy rescued the cells from undergoing the challenging dose induced apoptosis. (Figure 4) $CoCl_2$ treatment in place of priming dose also resulted in the decrease in radiation induced percent apoptosis when exposed to 2 Gy challenging dose. There was no change in percent apoptosis in cells exposed to either 0.1 Gy radiation or cobalt chloride treatment alone (results not shown)

Modulation of γ-radiation induced HIF transcription activity:

To increase the sensitivity to measure the HIF regulation at lower dose, transiently transfected MCF-7 cells with p-HRE3-tk-luc were exposed to various doses of γ-radiation. The plasmid

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

contains luciferase gene under control of HIF consensus signal. Therefore, the luciferase signal is directly proportional to the transcription activity of HIF. After 8 h the cells were lysed and analyzed for luciferase activity. There was dose dependent increase in luciferase signal, there was significantly increase in luciferase activity at 0.1 Gy compared to un-irradiated control. The signal saturated at 0.5 Gy as there was no statistically significant increase in luciferase activity between 0.5 Gy and 2 Gy. $CoCl_2$ treatment alone (used as positive control) gave very strong increase in luciferase activity (Figure 5).

Discussion

In the present study, the adaptive response in MCF-7 cells is described. Viability, apoptosis and DNA damage outcomes have suggested that there is a significant reduction of the radiation induced damages to cells pre-exposed to 0.1 Gy γ-radiation. Further, the results also suggest that when 0.1 Gy priming dose was replaced with $CoCl_2$ treatment resulted in reduction of damages after the challenging dose.

The chemical toxicity of Cobalt chloride treatment is minimal as shown in Figure 1. However, there is statistically significant adaptive response when priming dose if replaced with Cobalt chloride treatment. The toxicity induced by cobalt chloride could not be attributable to the adaptive response as any chemical toxicity induced adaptive response induction takes longer time duration (if at all) cobalt chloride treatment is given for 8 h, time not sufficient to make the global effect of chemical stress induced adaptive response. These results indicate that $CoCl_2$ treatment (and thus HIF stabilization) can mimic the effects of priming dose. This finding was corroborated as when transiently transfected cells with p-HRE-tk-luc plasmid was treated with different radiation doses; there was dose dependent increase in HIF transcription activity.

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Further, at 0.1 Gy there was significant increase in transcription activity of HIF indicating the activation of HIF. In spite of several reports which describe the γ-radiation induced adaptive response in cells; these observations could not translate to the reduction of stringent radiological guidelines. One of the reasons may be due to the lack of understanding in molecular mechanism which can explain the adaptive response. Lall et al. (2014) have shown the adaptive response in human fibroblasts is due to the increased in glucose metabolism. Their results also suggest that at higher doses of γ-radiation p53 gets activated which in turn shuts off the HIF stabilization, contrary to findings reported here, the difference of observations is assumed to stem from the cell type.

There are reports on studies on adaptive response in MCF-7 cells (Min et al. 2005; and Suwimol et al. 2008) where authors have shown the activation of MnSOD as one of the mechanism of adaptive response. Alexander et al. (2003) have demonstrated that NF-κB regulates the expression of MnSOD, which in turn increases the expression of genes that participate in radiation-induced adaptive response in MCF-7 cells.

Zhou et al. (2013) have shown that the combination of radiation and hypoxia could promote the glucose metabolism. Further, HIF-1(α) might inhibit the promoting effect of radiation on glycolysis in hypoxic MCF-7 cells by regulating the glucose metabolism. MCF-7 has wild type p53 status. There is the direct correlation with p53 and HIF therefore it was essential to study the radiation induced modulation of HIF in MCF-7 cells (Murley et al. 2017).

One of the observation that exposing the cells, when exposed to 0.1 Gy alone resulted in statistically significant increase in cell proliferation compare to non-irradiated control cells. This could be additional evidence that ionizing radiation can enhance the proliferation of cells and

