Parasite intensity is driven by temperature in a wild passerine bird

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Abstract

Increasing awareness that parasitism is an essential component of nearly all aspects of ecosystem functioning, as well as a driver of biodiversity, has led to rising interest in the consequences of climate change in terms of parasitism and disease spread. Yet empirical knowledge of the extent and ways in which climatic factors affect parasite prevalence and intensities remains scarce. In an 18-year, multi-site, correlative study we investigated the contributions of weather variables and other factors to spatio-temporal variation in infestation by blowfly parasitic larvae (Protocalliphora spp.) in nests of Corsican blue tits (Cyanistes caeruleus). We found that ambient temperature during the nestling stage is strongly and positively related to parasite load (number of parasites per chick), both across broods when controlling for year, and across years. In addition, annual mean parasite load also increased with minimal spring temperature, and decreased when average temperature was high in the previous summer. In this Mediterranean study system, parasite intensity thus appears to be primarily driven by ambient temperature during spring and summer. This suggests a major role for thermal tolerance at various stages in the blowfly life cycle, with implications for a possible northwards range shift for these parasites as climate keeps warming up. Finally, given that ambient temperature increases throughout the breeding season and that blowfly infestation has well-established consequences for survival and recruitment of blue tits, these results also suggest temporal variation in the selection that parasites may impose on their hosts, and in particular on host phenology.

Introduction

As global climate is very likely to keep warming up, it is of growing importance to understand how populations and communities respond to variations in temperature and precipitation. While milder, wetter winters are expected for Northern Europe, in the Mediterranean most of the expected climatic change will likely translate into warmer, drier summers (IPCC 2013). Increasing awareness that parasitism is an essential component of nearly all aspects of ecosystem functioning, as well as a driver of biodiversity (Hudson et al. 2006), has led to a rising interest in the consequences of climate change in terms of parasitism and disease spread (Harvell et al. 2002, Brooks and Hoberg 2007). Attempts were made to predict the direction of change in disease prevalence in response to climate warming (e.g. Møller et al. 2013). However, it seems that no single scenario is to be expected given the huge variation in parasite life histories and the complexity of their effects at multiple levels within ecosystems (Mas-Coma et al. 2009, Rohr et al. 2011, Altizer et al. 2013). Therefore, predictions of the effects of climate change on parasites and disease need to be supported by detailed empirical knowledge acquired regionally, and in relevant host-parasite systems (Hernandez et al. 2013, Roiz et al. 2014).

Amongst parasites and pathogens, invertebrates spending part of their life cycle as free-living (*i.e.* away from their host) are likely most affected by weather and climate (Hernandez et al. 2013, Rose et al. 2014, Charlier et al. 2016, Ogden and Lindsay 2016). In those ectoparasites developmental time, activity rates, and survival often display bell-shaped responses, *i.e.* highest at intermediate temperature and moisture values (reviewed in Ogden and Lindsay 2016). Local weather fluctuations may therefore result in either increased or decreased parasite intensities depending on the current position of populations relative to these optima (Stromberg 1997, Elderd and Reilly 2014, Eads and Hoogland 2016). In the longer run parasites could respond by means of adaptive evolution under directional climate change. In addition, invertebrates may be differently affected by climatic variability depending on their biology, and in particular on their ability to take refuge in micro-habitats that can buffer adverse climatic conditions (e.g. Roiz et al. 2014, Ogden and Lindsay 2016).

Within nesting cavities, birds are in contact with invertebrates, including nest-dwelling ectoparasites that use avian nest material as a habitat (Loye and Zuk 1991, Christe et al. 1994, Clayton and Moore 1997). The fitness costs of infestation as well as the fitness benefits of host defense traits are well documented in a range of bird species (e.g. Møller et al. 1990, Eeva and Nurmi 1994, Cantarero et al. 2013). This contrasts with the currently limited understanding of the factors driving the large temporal and spatial variation often found in

ectoparasite intensities (Hurtrez-Boussès et al. 1999, Heeb et al. 2000, Dudaniec et al. 2007, Moreno et al. 2009). It was suggested that nest ectoparasites might be influenced by weather, nest size or composition, or interactions with other invertebrate species within the nest microhabitat (Bennett and Whitworth in press, Heeb et al. 1996, Remeš and Krist 2005, Kleindorfer and Dudaniec 2009, Moreno et al. 2009), yet the relative contributions of these factors remain unclear.

