#### gwasurvivr: an R package for genome wide survival analysis

Abbas A Rizvi<sup>1\*</sup>, Ezgi Karaesmen<sup>1\*</sup>, Martin Morgan<sup>2</sup>, Leah Preus<sup>1</sup>, Junke Wang<sup>1</sup>, Theresa Hahn<sup>3</sup>, Michael Sovic<sup>1</sup>, Lara E Sucheston-Campbell<sup>1,4</sup>

<sup>1</sup>College of Pharmacy, The Ohio State University, Columbus, OH, <sup>2</sup>Department of Biostatistics and Bioinformatics, Roswell Park Comprehensive Cancer Center, <sup>3</sup>Department of Medicine, Roswell Park Comprehensive Cancer Center, Buffalo, NY, <sup>4</sup>College of Veterinary Medicine, The Ohio State University, Columbus, OH

\*These authors contributed equally

## ABSTRACT

**Summary:** To address the limited software options for performing survival analyses with millions of SNPs, we developed gwasurvivr, an R/Bioconductor package with a simple interface for conducting genome wide survival analyses using VCF (outputted from Michigan or Sanger imputation servers) and IMPUTE2 files. To decrease the number of iterations needed for convergence when optimizing the parameter estimates in the Cox model we modified the R package survival such that the covariates in the model are first fit without the SNP, and those parameter estimates are used as initial points. We benchmarked gwasurvivr with other GWAS software capable of conducting genome wide survival analysis (genipe, SurvivalGWAS\_SV, and GWASTools). gwasurvivr is significantly faster and shows better scalability as sample size and number of SNPs increase.

**Availability and implementation:** gwasurvivr, including source code, documentation, and vignette are available at: https://github.com/suchestoncampbelllab/gwasurvivr

**Contact:** Abbas Rizvi, <u>Rizvi.33@osu.edu</u>; Lara E Sucheston-Campbell, <u>sucheston-campbell.1@osu.edu</u>

Supplementary information: Supplementary data are available at <a href="https://github.com/suchestoncampbelllab/gwasurvivr\_manuscript">https://github.com/suchestoncampbelllab/gwasurvivr\_manuscript</a>

#### 1 Introduction

Genome-wide association studies (GWAS) are population-level experiments that investigate genetic variation in individuals to observe single nucleotide polymorphism (SNP) associations with a phenotype. Genetic variants tested for association are genotyped on an array and imputed from a reference panel of sequenced genomes eg 1000 Genomes Project or Haplotype Reference Consortium (HRC) (Das, et al., 2016; Genomes Project, et al., 2015). Imputation increases genome coverage from hundreds of thousands or a few million to upwards of 30 million SNPs, improves power to detect genetic associations, and/or homogenizes variant sets for meta-analyses (Das, et al., 2016). Imputed SNPs can be tested for association with binary outcomes (case/control) and quantitative outcomes (e.g., height) using a range of available software packages including SNPTEST (Marchini, et al., 2007) or PLINK (Purcell, et al., 2007). However, existing software options for performing survival analyses , genipe (Lemieux Perreault, et al., 2016), SurvivalGWAS\_SV (Syed, et al., 2017), and GWASTools (Gogarten, et al., 2012) across millions of imputed SNPs either require user interaction with raw output, were not initially designed for survival, have long run times and/or have other limitations

that could deter more introductory users. For these reasons, we developed an R/Bioconductor package, gwasurvivr, for genome wide survival analyses of imputed data in multiple file formats with flexible analysis and output options.

# 2 Implementation

### 2.1 Data structure

Gwasurvivr can analyze IMPUTE2 data or VCF files derived from Michigan or Sanger imputation servers. Data from each are prepared in gwasurvivr by leveraging existing Bioconductor packages GWASTools(Gogarten, et al., 2012) or VariantAnnotation (Obenchain, et al., 2014) depending on the imputation file format.

## IMPUTE2 IMPUTATION

IMPUTE2 format is a standard genotype (.gen) file which store genotype probabilities (GP). We utilized GWASTools in R to compress files into genomic data structure (GDS) format(Gogarten, et al., 2012). This allows for efficient, iterative access to subsets of the data while simultaneously converting GP into dosages (DS) for use in survival analyses.

## MICHIGAN or SANGER SERVER IMPUTATION

VCF files generated from these servers include a DS field and server-specific metafields (INFO score, reference panel allele frequencies) that are iteratively read in by VariantAnnotation(Obenchain, et al., 2014).

#### 2.2 Survival analysis

gwasurvivr implements a Cox proportional hazards regression model (Cox, 1992) to test each SNP with an outcome, with or without covariates and/or SNP-covariate interaction. To decrease the number of iterations needed for convergence when optimizing the parameter estimates in the Cox model we modified the R package survival (Therneau and Grambsch, 2000) such that the covariates in the model are first fit without the SNP, and those parameter estimates are used as initial points (supplementary data). If no additional covariates are added to the model, the parameter estimation optimization begins with null initial value.