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

promote long term resistance to multiple cytotoxic stresses (Anupama and Rajgopal 2013) Ionizing radiation exposure cause the disruption in the cellular functions or cause death to the cells depending on the dose of radiation. Ionizing radiation damages the cellular functions by generating free radicals and ROS through breakdown of water molecule. Radiation exposure results in cellular damages to DNA, protein and lipids. Those damages result in changes in biochemical and molecular signalling in the surviving fraction of cells (Azzam et al. 2011). One of the factor that get modulate is HIF.HIF is primarily an oxygen sensing transcription factor, under the continuous degradation pathway in the presence of oxygen; however, under the hypoxic conditions HIF is stabilized and regulates almost 800 genes (Semenza 2012b) It is also known that hypoxia is not the only cause of stabilization of HIF. Even in the presence of oxygen HIF can get stabilize through the effect of decrease in Fe2+ (Bianchi et al. 1999) or the presence of intermediate of TCA cycle like alpha-ketoglutarate (Pan et al 2007) The significant increase in HIF transcription activity at 0.1 Gy radiation was observed. As exposure to γ-radiation results in activation of MAP kinase pathway through EGFR signalling (Michael et al 2000) Further, Mylonis et al. (2006) have shown that MAPK-dependent phosphorylation of HIF is required for its efficient accumulation inside the nucleus. Taken together this could explain the increase in the transcription activity due to enhance in nuclear translocation of HIF-1(α).

The background radiation dose was greater than eight times when life was evolved, compared to the present day (Karam and Leslie 1999). Which could have resulted in evolution of efficient DNA repair system, which is evolutionary conserved till the present day this could help the humans to repair the low dose of radiation induced damages efficiently. Therefore, we have to review the notion that even a very low dose of radiation is harmful to the life. Thus, the debate regarding LNT model needs to be reviewed.

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Taken together, based on the findings presented here, it is proposed that the low dose of γ-radiation results in the increased transcription activity of HIF and subsequent activation of survival pathways. Therefore, when the cells were primed with low doses of radiation, it resulted in up-regulation of survival machinery protecting the cells when they were challenged with higher doses of radiation.

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

References

Alexander K, Zhongkui L, Christine W, Jeffrey AL, Ting-Ting H, Douglas RS, Larry WO,and Jian JL. 2003. Manganese Superoxide Dismutase-Mediated Gene Expression in Radiation-Induced Adaptive Responses Mol. Cellular Biol. 23: 2362-2378.

Anupama M and Rajagopal R. 2013.Mitogen-Activated Protein Kinases and Their Role in Radiation Response Genes & Cancer 4:401–408.

Azzam EI, Jay-Gerin JP, Pain D. 2011. Ionizing radiation-induced metabolic oxidative stress and prolonged cell injury. Cancer Lett 327:48-60. doi: 10.1016/j.canlet.2011.12.012.

Bianchi L, Tacchini L, Cairo G. 1999. HIF-1-mediated activation of transferrin receptor gene transcription by iron chelation. Nucleic Acids Res 27:4223–4227.

Feinendegen LE, Pollycove M, and Neumann RD. 2013. Hormesis by low dose radiation effects: low-dose cancer risk modelling must recognize up-regulation of protection In: Baum RP, editor. Therapeutic Nuclear Medicine. Berlin Heidelberg, Germany: Springer-Verlag; ISBN 973-3-540-36718-5.

Finkle T 2012. Signal transduction by mitochondrial oxidants. J. Biol. Chem 287: 4434-4440

Gade S and Gandhi NM. 2015. Baicalein Inhibits MCF-7 Cell Proliferation In Vitro, Induces Radiosensitivity, and Inhibits Hypoxia Inducible Factor. J Environ Pathol Toxicol Oncol. 34(4):299-308.

Karam PA and Leslie SA. 1999. Calculations of background beta-gamma radiation dose through geologic time Health Phys 77:662-667.

Konca K, Larkoff A, Banasik A, Lisowska H, Kuszewski T, Gozdz S et al. 2000. A cross platform public domain PC image analysis program for the comet assay. Mutat. Res 534:15–20.

Lall R, Ganapathy S, Yang M, Xiao S, Xu T, Su H, Shadfan M, Asara JM, Ha CS, Ben-Sahra I, Manning BD, Little JB, Yuan ZM. 2014. Low-dose radiation exposure induces a HIF-1-mediated adaptive and protective metabolic response Cell Death Differ 21:836-44. doi: 10.1038/cdd.2014.24

Little MP, Kukush AG, Masiuk SV, Shklyar S, Carroll RJ, Lubin JH, et al. 2014. Impact of uncertainties in exposure assessment on estimates of thyroid cancer risk among Ukrainian children and adolescents exposed from the Chernobyl accident. PLoS One e85723:9.

Michael H, Li W, Hanley JR, Jong SP, and Paul D. 2000. Ionizing Radiation-Induced Mitogen-Activated Protein (MAP) Kinase Activation in DU145 Prostate Carcinoma Cells: MAP Kinase Inhibition Enhances Radiation-Induced Cell Killing and G2/M-Phase Arrest Radiat. Res. 153:371.