Among the best-studied nest ectoparasites in free-ranging birds are the haematophagous larvae of *Protocalliphora* blowflies that feed on bird nestlings' blood (Bennett and Whitworth in press, Owen and Ash 1955, Møller et al. 1990, Eeva and Nurmi 1994, Hurtrez-Boussès et al. 1997, Dawson et al. 2005, Remeš and Krist 2005, Cantarero et al. 2013). The first eggs deposited by adult *Protocalliphora* blowflies hatch from the beginning of the bird nestling stage onwards, and develop into three larval stages before pupating. Adult blowflies are free-living and likely overwinter in litter and old nest materials (Matyukhin and Krivosheina 2008). The host spectrum of *Protocalliphora* is quite large, although avian hole-nesters seem to be especially exposed to this ectoparasite (Owen and Ash 1955, Jamriska et al. 2010). Nest-box studies showed that bird nestlings developing in nests with more *Protocalliphora* larvae have lower body condition, reduced growth or smaller body size (Merino and Potti 1996, Hurtrez-Boussès et al. 1997, Charmantier et al. 2004), decreased resting time (Simon et al. 2005) and lower physiological performance (Simon et al. 2004, Thomas et al. 2007). In addition, *Protocalliphora* abundance was found positively related to bacterial loads measured on blue tit (*Cyanistes caeruleus*) nestlings, suggesting higher risks of bacterial infection in nests heavily infested by *Protocalliphora* (Mennerat et al. 2009). *Protocalliphora* do not parasitize adult birds, but indirectly affect avian reproductive effort as reflected in increased feeding rates by males or increased investment into nest-sanitation behaviour by females (Hurtrez-Bousses et al. 1998, Bańbura et al. 2004). On the island of Corsica, blue tit nestlings (*Cyanistes caeruleus*) are exposed to the highest *Protocalliphora* loads reported in European study sites, while other types of nest ectoparasites are rarely found (Hurtrez-Boussès et al. 1997, 1999, Mennerat et al. 2008).

In an 18-year, multi-site, correlative study, we investigated how *Protocalliphora* infestation intensities varied both within and across years, with the main objective to understand the relative contributions of climatic and other factors to spatio-temporal variation in parasite intensities. We more specifically explored within-year variability in relation to host life history, nest characteristics and weather during the nestling stage, while accounting for spatial variation. We also explored how interannual variability in mean parasite intensities

relates to temperature and rainfall during the summer, autumn, winter and spring preceding

each breeding season.

Methods

Study sites and monitoring

Data from uniquely ringed female breeders were obtained for years 1997-2014 from seven

study plots situated in two valleys on the island of Corsica (plots Avapessa AVA, Arinelle

ARI, Feliceto FEL, Filagna FIL, Grassa GRA and Muro MUR in the Muro valley; plot Pirio

PIR in the Fango valley) (Blondel 1985, Lambrechts et al. 2004). The broad-leaved deciduous

oak *Quercus humilis* favouring the production of earlier and larger blue tit broods was the

dominant tree species in plots AVA, FEL, and MUR. The evergreen oak *Q. ilex* was the

dominant tree species in the other four study plots where blue tit broods were smaller and

reared later in the season (Lambrechts et al. 2004, Blondel et al. 2006). Breeding blue tits used

either wood-concrete Schwegler B1 boxes (Schorndorf, Germany) or concrete boxes of

similar dimensions (nest-chamber size of *ca.* 113 cm²).

Following basic protocols (Blondel et al. 2006) boxes were visited at least once a week

to check the progress of each avian breeding attempt, the initiation and progress of nest

construction, the first egg-laying dates expressed in Julian dates, and the number of eggs and

nestlings in the nest. Nest thickness, *i.e.* the vertical distance between the bottom floor and the

top of the external nest wall (Hurtrez-Boussès et al. 1999), was measured either shortly before

or during the egg-laying period. Given that all nestboxes in this study have similar internal

diameters, nest thickness is an appropriate proxy for nest volume, *i.e.* habitat size for nest

ectoparasites.

Because nests were not visited every day at the end of the incubation stage, the onset

of hatching was calculated based on the physical development of the nestlings within a few

days after hatching (Descamps et al. 2002). When nestlings were between nine and 15 days

old, adult breeders were trapped inside the nestbox. The age of the female parent (yearling *vs*

older) was determined either by using the wing plumage colors or, for previously ringed birds,

from the monitoring records (cf. Blondel et al. 2006). The age of female breeders may

contribute to variation in parasite loads by affecting their sanitation behaviour. Breeding

attempts for which female age could not be identified were not considered in this study.

Meteorological data

Records of daily minimum temperature (°C), daily maximum temperature (°C), and daily

rainfall (mm) were obtained from the meteorological station of Calvi in Corsica. This weather

station is situated at a maximum of 20 km from each of the study plots and thus informs on

regional climatic variation (see also Grosbois et al. 2006). We averaged the daily minimum

and maximum ambient temperature to obtain a measure of daily average ambient temperature.

Based on this, for each nest during the two weeks following hatching we calculated the

average ambient temperature and the average amount of rainfall. This period corresponds to

the time when *Protocalliphora* larvae develop in blue tit nests by intermittently feeding on

nestling blood, while spending the rest of the time hidden amongst nest materials (Hurtrez-

Boussès et al. 1999).

To explore the effect of interannual climatic variation on mean *Protocalliphora*

intensities we used meteorological archives of minimal, maximal and average monthly

temperatures, as well as monthly rainfall. We calculated average temperature and total rainfall

over three-months periods corresponding to summer (June-August), autumn (September-

November), winter (December-February), and spring (March-May) preceding each breeding

season.