Survival analyses are run using genetic data in either VCF or IMPUTE2 formats, which store survival time, survival status and additional covariates in a phenotype file indexed by sample ID. The VCF files contain both genomic data and sample IDs, while IMPUTE2 requires .gen and sample files. Gwasurvivr functions for IMPUTE2 (impute2CoxSurv) and VCF (michiganCoxSurv or sangerCoxSurv) include arguments for the survival model (event of interest, time to event, and covariates), and arguments for quality control that filter on minor allele frequency (MAF) and INFO score. Users can also provide a list of sample IDs for gwasurvivr to internally subset the data, an option not available for SurvivalGWAS\_SV. gwasurvivr outputs two files: (1) .snps\_removed, listing all SNPs that failed QC parameters, including their MAF and INFO score (2) a .coxph file with the results from the analyses, including parameter estimates, p-values, MAF, INFO score for sample set analyzed, the number of events and total sample N for each SNP. gwasurvivr is well suited for multi-core processors and the number of cores

used during computation on Windows and Linux can be specified. gwasurvivr overcomes potential memory limitations that are often attributed to R by iteratively reading in data on subsets of the entire data making it possible to conduct genome-wide analyses on a typical laptop computer.

#### 3 Simulations and benchmarking

We benchmarked gwasurvivr with genipe (Lemieux Perreault, et al., 2016), SurvivalGWAS\_SV (Syed, et al., 2017), and GWASTools (Gogarten, et al., 2012) using data in IMPUTE2 format (the comparison packages do not take VCF) for varying sample sizes (n=100, n=1000, n=5000) and number of SNPs (p=1,000, p=10,000, p=100,000), and including three non-genetic covariates. Survival time and event (alive/dead) were simulated using a normal and binomial distribution, respectively; covariates were simulated using normal distributions. Genetic data were simulated using HAPGENv2 (Su, et al., 2011) with SNP sets selected from chromosome 18 (1000 Genomes NCBI build 36). Benchmarking was performed using identical CPU constraints, 1 node (2.27 GHz Clock Rate) and 8 cores with 24 GB of RAM, on the University at Buffalo Center for Computational Research supercomputer. genipe (Lemieux Perreault, et al., 2016), SurvivalGWAS\_SV (Syed, et al., 2017), and GWASTools (Gogarten, et al., 2012) were performed as specified by the authors in the online manual or vignette (supplementary data).

## 4 Results

All 4 software packages showed excellent agreement between MAF estimates, coefficient estimates, and p-values. For the smallest dataset, the mean runtimes do not differ significantly between the 4 programs (Figure 1, left panel). However, with increasing n and p the mean runtime differs substantially. For the largest dataset, gwasurvivr was faster than genipe (Lemieux Perreault, et al., 2016), SurvivalGWAS\_SV (Syed, et al., 2017), and GWASTools (Gogarten, et al., 2012) by 9.4, 136.3, and 2.5-fold, respectively (Figure 1, right panel).

## FUNDING

This work was supported by the National Institutes of Health/National Heart, Lung, and Blood Institute R01HL102278 [to LSC and TH], The Ohio State University and the Translational Data Analytics Initiative.

## ACKNOWLEDGEMENTS

The authors would like to thank Amy Webb and Guy Brock at The Ohio State University Department of Biomedical Informatics for listening to presentations on and offering suggestions about gwasurvivr. This work was performed in part at the University at Buffalo's Center for Computational Research.

# REFERENCES

Cox, D.R. Regression Models and Life-Tables. Springer; 1992.

Das, S., *et al.* Next-generation genotype imputation service and methods. *Nat Genet* 2016;48(10):1284-1287.

Genomes Project, C., *et al.* A global reference for human genetic variation. *Nature* 2015;526(7571):68-74.

Gogarten, S.M., *et al.* GWASTools: an R/Bioconductor package for quality control and analysis of genome-wide association studies. *Bioinformatics* 2012;28(24):3329-3331. Lemieux Perreault, L.P., *et al.* genipe: an automated genome-wide imputation pipeline with automatic reporting and statistical tools. *Bioinformatics* 2016;32(23):3661-3663. Marchini, J., *et al.* A new multipoint method for genome-wide association studies by imputation of genotypes. *Nat Genet* 2007;39(7):906-913.

Obenchain, V., et al. VariantAnnotation: a Bioconductor package for exploration and annotation of genetic variants. *Bioinformatics* 2014;30(14):2076-2078.

Purcell, S., *et al.* PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 2007;81(3):559-575.

Su, Z., Marchini, J. and Donnelly, P. HAPGEN2: simulation of multiple disease SNPs. *Bioinformatics* 2011;27(16):2304-2305.

Syed, H., Jorgensen, A.L. and Morris, A.P. SurvivalGWAS\_SV: software for the analysis of genome-wide association studies of imputed genotypes with "time-to-event" outcomes. *BMC Bioinformatics* 2017;18(1):265.

Therneau, T.M. and Grambsch, P.M. Modeling Survival Data: Extending the Cox Model. Springer; 2000.

**Figure 1. Runtime for survival analyses**. The x-axis shows the three sample sizes (n=100, 1000, 5000) with SNP set sizes across the top (p=1000, 10000 and 100000). The y-axis is the total runtime in  $log_{10}$ seconds. The mean runtime and 95% confidence intervals (CI) for each n and p combination are show for genipe (red), GWASTools (green), SurvivalGWAS\_SV (purple) and gwasurvivr (cyan). Each of the nine simulations were run in triplicate with identical CPU constraints (1 node, 8 cores).