Min W, Jeanie SK, et al 2005. Manganese superoxide dismutase suppresses hypoxic induction of hypoxia-inducible factor-1a and vascular endothelial growth factor Oncogene 24: 8154-8166.

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Morgan WF and Bair WJ. Issues in Low Dose Radiation Biology: The Controversy Continues. A Perspective Radiat Res 2013; 179: 501-510 DOI: 10.1667/RR3306.1.

Mullenders L, Atkinson M, Paretzke H, Sabatier l, and Bouffler S. 2009. Assessing cancer risks of low-dose radiation. Nat Rev Cancer 9:596–604.

Murley JS, Miller RC, Weichselbaum RR and Djrdina DJ. 2017. TP53 Mutational Status and ROS Effect the Expression of the Survivin-Associated Radio-Adaptive Response Radiat. Res. 188: 579-590

Mylonis I, Chachami G, Samiotaki M, Panayotou G, Paraskeva E, Kalousi A, Georgatsou E, Bonanou S, Simos G. 2006. Identification of MAPK phosphorylation sites and their role in the localization and activity of hypoxia-inducible factor-1alpha. J Biol Chem 281:33095-106

Pan Y, Mansfield KD, Bertozzi CC et al. 2007. Multiple factors affecting cellular redox status and energy metabolism modulate hypoxia-inducible factor prolyl hydroxylase activity in vivo and in vitro. Mol Cell Biol 27:912–925.

Sasaki MS, Tachibana A, Takeda S. 2014. Cancer risk at low doses of ionizing radiation: artificial neural networks inference from atomic bomb survivors. J Radiat Res 55:391–406. doi: 10.1093/jrr/rrt133.

Semenza GL. 2012a. Hypoxia-inducible factors: mediators of cancer progression and targets for cancer therapy. Trends Pharmacol Sci. 33(4): 207–214.

Semenza GL. 2012b. Hypoxia-Inducible Factors in Physiology and Medicine Cell 148(3): 399–408. doi:10.1016/j.cell.2012.01.021.

Shore R, Walsh L, Azizova T, Rühm W. 2017. Risk of Solid Cancer in Low Dose-Rate Radiation Epidemiological Studies and the Dose-Rate Effectiveness Factor. Int J Radiat Biol 19:1-42.

Singh NP. 2000. Microgels for estimation of DNA strand breaks, DNA protein crosslinks and apoptosis. Mutat Res 455:111–127.

Suwimol K, Sujatha V, Garry RB, and Larry WO. 2008. Manganese Superoxide Dismutase Modulates Hypoxia-Inducible Factor-1A Induction via Superoxide Can. Res. 68:27818.

Tian H, McKnight SL, Russell DW. 1997.Endothelial PAS domain protein 1 (EPAS1), a transcription factor selectively expressed in endothelial cells Genes Dev 11:72-82.

Woo KJ, Lee TJ, Park JW, Kwon TK. 2003.Desferrioxamine, an iron chelator, enhances HIF-1alpha accumulation via cyclooxygenase-2 signalling pathway. Biochem Biophys Res Commun 343:8-14.

Yuan Y, Hilliard G, Ferguson T, Millhorn DE. 2003. Cobalt inhibits the interaction between hypoxia-inducible factor-alpha and von Hippel-Lindau protein by direct binding to hypoxia-inducible factor-alpha. J. Biol. Chem 278:15911–15916.

Zhao CB, Shi L, Pu HH, Zhang QY. 2017. The Promoting Effect of Radiation on Glucose Metabolism in Breast Cancer Cells under the Treatment of Cobalt Chloride Pathol. Oncol.Res 23: 47-53.

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Acknowledgements

This work was supported by Department of Atomic Energy, Government of India. Author will like to thank Dr. PV Babu for the critical review of the manuscript. Dr. Steve Mcknight for generously providing the plasmid. Ms. Shruti Gade and Ms. Krishna for helping in some of the experiments. Mr. Prayag Amin for conducting the FACS analysis.