Protocalliphora abundance

Nests were collected 15 days post-hatching, stored in hermetic plastic bags, and replaced by

similar amounts of new nest material, mainly moss. In the laboratory, *Protocalliphora* larvae

and pupae were sorted out of the nest material and counted. *Protocalliphora* larvae develop

into three successive larval stages before pupating. Our counts included the total number of

second-stage larvae, third-stage larvae and pupae (excluding first-stage larvae that are difficult

to detect due to their small size), following the protocols presented in Hurtrez-Boussès (1996),

Heeb et al. (2000) and Mennerat et al. (2008; 2009). In some study years (e.g. Mennerat et al.

2008; 2009), nests were enclosed in cotton bags to facilitate their collection (without: 274

nests; with: 261 nests). Cotton bags were first inserted under blue tit nests around hatching

time, so that the blue tit parents could habituate to the presence of the bag. A few days later

the edge of the bag was pulled up to reach the same height as that of the nest.

Statistical analysis

This study includes data from 535 broods covering an 18-year period (Table 1). Our study

only focused on first-clutch broods for which the contents of the nests were not manipulated,

nests that were not experimentally treated against *Protocalliphora*, and nests for which there

was no evidence of predation. We used the mean number of *Protocalliphora* larvae per

nestling (*i.e.* the number of larvae found in a nest divided by the number of chicks present at

time of sampling) as a measure of parasite load. All analyses were performed in the statistical

programming environment *R 3.2.2* (http://r-project.org). Model validation was performed by

visual inspection of residuals.

Within-year variation in parasite intensity

To investigate the relation between parasite intensity and current biotic or abiotic factors, we

used linear mixed-effect models with log-transformed parasite load as a dependent variable

(*lmer* from the lme4 package). We applied forward, AIC-based model selection starting with a

set of models with nest thickness, cotton bag treatment (with *vs* without a cotton bag), egg-

laying date (in Julian dates), and weather during the nestling stage (average ambient

temperature and rainfall) as explanatory variables. Female age was also included because nest

sanitation behaviour (exclusively performed by females), and thus parasite intensities, may be

affected by breeding experience (Banbura et al. 2001). Year (n=18), study site (n=7), nestbox

identity (n=243), and female identity (n=385) were included as random effect factors.

Interannual variation in parasite intensity

We further investigated the interannual variation in mean parasite load in relation to weather

during the summer, autumn, winter and spring preceding each breeding period. The study

sites are located in two distinct valleys (Table 1) that differ markedly in a range of factors

(Blondel et al. 2006). We used linear models with mean (yearly average, calculated for each

valley) *Protocalliphora* abundance per chick as a dependent variable. We applied forward,

AIC-based model selection with an initial set of explanatory models with valley as a factor (to

account for potential valley-specific relations between parasite intensities and weather) and

yearly mean values of egg laying-date, ambient temperature during the nestling period,

minimum, maximum, and average ambient temperature and total rainfall during each three-

month period (*i.e.* season) preceding the breeding season when parasite intensities were

sampled. Interactions between valley and all other covariates were also included.

Results

Within-year variation in parasite intensity

The final model for brood parasite intensity included only ambient temperature during the

nestling stage and cotton bag treatment as explanatory variables, and its fit was further

improved by adding a quadratic term for ambient temperature. Parasite load increased with

mean ambient temperature (linear term: $P < 10^{-4}$), with some degree of saturation at higher

temperatures (quadratic term: $P < 10^{-4}$; Table 2; Figure 1). In addition, parasite load was

higher in nests surrounded by a cotton bag than in nests collected without a cotton bag (P =

0.02).

Interannual variation in parasite intensity

Across years, mean parasite load was strongly and positively correlated with ambient

temperature during breeding ($P < 10^{-4}$; Figure 2), and to a lesser extent negatively correlated

with average temperature in the previous summer (P = 0.009; Figure 3) and positively

correlated with minimal spring temperature (P = 0.03; Figure 4). No other variable was

retained in the final model (Table 3).

Discussion

In this study we found that ambient temperature during the nestling stage is strongly related to

variation in parasite load, both across broods when controlling for year, and across years. In

addition, annual mean parasite load also increased with minimal spring temperature, and

decreased when average temperature was high in the previous summer. In this Mediterranean

study system, parasite intensity thus appears to be primarily driven by ambient temperatures

during spring and summer, similarly in all seven study plots. We also confirm that parasite

loads are higher in nests surrounded by a cotton bag compared to nests without a bag,

probably because these bags prevent the loss of nest-dwelling parasites, which was the main

argument for using them (Hurtrez-Boussès et al. 1999, Mennerat et al. 2008, 2009).

Our results indicate that thermal dependence is an important mechanism at various

stages in the life cycle of *Protocalliphora* blowflies. The main factor found to affect parasite

load is ambient temperature during the larval stage (*i.e.* the nestling stage of blue tit hosts). In

insects, temperature dependence of larval growth and survival has been extensively studied in

the laboratory (overview in e.g. Chown and Nicolson 2004), even though the link between

lab-measured thermal limits and variation in insect abundance in the field is not always

straightforward. Our study confirms, as observed in another host species the tree swallow

Tachycineta bicolor, that blowfly larval abundance appears driven by temperature (Dawson et

al. 2005). In that previous study, parasite abundance increased across the natural temperature

range in a curvilinear way very similar to what we report here (see Figure 1), and decreased

when temperatures were experimentally raised above natural levels. Taken together, these two

studies point to an optimal temperature for larval development around 23-25 °C and suggest that larval survival is significantly reduced below 20 °C. This is consistent with observations from other species within the *Calliphoridae* family (Wall et al. 2001, Lecheta et al. 2015).

We also found a negative effect of summer heat on *Protocalliphora* loads in the following spring, possibly as a consequence of increased mortality of adult blowflies at high temperatures. While heat tolerance limits in adult *Protocalliphora* are currently unknown, in other dipteran species they appear to range from 30 °C and above (Feder et al. 1997, Berrigan et al. 2000, Chown and Nicolson 2004, Enriquez and Colinet 2017). In the willow beetle *Chrysomela aeneicolli* the expression of heat shock protein Hsp70, indicative of physiological stress, increased in field samples along with temperature and reached high levels around 30-35 °C (Dahlhoff and Rank 2000). Furthermore, there is evidence of seasonal variation in selection at the phosphoglucose isomerase (PGI) locus inducing Hsp expression (Dahlhoff et al. 2008, Dick et al. 2013). These studies, which were carried out at a latitude close to that of our study sites, reveal that the temperatures measured on sunlit soil can approach 40 °C even when the maximum air temperature recorded by a nearby weather station did not exceed 26 °C. Considering that in our study area the maximal summer (air) temperature ranged from 34.0 up to 40.6 °C (average summer temperature 21.1 – 25.4 °C, see Figure S1), this means that adult *Protocalliphora* blowflies could in some years be exposed to temperatures above their thermal tolerance levels, resulting in high adult mortality. Minimal spring temperature (ranging from -4.6 °C to 3.4 °C, Figure S1) also seems to constrain parasite loads. Given that all minimal temperatures were recorded in March, *i.e.* before the reproductive period for *Protocalliphora*, this likely reflects cold-induced mortality of adults shortly after post-winter emergence. The selective effect of temperature extremes on parasites in this system is an aspect that deserves further attention.

Because mean ambient temperature during the nestling stage increases linearly with egg-laying date, our results help explain why parasite loads are higher in late broods, and suggest temporal variation in the selection that parasites may impose on their hosts, and in particular on host phenology (Figure S2). In the focal study populations as in generally all temperate insectivorous birds, laying date is persistently under negative natural selection (Porlier et al. 2012) and the strength of selection is stronger during very warm springs (Marrot et al. 2017, 2018). This strong selection favouring early breeding females is always discussed in the context of the phenological mismatch between birds and they main caterpillar preys (e.g. van Noordwijk et al. 1995, Visser et al. 2006), yet the present study shows that nest ectoparasites may also drive selection on laying date. *Protocalliphora* larvae are detrimental

to blue tit nestlings since they induce reduced growth, lower metabolic capacity and body condition, and eventually lower recruitment in the breeding population (Hurtrez-Boussès et al. 1997, Simon et al. 2004, Thomas et al. 2007). Prevalence is very high in Corsican populations of blue tits (Hurtrez et al 1997), yet no specific behaviour was found to either prevent these parasites from accessing the nest (Mennerat et al. 2008) or remove them (Hurtrez-Bousses et al. 1998). It appears, however, that adult blue tits can compensate for some of the harmful effects of parasites by increasing rates of chick provisioning (Hurtrez-Bousses et al. 1998). The level of compensation depends on food availability (Simon et al. 2004), and as a result parasites and food interact to determine post-fledging survival and recruitment (path analysis, Thomas et al. 2007). Based on this it can be inferred that parasite-mediated selection should be strongest in evergreen oak-dominated habitats, characterised by a combination of low caterpillar abundance and late breeding, and where parasite loads are highest. On the contrary in deciduous holm oak-dominated habitats, parasites might exert relatively weak selection on blue tits due to a combination of plentiful food, early breeding, and low parasite loads. The nature of traits in blue tits that could be under parasite-mediated selection has not yet been established, nor the strength of such selection; however given that parasite loads increase with temperature and thus egg laying date, breeding phenology appears as a likely candidate, as well as parental behaviours that improve nestling growth or survival under conditions of high parasite abundance (Hurtrez-Bousses et al. 1998, Mennerat et al. 2009).