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Legends to the figures:

Figure 1. Toxicity of $CoCl_2$ on MCF-7 cells. MCF-7 cells were grown in 96 wells plate with different concentration of $CoCl_2$ in triplicate. After 48 hours, viability was determined using CCK-8 kit (Sigma). Figure shows the percent viability of MCF-7 cells at different concentration of $CoCl_2$ compared to control (without $CoCl_2$)

Figure 2. Effect of priming dose on viability of cells when challenged with higher dose assayed by MTT assay MCF-7 cells were first primed with 0.1 Gy of γ-radiation, with one set cells were treated with 100 µM of $CoCl_2$ at time T=0, followed by exposure to 2 Gy, after 8 h. MTT assay was performed 48 h post challenging dose. All the sets were done in triplicate. (* $p<0.05$; * compared to 2 Gy radiation alone and ns= not significant compared to control)

Figure 3. Effect of priming dose on challenging dose induced DNA damage assayed by alkaline comet assay MCF-7 cells were first exposed to the priming dose of 0.1 Gy, after 8 h those cells were exposed to 2 Gy γ-radiation and immediately assayed for DNA strand breaks by comet assay. In one group, cells were treated with 200 µM of $CoCl_2$ 7 h before exposing to 2 Gy γ-radiation. All the sets were repeated three times. Statistical significance was measured using Mann-Whitney test (*$p<0.05$; #$p<0.001$)

Figure 4. Effect of priming dose on apoptotic cell death after exposure to challenging dose of 2 Gy MCF-7 cells were first primed with 0.1 Gy of priming dose. After 8 h of incubation cells were exposed to 2 Gy γ-radiation. After challenging dose cells were further incubated for 48 h, harvested and stained with FITC-Annexin V/Pi and analyzed by FACS. One group was treated with $CoCl_2$ (200 µM) in place of priming dose. (#$p<0.05$ with student's t test)

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

Figure 5. Dose dependant induction of transcription activity.MCF-7 cells were plated on 96 well plates 24 h prior to transfection. After washing out the media, the cells were incubated with 50 ng of p-HRE3-tk-luc plasmid and lipofectamine reagent in media without FCS. After 4 h of incubation 10 % FCS was added the cells were further incubated for 16 h. The cells were then exposed to different doses of γ-radiation and incubated for 8 h. In one set CoCl2 (200 µM) was added as positive control. All the experiment set were done in triplicate. At the end of the incubation the cells were harvested, counted for the cell number and were snap-frozen in liquid nitrogen, and then stored at -80 ºC. Luciferase assay was performed using LUC-1 kit (Sigma). Significance was assumed if $p<0.05$. (* compared to non-irradiated; 0 Gy control and, ns= not significant, compared to each other.)

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

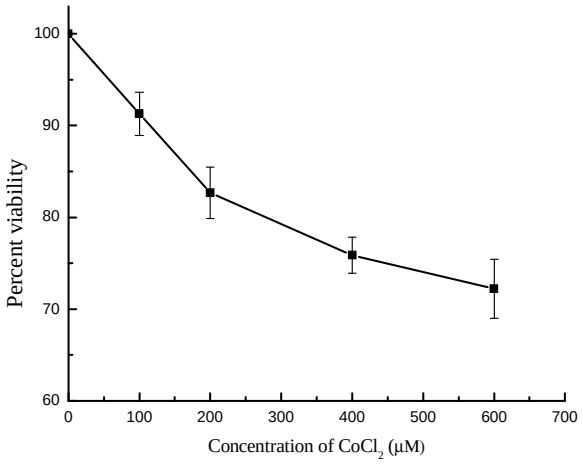

Figure 1. Toxicity of $CoCl_2$ on MCF-7 cells. MCF-7 cells were grown in 96 wells plate with different concentration of $CoCl_2$ in triplicate. After 48 hours, viability was determined using CCK-8 kit (Sigma). Figure shows the percent viability of MCF-7 cells at different concentration of $CoCl_2$ compared to control (without $CoCl_2$)