The apparent link between parasite loads and temperature fluctuations in this Mediterranean area is relevant for host-parasite interactions at broader spatial scales. Geographical distributions are expected to be limited by climate, and especially by temperature in insects (David et al. 2003, Kingsolver and Buckley 2017), where in a number of cases the amplitude of thermal tolerance seems reflected in species latitudinal ranges (e.g. Calosi et al. 2010). The parasitic larvae in the present study belong to two species, *P. azurea* and *P. falcozi*. Both species appear widespread over the Palaearctic, but whether one is more abundant than the other remains unclear (Wesolowski 2001, Matyukhin and Krivosheina 2008). *P. azurea* was reported from Spain to Scandinavia (Potti 2008, Eeva et al. 2015), and *P. falcozi* mostly from Central Europe, but also Germany and Corsica where both species coexist (Wesolowski 2001, Janoskova et al. 2010). Strikingly, of all reports from the Palaearctic it is on Corsica that prevalences and intensities are found highest (Hurtrez-Boussès et al. 1997), and this is consistent with the idea that parasite abundances increase with minimal and average spring temperatures, as both are higher on Corsica than in most other locations in Europe. Furthermore parasite loads relate negatively to average summer

temperature; this suggests that heat in areas located further south in the Mediterranean may

limit their southern range.

Current projections (IPCC 2013) indicate that regardless of the scenario, by 2100

mean spring and summer temperatures will likely have increased not only in the

Mediterranean (spring: +1 °C to +5 °C; summer: +1.5 °C to +7 °C) but also in Central

(spring: +1 °C to +5 °C; summer: +2 °C to +6 °C) and Northen Europe (spring: +2 °C to +5

°C; summer: +2 °C to +5 °C). Blowfly prevalence and infestation intensities in Central and

Northern Europe seem moderate at the moment (Wesolowski 2001, Eeva et al. 2015), yet our

results suggest that they might increase in the future, and that the range of *Protocalliphora*

spp. might shift northwards. A more comprehensive understanding of the relationships

between climate, spring phenology, but also selection and adaptive evolution in bird hosts as

well as their parasites is now needed.

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Ethical statement

Captures were performed under personal ringing permits delivered by the CRBPO (Centre de

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approved by the ethics committee for animal experimentation of Languedoc Roussillon (305-

CEEA-LR-12066 approved in 2012) as well as by Regional Institutions (bylaw issued by the

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References

Altizer, S. et al. 2013. Climate Change and Infectious Diseases: From Evidence to a

Predictive Framework. - Science (80-.). 341: 514–519.

Banbura, J. et al. 2001. Sex differences in parental care in a Corsican Blue Tit Parus caeruleus
population. - Ardea 89: 517–526.

Bańbura, J. et al. 2004. Effects of Protocalliphora Parasites on Nestling Food Composition in
Corsican Blue Tits Parus caeruleus: Consequences for Nestling Performance. - Acta
Ornithol. 39: 93–103.

Bennett, G. and Whitworth, T. . Studies on the life history of some species of Protocalliphora
(Diptera: Calliphoridae). - Can. J. Zool.: 2048–2058.

Berrigan, D. et al. 2000. Resistance to temperature extremes between and within life cycle
stages in Drosophila serrata, D. birchii and their hybrids: intraspecific and interspecific
comparisons. - Biol. J. Linn. Soc.: 403–416.

Blondel, J. 1985. Breeding Strategies of the Blue Tit and Coal Tit (Parus) in Mainland and
Island Mediterranean Habitats: A Comparison. - J. Anim. Ecol. 54: 531–556.

Blondel, J. et al. 2006. A Thirty-Year Study of Phenotypic and Genetic Variation of Blue Tits
in Mediterranean Habitat Mosaics. - Bioscience 56: 661.

Brooks, D. R. and Hoberg, E. P. 2007. How will global climate change affect parasite-host
assemblages? - Trends Parasitol. 23: 571–574.

Calosi, P. et al. 2010. What determines a species' geographical range? Thermal biology and
latitudinal range size relationships in European diving beetles (Coleoptera: Dytiscidae). -
J. Anim. Ecol. 79: 194–204.

Cantarero, A. et al. 2013. Behavioural responses to ectoparasites in pied flycatchers Ficedula
hypoleuca: An experimental study. - J. Avian Biol. 44: 591–599.

Charlier, J. et al. 2016. Climate-driven longitudinal trends in pasture-borne helminth
infections of dairy cattle. - Int. J. Parasitol. 46: 1–8.

Charmantier, A. et al. 2004. Parasitism reduces the potential for evolution in a wild bird
population. - Evolution (N. Y). 58: 203–206.

Chown, S. and Nicolson, S. 2004. Insect physiological ecology. - Oxford University Press.

Christe, P. et al. 1994. Ectoparasite affects choice and use of roost sites in the great tit, Parus
major. - Anim. Behav.: 895–898.

1997. Host parasite evolution: general principles and avian models (D Clayton and J Moore,
Eds.). - Oxford University Press.

Dahlhoff, E. P. and Rank, N. E. 2000. Functional and physiological consequences of genetic
variation at phosphoglucose isomerase: heat shock protein expression is related to
enzyme genotype in a montane beetle. - Proc. Natl. Acad. Sci. U. S. A. 97: 10056–61.

Dahlhoff, E. P. et al. 2008. Effects of Temperature on Physiology and Reproductive Success
of a Montane Leaf Beetle: Implications for Persistence of Native Populations Enduring
Climate Change. - Physiol. Biochem. Zool. 81: 718–732.

David, J. R. et al. 2003. The fly that came in from the cold : geographic variation time from
low-temperature of recovery exposure in <i>Drosophila subobscura. 17: 425–430.

Dawson, R. D. et al. 2005. Effects of Experimental Variation in Temperature on Larval
Densities of Parasitic Protocalliphora (Diptera : Calliphoridae) in Nests of Tree
Swallows (Passeriformes : Hirundinidae) Effects of Experimental Variation in
Temperature on Larval Densities o. - Physiol. Ecol. 34: 563–568.

Descamps, S. et al. 2002. Asynchronous hatching in a blue tit population: a test of some
predictions related to ectoparasites. - Can. J. Zool. 80: 1480–1484.

Dick, C. A. et al. 2013. Effects of Temperature Variation on Male Behavior and Mating
Success in a Montane Beetle. - Physiol. Biochem. Zool. 86: 432–440.

Dudaniec, R. Y. et al. 2007. Interannual and interspecific variation in intensity of the parasitic
fly, Philornis downsi, in Darwin's finches. - Biol. Conserv. 139: 325–332.

Eads, D. and Hoogland, J. 2016. Factors that affect parasitism of black-tailed prairie dogs by

fleas. - Ecosphere 7: e01372.

Eeva, T. L. E. and Nurmi, J. 1994. Effects of ectoparasites on breeding success of great tits
(Parus major) and pied flycatchers (Ficedula hypoleuca) in an air pollution gradient. -
Can. J. Zool. 72: 624–635.

Eeva, T. et al. 2015. Species and abundance of ectoparasitic flies (Diptera) in pied flycatcher
nests in Fennoscandia. - Parasites and Vectors 8: 1–9.

Elderd, B. D. and Reilly, J. R. 2014. Warmer temperatures increase disease transmission and
outbreak intensity in a host-pathogen system. - J. Anim. Ecol. 83: 838–849.

Enriquez, T. and Colinet, H. 2017. Basal tolerance to heat and cold exposure of the spotted
wing drosophila, Drosophila suzukii. - PeerJ 5: e3112.

Feder, M. E. et al. 1997. Natural thermal stress and heat-shock protein expression in
Drosophila larvae and pupae. - Funct Ecol 11: 90–100.

Grosbois, V. et al. 2006. Climate impacts on Mediterranean blue tit survival: an investigation
across seasons and spatial scales. - Glob. Chang. Biol. 12: 2235–2249.

Harvell, C. et al. 2002. Climate Warming and Disease Risks for Terrestrial and Marine Biota.
- Science (80-.). 296: 2158–2163.

Heeb, P. et al. 1996. Horizontal Transmission and Reproductive Rates of Hen Fleas in Great
Tit Nests. - J. Anim. Ecol. 65: 474–484.

Heeb, P. et al. 2000. Bird–Ectoparasite Interactions, Nest Humidity, and Ectoparasite
Community Structure. - Ecology 81: 958–968.

Hernandez, A. D. et al. 2013. Climate changes influence free-living stages of soil-transmitted
parasites of European rabbits. - Glob. Chang. Biol. 19: 1028–42.

Hudson, P. J. et al. 2006. Is a healthy ecosystem one that is rich in parasites? - Trends Ecol.
Evol. 21: 381–5.

Hurtrez-Bousses, S. et al. 1998. Chick parasitism by blowflies affects feeding rates in a
Mediterranean population of blue tits. - Ecol. Lett. 1: 17–20.

Hurtrez-Boussès, S. et al. 1997. High blowfly parasitic loads affect breeding success in a
Mediterranean population of blue tits. - Oecologia: 514–517.

Hurtrez-Boussès, S. et al. 1999. Variations in prevalence and intensity of blow fly infestations
in an insular Mediterranean population of blue tits. - Can. J. Zool. 77: 337–341.

IPCC 2013. Annex I: Atlas of Global and Regional Climate Projections. - In: van Oldenborgh,
G. J. et al. (eds), Climate Change 2013: The Physical Science Basis. Contribution of
Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on
Climate Change. Cambridge University Press, in press.

Jamriska, J. et al. 2010. Host spectrum of bird blow flies of the genus Protocalliphora Hough ,
1899 (Diptera , Calliphoridae) in Slovakia. - Sylvia 46: 125–132.

Janoskova, V. et al. 2010. Pre-imaginal stages of the blowfly Protocalliphora falcozi in nests
of the tree sparrow (Passer montanus). - Entomol. Fenn. 21: 107–116.

Kingsolver, J. G. and Buckley, L. B. 2017. Quantifying thermal extremes and biological
variation to predict evolutionary responses to changing climate. - Philos. Trans. R. Soc.
B Biol. Sci. 372: 20160147.

Kleindorfer, S. and Dudaniec, R. Y. 2009. Love thy neighbour? Social nesting pattern, host
mass and nest size affect ectoparasite intensity in Darwin's tree finches. - Behav. Ecol.
Sociobiol. 63: 731–739.