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

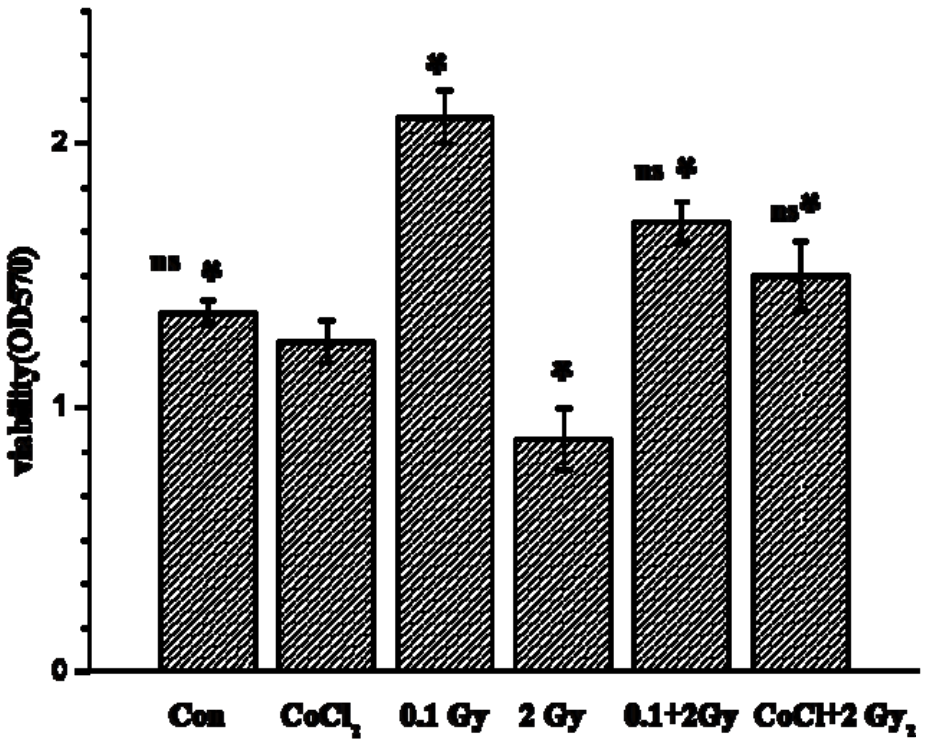

Figure 2. Effect of priming dose on viability of cells when challenged with higher dose assayed by MTT assay MCF-7 cells were first primed with 0.1 Gy of γ-radiation, with one set cells were treated with 100 µM of $CoCl_2$ at time T=0, followed by exposure to 2 Gy, after 8 h. MTT assay was performed 48 h post challenging dose. All the sets were done in triplicate. (* $p<0.05$; * compared to 2 Gy alone and ns= not significant compared to control)

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

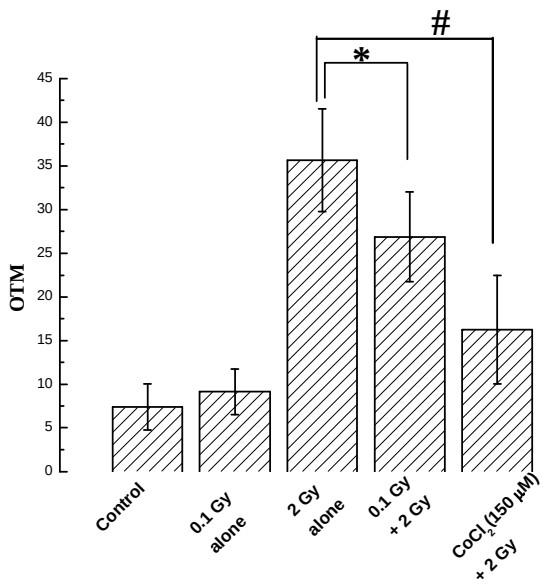

Figure 3. Effect of priming dose on challenging dose induced DNA damage assayed by alkaline comet assay MCF-7 cells were first exposed to the priming dose of 0.1 Gy, after 8 h those cells were exposed to 2 Gy γ-radiation and immediately assayed for DNA strand breaks by comet assay. In one group, cells were treated with 200 µM of $CoCl_2$ 7 h before exposing to 2 Gy γ-radiation. All the sets were repeated three times. Statistical significance was measured using Mann-Whitney test (*$p<0.05$; #$p<0.001$)