Lambrechts, M. M. et al. 2004. Habitat quality as a predictor of spatial variation in blue tit
reproductive performance: A multi-plot analysis in a heterogeneous landscape. -
Oecologia 141: 555–561.

Lecheta, M. C. et al. 2015. The effect of temperature on development of Sarconesia
chlorogaster, a blowfly of forensic importance. - Forensic Sci. Med. Pathol. 11: 538–543.

Loye, J. and Zuk, M. 1991. Bird - parasite interactions: ecology, evolution and behaviour. -

Oxford University Press.

Marrot, P. et al. 2017. Multiple extreme climatic events strengthen selection for earlier

breeding in a wild passerine. - Philos Trans R Soc L. B Biol Sci 372: 20160372.

Marrot, P. et al. 2018. Current spring warming as a driver of selection on reproductive timing

in a wild passerine. - J. Anim. Ecol. 87: 754–764.

Mas-Coma, S. et al. 2009. Climate change effects on trematodiases, with emphasis on

zoonotic fascioliasis and schistosomiasis. - Vet. Parasitol. 163: 264–280.

Matyukhin, A. and Krivosheina, M. 2008. Contribution to the knowledge of Diptera (Insecta)

parasitizing on birds. - Entomol. Rev. 88: 258–259.

Mennerat, A. et al. 2008. Aromatic plants in blue tit Cyanistes caeruleus nests: no negative

effect on blood-sucking Protocalliphora blow fly larvae. . - J. Avian Biol. 39: 127–132.

Mennerat, A. et al. 2009. Aromatic plants in nests of the blue tit Cyanistes caeruleus protect

chicks from bacteria. - Oecologia 161: 849–855.

Merino, S. and Potti, J. 1996. Weather dependent effects of nest ectoparasites on their bird

hosts. - Ecography (Cop.). 19: 107–113.

Møller, A. et al. 1990. Fitness effects of parasites on passerine birds: a review. - In: Blondel,

J. et al. (ed), Population biology of passerine birds. NATO ASI S. Springer-Verlag, pp.

269–280.

Møller, A. P. et al. 2013. Assessing the effects of climate on host-parasite interactions: A

comparative study of european birds and their parasites. - PLoS One in press.

Moreno, J. et al. 2009. Nest-dwelling ectoparasites of two sympatric hole-nesting passerines

in relation to nest composition: An experimental study. - Ecoscience 16: 418–427.

Ogden, N. H. and Lindsay, L. R. 2016. Effects of Climate and Climate Change on Vectors

and Vector-Borne Diseases: Ticks Are Different. - Trends Parasitol. 32: 646–656.

Owen, D. and Ash, J. 1955. Additional records of protocalliphora (Diptera) in birds' nests. -

Br. Birds XLVIII: 225–229.

Porlier, M. et al. 2012. Variation in phenotypic plasticity and selection patterns in blue tit

breeding time: Between- and within-population comparisons. - J. Anim. Ecol. 81: 1041–

1051.

Potti, J. 2008. Blowfly Infestation at the Nestling Stage Affects Egg Size in the Pied

Flycatcher *Ficedula hypoleuca*. - Acta Ornithol. 43: 76–82.

Remeš, V. and Krist, M. 2005. Nest design and the abundance of parasitic Protocalliphora

blow flies in two hole-nesting passerines. - Ecoscience 12: 549–553.

Rohr, J. R. et al. 2011. Frontiers in climate change-disease research. - Trends Ecol. Evol. 26:

270–277.

Roiz, D. et al. 2014. Climatic effects on mosquito abundance in Mediterranean wetlands. -

Parasit. Vectors 7: 333.

Rose, H. et al. 2014. Exploiting parallels between livestock and wildlife: Predicting the

impact of climate change on gastrointestinal nematodes in ruminants. - Int. J. Parasitol.

Parasites Wildl. 3: 209–219.

Simon, A. et al. 2004. Physiological ecology of Mediterranean blue tits (Parus caeruleus L.):

effects of ectoparasites (Protocalliphora spp.) and food abundance on metabolic capacity

of nestlings. - Physiol. Biochem. Zool. 77: 492–501.

Simon, A. et al. 2005. Impact of Ectoparasitic Blowfly Larvae (Protocalliphora Spp.) on the

Behavior and Energetics of Nestling Blue Tits. - J. F. Ornithol. 76: 402–410.

Stromberg, B. E. 1997. Environmental factors influencing transmission. - Vet. Parasitol. 72:

247–264.

Thomas, D. et al. 2007. Common paths link food abundance and ectoparasite loads to

physiological performance and recruitment in nestling blue tits. - Funct. Ecol. 21: 947–

955.

van Noordwijk, A. J. et al. 1995. Selection for the Timing of Great Tit Breeding in Relation to
Caterpillar Growth and Temperature. - J. Anim. Ecol. 64: 451.
Visser, M. E. et al. 2006. Shifts in Caterpillar Biomass Phenology Due to Climate Change and
Its Impact on the Breeding Biology of an Insectivorous Bird. - Oecologia 147: 164–172.
Wall, R. et al. 2001. Pre-adult mortality in the blowfly Lucilia sericata. - Med. Vet. Entomol.
15: 328–334.
Wesolowski, T. 2001. Host - parasite interactions in natural holes: marsh tits (Parus palustris)
and blow flies (Protocalliphora falcozi). - J. Zool.: 495–503.
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Table 1. Yearly fluctuations in the number (mean ± SD) of *Protocalliphora* larvae per blue tit nestling on Corsica from 1997 to 2014.

Year	Muro valley	Fango valley
1997	-	12.15 ± 7.01 (10)
1998	6.98 ± 0.84 (2)	10.38 ± 6.72 (8)
1999	6.15 ± 1.21 (2)	11.07 ± 4.53 (4)
2000	3.66 ± 2.63 (6)	13.79 ± 5.20 (5)
2001	3.07 ± 3.48 (23)	13.12 ± 5.51 (16)
2002	-	9.54 ± 3.29 (3)
2003	-	9.00 ± 3.59 (11)
2004	-	5.78 ± 4.71 (19)
2005	5.44 ± 3.65 (53)	10.23 ± 5.41 (35)
2006	-	12.44 ± 5.47 (6)
2007	3.37 ± 2.97 (56)	-
2008	5.68 ± 5.81 (52)	15.31 ± 8.58 (23)
2009	3.55 ± 2.73 (39)	11.39 ± 6.36 (33)
2010	2.94 ± 2.62 (29)	8.11 ± 4.14 (35)
2011	-	10.64 ± 6.71 (24)
2012	-	12.90 ± 7.27 (13)
2013	-	2.12 ± 1.49 (5)
2014	-	12.08 ± 10.21 (23)
Total	4.26 ± 3.93 (262)	10.70 ± 6.77 (273)

Table 2. Within-year variation in parasite load per chick in blue tit nests infested by

hematophageous *Protocalliphora* larvae, in relation to mean ambient temperature during the

nestling stage (*i.e.* in the period when parasite larvae develop into blue tit nests) and cotton

bag treatment (*i.e.* presence or absence of a cotton bag around the nest). Results from the final

linear mixed effects model, obtained after forward selection from an initial set of models with

nest thickness (*i.e.* height), cotton bag treatment, egg-laying date, female age, and average

ambient temperature and rainfall during the nestling stage as explanatory variables. Year,

study site, nestbox identity, and female identity were included as random effect factors.

	d.f.	Estimate	SE	Chisq	P
Mean ambient temperature during nesting (linear)	1	1.58	0.29	29.53	$< 10^{-4}$
Mean ambient temperature during nesting (quadratic)	1	-0.04	0.008	24.23	$< 10^{-4}$
Cotton bag treatment	1	0.22	0.09	5.56	0.02

Table 3. Interannual variation in mean parasite load per chick (yearly average) in nests of blue

tits infested by hematophageous *Protocalliphora* larvae, in relation to yearly average values

of mean temperature during nesting, mean temperature in the previous summer, and minimal

spring temperature. Results from the final linear model, obtained after forward selection from

an initial set of models with valley, egg laying-date, ambient temperature during the nestling

period, minimum, maximum, and average ambient temperature and total rainfall during each

preceding season as explanatory variables, as well as interactions between valley and all other

covariates.

	d.f.	Estimate	SE	Sum of squares	P
Mean temperature during nesting	1	1.60	0.17	272.1	$< 10^{-4}$
Mean temperature in previous summer	1	-0.89	0.34	20.40	0.009
Minimal spring temperature	1	0.37	0.17	14.32	0.03

Figure 1

Figure 1. Across broods, parasite load increases with mean ambient temperature during the
nestling stage (*i.e.* during the developmental period of *Protocalliphora* larvae) in nests of blue
tits on Corsica. Each dot represents a brood. The grey curve shows predicted values from the
final model (see Table 2).

Figure 2

Figure 2. Across years, parasite load correlates positively with ambient temperature during the
nestling stage (*i.e.* during the developmental period of *Protocalliphora* larvae) in nests of blue
tits on Corsica. Dots represent yearly average values.

Figure 3

Figure 3. Across years, parasite load (corrected for ambient temperature during nesting)
decreases with increasing mean temperatures in the previous summer (i.e. during the adult,
free-living stage of *Protocalliphora* blow flies). Dots represent yearly mean values.

Figure 4

Figure 4. Across years, parasite load (corrected for ambient temperature during nesting)
increases with increasing minimal temperatures in early spring (*i.e.* during the adult, free-
living stage of *Protocalliphora* blow flies). Dots represent yearly mean values.

Supplementary material

Figure S1. Yearly variation over the 18 years of the study in (A) minimal spring temperature,
(B) average spring temperature, and (C) average summer temperature.

Figure S2. Both average ambient temperature (A) and parasite load (B) during the nestling
stage increase with egg-laying date.