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

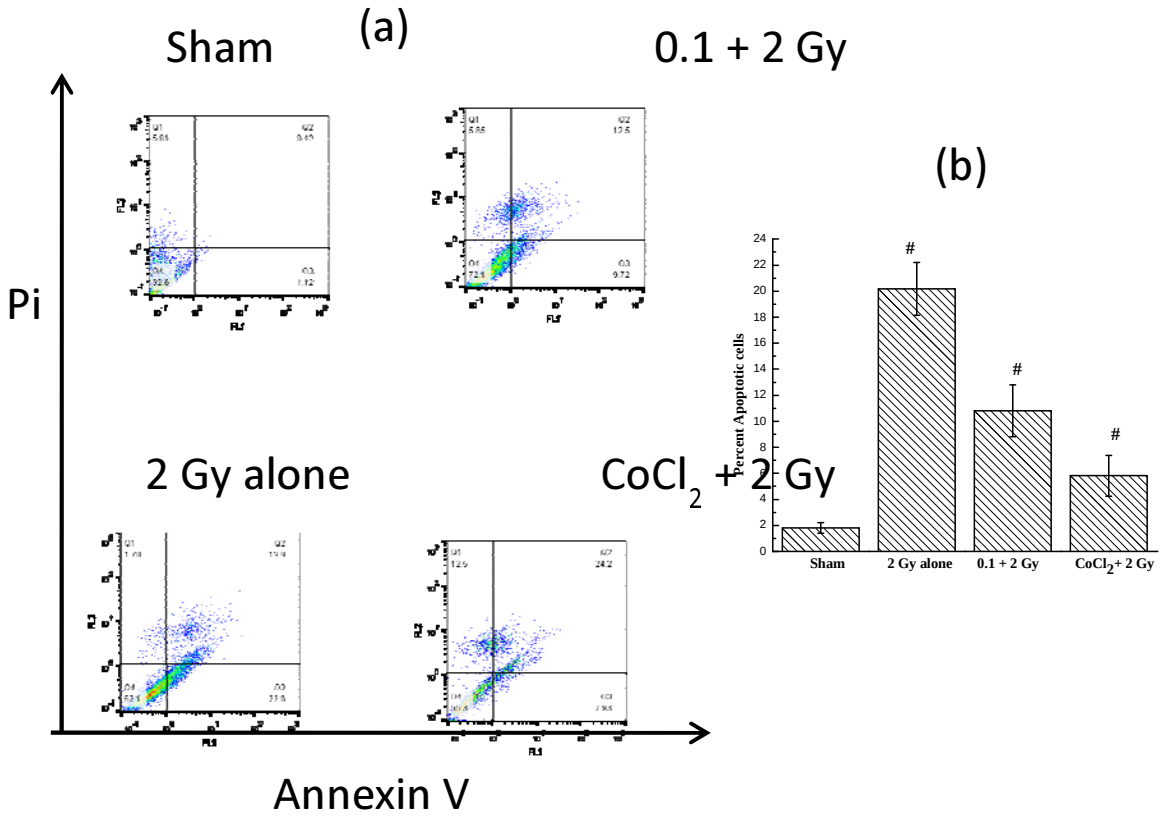

Figure 4. Effect of priming dose on apoptotic cell death after exposure to challenging dose of 2 Gy MCF-7 cells were first primed with 0.1 Gy of priming dose. After 8 h of incubation cells were exposed to 2 Gy γ-radiation. After challenging dose cells were further incubated for 48 h, harvested and stained with FITC-Annexin V/Pi and analyzed by FACS. One group was treated with $CoCl_2$ (200 µM) in place of priming dose. (#$p<0.05$ with student's t test compared to control)

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

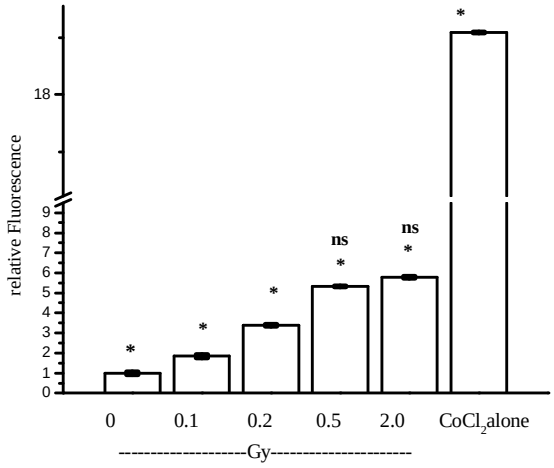

Figure 5. Dose dependant induction of transcription activity.MCF-7 cells were plated on 96 well plates 24 h prior to transfection. After washing out the media, the cells were incubated with 50 ng of p-HRE3-tk-luc plasmid and lipofectamine reagent in media without FCS. After 4 h of incubation 10 % FCS was added the cells were further incubated for 16 h. The cells were then exposed to different doses of γ-radiation and incubated for 8 h. In one set CoCl2 (200 µM) was added as positive control. All the experiment set were done in triplicate. At the end of the incubation the cells were harvested, counted for the cell number and were snap-frozen in liquid nitrogen, and then stored at -80 ºC. Luciferase assay was performed using LUC-1 kit (Sigma). Significance was assumed if $p<0.05$. (* compared to non-irradiated; 0 Gy control and, ns= not significant, compared to each other.)

bioRxiv preprint doi: https://doi.org/10.1101/316943; this version posted June 20, 2018. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.